

## A rapid and integrated pyramid screening method to classify and identify complex endogenous substances with UPLC/Q-TOF MS-based metabolomics

**Yubo Li,<sup>a</sup> Zhenzhu Zhang,<sup>a</sup> Zhiguo Hou,<sup>a</sup> Lei Wang,<sup>a</sup> Xin Wu,<sup>a</sup> Liang Ju,<sup>a</sup> Xiuxiu Zhang<sup>a</sup> and Yanjun Zhang<sup>\*a</sup>**

<sup>a</sup>Tianjin State Key Laboratory of Modern Chinese Medicine, School of Traditional Chinese Materia Medica, Tianjin University of Traditional Chinese Medicine, 312 Anshan west Road, Tianjin 300193, China.

\*Author for correspondence:

E-mail: tianjin\_tcm001@sina.com.

Tel and Fax number: +86-22-59596223.

**Table S1.** Summary of methods for the data processing of different classes of substances and corresponding references depending on LC-MS.

Compound classification	Subclass	Parent ion	Diagnostic fragments (m/z)	Diagnostic fragment mass defect range ( $\pm 10\text{ppm}$ )	Neutral loss (DA)	Mass rang (DA)	Mass defect range	References
Purines and Pyrimidines		[M+H] <sup>+</sup>		17.02(NH <sub>3</sub> ) 43.00(HNCO)		111.0-181.1	0.0272-0.0702	1-4
Nucleosides		[M+H] <sup>+</sup>		116.04(C <sub>5</sub> H <sub>8</sub> O <sub>3</sub> ) <sup>a</sup> 132.04(C <sub>5</sub> H <sub>8</sub> O <sub>4</sub> ) <sup>b</sup>		227.0-311.2	0.0695-0.1230	
Nucleic acids	Nucleotides					Contains a phosphate group 304.0-368.1	0.0253-0.0682	
						Contains two phosphate groups 387.0-443.1	0.0021-0.0346	
						Contains three phosphate groups 466.9-524.0	-0.0314-0.0009	
Steroid hormones	Estrogens	[M-H] <sup>-</sup>	145.06[C <sub>10</sub> H <sub>9</sub> O] <sup>c</sup>	0.0639-0.0667		270.1-288.2	0.1412-0.1776	5-7
	Androgens		97.06[C <sub>6</sub> H <sub>9</sub> O] <sup>d</sup>	0.0643-0.0663		286.1-304.3	0.1725-0.2090	
	Progesteragens							
	Glucocorticoids	[M+H] <sup>+</sup>	109.06[C <sub>7</sub> H <sub>9</sub> O] <sup>e</sup>	0.0642-0.0664		312.2-378.3	0.1885-0.2403	8-11
	Mineralocorticois		121.06[C <sub>8</sub> H <sub>9</sub> O] <sup>f</sup>	0.0640-0.0666				
Amino acids		[M+H] <sup>+</sup>			17.02(NH <sub>3</sub> ) 46.00(HCOOH) 18.01(H <sub>2</sub> O)	75.0-292.2	-0.0081-0.2100	12-14

		[M-H] <sup>-</sup>		17.02(NH <sub>3</sub> ) 43.98(CO <sub>2</sub> )			
Monosaccharides	Aldoses and Ketoses	[M-H] <sup>-</sup>		18.01(H <sub>2</sub> O) <sup>g</sup> 60.02(C <sub>2</sub> H <sub>4</sub> O <sub>2</sub> ) <sup>g</sup>	150.0-180.1	0.0527-0.0686	15
Organic acids	Carboxylic acids <sup>h</sup>	[M-H] <sup>-</sup>		43.98(CO <sub>2</sub> )	60.0-230.2	0.0003-0.1519	16,17
	Lysophosphatidylcholine (LPC)	[M+H] <sup>+</sup>	184.07[C <sub>5</sub> H <sub>15</sub> NO <sub>4</sub> P] <sup>+</sup> 104.10[C <sub>5</sub> H <sub>14</sub> NO] <sup>+</sup>	0.0720-0.0758 0.1064-0.1086	412.2-607.5	0.2464-0.4577	18,20,21, 24,26
	Phosphatidylcholine (PC)	[M+H] <sup>+</sup>	184.07[C <sub>5</sub> H <sub>15</sub> NO <sub>4</sub> P] <sup>+</sup> 104.10[C <sub>5</sub> H <sub>14</sub> NO] <sup>+</sup>	0.0720-0.0758 0.1064-0.1086	673.4-957.9	0.4682-0.8126	18,20-22, 24,26,27
	Sphingomyelin (SM)	[M+H] <sup>+</sup>	184.07[C <sub>5</sub> H <sub>15</sub> NO <sub>4</sub> P] <sup>+</sup> 104.10[C <sub>5</sub> H <sub>14</sub> NO] <sup>+</sup>	0.0720-0.0758 0.1064-0.1086	646.5-843.8	0.5049-0.7320	19-22,24
	Lysophosphatidylethanolamine (LPE)	[M+H] <sup>+</sup>		141.01(C <sub>2</sub> H <sub>8</sub> NO <sub>4</sub> P)	423.2-565.5	0.2385-0.4108	18,20,21
	Phosphatidylethanolamine (PE)	[M+H] <sup>+</sup>		141.01(C <sub>2</sub> H <sub>8</sub> NO <sub>4</sub> P)	631.4-915.8	0.4213-0.7657	18,20-23, 26,27
Phospholipids	Lysophosphatidic acid (LPA)	[M-H] <sup>-</sup>	152.99[C <sub>3</sub> H <sub>6</sub> PO <sub>5</sub> ] <sup>-</sup>	0.9937-0.9968	410.2-438.3	0.2433-0.2954	27,28
	Phosphatidic acid (PA)	[M-H] <sup>-</sup>	152.99[C <sub>3</sub> H <sub>6</sub> PO <sub>5</sub> ] <sup>-</sup>	0.9937-0.9968	648.4-700.6	0.4730-0.5251	27,28
	Phosphatidylserine (PS)	[M+H] <sup>+</sup>		185.00(C <sub>3</sub> H <sub>8</sub> NO <sub>6</sub> P)	675.4-879.6	0.4111-0.5990	20,22,24, 29
		[M-H] <sup>-</sup>		87.07(C <sub>3</sub> H <sub>5</sub> NO <sub>2</sub> )			22,25,27, 30
	Phosphatidylinositol (PI)	[M+H] <sup>+</sup>		260.02(C <sub>6</sub> H <sub>13</sub> PO <sub>9</sub> )	806.4-914.6	0.4945-0.5885	20,21,24, 26
		[M-H] <sup>-</sup>	241.01[C <sub>6</sub> H <sub>10</sub> PO <sub>8</sub> ] <sup>-</sup>	0.0088-0.0138			22,23,27

Phosphatidylglycerol (PG)	$[M+H]^+$		172.01( $C_3H_9PO_6$ )			20,24
	$[M-H]^-$	152.99( $C_3H_6PO_5$ ) <sup>c</sup>	0.9937-0.9968	718.4-826.6	0.4784-0.5724	27
Ceramide				481.4-679.7	0.4494-0.6843	

<sup>a</sup> Deoxynucleosides produce the neutral fragment of m/z 116.04 ( $C_5H_8O_3$ ) after collision induced dissociation (CID) process in the positive mode.

<sup>b</sup> Nucleosides produce the neutral fragment of m/z 132.04 ( $C_5H_8O_4$ ) after the CID process in the positive mode.

<sup>c</sup> Estrogen that A ring is a phenyl group and the C<sub>3</sub>-position that only replaces the phenolic hydroxyl group can generate m/z 145.06 [ $C_{10}H_9O$ ]<sup>-</sup> in the negative mode.

<sup>d</sup> Androgens, progestagens, glucocorticoids, and mineralocorticoids that A ring have  $\Delta^4$ -3-ketone group and C<sub>4</sub>-position containing ethylenic bonds generate m/z 97.06 [ $C_6H_9O$ ]<sup>+</sup> in the positive mode, except C<sub>6</sub>-position.

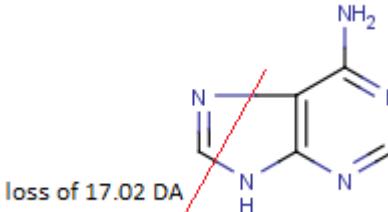
<sup>e</sup> Compound that meets "d" and C<sub>6</sub>-position with methyl generate m/z 109.06 [ $C_7H_9O$ ]<sup>+</sup> in the positive mode.

<sup>f</sup> Compound that meets "d" and C<sub>6</sub>-position without methyl generate m/z 121.09 [ $C_8H_9O$ ]<sup>+</sup> in the positive mode.

<sup>g</sup> Ketose and aldose (5–6 carbon atoms) can produce m/z 18.01 ( $H_2O$ ) and m/z 60.02 ( $C_2H_4O_2$ ) in the negative mode.

<sup>h</sup> Short-chain and medium-chain of carboxylic acids

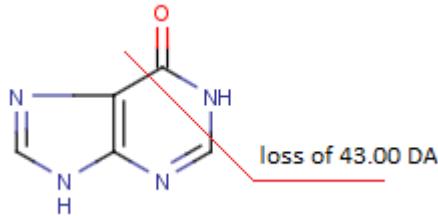
**Table S2.** Fragmentation rules of different classification of substances.

NO.	Endogenous substance	Mode	Structure
1	Adenine	Positive	

---

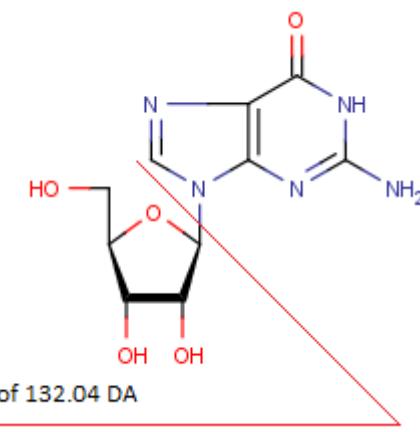
2      Hypoxanthine

Positive



3      Guanosine

Positive

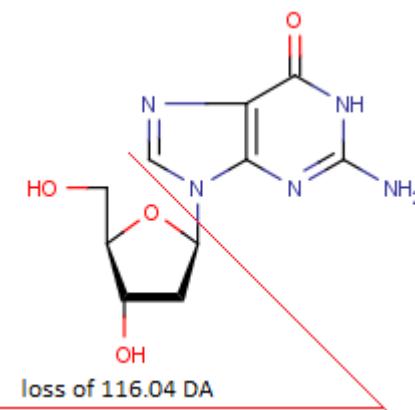


---

4

2'-Deoxyguanosine

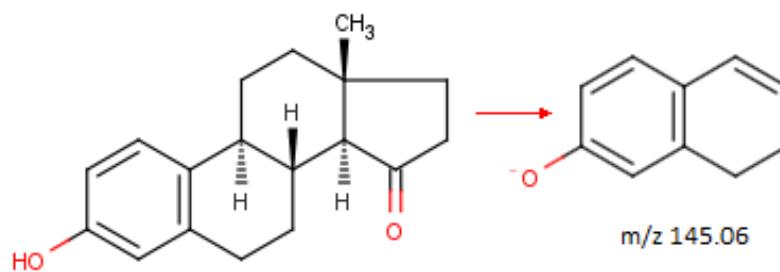
Positive



5

Estrone

Negative

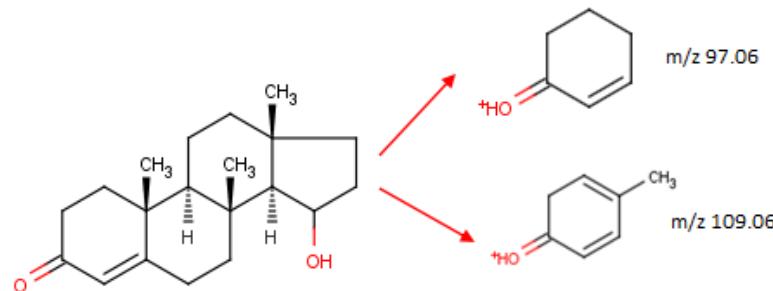


---

6

Testosterone

Positive

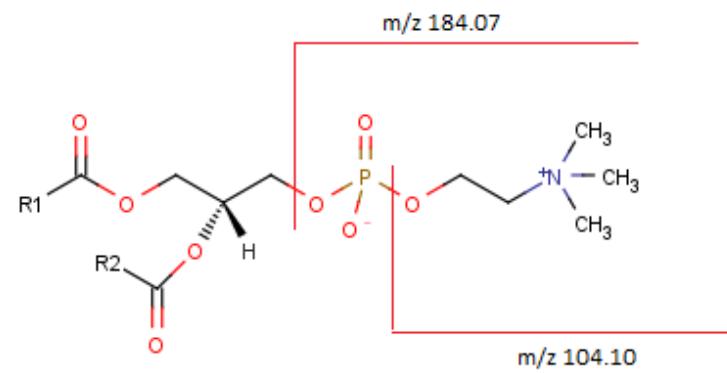


---

7

Phosphatidylcholine  
(PC)

Positive

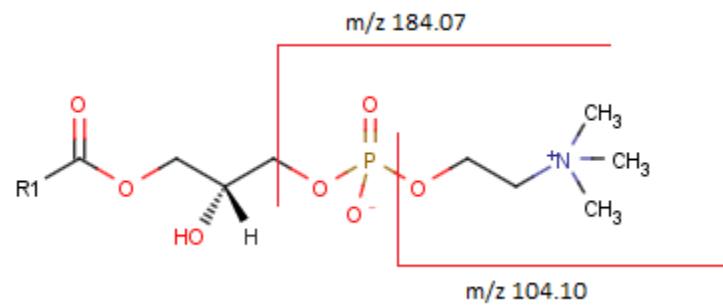


---

8

Plasmenylcholine  
(LPC)

Positive

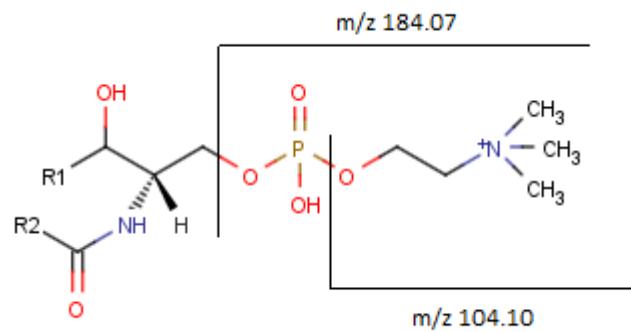


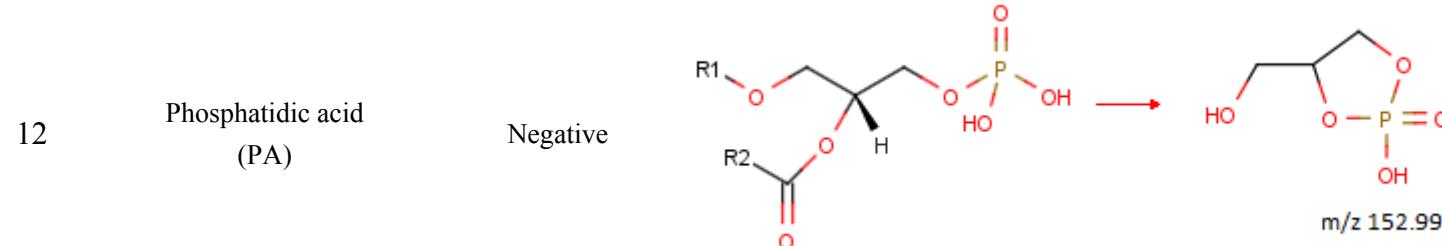
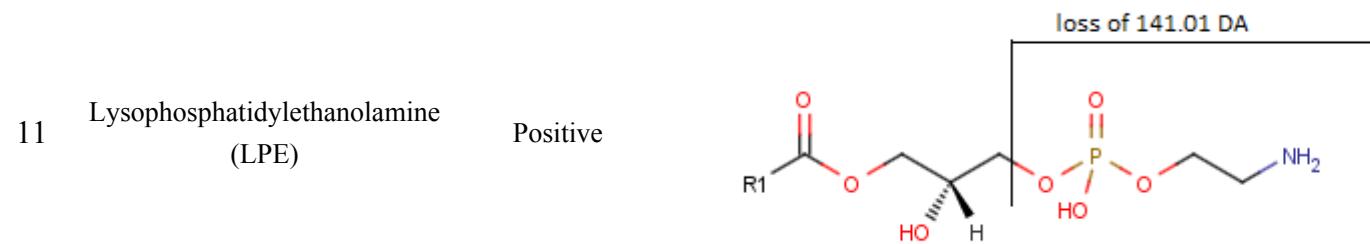
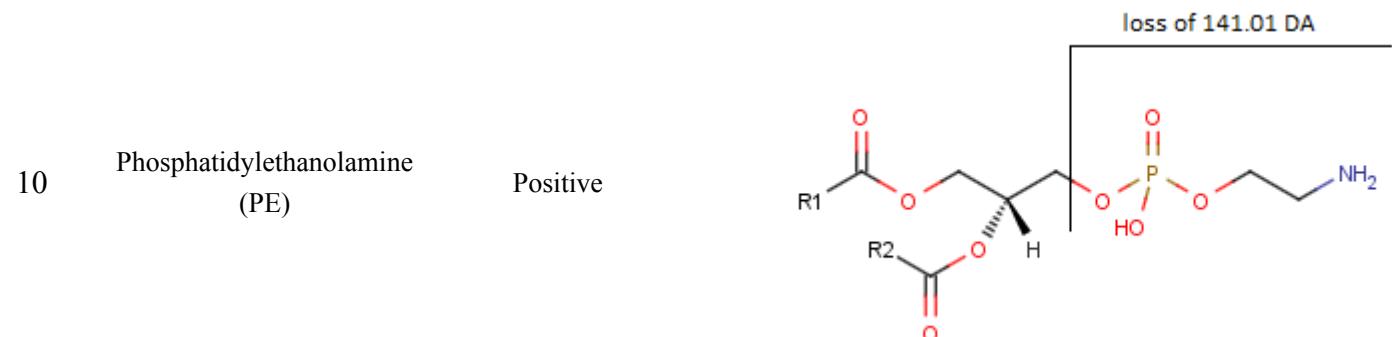
---

9

Sphingomyelin  
(SM)

Positive

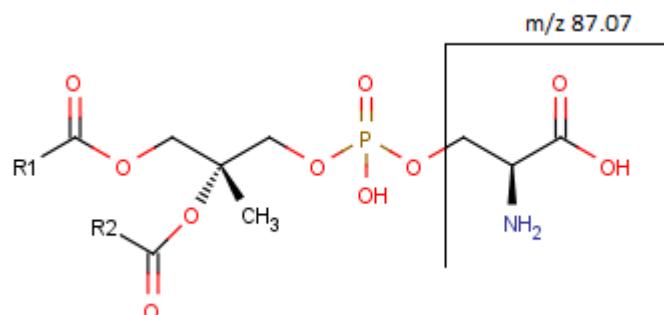




13

Phosphatidylserine  
(PS)

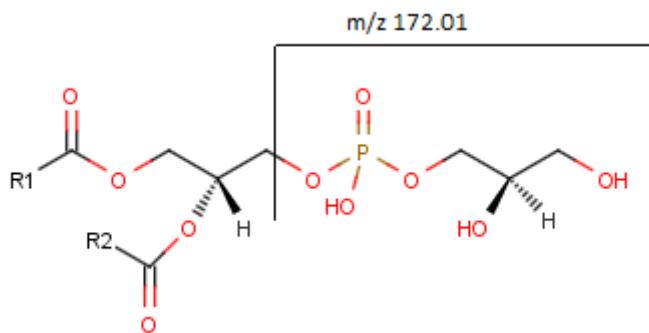
Negative



14

Phosphatidylglycerol  
(PG)

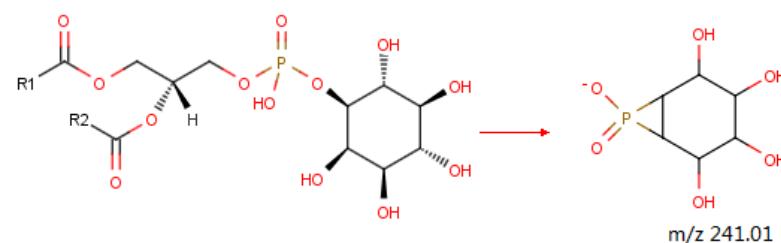
Positive



15

Phosphatidylinositol  
(PI)

Negative



## References

1. S. Guo, J. A. Duan, D. W. Qian, H. Q. Wang, Y. P. Tang, Y. F. Qian, D. W. Wu, S. L. Su and E. X. Shang, *J. Chromatogr. A.*, 2013, **1301**, 147-155.
2. S. Al-Shehri, M. Henman, B. G. Charles, D. Cowley, P. N. Shaw, H. Liley, A. Tomarchio, C. Punyadeera and J. A. Duley, *J. Chromatogr. B.*, 2013, **931**, 140-147.
3. H. Boudra, M. Doreau, P. Noziere, E. Pujos-Guillot and D. P. Morgavi, *J. Chromatogr. A.*, 2012, **1256**, 169-176.
4. W. Y. Hsu, W. D. Lin, Y. Tsai, C. T. Lin, H. C. Wang, L. B. Jeng, C. C. Lee, Y. C. Lin, C. C. Lai and F. J. Tsai, *Clin. Chim. Acta.*, 2011, **412**, 1861-1866.
5. M. J. Lopez de Alda, S. Díaz-Cruz, M. Petrovic and D. Barceló, *J. Chromatogr. A.*, 2003, **1000**, 503-526.
6. F. F. Sodré, I. C. Pescara, C. C. Montagner and W. F. Jardim, *Microchem. J.*, 2010, **96**, 92-98.
7. Y. G. Zhao, X. H. Chen, S. D. Pan, H. Zhu, H. Y. Shen and M. C. Jin, *Talanta.*, 2013, **115**, 787-797.
8. F. Y. Guan, C. E. Uboh, L. R. Soma, Y. Luo, J. Rudy and T. Tobin, *J. Chromatogr. B.*, 2005, **829**, 56-68.
9. N. Janzen, S. Sander, M. Terhardt, U. Steuerwald, M. Peter, A. M. Das and J. Sander, *Steroids.*, 2011, **76**, 1437-1442.
10. B. Á. Sánchez, F. P. Capote, J. Ruiz Jiménez and M. D. Luque de Castro, *J. Chromatogr. A.*, 2008, **1207**, 46-54.
11. J. Abdel-Khalik, E. Björklund and M. Hansen, *J. Chromatogr. B.*, 2013, **935**, 61-69.
12. A. Le, A. Ng, T. Kwan, K. Cusmano-Ozog and T. M. Cowan, *J. Chromatogr. B.*, 2014, **944**, 166-174.
13. K. Petritis, P. Chaimbault, C. Elfakir and M. Dreux, *J. Chromatogr. A.*, 2000, **896**, 253-263.
14. C. Y. Wang, H. B. Zhu, Z. F. Pi, F. R. Song, Z. Q. Liu and S. Y. Liu, *J. Chromatogr. B.*, 2013, **935**, 26-31.
15. L. A. Hammad, M. M. Saleh, M. V. Novotny and Y. Mechref, *J. Am. Soc. Mass. Spectr.*, 2009, **20**, 1224-1234.
16. Y. Huang, Y. Tian, Z. J. Zhang and C. Peng *J. Chromatogr. B.*, 2012, **905**, 37-42.
17. P. Flores, P. Hellín and J. Fenoll, *Food Chem.* 2012, **132**, 1049-1054.
18. T. Xie, Y. Liang, J. Y. A, H. P. Hao, L. S. Liu, X. Zheng, C. Dai, Y. Y. Zhou, T. Y. Guan, Y. N. Liu, L. Xie and G. J. Wang, *J. Chromatogr. B.*, 2012, **905**, 43-53.
19. J. Li, C. X. Hu, X. J. Zhao, W. D. Dai, S. L. Chen, X. Lu and G. W. Xu, *J. Chromatogr. A.*, 2013, **1320**, 103-110.
20. Y. P. Xiong, Y. Y. Zhao, S. Goruk, K. Oilund, C. J. Field, R. L. Jacobs and J. M. Curtis, *J. Chromatogr. B.*, 2012, **911**, 170-179.

21. Y. Y. Zhao, Y. P. Xiong and J. M Curtis, *J. Chromatogr. A.*, 2011, **1218**, 5470-5479.
22. C. M. Spickett, A. Reis and A. R. Pitt, *Free. Radical. Bio. Med.*, 2011, **51**, 2133-2149.
23. J. F. Brouwers, *Biochimica et Biophysica Acta.*, 2011, **1811**, 763-775.
24. O. A. Ismaiel, T. Y. Zhang, R. G. Jenkins and H. T. Karnes, *J. Chromatogr. B.*, 2010, **878**, 3303-3316.
25. E. Maciel, R. Nunes da Silva, C. Simões, T. Melo, R. Ferreira, P. Domingues and R. M. Domingues, *Chem. Phys. Lipids.*, 2013, **174**, 1-7.
26. A. K. Nilsson, O. N. Johansson, P. Fahlberg, F. Steinhart, M. B. Gustavsson, M. Ellerström and M. X. Andersson, *Phytochemistry.*, 2014, **101**, 65-75.
27. A. D. Postle, D. C. Wilton, A. N. Hunt and G. S. Attard, *Prog. Lipid. Res.*, 2007, **46**, 200-224.
28. N. Aaltonena, J. T. Laitinen and M. Lehtonen, *J. Chromatogr. B.*, 2010, **878**, 1145-1152.
29. L. W. Jia, C. Wang, S. M. Zhao, X. Lu and G. W. Xu, *J. Chromatogr. B.*, 2007, **860**, 134-140.
30. E. Maciel, R. Faria, D. Santinha, M. R. M. Domingues and P. Domingues, *J. Chromatogr. B.*, 2013, **929**, 76-83.