# Polyamine-functionalized carbon nanodots: a novelchemiluminescence probe for selective detection of iron (III) ions 

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## Experimental

Chemicals: Ethylene imine polymer $\left(\mathrm{C}_{2 \mathrm{x}+4 \mathrm{k}} \mathrm{H}_{5 \mathrm{x}+10 \mathrm{k}} \mathrm{N}_{\mathrm{x}+2 \mathrm{k}}\right.$, M.W. 1800, 99\%) was obtained from Aladdin Industrial Corporation. Citric acid monohydrate $\left(\mathrm{C}_{6} \mathrm{H}_{8} \mathrm{O}_{7} \cdot \mathrm{H}_{2} \mathrm{O}\right)$ and lead chloride $\left(\mathrm{PbCl}_{2}\right.$, chemical pure) were purchased from Beijing Chemical Reagents Company. EDTA$2 \mathrm{Na} \cdot \mathrm{H}_{2} \mathrm{O}$ was obtained from Beijing Xin Jing Ke Biotechnology Co, Ltd (Beijing, China). $\mathrm{NaOH}, \mathrm{HCl}(36 \% \sim 38 \%)$, $\mathrm{Fe}(\mathrm{III})$ chloride hexahydrate $\left(\mathrm{FeCl}_{3} \bullet 6 \mathrm{H}_{2} \mathrm{O}\right)$, Manganese( II )chloride tetrahydrate $\left(\mathrm{MnCl}_{2} \bullet 4 \mathrm{H}_{2} \mathrm{O}\right)$, Aluminum chloride hexahydrate $\left(\mathrm{AlCl}_{3} \cdot 6 \mathrm{H}_{2} \mathrm{O}\right)$, Copper( II ) sulfate pentahydrate $\left(\mathrm{CuSO}_{4} \cdot 5 \mathrm{H}_{2} \mathrm{O}\right)$, Iron( II ) sulfate heptahydrate $\left(\mathrm{FeSO}_{4} \cdot 7 \mathrm{H}_{2} \mathrm{O}\right)$, Magnesium chloride hexahydrate $\left(\mathrm{MgCl}_{2} \cdot 6 \mathrm{H}_{2} \mathrm{O}\right)$, Cobalt chloride hexahydrate $\left(\mathrm{CoCl}_{2} \bullet 6 \mathrm{H}_{2} \mathrm{O}\right)$, cadmium chloride $\left(\mathrm{CdCl}_{2} \bullet 2.5 \mathrm{H}_{2} \mathrm{O}\right)$, Nickel(II) nitrate hexahydrate $\left(\mathrm{Ni}\left(\mathrm{NO}_{3}\right)_{2} \cdot 6 \mathrm{H}_{2} \mathrm{O}\right)$ were purchased from Sinopharm Chemical Reagent Co., Ltd (Beijing, China). Calcium chloride anhydrate $\left(\mathrm{CaCl}_{2}\right)$ was from Tianjin jinke chemical research institute. Mercuric chloride $\left(\mathrm{HgCl}_{2}\right)$ was from Jiangyan universal reagents (Jiangsu, China). Purified deionised water (Millipore Milli-Q, $\geq 18.2 \mathrm{M} \Omega \mathrm{cm}-1$ ) was used for the preparation of all aqueous

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solutions and rinsing of all laboratory ware. Stock standard solution of Fe (III) was prepared by dissolving $0.2703 \mathrm{~g} \mathrm{FeCl} 3 \cdot 6 \mathrm{H} 2 \mathrm{O}$ in 10 ml 0.005 M HCl , both of them were prepared weekly.

## Preparation of BPEI-Carbon Dots

The polyamine-functionalized carbon dots (BPEI-CDs) was synthesized by the pyrolysis of citric acid and branched poly(ethylenimine) (BPEI). Firstly, 2.0g BPEI and 4.0 g citric acid were dissolved uniformly with 40 ml hot water in a 100 ml beaker, and then the above admixture was poured into a stainless steel autoclave with a Teflon liner of 100 ml capacity and heated at $195^{\circ} \mathrm{C}$ for 3 h . Finally, the reactor was automatically cooled to room temperature. The resulting brown solution was centrifuged at 7000 rpm for 10 min to remove the weight precipitate and agglomerated particles. Then the brown aqueous solution of BPEI-CDs was dialyzed for 8 h in super-purified water. The cutoff of the dialysis membrane is equal to molecular weight of 2000 . Then the concentration of the acquired uniform aqueous which is about $30 \mathrm{mg} / \mathrm{ml}$ was determined by freeze-dry method.

## Instruments and characterization

The CL kinetic curves were recorded by a BPCL Ultra-Weak Luminescence Analyzer (Institute of Biophysics, Chinese Academy of Sciences, Beijing, China). Fluorescence measurements were performed on a FluoroMax-4 spectrofluorometer (Horiba JobinYvon, Edison, NJ, USA), using 300-500nm excitation and a slit width of 2 nm . UV-vis absorption spectra were measured on an Agilent 8453 UV-visible spectrophotometer (Palo Alto, CA, USA).

EPR spectra were collected on a Bruke spectrometer (A300-10/12, Bruker, Germany). Highresolution transmission electron microscopy (HRTEM) images were recorded by an electron microscope operating at 120 kV (JEM-2010, JEOL, Japan). Surface chemical bonding state was analyzed by X-ray photoelectron spectroscopy (ESCALAB250Xi, Thermo Scientific, USA). Fourier transform infrared (FT-IR) measurements were carried out with a FT-IR spectrometer (6100, JASCO, Japan). The CL spectra were examined by a series of high-energy optical filters (440,460, 475, 490, 505,535, 555, 575, 590, 605nm).

## CL Measurements

The CL kinetic characteristics of carbon dots were obtained by batch experiments, which were achieved by a static system consisted of a glass cuvette and the BPCL Ultra-Weak Luminescence Analyzer. $100 \mu \mathrm{~L}$ of carbon dots and $100 \mu \mathrm{Lsample}$ were added into the glass cuvette, and then NaOH was injected by a $100 \mu \mathrm{~L}$ micro syringe from the upper injection pore.

## Analysis of water samples

Two river water samples and a tap water sample were collected from Qing River (Beijing, China) and our laboratory respectively. All the water samples were filtered through $0.22 \mu \mathrm{~m}$ membrane (Millipore) and acidified with $\mathrm{HNO}_{3}$ to pH 2.3. Then the CL measurements were taken to determine iron concentration within the test samples and spiked samples.


Fig. S1 XPS spetra(A) and the C1S peaks (B) of BPEI-CDs


Fig. S2 FTIR spectrum of CA, BPEI and BPEI-CDs


Fig. S3 HRTEM image of BPEI-CDs (A) and the size distribution of BPEI-CDs (B)


Fig. S4 The PL spectra for BPEI-CDs excited at wavelengths from 300 nm to 500 nm with 20 nm increment.


Fig. S5 The UV-vis absorption of BPEI, BPEI+Fe(III), BPEI-CDs, BPEI-CDs+Fe(III), Fe(III)


Fig.S6 Effect of different NaOH concentration and dilution ratio of BPEI-CDs on the improved CL intensity by Fe(III)


Fig S7 The EPR spectrum of BPEI-CDs, NaOH-treated BPEI-CDs and BPEI-CDs after reaction with NaOH and $\mathrm{Fe}(\mathrm{III})$


Fig S8 Fe 2p XPS spectra of BPEI-CDs after reaction with NaOH and Fe (III).

Table S1 Determination of Fe(III) in three water samples

| Samples | Blank ( <br> M) | $\begin{aligned} & \text { CV\% } \\ & \mathrm{n}=3) \end{aligned}$ | Fe(III)added <br> (M) | mesured <br> (M) | $\begin{aligned} & \text { CV\% } \\ & \mathrm{n}=3 \text { ) } \end{aligned}$ | Recovery (\%) |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Tap water | $4.25 \times 10^{-7}$ | 5.00 | $5 \times 10^{-6}$ | $5.45 \times 10^{-6}$ | 8.58 | 100.88 |
| River 1 | $4.18 \times 10^{-8}$ | 12.80 | $5 \times 10^{-6}$ | $5.24 \times 10^{-6}$ | 3.61 | 103.98 |
| River 2 | $2.87 \times 10^{-8}$ | 6.67 | $5 \times 10^{-6}$ | $4.67 \times 10^{-6}$ | 11.66 | 92.89 |

