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Polyamine-functionalized carbon nanodots: a

novelchemiluminescence probe for selective detection of iron (III) ions

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Experimental

Chemicals: Ethylene imine polymer($C_{2x+4k}H_{5x+10k}N_{x+2k}$, M.W. 1800, 99%) was obtained from Aladdin Industrial Corporation. Citric acid monohydrate (C₆H₈O₇•H₂O) and lead chloride (PbCl₂, chemical pure) were purchased from Beijing Chemical Reagents Company. EDTA-2Na•H₂O was obtained from Beijing Xin Jing Ke Biotechnology Co, Ltd (Beijing, China). NaOH, HCl (36%~38%), Fe(III)chloride hexahydrate (FeCl₃•6H₂O), Manganese(II)chloride tetrahydrate (MnCl₂•4H₂O), Aluminum chloride hexahvdrate (AlCl₃•6H₂O), Copper(II) sulfate pentahydrate (CuSO₄•5H₂O), Iron(II) sulfate heptahydrate (FeSO₄•7H₂O), Magnesium chloride hexahydrate (MgCl₂•6H₂O), Cobalt chloride hexahydrate $(CoCl_2 \bullet 6H_2O),$ cadmium chloride $(CdCl_2 \cdot 2.5H_2O)$, Nickel(II) nitrate hexahydrate (Ni(NO₃)₂•6H₂O) were purchased from Sinopharm Chemical Reagent Co., Ltd (Beijing, China). Calcium chloride anhydrate (CaCl₂) was from Tianjin jinke chemical research institute. Mercuric chloride (HgCl₂) was from Jiangyan universal reagents (Jiangsu, China). Purified deionised water (Millipore Milli-Q, $>18.2M\Omega$ cm-1) was used for the preparation of all aqueous

solutions and rinsing of all laboratory ware. Stock standard solution of Fe(III) was prepared by dissolving 0.2703g FeCl3•6H2O in 10ml 0.005M HCl, both of them were prepared weekly.

Preparation of BPEI-Carbon Dots

The polyamine-functionalized carbon dots (BPEI-CDs) was synthesized by the pyrolysis of citric acid and branched poly(ethylenimine) (BPEI). Firstly, 2.0g BPEI and 4.0g citric acid were dissolved uniformly with 40ml hot water in a 100ml beaker, and then the above admixture was poured into a stainless steel autoclave with a Teflon liner of 100ml capacity and heated at 195°C for 3h. Finally, the reactor was automatically cooled to room temperature. The resulting brown solution was centrifuged at 7000 rpm for 10 min to remove the weight precipitate and agglomerated particles. Then the brown aqueous solution of BPEI-CDs was dialyzed for 8h in super-purified water. The cutoff of the dialysis membrane is equal to molecular weight of 2000. Then the concentration of the acquired uniform aqueous which is about 30mg/ml was determined by freeze-dry method.

Instruments and characterization

The CL kinetic curves were recorded by a BPCL Ultra-Weak Luminescence Analyzer (Institute of Biophysics, Chinese Academy of Sciences, Beijing, China). Fluorescence measurements were performed on a FluoroMax-4 spectrofluorometer (Horiba JobinYvon, Edison, NJ, USA), using 300-500nm excitation and a slit width of 2 nm. UV–vis absorption spectra were measured on an Agilent 8453 UV–visible spectrophotometer (Palo Alto, CA, USA).

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EPR spectra were collected on a Bruke spectrometer (A300-10/12, Bruker, Germany). Highresolution transmission electron microscopy (HRTEM) images were recorded by an electron microscope operating at 120 kV (JEM-2010, JEOL, Japan). Surface chemical bonding state was analyzed by X-ray photoelectron spectroscopy (ESCALAB250Xi, Thermo Scientific, USA). Fourier transform infrared (FT-IR) measurements were carried out with a FT-IR spectrometer (6100, JASCO, Japan). The CL spectra were examined by a series of high-energy optical filters (440,460, 475, 490, 505,535, 555, 575, 590, 605nm).

CL Measurements

The CL kinetic characteristics of carbon dots were obtained by batch experiments, which were achieved by a static system consisted of a glass cuvette and the BPCL Ultra-Weak Luminescence Analyzer. 100µL of carbon dots and 100µLsample were added into the glass cuvette, and then NaOH was injected by a 100µL micro syringe from the upper injection pore.

Analysis of water samples

Two river water samples and a tap water sample were collected from Qing River (Beijing, China) and our laboratory respectively. All the water samples were filtered through $0.22\mu m$ membrane (Millipore) and acidified with HNO₃ to pH 2.3. Then the CL measurements were taken to determine iron concentration within the test samples and spiked samples.



Fig. S1 XPS spetra(A) and the C1S peaks (B) of BPEI-CDs



Fig. S2 FTIR spectrum of CA, BPEI and BPEI-CDs



Fig. S3 HRTEM image of BPEI-CDs (A) and the size distribution of BPEI-CDs (B)



Fig. S4 The PL spectra for BPEI-CDs excited at wavelengths from 300nm to 500nm with 20 nm

increment.



Fig. S5 The UV-vis absorption of BPEI, BPEI+Fe(III), BPEI-CDs, BPEI-CDs+Fe(III), Fe(III)



Fig.S6 Effect of different NaOH concentration and dilution ratio of BPEI-CDs on the improved CL intensity by Fe(III)



Fig S7 The EPR spectrum of BPEI-CDs, NaOH-treated BPEI-CDs and BPEI-CDs after reaction with NaOH and Fe(III)



Fig S8 Fe 2p XPS spectra of BPEI-CDs after reaction with NaOH and Fe(III).

Samples	Blank (M)	CV% (n=3)	Fe(Ⅲ)added (M)	mesured (M)	CV% (n=3)	Recovery (%)
Tap water	4.25×10 ⁻⁷	5.00	5×10 ⁻⁶	5.45×10 ⁻⁶	8.58	100.88
River 1	4.18×10 ⁻⁸	12.80	5×10 ⁻⁶	5.24×10 ⁻⁶	3.61	103.98
River 2	2.87×10 ⁻⁸	6.67	5×10 ⁻⁶	4.67×10 ⁻⁶	11.66	92.89

 Table S1
 Determination of Fe(III) in three water samples