

1 Supporting information

2 **Quantifying the Distribution of Ceria Nanoparticles in**  
3 **Cucumber Plants: the Influence of Labeling**

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19 The supporting information includes:

20 1. Number of pages: 5

21 2. Number of figures: 4

22 **1. Preparation and Characterization of Nano-ceria**

23 In the present work, 0.5 M solution of hexamethyleneteramine (HMT) and 0.0375M  
24 solution of  $\text{Ce}(\text{NO}_3)_3 \cdot 6\text{H}_2\text{O}$  were mixed and stirred in 75°C water bath for 3 h. The  
25 particles obtained were washed three times by ultrapure water. The morphology and  
26 size distribution of the as-synthesized nano-ceria were characterized by TEM (FEI  
27 Co., Eindhoven, The Netherlands). The samples were dispersed in ultrapure water and  
28 sonicated for 15 min before dropped on a Cu grid for observation. X-ray diffraction  
29 (XRD) of nano-ceria was obtained on a Panalytical's X'Pert PRO diffractometer  
30 (Panalytical BV, the Netherlands) using monochromatized Cu  $K\alpha$  radiation at 30 kV  
31 and 40 mA. The XRD patterns were scanned from 5-90°  $2\theta$  range using 0.02° step  
32 intervals. The size distribution and the surface charge density (in terms of zeta  
33 potential) of as-synthesized nano-ceria (AS-ceria) and diiodofluorescein-coated nano-  
34 ceria (DIF-ceria) in ultrapure water were evaluated by dynamic light scattering (DLS)  
35 analysis (Zeta-Sizer, Malvern Instruments, UK). The hydrodynamic sizes of AS-ceria  
36 and DIF-ceria were  $34.2 \pm 9.7$  nm (PDI=0.218) and  $39.5 \pm 13.4$  nm (PDI=0.241),  
37 respectively; their zeta-potentials were  $36.6 \pm 5.9$  mV and  $34.3 \pm 7.0$  mV.

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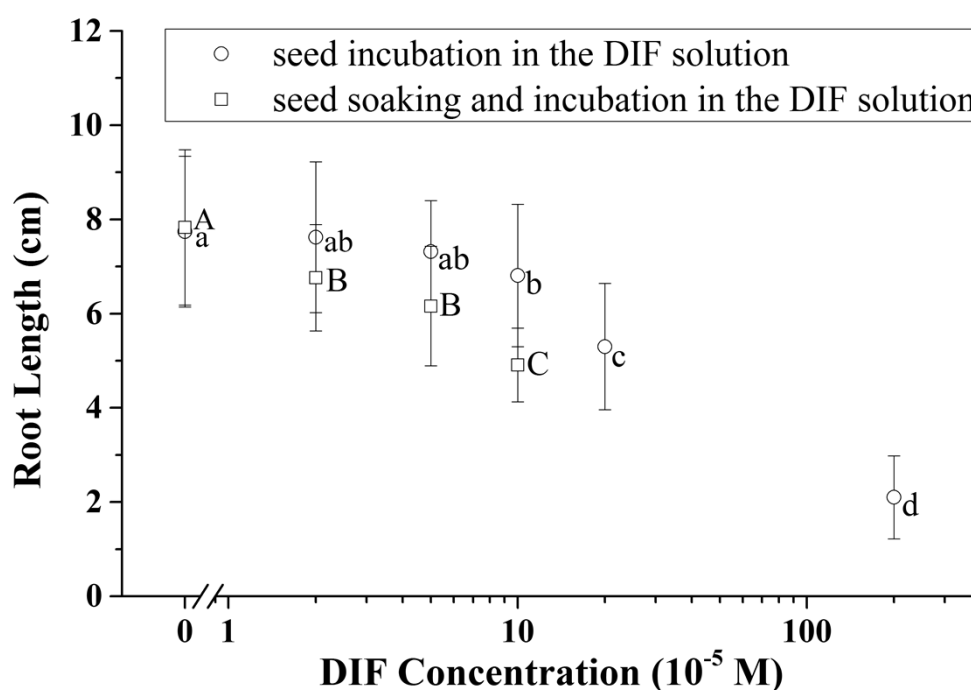
39 Figure S1. The XRD pattern of nano-ceria.

40 The specific surface area of nano-ceria was  $86.85 \text{ m}^2 \cdot \text{g}^{-1}$ , measured by the BET

41 method using a surface area analyzer (Autosorb-1 model, Quantachrome, USA).  
42 Assuming that each DIF molecule could cover 0.9 nm<sup>2</sup> of the particulate surface  
43 (fluorescein is a nonspherical solute with an approximate size of 0.47, 0.81, and 1.09  
44 nm in different directions)<sup>23</sup>, 25% of the surface of nano-ceria was covered with DIF  
45 when 25 mg nano-ceria were mixed with 50 mL 2×10<sup>-5</sup> M DIF solution.

## 46 2. Effect of DIF exposure on the root elongation of cucumber

47 The root elongation of cucumber was determined after a 5-day incubation in DIF  
48 solution at a concentration of 2×10<sup>-5</sup> M. In a parallel study, the seeds were pre-soaked  
49 in the respective DIF solution for 2 h before the 5-day germination test.



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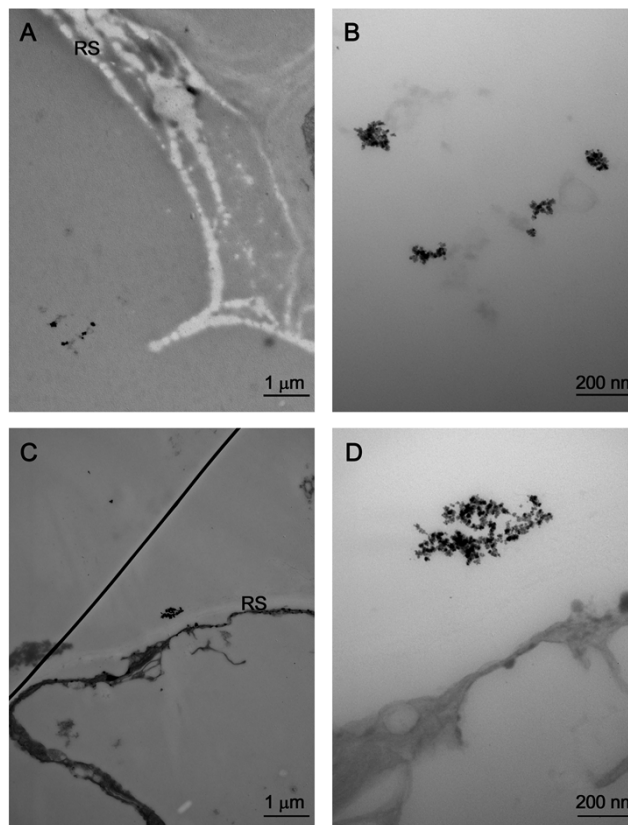
51 Figure S2. Cucumber root elongation after a 5-day incubation in DIF solution or a 2-hour pre-soaking followed by  
52 a 5-day incubation in DIF solution. The cucumber root length is expressed as mean±SD of 40 replicates. In the  
53 seed incubation treatment, data labeled with different lowercase letters are significantly different at  $p < 0.05$ ; in the  
54 seed soaking and incubation treatment, data labeled with different uppercase letters are significantly different at  $p$   
55  $< 0.05$ .

## 56 3. TEM images of the cross sections of cucumber roots treated with AS-ceria and

57 **DIF-ceria**

58 After a 5-day germination, cucumber roots were washed with deionized water  
59 thoroughly and the root apices were cut and fixed in 2.5% glutaraldehyde solution.  
60 Then they were dehydrated in a graded acetone series and embedded in Spurr's resin.  
61 Ultrathin sections (90 nm) were obtained using an UC6i ultramicrotome (Leica,  
62 Austria) with a diamond knife. The sections were collected on copper grids and  
63 observed on a JEM-1230 (JEOL, Japan) transmission electron microscope operating  
64 at 80 kV without staining.

65 As shown in figure S3, most of the AS-ceria and DIF-ceria agglomerates were found  
66 on the surface of roots though the seedlings had been thoroughly washed.



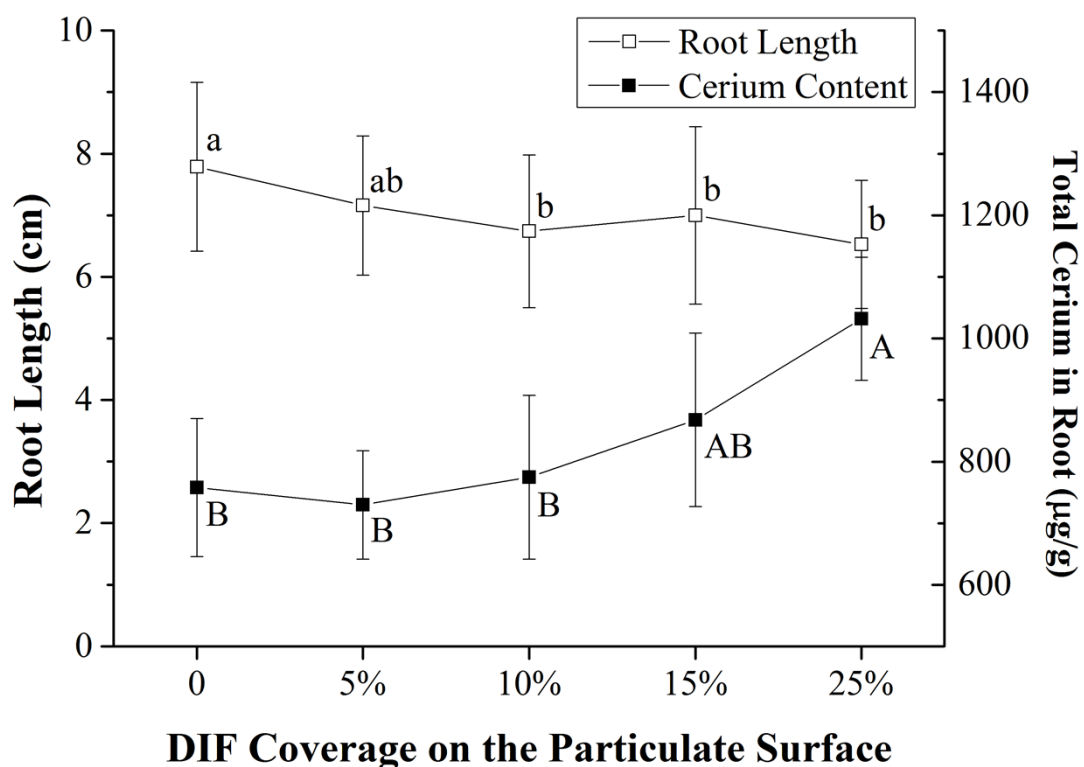
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68 Figure S3. TEM images of the cross sections of cucumber roots treated with As-ceria (A&B) and

69 DIF-ceria (C&D). RS: root surface.

70 **4. The influence of DIF coverage**

71 Our results showed that fluorescent labeling could change the behaviors and bio-  
72 effects of nano-ceria when 25% of the particulate surface was coated with DIF. We  
73 further determined the labeling-caused changes by different DIF-coverage rates. The  
74 results suggest that to minimize the labeling-caused changes, the DIF coverage rate  
75 should not be larger than 5%.



76

77 Figure 4. The changes in the root length and root cerium content caused by the different DIF coverage rates on the  
78 surface of nano-ceria. Cucumber root lengths labeled with different lowercase letters are significantly different at  $p$   
79  $< 0.05$ ; As for the contents of total cerium in roots, data labeled with different uppercase letters are significantly  
80 different at  $p < 0.05$ .