

**Electronic Supplementary Information (ESI)**

**Design, synthesis and biocidal effect of novel amine N-halamine microspheres based on 2,2,6,6-tetramethyl-4-piperidinol as promising antibacterial agents**

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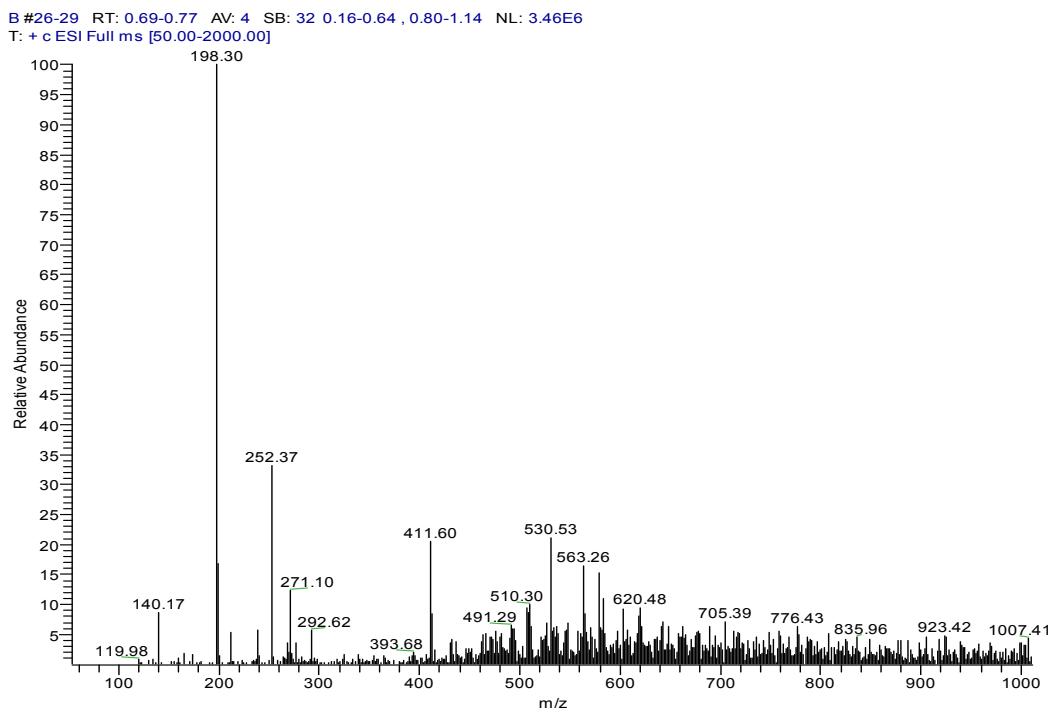
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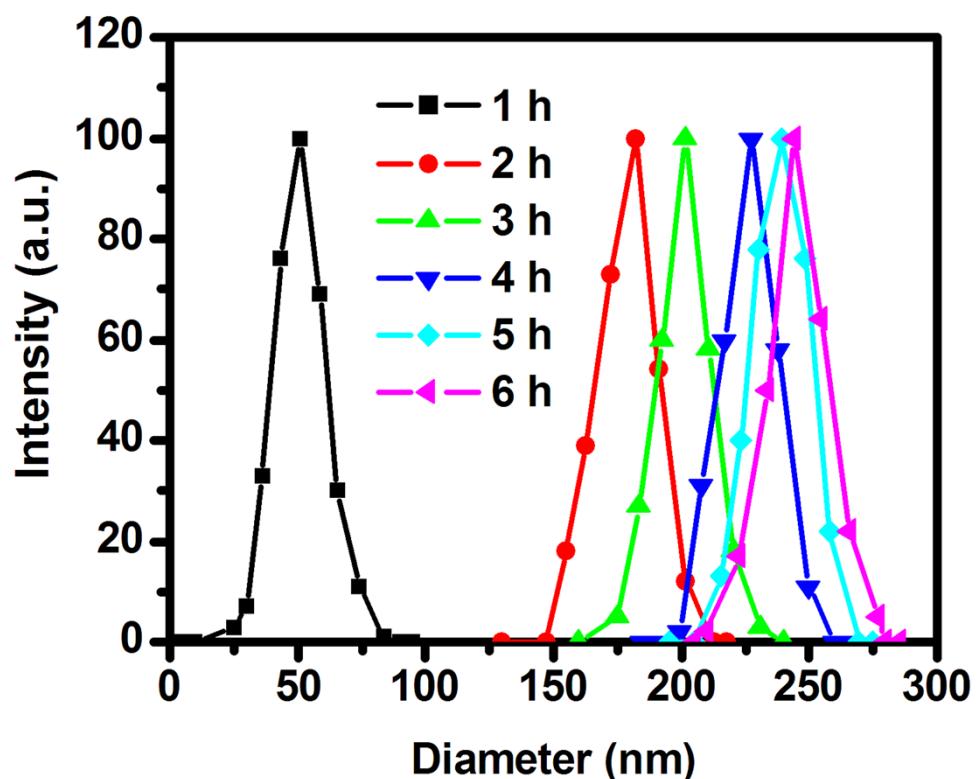
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**Fig. S1.** Mass spectrogram of ATMP



**Fig. S2.** Particle size distribution of poly(ATMP-*co*-MMA) microspheres with different copolymerization period.

**Table S1.** Dissociation constant of different *N*-halamine in aqueous solution

<i>N</i> -halamine type	Dissociation reaction	Dissociation constant <sup>a</sup>
<b>Imide</b>		< 10 <sup>-4</sup>
<b>Amide</b>		< 10 <sup>-9</sup>
<b>Amine</b>		< 10 <sup>-12</sup>

<sup>a</sup>Dissociation constant was from references.<sup>1-4</sup>

**Table S2.** Particle size and surface area characteristics of poly(ATMP-*co*-MMA) microspheres formed with different copolymerization period

Sample	Copolymerization period (h)	Particle size (nm) <sup>a</sup>		Surface area <sup>b</sup>
		Size distribution	Average size	(m <sup>2</sup> ·g <sup>-1</sup> )
S1	1	20-90	50.7	118.3
S2	2	150-210	181.8	33.0
S3	3	170-240	201.9	29.7
S4	4	190-250	227.5	26.4
S5	5	210-260	239.2	25.1
S6	6	210-280	244.5	24.5

<sup>a</sup>Particle size was determined by TEM images.

<sup>b</sup>The surface area was calculated based on the assumption that the particles are non-porous spheres with density of

1.0 g·cm<sup>-3</sup>. The calculation was performed according to the following equation:  $S = 6(D \cdot d)^{-1}$ , wherein  $S$  is the

surface area (m<sup>2</sup>·g<sup>-1</sup>);  $D$  is the diameter (μm); and  $d$  is the density (g·cm<sup>-3</sup>) of the particles.<sup>5</sup>

**Table S3.** Minimum inhibitory concentration (MIC) of different products against *S. aureus*, *B. subtilis*, *E. coli*, and *P. aeruginosa*

Sample	MIC (mg/mL)				Reference	
	Gram-positive bacteria		Gram-negative bacteria			
	<i>S. aureus</i>	<i>B. subtilis</i>	<i>E. coli</i>	<i>P. aeruginosa</i>		
PSA@Fe <sub>3</sub> O <sub>4</sub> @SiO <sub>2</sub> - <i>N</i> -halamine <sup>a</sup>	80	-	-	60	6	
HMNH NPs <sup>b</sup>	160	160	160	80	7	
BAMNH NPs <sup>c</sup>	80	80	80	40	7	
H-NHFS NPs <sup>d</sup>	160	-	-	80	8	
BA-NHFS NPs <sup>e</sup>	40	-	-	40	8	
Amine <i>N</i> -halamine microspheres	10	-	10	-	This study	

<sup>a</sup>Amide *N*-halamine-immobilized PSA@Fe<sub>3</sub>O<sub>4</sub>@SiO<sub>2</sub> nanoparticles.

<sup>b</sup>Hydantoin-structural magnetic amide *N*-halamine nanoparticles.

<sup>c</sup>Barbituric acid-based magnetic imide *N*-halamine nanoparticles.

<sup>d</sup>Hydantoin-originated amide *N*-halamine-functionalized silica nanoparticles.

<sup>e</sup>Barbituric acid-originated imide *N*-halamine-functionalized silica nanoparticles.

**Table S4.** Oxidative chlorine content of amine *N*-halamine microspheres with different soaking period

Soaking period (h) <sup>a</sup>	Oxidative chlorine (%) <sup>b</sup>	Reduction (%) <sup>c</sup>
0	1.17	0
6	1.16	0.85
12	1.16	0.85
18	1.14	2.56
24	1.11	5.13
30	1.10	5.98
36	1.08	7.69
42	1.08	7.69
48	1.07	8.55

<sup>a</sup>Period since initial soaking.

<sup>b</sup>Oxidative chlorine content was determined by the iodometric/thiosulfate titration after a certain soaking age.

<sup>c</sup>Reduction percentage of oxidative chlorine after a certain soaking age.

### **Supplementary References**

- 1 K. Tan and S. K. Oberndorf, *J. Membrane Sci.*, 2007, **305**, 287-298.
- 2 A. Akdag, S. Okur, M. L. McKee and S. D. Worley, *J. Chem. Theory Comput.*, 2006, **2**, 879-884.
- 3 K. Barnes, J. Liang, R. Wu, S. D. Worley, J. Lee, R. M. Broughton and T. S. Huang, *Biomaterials*, 2006, **27**, 4825-4830.
- 4 O. Gutman, M. Natan, E. Banin and S. Margel, *Biomaterials*, 2014, **35**, 5079-5087.
- 5 M. Omer-Mizrahi and S. Margel, *J. Colloid Interface Sci.*, 2009, **329**, 228-234.
- 6 A. Dong, S. Lan, J. Huang, T. Wang, T. Zhao, L. Xiao, W. Wang, X. Zheng, G. Gao and Y. Chen, *ACS Appl. Mater. Interfaces*, 2011, **3**, 4228-4235.
- 7 A. Dong, Y. Sun, S. Lan, Q. Wang, Q. Cai, X. Qi, Y. Zhang, G. Gao, F. Liu and C. Harnoode, *ACS Appl. Mater. Interfaces*, 2013, **5**, 8125-8133.
- 8 A. Dong, M. Xue, S. Lan, Q. Wang, Y. Zhao, Y. Wang, Y. Zhang, G. Gao, F. Liu and C. Harnoode, *Colloid Surf. B*, 2014, **113**, 450-457.