

Supporting Information

Enantioselective Cyanosilylation of Aldehydes Catalyzed by Multistereogenic Salen-Mn(III) Complex With Rotatable Benzylic Group As a Helping Hand

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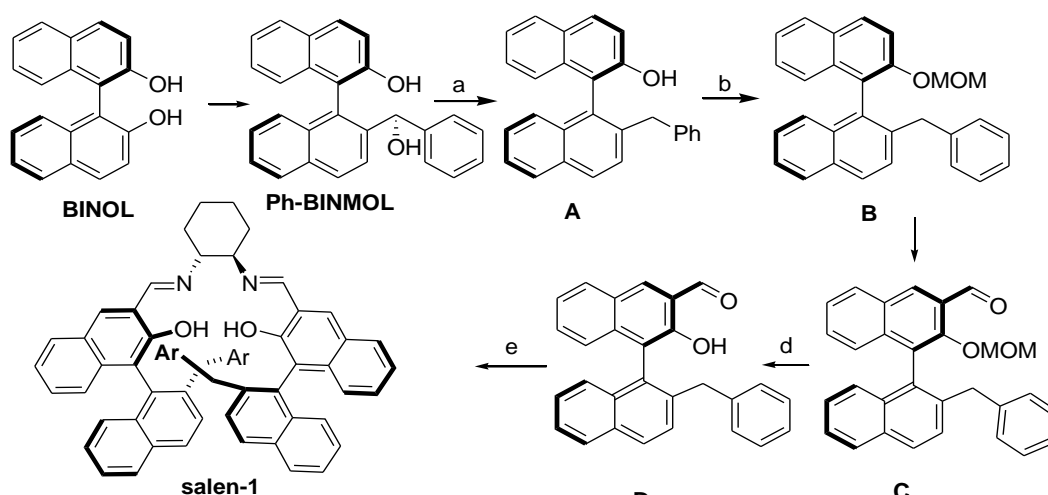
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S-1. General information

All reagents and solvents were used directly without purification. Flash column chromatography was performed over silica (200-300 mesh). Reactions were monitored by thin layer chromatography using silica gel. $^1\text{H-NMR}$, $^{13}\text{C-NMR}$, $^{19}\text{F-NMR}$ were respectively recorded at 400 or 500 MHz, 101 or 126 MHz and 376 or 471 MHz respectively, ^1H NMR chemical shifts are reported in ppm relative to tetramethylsilane (TMS) with the solvent resonance employed as the internal standard (CDCl_3 at 7.26 ppm, $(\text{CD}_3)_2\text{CO}$ at 2.05 ppm). ^{13}C NMR chemical shifts are reported in ppm from tetramethyl silane (TMS) with the solvent resonance as the internal standard (CDCl_3 at 77.20 ppm). Thin layer chromatography was performed using silica gel; F_{254} TLC plates and visualized with ultraviolet light. HPLC was carried out with a Waters 2695 Millennium system equipped with a photodiode array detector. The ESI-MS analysis of the samples was operated on an LCQ advantage mass spectrometer (ThermoFisher Company, USA), equipped with an ESI ion source in the positive ionization mode, with data acquisition using the Xcalibur software.

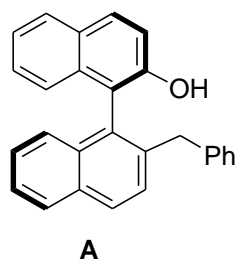
S-2. The Synthesis of Salen Ligands

2.1. Schematic synthesis of Salen 1-3



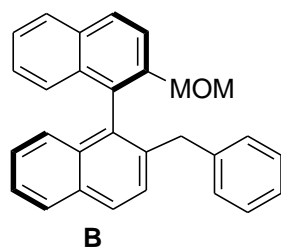
salen-1a: Ar = Ph
salen-1b: Ar = *p*(*t*-Bu)Ph
salen-1c: Ar = *o*-MePh
salen-1d: Ar = 1-naphthyl

1.2 The synthesis of BINOL-derived product salen-1a from Ph-BINMOL:



Step a: To a solution of (*S, R*)-Ar-BINMOL (9.4g, >99% *ee*, 25 mmol) and NaI (22.5 g, 150 mmol) in dry acetonitrile (100 mL) were added TMSCl (13 mL, 150 mmol) at room temperature under Argon atmosphere. After the addition, the solution was allowed to stir at room temperature overnight. Then saturated sodium thiosulfate solution was added until the mixture turned colorless. The resultant mixture was extracted with DCM, and washed with water and saturated NaCl solution. The organic layer was dried over Na₂SO₄, concentrated, and the residue was purified by column chromatography (hexanes/ethyl acetate = 5/1) to give the desired product A (9 g, 100% yield) as a white solid. ¹H NMR (CDCl₃, 400 MHz) δ 7.91 (m, 3H), 7.52 (d, *J* = 8.4 Hz, 1H), 7.45 (t, *J* = 6.8 Hz, 1H), 7.52-7.25 (m, 8H), 7.12 (d, *J* = 7.2 Hz, 2H), 6.99 (d, *J* = 8.0 Hz, 1H), 6.90 (d, *J* = 6.0 Hz, 1H), 4.62 (s, 1H), 3.77 (s, 2H). ¹³C NMR (CDCl₃, 100 MHz) δ 151.2, 140.5, 140.3, 133.7, 133.2, 132.8, 130.1, 129.3, 129.1, 129.1, 128.4, 128.3, 128.2, 127.0, 126.8, 126.1, 126.0, 125.8, 124.7, 124.7, 123.5, 117.5, 117.3, 77.4, 77.1, 76.8, 39.8 ppm. IR (neat, cm⁻¹): 3488, 3439, 3052, 3020, 2957, 2925, 1924, 1810, 1685, 1617, 1596, 1517, 1505, 1493, 1468, 1450, 1438, 1401, 1379, 1361, 1332, 1301, 1271, 1260, 1236, 1220, 1202, 1173, 1142, 1126, 1074, 1026, 975, 968, 948, 935, 910, 875, 868, 837, 816, 790, 769, 755, 717, 698, 684 cm⁻¹. Exact mass calcd for C₂₇H₂₀O [M+H]⁺, 361.15, Found 361.15.

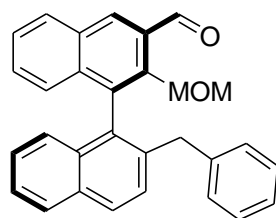
1.3 The synthesis of MOM-protected product (Step b):



To a solution of NaH (0.96 g, 40 mmol) in dry THF (150 mL) were added compound A (7.2 g, 20 mmol) which was dissolved in dry THF at 0 °C temperature under Argon

atmosphere. The mixture was stirred for 1h, then MOMCl (3.06 mL, 40 mmol) was added at 0 °C. The mixture was allowed to stir at room temperature for 4h. After quenching with water, the resultant mixture was extracted with ethyl acetate, and washed with water and saturated NaCl solution. The organic layer was dried over MgSO₄, concentrated, and the residue was purified by column chromatography (hexanes/ethyl acetate = 10/1) to give the desired product **B** (7.28 g, 90% yield) as a pale yellow solid. ¹H NMR (CDCl₃, 400 MHz) δ 7.93 (m, 4H), 7.59 (d, *J* = 9.2 Hz, H), 7.47-7.34 (m, 3H), 7.22 (s, 3H), 7.12-7.06 (m, 4H), 6.94 (s, 2H), 4.91 (m, 2H), 3.78 (s, 2H), 3.15 (s, 3H). ¹³C NMR (CDCl₃, 100 MHz) δ 152.6, 141.1, 138.0, 134.0, 133.3, 132.7, 132.3, 129.7, 129.6, 129.2, 128.1, 128.0, 127.9, 127.9, 127.8, 126.5, 126.3, 126.0, 125.7, 125.4, 125.3, 124.1, 116.4, 94.6, 77.4, 77.1, 76.8, 55.8, 39.9 ppm. IR (neat, cm⁻¹): 3419, 3060, 3025, 2951, 2920, 2900, 2848, 2839, 1910, 1754, 1621, 1592, 1504, 1492, 1471, 1450, 1435, 1401, 1355, 1335, 1300, 1269, 1246, 1200, 1166, 1148, 1088, 1069, 1032, 1013, 946, 923, 897, 861, 843, 829, 812, 794, 784, 762, 747, 740, 718, 696, 683 cm⁻¹. Exact mass calcd for C₂₉H₂₄O₂ [M+Na]⁺, 427.17, Found 427.17.

1.4 The synthesis of BINMOL-derived aldehyde (Step c):

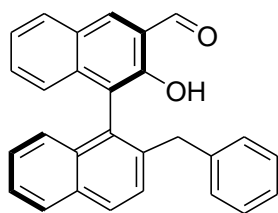


C

To a solution of compound **B** (4.04 g, 10 mmol) in the dry THF (100 ml) were added TMEDA (1.8 ml, 12 mmol) in dry THF at -78 °C under Argon atmosphere. Then *n*-BuLi (9 ml, 14 mmol, 1.5 M in hexane) was added slowly in 1h. After the addition, the solution was allowed to stir at 0 °C for 30 min. The solution was cooled to -78 °C. A solution of dry DMF (10 ml, 13 mmol) in dry THF (10 ml) were added. The solution was stirred for 30 min at -78 °C, then warmed to 0 °C. At 0 °C, it was stirred for 1.5 h. After quenching with saturated NH₄Cl solution, 1 M HCl aqueous solution was added. The resultant mixture was extracted with ethyl acetate, and washed with

saturated NaHCO₃, water and saturated NaCl solution. The organic layer was dried over Na₂SO₄, concentrated, and the residue was purified by column chromatography (hexanes/ethyl acetate = 20/1) to give the desired product **C** (3.24 g, 75% yield) as a pale yellow power. ¹H NMR (CDCl₃, 400 MHz) δ 10.60 (s, H), 8.60 (s, H), 8.04 (d, *J*=8.0 Hz, H), 7.92 (t, *J*=8.0 Hz, 3H), 7.49-7.43 (m, 3H), 7.27 (s, 3H), 7.19 (d, *J*=8.4, H), 7.08-7.03 (m, 4H), 6.89 (d, *J*=7.6 Hz, 2H), 4.62 (m, 2H), 3.86 (m, 2H), 2.93 (s, 3H). ¹³C NMR (CDCl₃, 100 MHz) δ 191.1, 153.1, 140.2, 138.9, 136.9, 133.3, 132.2, 131.9, 131.0, 130.1, 130.1, 129.2, 129.2, 129.1, 128.9, 128.6, 128.1, 128.1, 126.6, 126.2, 126.2, 126.1, 125.9, 125.5, 99.9, 77.4, 77.1, 76.8, 57.0, 40.0 ppm. IR (neat, cm⁻¹): 3057, 2920, 1689, 1618, 1586, 1506, 1496, 1451, 1382, 1354, 1330, 1259, 1182, 1156, 1104, 1073, 1042, 963, 925, 879, 832, 807, 787, 754, 712, 698, 665 cm⁻¹. Exact mass calcd for C₃₀H₂₄O₃ [M+Na]⁺, 455.16, Found 455.16.

1.5 The synthesis of deprotected BINMOL-derived aldehyde (Step d):

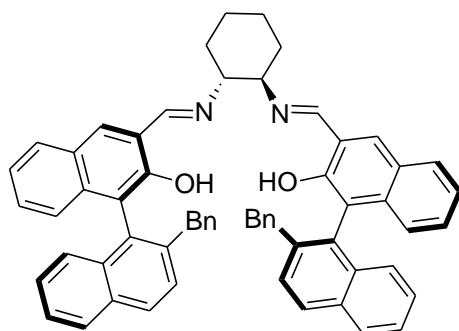


D

To a solution of compound **C** (3.03 g, 7 mmol) in THF (75 ml) were added slowly Conc. HCl (75 ml) at 0 °C. The mixture was stirred for 2 h at room temperature. Then water was added, The resultant mixture was extracted with ethyl acetate, and washed with saturated NaHCO₃, water and saturated NaCl solution. The organic layer was dried over Na₂SO₄, concentrated, and the residue was purified by column chromatography (hexanes/ethyl acetate = 10/1) to give the desired product **D** (2.58 g, 95% yield) as a yellow power. ¹H NMR (CDCl₃, 400 MHz) δ 10.04 (s, H), 10.18 (s, H), 8.42 (s, H), 7.87 (d, *J*=8.8 Hz, 3H), 7.70 (s, H), 7.46-7.43 (m, 2H), 7.36 (d, *J*=8.0 Hz, 2H), 7.36 (d, *J*=8.0 Hz, 2H), 7.06 (s, 4H), 6.94 (d, *J*=8.4 Hz, 3H), 3.78 (d, *J*=5.6 Hz, 2H). ¹³C NMR (CDCl₃, 100 MHz) δ 196.8, 153.4, 140.7, 138.5, 138.0, 137.7, 132.9, 132.5, 130.6, 129.8, 129.2, 128.5, 128.2, 128.2, 128.1, 127.5, 126.4, 125.8, 125.7,

125.4, 125.2, 124.5, 122.1, 120.8, 77.4, 77.1, 76.8, 39.9 ppm. **IR (neat, cm⁻¹):** 3428, 3150, 3059, 2838, 1792, 1655, 1629, 1577, 1506, 1452, 1443, 1412, 1384, 1292, 1254, 1225, 1212, 1179, 1142, 1115, 1073, 1054, 1025, 952, 936, 923, 895, 879, 856, 809, 796, 778, 757, 711, 697, 681 cm⁻¹. Exact mass calcd for C₂₈H₂₀O₂ [M]⁺, 388.15. Found 388.15;

1.6 The synthesis of BINMOL-derived salen:

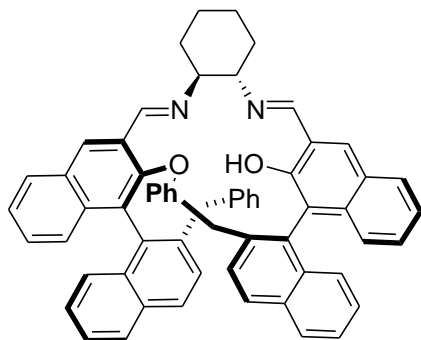


BINMOL-derived Salen

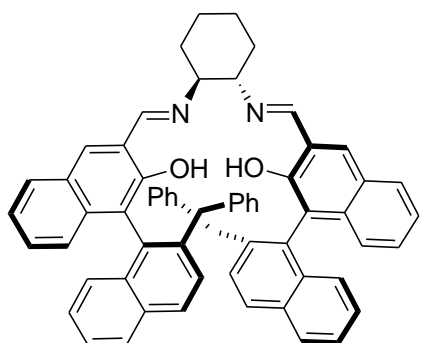
To a solution of (1R,2R)- diaminocyclohexane tartrate (0.66 g , 2.5 mmol) and K₂CO₃ (0.7 g , 5 mmol) in 50% ethanol(12 ml) were added slowly compound **D** (1.94 g , 5 mmol) in ethanol(24ml) at room temperature. After the addition, the solution was allowed to stir at room temperature for 48h. The precipitate was collected by filtration, washed with ice ethanol, and dried in vacuum oven to obtain the product **E** (2.03 g , 95% yield) as a pale yellow power. ¹H NMR(CDCl₃, 400 MHz) δ 8.45(s, H), 8.34 (s, H), 8.01 (t, *J* = 7.2 Hz, H), 7.90 (d, *J* = 8.8 Hz, 4H), 7.76 (t, *J* = 8.0 Hz, 4H), 7.49-7.46 (m, 3H), 7.40 (s, H), 7.38 (s, H), 7.30 (s, H), 7.22 (s, H), 7.11-7.08 (m, 7H), 7.00 (s, H), 6.97 (d, *J* = 8.0 Hz, 5H), 6.94 (s, H), 6.92 (s, H), 3.81 (d, *J* = 5.6 Hz, 4H), 3.32-3.30 (m, 2H), 1.99 (d, *J* = 13.6 Hz, 2H), 1.89 (d, *J* = 8.8 Hz, 2H), 1.70 (d, *J* = 11.2 Hz, 2H), 1.46 (t, *J* = 10.0 Hz, 2H). ¹³C NMR (CDCl₃, 100 MHz) δ 196.8, 165.0, 154.4, 141.0, 138.4, 138.0, 133.3, 129.3, 129.2, 128.9, 128.3, 128.2, 128.2, 128.0, 126.4, 126.1, 125.8, 125.8, 125.7, 125.6, 125.5, 125.2, 125.2, 124.7, 124.5, 123.3, 120.4, 119.3, 77.4, 77.1 , 76.8, 39.9 ppm. **IR(neat, cm⁻¹):** 3403, 3055, 2929, 2856, 1629, 1597, 1506, 1494, 1443, 1380, 1348, 1294, 1258, 1180, 1143, 1115, 1062, 1024, 941, 915, 808, 746, 712, 697 cm⁻¹. Exact mass calcd for C₆₂H₅₀N₂O₂ [M+H]⁺, 855.39, Found 855.39.

1.7 The Synthesis of Salen Ligands:

Similarly to the synthesis of salen ligands (2-3):

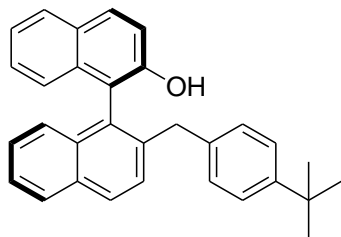


Salen-2: $^1\text{H-NMR}$ (500 MHz, CDCl_3) δ ppm: 7.53-7.54 (m, 2H), 7.97 (s, 4H), 7.81-7.84 (m, 4H), 7.55-7.58 (m, 2H), 7.43-7.45 (m, 2H), 7.34-7.37 (m, 2H), 7.29-7.31 (m, 2H), 7.09-7.20 (m, 4H), 6.99-7.02 (m, 10H), 4.22-4.25 (m, 2H), 3.85-3.92 (m, 4H), 3.40-3.41 (d, 2H), 2.33-2.34 (d, 6H), 1.95-1.96 (d, 2H), 1.81 (s, 2H), 1.35-1.41 (m, 4H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ ppm: 165.2, 154.5, 138.7, 136.1, 135.4, 135.0, 133.4, 133.2, 132.6, 132.0, 129.3, 129.0, 128.9, 128.4, 128.1, 127.5, 126.2, 126.0, 125.2, 124.9, 123.4, 120.6, 119.5, 73.2, 60.5, 39.5, 32.9, 24.2, 21.1, 14.3. **FTIR** ν_{max} (**KBr**)/ cm^{-1} : 3053, 2930, 2854, 2659, 1946, 1628, 1507, 1445, 1347, 1257, 1147, 1099, 943, 890, 804, 745, 713, 698, 496, 434.

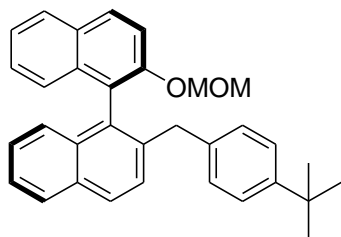


Salen-3: $^1\text{H NMR}$ (500 MHz, CDCl_3) δ ppm: 13.09- 13.11 (m, 2H), 8.49 - 8.51 (m, 2H), 7.93-7.94 (m, 4H), 7.78 - 7.81 (m, 4H), 7.52 - 7.55 (m, 2H), 7.42-7.43 (m,2H), 7.33-7.34 (m, 2H), 7.26-7.27 (m, 2H), 6.94 -7.16 (m, 16H), 3.87-3.91 (m, 4H), 3.37 (s, 2H), 2.03-2.06 (d, $J = 12.0$ Hz, 2H), 1.93 (s, 2H), 1.77 (s, 2H), 1.51 (s, 2H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ ppm : 165.1, 154.5, 141.1, 138.4, 135.3, 135.3, 135.3, 133.1, 132.1, 129.4, 128.9, 128.4, 128.1, 128.06, 127.4, 126.2, 125.9, 125.7, 125.3, 124.8, 123.3, 120.5, 119.4, 73.2, 40.0, 32.8, 24.2.

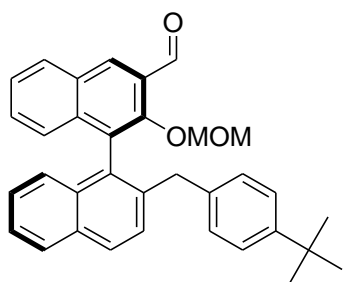
1.8 The synthesis and characterization of product Salen-1b:



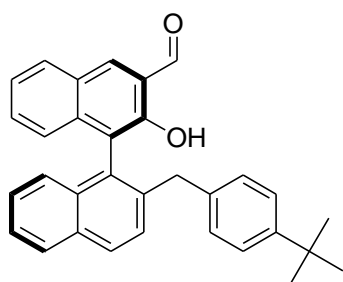
The characterization of product Salen1b-A: ¹H NMR (500 MHz, CDCl₃) δppm: 7.94-8.04 (m, 4H), 7.59-7.61 (dd, J = 2.0, 8.5 Hz, 1H), 7.53-7.55 (m, 1H), 7.45-7.47 (m, 1H), 7.36-7.42 (m, 3H), 7.23-7.29 (m, 3H), 7.08-7.10 (d, J = 8.5Hz, 1H), 6.94-6.96 (m, 2H), 5.00 (s, 1H), 3.86 (s, 2H), 1.37-1.38 (d, J = 4.0 Hz, 9H); ¹³C NMR (125 MHz, CDCl₃) δppm : 151.3, 148.8, 140.7, 137.3, 133.9, 133.5, 132.8, 130.1, 129.3, 129.2, 128.9, 128.6, 128.2, 127.0, 126.04, 126.0, 125.3, 124.9, 123.5, 117.7, 117.5, 39.4, 34.4, 31.5; FTIR ν_{\max} (KBr)/cm⁻¹: 3508, 3055, 2961, 2867, 2355, 1907, 1619, 1595, 1515, 1468, 1266, 1201, 1175, 1143, 1022, 816, 746, 561, 432.



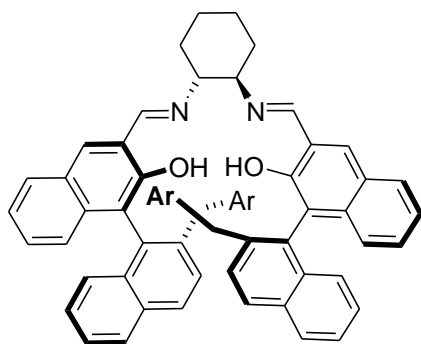
The characterization of product Salen1b-B: ¹H NMR (500 MHz, CDCl₃) δppm: 13.3 (s, 2H), 8.39 (s, 2H), 7.89-7.90 (m, 4H), 7.38-7.42 (m, 6H), 7.32-7.34 (d, J = 8.0 Hz, 2H), 7.25- 7.26 (t, J = 3.0 Hz, 4H), 7.10-7.14 (m, 10H), 7.01-7.02 (d, J = 7.0 Hz, 4H), 6.91-6.92 (d, J = 8.0 Hz, 2H), 3.72-3.75 (d, J = 15.0 Hz, 2H), 3.58-3.61 (d, J = 15.0 Hz, 2H), 3.32-3.35 (d, J = 13.5 Hz, 2H), 2.04-2.07 (d, J = 13.5 Hz, 2H), 1.47-1.51 (t, J = 10.0 Hz, 2H); ¹³C NMR (125 MHz, CDCl₃) δppm : 165.1, 154.2, 141.3, 138.6, 135.3, 133.2, 133.0, 132.5, 131.9, 129.6, 128.8, 128.1, 128.06, 128.0, 127.2, 126.2, 125.2, 124.6, 123.2, 120.3, 119.3, 73.2, 39.6, 32.5, 24.1; FTIR ν_{\max} (KBr)/cm⁻¹: 3458, 3054, 2960, 2902, 2358, 1914, 1622, 1593, 1507, 1362, 1244, 1197, 1067, 1034, 1015, 922, 812, 749, 561.



The characterization of product Salen1b-C: ¹H NMR (400 MHz, CDCl₃) δppm: 10.64 (s, 1H), 8.62 (s, 1H), 8.04 (s, 1H), 7.92-8.02 (m, 2H), 6.98-7.31 (m, 9H), 6.77 (d, J = 8.4 Hz, 2H), 4.57-4.71 (m, 2H), 3.80-3.96 (m, 2H), 2.96 (s, 3H), 1.24 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δppm: 191.1, 153.1, 148.5, 139.2, 127.1, 137.0, 133.4, 132.3, 131.8, 131.1, 130.0, 129.1, 129.0, 128.7, 128.6, 128.4, 128.1, 126.3, 126.2, 126.0, 125.4, 124.9, 99.9, 39.7, 34.3, 31.4. FTIR ν_{max} (KBr)/cm⁻¹: 3456, 3054, 2960, 2903, 2868, 2359, 1911, 1690, 1619, 1587, 1508, 1450, 1268, 1157, 1075, 966, 925, 815, 756, 563.



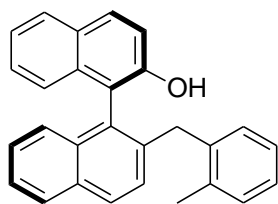
The characterization of product Salen1b-D: ¹H NMR (500 MHz, CDCl₃) δppm: 10.43-10.45 (m, 1 H), 10.91 (s, 1 H), 8.31 (s, 1 H), 7.96-7.99 (s, 3 H), 6.87-7.60 (m, 11 H), 3.84-3.85 (m, 2 H), 1.29-1.31 (m, 9 H); ¹³C NMR (125 MHz, CDCl₃) δppm : 196.7, 153.4, 148.5, 138.8, 137.95, 137.88, 137.6, 133.0, 132.6, 130.7, 130.4, 129.8, 128.8, 128.5, 128.3, 127.5, 12.4, 125.8, 125.3, 125.0, 122.1, 120.9, 39.6, 34.3, 31.4. FTIR ν_{max} (KBr)/cm⁻¹: 3202, 3054, 2960, 2859, 1918, 1653, 1629, 1504, 1411, 1386, 1339, 1295, 1116, 1055, 936, 838, 815, 777, 754, 604, 434.



1b: Ar = *p*-*t*-BuPh

The characterization of product Salen-1b: $^1\text{H NMR}$ (500 MHz, CDCl_3) δ ppm: 13.4 (s, 2H), 8.35 (s, 2H), 7.88-7.90 (dd, $J = 3.0$ Hz, 8.5 Hz, 4H), 7.47-7.49 (d, $J = 8.5$ Hz, 2H), 7.38- 7.42 (m, 2H), 7.29 (s, 2H), 7.22-7.23 (d, $J = 3.5$ Hz, 4H), 7.10-7.12 (dd, $J = 2.0$ Hz, 8.5Hz, 6H), 6.90-7.04 (m, 8H), 3.78-3.82 (d, $J = 15.5$ Hz, 2H), 3.64-3.67 (d, $J = 15.5$ Hz, 2H), 3.31-3.33 (dd, $J = 4.0$ Hz, 6.0 Hz, 2H), 2.07-2.10 (d, $J = 13.5$ Hz, 2H), 1.90-1.92 (d, $J = 8.5$ Hz, 2H), 1.77-1.79 (m, 2H), 1.47-1.51 (t, $J = 5.0$ Hz, 2H), 1.27 (s, 18H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ ppm :165.2, 154.0, 148.4, 138.8, 138.0, 135.3, 133.2, 132.5, 131.9, 129.2, 128.7, 128.1, 127.2, 126.1, 126.09, 125.1, 125.0, 124.6, 123.0, 120.2, 119.4, 39.4, 34.3, 32.3, 31.5, 21.1. **FTIR** ν_{max} (KBr)/ cm^{-1} : 3053, 2959, 2861, 2361, 1910, 1630, 1507, 1442, 1344, 1258, 1145, 1117, 1022, 943, 812, 747, 566, 433.

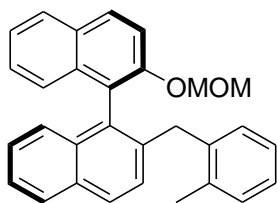
1.9 The synthesis and characterization of product Salen 1c:



The characterization of product Salen1c-A:

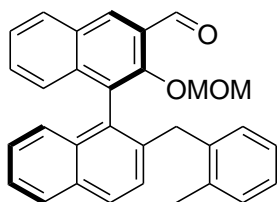
$^1\text{H NMR}$ (500 MHz, CDCl_3) δ ppm: 7.9-8.03(m, 4 H), 7.54-7.57(td, $J=1, 6.5$ Hz, 1 H), 7.33-7.48(m, 6 H), 7.16-7.21(m, 4 H), 7.08-7.09(d, $J=7\text{Hz}$, 1 H), 4.96(s, 1 H), 3.86-3.96(q, $J=16.5\text{Hz}$, 2 H), 2.09(s, 3H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ ppm: 151.2, 139.9, 138.3, 136.7, 13.6, 133.4, 132.7, 130.3, 130.3, 130.1, 129.5, 129.4, 129.2, 128.3, 128.2, 127.6, 127.1, 126.8, 126.3, 126.1, 126.0, 125.8, 124.4, 123.6, 117.8, 117.5, 37.1, 19.7; **FTIR** ν_{max} (KBr)/ cm^{-1} : 3454, 2084, 1632, 1508, 1457, 1340, 1178,

1143, 494.



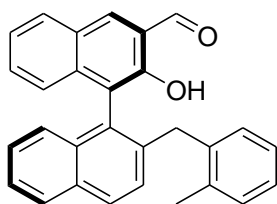
The characterization of product Salen1c-B:

¹H NMR (500 MHz, CDCl₃) δppm: 8.05-8.16(m, 4 H), 7.80-7.83(m, 1 H), 7.70-7.74(m, 1 H), 7.20-7.59(m, 7 H), 7.01-7.10(dd, J=7 Hz, 2 H), 6.91(s, 1H), 5.16-5.18(m, 1H), 5.06-5.08(m, 1H), 4.01(s, 2H), 3.33-3.45(m, 3H), 2.37(s, 3H); **¹³C NMR** (125 MHz, CDCl₃) δppm: 152.8, 141.1, 138.4, 137.6, 134.2, 133.6, 132.9, 132.6, 130.3, 130.0, 129.8, 128.4, 128.0, 126.7, 126.6, 126.5, 126.4, 125.7, 125.4, 124.3, 123.7, 116.7, 94.9, 55.9, 40.1, 21.5; **FTIR** ν_{\max} (KBr)/cm⁻¹: 3529, 3485, 3044, 2958, 2924, 1905, 1594, 1507, 1304, 1196, 1147, 1067, 1034, 924, 808, 745, 440.



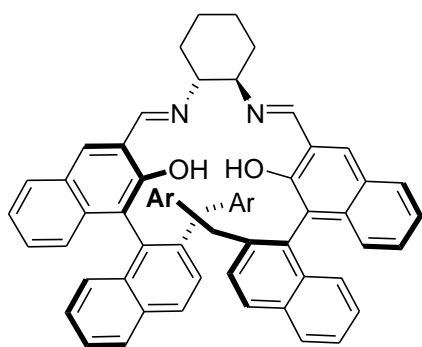
The characterization of product Salen1c-C:

¹H NMR (500 MHz, CDCl₃) δppm: 10.65 (s, 1H), 8.4(s, 1H), 8.07-8.09(d, J =8 Hz, 1H), 7.91-7.95 (t, J= 9Hz, 2H), 7.46-7.51(dd, J=7.5, 14.5Hz, 2 H), 7.24-7.37(m, 4H), 7.18-7.19(d, J=8.5Hz, 1H), 7.00-7.08(m, 4H), 4.76-4.77(d, J=6Hz, 1H), 4.61-4.63(d, J=Hz, 1H), 4.02-4.06(d, J=16.5Hz, 1H), 3.76-3.79(d, J=17Hz, 1H), 2.97(s, 3H), 2.04(s, 3H); **¹³C NMR** (125 MHz, CDCl₃) δppm: 190.1, 152.3, 138.1, 136.7, 136.7, 133.4, 133.2, 131.9, 131.2, 130.4, 130.3, 130.1, 129.3, 129.2, 129.1, 128.6, 128.1, 127.2, 126.5, 126.4, 126.2, 126.1, 125.9, 125.4, 100.0, 57.0, 37.4, 19.7; **FTIR** ν_{\max} (KBr)/cm⁻¹: 3458, 2359, 1687, 1654, 1581, 1508, 1355, 1260, 1180, 1155, 1104, 964, 810, 755, 554, 500.



The characterization of product Salen1c-D:

^1H NMR (500 MHz, CDCl_3) δ ppm: 10.66(s, 1H), 8.66(s, 1H), 8.0-8.08(d, $J=8\text{Hz}$, 1H), 7.94-7.98(dd, $J=9, 11.5\text{Hz}$, 2H), 7.45-7.55(m, 3H), 7.25-7.33(m, 3H), 7.08-7.12(m, 4H), 6.93-6.95(d, $J=8\text{Hz}$, 2H), 4.74-4.75(d, $J=5.5\text{Hz}$, 1H), 4.61-4.62(d, $J=5.5\text{Hz}$, 1H), 3.98-4.00(d, $J=15.5\text{Hz}$, 1H), 3.84-3.88(d, $J=5.5\text{Hz}$, 1H), 2.98(s, 3H); **^{13}C NMR** (125 MHz, CDCl_3) δ ppm: 191.0, 153.2, 138.9, 136.9, 133.4, 132.3, 132.2, 131.1, 130.2, 129.2, 129.1, 129.0, 128.6, 128.2, 126.6, 126.3, 126.0, 125.9, 125.5, 99.9, 57.0, 40.1; **FTIR** ν_{max} (KBr)/ cm^{-1} : 3478, 3054, 2855, 2354, 1658, 1631, 1504, 1338, 1296, 1118, 933, 895, 815, 749, 507, 434.

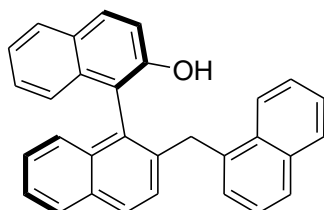


salen1c: Ar = *o*-MePh

The characterization of product Salen1c:

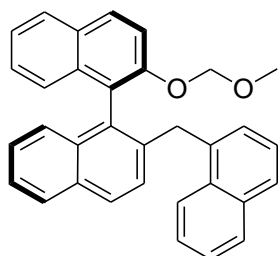
^1H NMR (400 MHz, CDCl_3) δ ppm: 13.03(s, 2H), 8.40(s, 2H), 7.80-7.86(m, 4H), 7.68-7.72(t, $J=8.8\text{Hz}$, 4H), 7.31-7.35(t, $J=8.8\text{Hz}$, 2H), 7.17-7.26(m, 8H), 6.98-7.05(m, 12H), 6.87-6.91(t, $J=8\text{Hz}$, 2H), 3.81-3.84(d, $J=16\text{Hz}$, 2H), 3.68-3.72(d, $J=16\text{Hz}$, 2H), 3.26-3.28(m, 2H), 1.95(s, 6H), 1.83-1.85(d, $J=8.8\text{Hz}$, 2H), 1.66-1.68(d, $J=6\text{Hz}$, 2H), 1.36-1.48(m, 2H); **^{13}C NMR** (100 MHz, CDCl_3) δ ppm: 165.0, 154.3, 138.7, 137.8, 136.7, 135.0, 133.2, 133.0, 132.4, 132.0, 130.5, 129.8, 128.8, 128.2, 128.0, 127.9, 127.4, 127.1, 126.1, 126.0, 125.7, 125.6, 125.0, 124.7, 123.2, 120.5, 119.3, 73.0, 37.1, 32.7, 24.1, 19.6; **FTIR** ν_{max} (KBr)/ cm^{-1} : 3054, 2934, 2858, 2836, 1633, 1508, 1462, 1301, 1259, 1215, 1143, 947, 882, 859, 769, 747, 619, 434.

The synthesis and characterization of product Salen-1d:



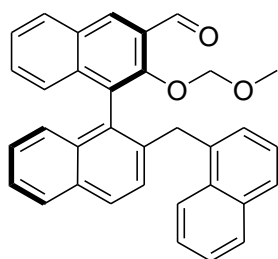
The characterization of product Salen1d-A:

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ ppm:7.81-7.86(dd, $J=8\text{Hz}$, 2H), 7.76-7.78(d, $J=8.4\text{Hz}$, 1H), 7.71-7.73(d, $J=8\text{Hz}$, 1H), 7.72-7.9(m, 3H), 7.34-7.38(t, $J=7.2\text{Hz}$, 1H), 7.12-7.32(m,11H), 5.19(s, 1H), 3.61-3.64(t, $J=6.4\text{Hz}$, 2H); $^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ ppm:151.4, 139.9, 136.1, 133.9, 133.8, 133.3, 132.7, 132.2, 130.1, 129.4, 129.2, 129.1, 128.6, 128.4, 128.2, 128.0, 127.5, 127.2, 127.0, 126.8, 125.9, 125.8, 125.5, 125.5, 124.8, 124.4, 123.6, 117.8, 117.4, 68.0, 36.7, 25.6; **FTIR** ν_{max} (**KBr**)/ cm^{-1} : 3486, 3055, 2359, 1620, 1595, 1507, 1379, 1262, 1201, 1174, 1143, 817, 749, 419.



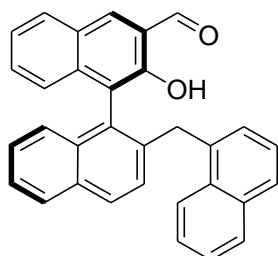
The characterization of product Salen1d-B:

$^1\text{H NMR}$ (500 MHz, CDCl_3) δ ppm:7.82-7.84(d, $J=9.2\text{Hz}$, 1H), 7.76-7.78(d, $J=8\text{Hz}$, 1H), 7.62-7.73 (m, 4H), 7.55-7.57 (d, $J=8.4\text{Hz}$, 1H), 7.48-7.50(d, $J=9.2\text{Hz}$, 1H), 7.08-7.26(m, 11H), 4.94-4.96(d, $J=6.8\text{Hz}$, 1H), 4.86-4.87(d, $J=6.8\text{Hz}$, 1H), 4.16-4.20(d, $J=16.4$, 1H), 4.03-4.07(d, $J=16.8\text{Hz}$, 1H), 3.04(s, 3H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ ppm: 152.7, 137.9, 136.6, 133.9, 133.4, 132.6, 132.4, 132.0, 129.8, 128.5, 128.2, 128.0, 127.9, 127.8, 127.3, 127.0, 126.7, 126.3, 126.0, 125.6, 125.5, 125.4, 125.2, 124.8, 124.3, 122.9, 116.7, 95.2, 55.9, 37.2; **FTIR** ν_{max} (**KBr**)/ cm^{-1} : 3462, 1636, 1507, 1472, 1397, 1259, 1240, 1196, 1066, 1032, 749.



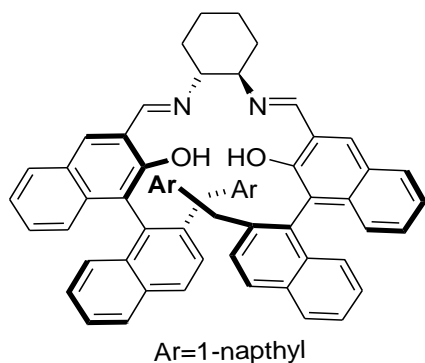
The characterization of product Salen1d-C:

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ ppm: 10.56(s,1H), 8.52(s, 1H), 7.97-7.99(d, J=8Hz, 1H), 7.82-7.84(d, J=8Hz, 1H), 7.68-7.77(m,3H), 7.59-7.61(d, J=8Hz, 1H), 7.37-7.44(m, 2H), 7.28-7.34(dd, J=6.8, 15.2Hz, 2H), 7.13-7.28(m, 7H), 4.73-4.74(d, J=6Hz, 1H), 4.58-4.59(d, J=6Hz, 1H), 4.37-4.42(d, J=16.8Hz, 1H), 4.14-4.18(d, J=16.4Hz, 1H), 2.9(s, 3H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ ppm: 190.9, 153.2, 138.7, 136.6, 135.9, 133.8, 133.3, 133.2, 133.0, 132.2, 132.0, 130.8, 129.2, 129.0, 128.5, 128.5, 128.1, 128.0, 127.3, 127.0, 126.5, 126.1, 125.5, 125.3, 124.4, 100.1, 57.0, 37.1; **FTIR** ν_{max} (KBr)/ cm^{-1} : 3453, 2280, 1685, 1636, 1356, 1157, 1074, 962, 749.



The characterization of product Salen1d-D:

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ ppm: 10.50(s, 1H), 10.04(s, 1H), 8.14(s, 1H), 7.85-7.88(dd, J=3.6, 6.8Hz, 1H), 7.79-7.81(d, J=8Hz, 1H), 7.69-7.74(m, 2H), 7.58-7.60(d, J=8Hz, 1H), 7.28-7.37(m, 4H), 7.12-7.24(m, 7H), 4.26-4.30(d, J=16.4Hz, 1H), 4.11-4.15(d, J=16.4Hz, 1H); $^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ ppm: 196.8, 153.3, 138.3, 138.0, 137.6, 136.2, 133.6, 132.9, 132.5, 132.2, 130.5, 130.4, 129.9, 128.4, 128.2, 128.0, 127.6, 126.9, 126.3, 125.7, 125.4, 125.2, 124.6, 124.5, 122.1, 37.0. **FTIR** ν_{max} (KBr)/ cm^{-1} : 3184, 3049, 2913, 2842, 1943, 1653, 1626, 1504, 1384, 1292, 1177, 895, 827, 793, 774, 752, 507, 431.



The characterization of product Salen 1d:

¹H NMR (400 MHz, CDCl₃) δppm: 13.19(s, 2H), 8.42(s, 2H), 7.79-7.87(m, 4H), 7.62-7.74(m, 10H), 7.47-7.55(m, 2H), 7.18-7.36 (m, 16H), 7.09-7.11 (m, 4H), 6.88-6.94(m, 2 H), 4.28-4.35(m, 2 H), 4.12-4.17(m, 2 H), 3.12-3.32(m, 2 H), 1.40-2.00(m, 8H); **¹³C NMR** (101 MHz, CDCl₃) δ ppm: 162.6, 151.9, 135.7, 134.1, 132.6, 131.3, 130.9, 130.6, 129.9, 129.3, 128.1, 127.4, 126.5, 126.4, 126.0, 125.9, 125.8, 125.7, 125.6, 125.1, 125.0, 124.9, 124.7, 124.3, 123.7, 123.4, 123.2, 123.1, 123.0, 122.9, 122.8, 122.7, 122.5, 122.4, 122.3, 120.9, 118.1, 116.9, 70.7, 34.5, 30.3, 21.7; **FTIR** ν_{\max} (KBr)/cm⁻¹: 3179, 3050, 3008, 2929, 2857, 2302, 1919, 1629, 1507, 1442, 1383, 1342, 1295, 1115, 944, 858, 776, 749, 432.

S-3. The synthesis of (Salen)Mn(III)-X catalysts

3.1 Typical procedure of synthesis of (Salen)Mn(III)-Cl:

Salen(1.57g, 1.83 mmol, 1eq) and manganese(II) acetate tetrahydrate (1.34, 5.47mmol, 3eq) were added to 20 mL of ethanol, and the mixture was refluxed for 3 h in air. Upon cooling, 20 mL of an aqueous saturated sodium chloride solution was added, and after stirring for 2h, an additional 10 mL of water was added. The mixture was then extracted with 20 mL of dichloromethane. The combined organic portions were then washed with 5 mL of water and dried over MgSO₄. The solvent was removed by rotary evaporation, the dark brown solid is dissolved in 20 mL of methylene chloride in a 1-L, round-bottomed flask. Heptane (100 mL) is added, and the methylene chloride is removed by reduced pressure rotary evaporation. After complete removal of the methylene chloride, the brown slurry is stirred for 1 hr at ≤5°C in an ice bath. The brown solid is collected by vacuum filtration and allowed to air dry. Heating of the solid at 50-60 °C under high vacuum removes any residual solvent to yield the brown product (1.60 g, 93% yield). Element Analysis: Calcd for C₆₂H₄₈MnN₂O₂: C 78.88, H: 5.13, N: 2.99 Found: C 78.29, H: 4.63, N: 2.80

C1a: FTIR ν_{\max} (KBr)/cm⁻¹: 3644, 3054, 2925, 2857, 1948, 1610, 1492, 1309, 1189, 952, 800, 758, 689, 537. **ESI-MS (m/z):** [M]⁺ calcd for C₆₂H₄₈MnN₂O₂ 907.3,

found 907.3.

C1b: FTIR ν_{\max} (KBr)/ cm^{-1} : 3645, 3052, 2955, 2902, 2862, 1909, 1609, 1508, 1310, 952, 860, 757, 689, 567; **ESI-MS (m/z):** $[\text{M}]^+$ calcd for $\text{C}_{70}\text{H}_{64}\text{MnN}_2\text{O}_2$ 1019.4, found 1019.4.

C1c: FTIR ν_{\max} (KBr)/ cm^{-1} : 3645, 3051, 2931, 2858, 1914, 1609, 1449, 1320, 1222, 1189, 952, 760, 688, 569; **ESI-MS (m/z):** $[\text{M}]^+$ calcd for $\text{C}_{64}\text{H}_{52}\text{MnN}_2\text{O}_2$ 935.3, found 935.3.

C1d: FTIR ν_{\max} (KBr)/ cm^{-1} : 3436, 3048, 2934, 2858, 1940, 1609, 1555, 1507, 1447, 1347, 1371, 1026, 951, 758, 689, 570; **ESI-MS (m/z):** $[\text{M}]^+$ calcd for $\text{C}_{70}\text{H}_{52}\text{MnN}_2\text{O}_2$ 1007.3, found 1007.3.

C2: FTIR ν_{\max} (KBr)/ cm^{-1} : 3645, 3045, 3025, 2930, 2857, 1947, 1609, 1493, 1449, 1347, 951, 802, 745, 715, 568. **ESI-MS (m/z):** $[\text{M}]^+$ calcd for $\text{C}_{62}\text{H}_{48}\text{MnN}_2\text{O}_2$ 907.3, found 907.3.

C3: FTIR ν_{\max} (KBr)/ cm^{-1} : 3645, 3053, 2931, 2858, 1609, 1449, 1309, 1221, 1189, 951, 759, 690, 658, 537. **ESI-MS (m/z):** $[\text{M}]^+$ calcd for $\text{C}_{62}\text{H}_{48}\text{MnN}_2\text{O}_2$ 907.3, found 907.3.

3.2 Typical procedure of synthesis of (Salen)Mn-OAc:

An appropriate amount of salen ligands (**Salen-1a**, 0.253 mmol, 0.216 g) was separately dissolved in absolute ethanol in nitrogen atmosphere under vigorous stirring. Then was added $\text{Mn}(\text{OAc})_2 \cdot 4\text{H}_2\text{O}$ (0.253 mmol, 0.062 g), and the resulting mixture was refluxed for 7 h in nitrogen atmosphere until the complete reaction of ligand compound (checked by TLC). Finally, the precipitate complexes were recovered by filtration, and washed several times with absolute ethanol and dichloromethane, dried in vacuum at 65 °C for 5 h, and again purified by re-crystallization in acetone to get the brown product in a 88% yield. **FTIR ν_{\max} (KBr)/ cm^{-1} :** 3643, 3203, 3054, 2933, 2859, 1946, 1819, 1610, 1554, 1422, 1323, 1027, 952, 801, 757, 689, 567, 535. **ESI-MS (m/z):** $[\text{M}]^+$ calcd for $\text{C}_{62}\text{H}_{48}\text{MnN}_2\text{O}_2$ 907.3, found 907.3.

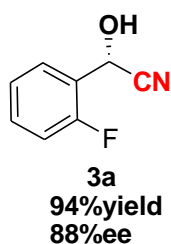
3.3 Typical procedure of synthesis of (Salen)Mn-OTf:

Manganese(II) acetate tetrahydrate (1.55 g, 6.32 mmol) was added to the solution of Salen 1a (2.7g, 3.16 mmol) in N,N-dimethylformamide (20 mL). The resulting solution was heated at 100 °C for 3 h. After cooling, the solvent was removed by evaporation under reduced pressure, and then dried in vacuo at 80 °C for 12 h. The residue dissolved in CH₂Cl₂ (200 mL) was washed with aqueous trifluoromethanesulfonic acid solution (0.5 M, 100 mL × 3). The organic layer was dried over MgSO₄, and the solvent was removed by evaporation under reduced pressure. The residue was then dissolved in CH₂Cl₂ (~10 mL), and the resulting solution was passed through a membrane filter (Millex®-FG, pore size 0.20 μm, diameter 25 mm, S22 Millipore). The addition of pentane (~50 mL) afforded Mn (III) (2.77, 2.62 mmol) as a brown solid in a 83% yield. **FTIR** ν_{\max} (KBr)/cm⁻¹: 3183, 3055, 2933, 2858, 1651, 1610, 1505, 1451, 1347, 1245, 1152, 1029, 804, 747, 638, 571, 510. **ESI-MS** (m/z): [M]⁺calcd for C₆₂H₄₈MnN₂O₂ 907.3, found 907.3.

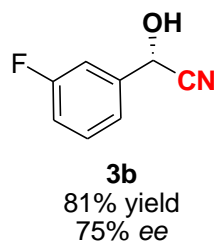
S-4. General procedure for the asymmetric cyanosilylation

General procedure of silylcyanation of the aldehydes catalyzed by Mn(salen) and POPh3: The aldehyde (0.5 mmol) was added to a stirred DCE solution of the catalyst [3 mol% Mn(salen), 20 mol% Ph₃PO] and the mixture stirred for 30 min at -10 °C. TMSCN (0.75 mmol) was then added with a syringe and the mixture was stirred at the same temperature for 24 h. The solvent was then evaporated, 2N HCl (10 mL) was added, and the mixture was stirred vigorously at room temp. for 2 h to hydrolyze the trimethylsilyl ether. After addition of ethylacetate (10 ml), the mixture was stirred for 30 min. The organic layer was separated and washed with H₂O. The aqueous layer was extracted with ethyl acetate (2 ×20 mL), and the combined organiclayers were washed with brine and dried with Na₂SO₄.The crude product was further purified by flash chromatography and ee % was determined by chiral high pressureliquid

chromatography (HPLC) column .

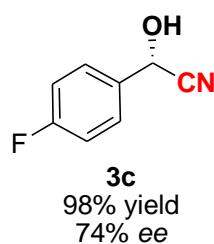


2-Hydroxy-2-(2- Fluorophenyl) acetonitrile (**3a**) (yellow oil) was obtained according to the general procedure (94% yield). Enantiomeric excess was determined by HPLC with a Chiralcel OD-H column(n-hexane-isopropanol 95:5 V/V, 0.8 mL/min, 210 nm); minor enantiomer $t_r = 18.693$ min, major enantiomer $t_r = 17.504$ min; 88% *ee*; $[\alpha]_D^{18} -28.44^\circ$ (c 1.0, CHCl_3) ; [lit.^[4] $[\alpha]_D^{20} +21.9$ (c 3.60 , CHCl_3), 84% *ee* (R)]. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ ppm: 7.60-7.4 (td, $J=1.2, 6.4\text{Hz}$, 1H) , 7.41-7.46 (m, 1H), 7.22-7.26 (t, $J= 7.8\text{Hz}$, 1H), 7.11-7.16(t, 9.2Hz, 1H), 5.77(s, 1H), 3.48(s, 1H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ ppm: 161.2, 158.7, 131.8-131.9 (s, $J_{\text{CF}}=8.2\text{Hz}$, 1C), 128.4, 124.9-125.0 (s, $J_{\text{CF}}=3.7\text{Hz}$, 1C), 122.8-122. 9 (s, $J_{\text{CF}}=4.2\text{Hz}$, 1C), 118, 115.9-116.1 (s, $J_{\text{CF}}=20.4\text{Hz}$, 1C), 57.9-58.0 (s, $J_{\text{CF}}=4.9\text{Hz}$, 1C); $^{19}\text{F NMR}$ (470 MHz, CDCl_3): -118.2; **IR (KBr)**: 3405, 2959, 2852, 2362, 1617, 1492, 1459, 1231, 1030, 759.

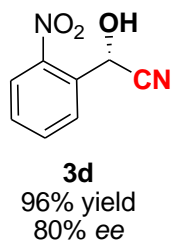


2-Hydroxy-2-(3- Fluorophenyl) acetonitrile (**3b**) (yellow oil) was obtained according to the general procedure (81% yield). Enantiomeric excess was determined by HPLC with a Chiralcel AD-H column(n-hexane-isopropanol 95:5 V/V, 0.8 mL/min, 210 nm); minor enantiomer $t_r = 21.36$ min, major enantiomer $t_r = 22.59$ min; 75% *ee* ; $[\alpha]_D^{18} -20.05^\circ$ (c 1.0, CHCl_3) ; $^1\text{H NMR}$ (400 MHz, CDCl_3) δ ppm: 7.11 (dt, $J = 8.4, 8. 3, 2.2$ Hz, 1H), 7.33-7.18 (m, 1H), 7.40 (dd, $J = 13.8, 7.9$ Hz, 1H), 5.53 (s, 1H), 3.93 (d, $J = 2.8$ Hz, 1H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ ppm: 164.21 (s, 1C), 161.74 (s, 1C), 137.46 (d, $J_{\text{CF}} = 7.31$ Hz, 1C), 130.87 (d, $J_{\text{CF}} = 8.30$ Hz, 1C), 122.14 (d, $J_{\text{CF}} = 3.03$ Hz,1C), 118.55 (s, 1C), 116.91 (s, 1C), 116.70 (s, 1C), 113.89 (s, 1C), 113.65 (s, 1C), 62.77 (s, 1C); $^{19}\text{F-NMR}$ (470 MHz, CDCl_3) δ ppm: -110.9; **IR(KBr)**: 3445, 2968,

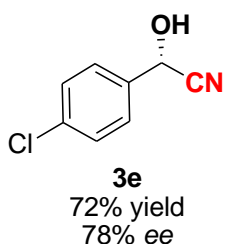
2923, 2851, 2061, 1619, 1453, 1265, 1137, 1051, 868, 757



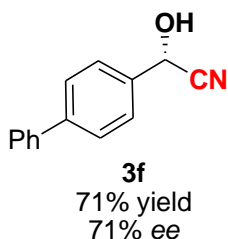
2-Hydroxy-2-(3-Fluorophenyl)acetonitrile (**3c**) (yellow oil) was obtained according to the general procedure (98% yield). Enantiomeric excess was determined by HPLC with a Chiralcel AD-H column(n-hexane-isopropanol 95:5 V/V, 0.8 mL/min, 210 nm); minor enantiomer $t_r = 20.464$ min, major enantiomer $t_r = 18.488$ min; 74% *ee*; $[\alpha]_D^{18} -26.4^\circ$ (c 1.0, CHCl_3); [lit. $^{[4]}[\alpha]_{20}^D +36.4$ (c 6.4, CHCl_3) for R-enantiomer in 94%*ee*]; $^1\text{H NMR}$ (400 MHz, CDCl_3) δ ppm: 7.50 (dd, $J = 8.4, 5.2$ Hz, 1H), 7.16-7.06 (m, 1H), 5.51 (s, 1H), 3.68 (s, 1H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ ppm: 164.7-164.2(d, $J_{\text{CF}}=48.1\text{Hz}$, 1C), 131.2-131.1(d, $J_{\text{CF}}=3.2\text{Hz}$, 1C), 128.7-128.6 (d, $J_{\text{CF}}=8.4\text{Hz}$, 1C), 118.8, 116.3-116.1(d, $J_{\text{CF}}=21.0\text{Hz}$, 1C), 62.9; $^{19}\text{F-NMR}$ (470 MHz, CDCl_3) δ ppm: -110.9; **IR (KBr)**: 3430, 2959, 2924, 2853, 2361, 1605, 1510, 1234, 1039, 836, 749.



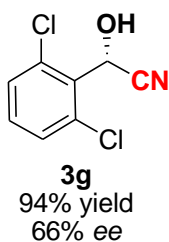
2-Hydroxy-2-(4-Nitrophenyl)acetonitrile (**3d**) was obtained according to the general procedure (96% yield). Enantiomeric excess was determined by HPLC with a Chiralcel AD-H column(n-hexane-isopropanol 95:5 V/V, 0.8 mL/min, 210 nm); minor enantiomer $t_r = 19.831$ min, major enantiomer $t_r = 18.258$ min; 80% *ee*; $[\alpha]_D^{18} -70.4^\circ$ (c 1.0, CHCl_3); $^1\text{H NMR}$ (400 MHz, CDCl_3) δ ppm: 8.16-8.18(dd, $J=1, 8\text{Hz}$, 1H), 7.97-7.99(d, $J=7.5\text{Hz}$, 1H), 7.77-7.80(t, $J=8\text{Hz}$, 1H), 7.61-7.65(t, $J=8\text{Hz}$, 1H), 6.20(s, 1H), 4.15(s, 1H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ ppm: 146.9, 134.8, 130.9, 130.8, 129.3, 125.8, 117.6, 60.4; **IR (KBr)**: 3492, 3399, 3111, 2966, 2934, 2854, 2249, 1524, 1346, 1186, 1040, 786, 741, 704.



2-Hydroxy-2-(4-Chlorophenyl)acetonitrile (**3e**) (yellow oil) was obtained according to the general procedure (72% yield). Enantiomeric excess was determined by HPLC with a Chiralcel AD-H column(n-hexane-isopropanol 95:5 V/V, 0.8 mL/min, 210 nm); minor enantiomer $t_r = 13.941$ min, major enantiomer $t_r = 12.299$ min; 78% *ee*; $[\alpha]_D^{18} -31.9^\circ$ (c 1.0, CHCl_3); [lit.^[1] $[\alpha]_D +27.2$ (c 1.50, CHCl_3) for R enantiomer in 66% *ee*]; $^1\text{H NMR}$ (500 MHz, CDCl_3) δ ppm: 7.39-7.46 (m , 4 H), 5.29 (s, 1 H), 3.65 (s, 1H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ ppm: 133.8 , 131.0 , 129.5, 129.4 , 128.0 , 118.7, 62.8; **IR(KBr)**: 3418, 2957, 2922, 2852, 2361, 2341, 1598, 1409, 1193, 1015, 837..

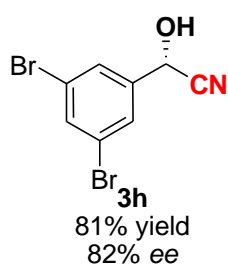


2-Hydroxy-2-(4-phenylphenyl) acetonitrile (**3f**) was obtained according to the general procedure (71% yield). Enantiomeric excess was determined by HPLC with a Chiralcel AD-H column(n-hexane-isopropanol 95:5 V/V, 0.8 mL/min, 210 nm); minor enantiomer $t_r = 21.36$ min, major enantiomer $t_r = 22.59$ min; 71% *ee*; $[\alpha]_D^{18} -25.3^\circ$ (c 1.0, CHCl_3); $^1\text{H NMR}$ (400 MHz , CDCl_3) δ ppm: 7.74-7.76(d, $J = 8\text{Hz}$, 2H), 7.67-7.70(m,4H),7.47-7.50(t, $J=8$ Hz, 2H), 7.38-7.41(t, $J=7.5\text{Hz}$, 1Hz), 6.04-6.05(d, $J=6.4\text{Hz}$, 1H), 5.84-5.86(d, $J=6\text{Hz}$, 1H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ ppm: 147.6, 145.4, 141.5, 134.1, 132.7, 132.5, 132.3, 132.2, 125.0, 67.6; **IR(KBr)**: 3413, 2963, 2922, 2852, 2239, 1487, 1409, 1273, 1193, 1027, 762.

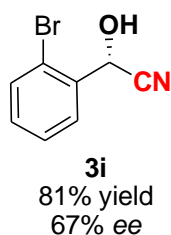


2-Hydroxy-2-(2,6-diChlorophenyl) acetonitrile (**3g**) (yellow solid) was obtained

according to the general procedure (94% yield). Enantiomeric excess was determined by HPLC with a Chiralcel AD-H column(n-hexane-isopropanol 95:5 V/V, 0.8 mL/min, 210 nm); minor enantiomer tr = 13.97 min, major enantiomer tr = 16.577 min; 66% *ee*; $[\alpha]_{\text{D}}^{18} -53.4^{\circ}$ (c 0.8, CHCl₃); $^1\text{H NMR}$ (400 MHz, CDCl₃) δ ppm: 7.27-7.39 (m, 3 H), 6.29 (s, 1 H), 3.64 (s, 1 H); $^{13}\text{C NMR}$ (100 MHz, CDCl₃) δ ppm :135.1 , 131.6 , 129.4, 117.1, 59.1 ; **IR (KBr)**: 3374, 3078, 2934, 2252, 1738, 1560, 1426, 1189, 1035, 861, 744, 664.

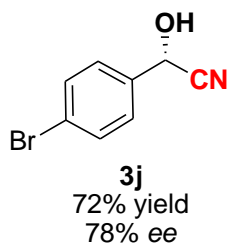


2-Hydroxy-2-(3,5-diBromophenyl)acetonitrile (**3h**) (yellow solid) was obtained according to the general procedure (81% yield). Enantiomeric excess was determined by HPLC with a Chiralcel AD-H column(n-hexane-isopropanol 95:5 V/V, 0.8 mL/min, 210 nm); minor enantiomer tr = 21.36 min, major enantiomer tr = 22.59 min; 82% *ee*. $[\alpha]_{\text{D}}^{18} -15.3^{\circ}$ (c 1.0, CHCl₃); $^1\text{H NMR}$ (400 MHz, CDCl₃) δ ppm:7.71 (s, 1 H), 7.60 (s, 2 H), 5.50 (s, 1 H), 4.10 (s, 1 H); $^{13}\text{C NMR}$ (100 MHz, CDCl₃) δ ppm 138.7 , 135.4 , 128.3, 123.6 , 118.1 , 61.9. **IR (KBr)**: 3388, 3085, 2928, 2852, 2685, 2255, 1564, 1438, 1045, 936, 784, 732, 603.

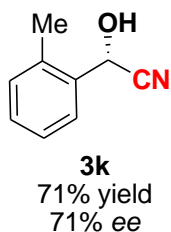


2-Hydroxy-2-(2-Bromophenyl) acetonitrile (**3i**) (yellow oil) was obtained according to the general procedure (81% yield). Enantiomeric excess was determined by HPLC with a Chiralcel OD-H column(n-hexane-isopropanol 95:5 V/V, 1 mL/min, 210 nm); minor enantiomer tr = 13.654 min, major enantiomer tr = 12.608 min; 67% *ee*; $[\alpha]_{\text{D}}^{18} 8.4^{\circ}$ (c 1.0, CHCl₃); $^1\text{H NMR}$ (400 MHz, CDCl₃) δ ppm 7.72 (d, J = 7.74 Hz, 1H), 7.61 (d, J = 7.99 Hz, 1H), 7.42 (t, J = 7.58 Hz, 1H), 7.29 (t, J = 7.70 Hz, 1H), 5.83 (s,

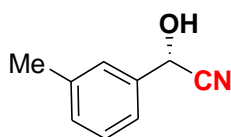
1H), 3.76 (s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ ppm: 134.45, 133.37, 131.31, 128.60, 128.33, 122.35, 118.05, 63.12; IR (KBr): 3389, 3058, 2913, 2805, 2249, 1591, 1571, 1243, 1031, 750, 637, 608.



2-Hydroxy-2-(2-diBromophenyl) acetonitrile (**3j**) was obtained according to the general procedure (72% yield). Enantiomeric excess was determined by HPLC with a Chiralcel AD-H column(n-hexane-isopropanol 95:5 V/V, 0.8 mL/min, 210 nm); minor enantiomer tr = 15.197 min, major enantiomer tr = 12.888 min; 78% ee. $[\alpha]_{\text{D}}^{18} -21.2^{\circ}$ (c 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ ppm: 7.56 (d, J = 8.35 Hz, 1H), 7.37 (d, J = 8.32 Hz, 1H), 5.49 (s, 1H), 4.42-3.14 (m, 1H); ¹³C NMR (100 MHz, CDCl₃) δ ppm: 134.2, 132.4, 128.2, 124.1, 118.6, 62.9; IR (KBr):3391, 1958, 2923, 2852, 2254, 1697, 1592, 1491, 1404, 1038, 841, 797, 615.

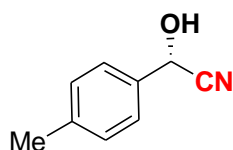


2-Hydroxy-2-(2-Methylphenyl)acetonitrile (**3k**) (yellow oil) was obtained according to the general procedure (71% yield). Enantiomeric excess was determined by HPLC with a Chiralcel AD-H column(n-hexane-isopropanol 95:5 V/V, 0.8 mL/min, 210 nm); minor enantiomer tr = 12.868 min, major enantiomer tr = 11.066 min; 71% ee. $[\alpha]_{\text{D}}^{18} -11.8^{\circ}$ (c1.0, CHCl₃); lit.^[1] $[\alpha]_{\text{D}}+21.3$ (c1.03, CHCl₃), 51% ee (R). ¹H NMR (400 MHz, CDCl₃) δ ppm: 7.22-7.32 (m, 4 H), 5.46 (s, 1 H), 3.60 (s, 1 H), 2.39 (s, 3 H); ¹³C NMR (100 MHz,CDCl₃) δ ppm :139.2, 135.2, 130.6, 129.0, 127.4, 123.7, 119.1, 63.5, 21.3; IR (KBr): 3412, 2957, 2924, 2853, 2247, 1692, 1600, 1491, 1462, 1038, 753.



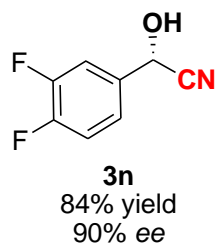
3l
71% yield
78% ee

2-Hydroxy-2-(3-Methylphenyl) acetonitrile (**3l**) (yellow oil) was obtained according to the general procedure (71% yield). Enantiomeric excess was determined by HPLC with a Chiralcel AD-H column(n-hexane-isopropanol 95:5 V/V, 0.8 mL/min, 210 nm); minor enantiomer tr = 13.093 min, major enantiomer tr = 11.292 min; 78% ee. $[\alpha]_{\text{D}}^{18}$ -14.1° (c 1.0, CHCl₃); lit.^[1] $[\alpha]_{\text{D}}^{+41.2}$ (c 2.29, CHCl₃), 91% ee (R). **¹H NMR** (400 MHz, CDCl₃) δ ppm: 7.22-7.33 (m, 4 H), 5.47 (s, 1 H), 3.49 (s, 1 H), 2.39 (s, 3 H); **¹³C NMR** (100 MHz, CDCl₃) δ ppm: 139.2, 135.3, 130.6, 129.1, 127.3, 123.7, 119.0, 63.58, 21.32; **IR (KBr)**: 3325, 2957, 2923, 2853, 2249, 1694, 1609, 1461, 1156, 1038, 793, 747, 700.

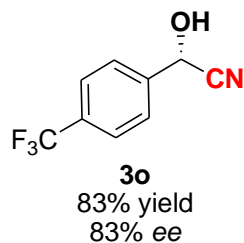


3m
72% yield
75% ee

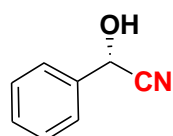
2-Hydroxy-2-(4-Methylphenyl) acetonitrile (**3m**) (yellow oil) was obtained according to the general procedure (72% yield). Enantiomeric excess was determined by HPLC with a Chiralcel AD-H column(n-hexane-isopropanol 95:5 V/V, 0.8 mL/min, 210 nm); minor enantiomer tr = 14.196 min, major enantiomer tr = 12.746 min; 75% ee. $[\alpha]_{\text{D}}^{18}$ -12.3° (c 1.0, CHCl₃); lit.^[1] $[\alpha]_{\text{D}}^{+47.4}$ (c 1.61, CHCl₃), 92% ee (R). **¹H NMR** (400 MHz, CDCl₃)δ ppm: 7.40 (d, J = 7.9 Hz, 2H), 7.24 (d, J = 7.9 Hz, 2H), 5.47 (s, 1H), 3.42 (s, 1H), 2.38 (s, 3 H); **¹³C NMR** (100 MHz, CDCl₃) δ ppm: 139.9, 132.6, 129.8, 126.7, 119.1, 63.4, 21.2; **IR (KBr)**: 3419, 2956, 2923, 2854, 2248, 1681, 1514, 1260, 1037, 929, 814, 762, 606, 528.



2-Hydroxy-2-(3,4-difluorophenyl)acetonitrile (**3n**) (yellow oil) was obtained according to the general procedure (84% yield). Enantiomeric excess was determined by HPLC with a Chiralcel AS-H column(n-hexane-isopropanol (97:3V/V, 0.8 mL/min, 210 nm); minor enantiomer tr = 21.36 min, major enantiomer tr = 33.22 min; 90% ee. $[\alpha]_D^{18}$ -25.6° (c 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ ppm: 7.34 (dd, J = 13.47, 5.31 Hz, 1H), 7.23 (m, 2H), 5.51 (s, 1H), 4.04 (s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ ppm: 151.7-152.4 (d, J_{CF}=14.8, 55Hz, 1C), 149.2 -149.9 (d, J_{CF}=14.8, 55Hz, 1C), 132-132.1 (t, J_{CF}=4.9Hz, 1C), 122.8-122.9 (t, J_{CF}= 8.4Hz, 1C) 118.0-118.4 (d, J_{CF}=16.6, 23.2Hz, 1C), 155.9-116.1 (d, J_{CF}=2, 17.6Hz, 1C), 62.3 (s, 1C); ¹⁹F NMR (470 MHz, CDCl₃) δ ppm: -135.18, -135.22, -135.32, -135.37; IR (KBr): 3417, 2957, 2924, 2854, 2253, 1707, 1616, 1521, 1437, 1289, 1119, 875, 825, 772.

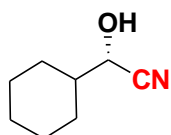


2-hydroxy-2-(4-(trifluoromethyl)phenyl)acetonitrile (**3o**) (yellow oil) was obtained according to the general procedure (83% yield). Enantiomeric excess was determined by HPLC with a Chiralcel AD-H column(n-hexane-isopropanol 95:5 V/V, 0.8 mL/min, 210 nm); minor enantiomer tr = 21.36 min, major enantiomer tr = 22.59 min; 83% ee. $[\alpha]_D^{18}$ -23.9° (c 1.0, CHCl₃); lit.^[1] $[\alpha]_D^{19}$ +19.0 (c1.76, CHCl₃), 80% ee (R). ¹H NMR (400 MHz, CDCl₃) δ ppm: 7.68-7.70 (d, J=8.4Hz, 2 H), 7.61-7.63 (d, J=8.4Hz, 2H), 5.61 (s, 1 H), 4.02 (s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ ppm: 138.7, 126.9, 126.2, 126.1, 118.5, 62.8; ¹⁹F NMR (470 MHz, CDCl₃) δ ppm: -62.90; IR (KBr): 3407, 2926, 2854, 2644, 2254, 1928, 1703, 1422, 1327, 1113, 850, 606.



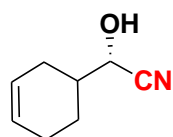
3p
65% yield
71% ee

2-Hydroxy-phenylacetonitrile (**3p**) was obtained according to the general procedure (65% yield). Enantiomeric excess was determined by HPLC with a Chiralcel OD-H column (n-hexane-isopropanol 95:5 V/V, 0.8 mL/min, 210 nm); minor enantiomer tr = 23.449 min, major enantiomer tr = 21.826 min; 71% ee. $[\alpha]_{\text{D}}^{18} -27.3^{\circ}$ (c 1.0, CHCl₃); lit.^[3] $[\alpha]_{\text{D}}^{24} +36.8$ (c 2.0, CHCl₃), 85% ee (R). ¹H NMR (400 MHz, CDCl₃) δ ppm: 7.42-7.52(m, 5H), 5.51(s, 1H), 3.59 (s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ ppm: 135.2, 129.8, 129.2, 126.7, 118.9, 63.6; IR(KBr): 3425, 3064, 2928, 2852, 1485, 1275, 1260, 1192, 1040, 932, 749, 697.



3q
83% yield

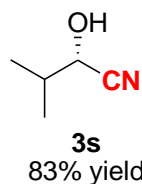
2-cyclohexyl-2-hydroxyacetonitrile (**3q**) was obtained according to the general procedure (83% yield). $[\alpha]_{\text{D}}^{21} -7.3^{\circ}$ (c 1.0, CHCl₃); lit.^[3] $[\alpha]_{\text{D}}^{24} +6.1$ (c = 3.80, CHCl₃) for R enantiomer in 65% ee; ¹H NMR (400 MHz, CDCl₃) δ ppm: 4.24-4.25(d, J=6Hz, 1H), 3.59(s, 1H), 1.68-1.89(m, 6H), 1.10-1.28(m, 5H); ¹³C NMR (100 MHz, CDCl₃) δ ppm: 119.5, 42.2, 28.2, 27.9, 25.9, 25.5, 25.4. IR(KBr): 3448, 2931, 2856, 1636, 1451, 1265, 1043, 747, 704.



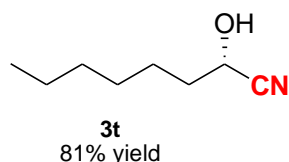
3r
92% yield

2-(cyclohex-3-enyl)-2-hydroxyacetonitrile (**3r**) was obtained according to the general procedure (92% yield). $[\alpha]_{\text{D}}^{21} -4.7^{\circ}$ (c 1.0, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ ppm: 5.64-5.72(dd, J=20.4, 11.6Hz, 2H), 4.34-4.36(dd, J=2.8, 6.8Hz, 1H), 3.41(s, 1H), 1.93-2.26(m, 6H), 1.24(s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ ppm: 127.1,

124.7, 119.4, 38.4, 38.3, 26.7, 26.7, 24.3, 23.9, 23.8. **IR(KBr)**: 3446, 3029, 2920, 2847, 2245, 2092, 1652, 1438, 1266, 1084, 1054, 741, 657.



2-hydroxy-3-methylbutanenitrile(**3s**)was obtained according to the general procedure (83% yield). $[\alpha]_D^{21} -5.3^\circ$ (c1.0,CHCl₃);lit.^[5] $[\alpha]_D^{19} -15.4$ (c=2.1,CHCl₃) ,90%ee,(S)-isomer]; ¹H NMR (400 MHz, CDCl₃) δ ppm: 4.26-4.28(d, J=6Hz, 1H), 3.02(s, 1H), 1.24(s, 1H), 1.05-1.10(m, 6H); ¹³C NMR (100 MHz, CDCl₃) δ ppm:119.2, 67.0, 33.1, 17.7, 17.2.**IR(KBr)**: 3432, 2964, 2917, 2849, 2248, 1651, 1470, 1275, 1260, 1064, 971, 763, 749.



2-hydroxyoctanenitrile(**3t**)was obtained according to the general procedure (81% yield). $[\alpha]_D^{21} -6.1^\circ$ (c 1.0, CHCl₃);lit.^[2] $[\alpha]_D^{26} +9.1$ (c 2.82 , CHCl₃), 66% ee (R). ¹H NMR (400 MHz, CDCl₃) δ ppm: 4.43-4.47(t, J=6.8Hz, 1H), 3.42(s, 1H), 1.80-1.85(dd, J=6.8Hz, 15.2Hz, 2H), 1.44-1.51(td, J=6.8, 11Hz, 2H), 1.24-1.37(m, 6H), 0.86-0.90(t, J=6.4Hz, 3H);¹³C NMR (300 MHz,CDCl₃) δ ppm:120.2, 61.3, 35.1, 31.5, 26.6, 24.5, 22.5, 14.0; **IR(KBr)**: 3448, 2956, 2928, 2829, 2359, 1635, 1457, 1275, 1261, 1070, 748.

Refereces:

[1] B. R. Matthews, W. R. Jackson, G. S. Jayatilake, C. Wilshire, H. A. Jacobs, Aust. J. Chem. 1988, 41, 1697–1709.

[2] H. Nitta, D. Yu, M. Kudo, A. Mori, S. Inoue, J. Am. Chem. Soc. 1992, 114, 7969–7975.

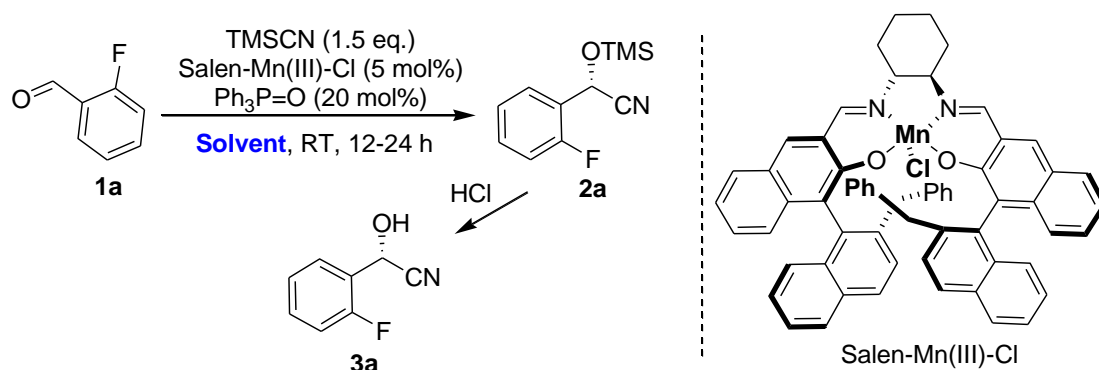
[3] M. Hayashi, Y. Miyamoto, T. Inoue, M. Oguni, J. Org. Chem. 1993,

58, 1515–1522.

[4] S. Han, P. Chen, G. Lin, H. Huang, Z. Li, *Tetrahedron: Asymmetry* 2001, 12, 843–846.

[5] Y. Hamashima, D. Sawada, M. Kanai, M. Shibasaki, *J. Am. Chem. Soc.* 1999, 121, 2641 - 2642.

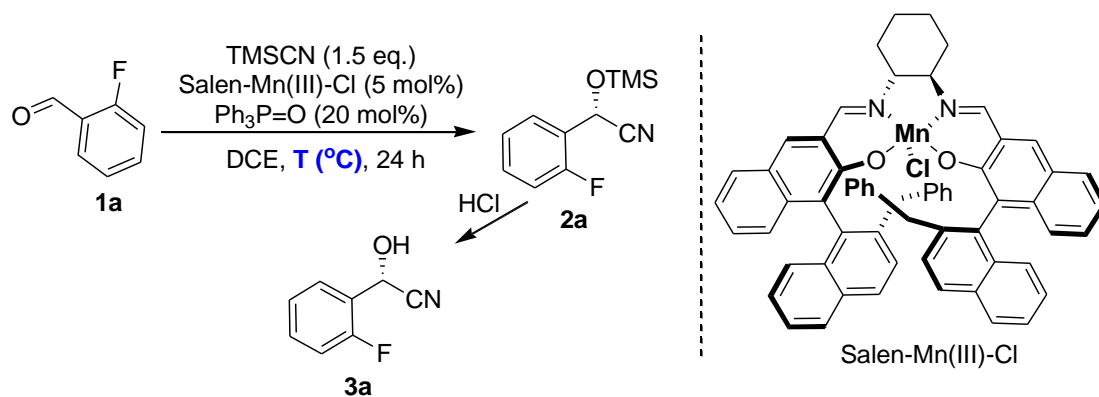
Table S1. Screening of solvents in the Salen-Mn catalyzed cyanosilylation reaction of aldehyde **1a**



Entry	Solvent	Time (h)	Yield (%)	<i>Ee</i> (%)
1	DCM	12	90	64
2	Toluene	12	54	30
3	THF	12	69	44
4	Et ₂ O	12	49	23
5	CH ₃ CN	12	56	11
6	EtOH	12	54	6
7	1,4-dioxane	12	56	6
8	PhCF ₃	12	82	45
9	CHCl ₃	12	41	38
10	Xylene	12	81	36
11	DCE	24	91	79
12	PhCl	24	89	60

13	PhBr	24	86	59
14	CH ₂ Br ₂	24	71	52
15	<i>i</i> -PrOH	24	51	-7

Table S2. The effect of temperature on the Salen-Mn catalyzed cyanosilylation reaction of aldehyde **1a**



Entry	Temperature (°C)	Yield (%)	<i>Ee</i> (%)
1	20	91	79
2	0	92	83
3	-5	90	82
4	-10	94	88
5	-20	82	67
6	-30	33	56

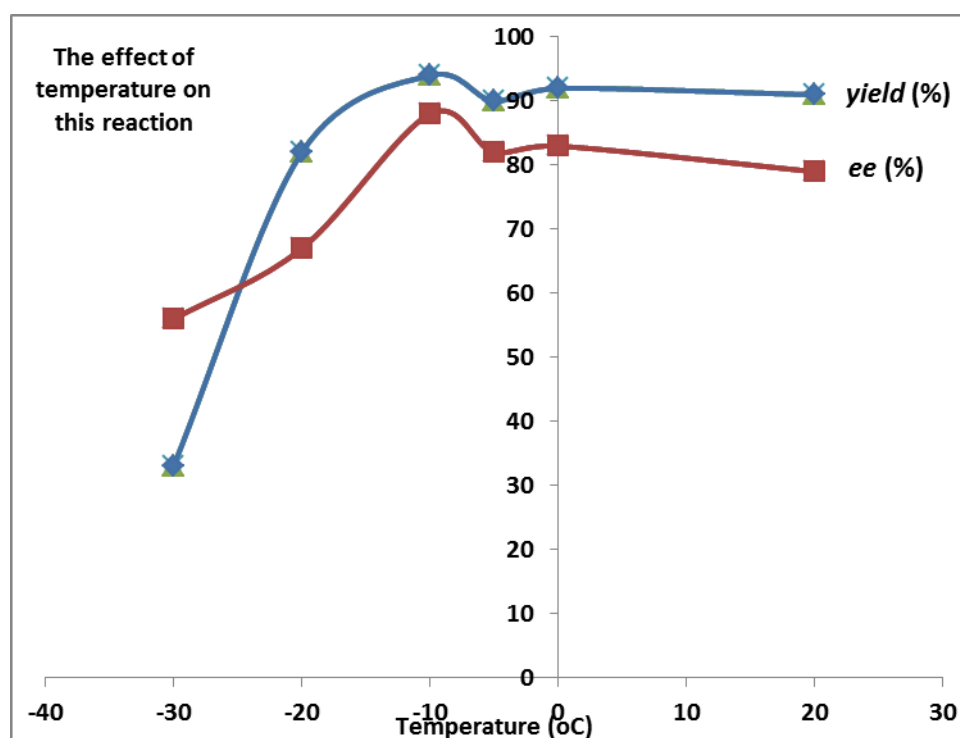
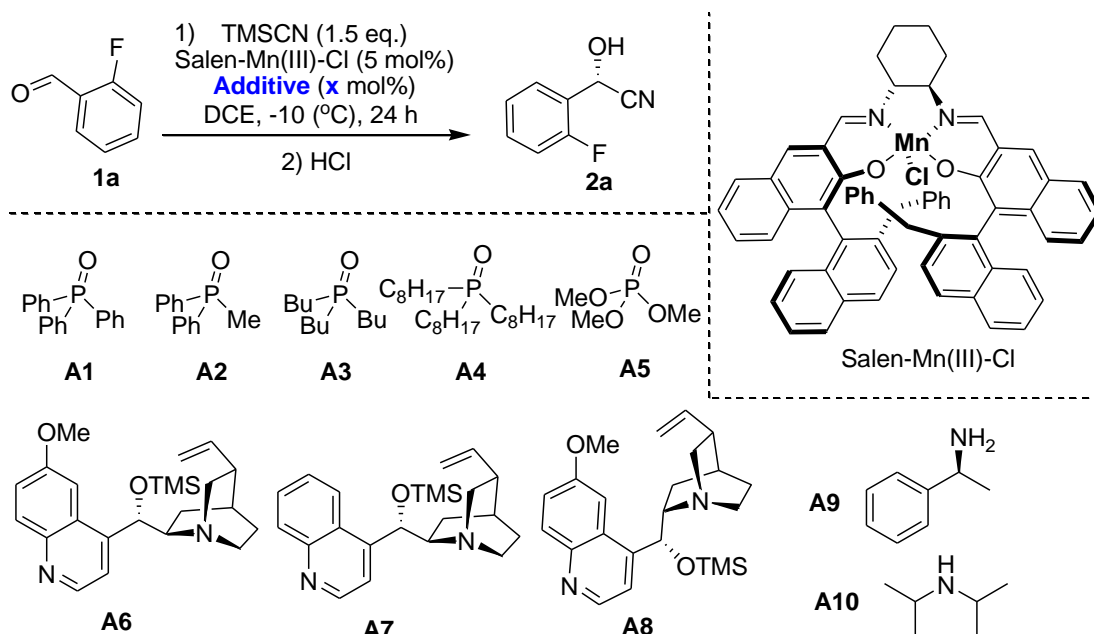


Table S3. The effect of phosphine oxide on the Salen-Mn catalyzed cyanosilylation reaction of aldehyde **1a**^[a]



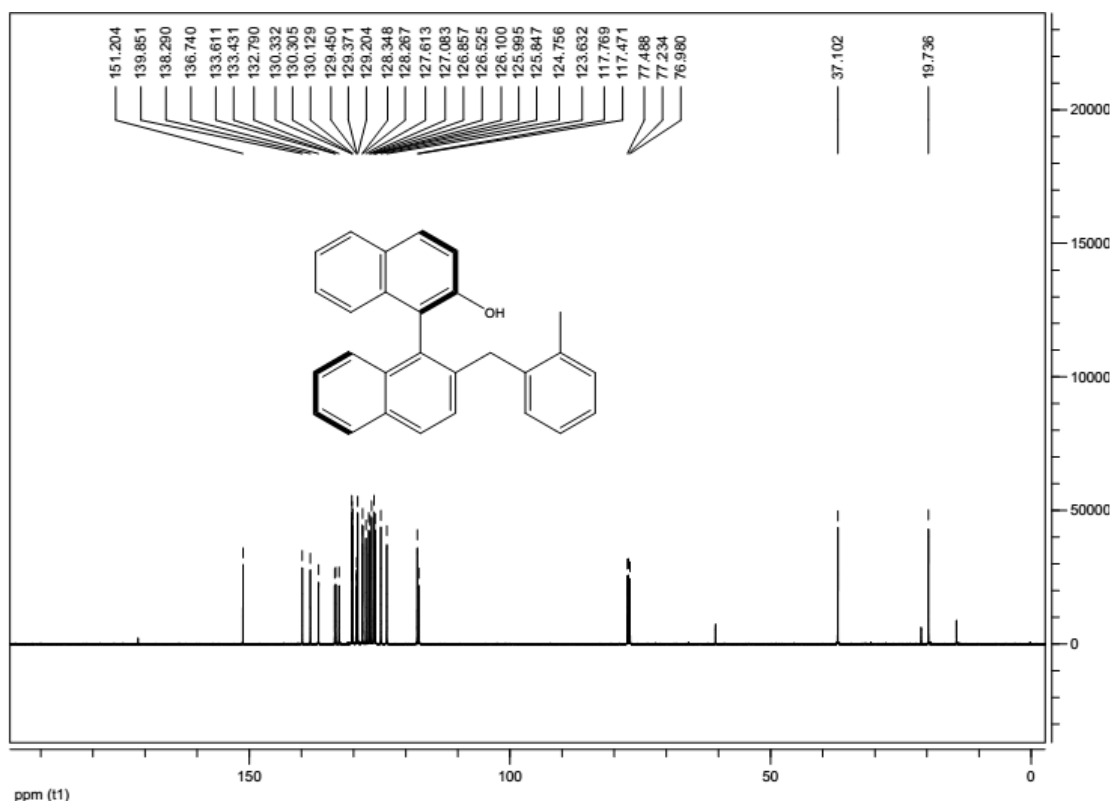
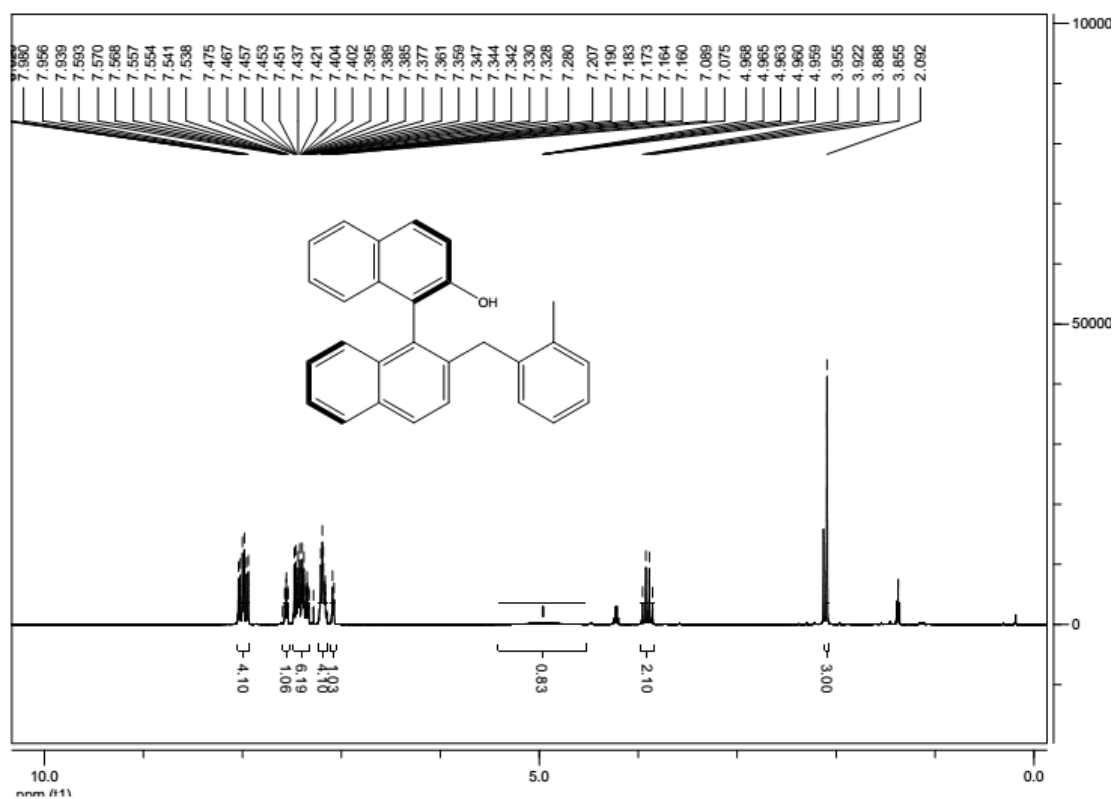
Entry	Additive (x mol%)	Yield (%) ^[b]	<i>Ee</i> (%) ^[c]
1	A1 (10)	90	85
2	A1 (20)	94	88
3	A1 (30)	95	86
4	A1 (40)	61	68
5	A1 (50)	95	85
6	A1 (75)	90	84
7	A1 (100)	78	80
8	A2 (20)	61	72
9	A3 (20)	71	71
10	A4 (20)	73	74
11	A5 (20)	79	73
12	AgF (10) ^[c]	<5	-
13	AgSbF ₆ (10) ^[c]	<5	-
14	AgClO ₄ (10) ^[c]	<5	-
15	AgOTf (10) ^[c]	<5	-

16	A6 (10)^[c]	73	67
17	A7 (10)^[c]	71	67
18	A8 (10)^[c]	70	64
19	A9 (10)^[c]	<20	50
20	A10 (10)^[c]	<20	65
21	DMAP (10)^[c]	<20	32
22	DIPEA (10)^[c]	<20	51
23	H₂O₂ (10)^[c]	<20	36
24	PhIO (5)^[c]	40	32
25	NFSI (10)^[c]	<10	72
26	Selectfluor (10)^[c]	<10	76

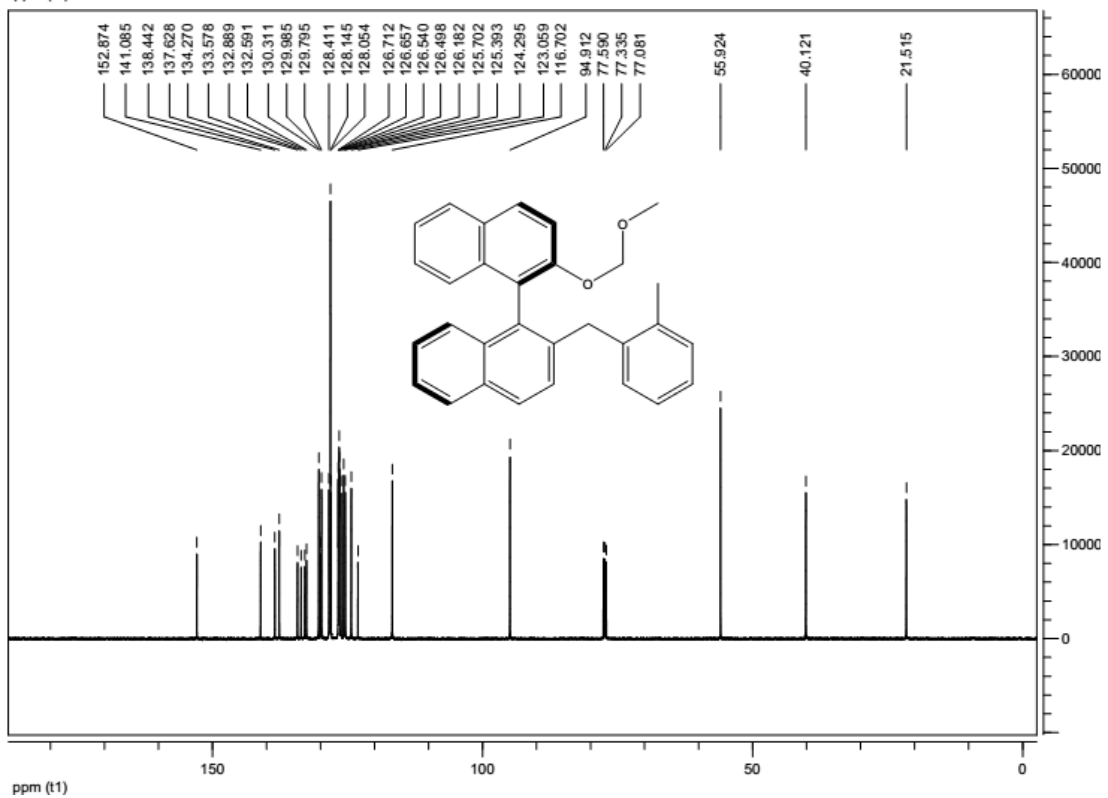
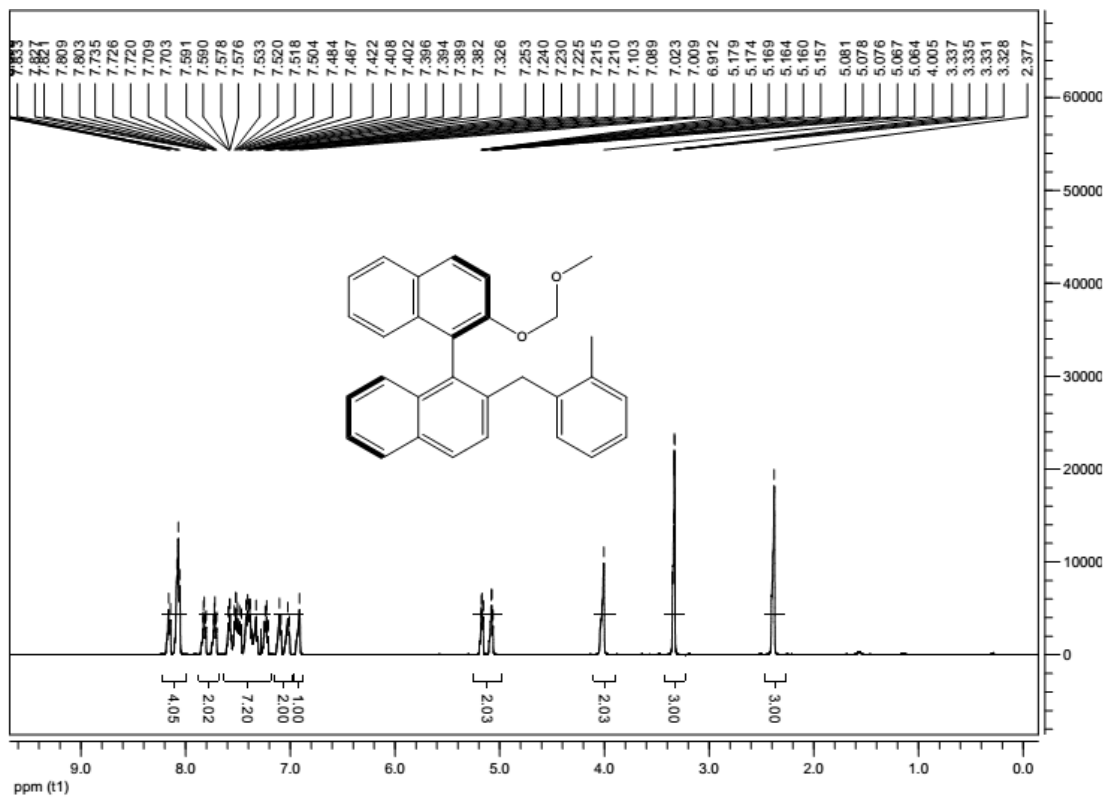
[c] Entries 12-26, except the second additive, the use of 20 mol% of triphenylphosphine oxide was also necessary.

S-5. NMR Charts of Salen ligands and products

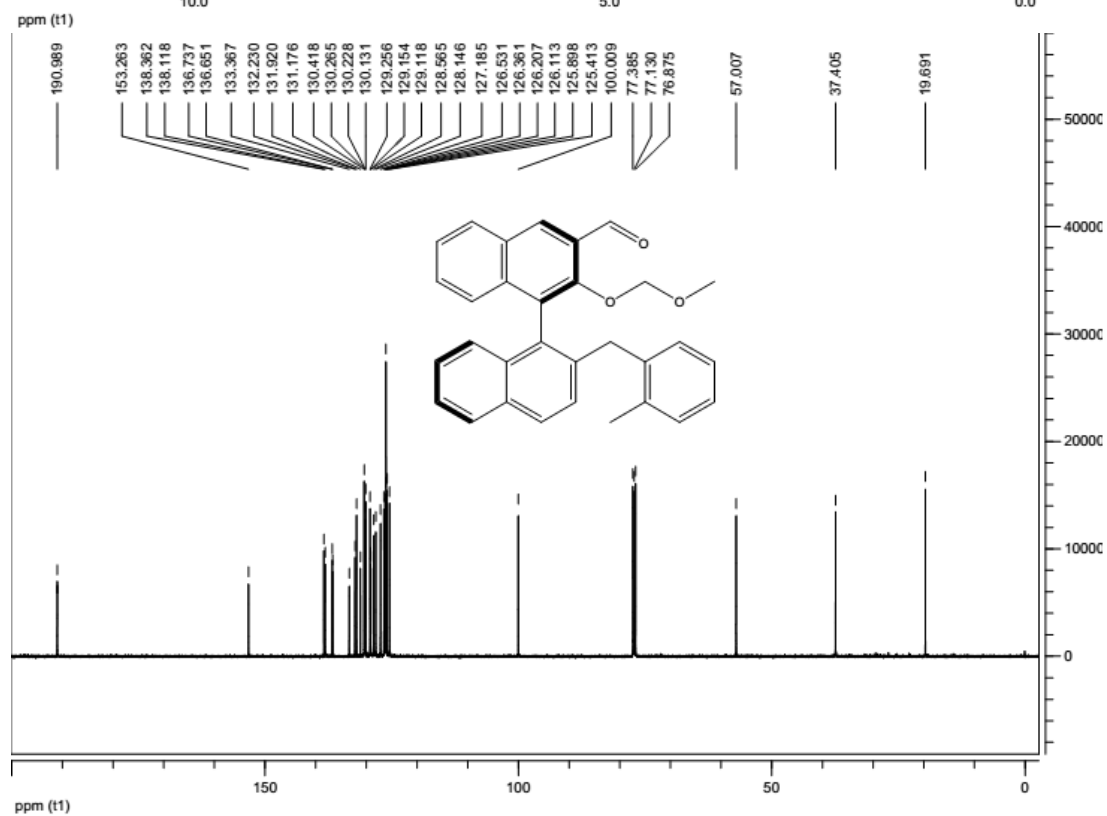
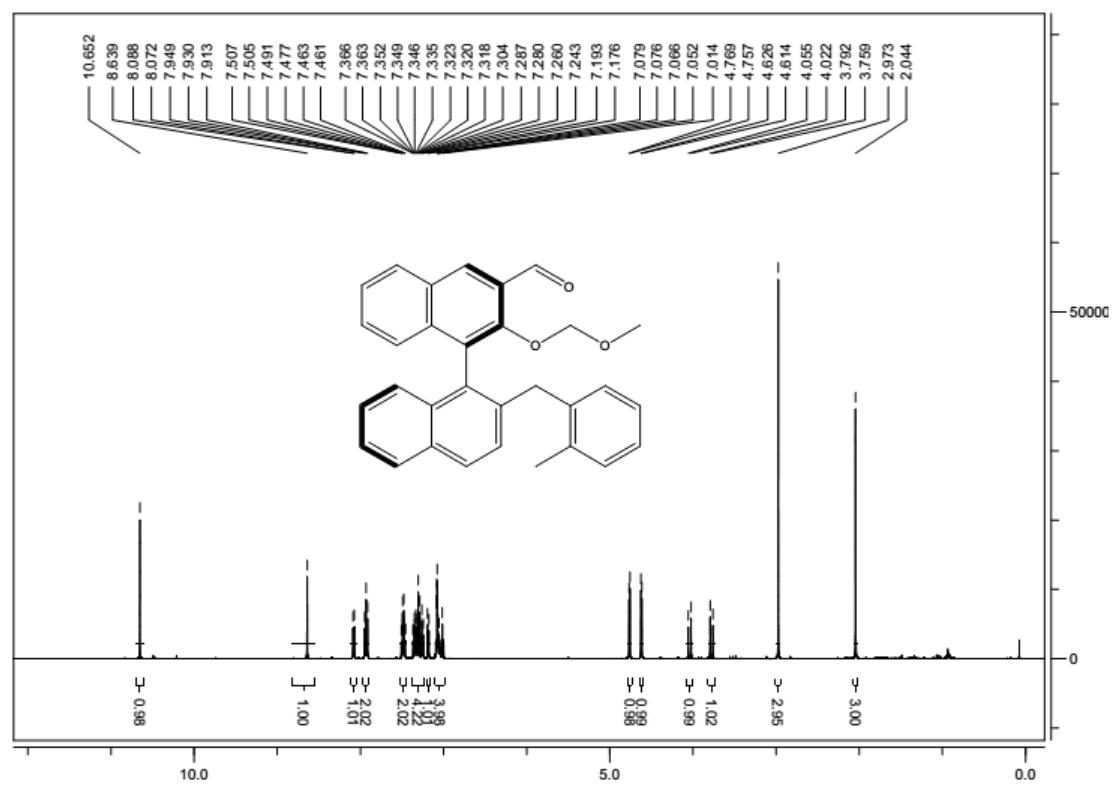
Salen 1c-A



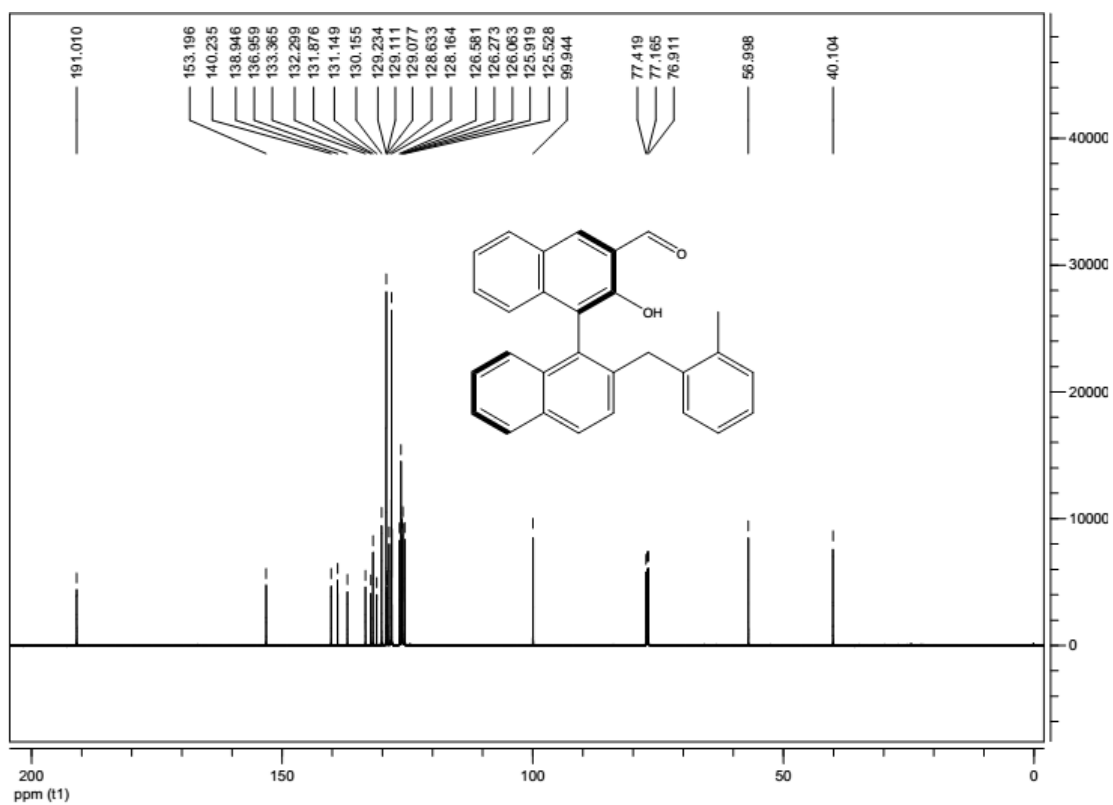
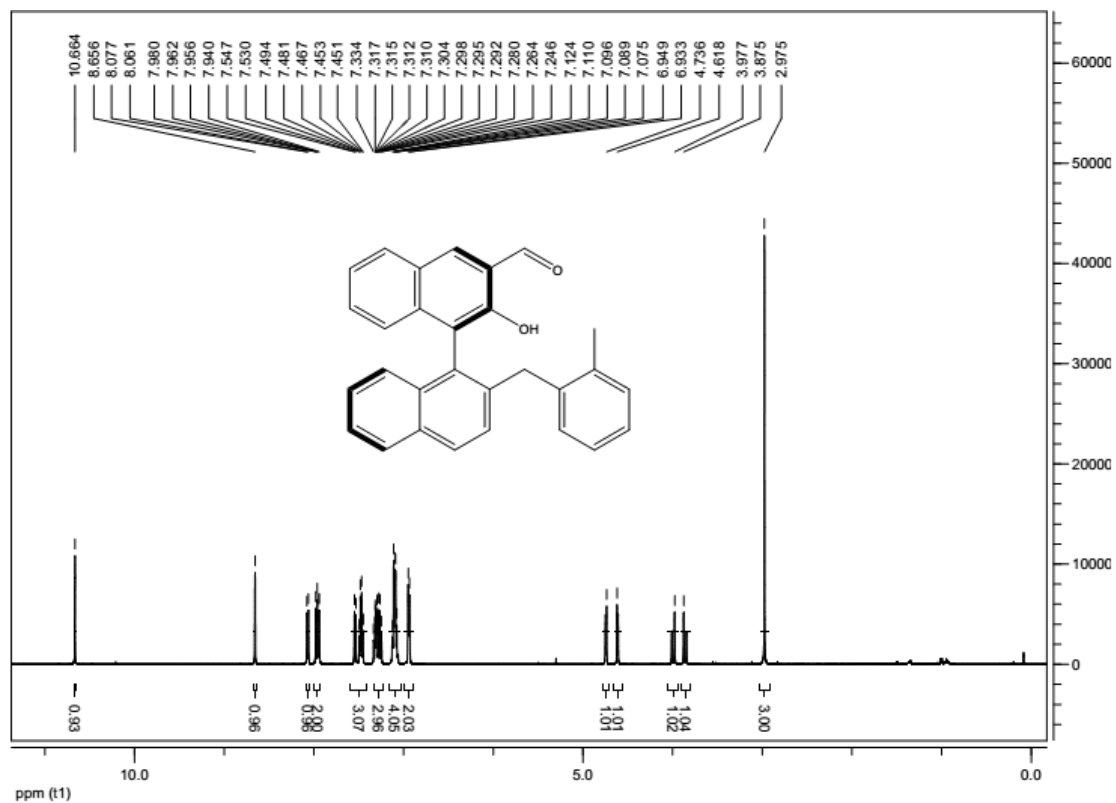
Salen 1c-B



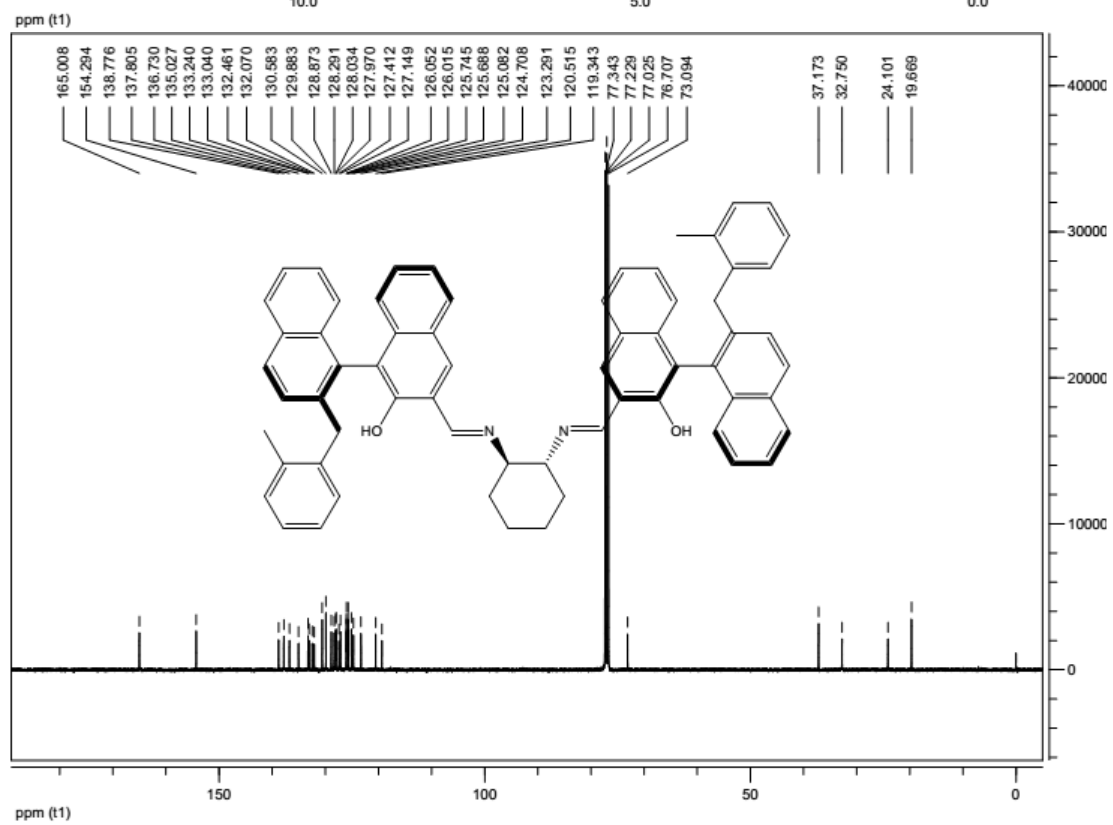
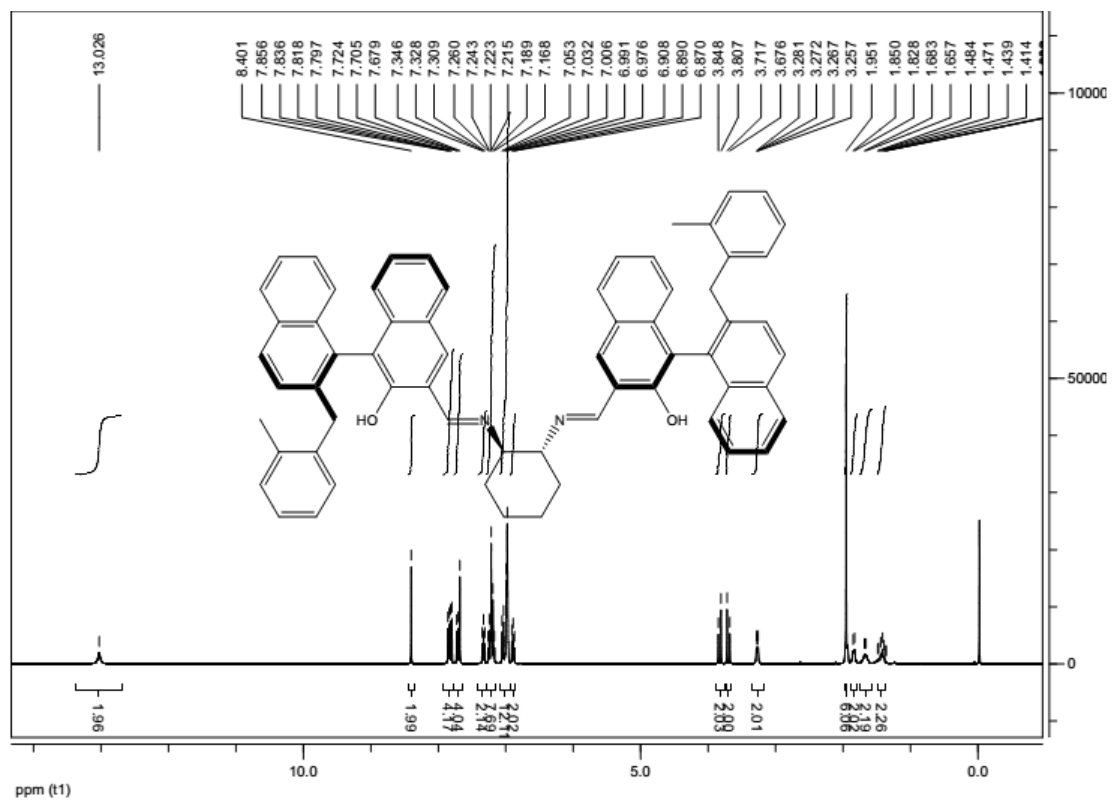
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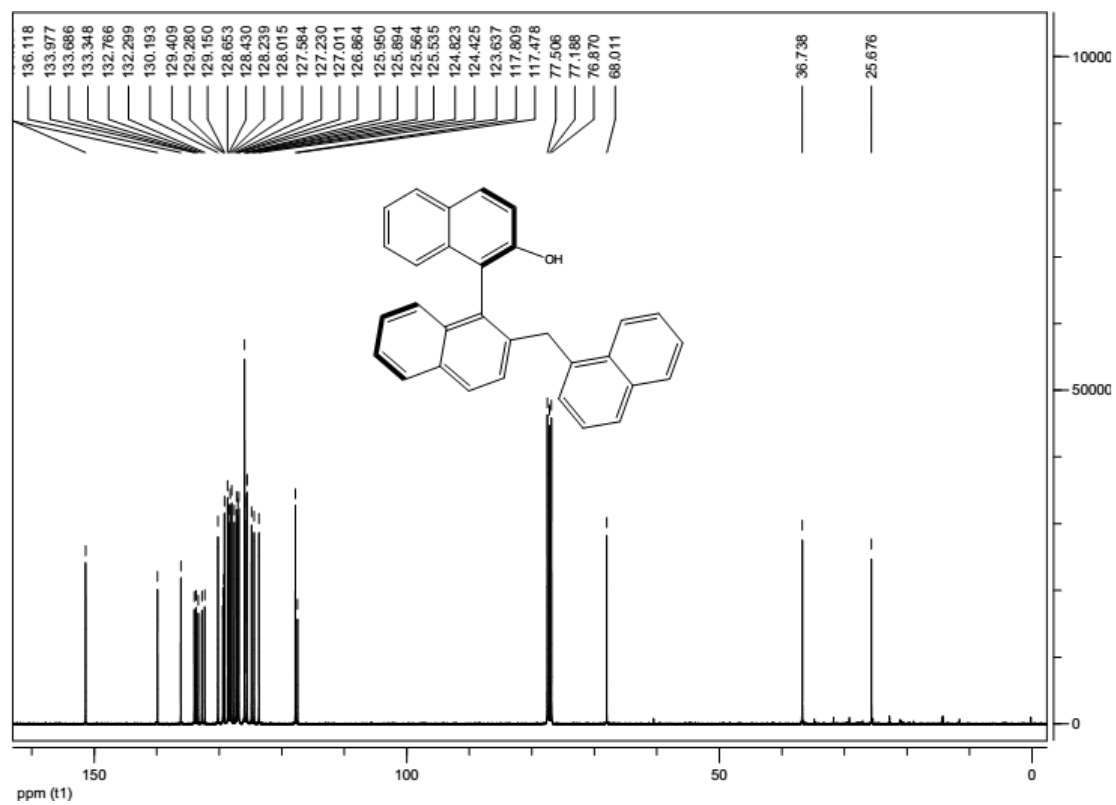
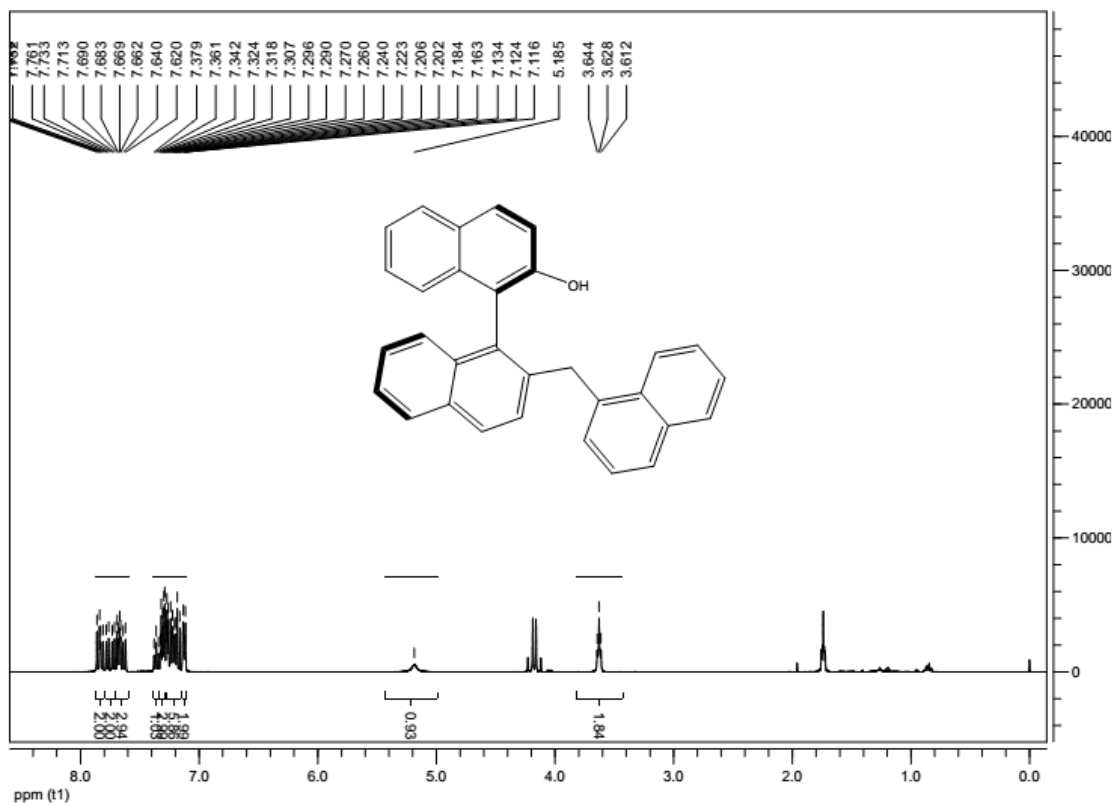
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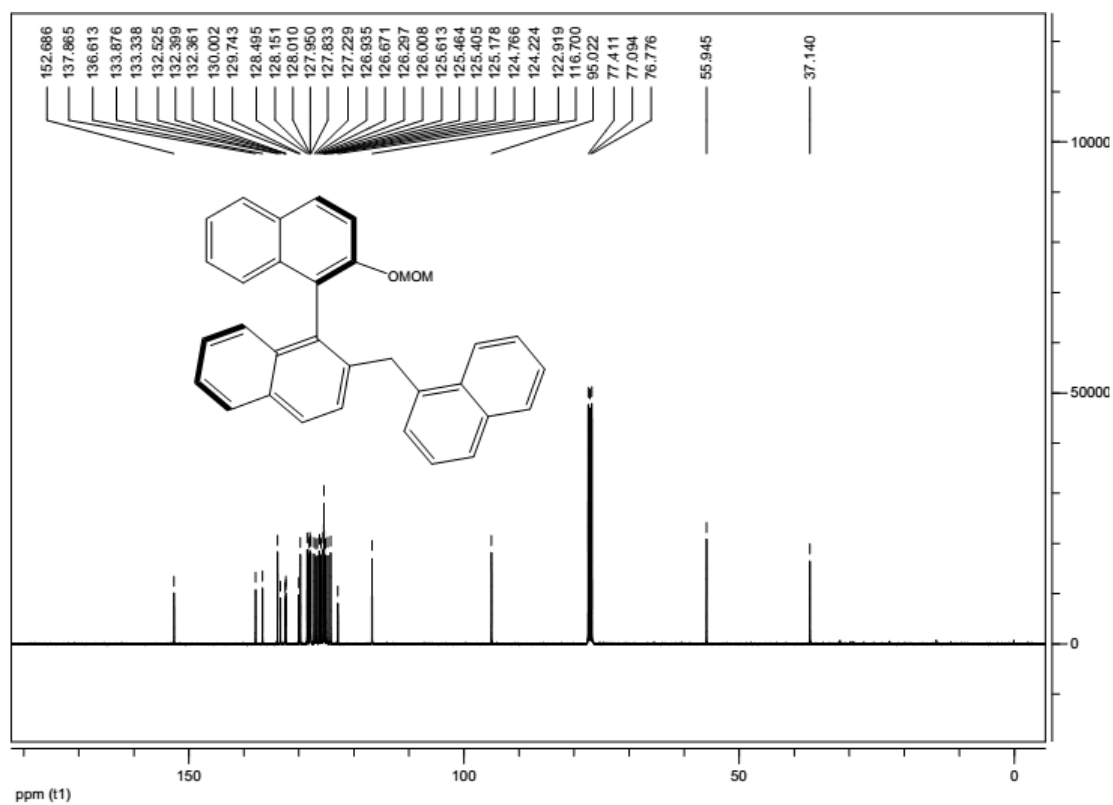
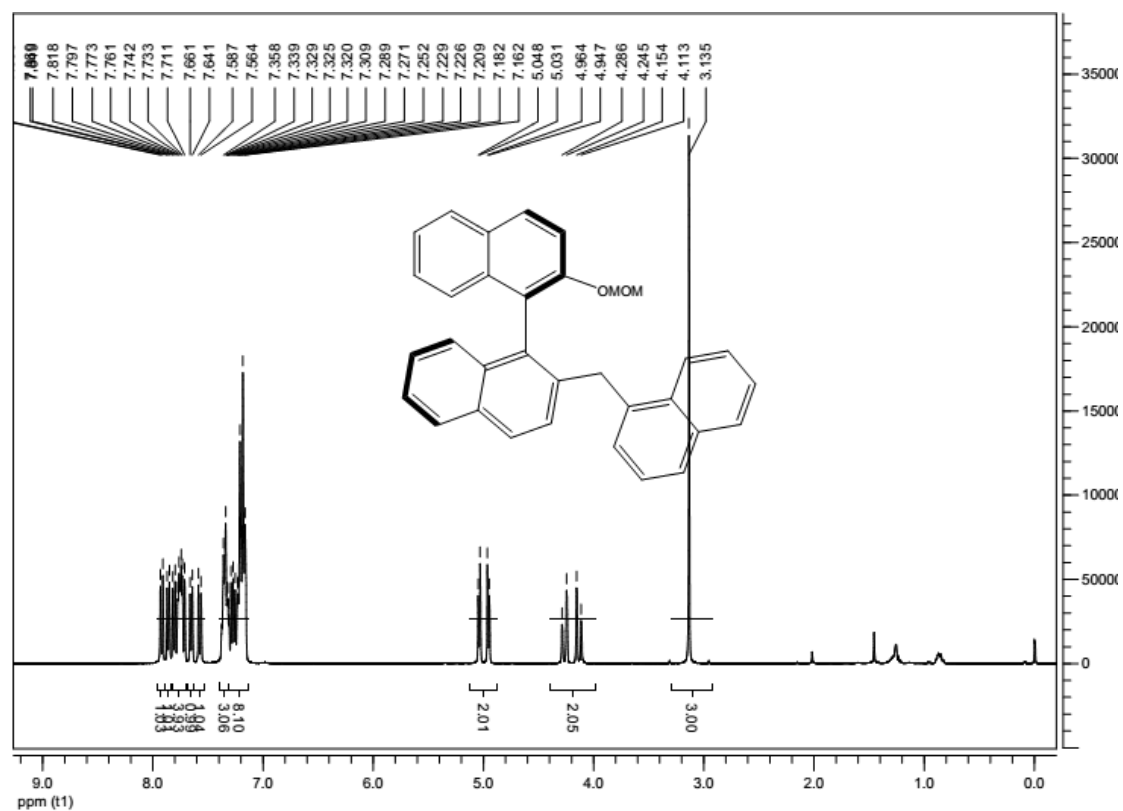
Salen 1c



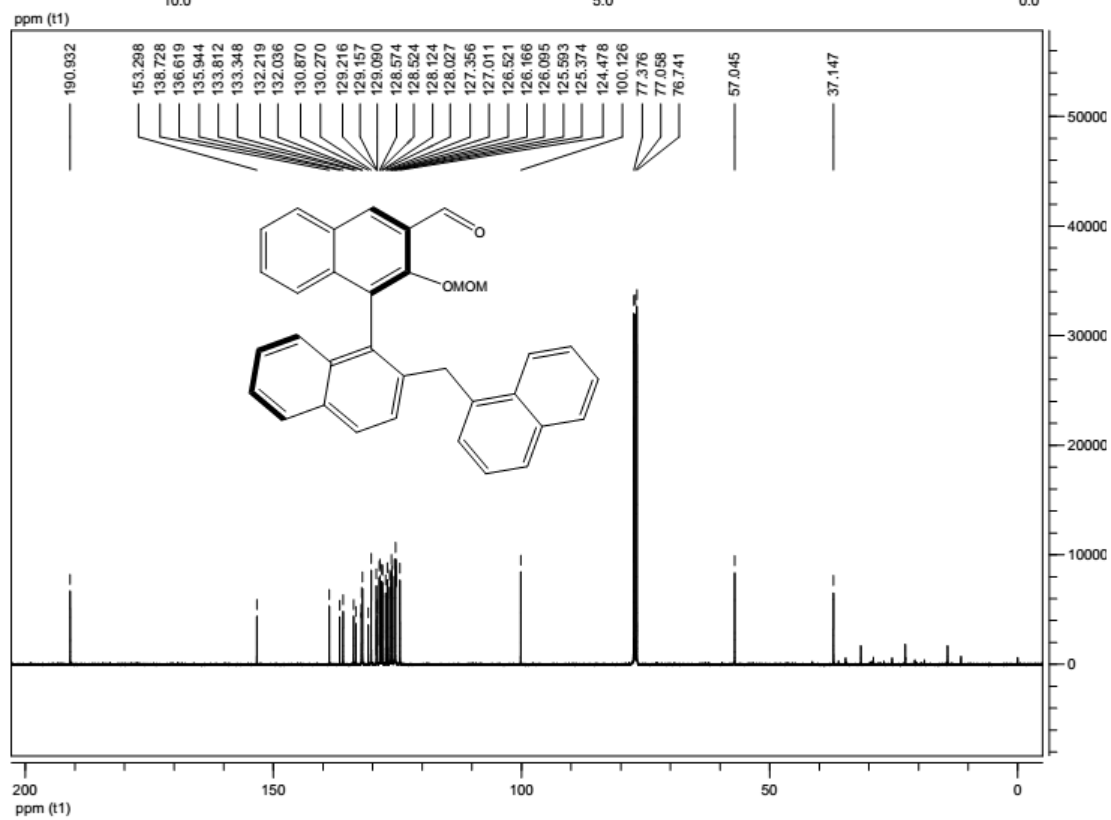
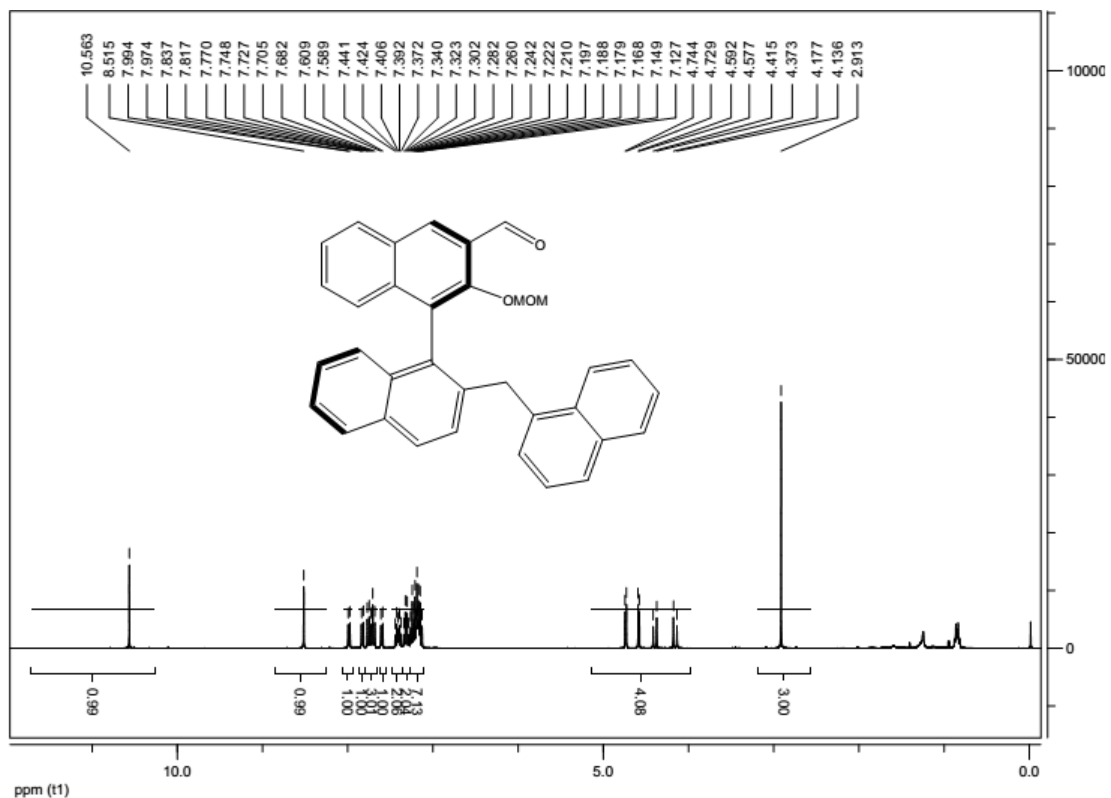
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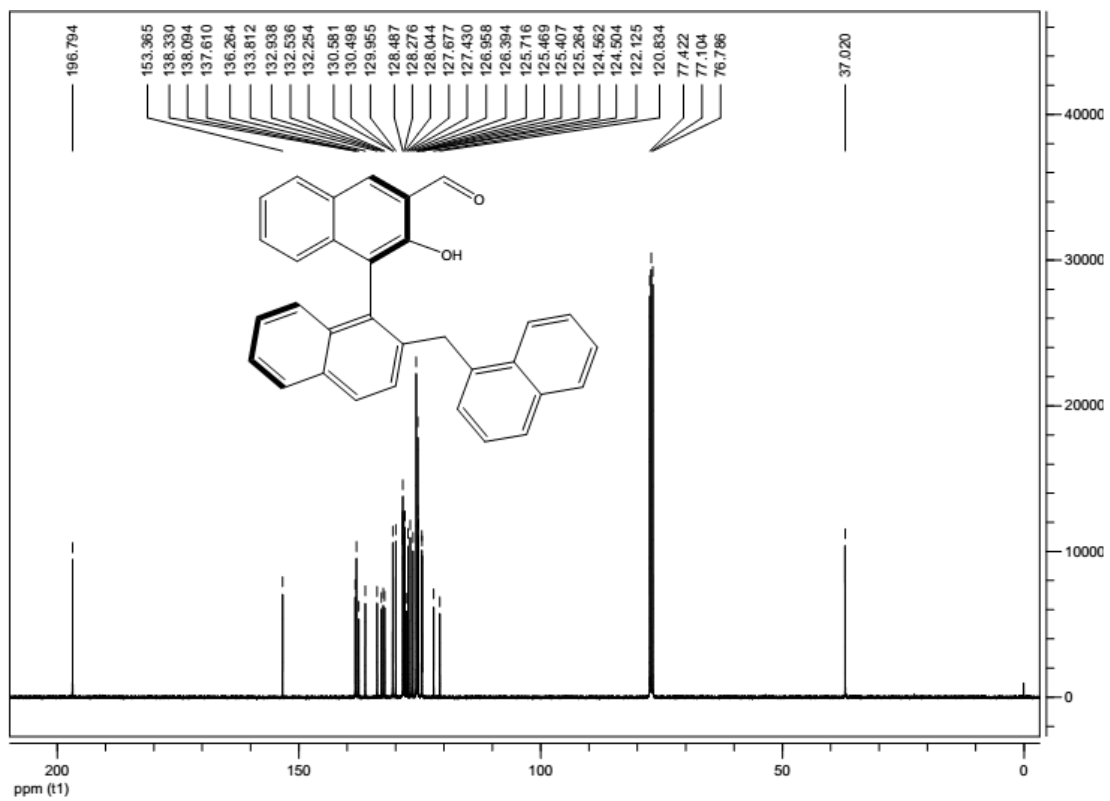
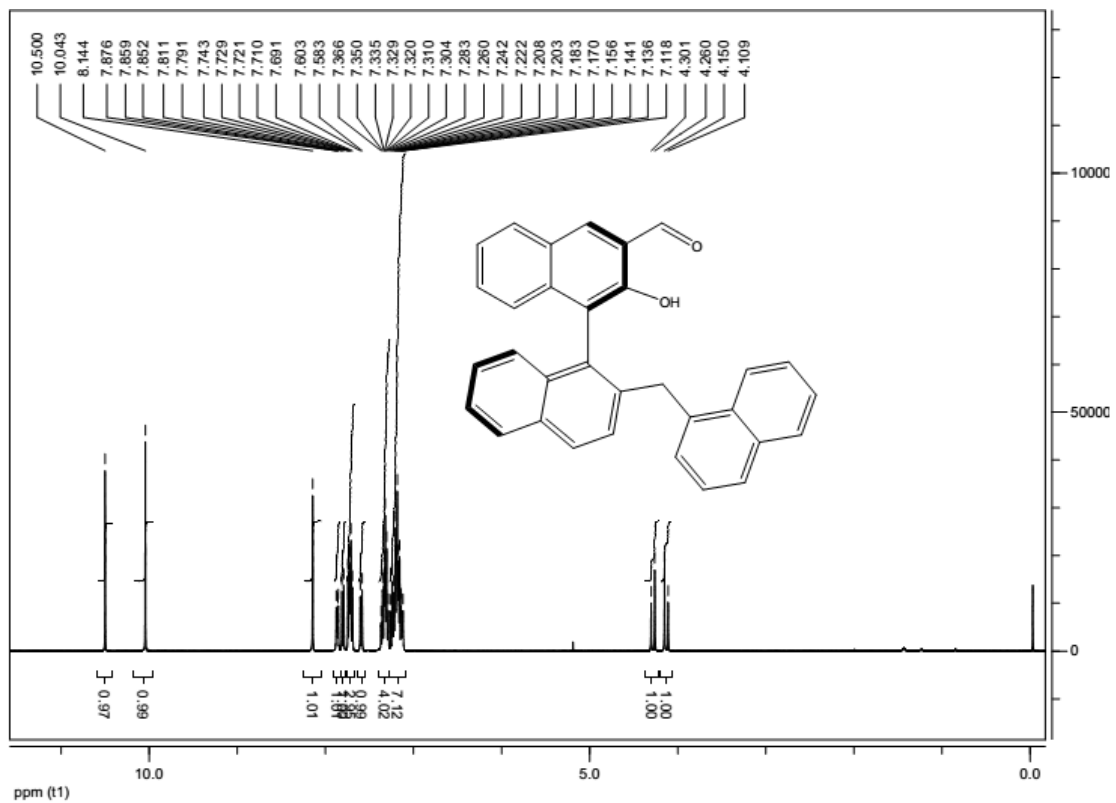
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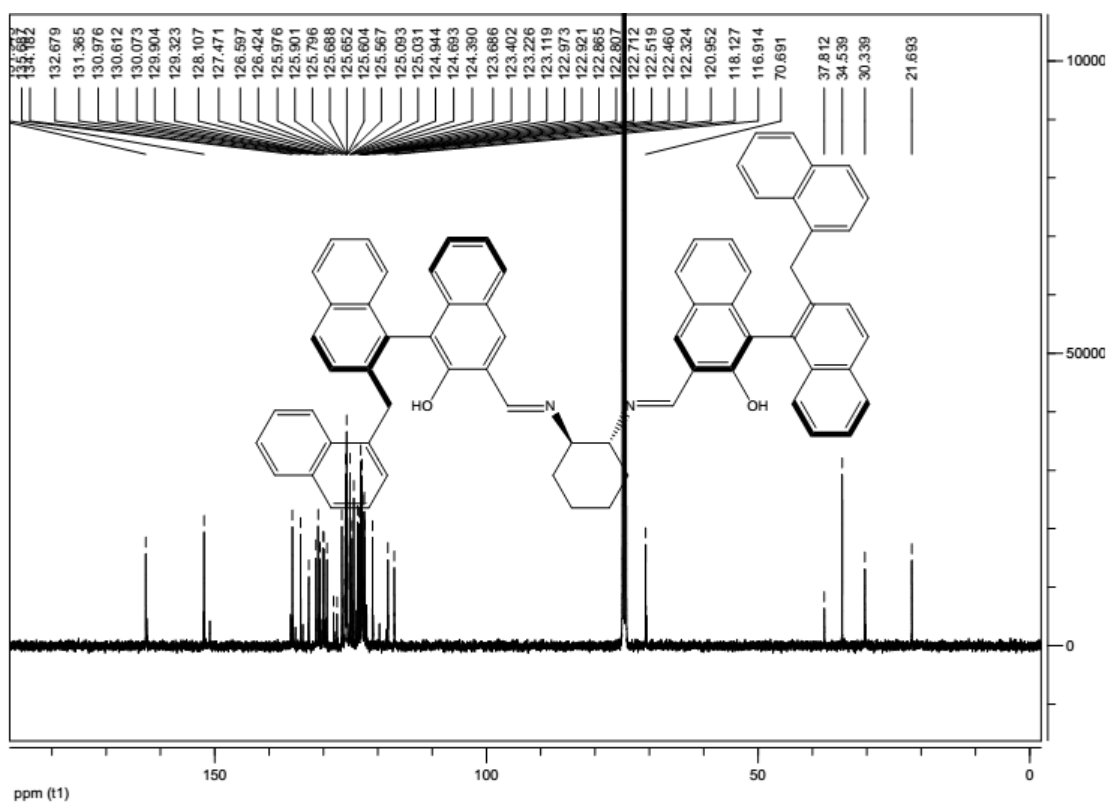
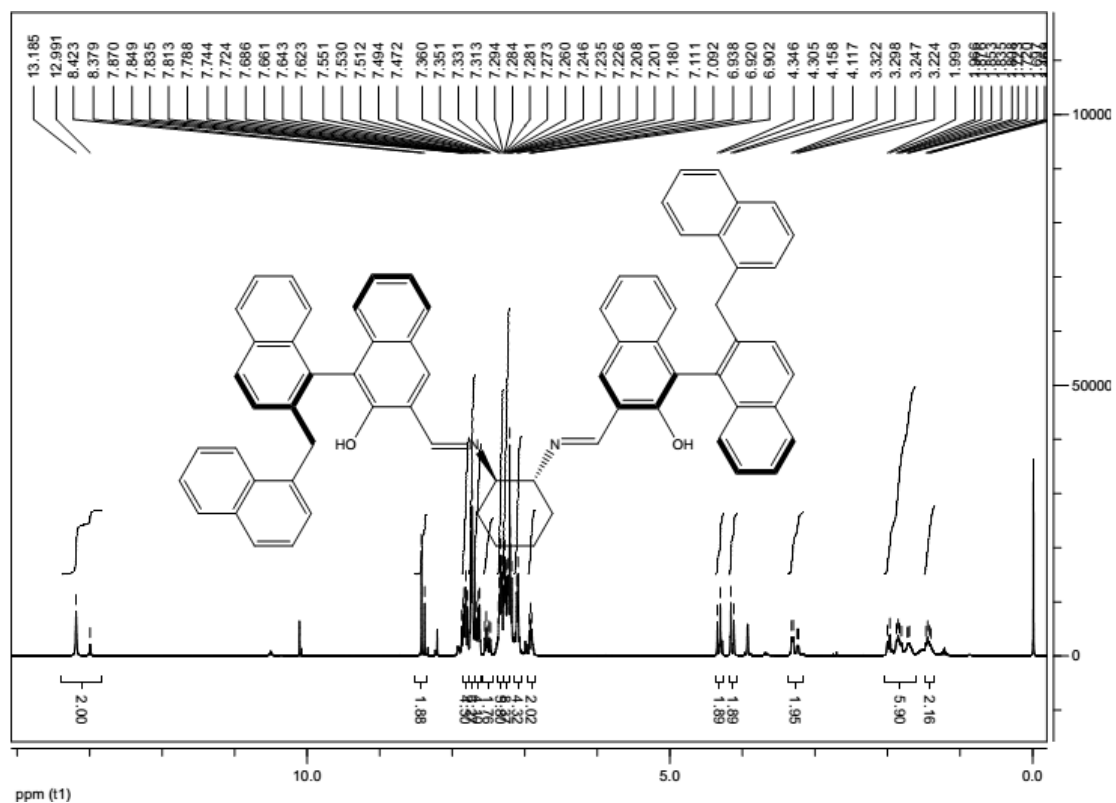
Salen 1d-C:



Salen 1d-D:

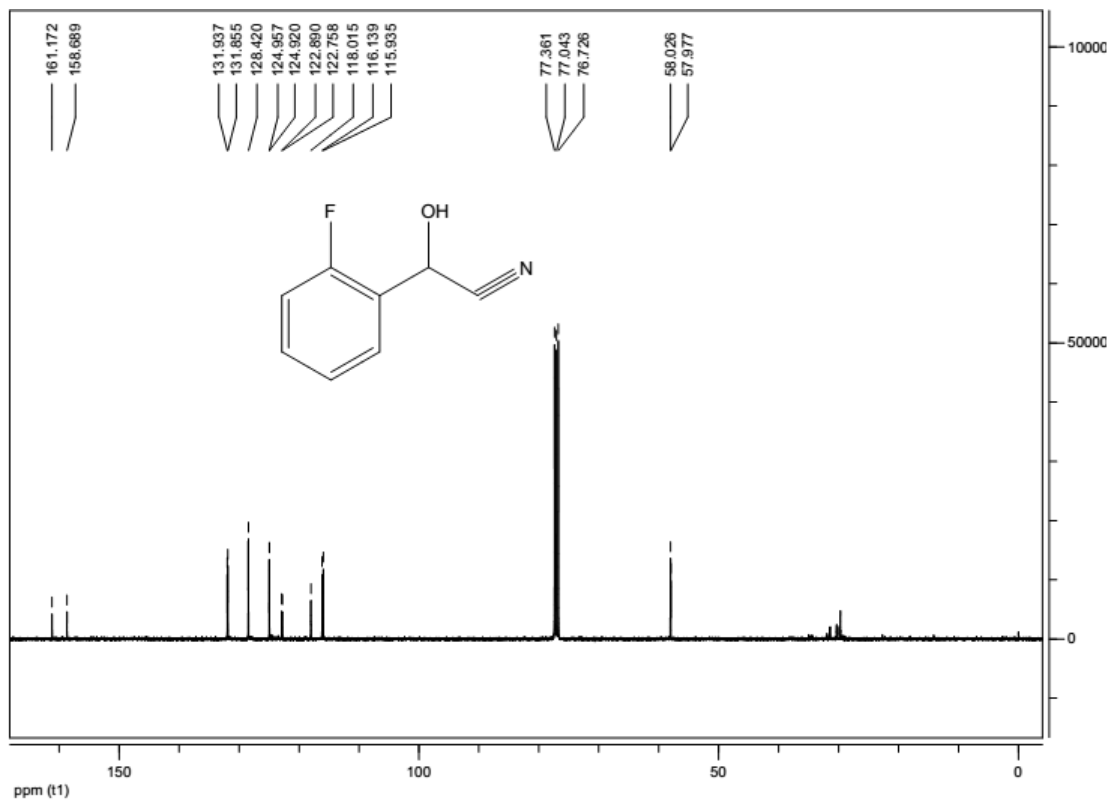
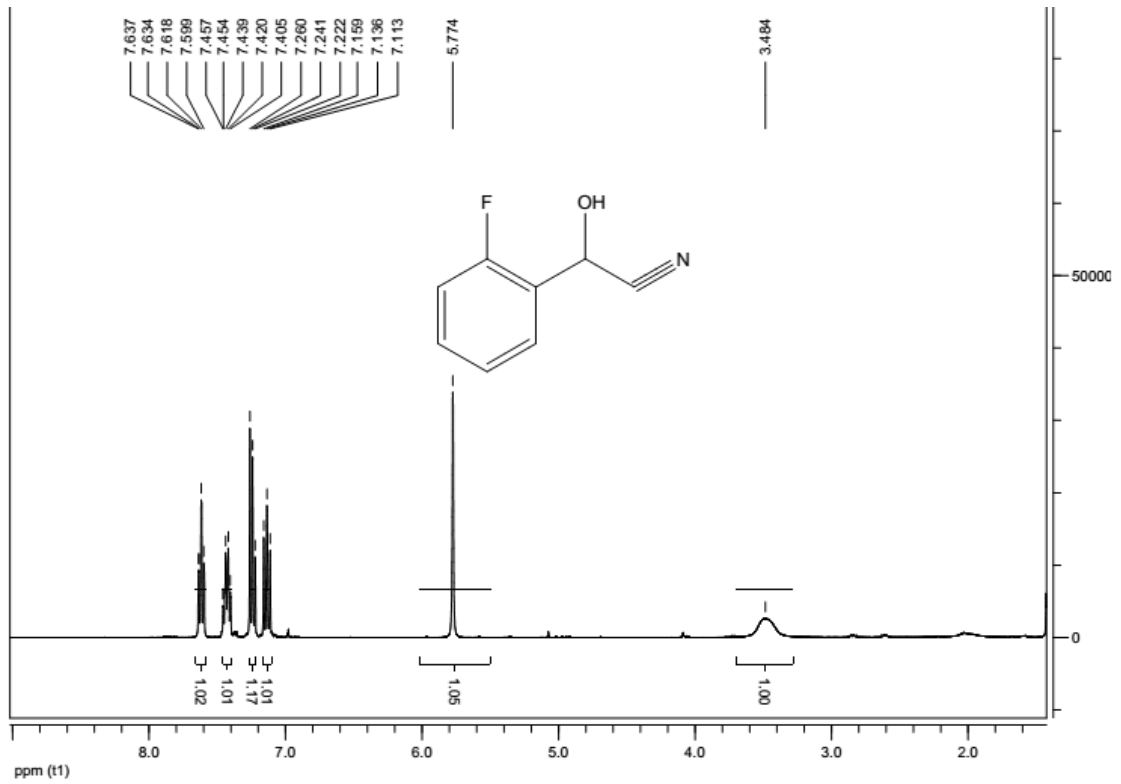


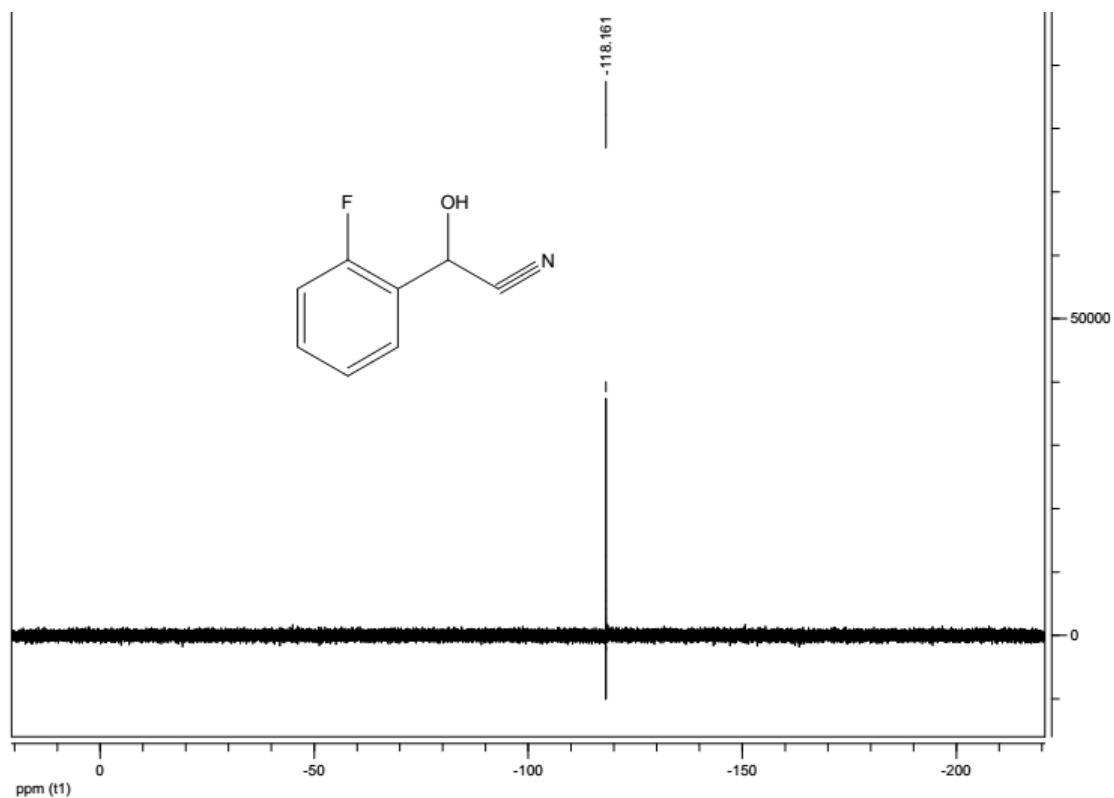
Salen 1d



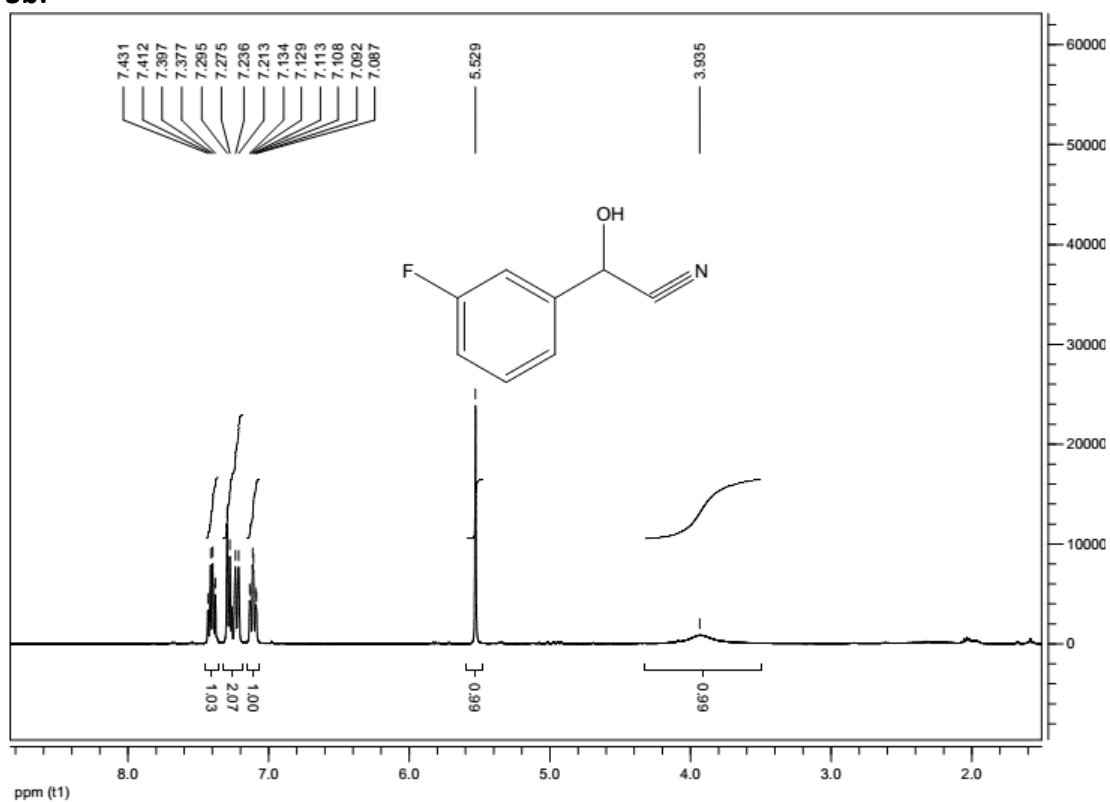
NMR Charts of Compounds:

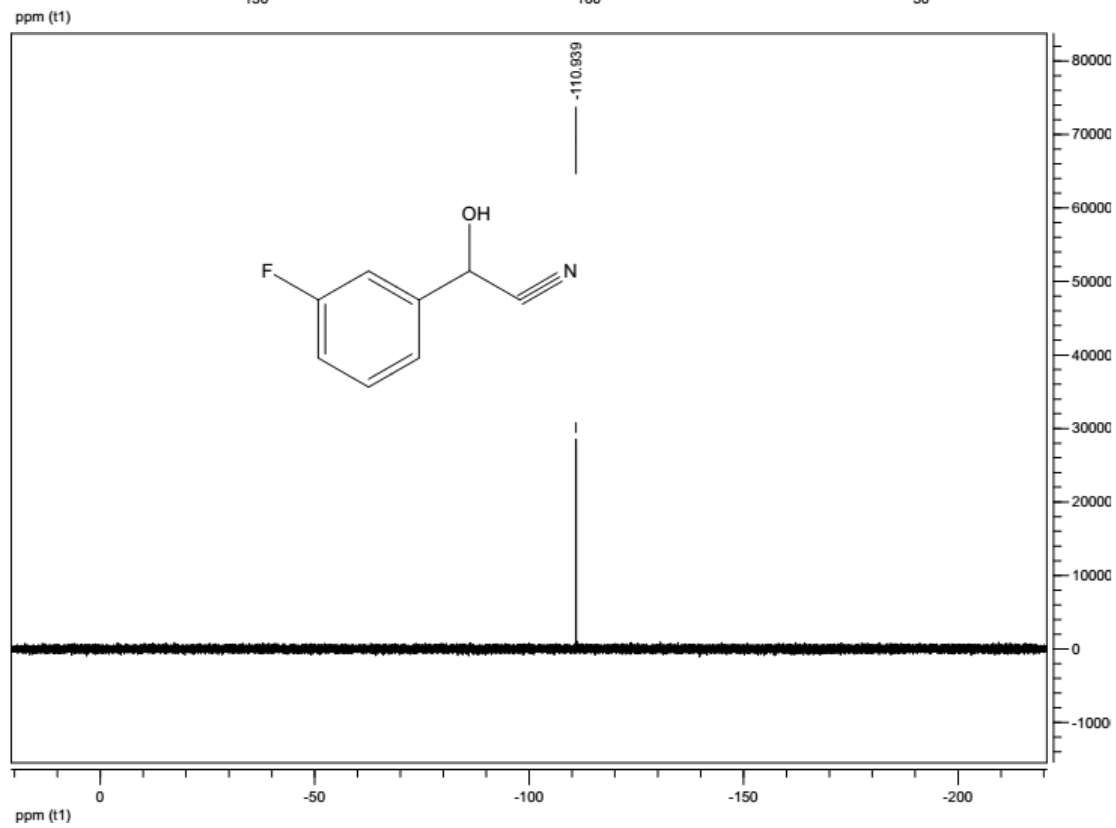
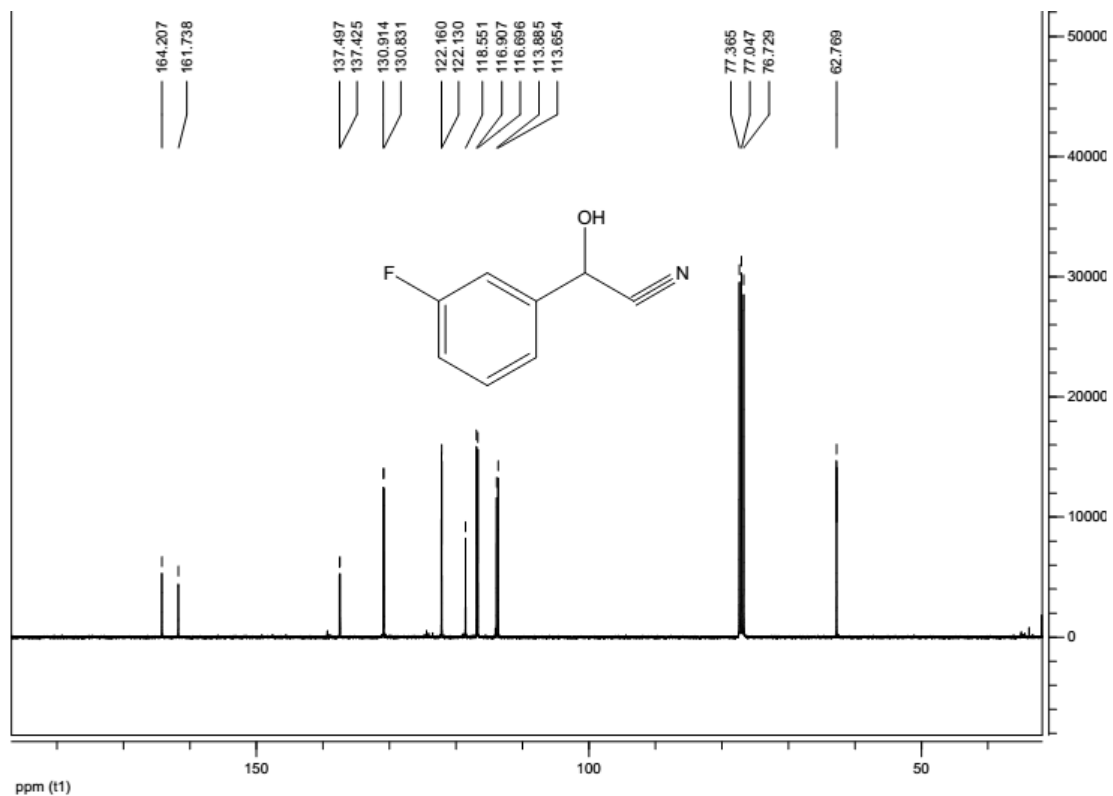
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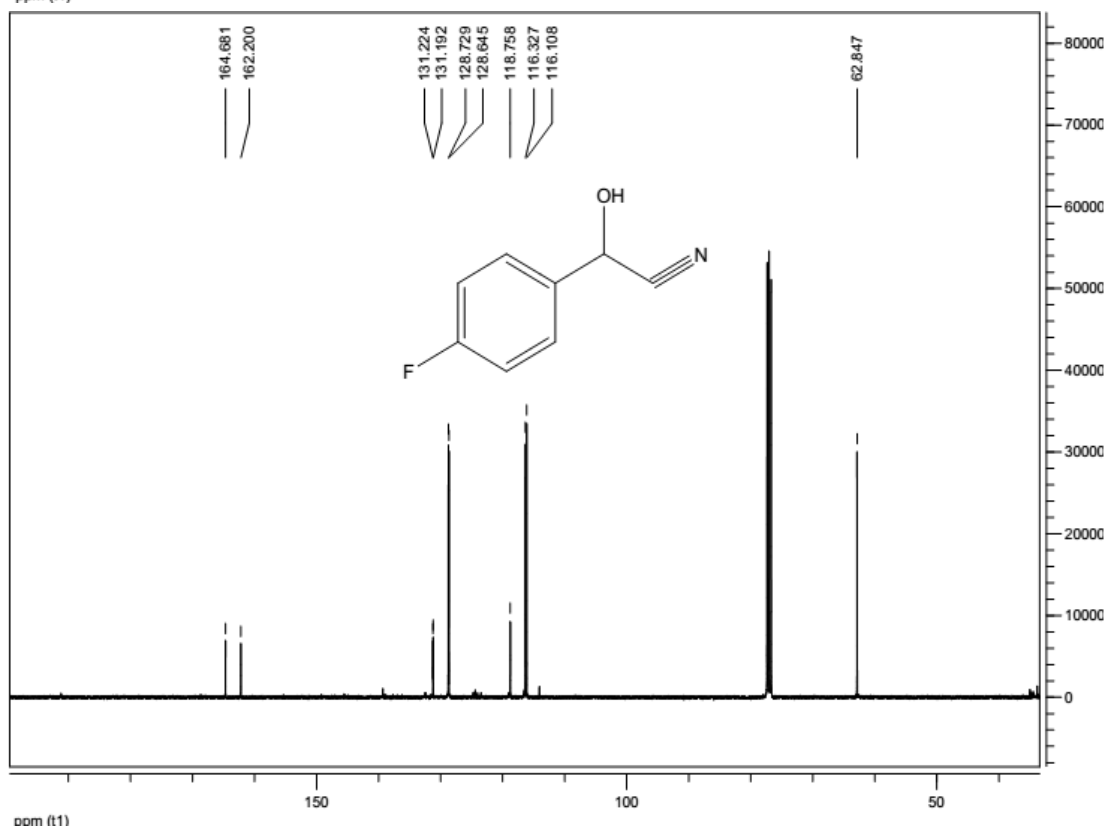
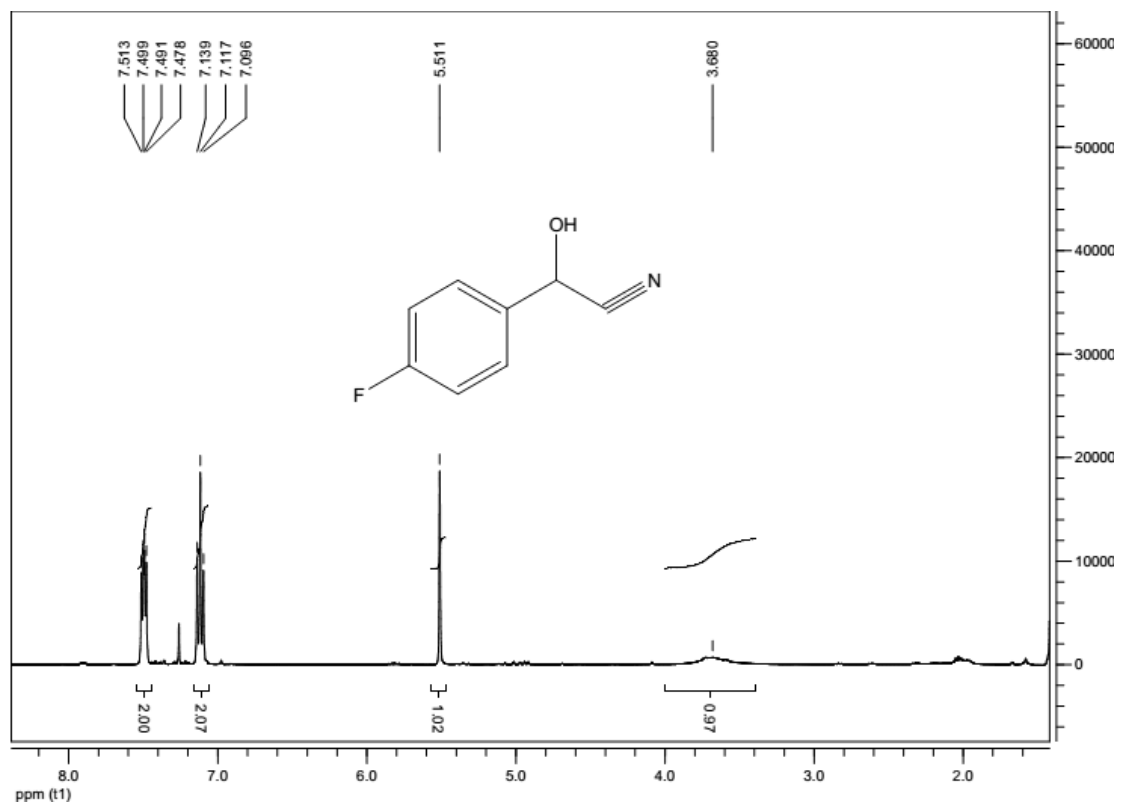


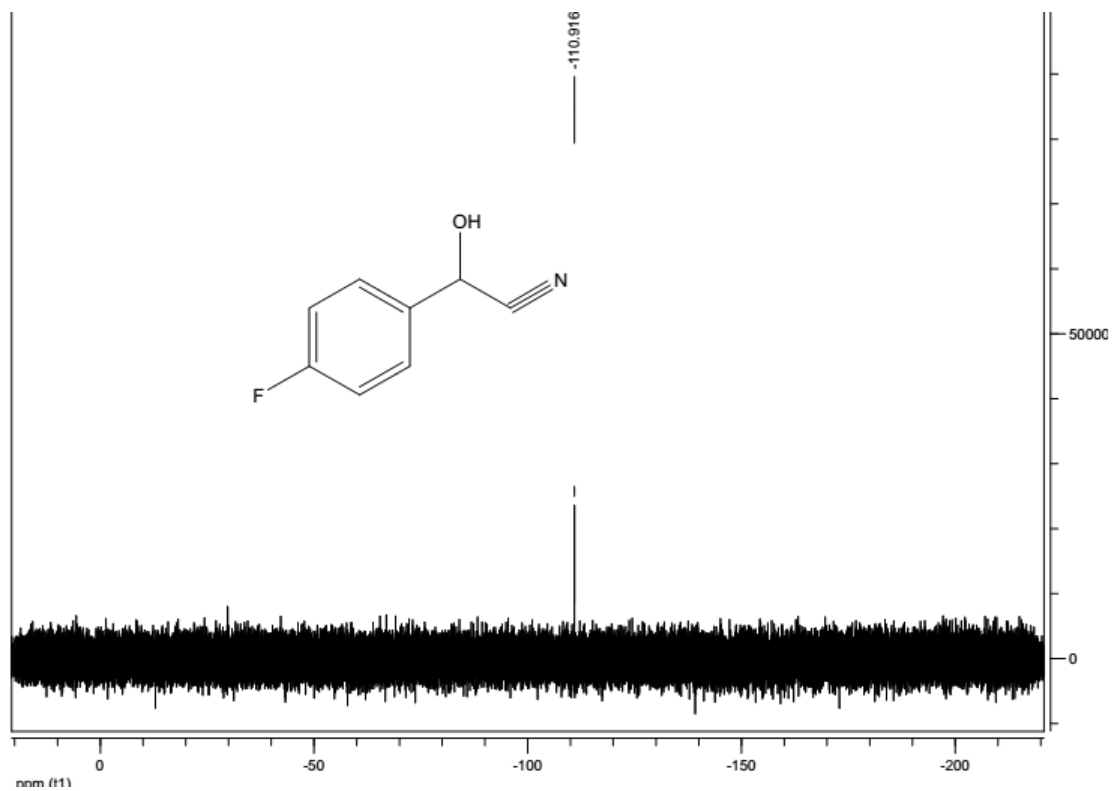
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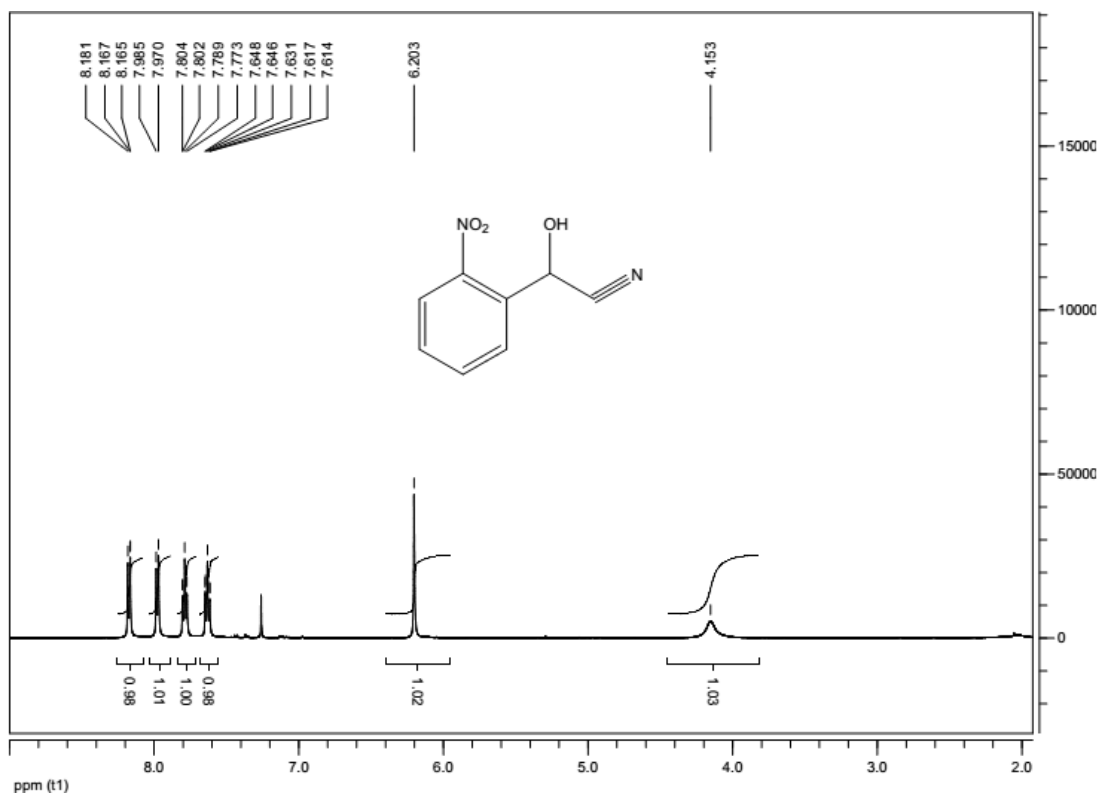


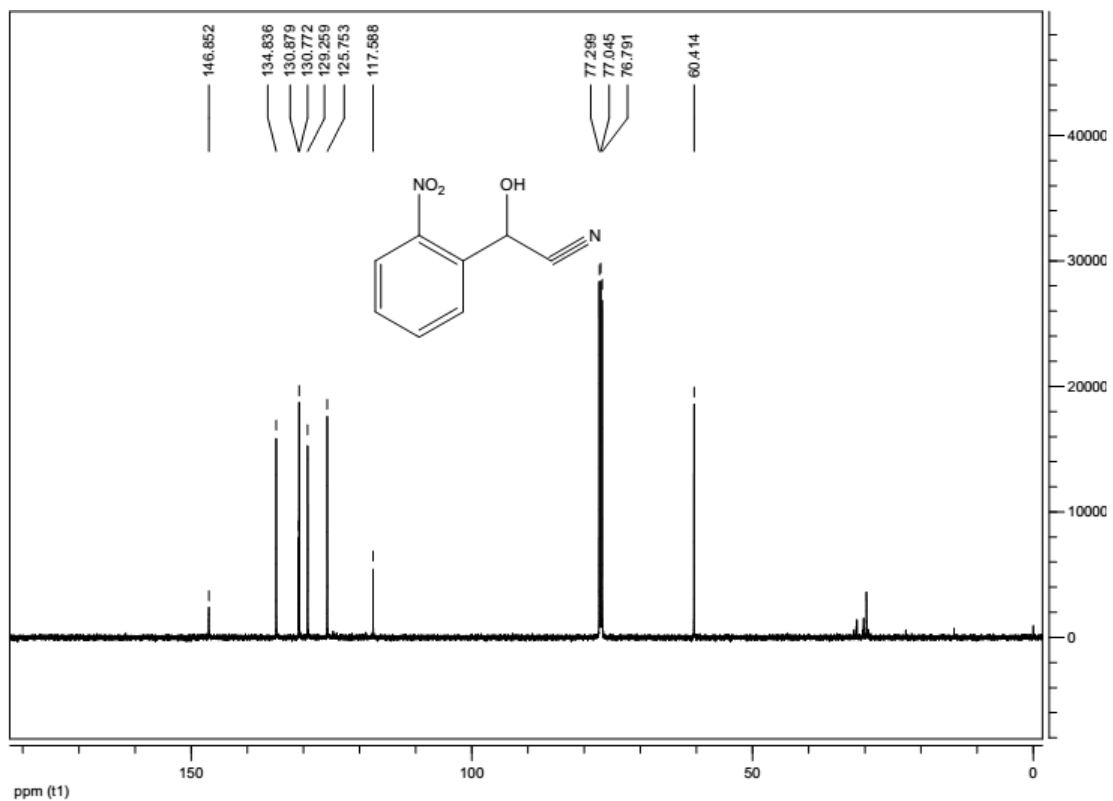
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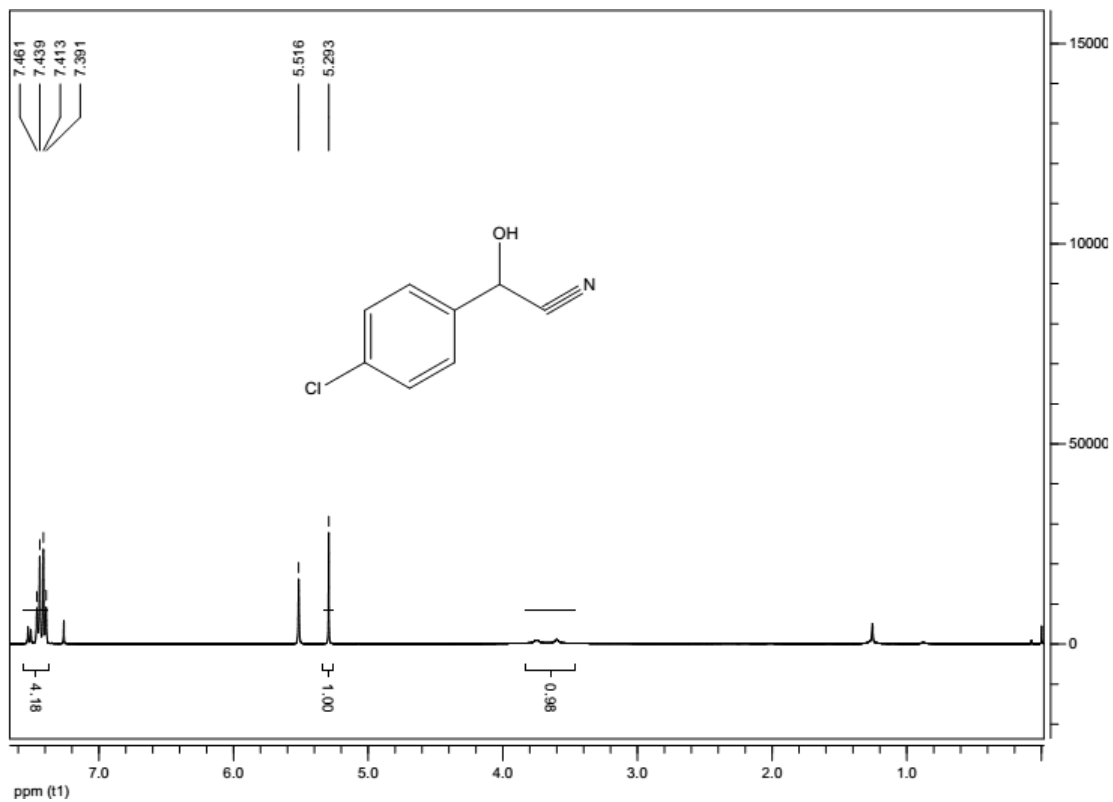


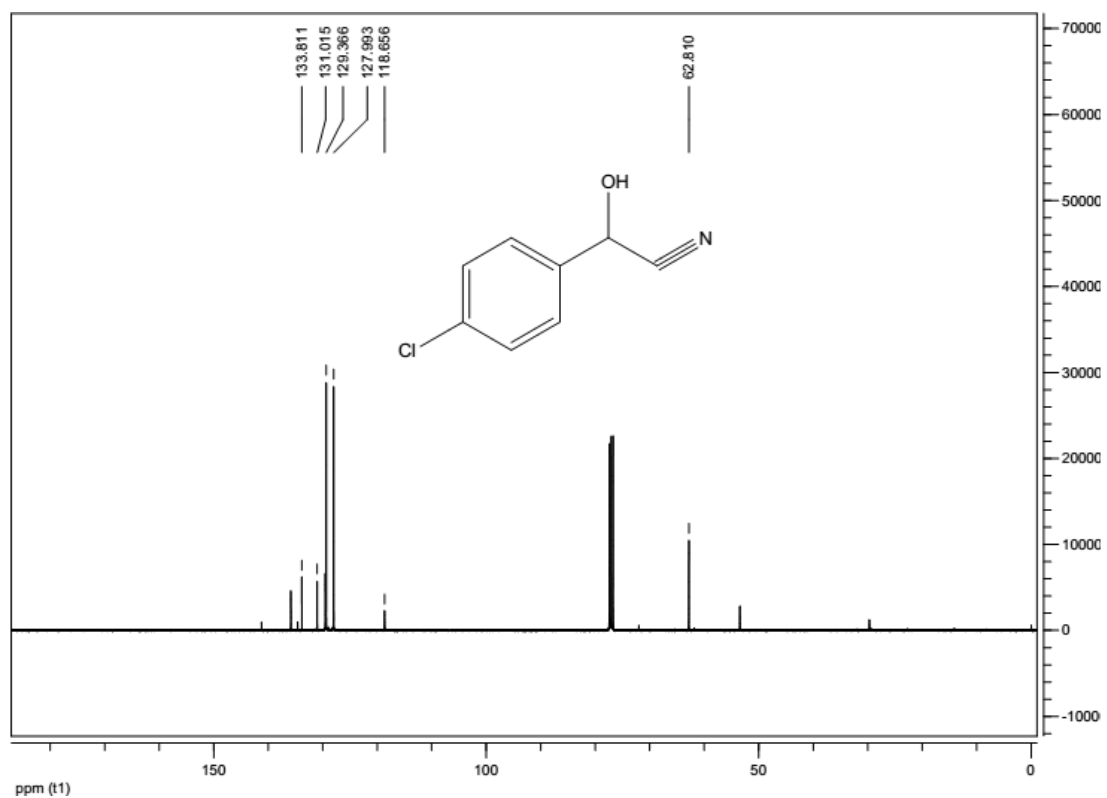
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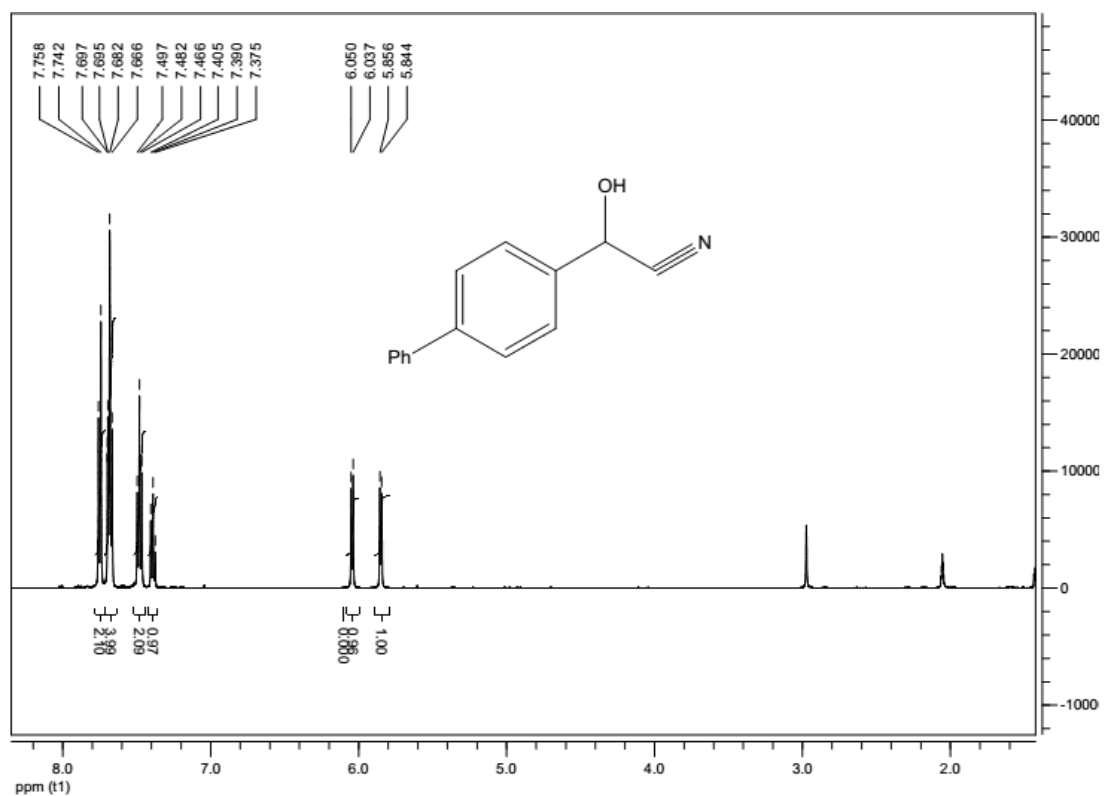


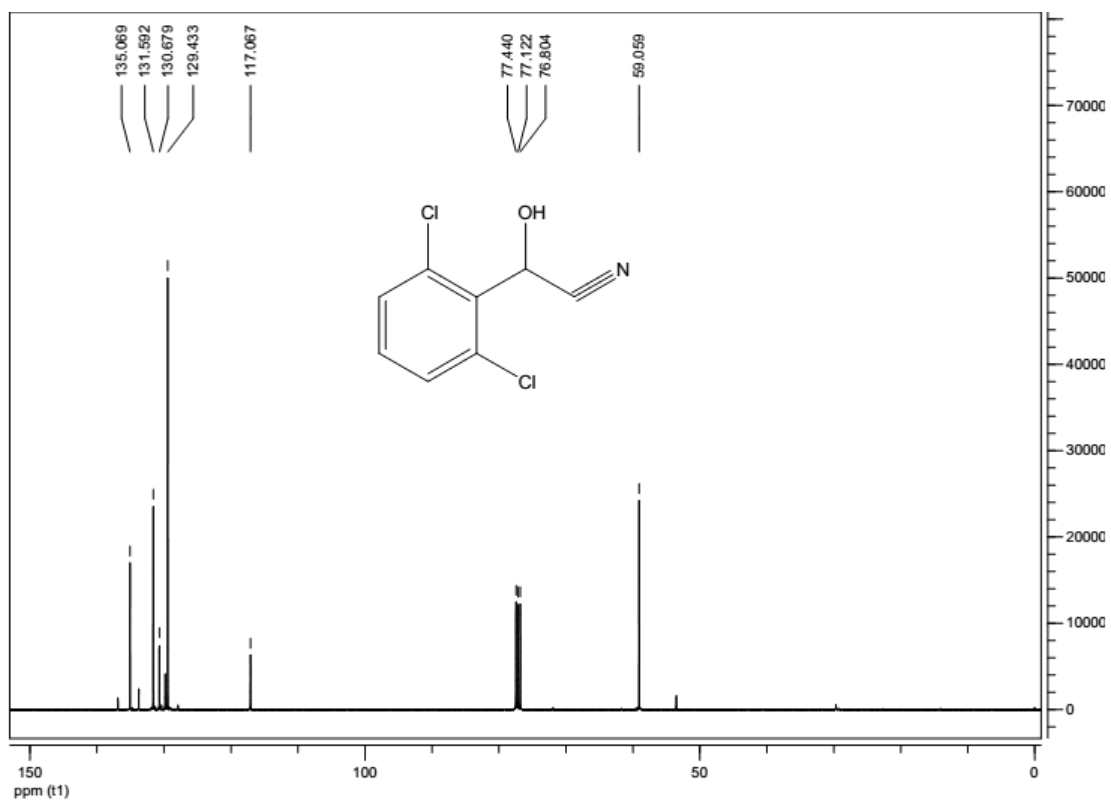
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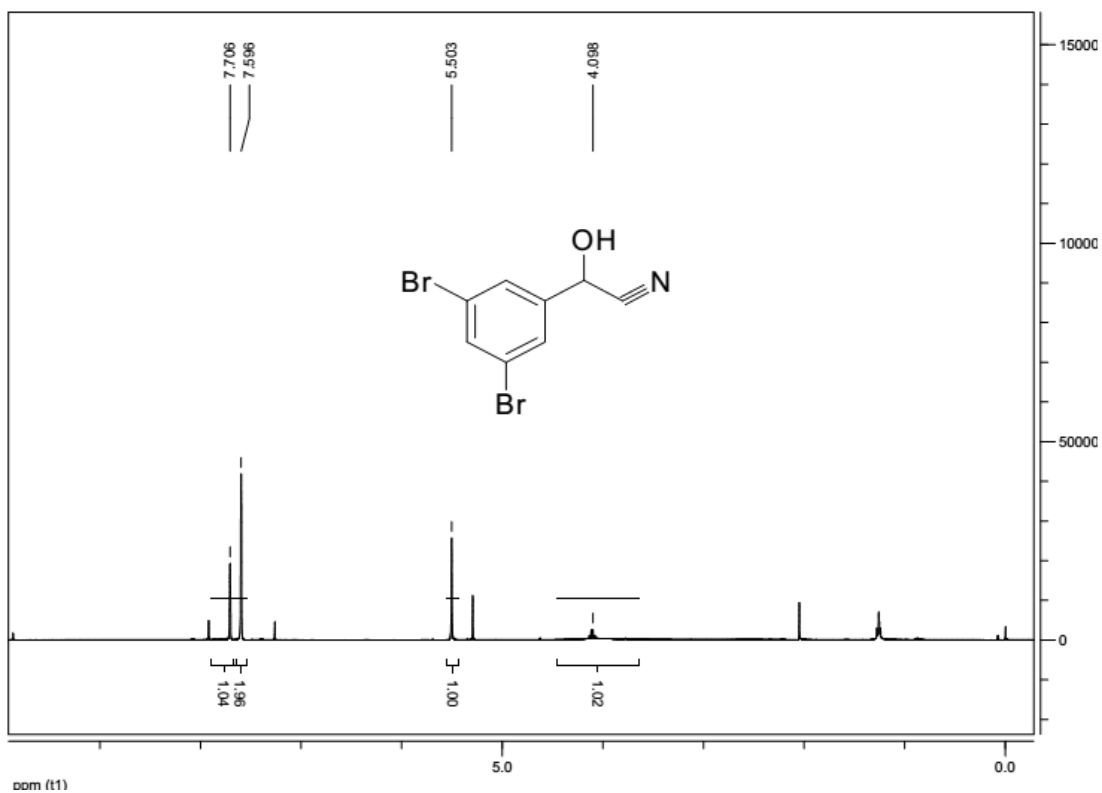


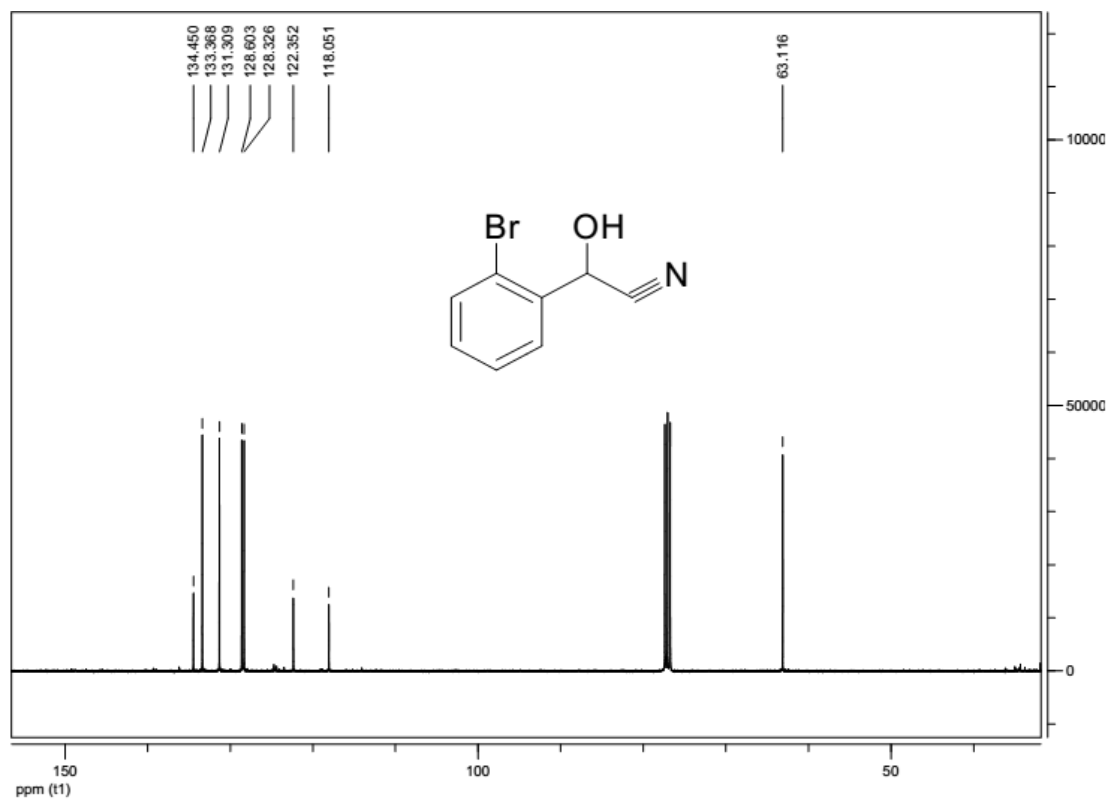
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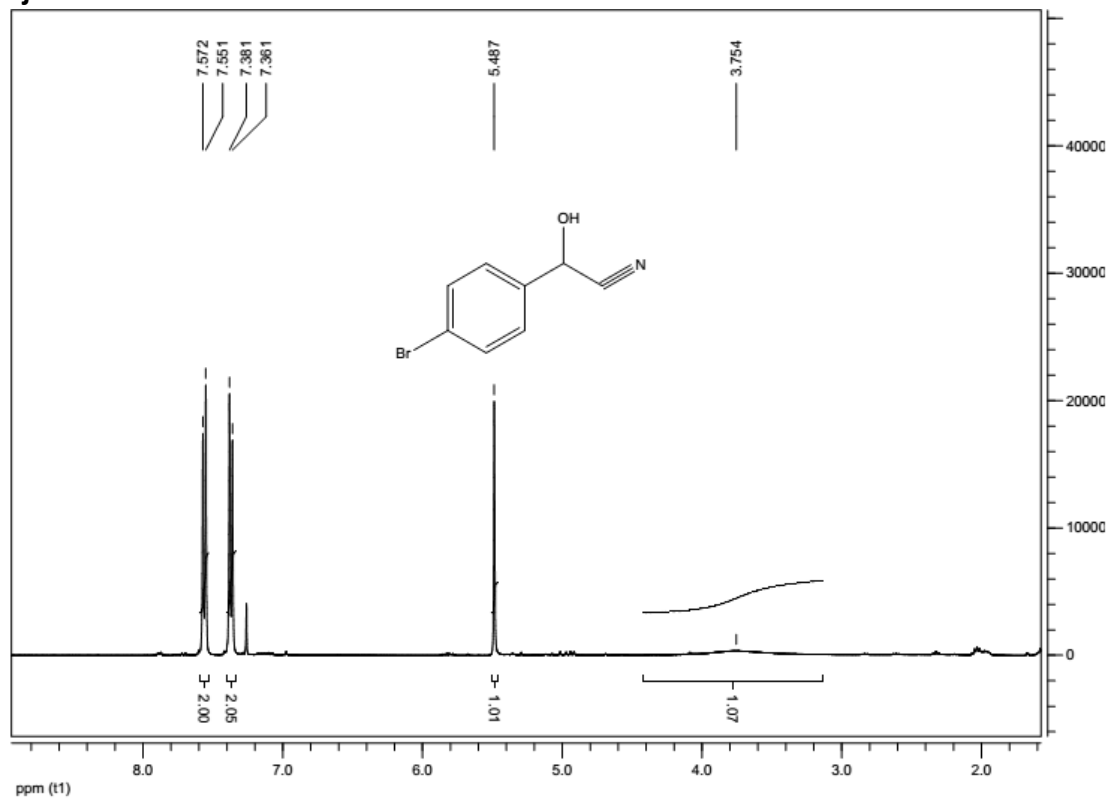


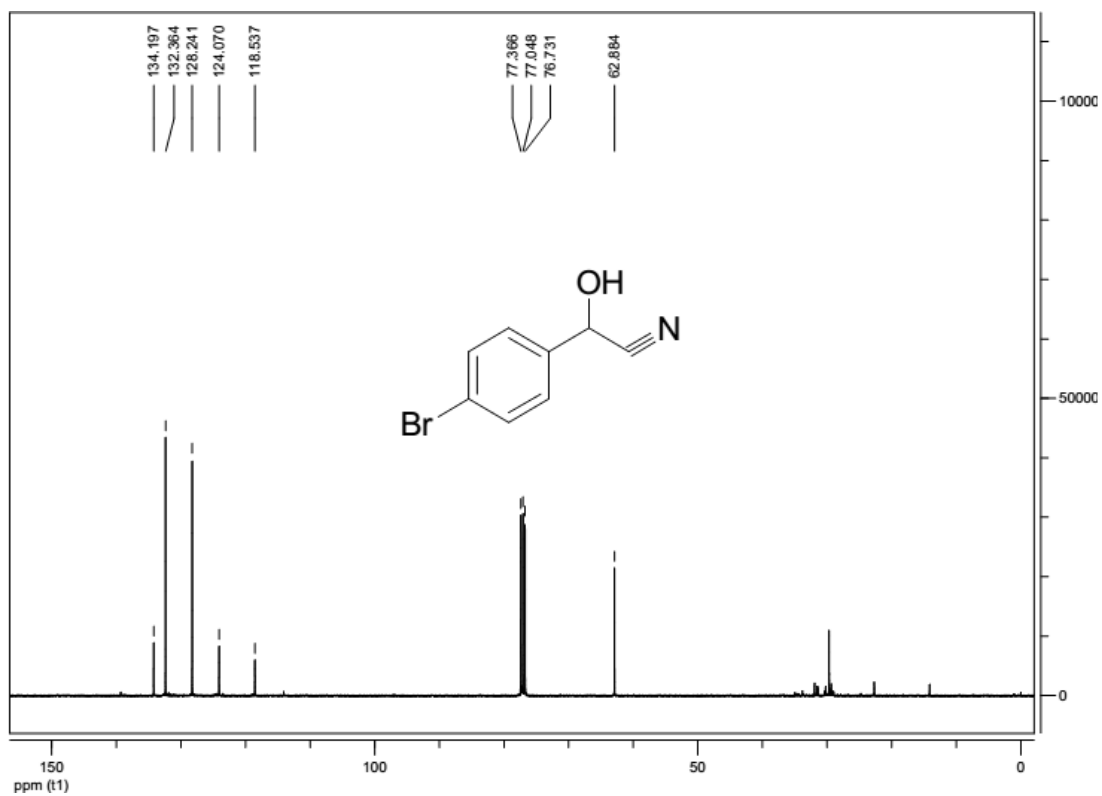
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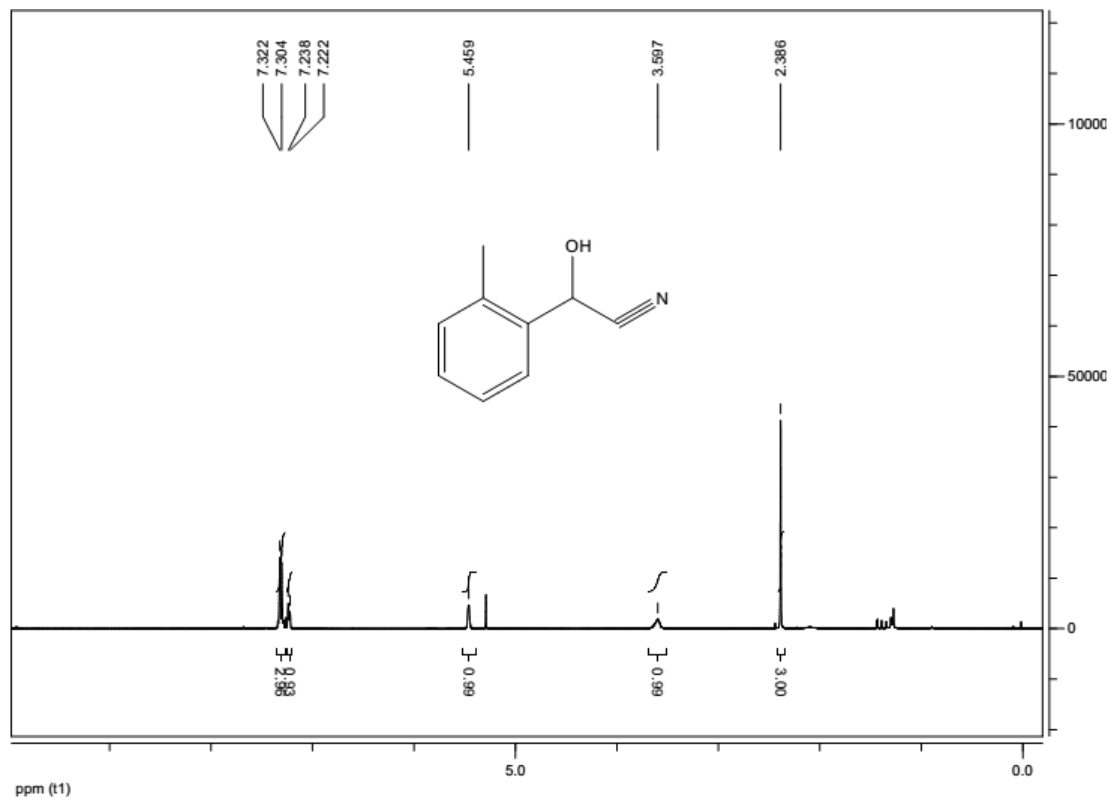


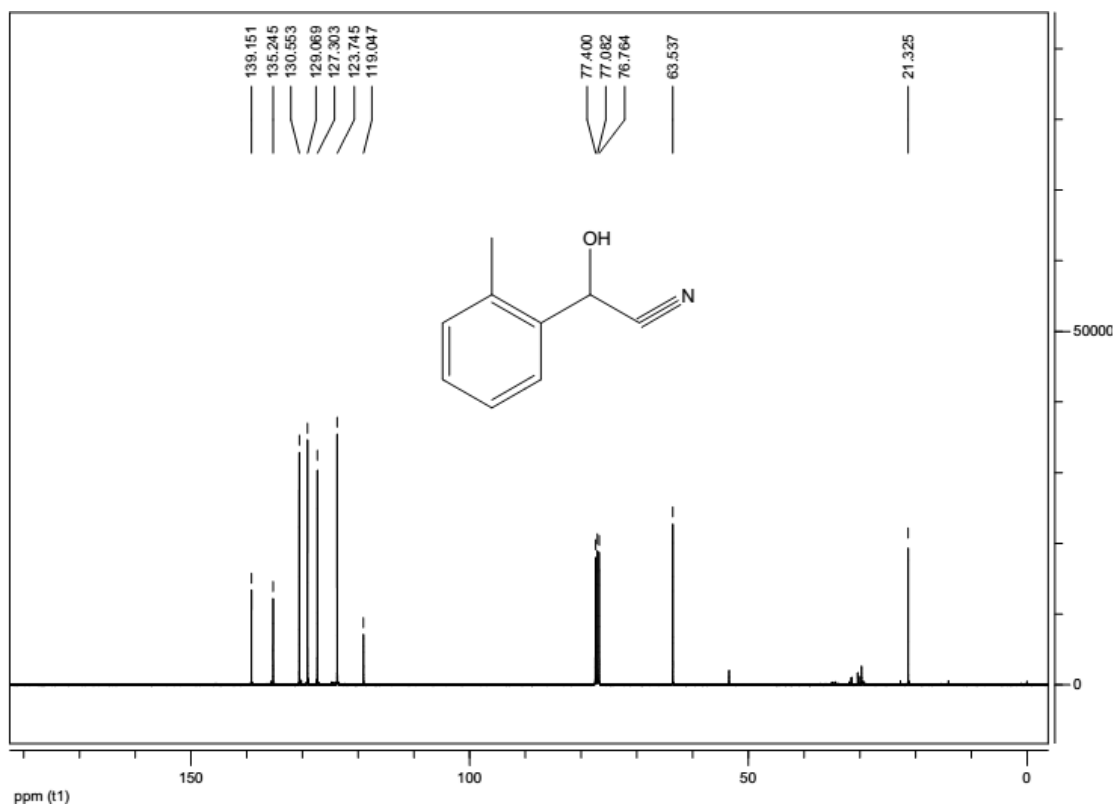
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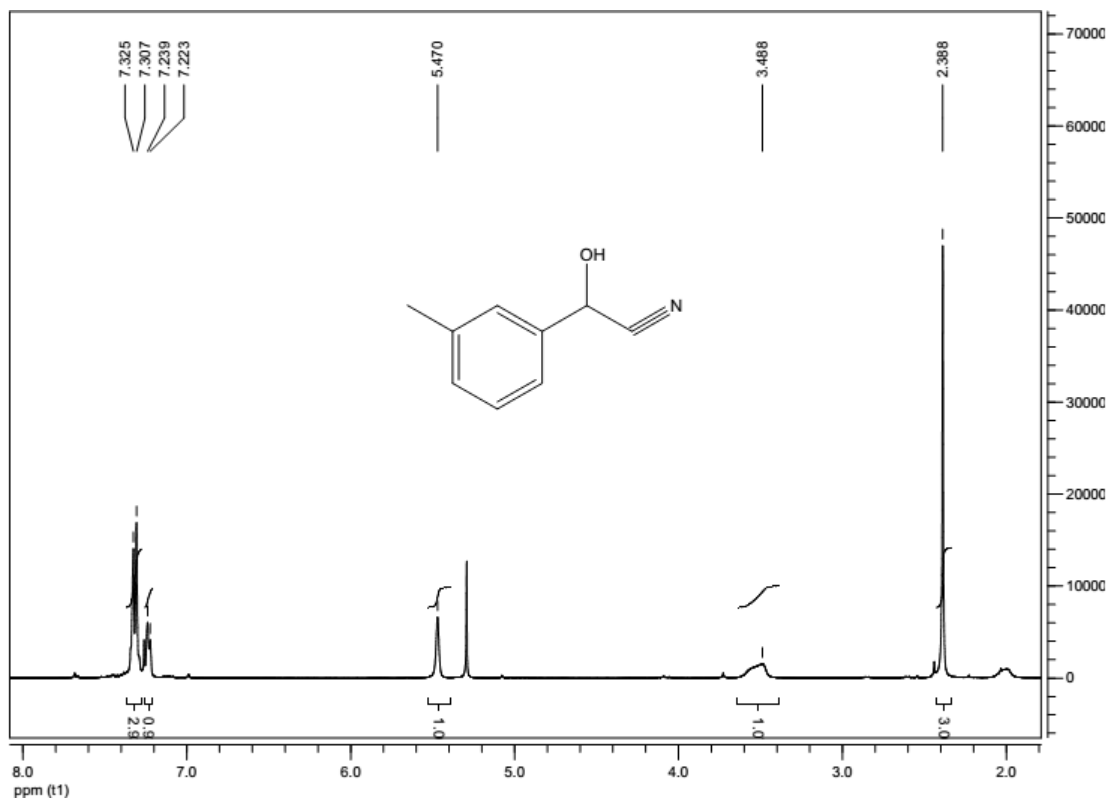


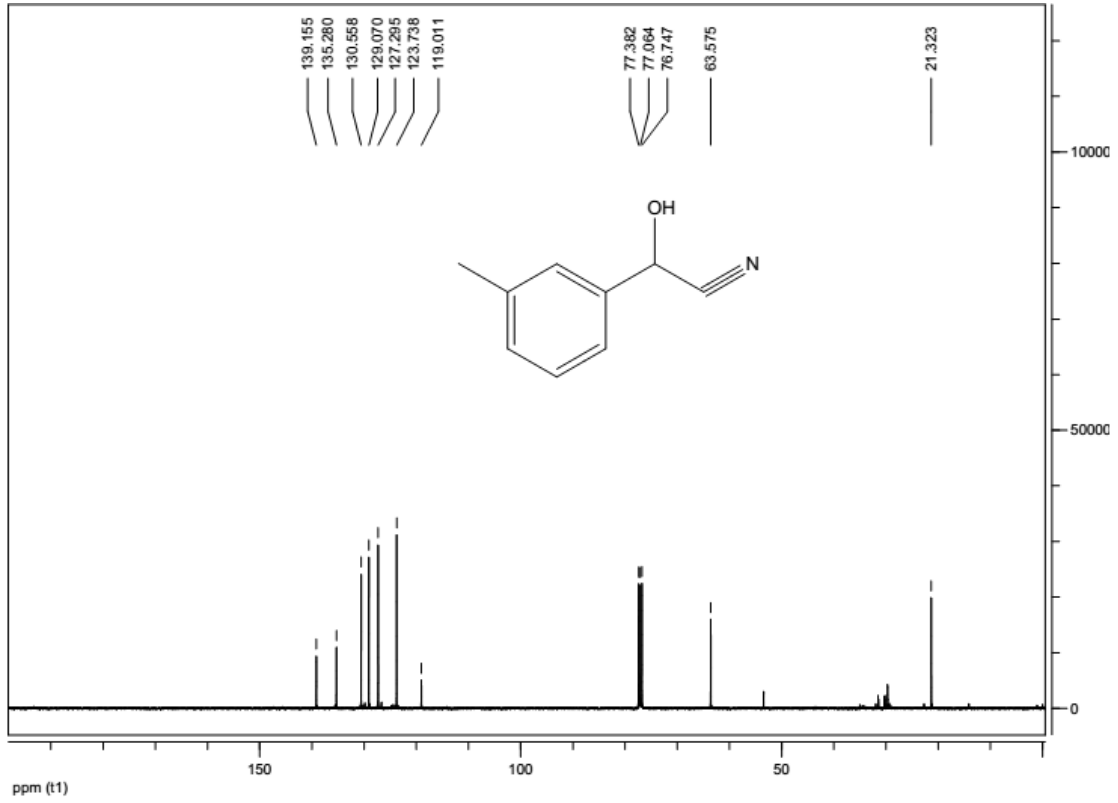
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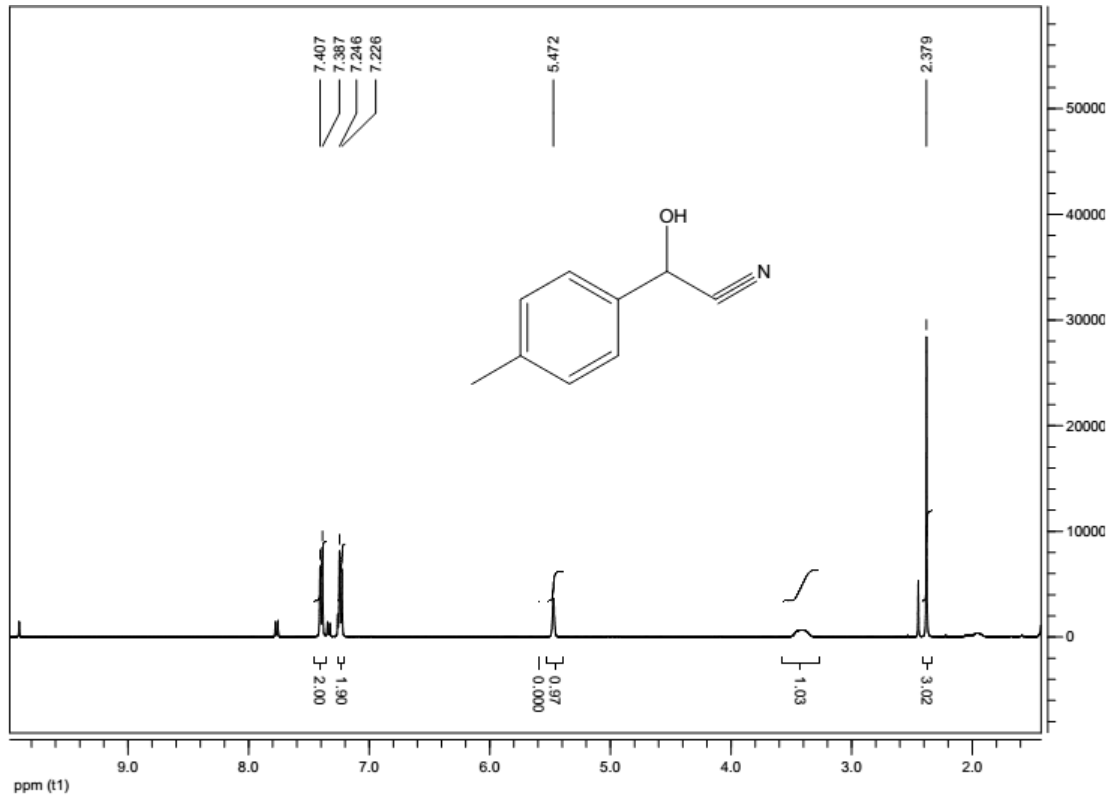


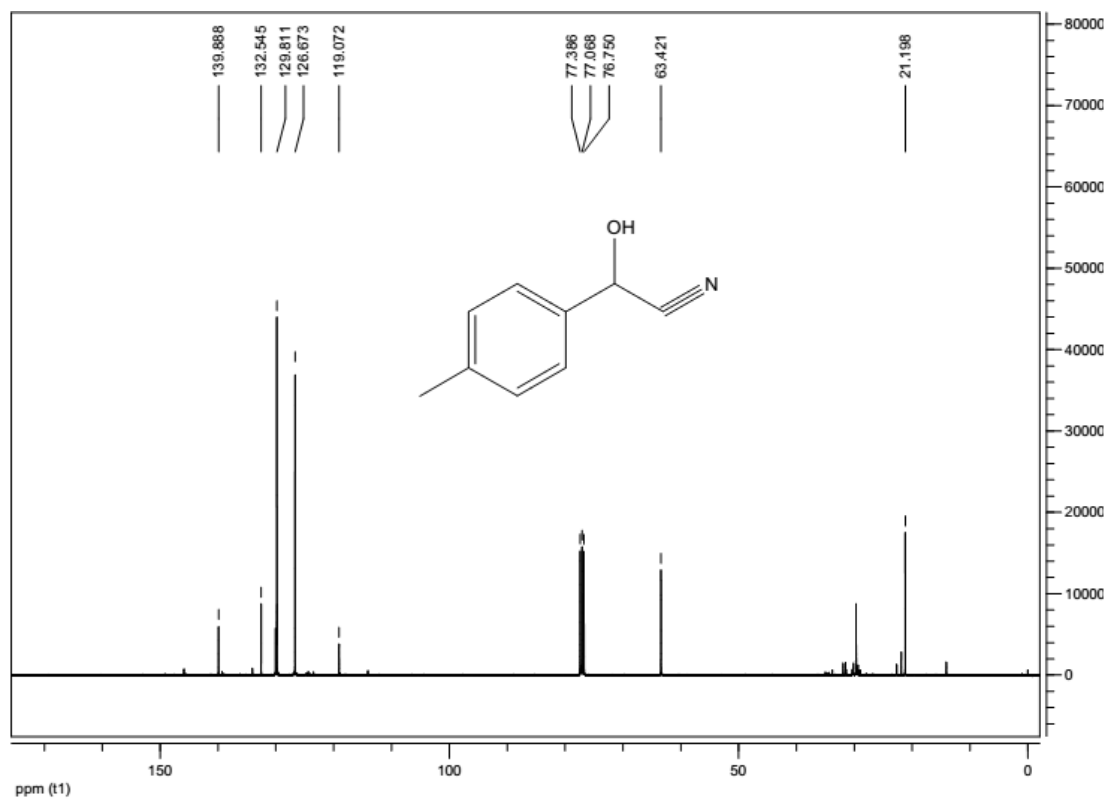
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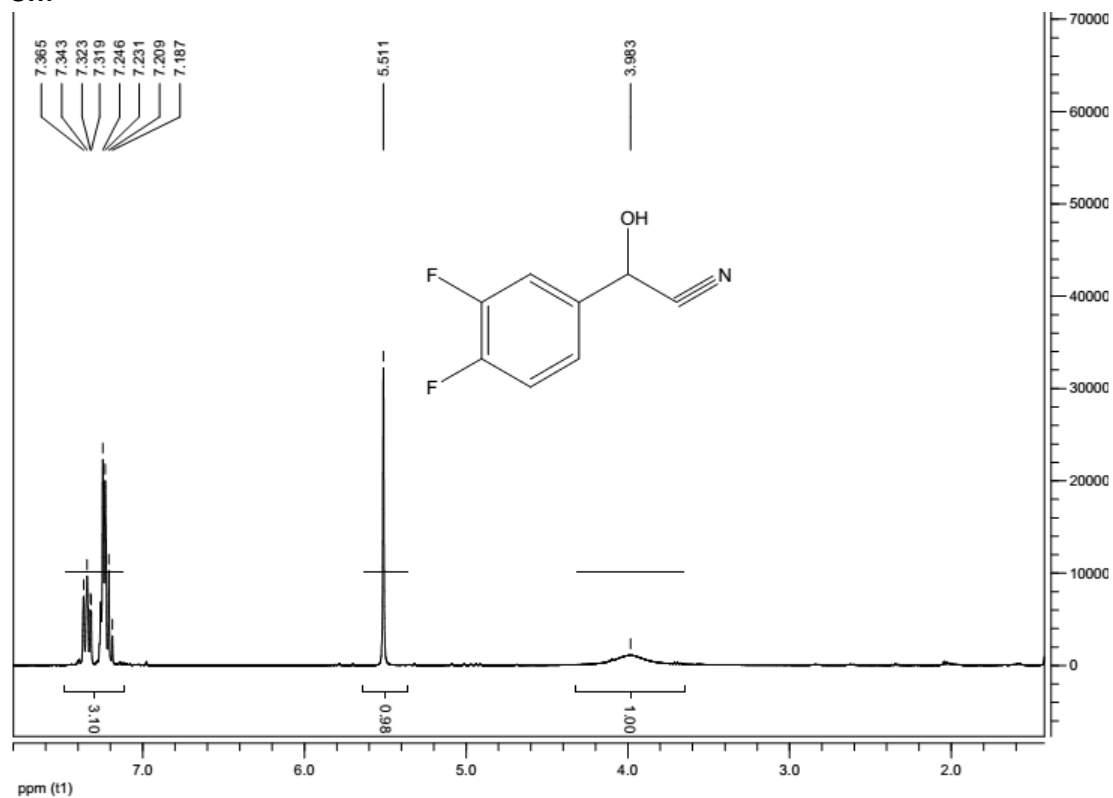


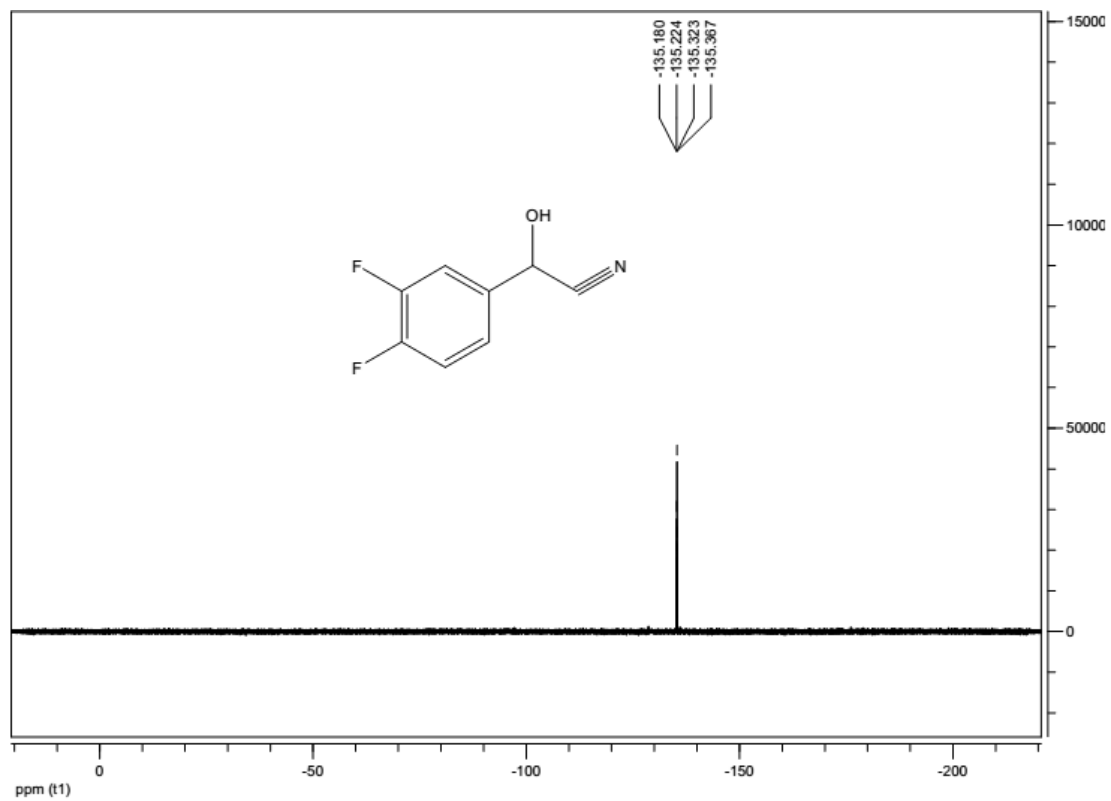
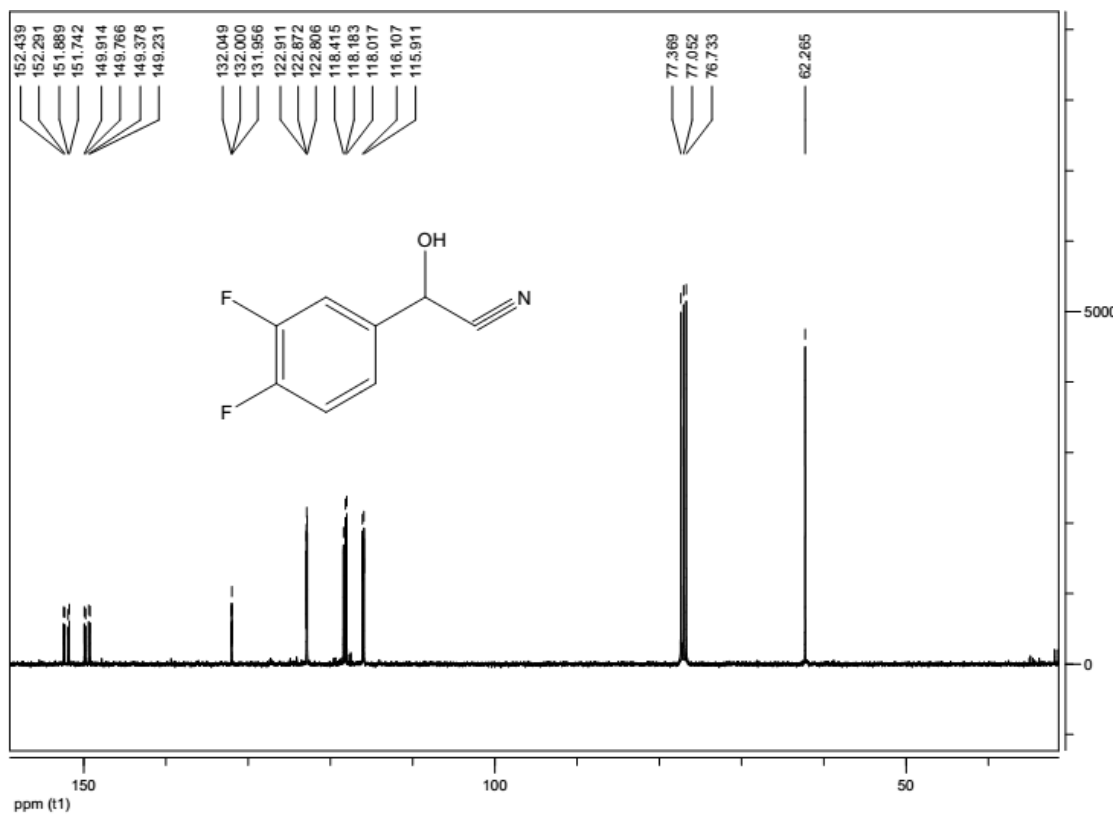
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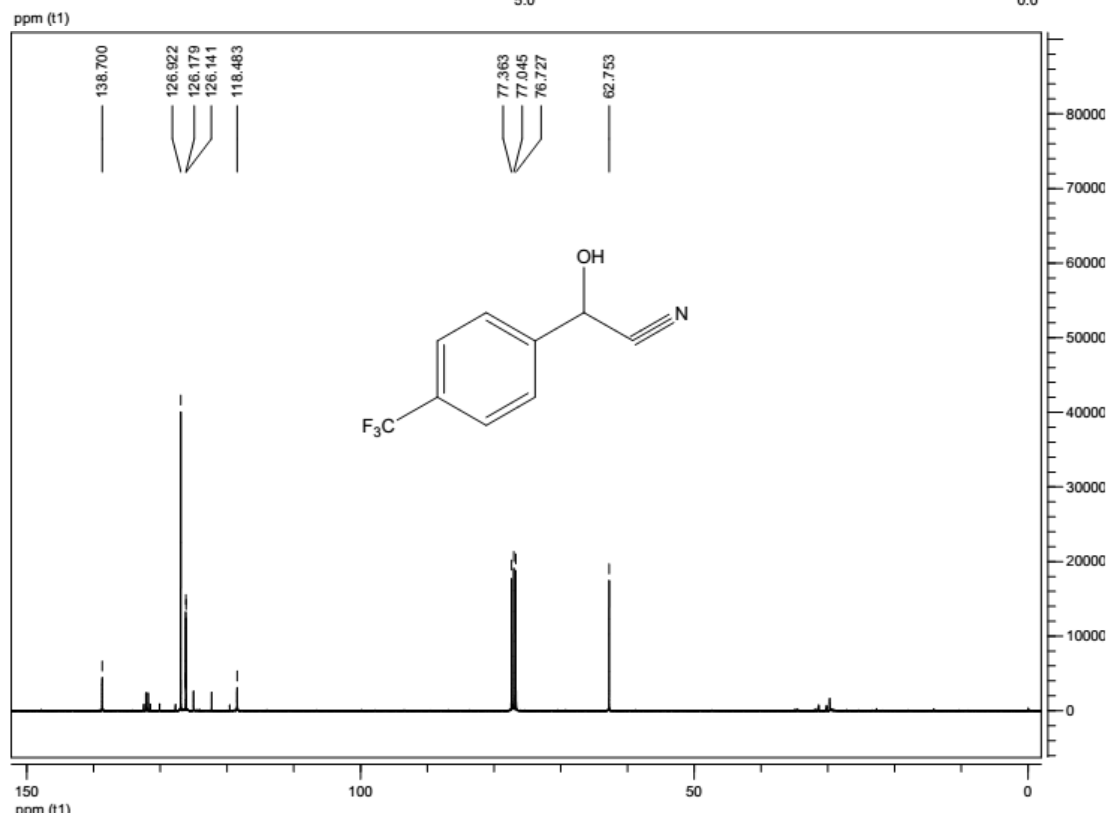
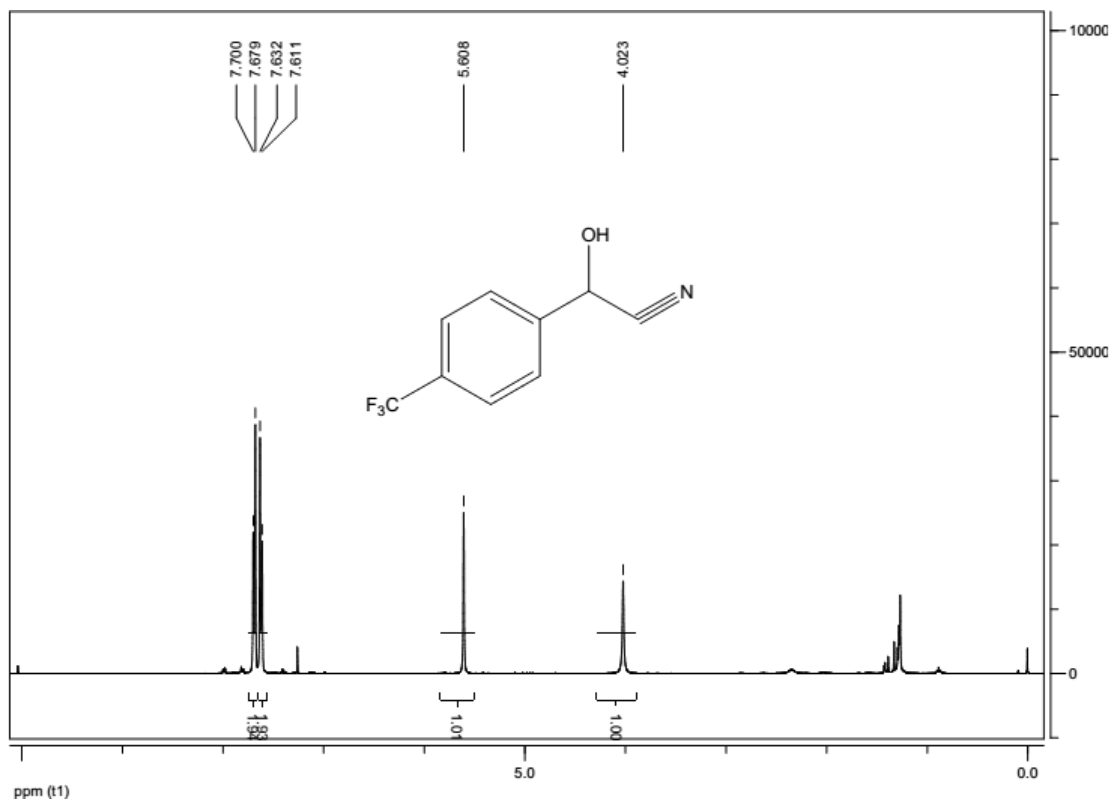


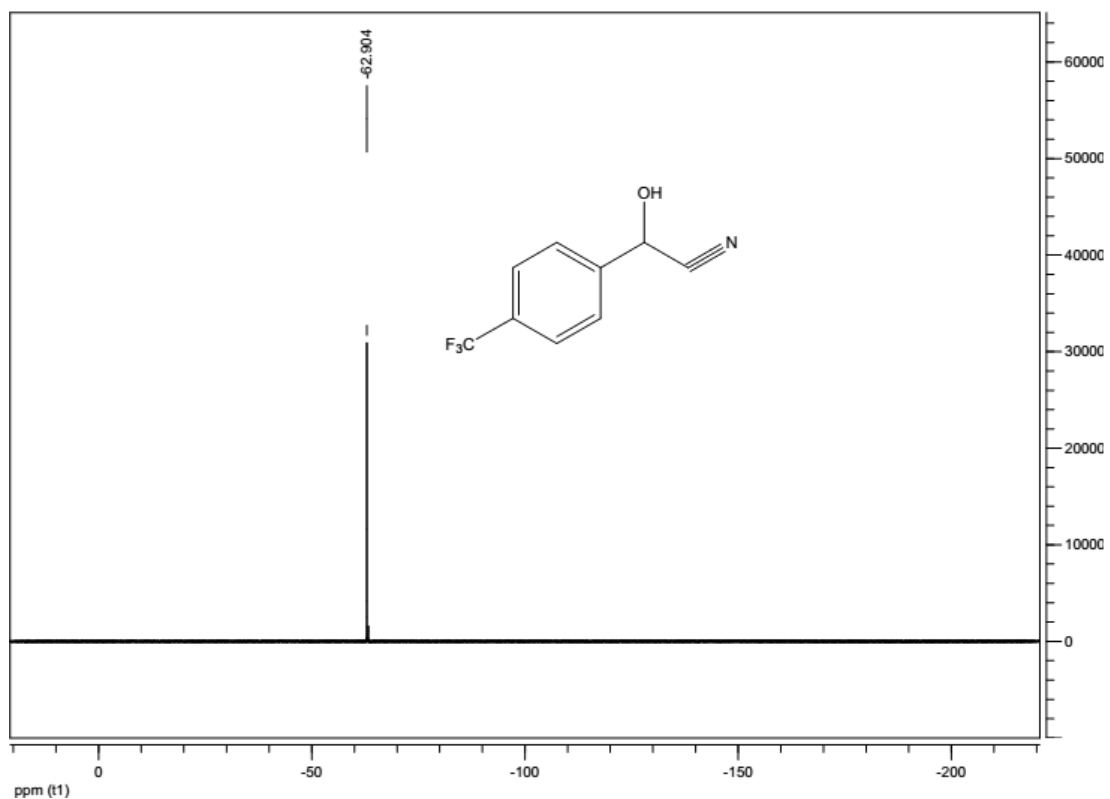
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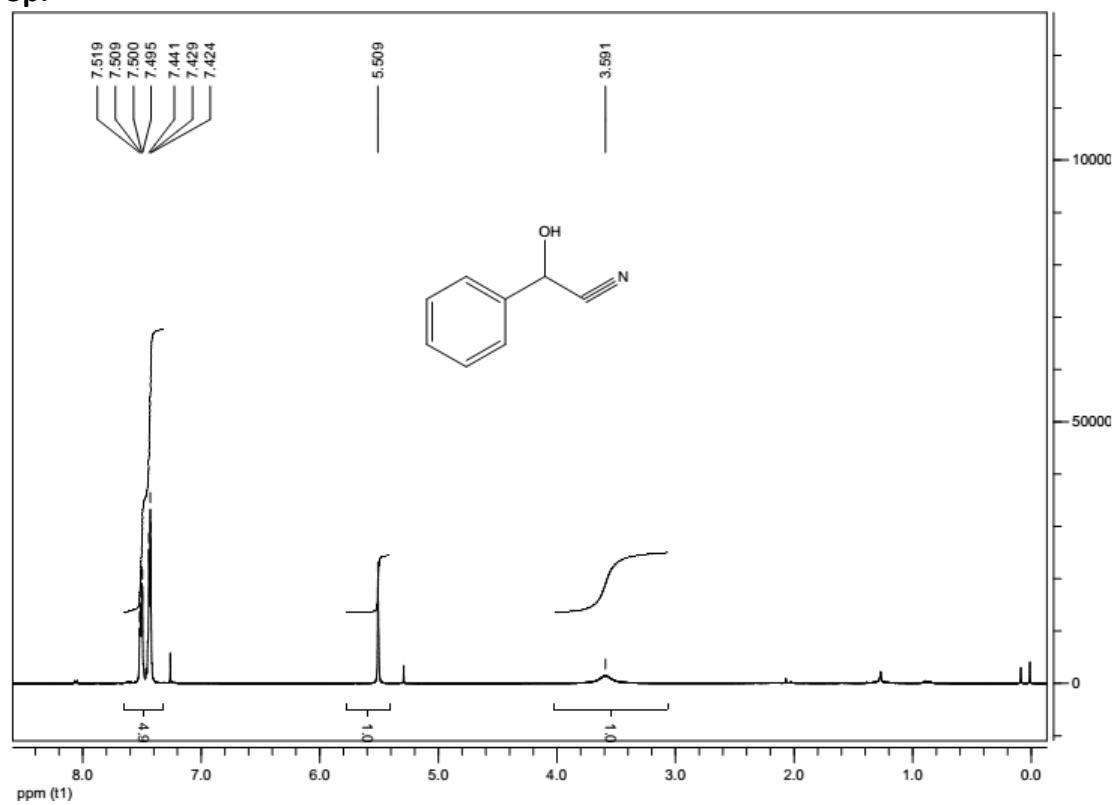


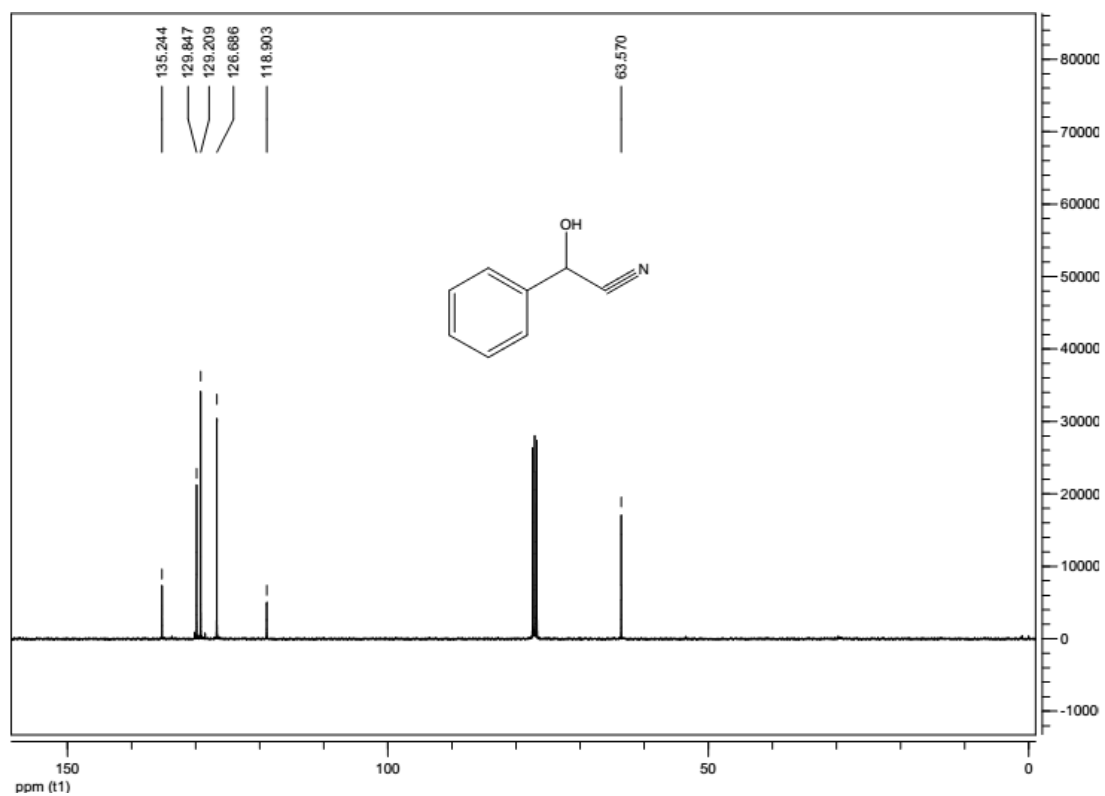
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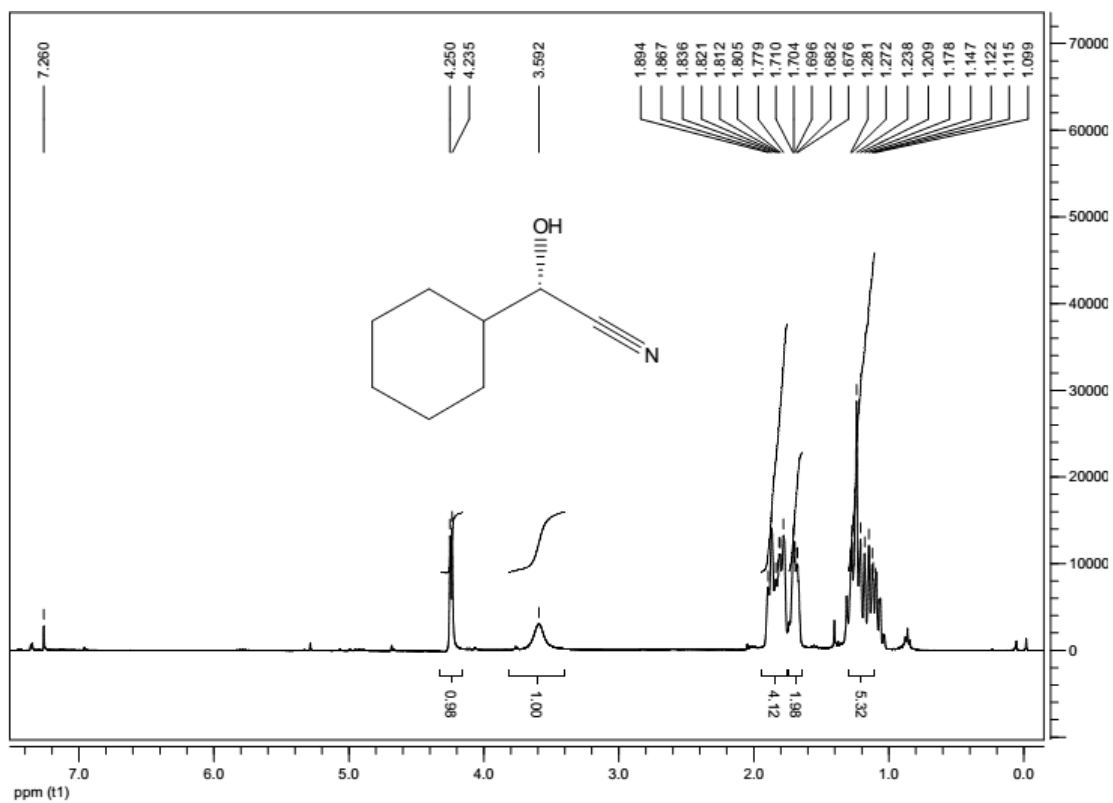


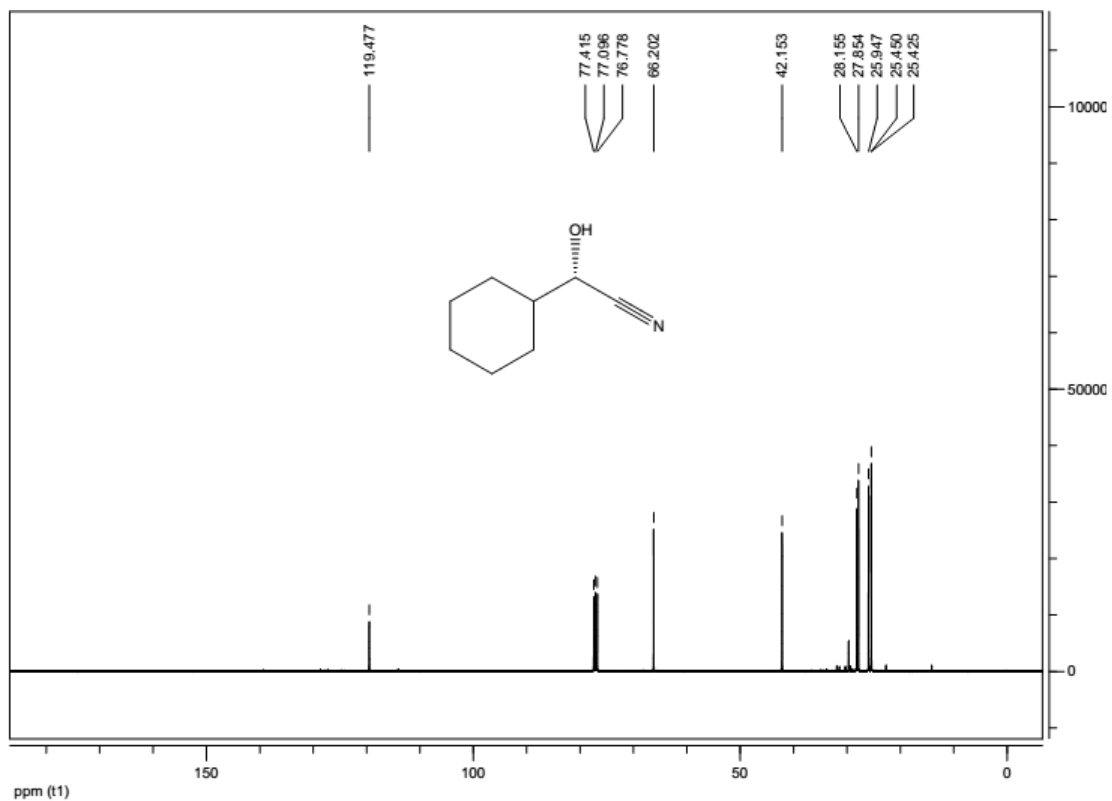
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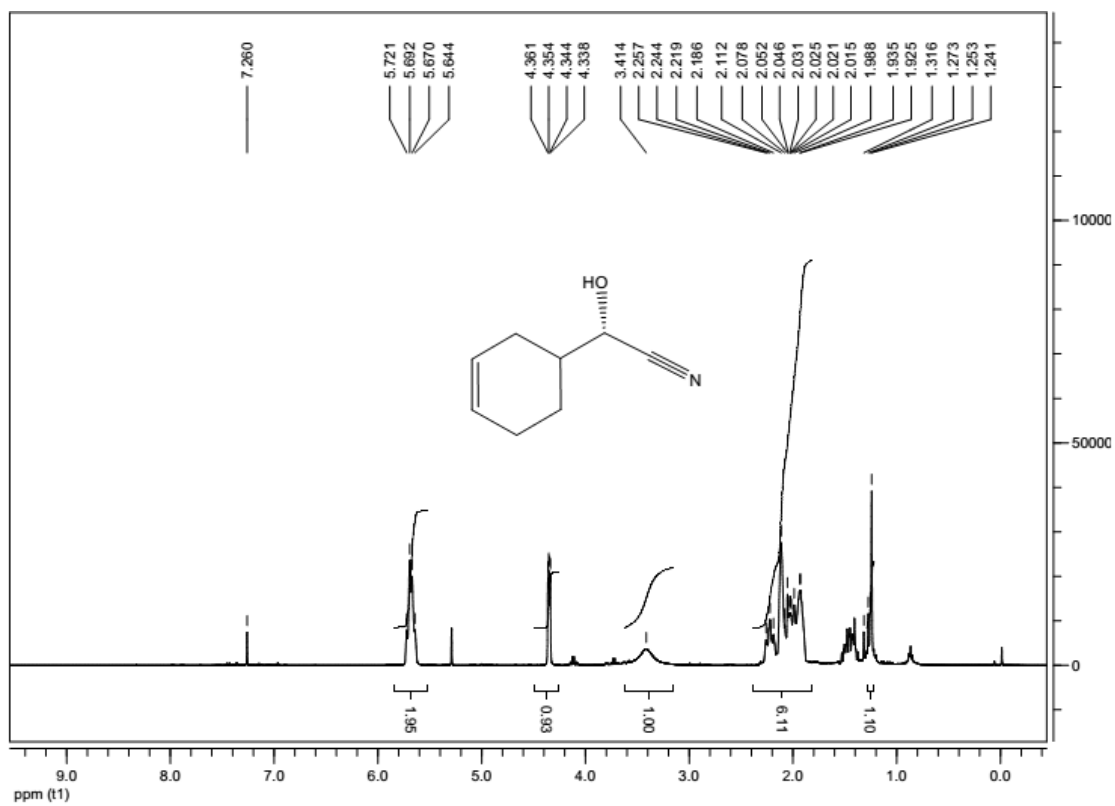


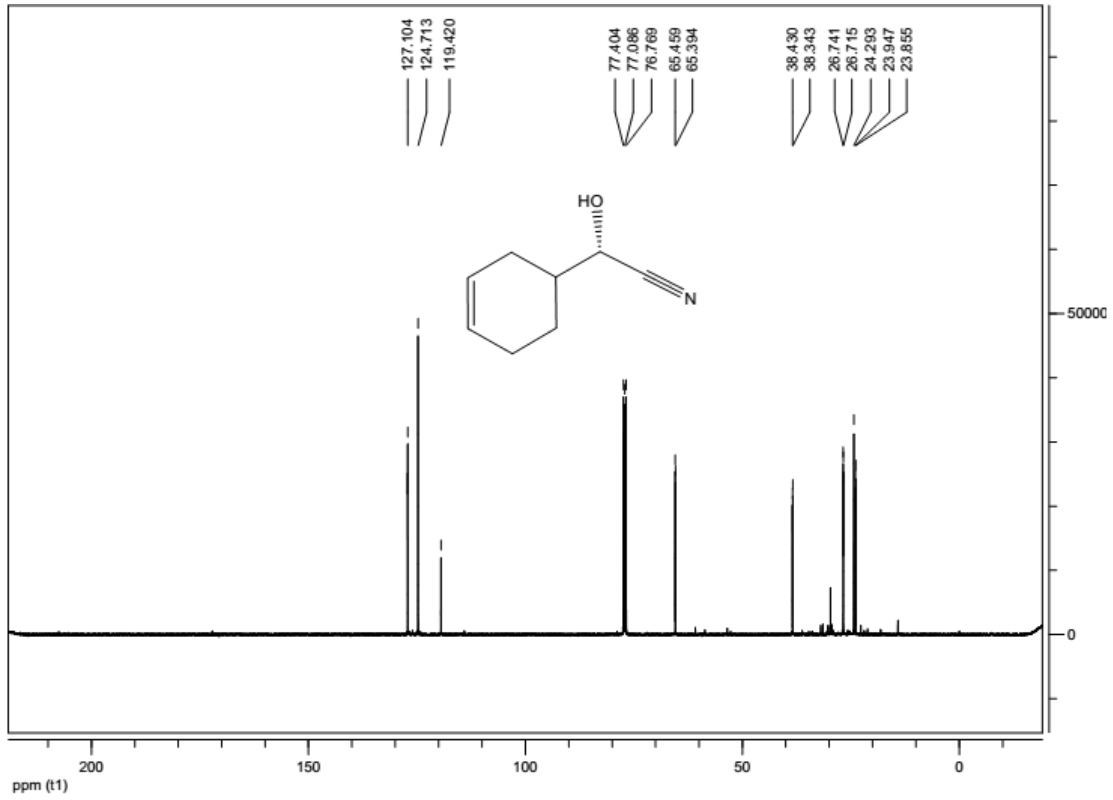
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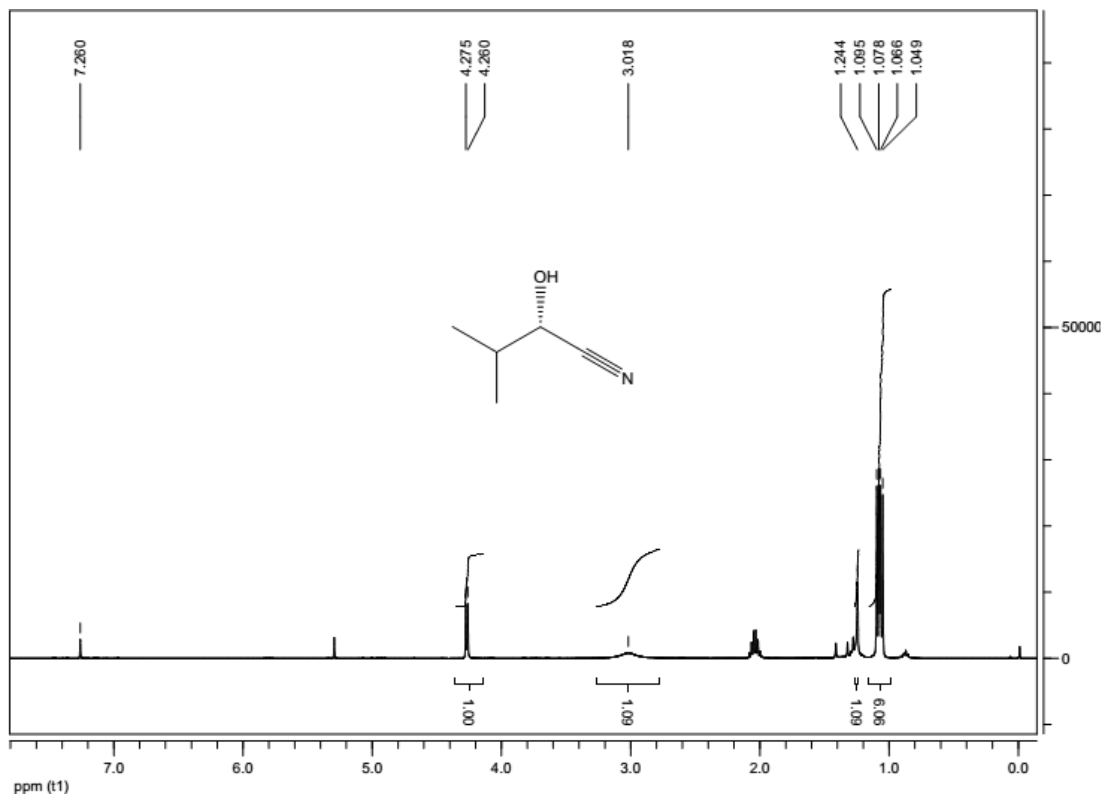


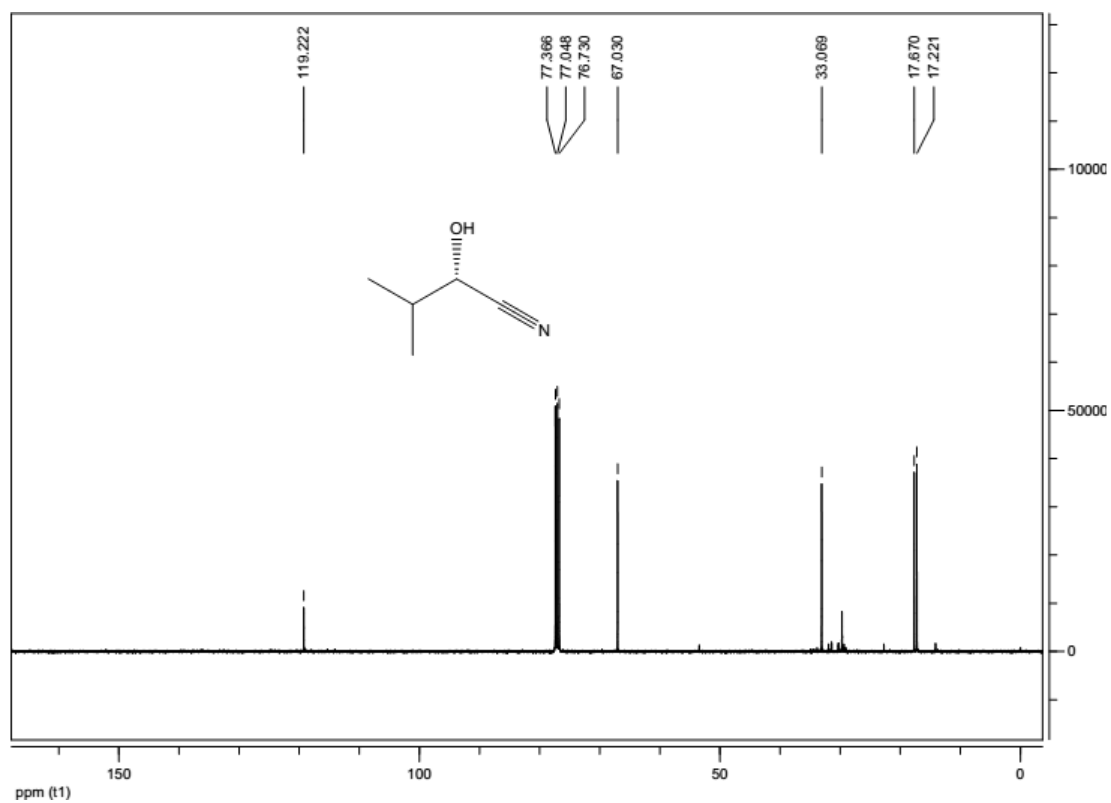
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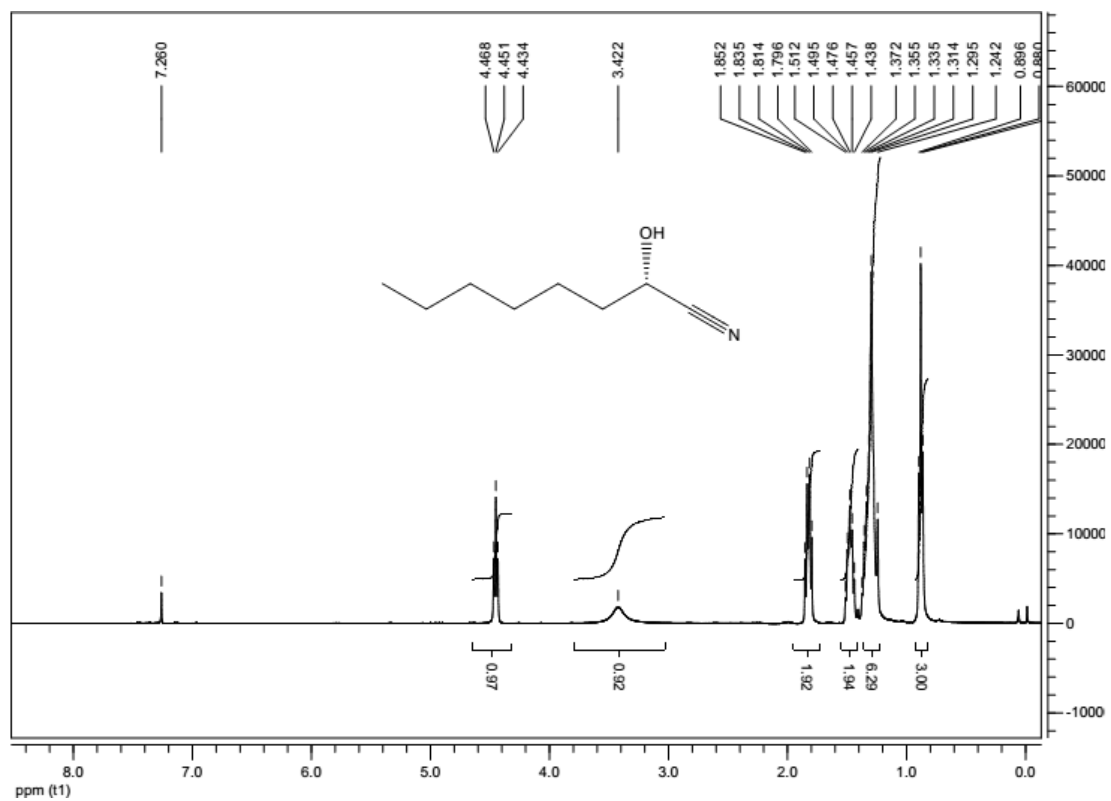


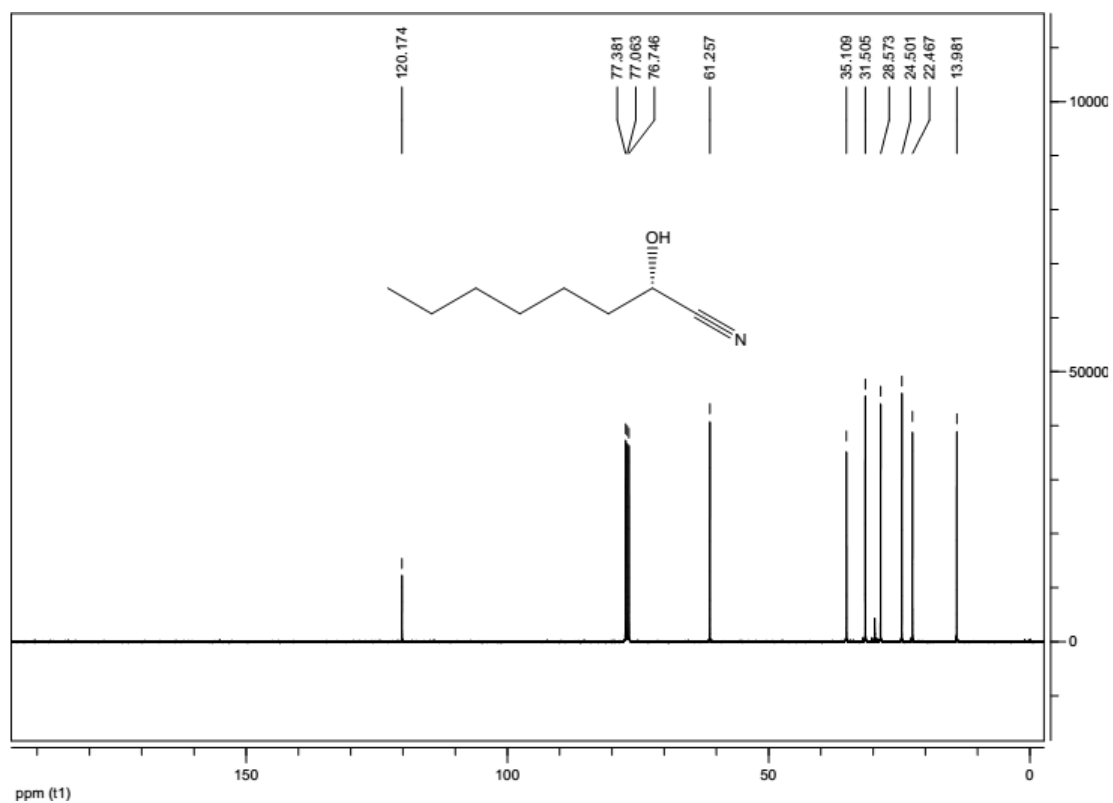
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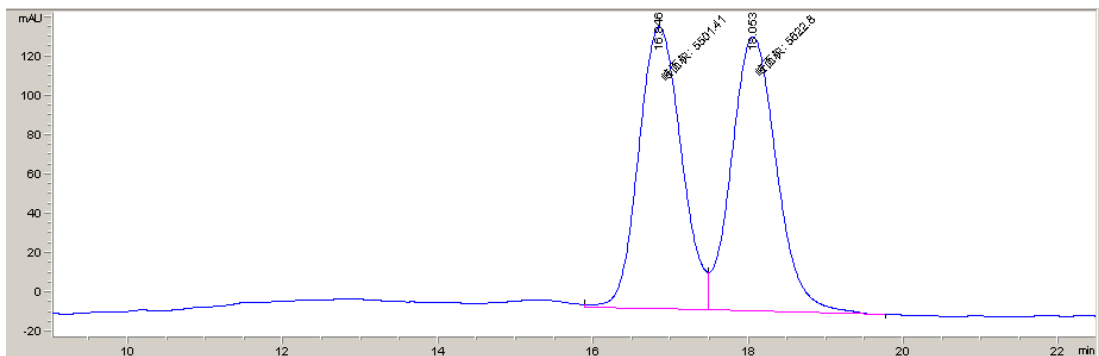
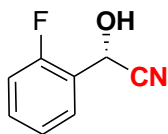


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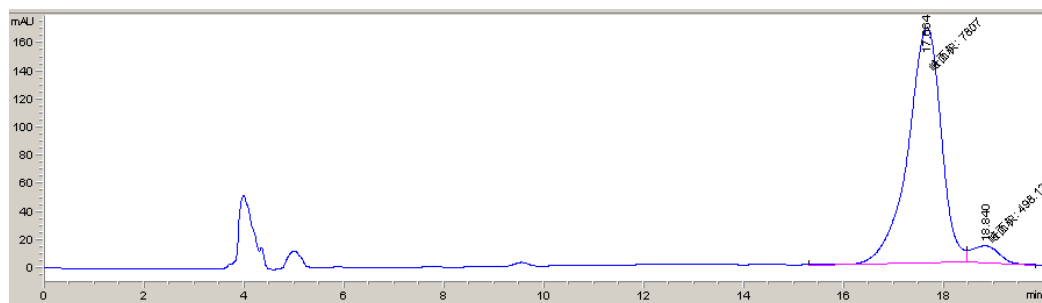




S-6. HPLC chromatograms of Compounds 3

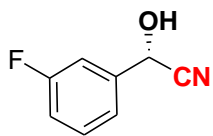


	Processed Chanel Descr	Time/min	Area	%Area	Height
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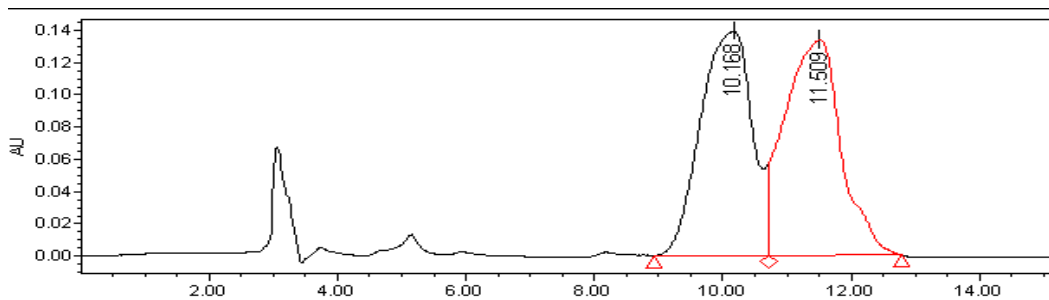


文件信息		#	时间	峰面积	峰高	峰宽	对称因子	峰面积 %
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文件路径	D:\EXAMPLES\WYL\BXF 2014-02-21 20:38:58\	2	18.84	498.1	12.2	0.6789	0.783	5.998
日期	21-Feb-14, 21:05:48							
样品	wyl-3							
样品信息								
条形码								
操作者	ZJ							
方法	BXF-SXYH.M							
仪器时间	10.000 min							

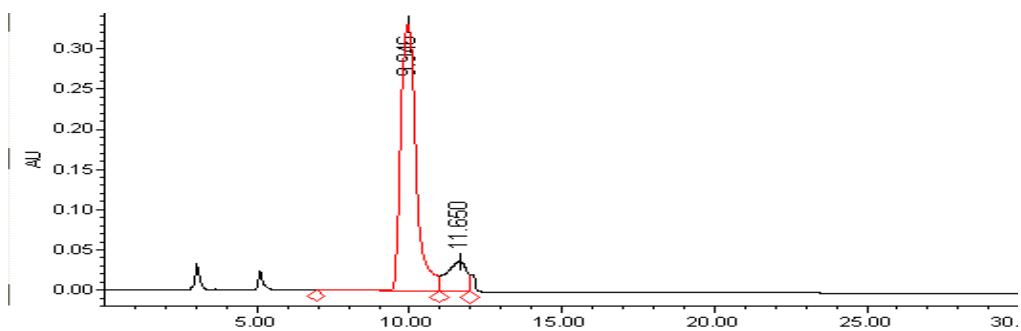
	Processed Chanel Descr	Time/min	Area	%Area	Height
1	210	17.664	7807	94.002	167.5
2	210	18.84	498.1	5.998	12.2



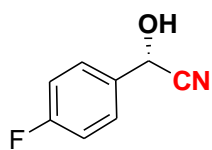
3b
81% yield
75% ee



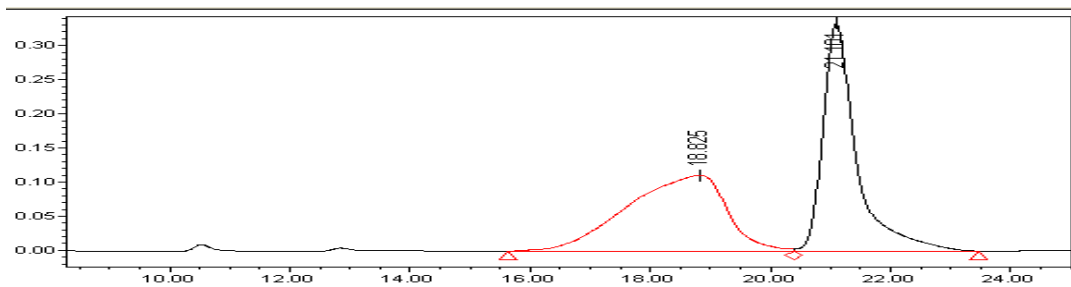
	Processed Chanel Descr	Time/min	Area	%Area	Height
1	210	10.169	7817667	49.07	138869
2	210	11.509	8115473	50.93	133215



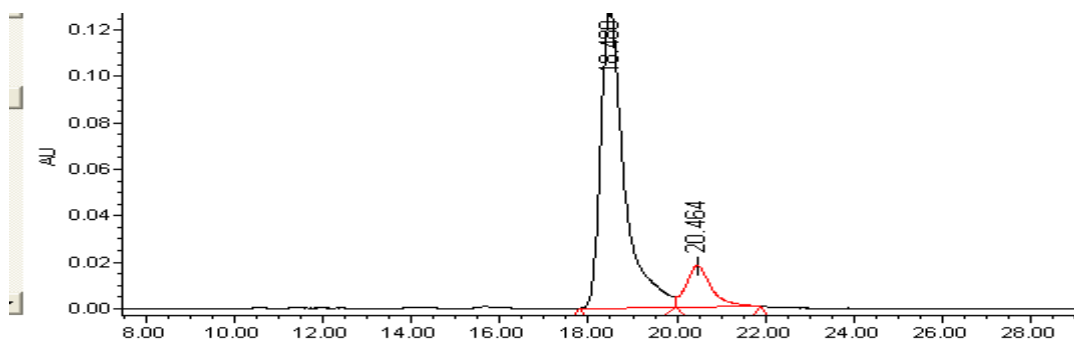
	Processed Chanel Descr	Time/min	Area	%Area	Height
1	210	9.946	11692001	86.32	138869
2	210	11.650	1698456	12.68	133215



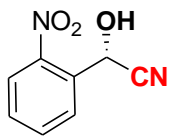
3c
98% yield
74% ee



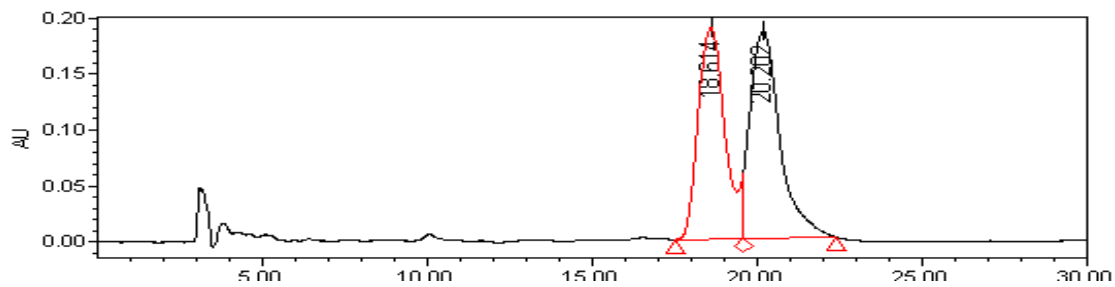
	Processed Chanel Descr	Time/min	Area	%Area	Height
1	210	18.825	12817440	49.66	111092
2	210	21.101	12994209	50.34	333222



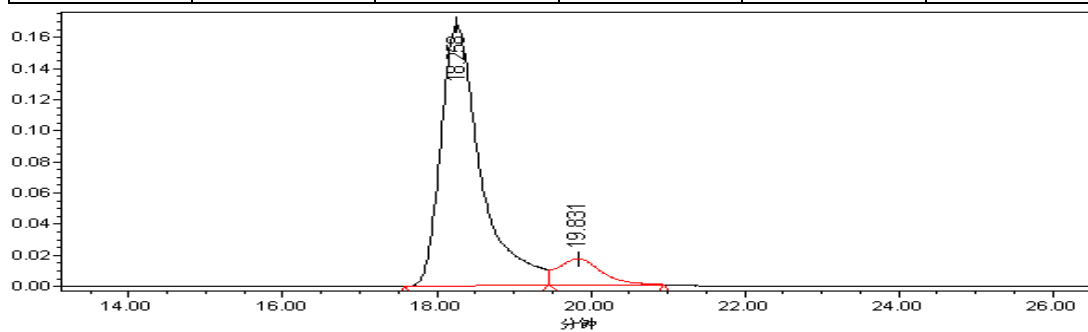
	Processed Chanel Descr	Time/min	Area	%Area	Height
1	210	18.488	4801350	87.05	183618
2	210	20.464	714078	12.95	25252



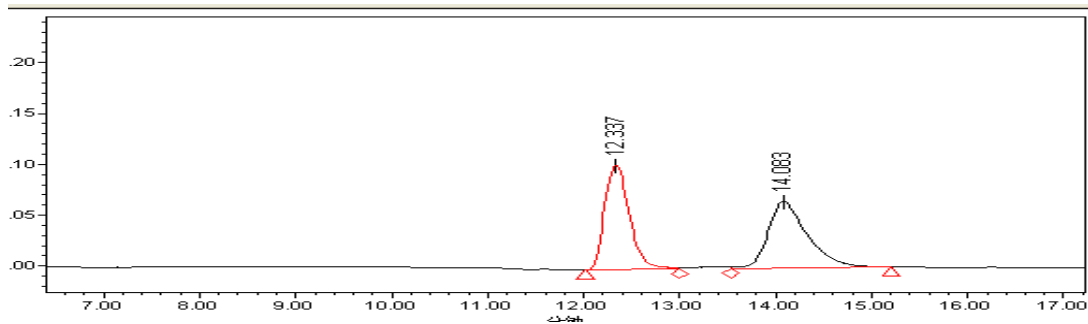
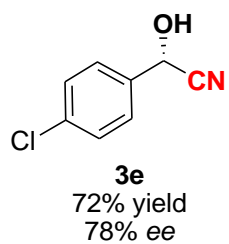
3d
96% yield
80% ee



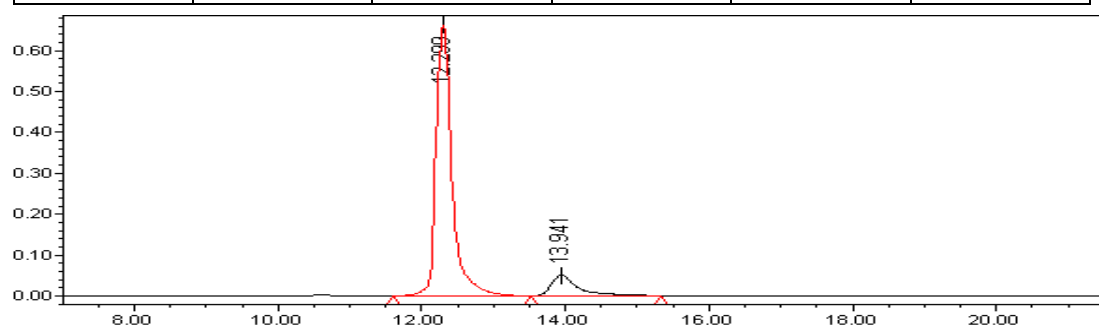
	Processed Chanel Descr	RT	Area	%Area	Height
1	210	18.614	10711425	48.10	188181
2	210	20.202	11559311	51.90	184655



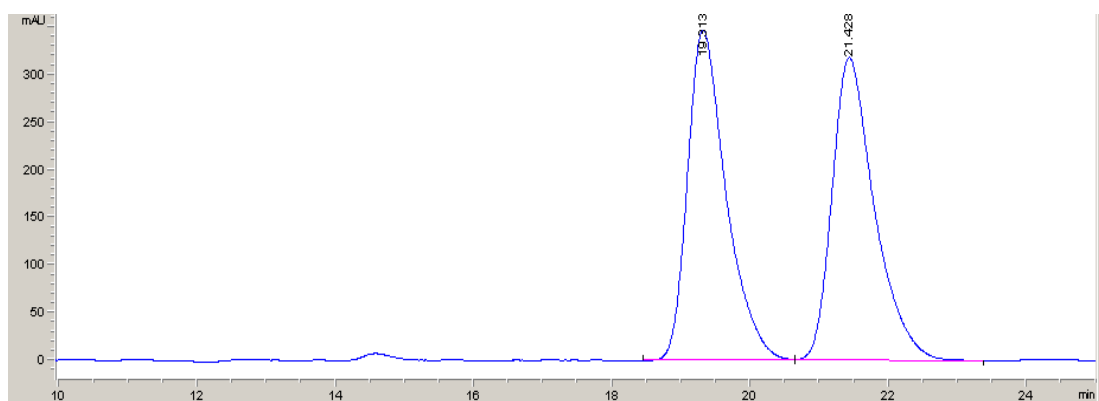
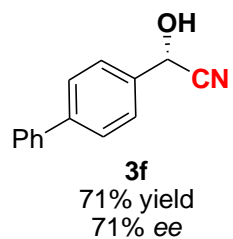
	Processed Chanel Descr	RT	Area	%Area	Height
1	210	18.258	6040570	90.04	167714
2	210	19.831	668192	9.96	17051



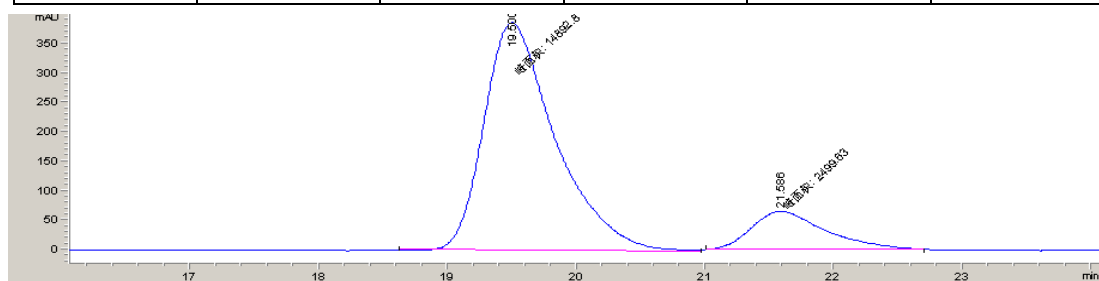
	Processed Chanel Descr	Time/min	Area	%Area	Height
1	210	12.337	1848249	49.94	102496
2	210	14.083	1852516	50.06	65181



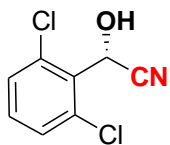
	Processed Chanel Descr	Time/min	Area	%Area	Height
1	210	12.299	10240711	88.93	661342
2	210	13.941	1288333	11.07	49857



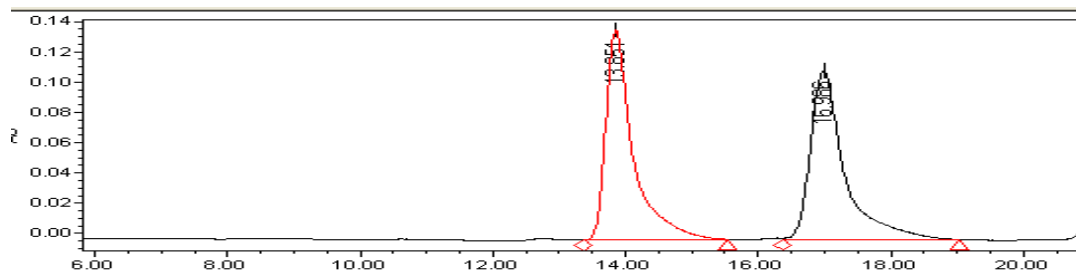
	Processed Chanel Descr	Time/min	Area	%Area	Height
1	210	19.313	135101	49.782	347
2	210	21428	136285	50.218	318.3



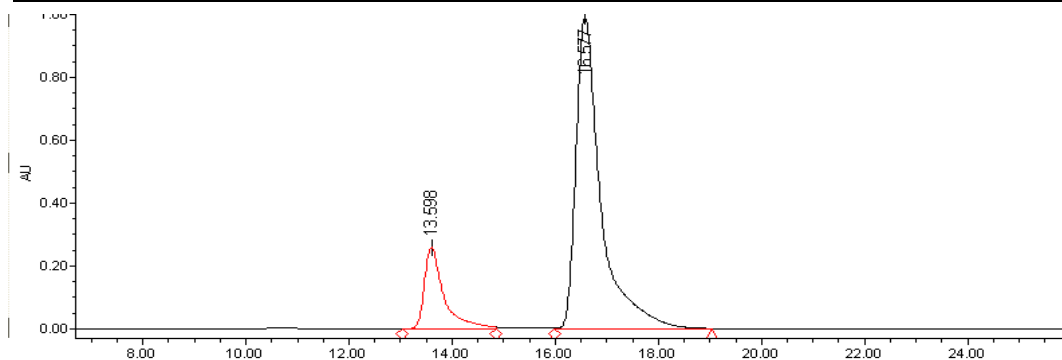
	Processed Chanel Descr	Time/min	Area	%Area	Height
1	210	19.5	14892.8	85.6283	386.9
2	210	21.586	2499.6	14.377	63.5



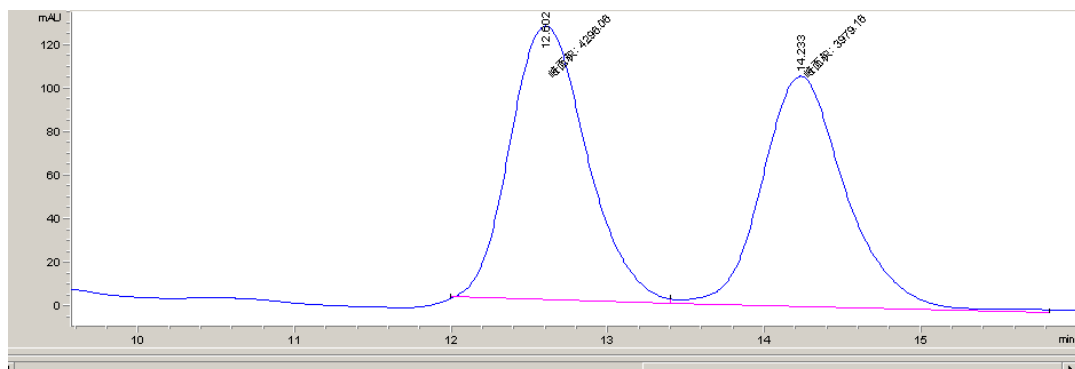
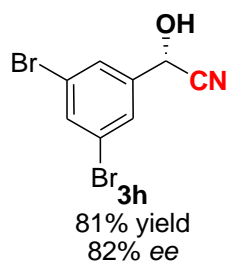
3g
94% yield
66% ee



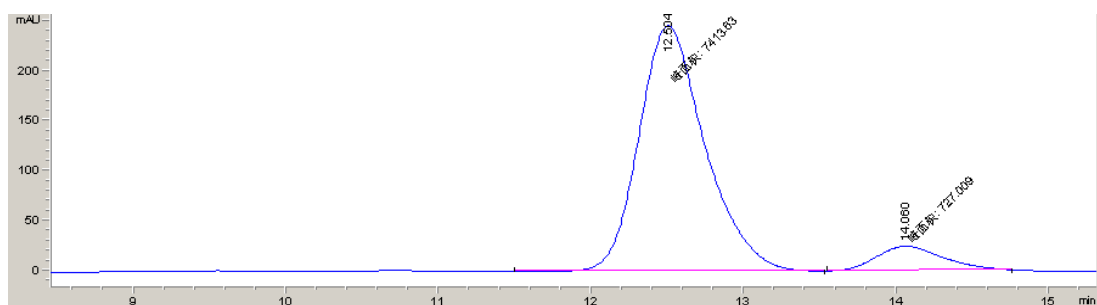
	Processed Chanel Descr	Time/min	Area	%Area	Height
1	210	13.851	14892.8	85.6283	386.9
2	210	16.986	2499.6	14.377	63.5



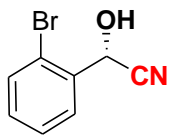
	Processed Chanel Descr	Time/min	Area	%Area	Height
1	210	13.598	6782412	16..97	258624
2	210	16.577	33181786	83.03	990106



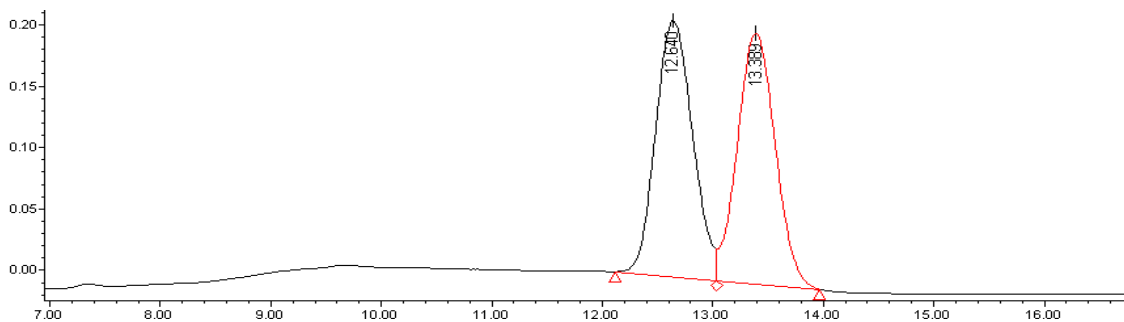
	Processed Chanel Descr	Time/min	Area	%Area	Height
1	210	12.602	4296.1	51.915	125.9
2	210	14.233	3979.2	48.085	105.9



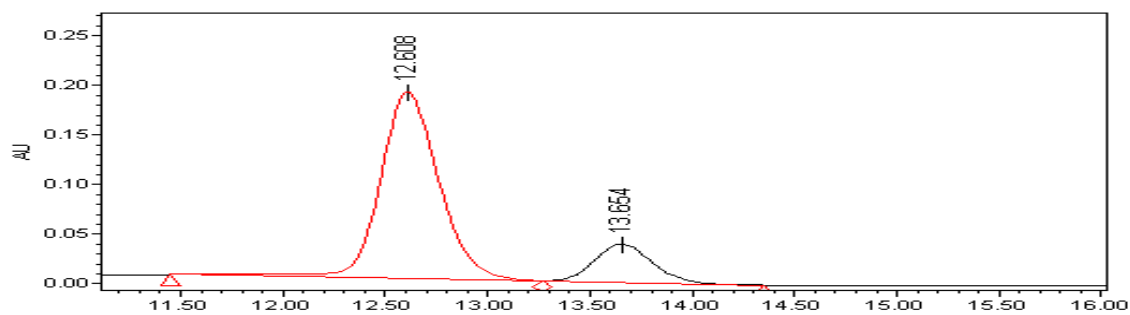
	Processed Chanel Descr	Time/min	Area	%Area	Height
1	210	12.504	7413.6	91.069	145.8
2	210	14.06	2499.6727	8.931	24.2



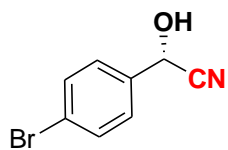
3i
81% yield
67% ee



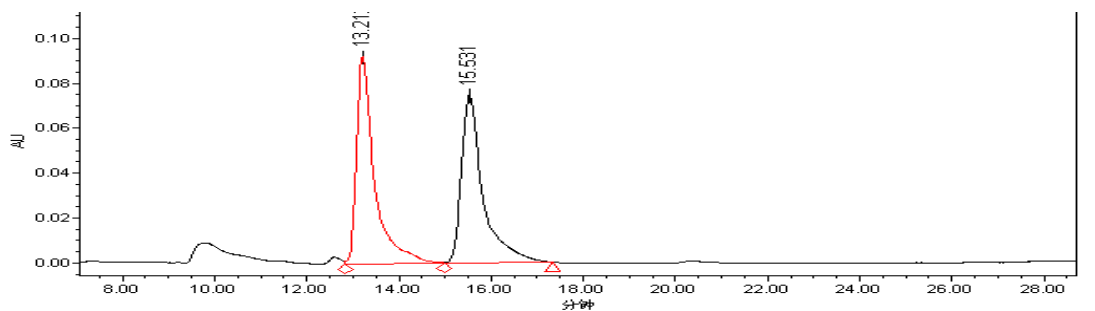
	Processed Chanel Descr	Time/min	Area	%Area	Height
1	210	12.640	4860600	50.06	208562
2	210	13.389	4849388	49.94	204079



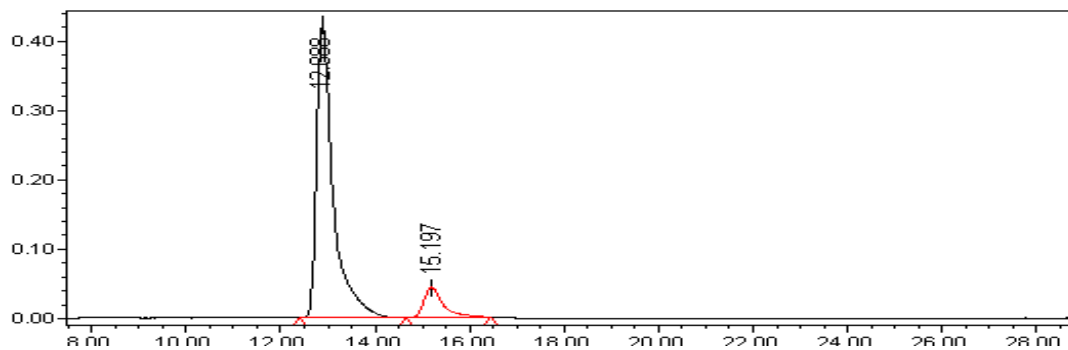
	Processed Chanel Descr	Time/min	Area	%Area	Height
1	210	12.608	3756088	83.46	188310
2	210	13.654	759342	16.54	18697



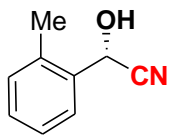
3j
72% yield
78% ee



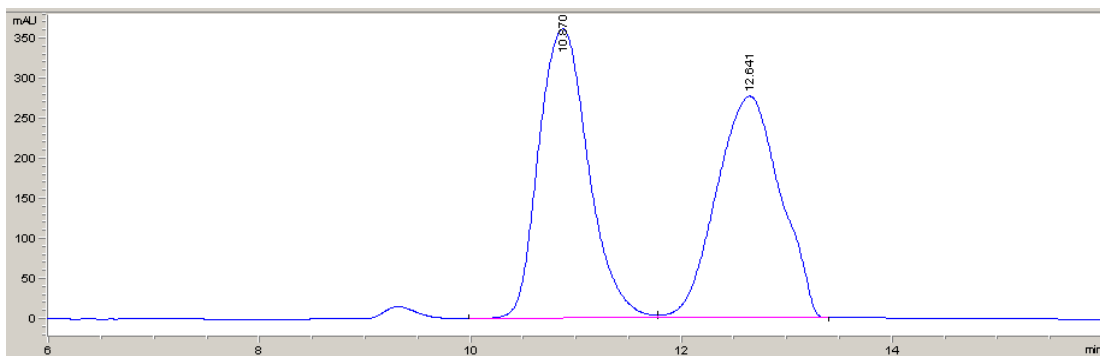
	Processed Chanel Descr	Time/min	Area	%Area	Height
1	210	13.212	2496844	50.73	92279
2	210	15.531	2424717	49.27	74772



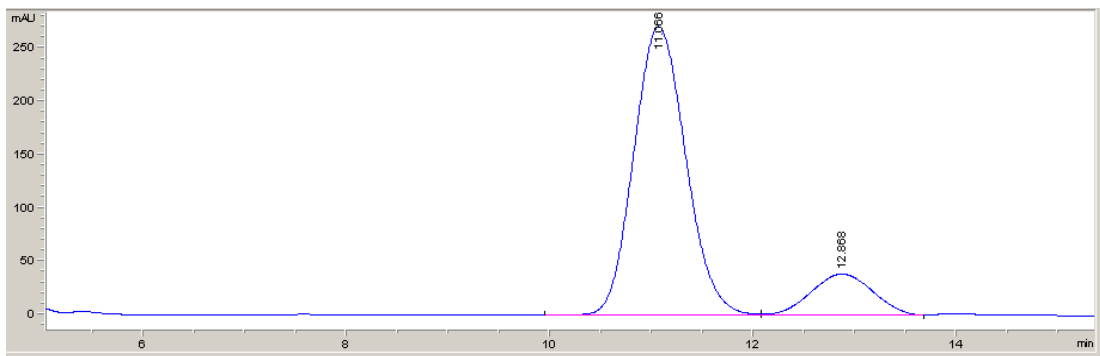
	Processed Chanel Descr	RT	Area	%Area	Height
1	210	12.888	10287943	89.11	188310
2	210	15.197	1257478	10.89	18697



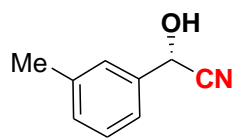
3k
71% yield
71% ee



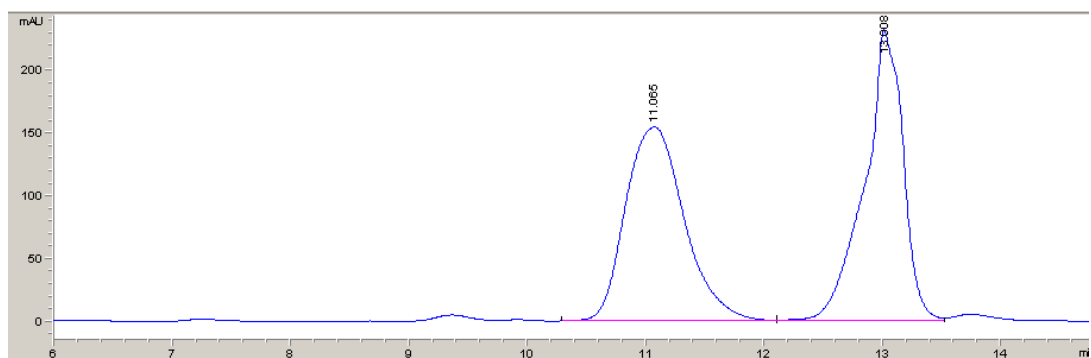
	Processed Chanel Descr	Time/min	Area	%Area	Height
1	210	10.87	11849.3	50.352	361.3
2	210	12.641	11683.4	49.648	276.8



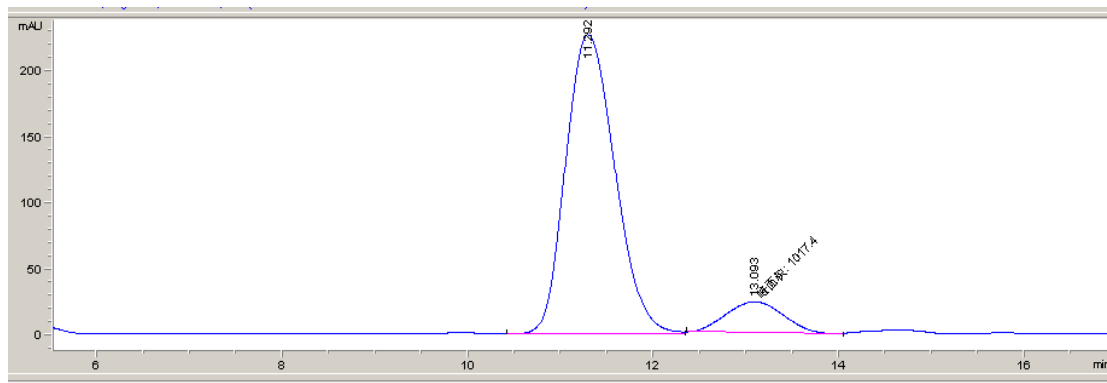
	Processed Chanel Descr	Time/min	Area	%Area	Height
1	210	11.066	9639.2	85.536	271.2
2	210	12.868	1629.9	14.464	38.7



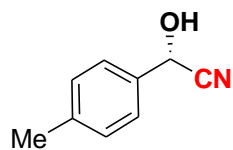
3I
71% yield
78% ee



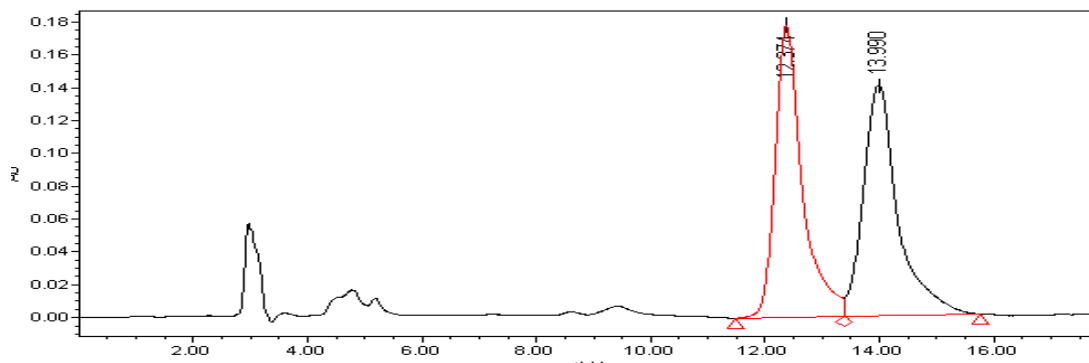
	Processed Chanel Descr	Time/min	Area	%Area	Height
1	210	11.065	5347.3	50.431	154.5
2	210	13.008	5255.9	49.569	231.9



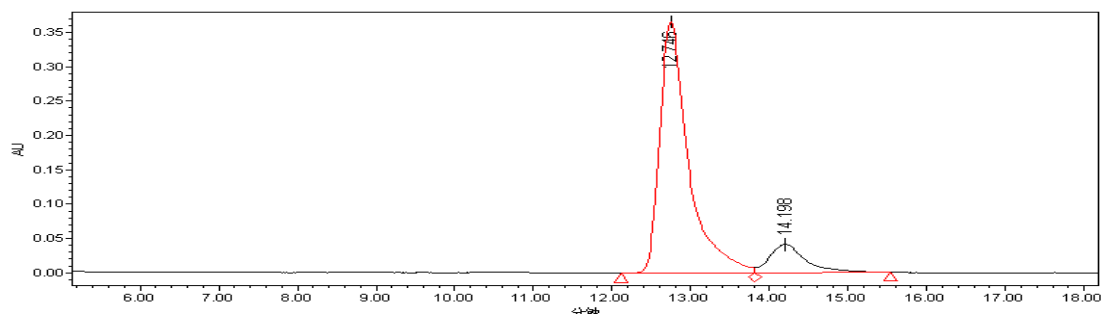
	Processed Chanel Descr	Time/min	Area	%Area	Height
1	210	11.292	8357.2	89.147	22.5
2	210	13.093	1017.4	10.853	23.7



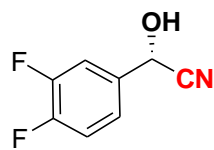
3m
72% yield
75% ee



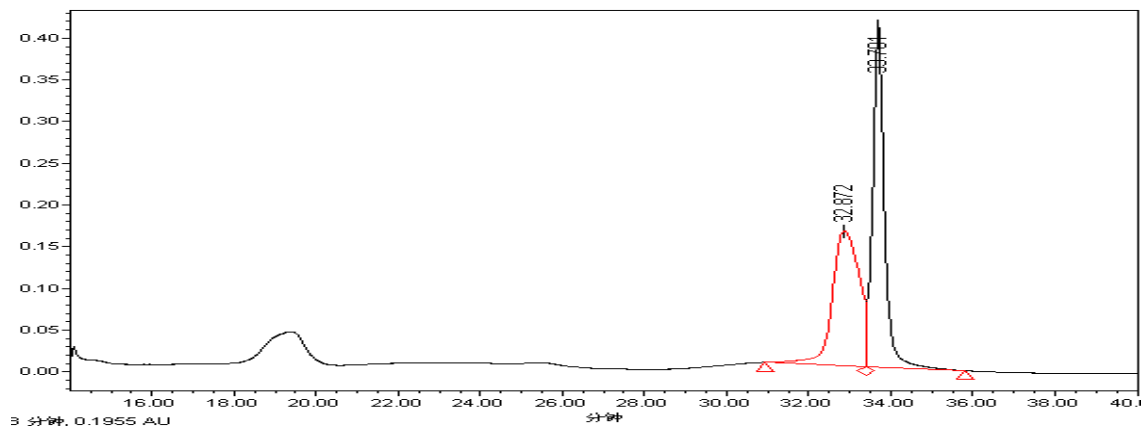
	Processed Chanel Descr	Time/min	Area	%Area	Height
1	210	12.374	5891259	49.89	177881
2	210	13.990	59.16406	50.11	140500



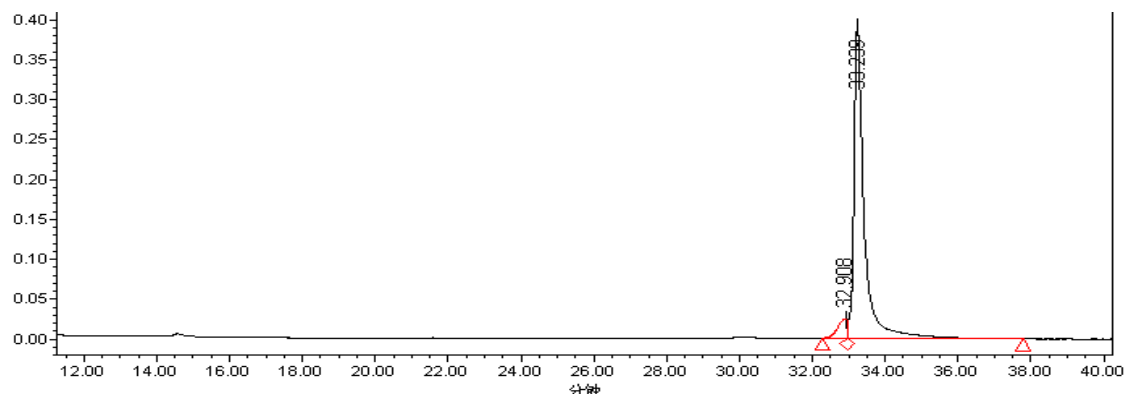
	Processed Chanel Descr	Time/min	Area	%Area	Height
1	210	12.746	9333394	87.56	365714
2	210	14.196	1326161	12.44	41386



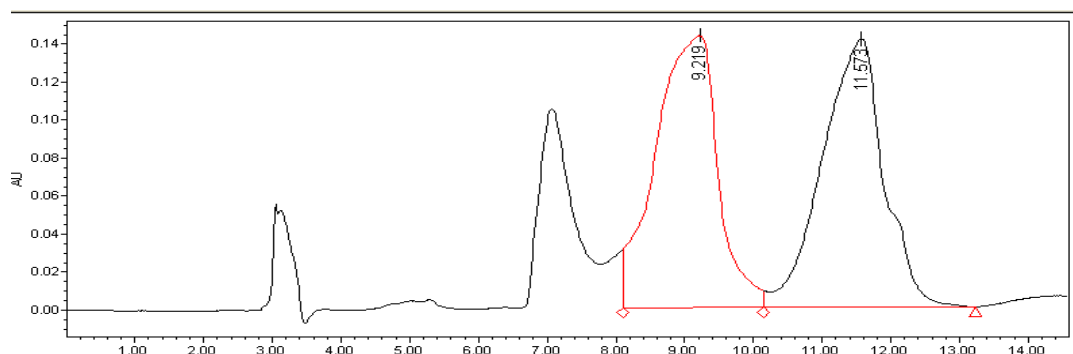
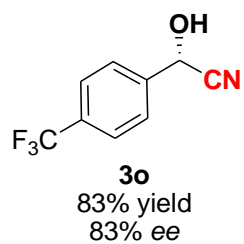
3n
84% yield
90% ee



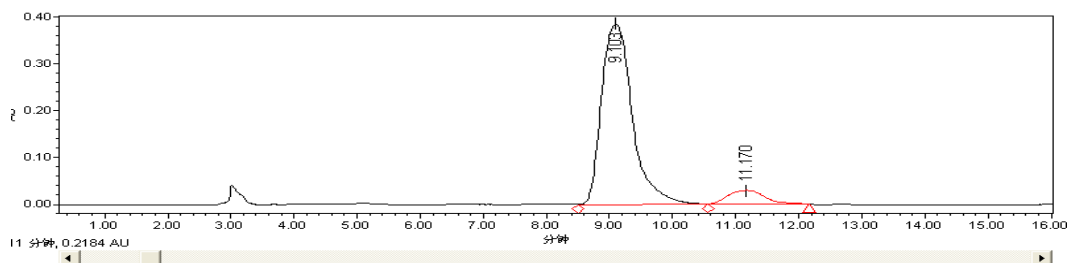
	Processed Chanel Descr	Time/min	Area	%Area	Height
1	210	32.872	7338301	47.19	161882
2	210	33.701	8212877	52.81	400403



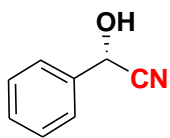
	Processed Chanel Descr	Time/min	Area	%Area	Height
1	210	32.908	410138	5.10	23967
2	210	33.239	7627099	94.90	390290



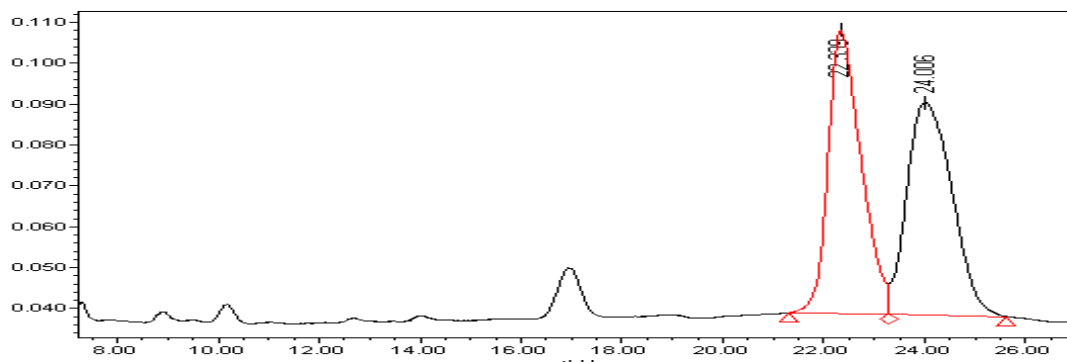
	Processed Chanel Descr	Time/min	Area	%Area	Height
1	210	9.219	9025977	50.65	143163
2	210	11.573	8792871	49.35	141162



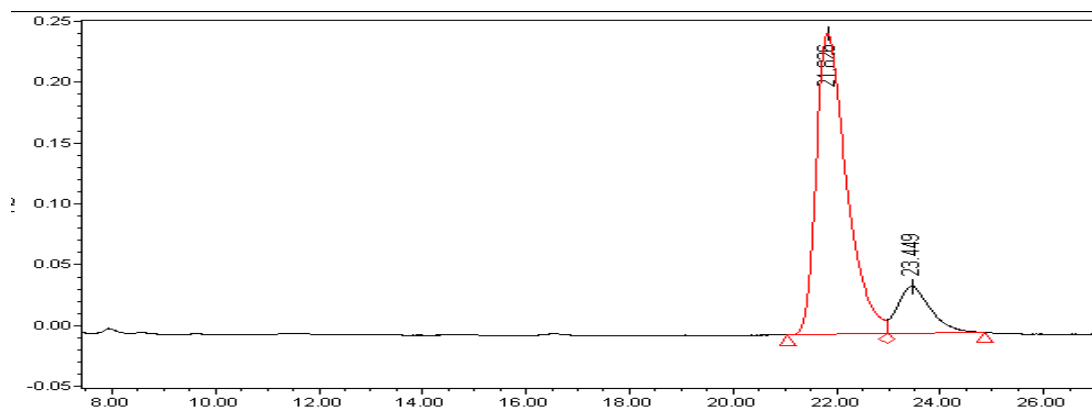
	Processed Chanel Descr	Time/min	Area	%Area	Height
1	210	9.103	12501667	91.44	384811
2	210	11.170	1226349	8.56	29639



3p
65% yield
71% ee



	Processed Chanel Descr	Time/min	Area	%Area	Height
1	210	22.339	3160665	49.3	59240
2	210	24.006	3250749	50.70	52047



	Processed Chanel Descr	Time/min	Area	%Area	Height
1	210	21.826	10000777	85.41	247014
2	210	23.449	1721857	14.59	38682