Host-guest assemble of adamantyl tethered squaraine in βcyclodextrin for monitoring pH in living cells

Wenjian Xu,^{a,b} Xiaochan Zhu,^{a,b} Guimei Wang,^{a,b} Chuanguo Sun,^{a,b} Qingfeng Zheng,^{a,b}

Huanghao Yang^{b,}* and Nanyan Fu^{*a,b,**}

^aResearch Institute of Photocatalysis, State Key Laboratory of Photocatalysis on Energy and

Environment, Fuzhou University, Fuzhou 350002, P. R. China.

^bKey Laboratory of Analysis and Detection for Food Safety, Ministry of Education & Fujian

Provincial Key Laboratory of Analysis and Detection Technology for Food Safety, and Department

of Chemistry, Fuzhou University, Fuzhou 350108, P. R. China.

*E-mail: nanyan_fu@fzu.edu.cn

Supplementary Supporting Information

Contents:

1.	Experimental details	S1-S2
2.	Characterization of 2 and SQ	S3-S6
3.	Change in absorbance and fluorescence of \boldsymbol{SQ} with increase of H_2O percentage in	n EtOH
	solution	S6-S7
4.	Dynamic light scattering experiments	S7-S9
5.	Change in absorbance of SQ in water solution containing varied concentration of β -CE) S9
6.	ESI mass spectrum of SQ $\subset\beta$ -CD inclusion complex	S10
7.	Trifloroacetic acid titrations of SQ in MeCN	S10
8.	¹ H NMR titration experiment	S11
9.	ESI mass spectrum of SQ &OH adduct⊂β-CD	S11
10.	Change in absorbance of $SQ \subset \beta$ -CD in varied pH buffers	S12
11.	Reversibility of pH titrations	S13
12.	Photobleaching experiments of SQ	S14
13.	Competition experiments	S15
14.	The study of SQ in different concentrations in pure water	S16
15.	The control experiment of pH-dependence of SQ fluorescence spectra in PB buffer a	solution
	without β-CD	S16
16.	The control experiment of fluorescence images of \boldsymbol{SQ} without $\beta\text{-}CD$ in pH 4.5 and 8.0	S17

1. Experimental details

1.1. Materials and general methods

The ¹H NMR spectra were recorded on Bruker AV-400 and JEOL JNM LA-500 spectrometer and the chemical shifts were measured relative to TMS (0.00 ppm). FTIR spectra were recorded on a Perkin Elmer Spectrum 2000 Fourier Transform Infrared Spectrophotometer. MS and HRMS were recorded on DECAX-30000 LCQ Deca XP Ion Trap Mass Spectrometer and Applied Biosystems (Sciex) QStar Mass Spectrometer by positive ESI-Q-TOF, respectively. Absorption spectra were detected on a Perkin Elmer Lambda750 UV spectrophotometer. Fluorescent emission spectra were collected on a Cary Edipse fluorescence spectrophotometer. Melting points of compounds were determined with SGW X-4 and were uncorrected. All the solvents were redistilled before use. Except for specific note, other chemicals and reagents were obtained from commercial suppliers and used without further purification. The syntheses and manipulations of squaraine dyes were carried out under dry N₂ atmosphere.

1.2. Synthesis

1.2.1. Synthesis of N-(2-adamantyl)aniline (2)

2-Adamantanone (1) (1.50 g, 10 mmol) was ground with aniline (0.93 g, 10 mmol) for 10 min in an agate mortar and a pestle at room temperature under solvent-free conditions. To the resulting mixture was added NaBH₄ (0.38 g, 10 mmol) and boric acid (0.62 g, 10 mmol), and then the mixture was ground under identical conditions until TLC showed complete disappearance of the starting ketone **1**. The reaction mixture was quenched with water (30 mL) and extracted with CH₂Cl₂. The combined extract was dried over anhydrous Na₂SO₄, filtered and concentrated. The crude products obtained were further purified by column chromatography on silica gel with ethyl acetate/petroleum ether (1/30 v/v) to afford a white solid (1.41 g, 6.20 mmol). Yield: 62%. m.p. 57-58 °C [1]. IR (KBr): 3419, 2903, 2842, 1599, 1498, 1445, 1420, 1311, 1128, 747, 688 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 1.58-1.61 (d, *J* = 12.3 Hz, 2H) , 1.75 (s, 2H), 1.81-1.84 (d, *J* = 10.4 Hz, 4H), 1.89-1.94 (dd, *J* = 14.1, 7.9 Hz, 5H), 2.03 (s, 2H), 3.54 (s, 1H), 6.60 (d, *J* = 8.0 Hz, 2H), 6.65 (t, *J* = 7.3 Hz, 1H), 7.15 (t, *J* = 7.8 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 27.35, 27.49, 31.62, 31.63, 37.43, 37.74, 56.80, 113.04, 116.70, 129.27, 147.41.

1.2.2. Synthesis of SQ

A mixture of 2 (114 mg, 0.5 mmol), squaric acid (29 mg, 0.25 mmol), toluene (10 mL) and n-

butanol (10 mL) was refluxed at 120 °C for 24 h under nitrogen while the water formed was azeotropically removed by using a Dean–Stark trap. After cooling to room temperature, the solvent was removed under reduced pressure and then the residue was purified by column chromatography on silica gel with ethyl acetate/petroleum ether (1:4, v/v) as eluent and eluting again with methanol/chloroform (1:100 to 1:30, v/v) to afford a blue solid (47 mg, 0.088 mmol). Yield: 35%. m.p. >300 °C. IR (KBr): 3296, 2898, 2845, 1611, 1583, 1538, 1485, 1465, 1384, 1163, 1107, 846, 804, 750, 518 cm⁻¹; ¹H NMR (400 MHz, CDCl₃/CD₃OD, 5:1, v/v): δ 1.67-1.70 (d, *J* = 12.6 Hz, 4H), 1.80 (s, 4H), 1.87-1.97 (dd, *J* = 29.6, 11.8 Hz, 18H), 2.08 (s, 4H), 3.76 (s, 2H), 6.73 (d, *J* = 7.7 Hz, 4H), 8.25 (d, *J* = 7.6 Hz, 4H); ¹³C NMR (100 MHz, CDCl₃/CD₃OD, 5:1, v/v) δ 27.22, 27.24, 31.52, 31.76, 37.21, 37.46, 57.04, 113.80, 119.92, 133.75, 154.49, 184.42, 184.92; ESI-MS: *m/z* 531.9 ([M-H]⁻). ESI-HRMS: Calcd for C₃₆H₄₁N₂O₂ ([M+H]⁺): 533.3168, Found: 533.3173.

[1] Averin, A. D. Palladium-catalyzed amination of isomeric dihalobenzenes with 1- and 2aminoadamantanes. *Russian Journal of Organic Chemistry* **2010**, *46*, 64-72.

2. Characterization of $\mathbf{2}$ and \mathbf{SQ}



Figure S1. The IR spectrum of 2 in KBr disc.



Figure S2. The ¹H NMR spectrum of 2 in CDCl₃ (400 MHz).



Figure S3. The ¹³C NMR spectrum of 2 in CDCl₃ (100 MHz).



Figure S4. The IR spectrum of SQ in KBr disc.



Figure S5. The ¹H NMR spectrum of SQ in CDCl₃/CD₃OD (5:1, v/v) (400 MHz).



Figure S6. The ¹³C NMR spectrum of SQ in CDCl₃/CD₃OD (5:1, v/v) (100 MHz).



Figure S7. The ESI mass spectrum of SQ.



Figure S8. The ESI-HR mass spectrum of SQ.

3. Change in absorbance and fluorescence of \mathbf{SQ} with increase of H_2O percentage in EtOH solution





Figure S9. Change in absorbance (A) and fluorescence (B) spectra of SQ (2.0 μ M) with increase of H₂O percentage in EtOH solution. (λ_{ex} =610 nm, slit: 5 nm/5 nm, PMT Volts: 650 V.).

4. Dynamic light scattering experiments

A:	SQ (1.0	μM)	in	pure	water
		· · · · · · · · · · · · · · · · · · ·				

Sample ID Operator ID Elapsed Time Mean Diam. Rel. Var. Skew RmsError	1 Unkn 00:02 57.4 0.077 0.581 2.658	own Op 2:04 (nm) 7 36e-03	erator			100 50 0 6.0		Diamo	500.0 eter (nm)
d 20.68 23.52 26.74 30.84 34.66 38.96 43.78 49.20 55.30 62.14 69.84	G(d) 0 16 31 52 75 93 100 93 75	C(d) 0 3 8 16 28 42 58 72 84	d 78.49 88.21 99.14 124.90 142.01 161.48	G(d) 52 31 16 0 0 0	C(c 92 97 100 100 100	i) d	G(d)	C(d)	Print Window Copy For Spreadsheet Copy to Clipboard Close

Sample ID Operator ID Elapsed Time Mean Diam. Rel. Var. Skew RmsError	2 Unkn 00:02 49.5 0.091 0.814 4.151	own Op 2:13 (nm) I 4 17e-03	erator			100 50 0 5.0		Diam	500.0 eter (nm)
d	G(d)	C(d)	d	G(d)	Crdh	d	G(d)	C(d)	
17.85	0	Ó	78.40	26	97				
20.51	0	0	89.05	12	99				
24.92	12	2	101.14	5	100				
28.30	26	6	124.85	0	100				
32.14	47	14	143.46	0	100				
36.51	72	26	164.84	0	100				Print Window
41.47	92	41							
47.10	100	58							Copy For Spreadsheet
53.50	92	73							Course Click and
60.77	72	85							Lopy to Clipboard
69.02	47	93							Close

B: SQ (1.0 μ M) with β -CD (0.1 mM) in pure water

C: **SQ** (1.0 μ M) with β -CD (1.0 mM) in pure water

Sample ID Operator ID Elapsed Time Mean Diam. Rel. Var. Skew RmsError	3 Unkn 00:09 2315 2.477 1.295 8.082	own O; 3:40 .5 (nm) 7 5 28e-03	perator			100 100 50 100 5.00	De-02	Diame	50000.0 eter (nm)
d 0.17 0.33 0.66 1.31 2.61 5.19 10.32 20.53 40.82 81.20 161.51	G(d) 0 25 100 11 0 0 0 0 0 0	C(d) 0 10 48 53 53 53 53 53 53 53	d 321.24 638.94 1270.86 2527.73 5027.66 10000.00	G(d) 23 23 0 0 38 38 38	C(62 70 70 70 85 100	(k	G(d)	C(d)	Print Window Copy For Spreadsheet Copy to Clipboard Close

Sample ID Operator ID Elapsed Time Mean Diam. Rel. Var. Skew RmsError	4 Unkn 00:00 275.8 29.14 5.700 2.541	own O;):38 3 (nm) 44 2 17e-02	perator			100 100 50 0 5.0	0e-02	Diamo	50000.0 eter (nm)
d	G(d)	C(d)	d	G(d)	C(d)	d	G(d)	C(d)	
0.10	14	9	273.84	0	96				
0.21	100	75	562.34	0	96				
0.42	32	96	1154.78	0	96				
0.87	0	96	2371.37	0	96				
1.78	0	96	4869.67	3	98				
3.65	0	96	10000.00	3	100				Print Window
7.50	0	96							
15.40	0	96							Copy For Spreadsheet
31.62	0	96							
64.94	0	96							Copy to Clipboard
133.35	0	96							Close

D: SQ (1.0 μ M) with β -CD (5.0 mM) in pure water

Figure S10. Solvodynamic diameters of SQ (1.0 μ M) (A), SQ (1.0 μ M) with β -CD (0.1 mM) (B), SQ (1.0 μ M) with β -CD (1.0 mM) (C), and SQ (1.0 μ M) with β -CD (5.0 mM) (D) in pure water determined by dynamic light scattering.

5. Change in absorbance of \boldsymbol{SQ} in water solution containing varied concentration of $\beta\text{-}$ CD



Figure S11. Absorbance spectra of SQ (2.0 μ M) in different concentrations of β -CD solution.



6. ESI mass spectrum of **SQ** $\subset\beta$ -CD inclusion complex

Figure S12. The ESI mass spectrum of SQ $\subset\beta$ -CD inclusion complex [SQ $\subset\beta$ -CD+Na⁺]⁺. (Calcd for C₁₂₀H₁₈₀N₂NaO₇₂ ([SQ+2 β -CD+Na⁺]⁺): 2824.0383, Found: 2824.0.).

7. Trifloroacetic acid titrations of SQ in MeCN



Figure S13. Change in absorbance of SQ (2.0 μ M) with the increase of TFA (0-10 μ L) in CH₃CN.



Figure S14. Change in absorbance of SQ (3.75 μ M) with the increase of TFA (0-90 μ L) in CH₃CN.

8. ¹H NMR titration experiment



Figure S15. The ¹H NMR spectra of SQ (3.0 mg) (1), and SQ (3.0 mg) and D_2SO_4 (1.0 μ L) (2) in 0.6 mL CDCl₃/CD₃OD (5:1, v/v) (400 MHz).



Figure S16. The ESI mass spectrum of SQ&OH adduct $\subset\beta$ -CD. (Calcd for C₇₈H₁₁₃N₂O₃₉ ([SQ+ β -CD+H₂O+OH⁻]⁻): 1701.6920, Found: 1701.8.).

10. Change in absorbance of **SQ** $\subset\beta$ -CD in varied pH buffers



Figure S17. (A) pH-Dependence of absorption spectra of **SQ** (2.0 μ M) in PB (20 mM) buffer solution with β -CD (2.0 mM). **(B)** pH-Dependence absorbance changes in PB (20 mM) buffer solution at 628 nm.

11. Reversibility of pH titrations



Figure S18. Fluorescence titration spectra of SQ $\subset\beta$ -CD with NaOH and HCl. (λ_{ex} =610 nm, λ_{em} =644 nm, slit: 5 nm/5 nm, PMT Volts: 650 V.).



Figure S19. The absorbance response of SQ⊂β-CD upon the alter addition of NaOH and HCl at 628 nm.



Figure S20. (A) Time dependence of absorption spectra of SQ (2.0 μM) in CHCl₃ irradiated under tungsten lamp (500 W) at 40 cm distance. (B) The absorption decay of SQ (2.0 μM) at 615 nm in CHCl₃ irradiated under tungsten lamp (500 W) at 40 cm distance.



Figure S21. The fluorescence response of SQ (2.0 μM) toward 100 μM different metal ions (A) and anions (B) in pH=5.0 (red bar) and 8.0 (black bar) PB buffer solution with β-CD (2.0 mM). $(\lambda_{ex}=610 \text{ nm}, \lambda_{em}=644 \text{ nm}, \text{slit: 5 nm/5 nm}, \text{PMT Volts: 650 V.}).$

14. The study of SQ in different concentrations in pure water



Figure S22. Change in absorbance spectra with increase the concentration of SQ (0.5-3.5 μ M) in pure water.



Figure S23. Change in flourescence spectra with increase the concentration of SQ (0.5-3.5 μ M) in pure water. (λ_{ex} =610 nm, slit: 5 nm/5 nm, PMT Volts: 650 V.).

15. The control experiment of pH-dependence of SQ fluorescence spectra in PB buffer solution without β -CD



Figure S24. The control experiment of pH-dependence of **SQ** (2.0 μ M) fluorescence spectra in PB (20 mM) buffer solution without β -CD. (λ_{ex} =610 nm, slit: 5 nm/5 nm, PMT Volts: 650 V.). 16. The control experiment of fluorescence images of **SQ** without β -CD in pH 4.5



Figure 25. Fluorescence images of living HeLa cells. (a) Bright-field image of HeLa cells incubated with SQ⊂β-CD in PBS medium at pH 4.5; (b) Fluorescence image of (a); (c) the overlay image of (a) and (b); (d) Bright-field image of HeLa cells incubated with SQ in the absence of β-CD in PBS medium at pH 4.5; (e) Fluorescence image of (d); (f) the overlay image of (d) and (e); (g) Bright-field image of HeLa cells incubated with SQ in the absence of β-CD in PBS medium at pH 8.0; (h) Fluorescence image of (g); (i) the overlay image of (g) and (h).