

Supporting Information

Indium(III)-catalyzed Tandem Synthesis of 2-Alkynyl-3,3-dichloropyrrolidines and their Conversion to 3-Chloropyrroles

Khushbu Kushwaha,^a Chandi C. Malakar,^a Sara Stas,^a Filip Lemièr^b and Kourosch

Abbaspour Tehrani^{a*}

^a *Organic Synthesis, Department of Chemistry, University of Antwerp, Groenenborgerlaan 171, B-2020 Antwerp, Belgium. Fax: (+32)32653233; phone: (+32)32653226.*

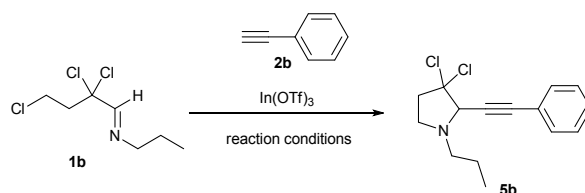
^b *Biomolecular & Analytical Mass Spectrometry and Center for Proteomics, Department of Chemistry, University of Antwerp, B-2020 Antwerp, Belgium.*

E-mail: kourosch.abbaspourtehrani@uantwerpen.be

Table of Contents

1. Optimization of the reaction conditions	2-3
2. Copies of ¹ H and ¹³ C spectra of starting imines 1a-f	4-12
3. Formation of side products 5c' and 5f'	13-15
4. Copies of ¹ H and ¹³ C spectra of 2-alkynylpyrrolidines 5a-q	15-45
5. Copies of ¹ H and ¹³ C spectra of 2-alkenylpyrroles 6a-f and 7	45-60
6. Mechanistic study for the borontrifluoride catalyzed synthesis of 5a	61-62
7. Mechanistic study for the indium triflate catalyzed synthesis of 5a	62-64
8. Copies of ¹ H and ¹³ C spectra of 3,3-dichloro-1-ethyl-2-(2-phenylvinyl)pyrrolidine (12a)	65-66
9. Copies of ¹ H and ¹³ C spectra of 13a and 14a	66-69
10. References	69

1. Optimization of the Reaction Conditions for the Synthesis of **5b** using In(OTf)₃^a



Entry	Catalyst/[mol%]	2b [equiv]	Solvent	<i>t</i> [°C]	Yield 5b [%]
1	In(OTf) ₃ /25	1.0	DCM	50	60 ^b
2	In(OTf) ₃ /25	1.0	DCE	70	5 ^c
3	In(OTf) ₃ /25	1.0	DCM/HFIP	rt	0
4	In(OTf) ₃ /25	1.0	THF	70	SM
5	In(OTf) ₃ /40	1.0	DCM	50	64 ^b
6	In(OTf) ₃ /10	1.0	DCM	50	34 ^b
7	In(OTf) ₃ /25	1.5	DCM	50	74 ^d
8	In(OTf)₃/25	2.0	DCM	50	80^d
9	In(OTf) ₃ /25	2.5	DCM	50	81 ^d

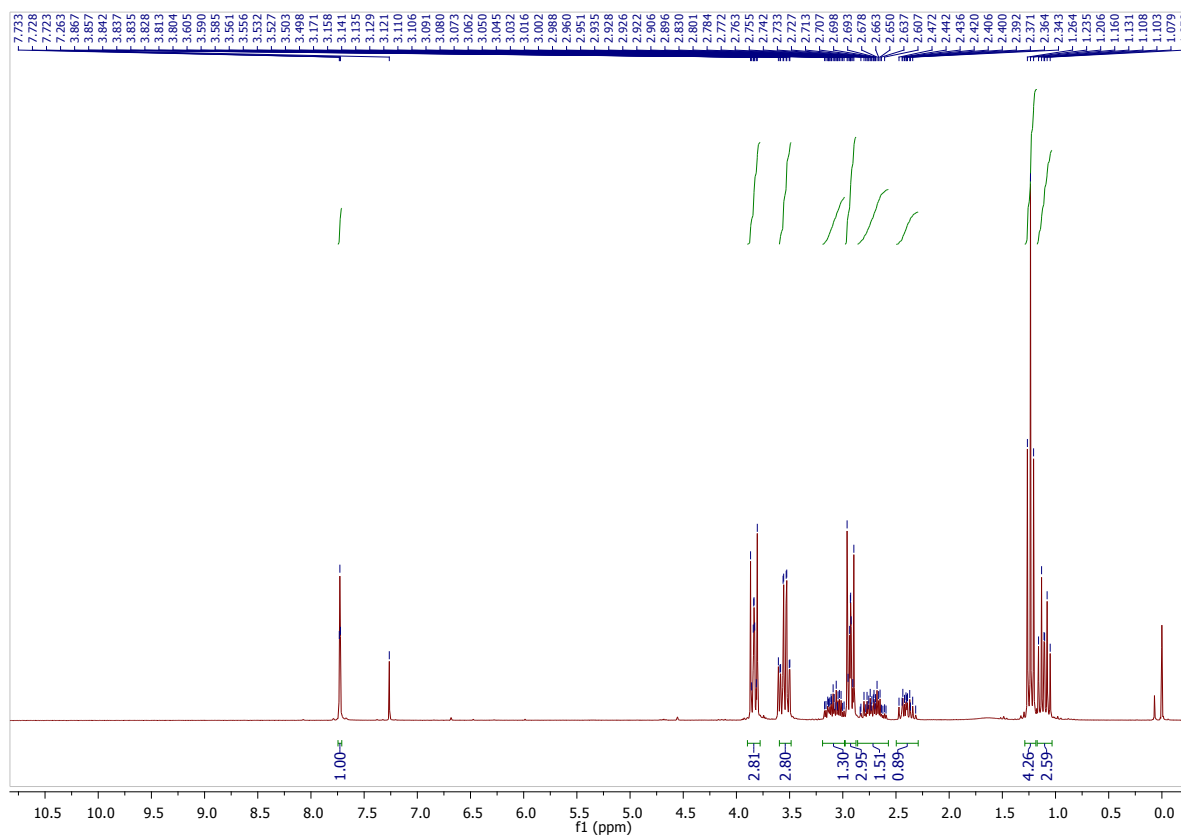
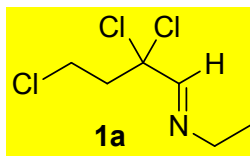
^aAll reactions were performed using 0.5 mmol **1b** in DCM (4 mL) in a sealed vial under air for 18h, unless otherwise stated. ^bYield after chromatography using a short column of silica gel. ^cYield calculated from ¹H NMR. ^dIsolated yield after basic workup.

First of all, we tried to isolate the product **5b**, obtained under the optimized reaction conditions, by means of column chromatography on silica gel, which resulted in a reasonably good yield of 60% (entry 1). The more convenient acid base workup yielded a more complex mixture and required still further column chromatography which reduced the yield even more. Replacing dichloromethane with 1,2-dichloroethane as solvent and heating the reaction mixture at 70 °C resulted in a complex mixture and yielded **5b** only in 5% yield (entry 2). Since there have been precedents¹ that the use of fluorinated solvents can result in extraordinary effects on the efficiency, regioselectivity and stereoselectivity of several chemical reactions, we also used hexafluoroisopropanol (HFIP) as co-solvent. This reaction gave a complex mixture with multiple unidentifiable side products. More polar solvents such as THF did not result in any reaction and only starting materials were recovered (entries 3 and 4). An increase in the amount of In(OTf)₃ from 25 mol% to 40 mol%, did not show a relevant improvement and product **5b** was isolated in 64% yield. Decreasing the amount of In(OTf)₃ to 10 mol% decreased the yield to 34% (entries 5 and 6). When the reaction was conducted with an excess of phenylacetylene (**2b**) (1.5 equiv) a 74% yield was obtained

(entry 7). Further improvement in the reaction was made by using 2 equivalents of phenylacetylene which delivered the product **5b** in 80% yield (entry 8). Under this condition the crude reaction mixture was purer and the formation of hemiaminal side product **5b'** could be completely avoided as it was not observed in the ^1H NMR. With 2.5 equivalents of phenylacetylene 81% yield was obtained, which is not a substantial improvement compared to that from the previous reaction (entry 9).

1. Copies of ^1H and ^{13}C spectra of starting imines

N-(2,2,4-Trichloro-1-butylidene)ethylamine (**1a**)



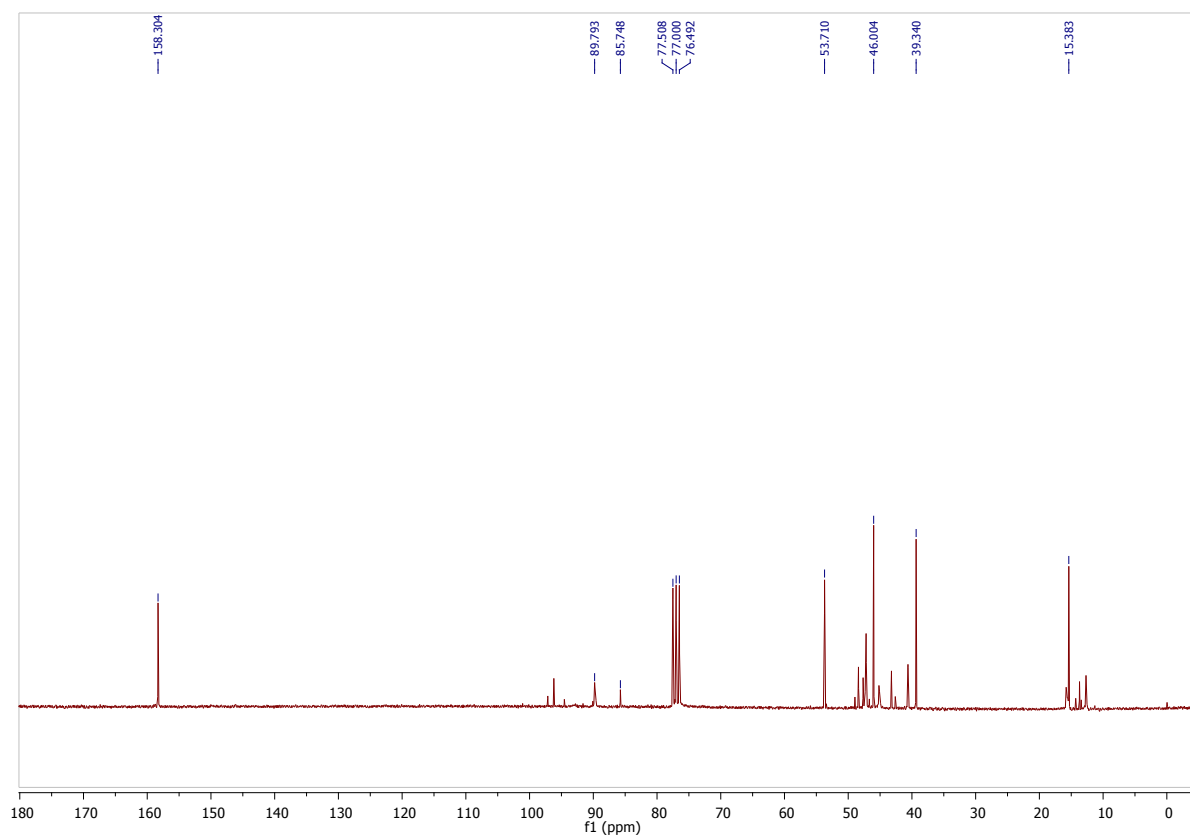
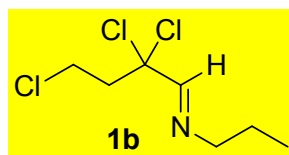


Figure 1. ^1H (250 MHz) and ^{13}C (62.90 MHz) NMR spectra of **1a** in CDCl_3 .

***N*-(2,2,4-Trichlorobutylidene)propan-1-amine (**1b**)**



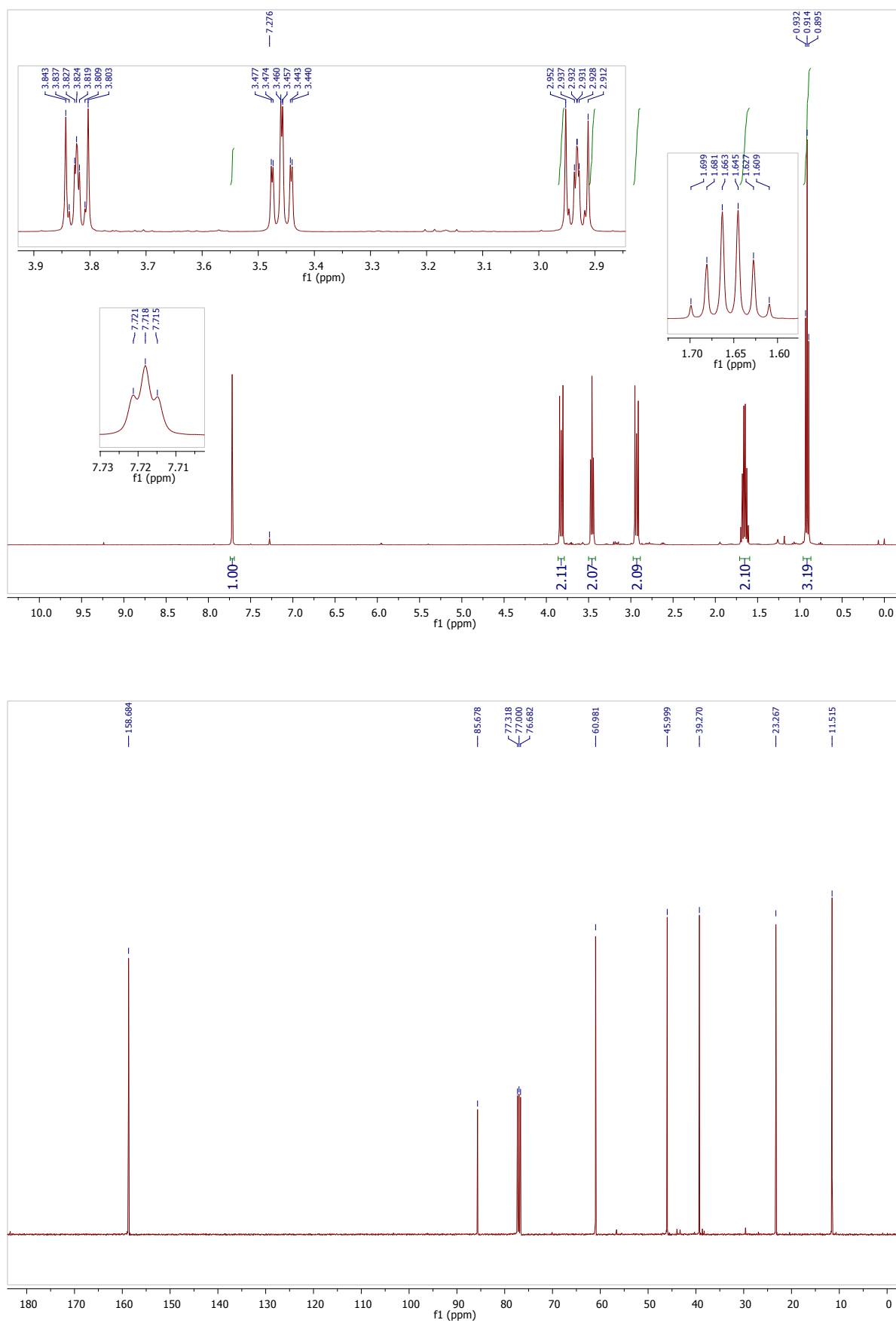
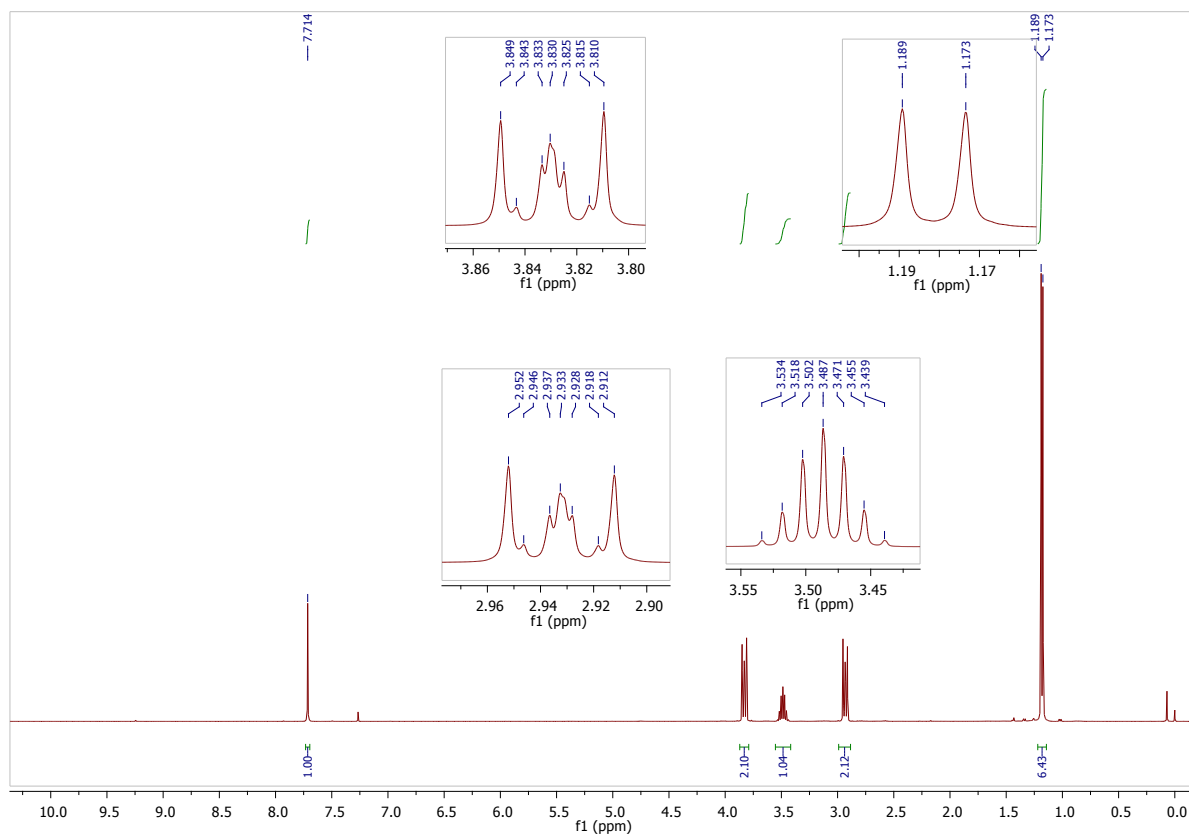
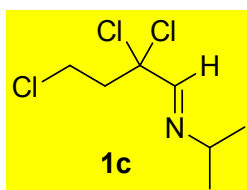


Figure 2. ¹H (400 MHz) and ¹³C (100 MHz) NMR spectra of **1b** in CDCl₃.

***N*-(2,2,4-Trichlorobutylidene)propan-2-amine (1c)**



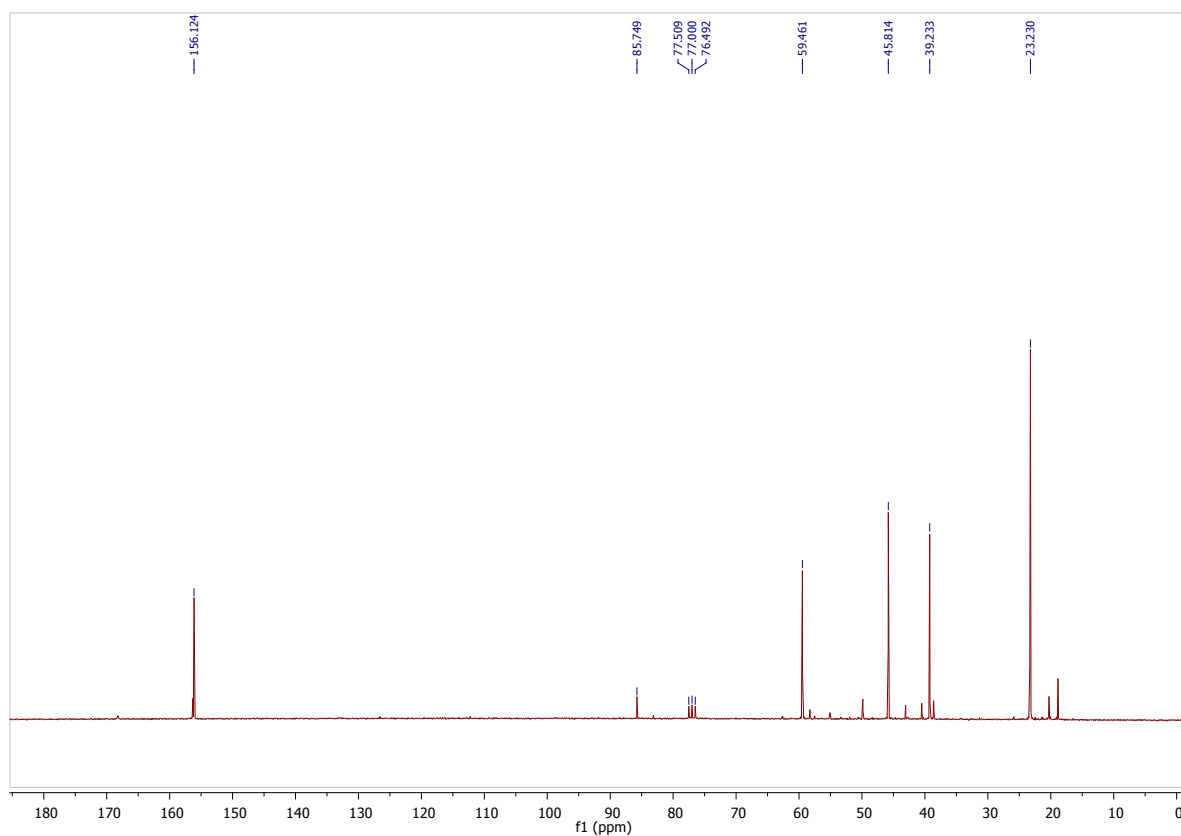
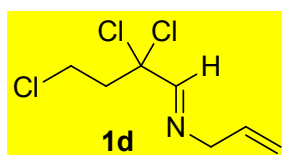
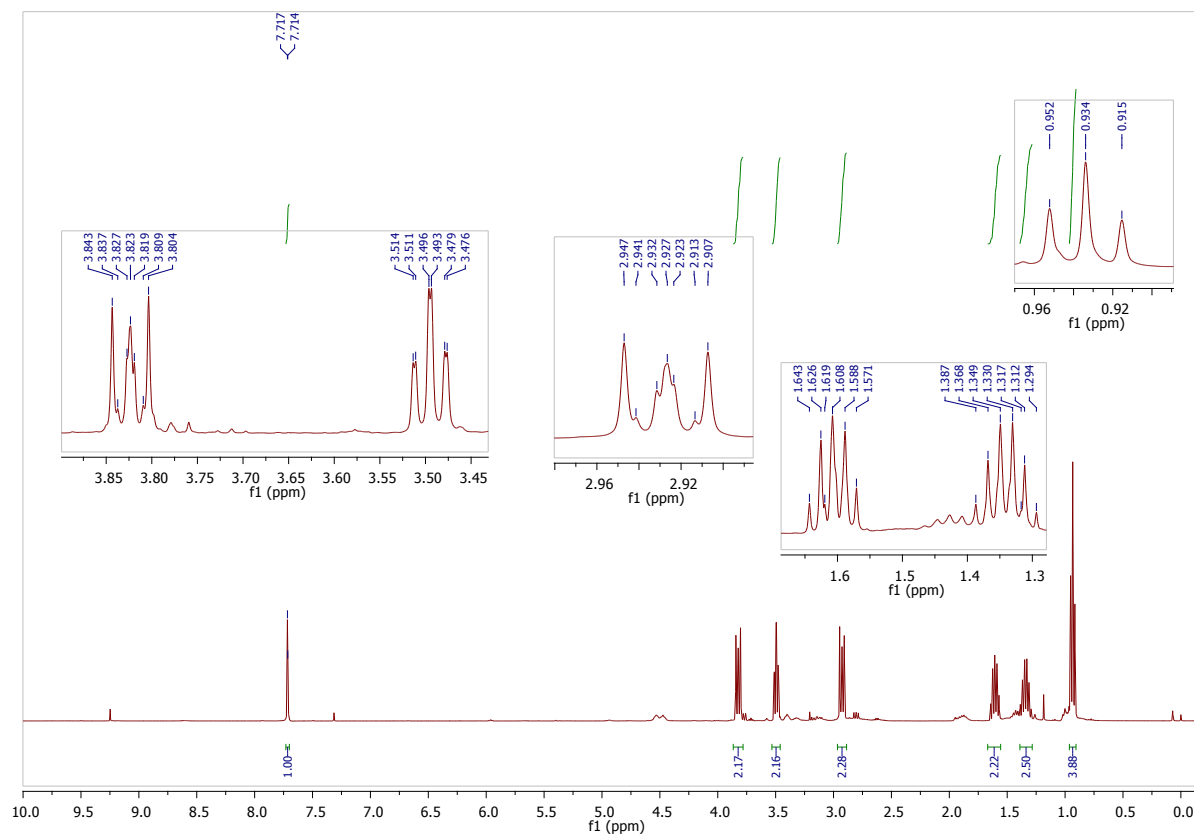
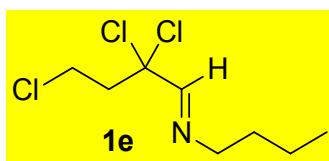


Figure 3. ^1H (400 MHz) and ^{13}C (62.90 MHz) NMR spectra of **1c** in CDCl_3 .

***N*-(2,2,4-Trichlorobutylidene)prop-2-en-1-amine (**1d**)**



***N*-(2,2,4-Trichlorobutylidene)butan-1-amine (1e)**



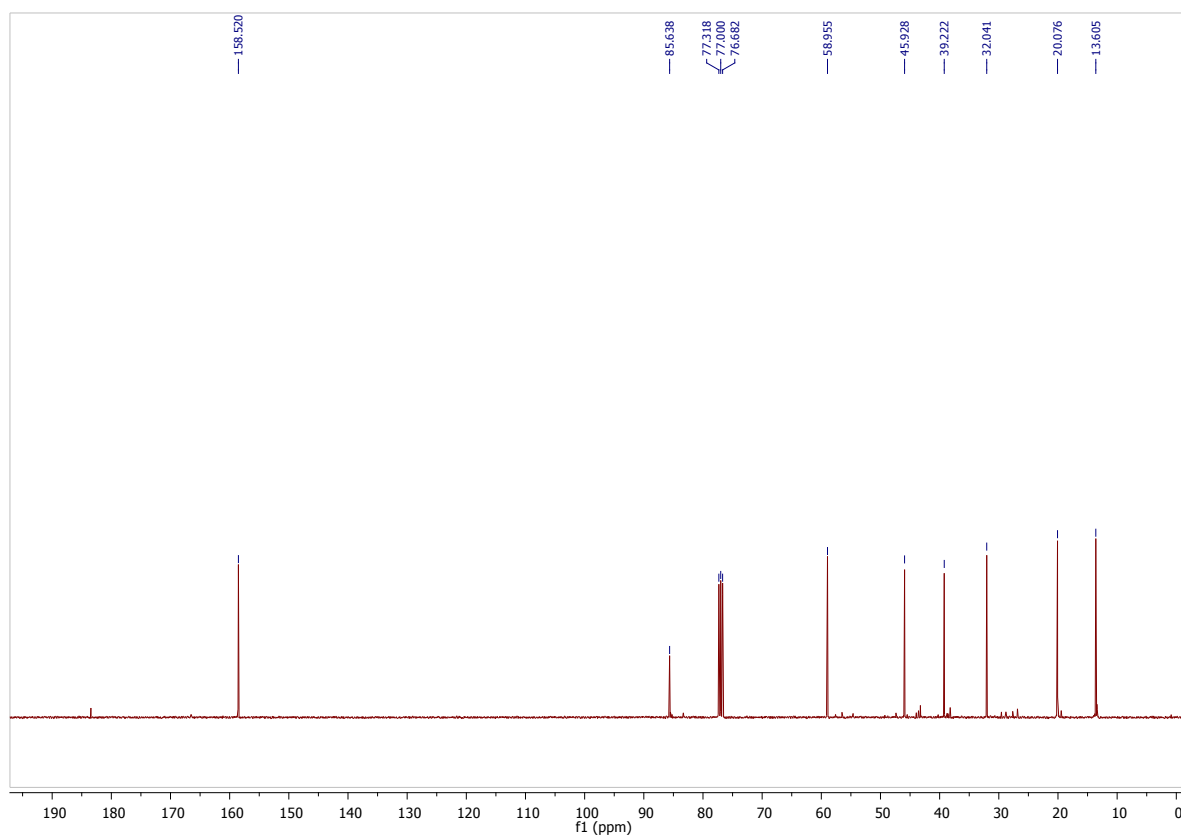
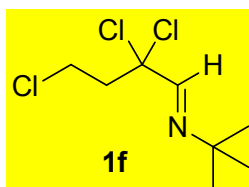


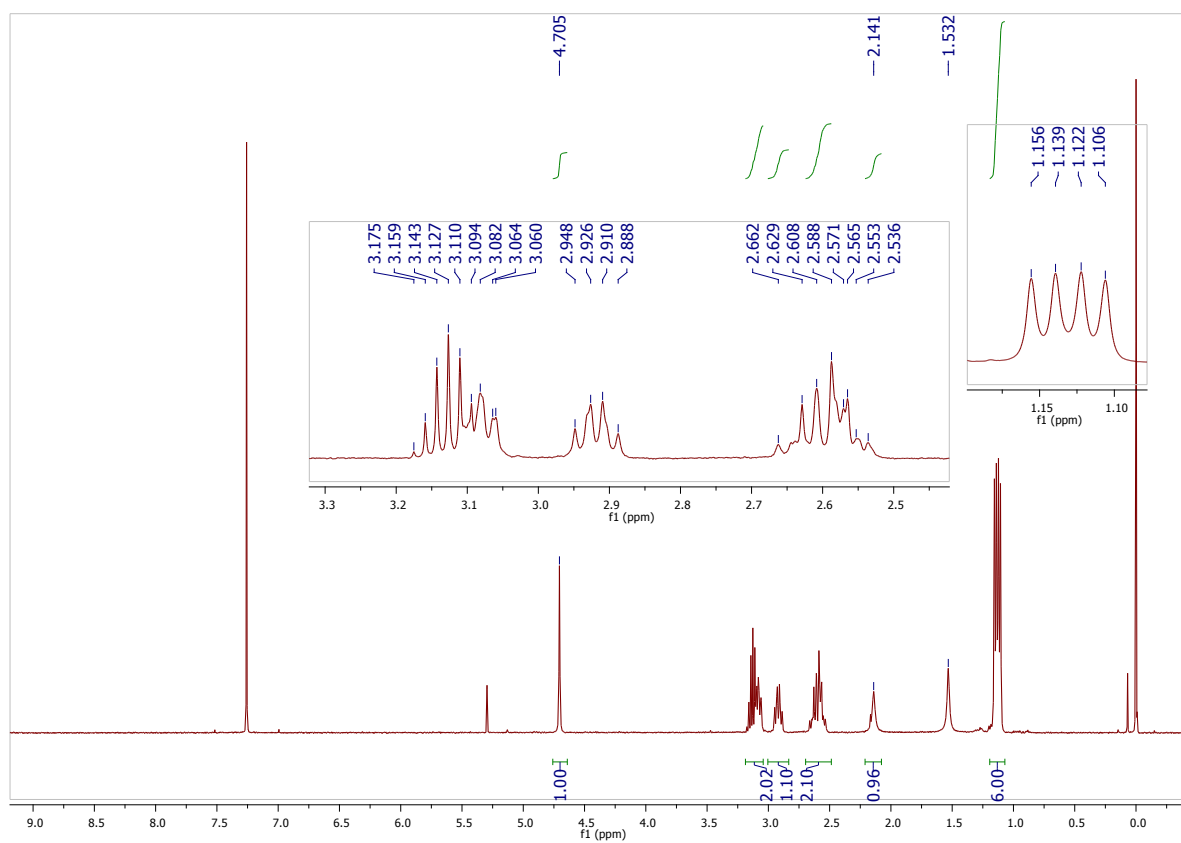
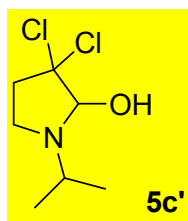
Figure 5. ^1H (400 MHz) and ^{13}C (100 MHz) NMR spectra of **1e** in CDCl_3 .

***N*-tert-Butyl-(2,2,4-trichloro-1-butyldene)amine (**1f**)**



2. Formation of side products

3,3-Dichloro-1-isopropylpyrrolidin-2-ol (5c')



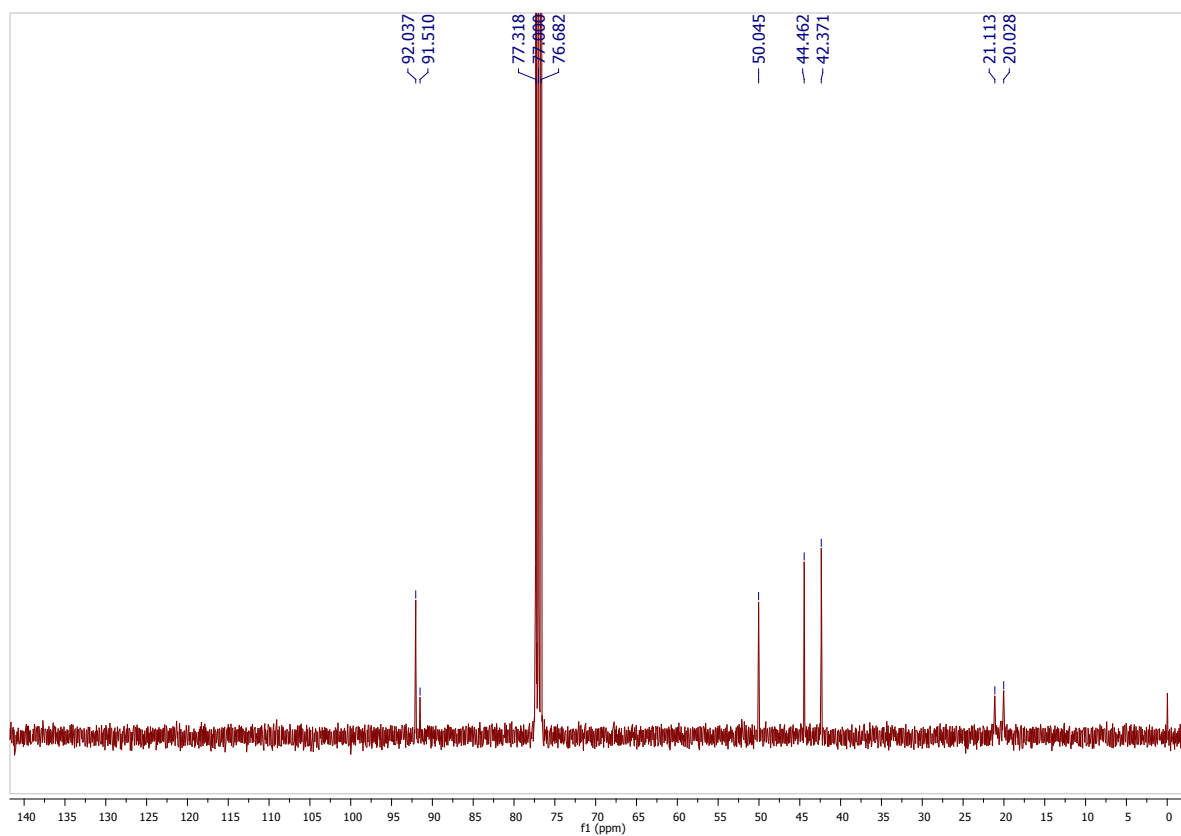
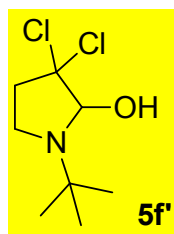


Figure 7. ^1H (400 MHz) and ^{13}C (100 MHz) NMR spectra of **5c'** in CDCl_3 .

1-*tert*-Butyl-3,3-dichloro-2-hydroxypyrrolidine (5f'**)**



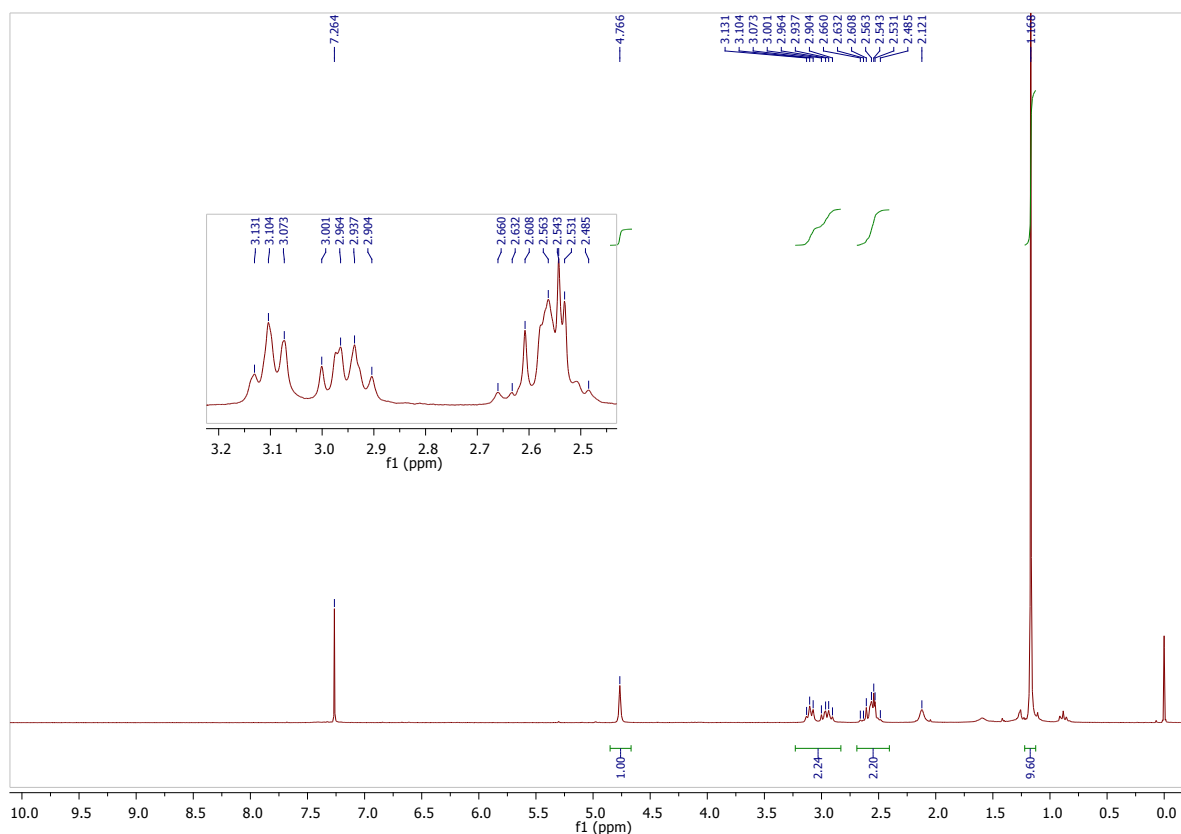
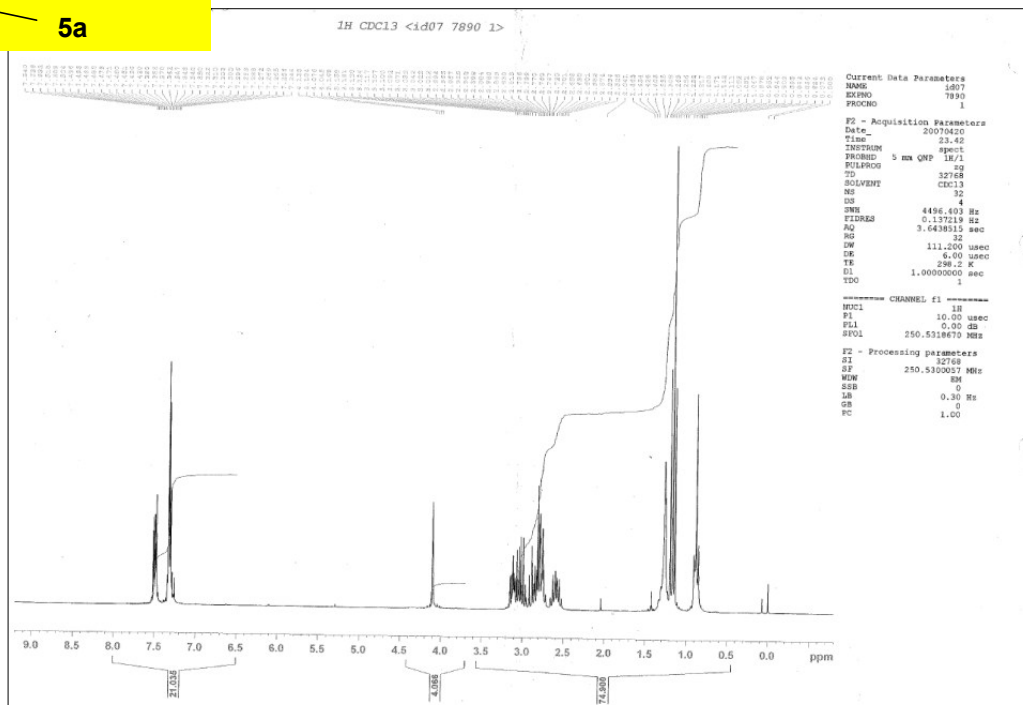
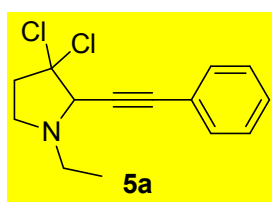
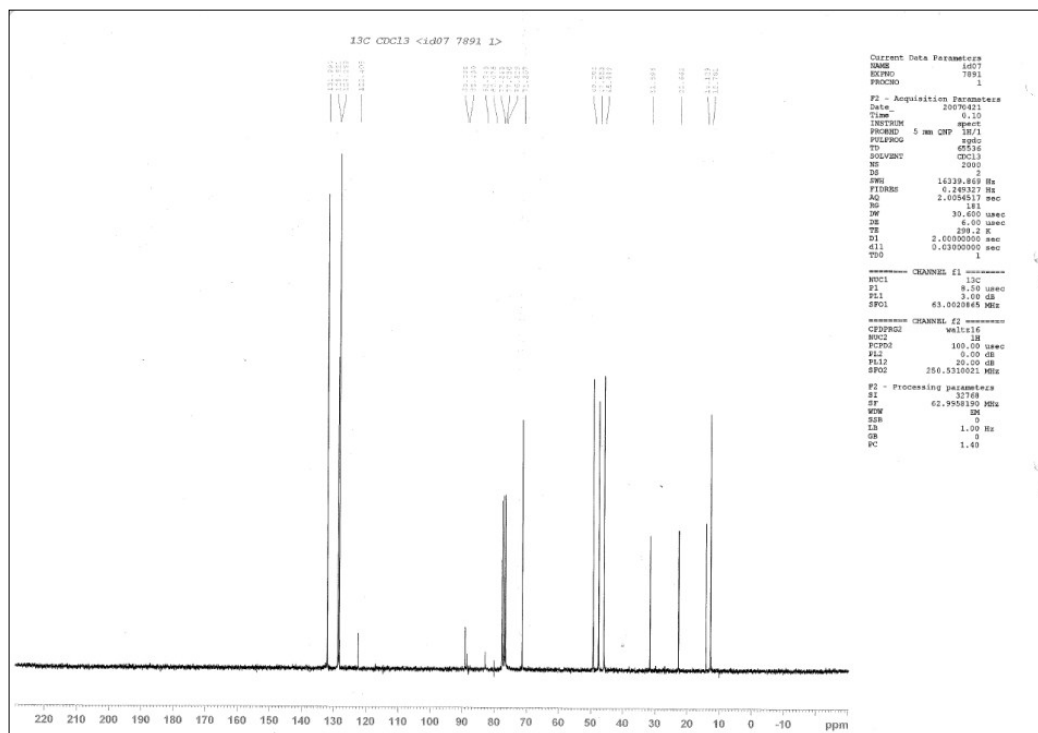


Figure 8. ^1H (400 MHz) NMR spectrum of **5f** in CDCl_3 .

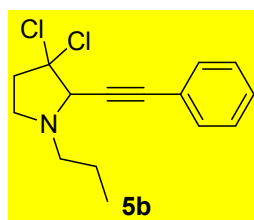
3. Copies of ^1H and ^{13}C spectra of 2-alkynylpyrrolidines **5a-q**

3,3-Dichloro-1-ethyl-2-(phenylethynyl)pyrrolidine (**5a**)





3,3-Dichloro-1-propyl-2-(phenylethynyl)pyrrolidine (5b)



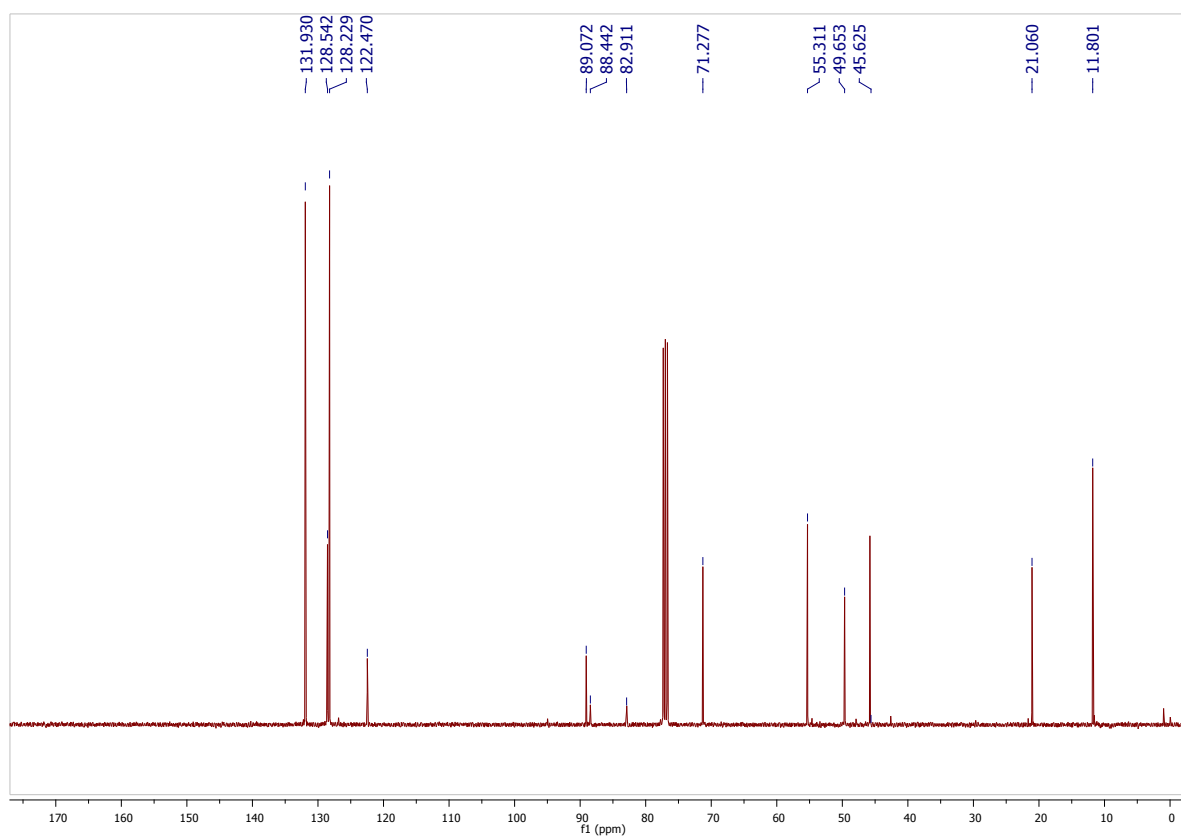
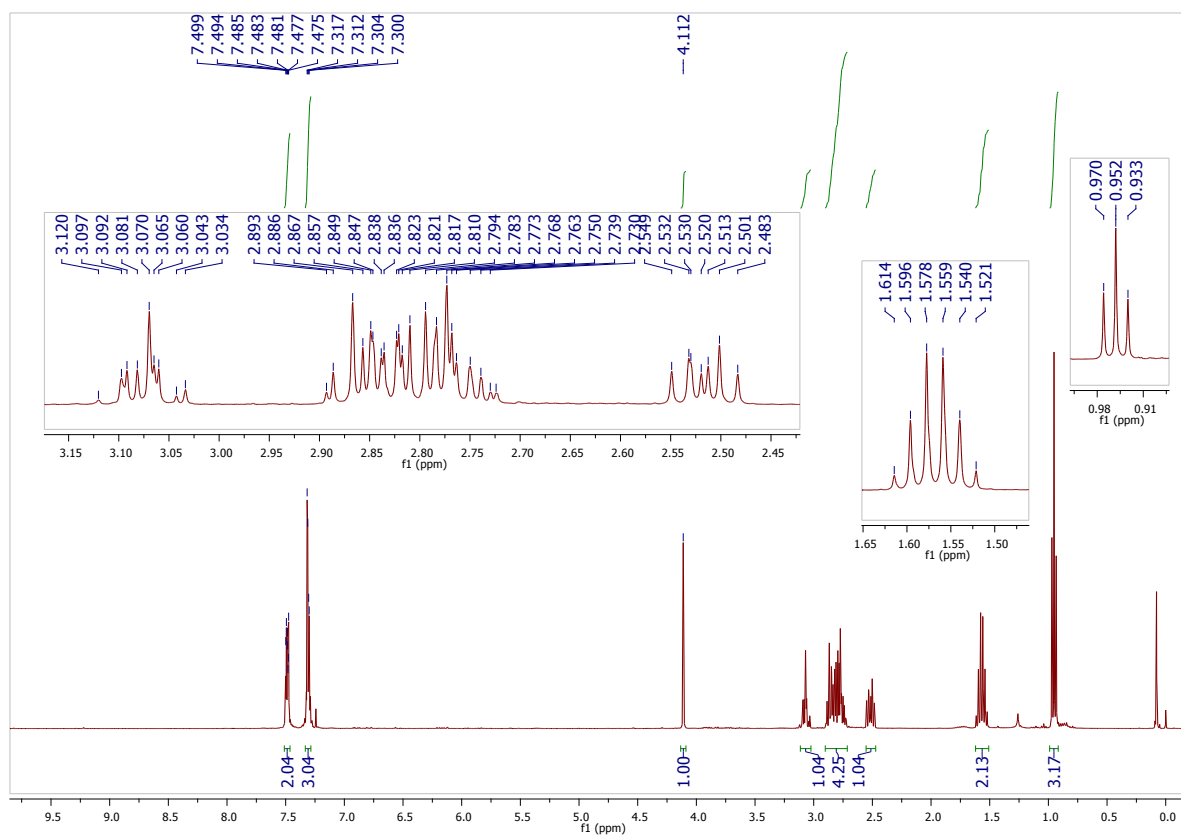
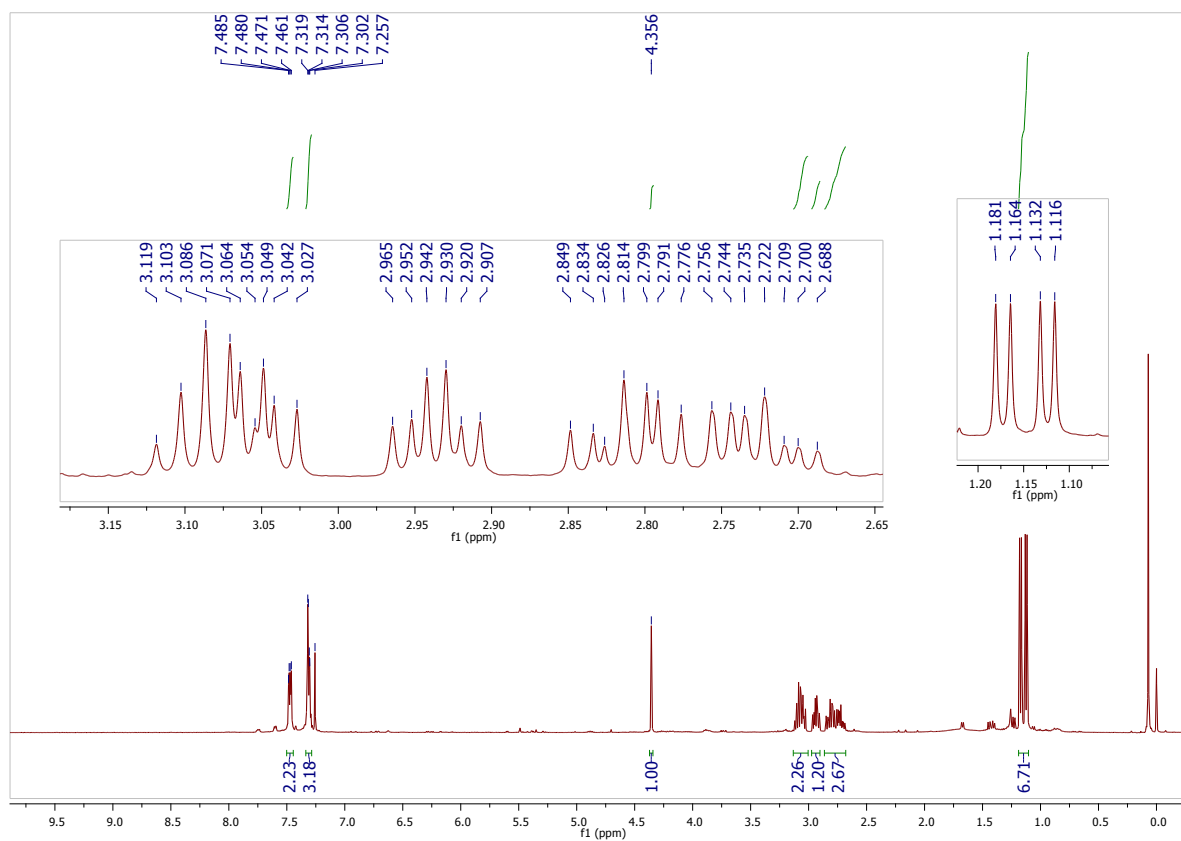
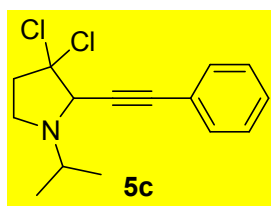


Figure 10. ¹H (400 MHz) and ¹³C (100 MHz) NMR spectra of **5b** in CDCl₃.

3,3-Dichloro-1-isopropyl-2-(phenylethynyl)pyrrolidine (5c)



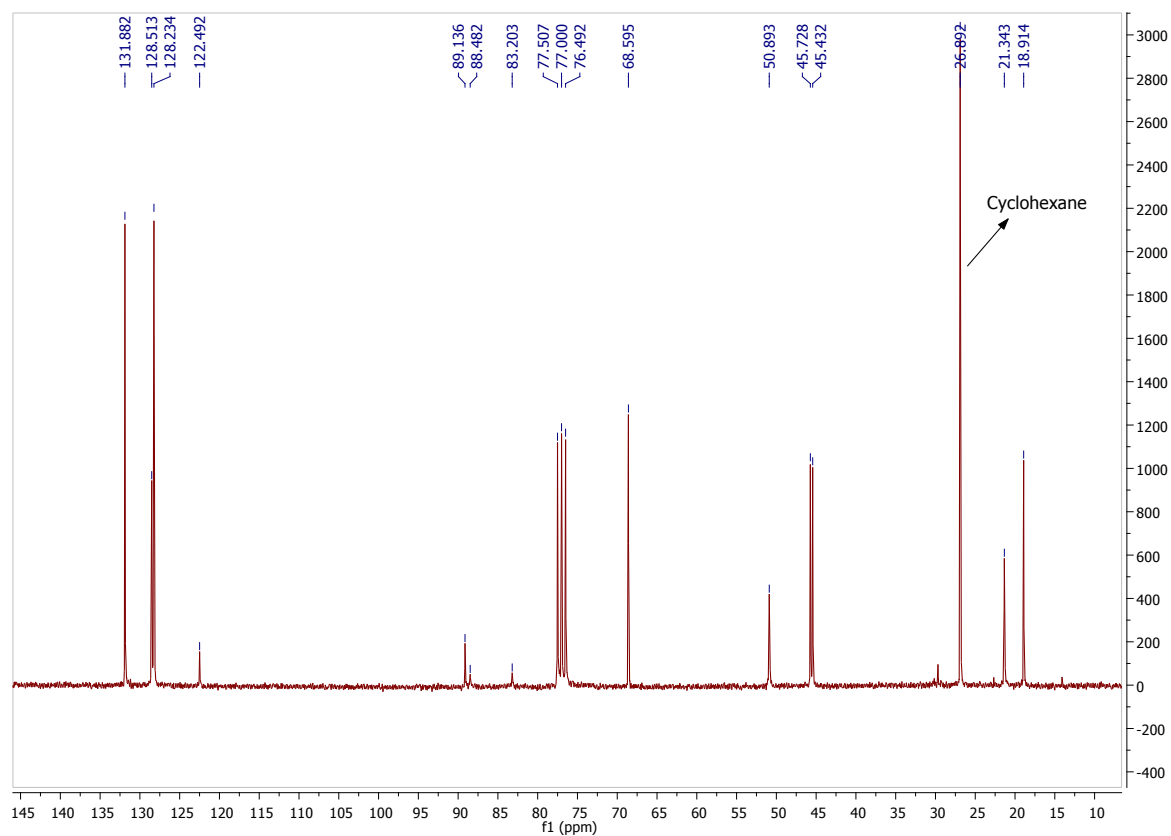
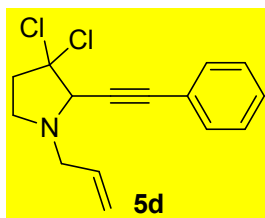


Figure 11. ^1H (400 MHz) and ^{13}C (100 MHz) NMR spectra of **5c** in CDCl_3 .

1-Allyl-3,3-dichloro-2-(phenylethynyl)pyrrolidine (5d**)**



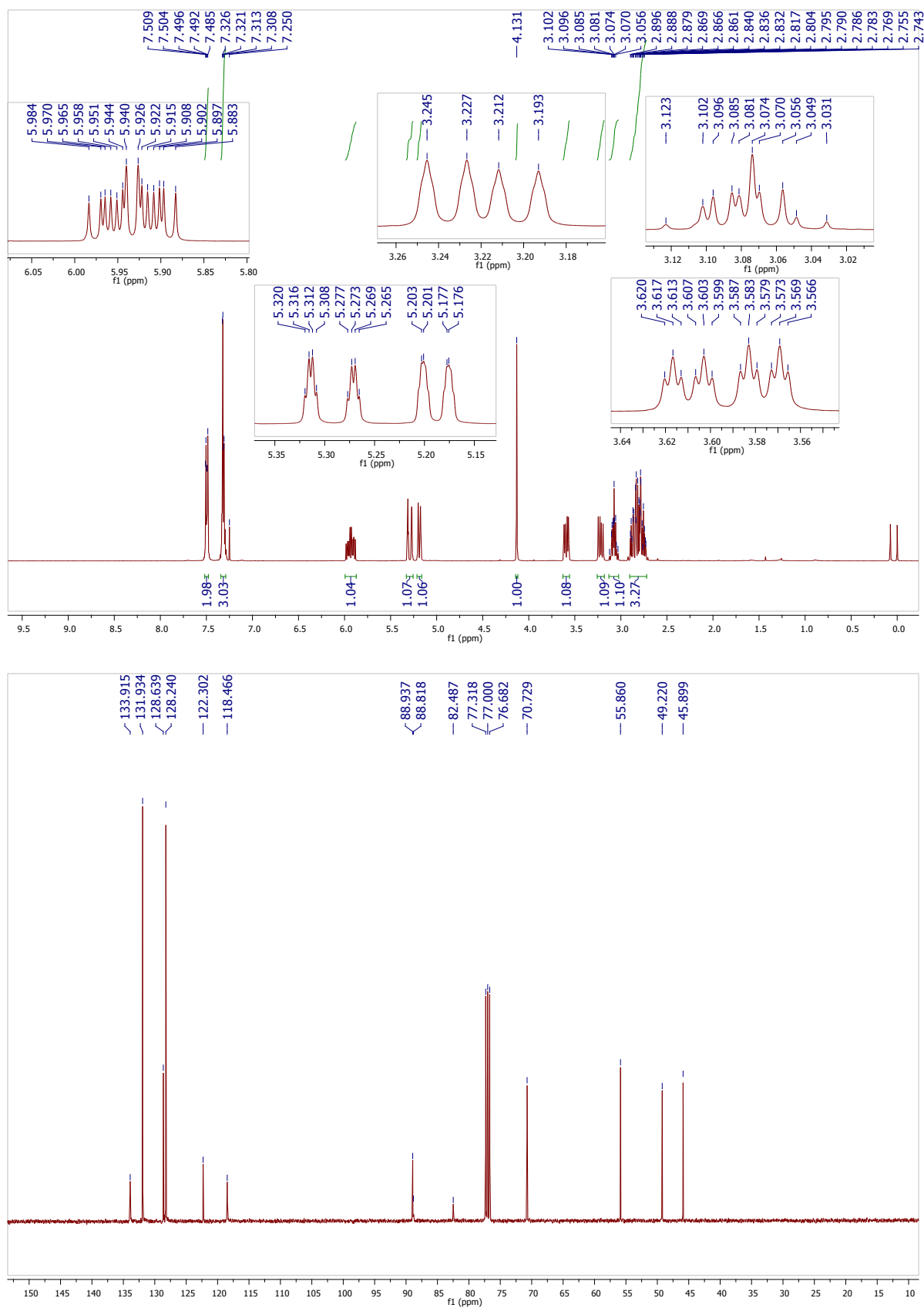
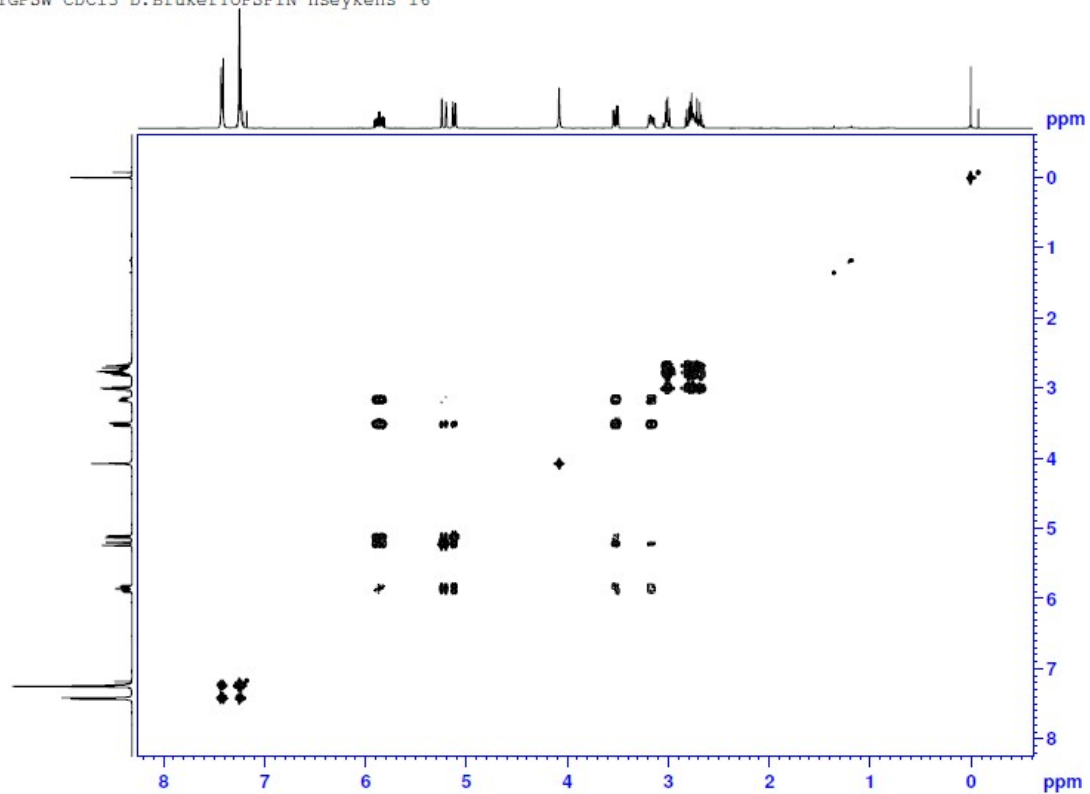
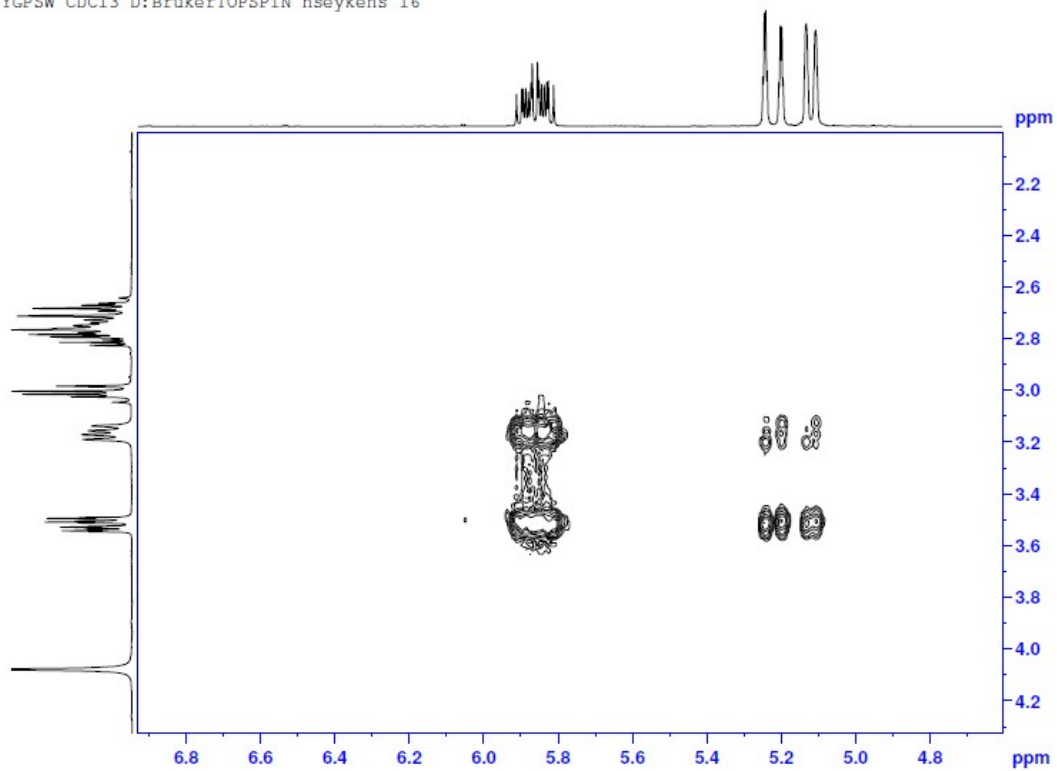


Figure 12. ¹H (400 MHz) and ¹³C (100 MHz) NMR spectra of **5d** in CDCl₃.

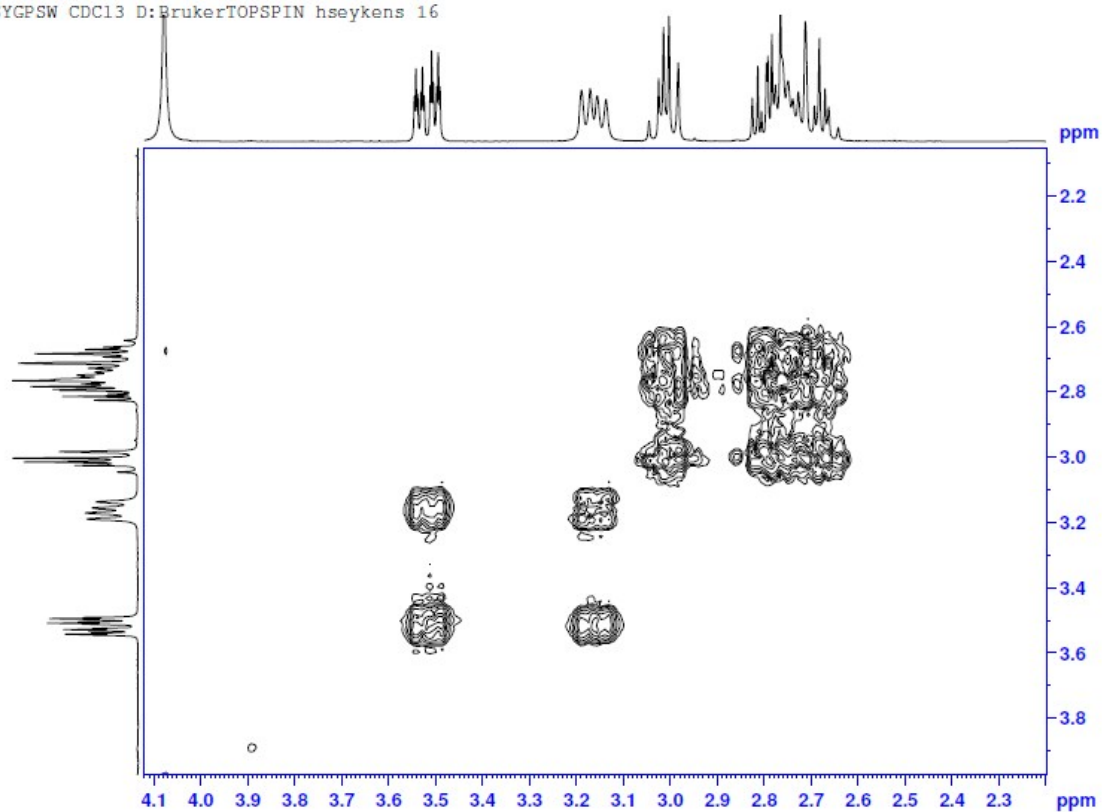
COSYGPSW CDC13 D:BrukerTOPSPIN hseykens 16



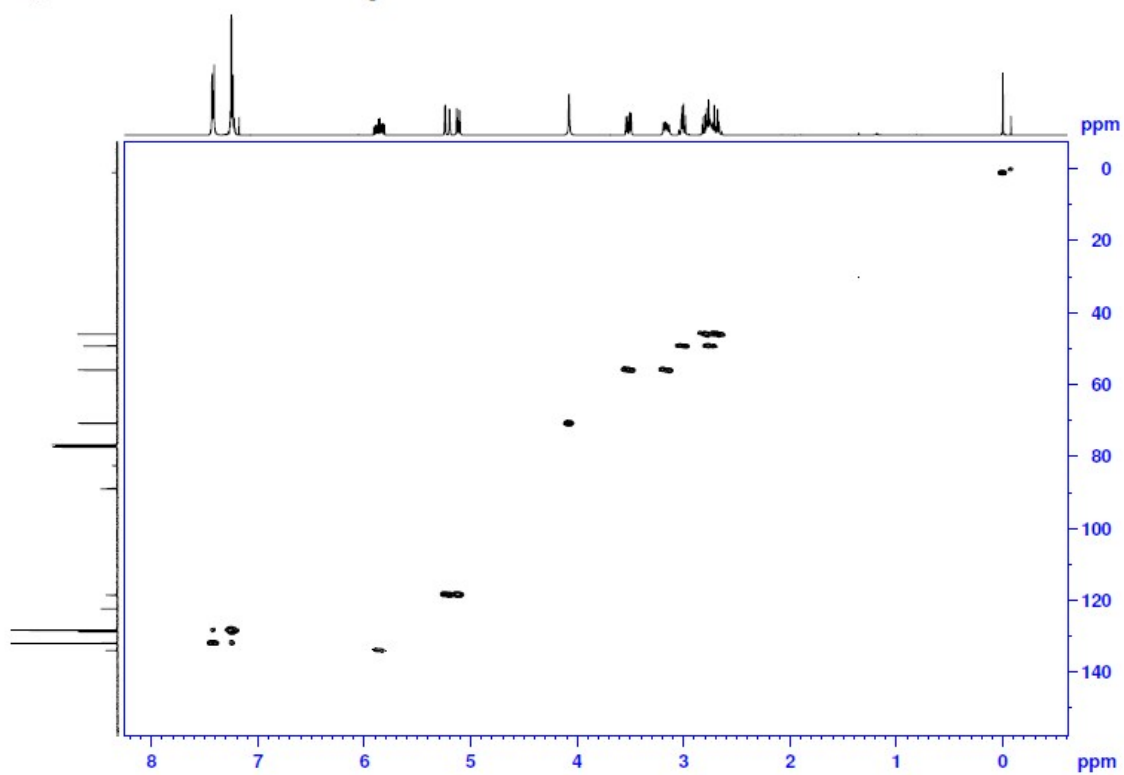
COSYGPSW CDC13 D:BrukerTOPSPIN hseykens 16



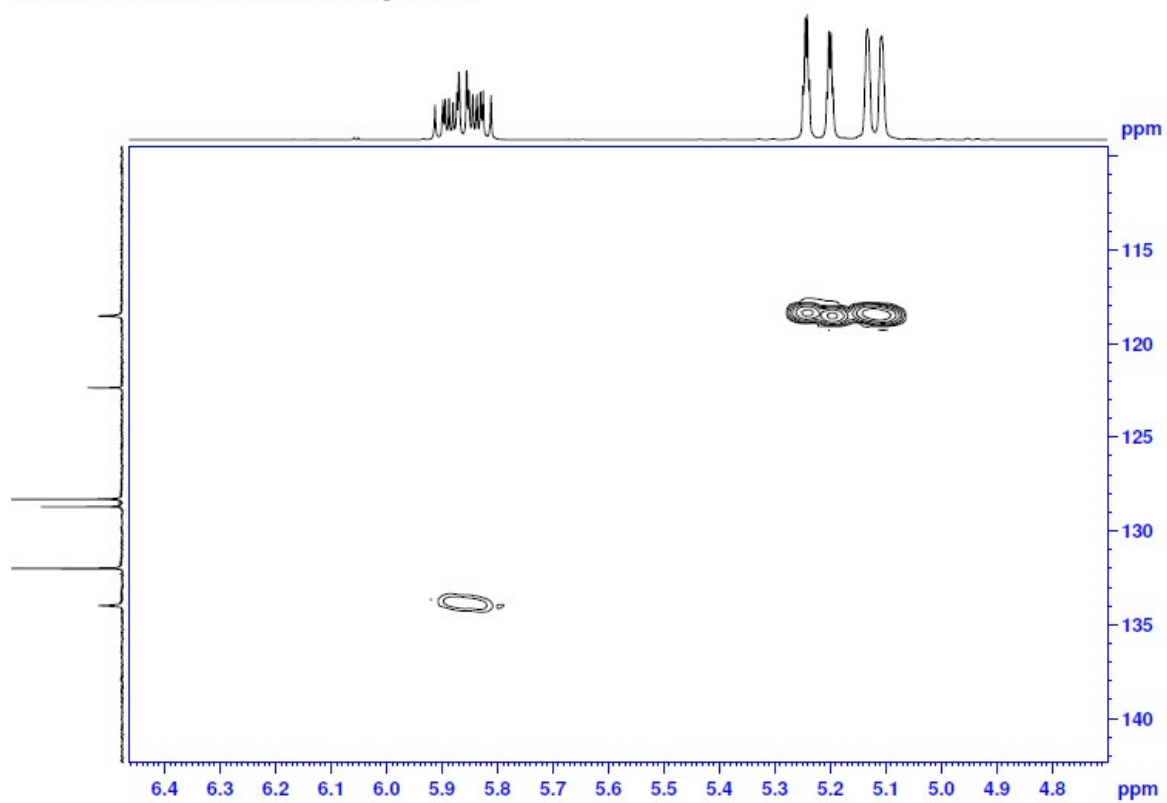
COSYGPSW CDC13 D:BrukerTOPSPIN hseykens 16



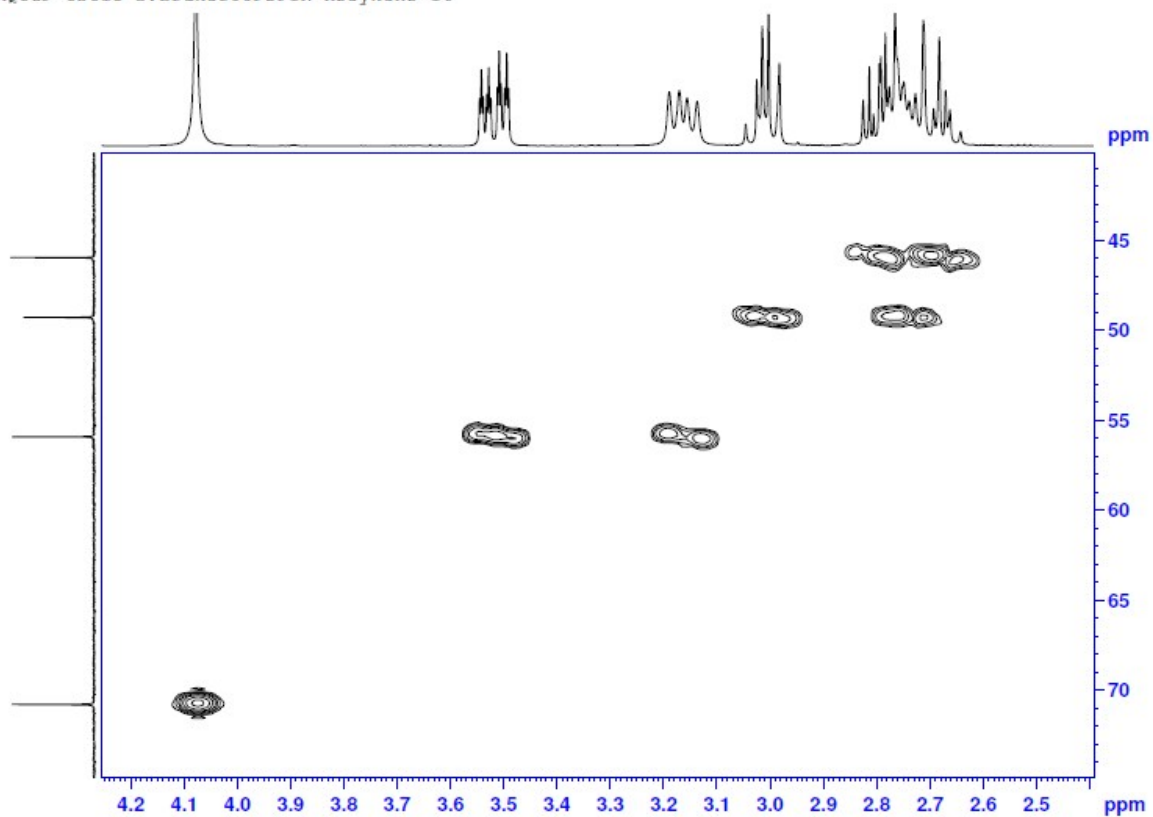
HMQCGP CDC13 D:BrukerTOPSPIN hseykens 16



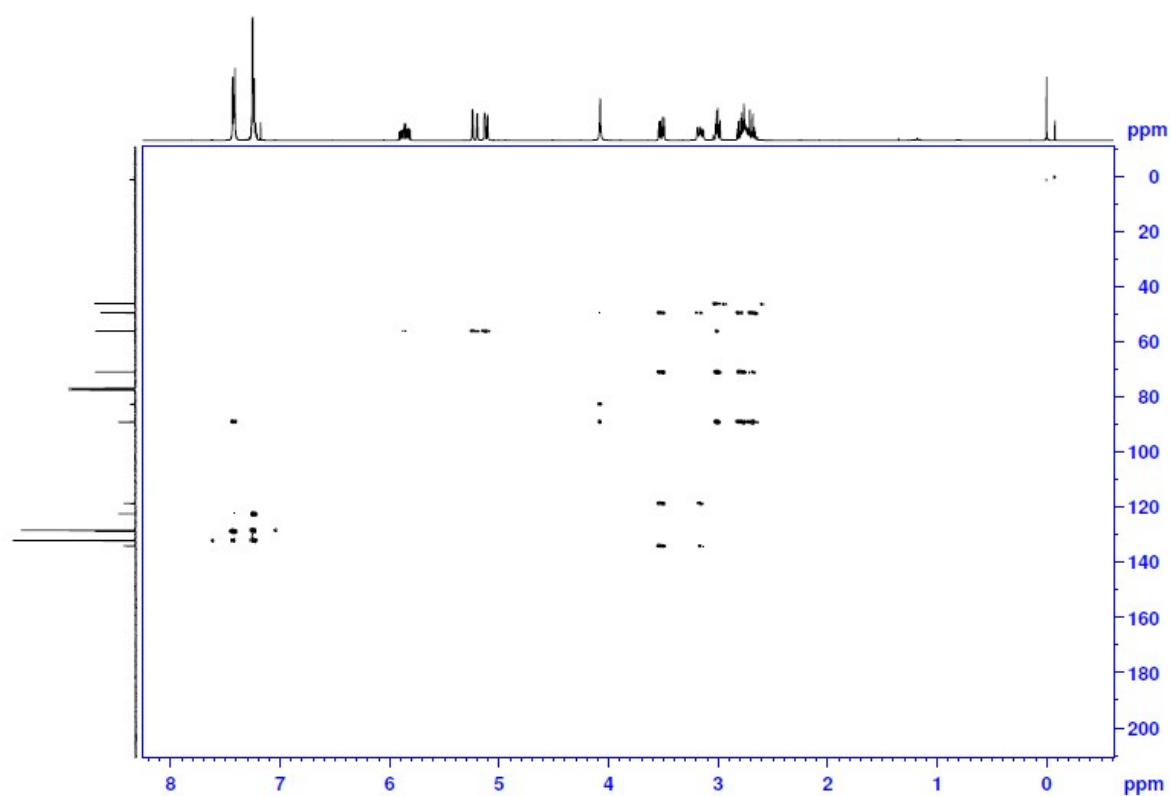
HMQCGP CDC13 D:BrukerTOPSPIN hseykens 16



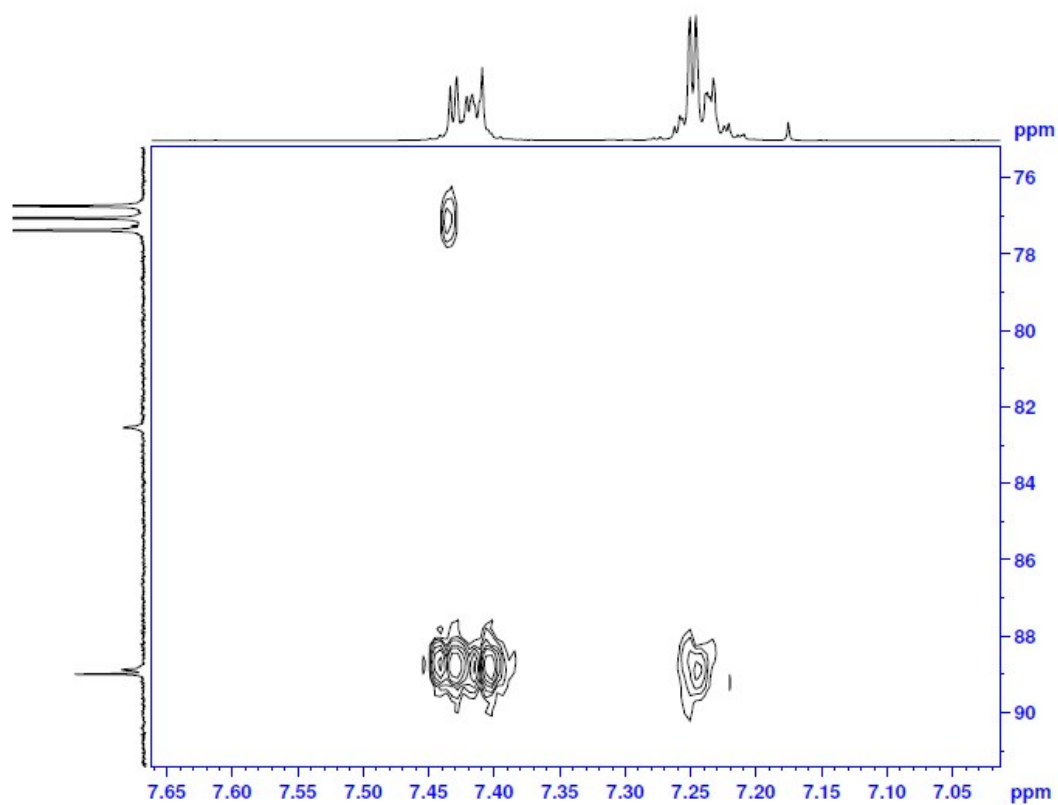
HMQCGP CDC13 D:BrukerTOPSPIN hseykens 16



HMBCGP CDC13 D:BrukerTOPSPIN hseykens 16



HMBCGP CDC13 D:BrukerTOPSPIN hseykens 16



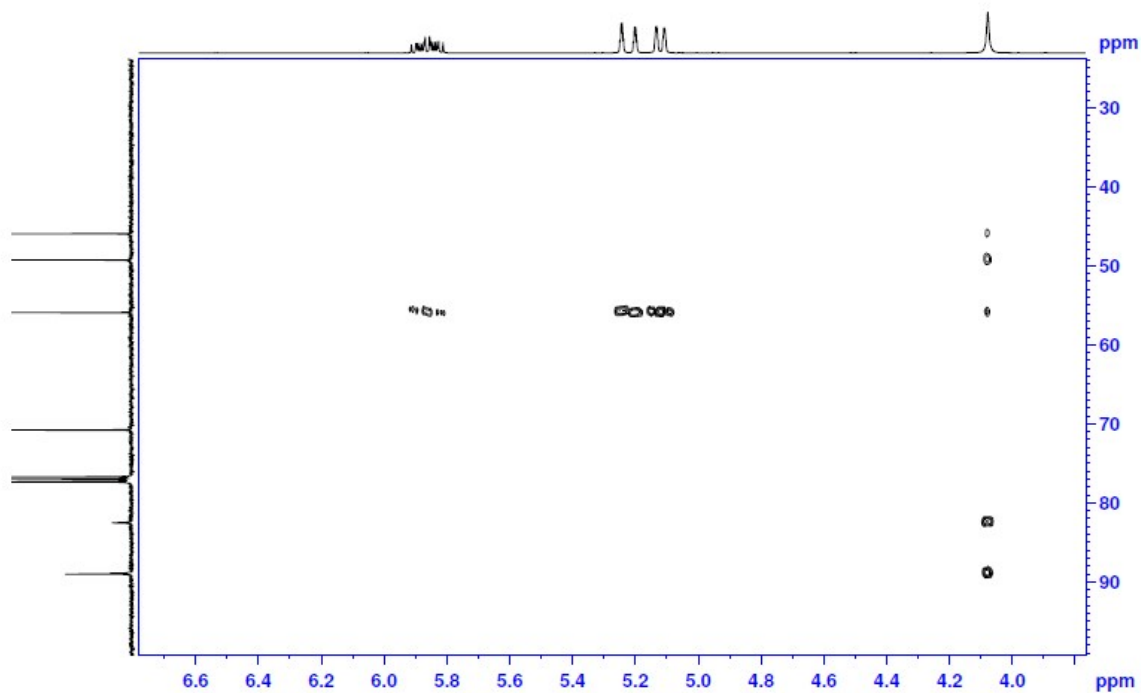
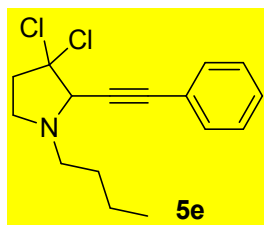


Figure 13. COSY, HMQC and HMBC spectra of **5d** in CDCl₃.

1-Butyl-3,3-dichloro-2-(phenylethynyl)pyrrolidine (5e**)**



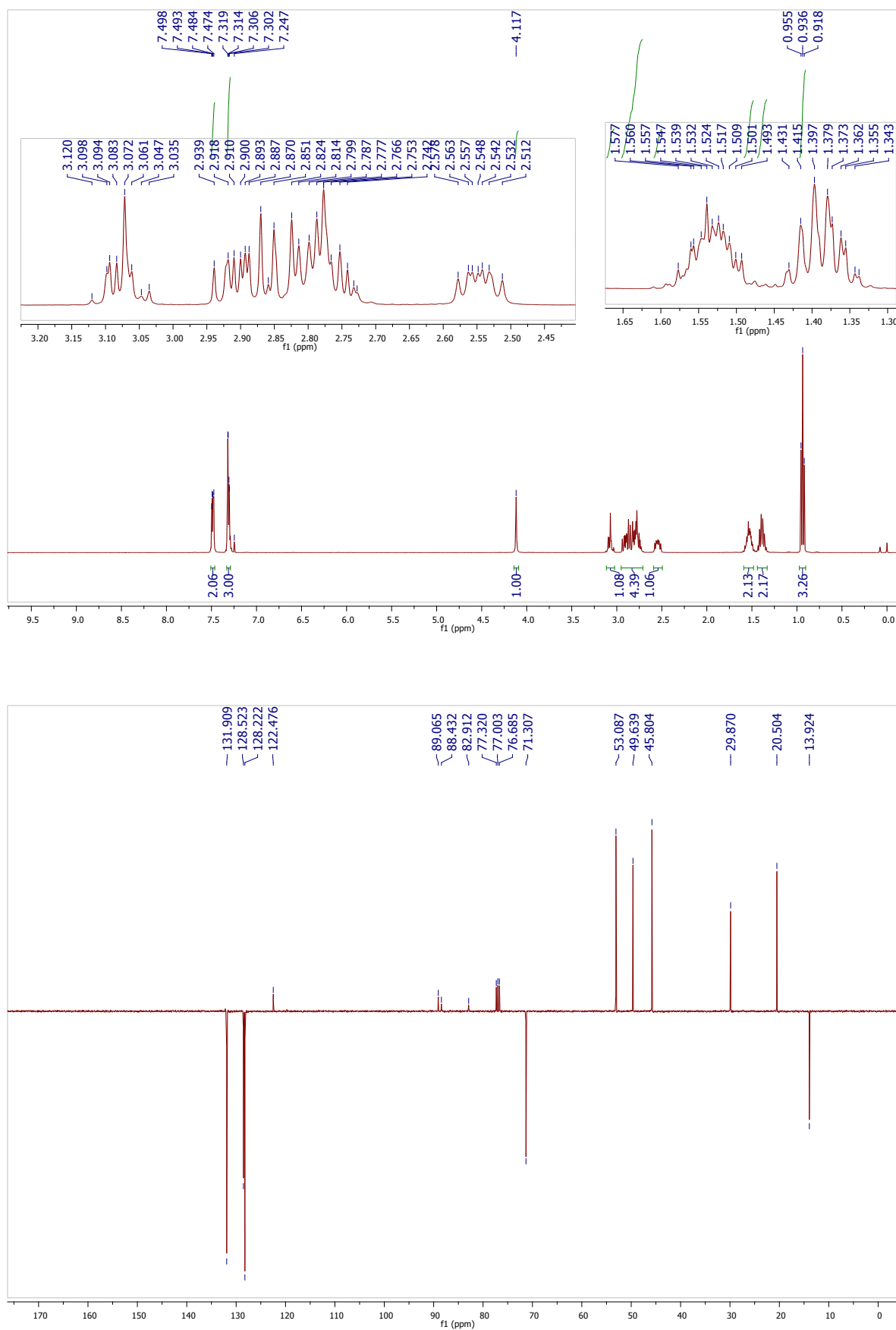
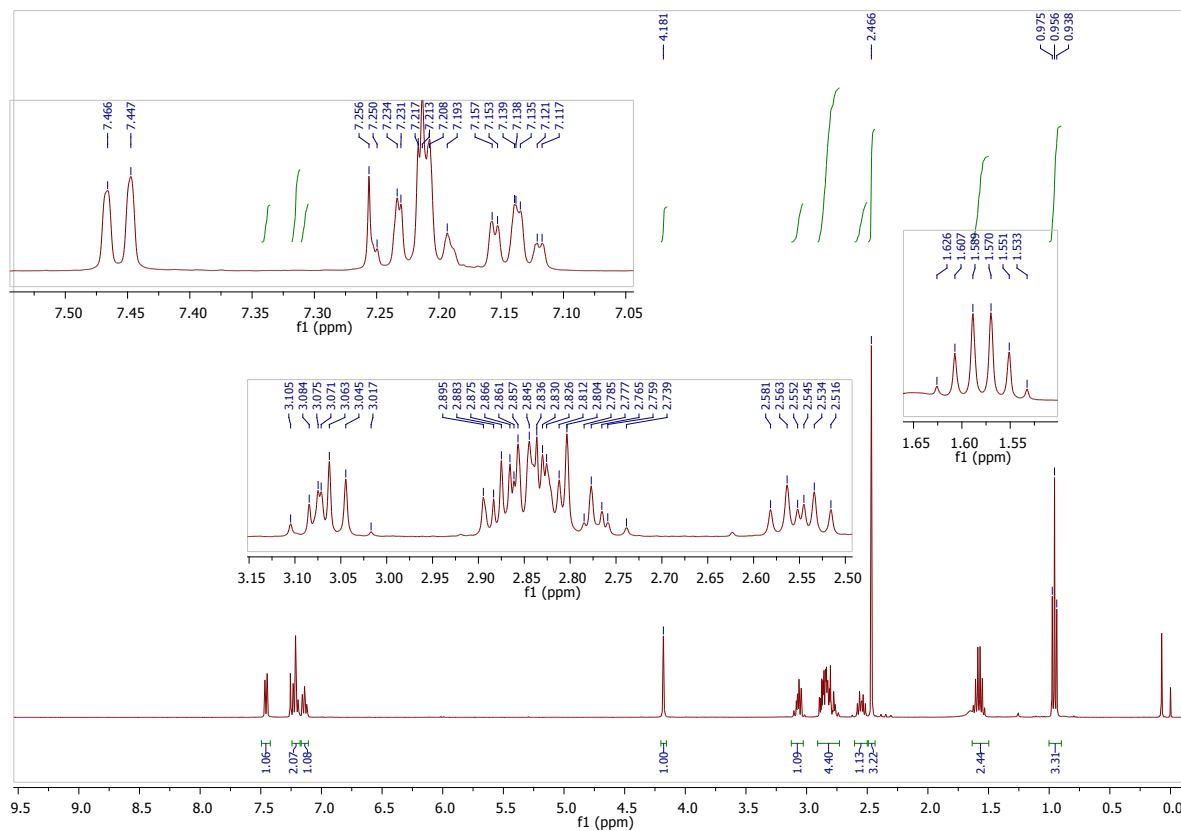
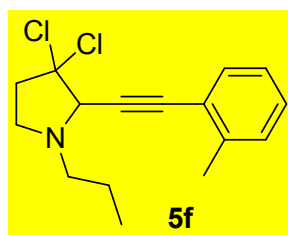


Figure 14. ¹H (400 MHz) and ¹³C (100 MHz) NMR spectra of **5e** in CDCl₃.

3,3-Dichloro-1-propyl-2-(*ortho*-tolylethynyl)pyrrolidine (**5f**)



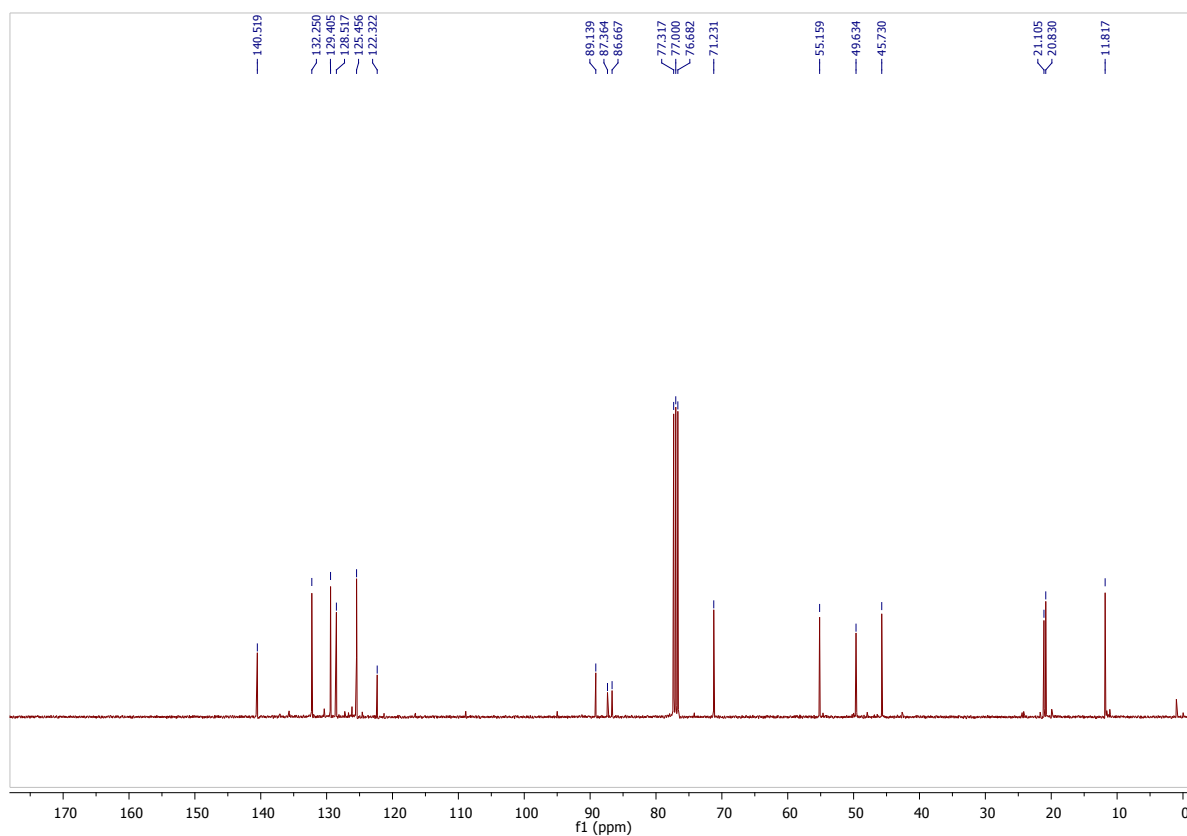
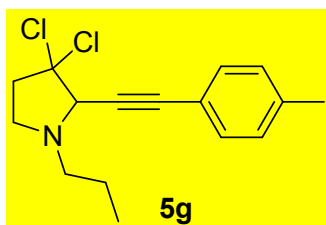


Figure 15. ^1H (400 MHz) and ^{13}C (100 MHz) NMR spectra of **5f** in CDCl_3 .

3,3-Dichloro-1-propyl-2-(*para*-tolylethynyl)pyrrolidine (5g**)**



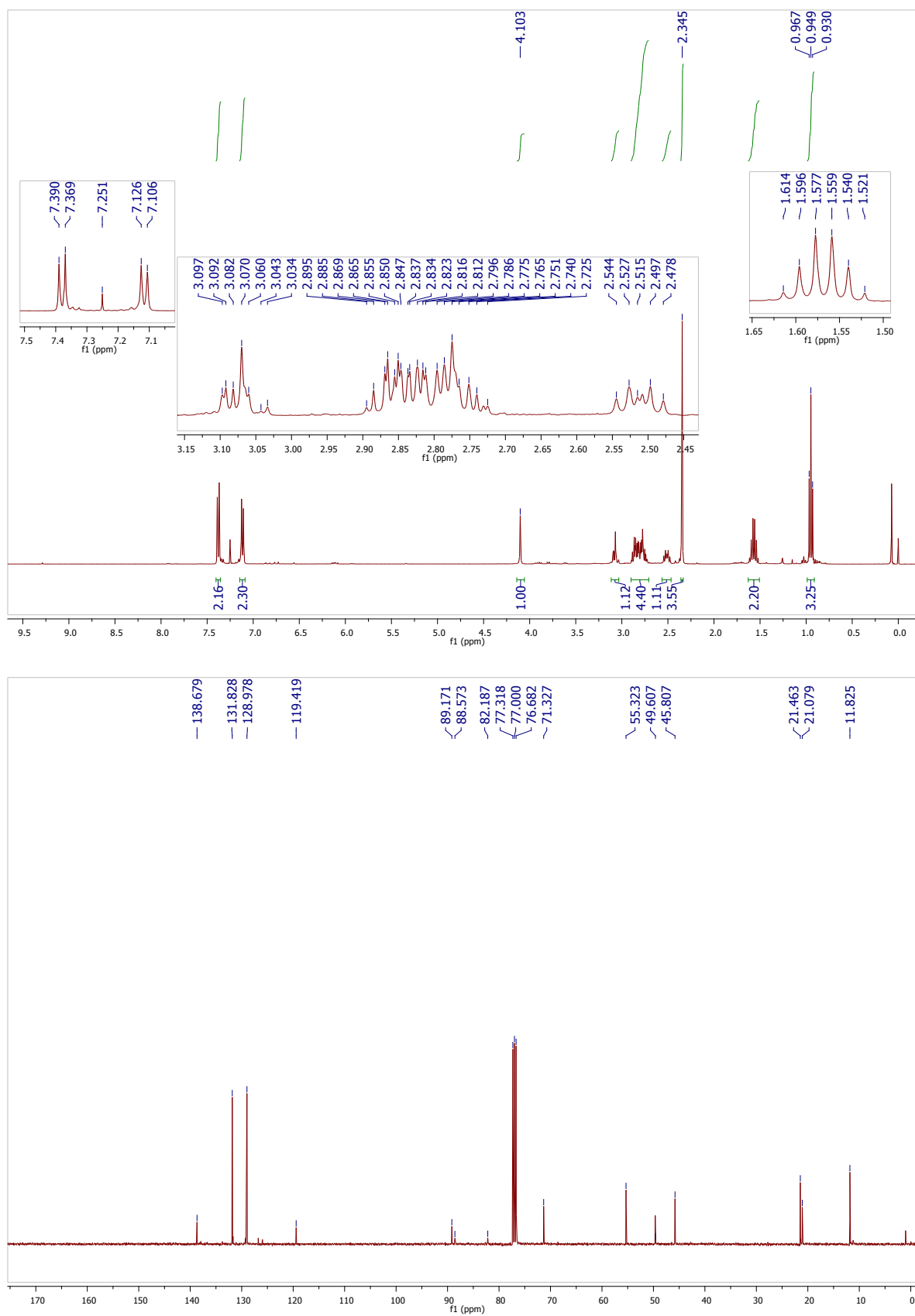
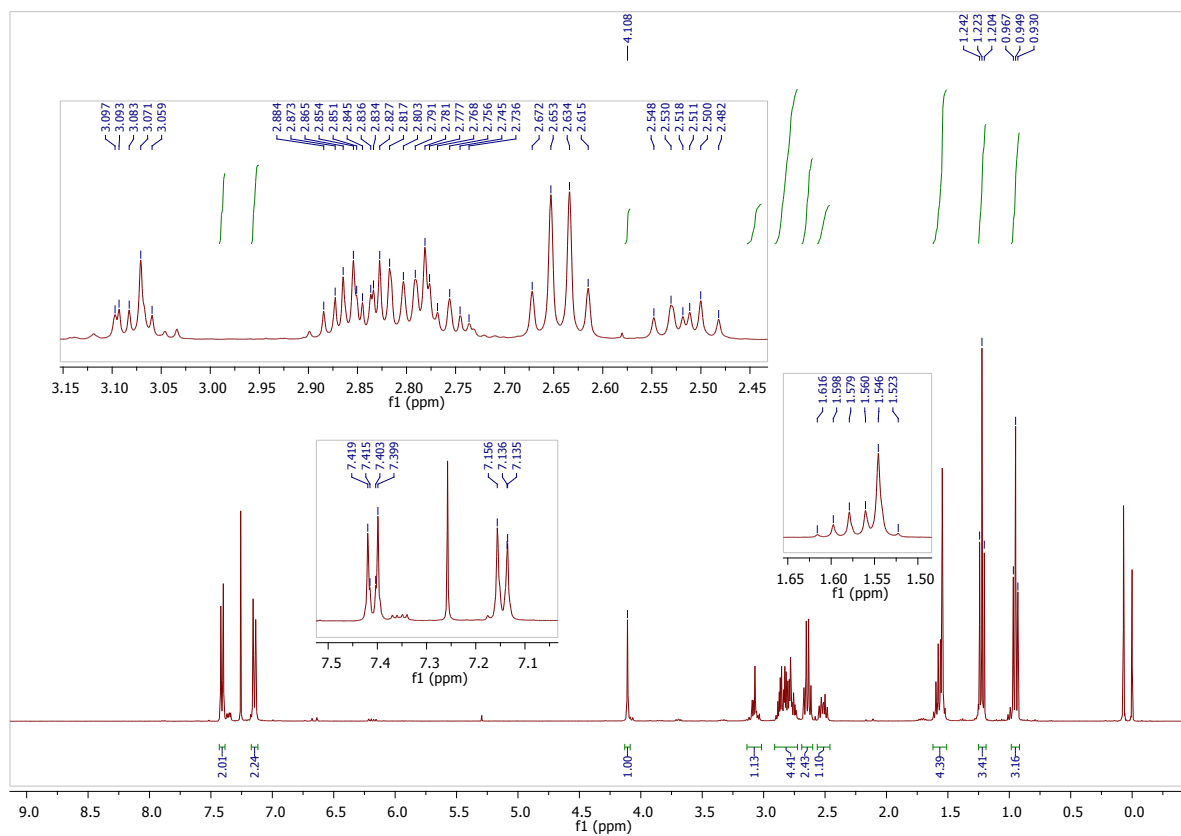
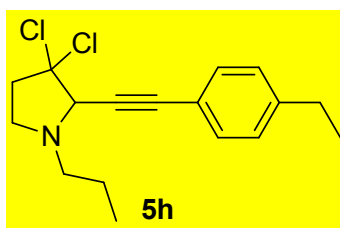


Figure 16. ¹H (400 MHz) and ¹³C (100 MHz) NMR spectra of **5g** in CDCl₃.

3,3-Dichloro-2-((4-ethylphenyl)ethynyl)-1-propylpyrrolidine (5h)



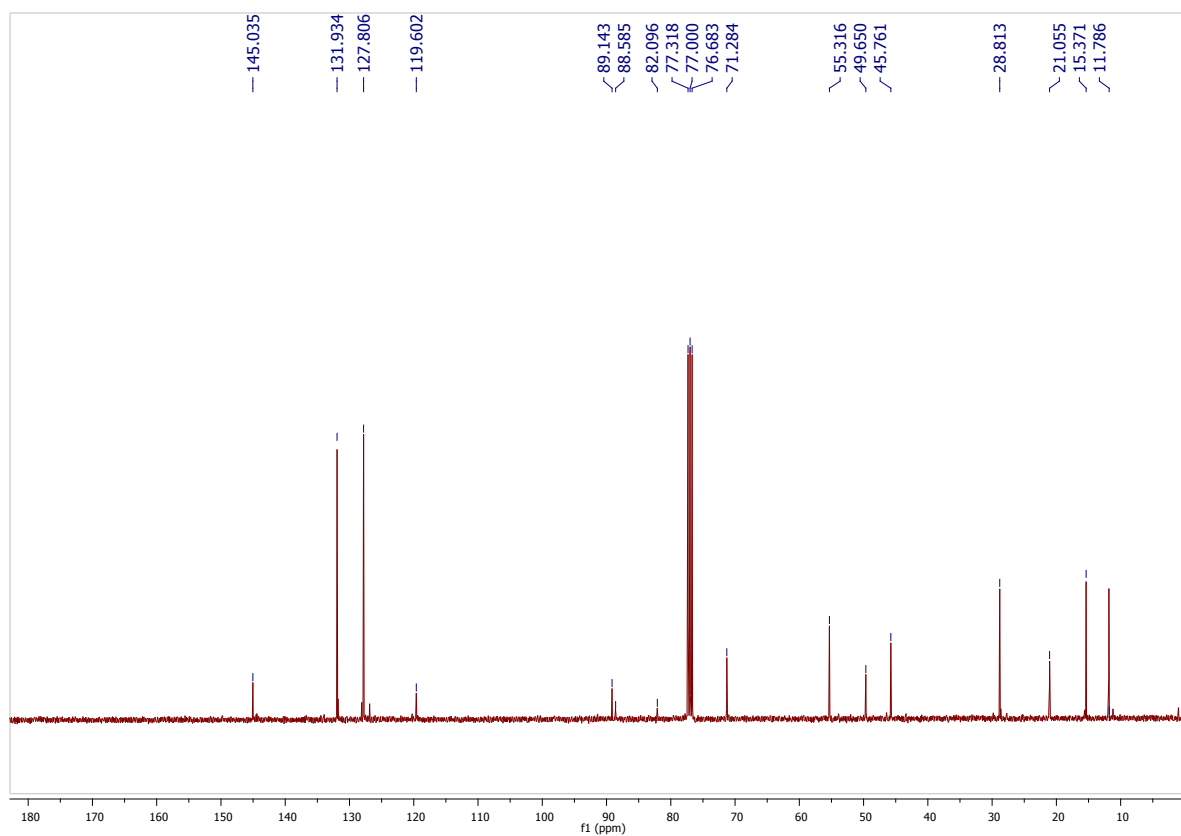
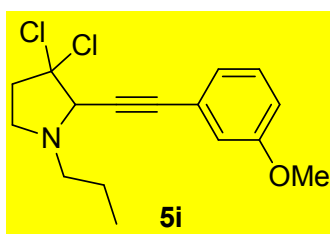


Figure 17. ^1H (400 MHz) and ^{13}C (100 MHz) NMR spectra of **5h** in CDCl_3 .

3,3-Dichloro-2-((3-methoxyphenyl)ethynyl)-1-propylpyrrolidine (**5i**)



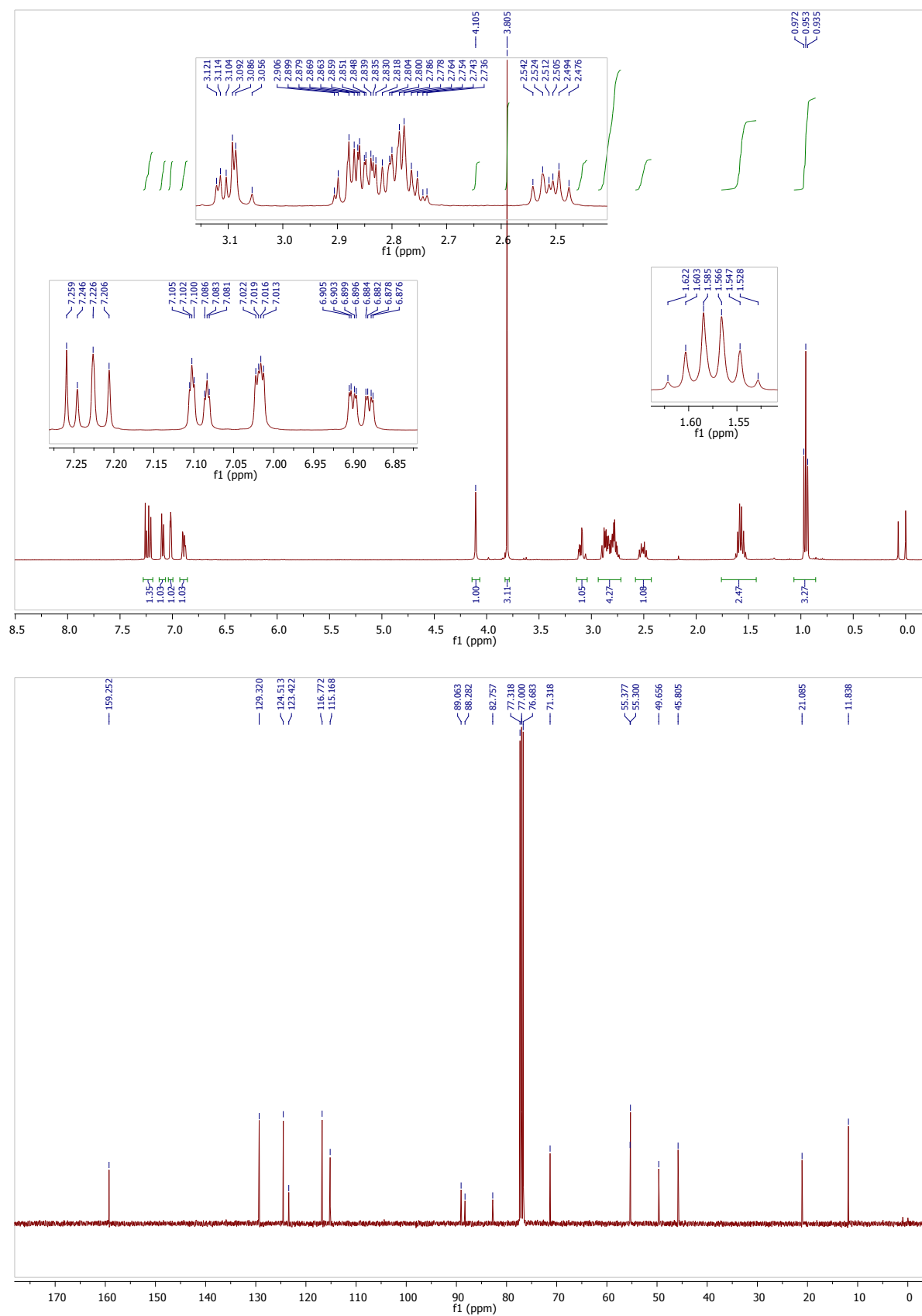
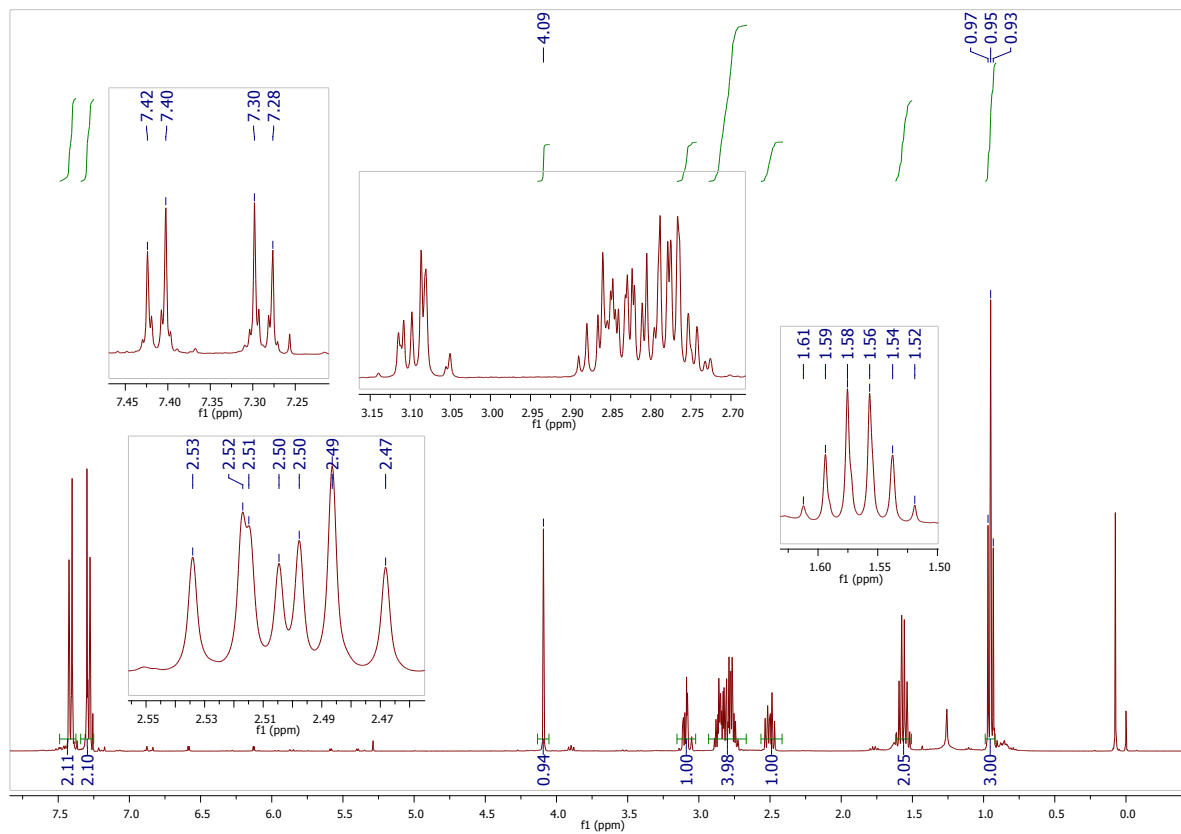
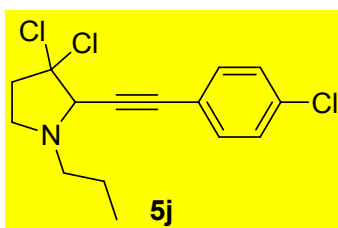


Figure 18. ¹H (400 MHz) and ¹³C (100 MHz) NMR spectra of **5i** in CDCl₃.

3,3-Dichloro-2-((4-chlorophenyl)ethynyl)-1-propylpyrrolidine (5j)



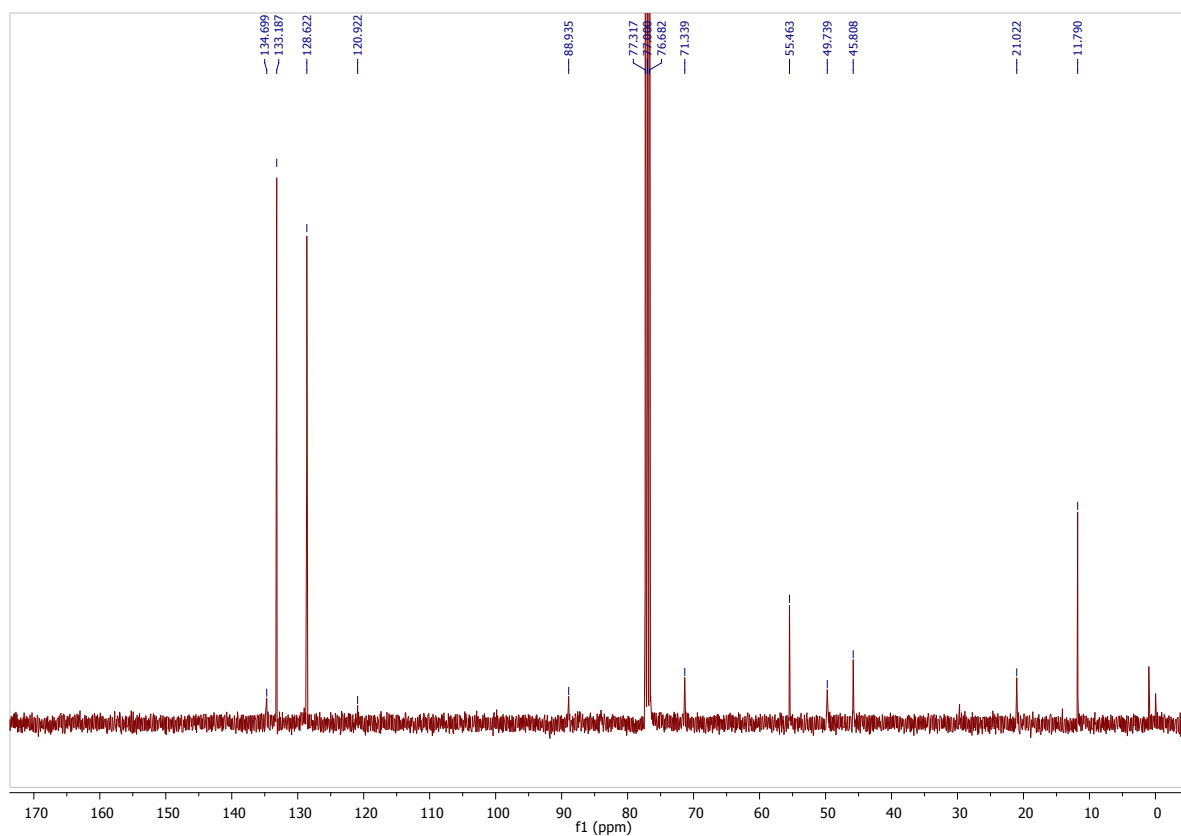
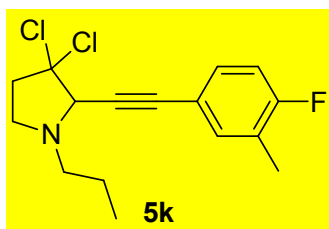


Figure 19. ^1H (400 MHz) and ^{13}C (100 MHz) NMR spectra of **5j** in CDCl_3 .

3,3-Dichloro-2-((4-fluoro-3-methylphenyl)ethynyl)-1-propylpyrrolidine (5k**)**



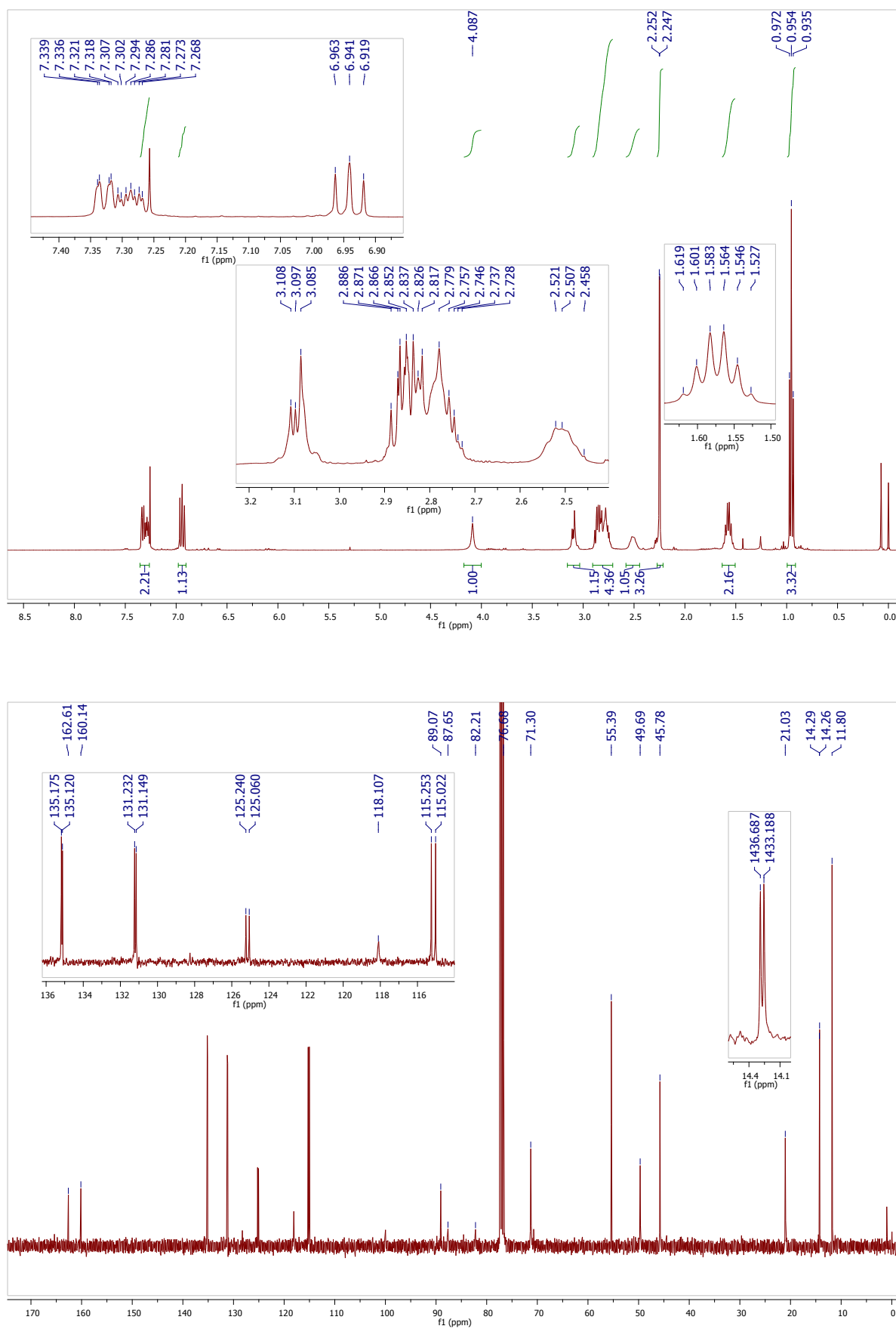
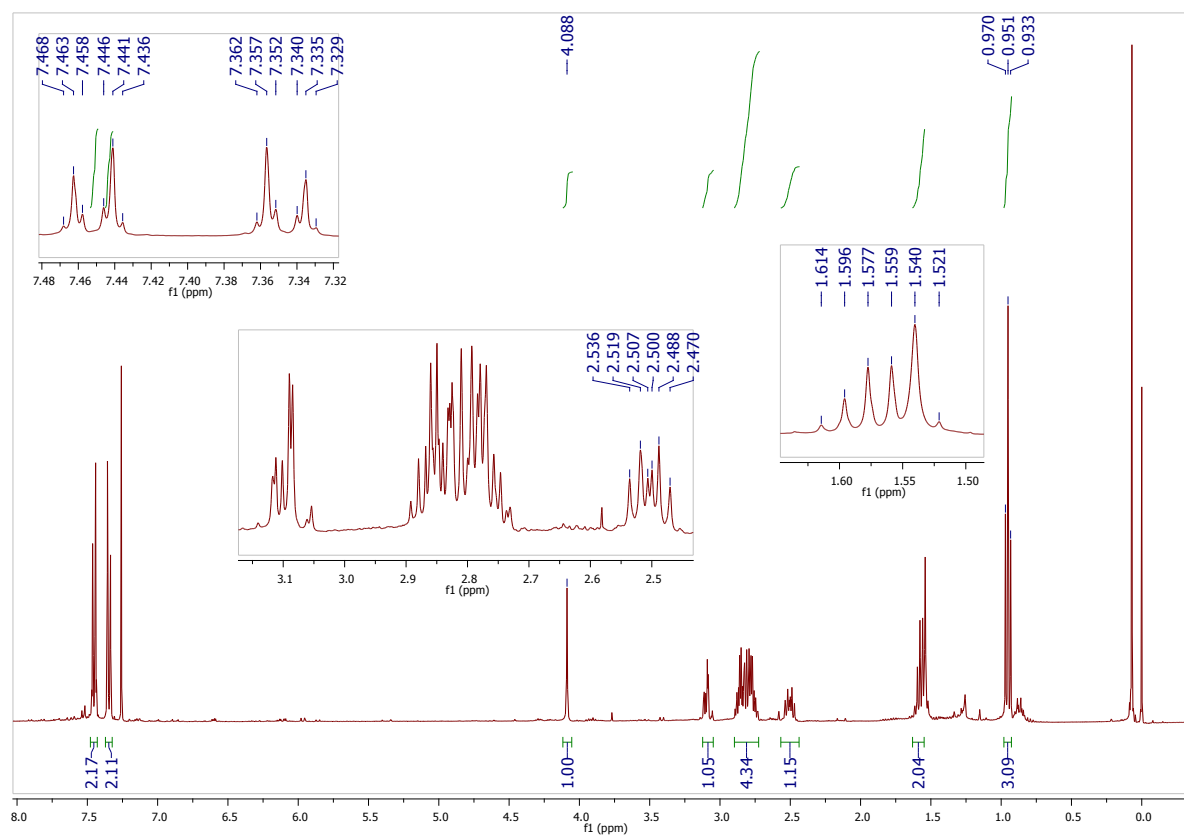
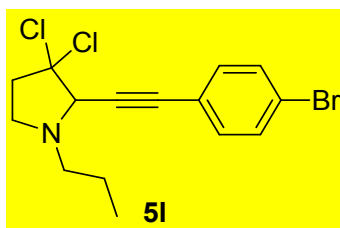


Figure 20. ¹H (400 MHz) and ¹³C (100 MHz) NMR spectra of **5k** in CDCl₃.

3,3-Dichloro-2-((4-bromophenyl)ethynyl)-1-propylpyrrolidine (**5l**)



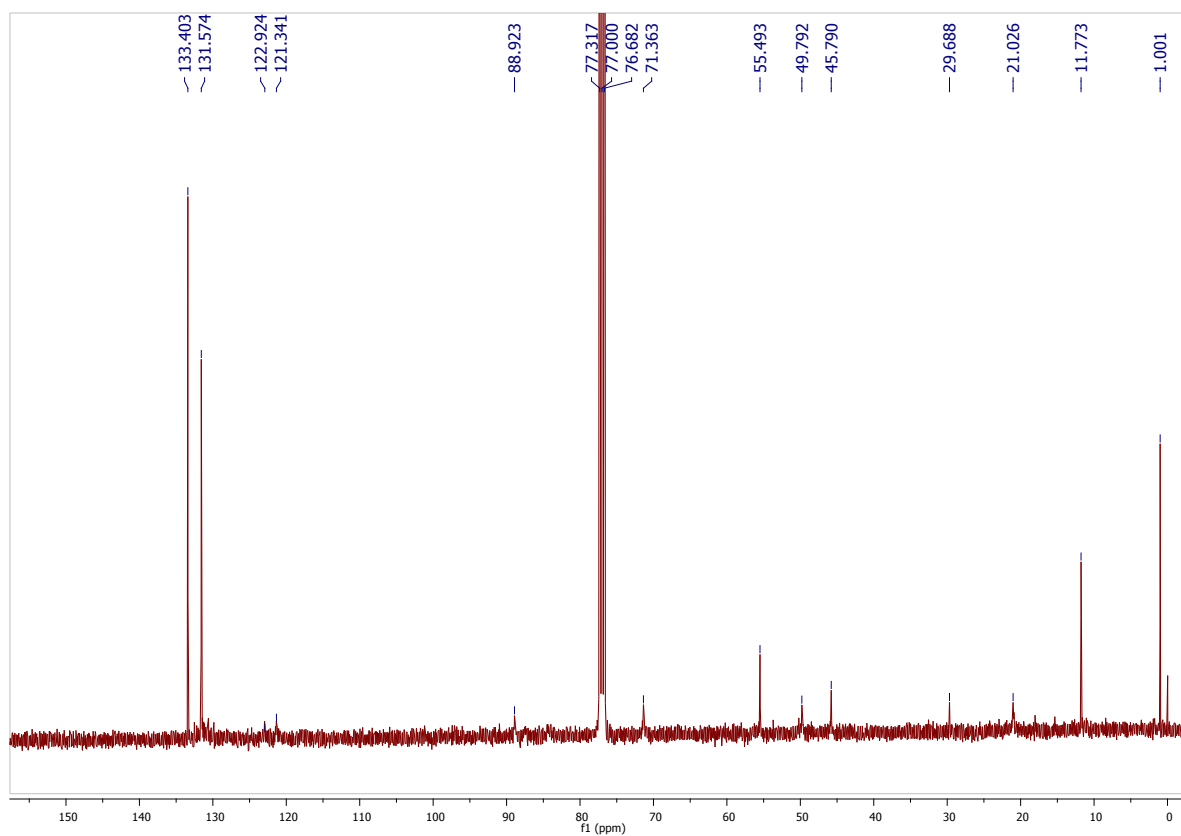
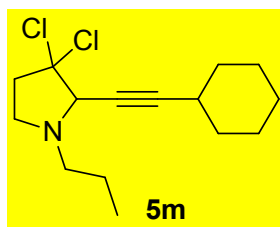


Figure 21. ^1H (400 MHz) and ^{13}C (100 MHz) NMR spectra of **5l** in CDCl_3 .

3,3-Dichloro-2-(cyclohexylethynyl)-1-propylpyrrolidine (**5m**)



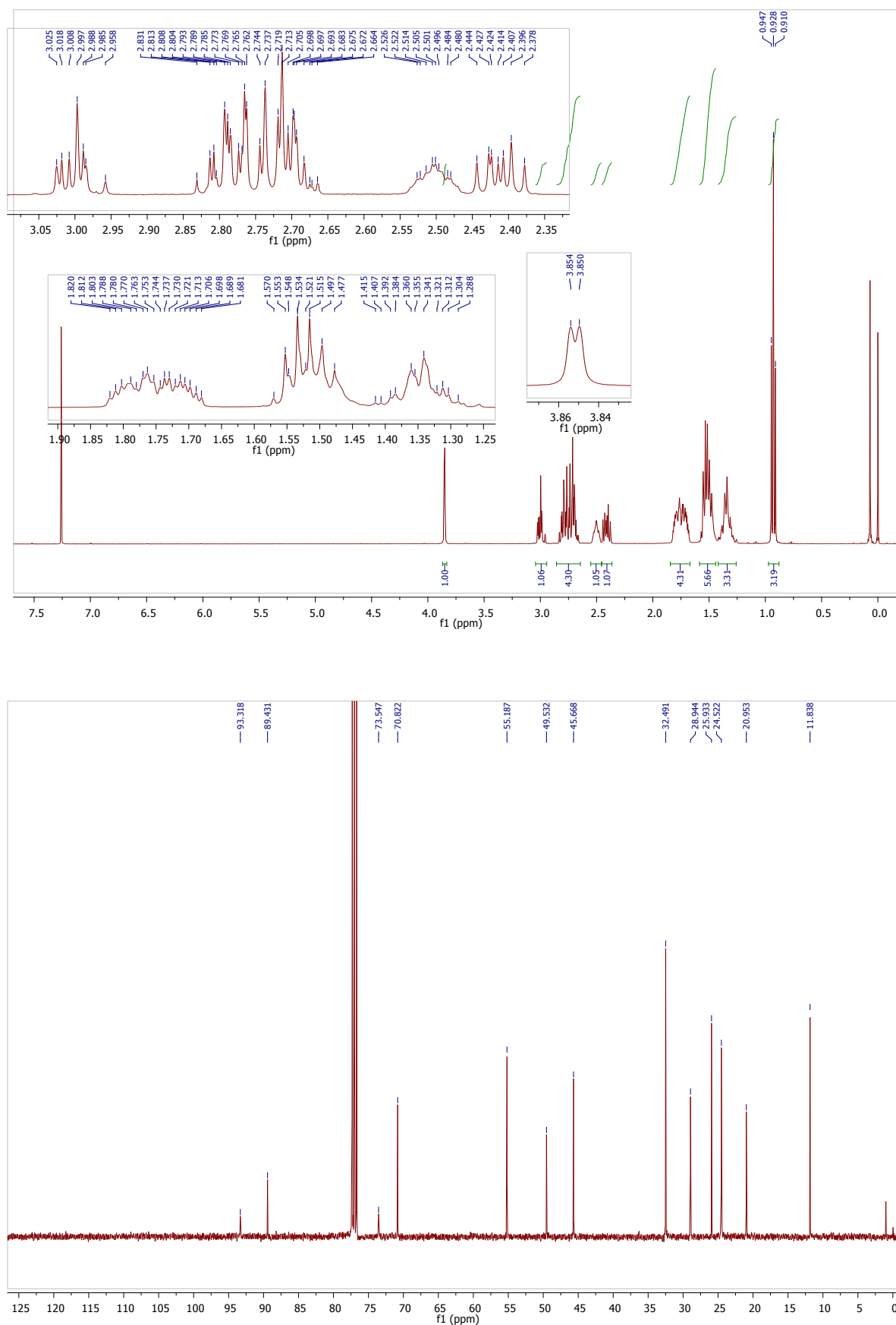
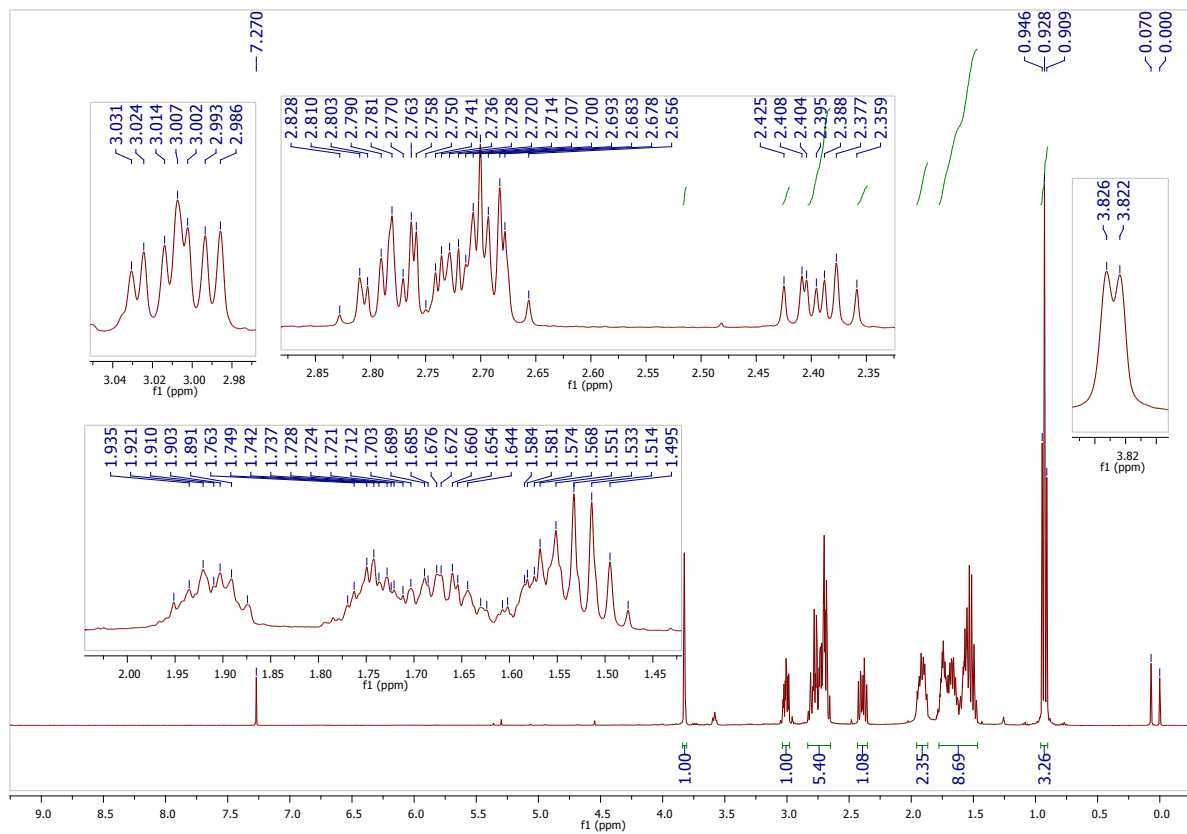
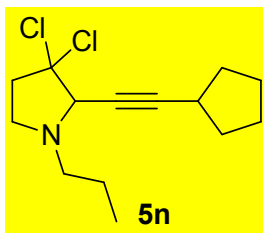


Figure 22. ¹H (400 MHz) and ¹³C (100 MHz) NMR spectra of **5m** in CDCl₃.

3,3-Dichloro-2-(cyclopentylethynyl)-1-propylpyrrolidine (5n)



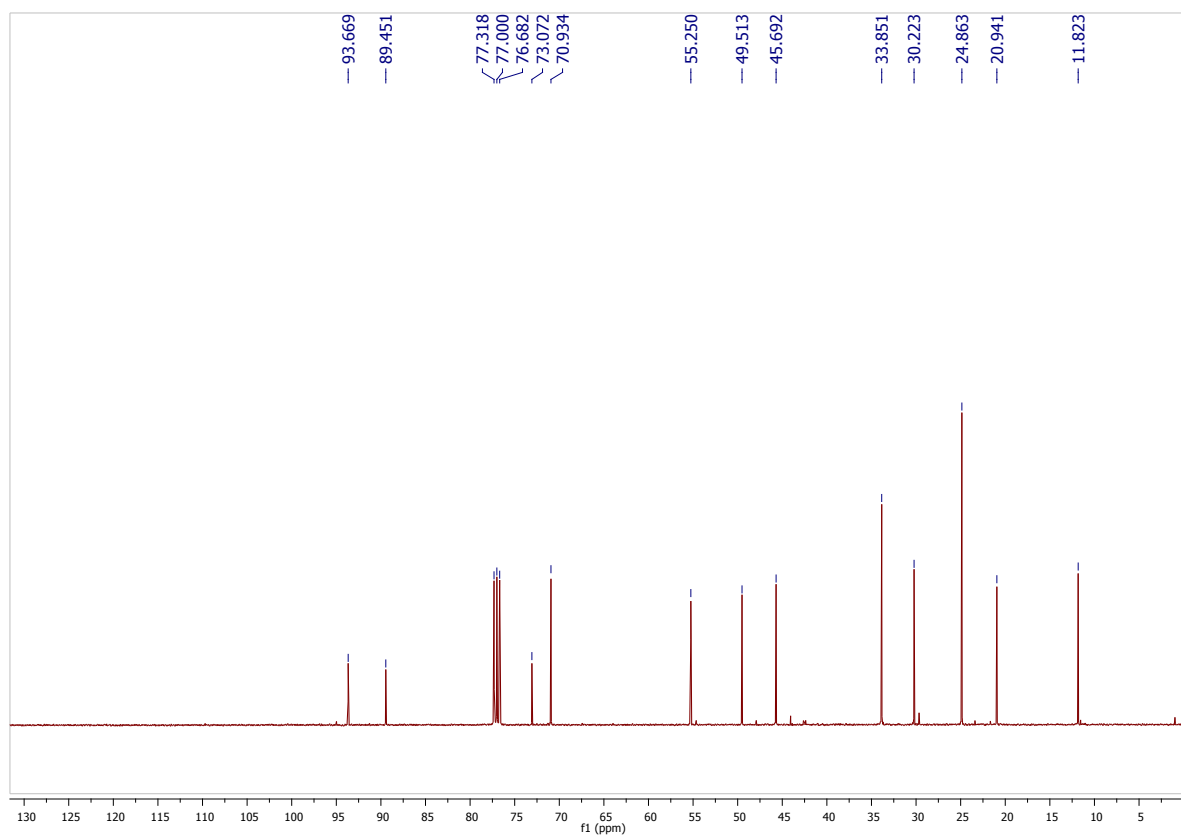
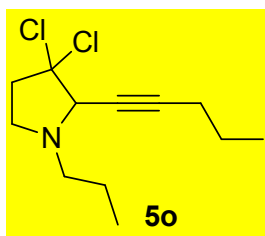


Figure 23. ^1H (400 MHz) and ^{13}C (100 MHz) NMR spectra of **5n** in CDCl_3 .

3,3-Dichloro-2-(pent-1-ynyl)-1-propylpyrrolidine (**5o**)



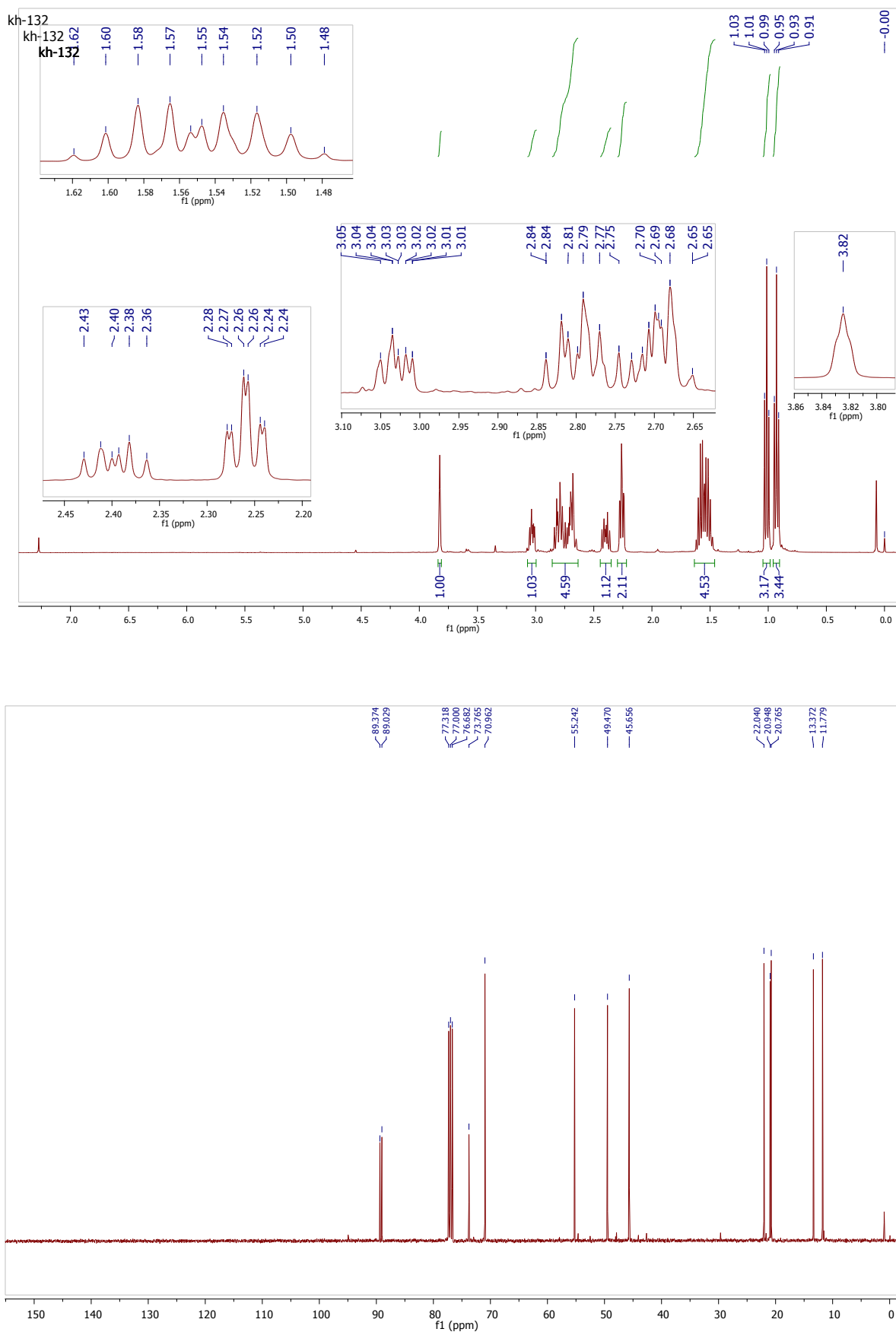
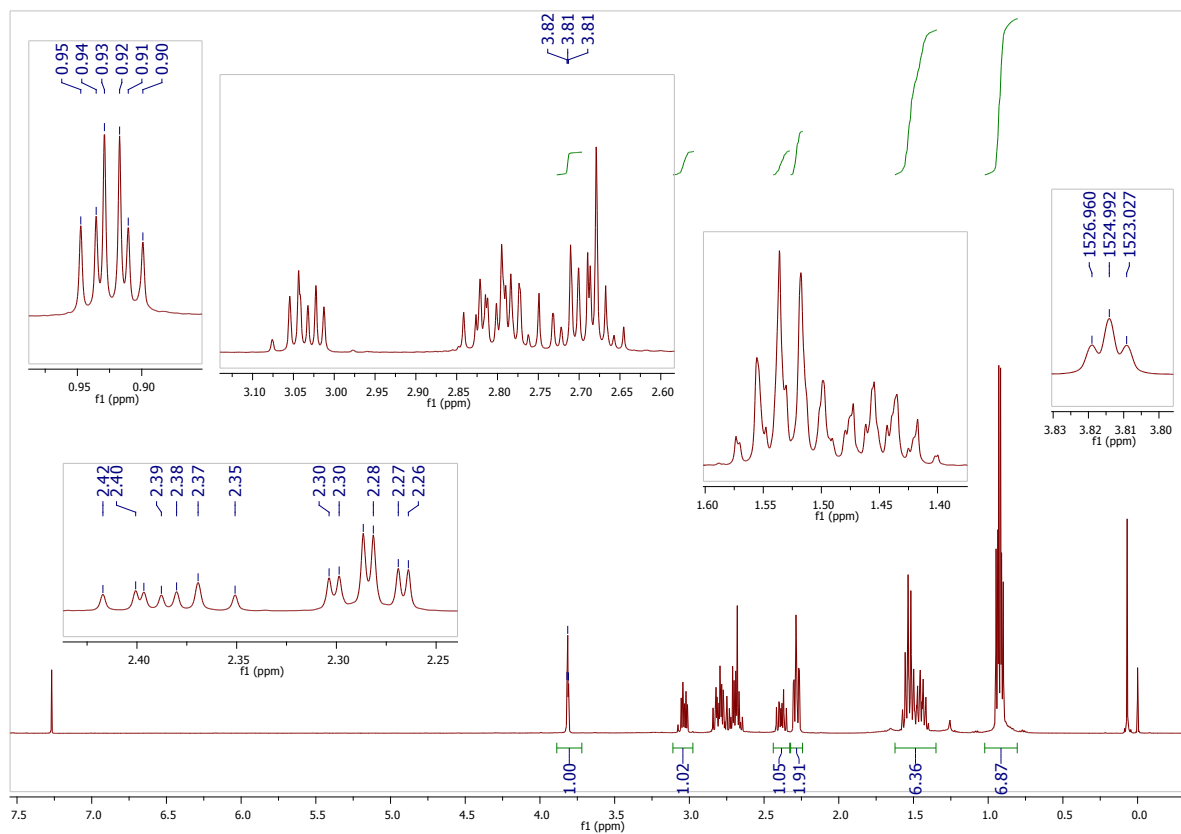
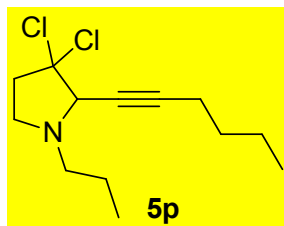


Figure 24. ^1H (400 MHz) and ^{13}C (100 MHz) NMR spectra of **5o** in CDCl_3 .

3,3-Dichloro-2-(hex-1-ynyl)-1-propylpyrrolidine (5p)



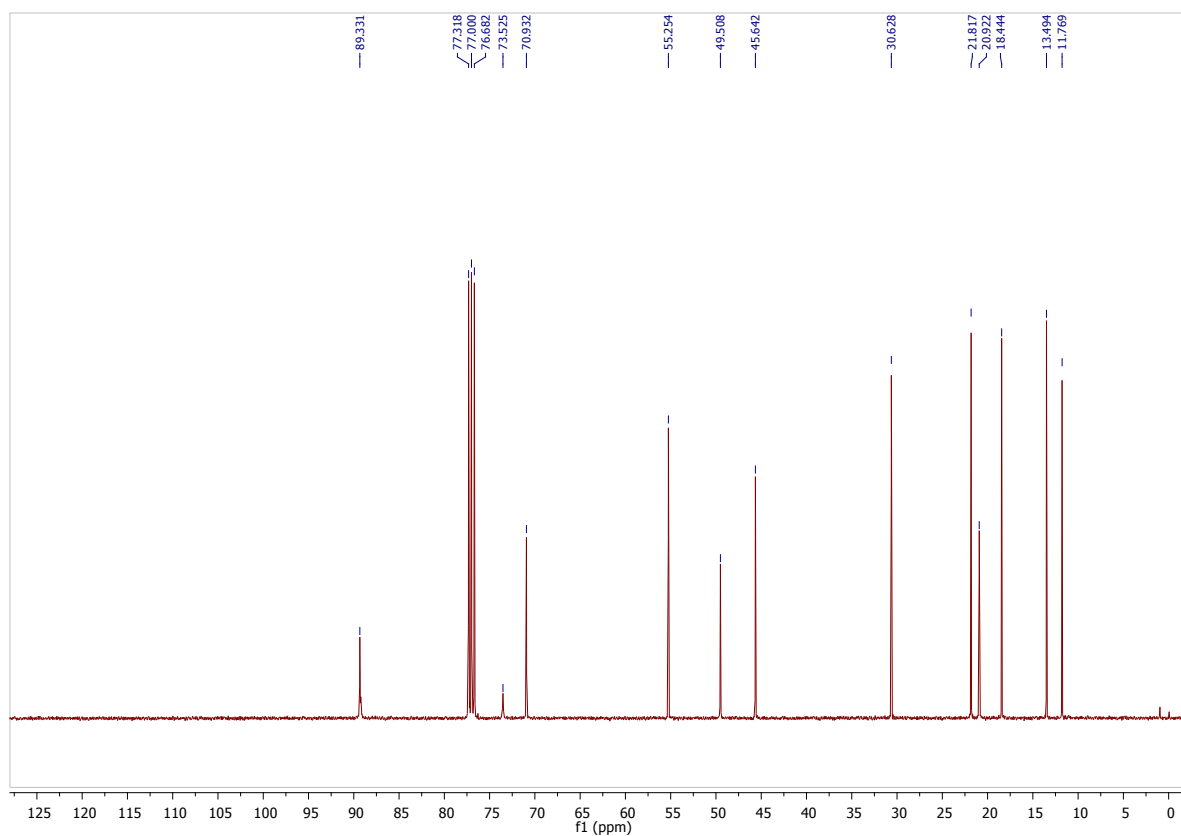
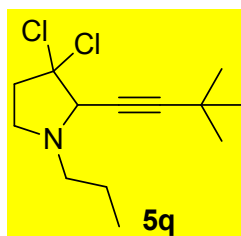


Figure 25. ^1H (400 MHz) and ^{13}C (100 MHz) NMR spectra of **5p** in CDCl_3 .

3,3-Dichloro-2-(3,3-dimethylbut-1-ynyl)-1-propylpyrrolidine (5q**)**



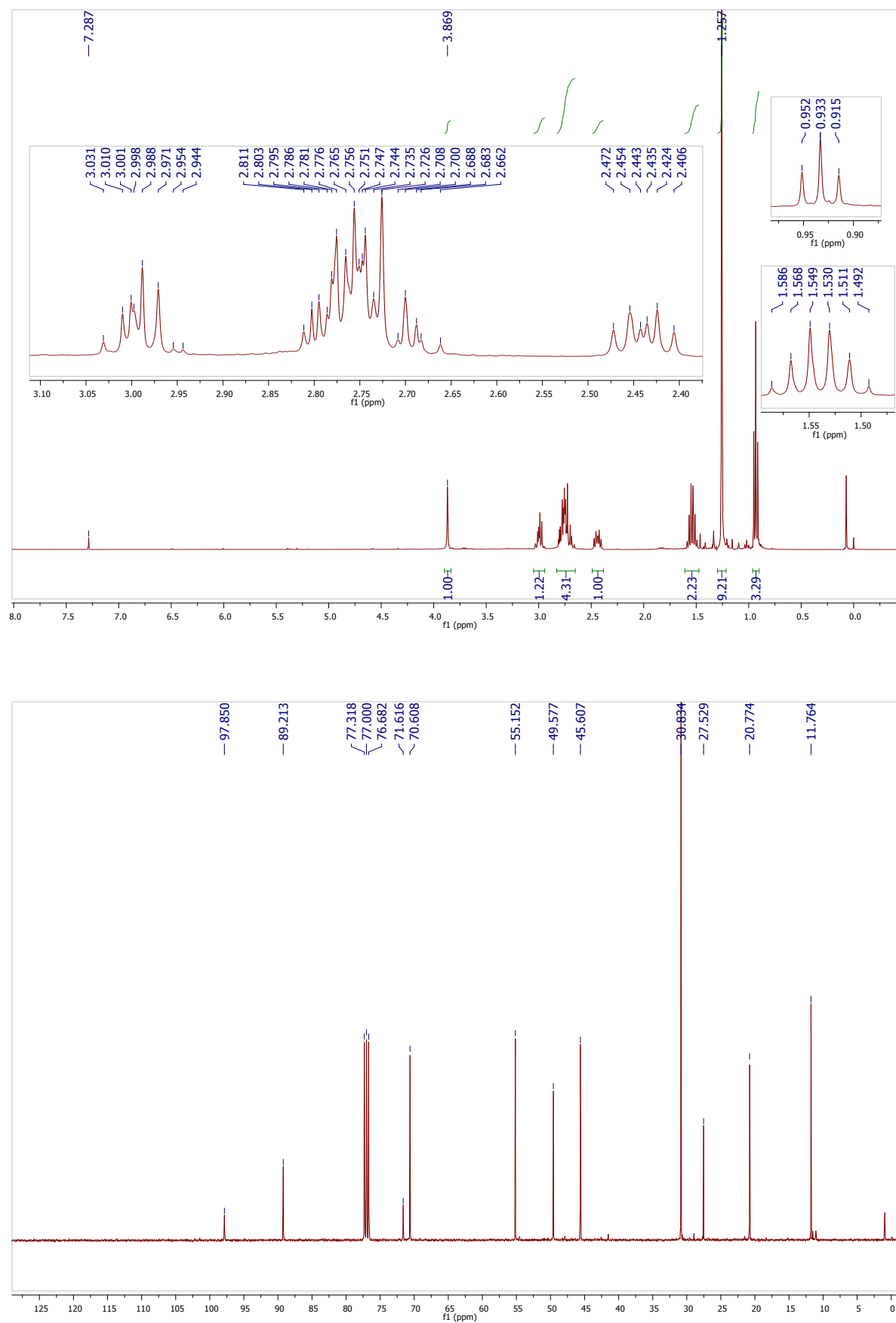


Figure 26. ¹H (400 MHz) and ¹³C (100 MHz) NMR spectra of **5q** in CDCl₃.

4. Copies of ^1H and ^{13}C spectra of 2-alkenylpyrroles **6a-f** and **7**

(*E/Z*)-3-Chloro-1-propyl-2-styryl-1*H*-pyrrole (**6a**)/(**6b**)

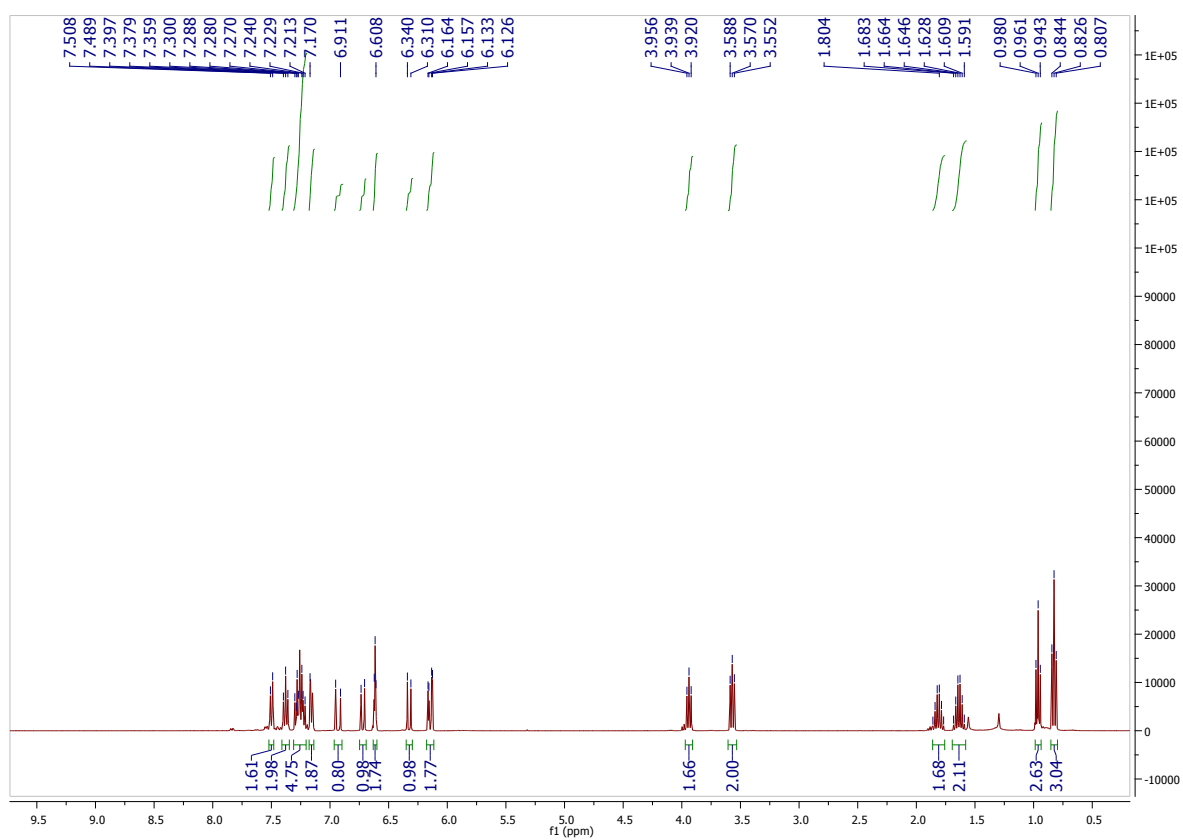
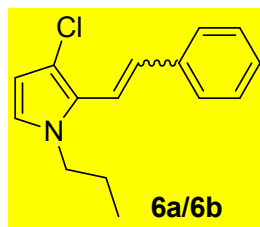
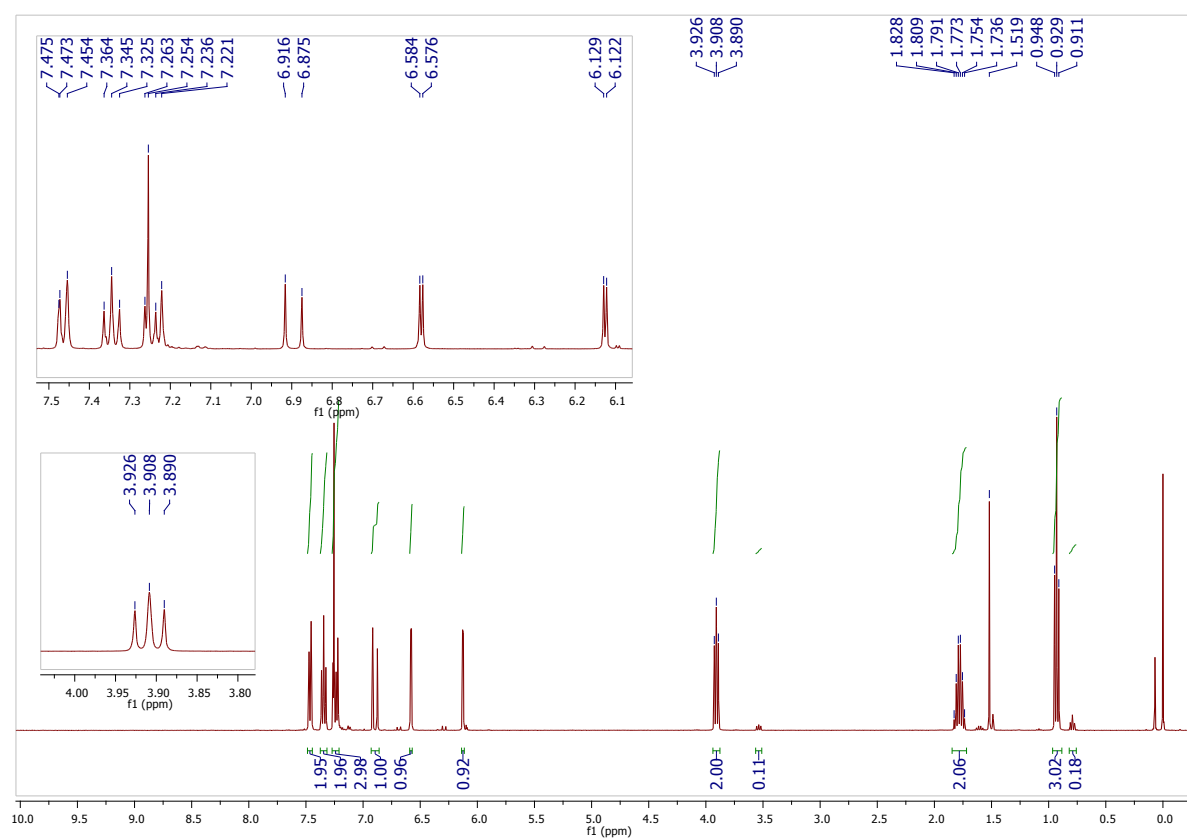


Figure 27. ^1H (400 MHz) NMR spectra of **6a/6b** (mixture of *E* and *Z*) in CDCl_3 .

(E)-3-Chloro-1-propyl-2-styryl-1*H*-pyrrole (6a)



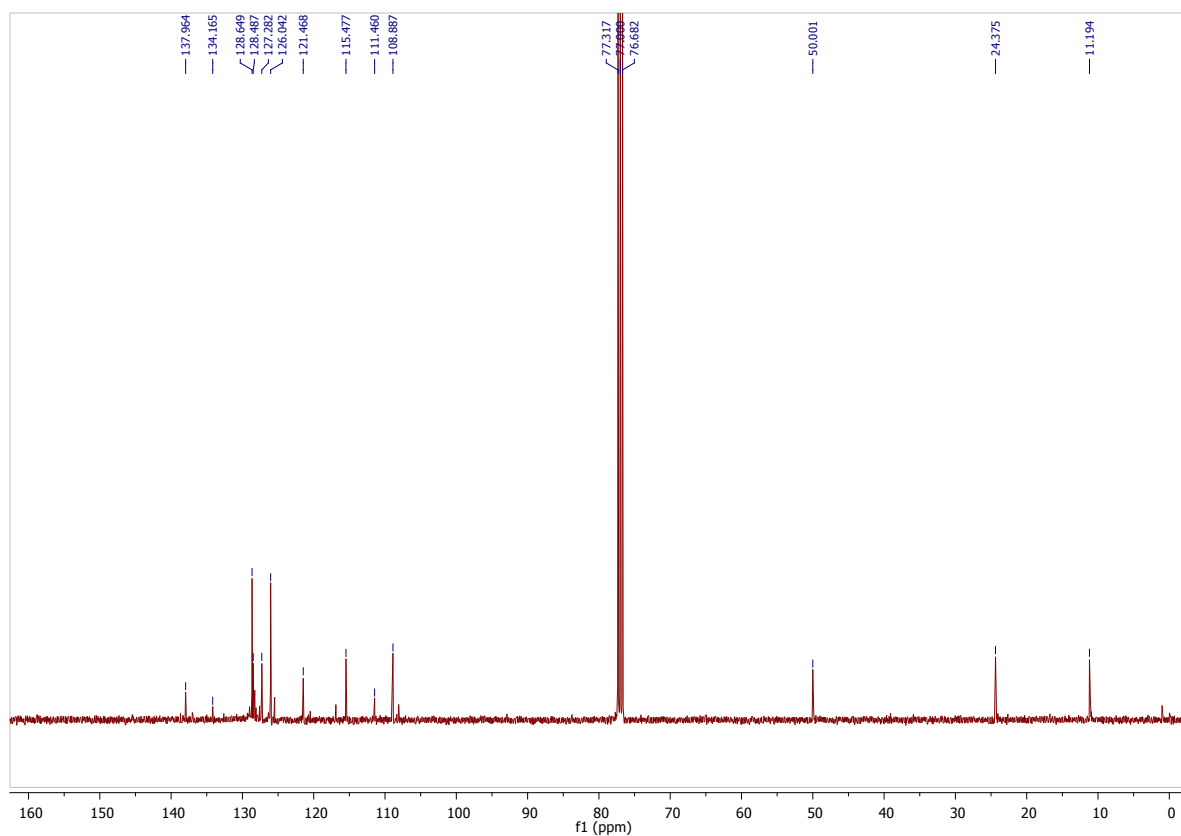
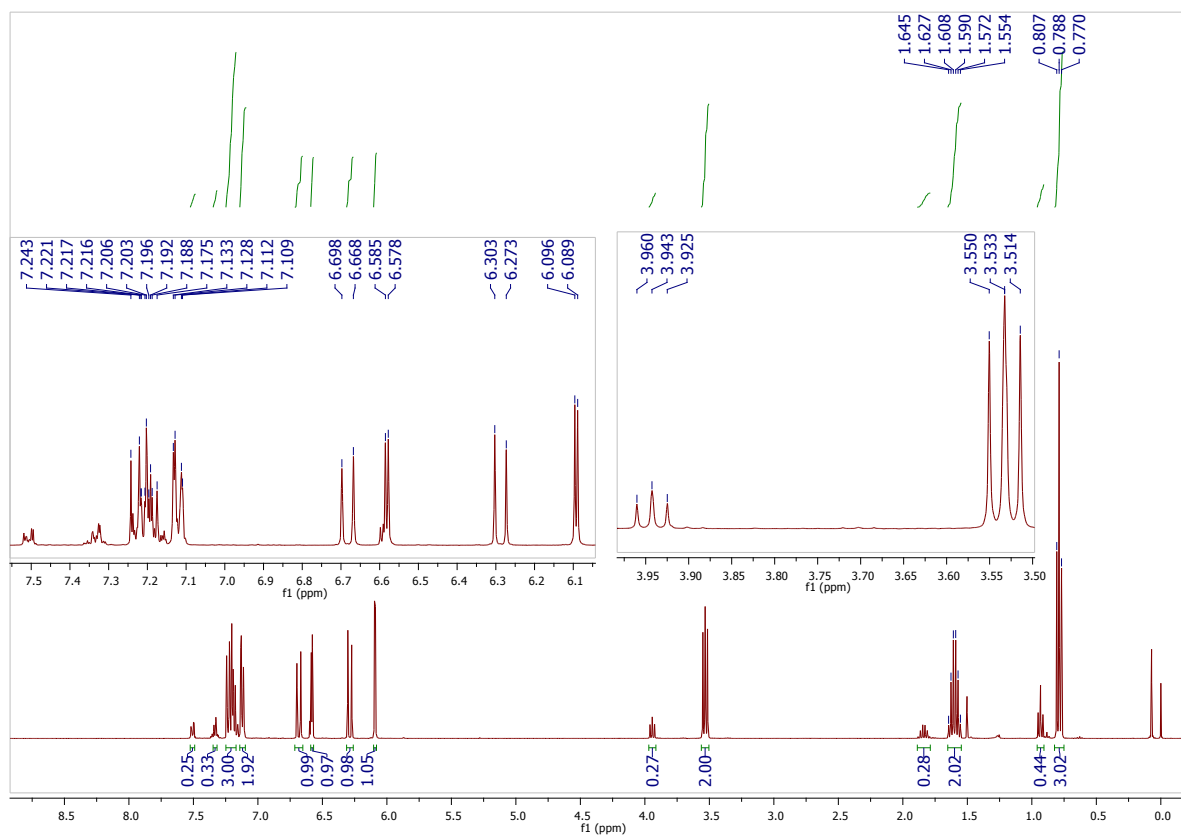


Figure 28. ^1H (400 MHz) and ^{13}C (100 MHz) NMR spectra of **6a** in CDCl_3 .

(Z)-3-Chloro-1-propyl-2-styryl-1H-pyrrole (6b)



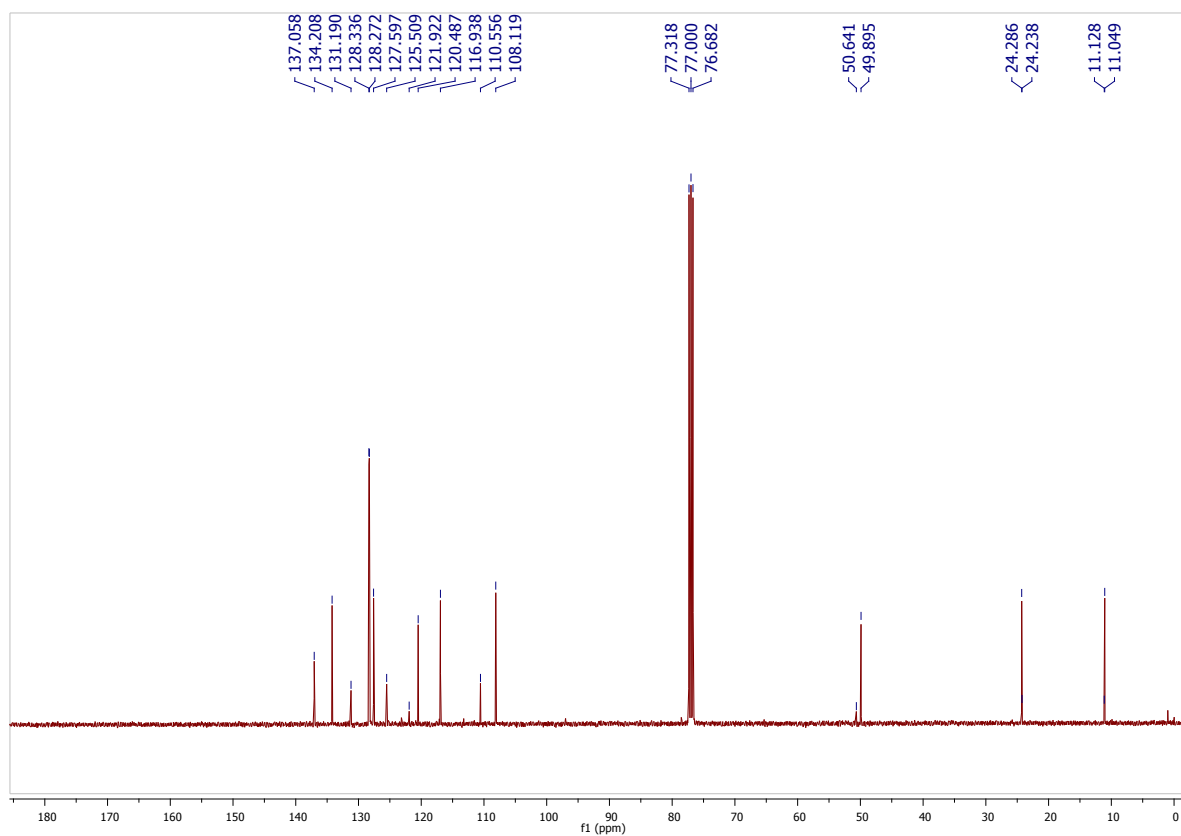
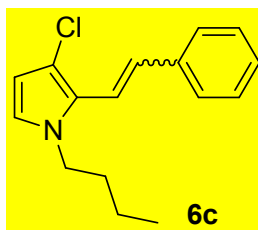
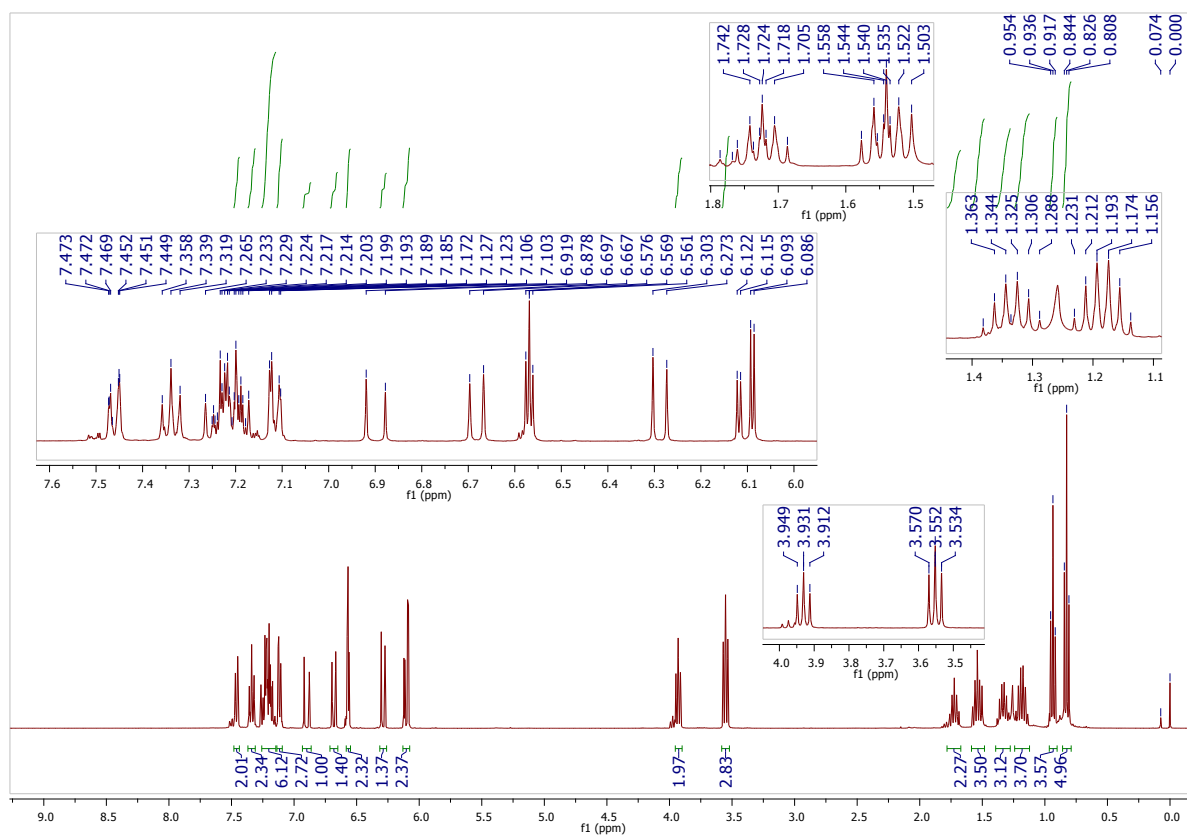


Figure 29. ^1H (400 MHz) and ^{13}C (100 MHz) NMR spectra of **6b** in CDCl_3 .

(E/Z)-1-Butyl-3-chloro-2-styryl-1H-pyrrole (6c)





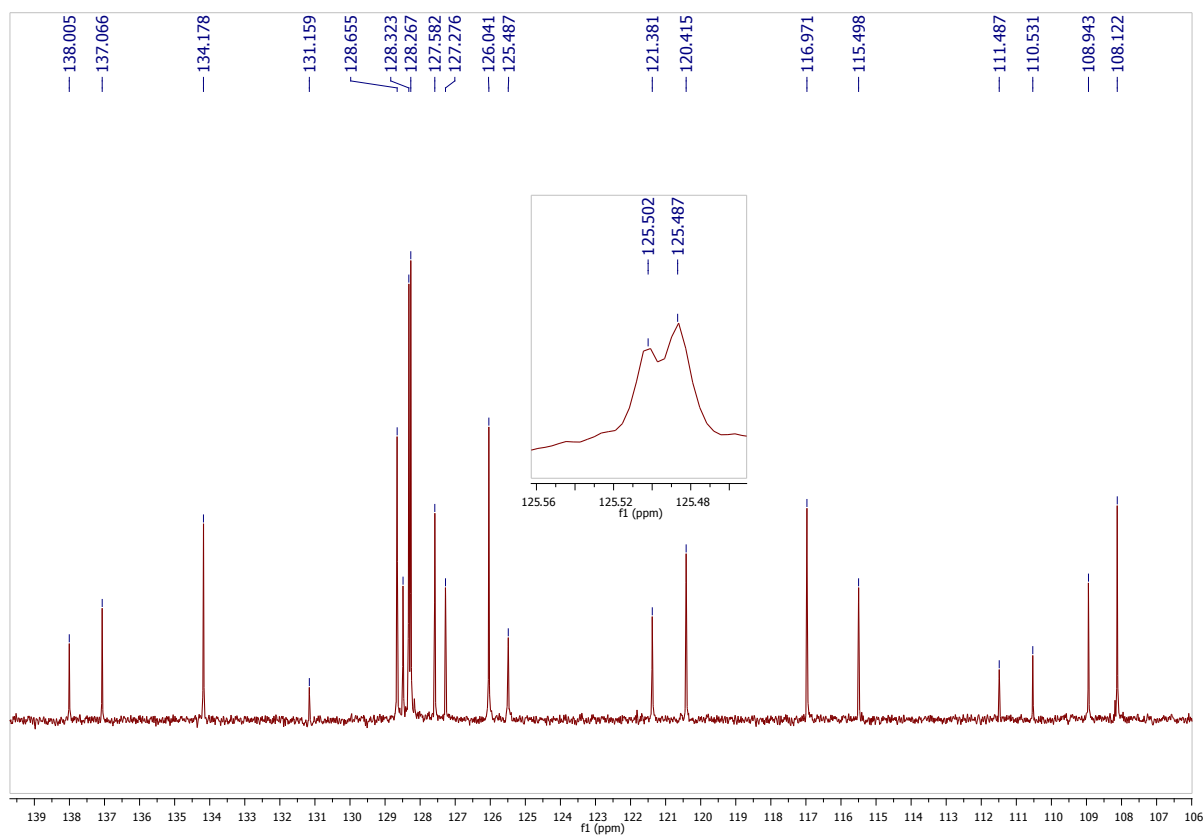
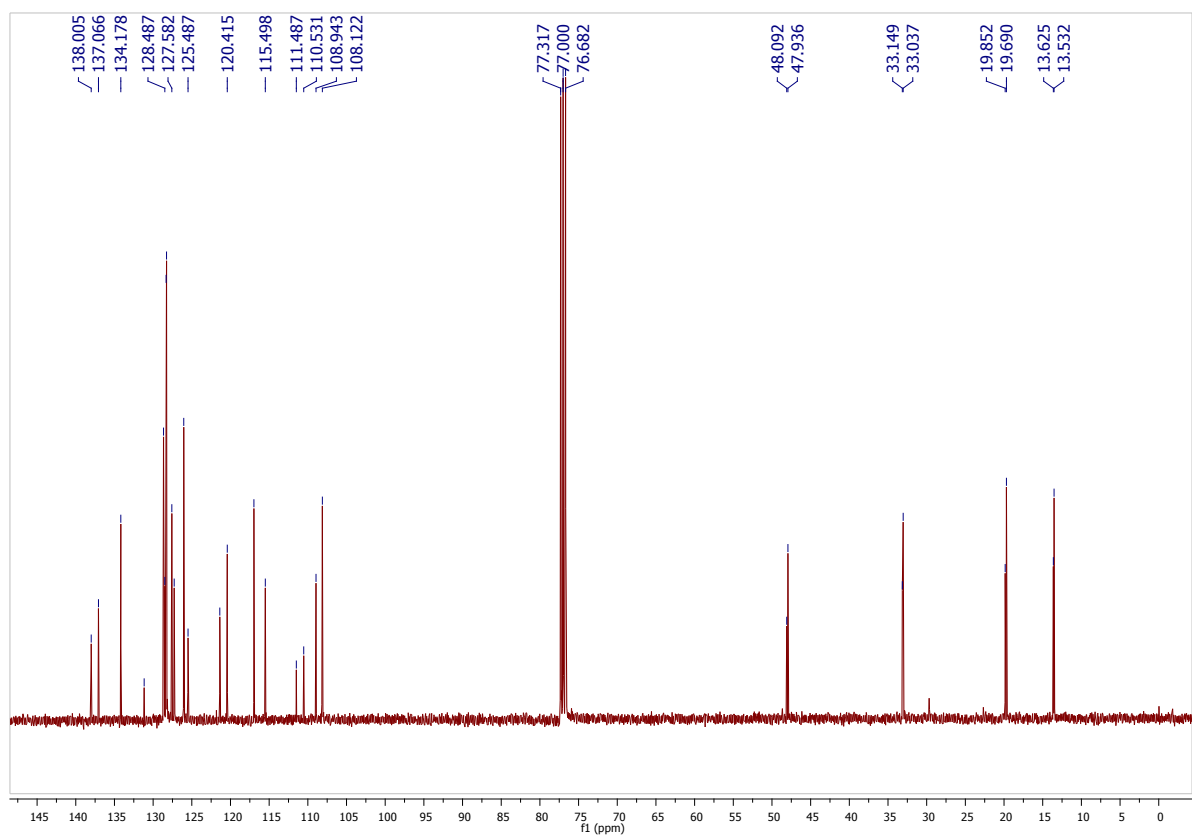
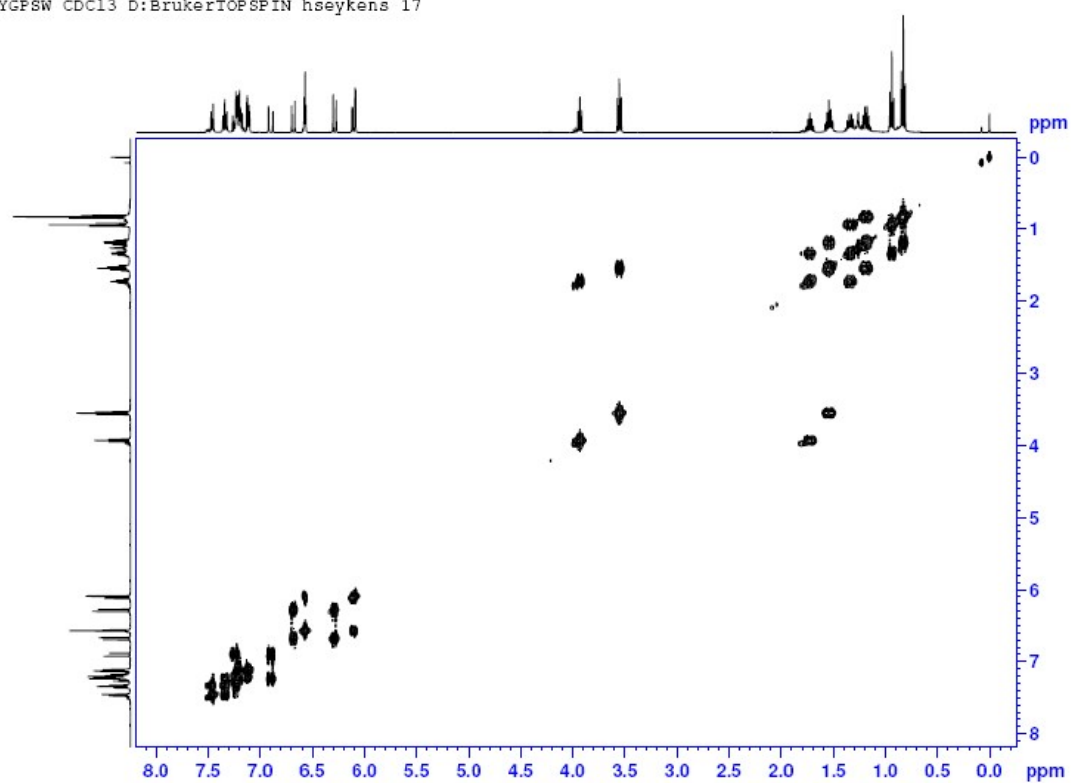
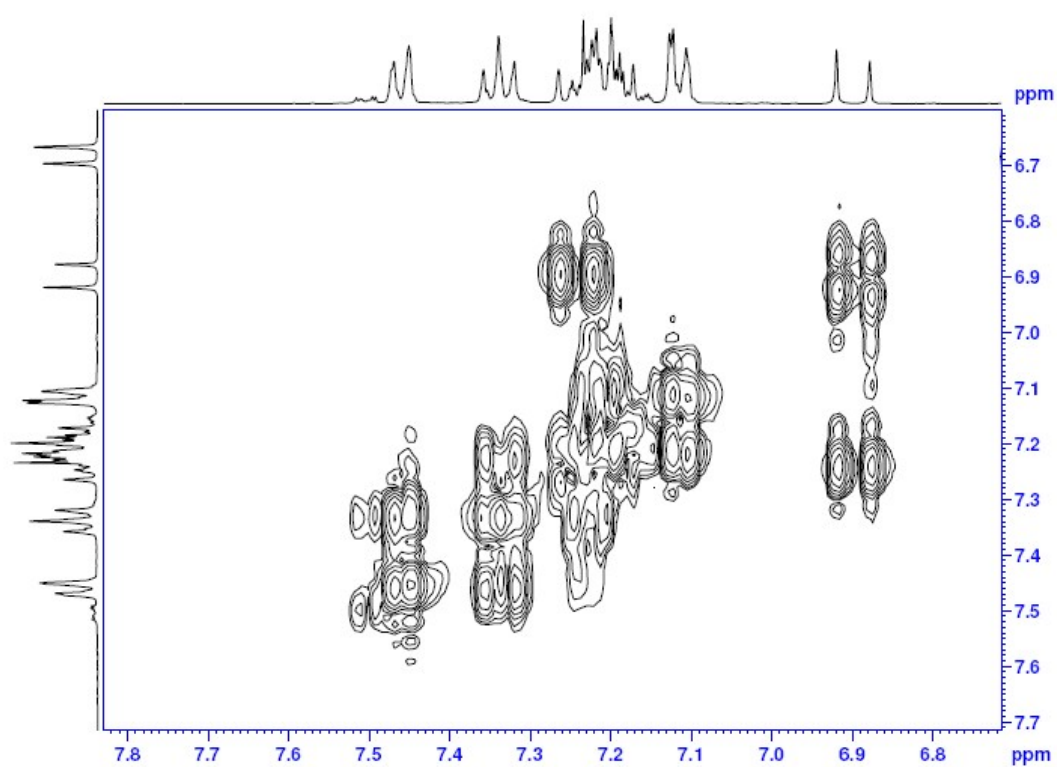


Figure 30. ¹H (400 MHz) and ¹³C (100 MHz) NMR spectra of **6c** (mixture of *E* and *Z*) in CDCl₃.

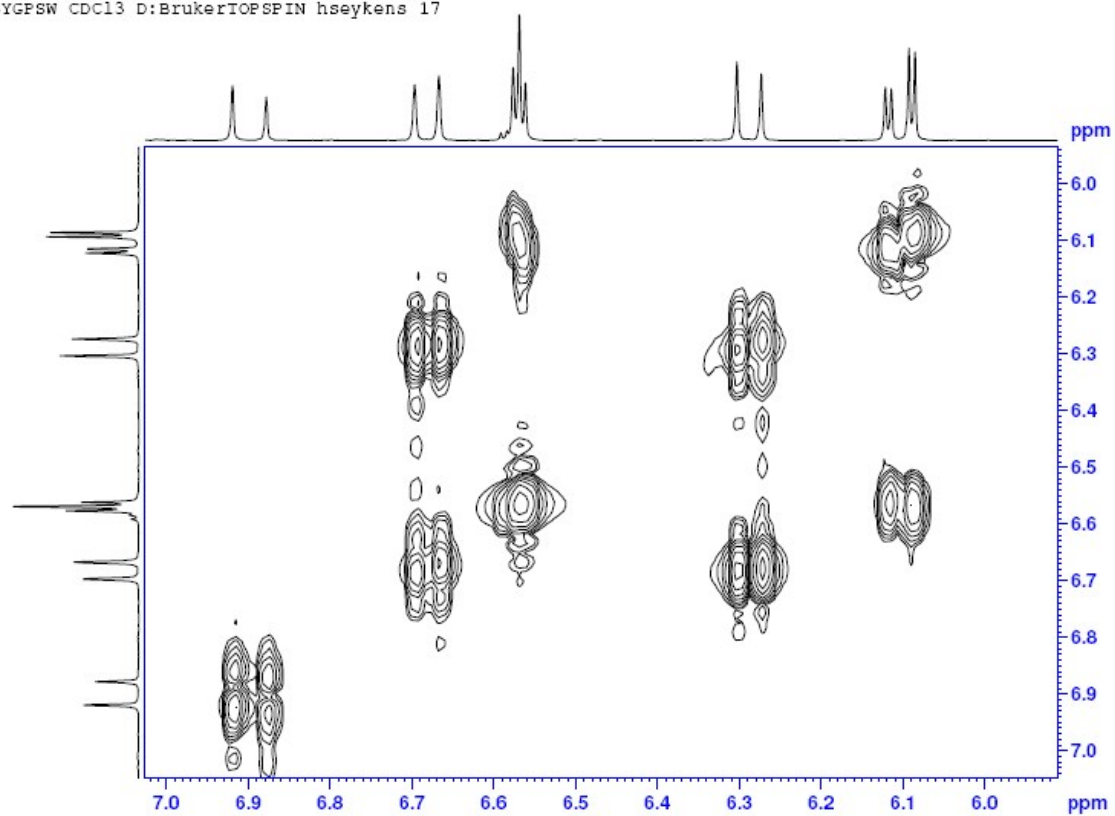
COSYGPSW CDC13 D:BrukerTOPSPIN hseykens 17



COSYGPSW CDCl3 D:BrukerTOPSPIN hseykens 17



COSYGPSW CDCl3 D:BrukerTOPSPIN hseykens 17



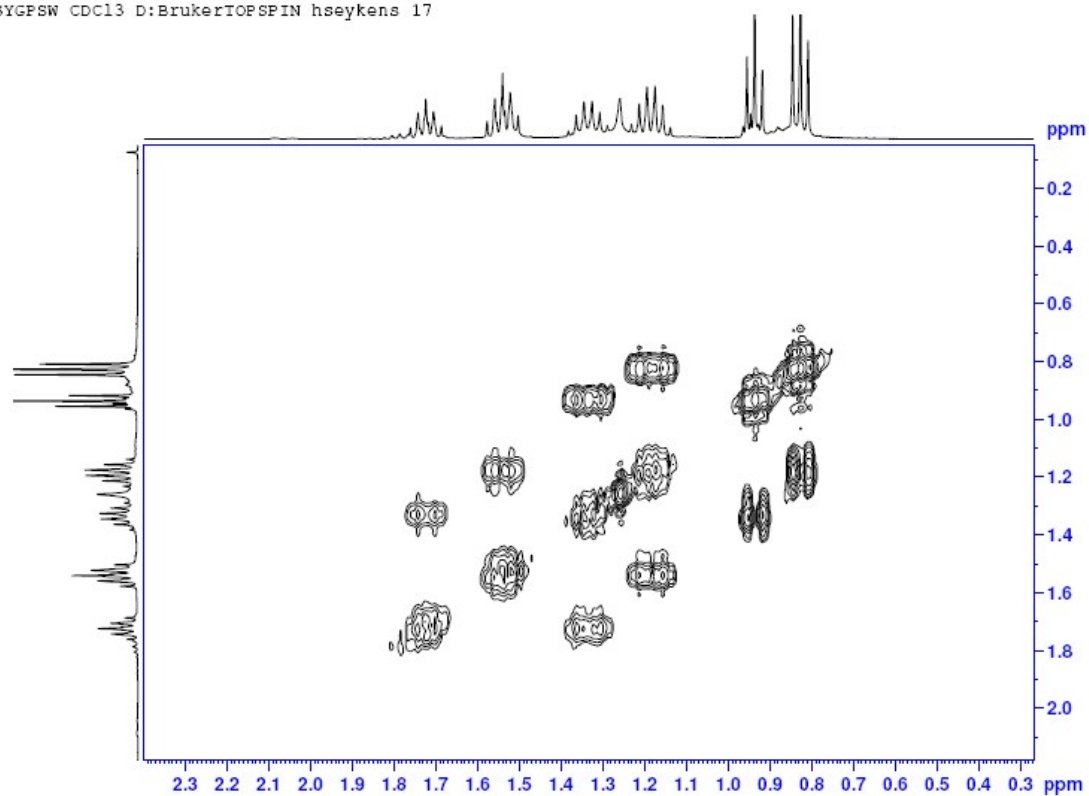
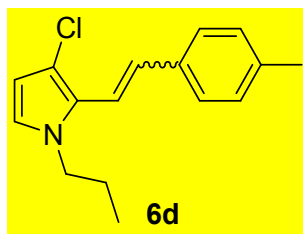
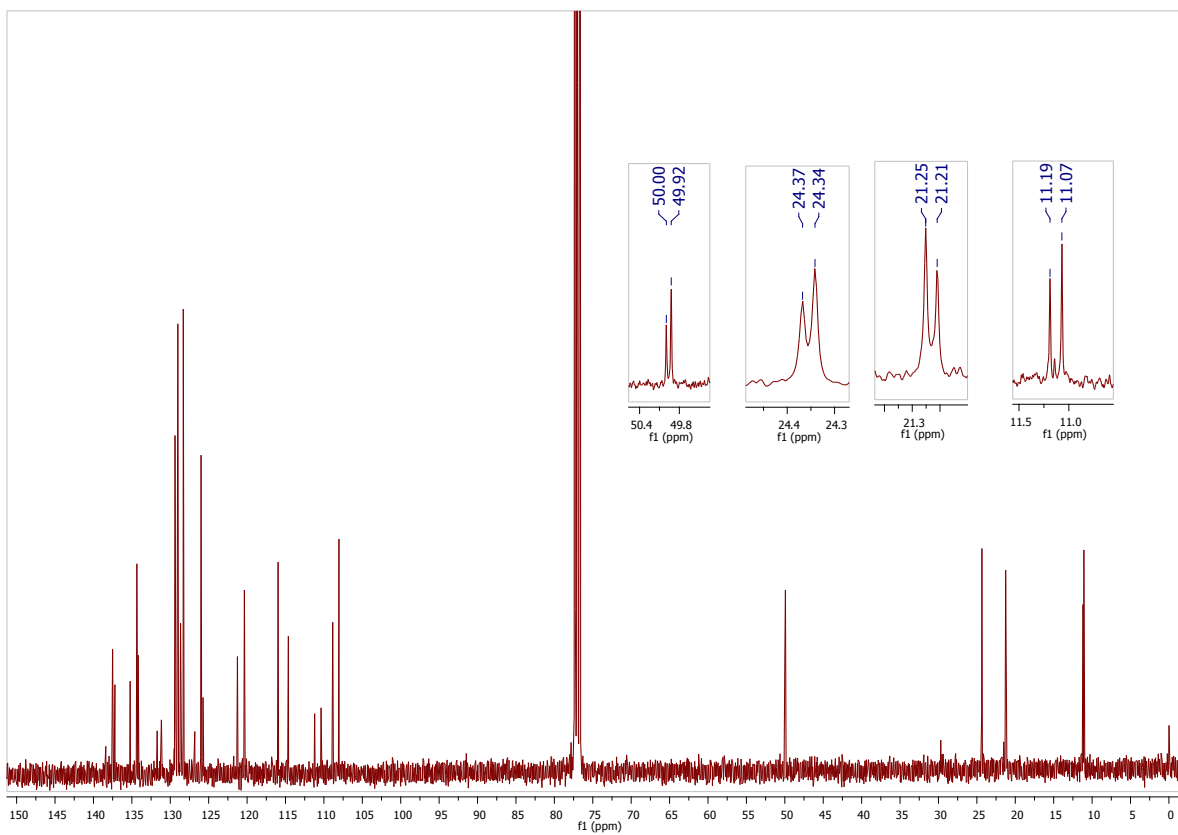
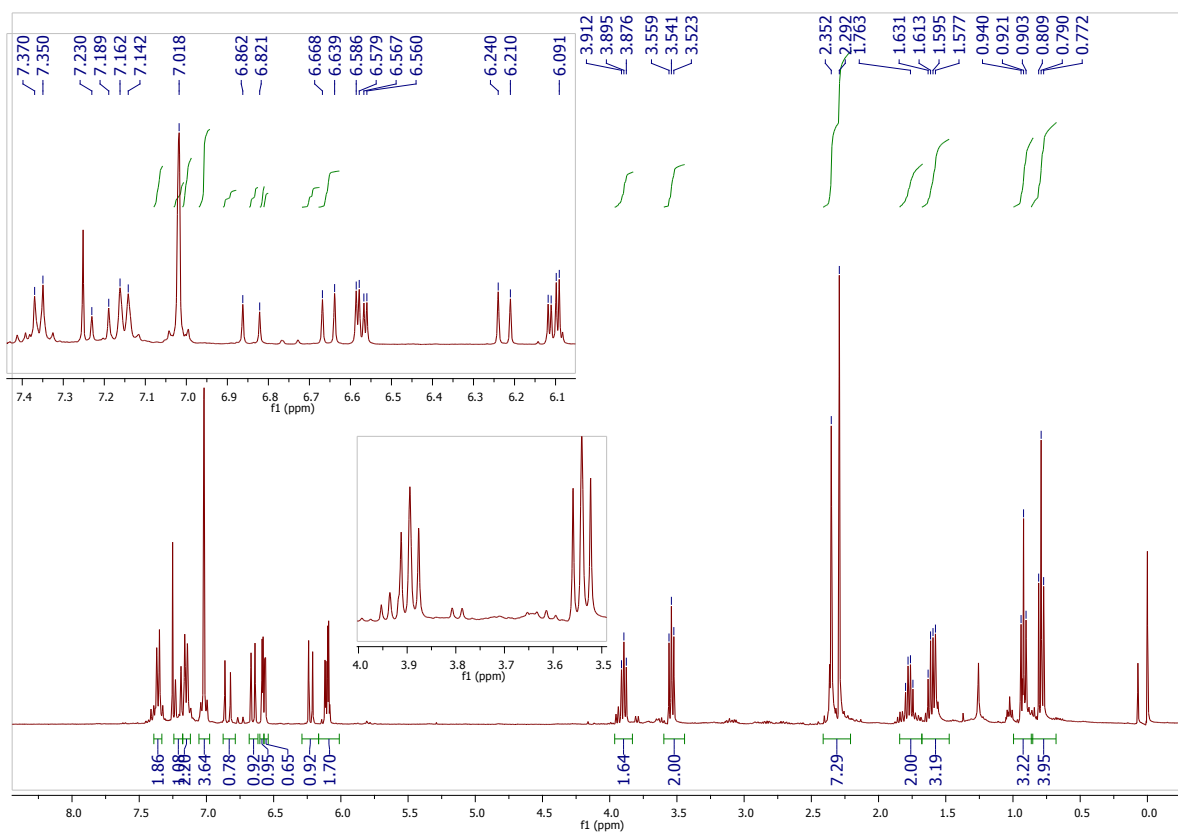


Figure 31. 2D-NMR spectra of **6c** in CDCl₃.

(*E/Z*)-3-Chloro-2-(4-methylstyryl)-1-propyl-1*H*-pyrrole (6d**)**





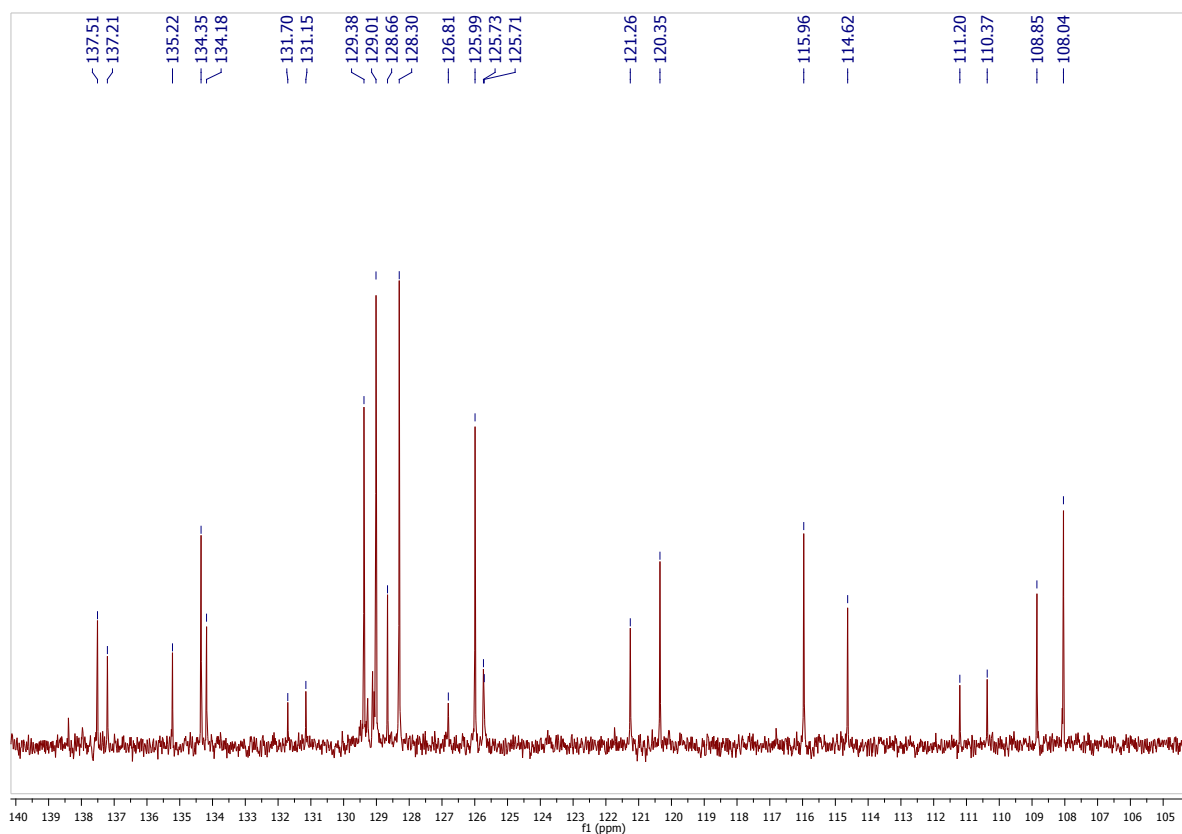
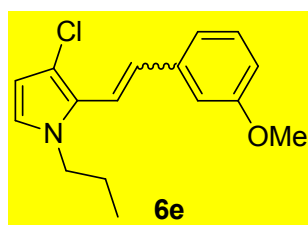
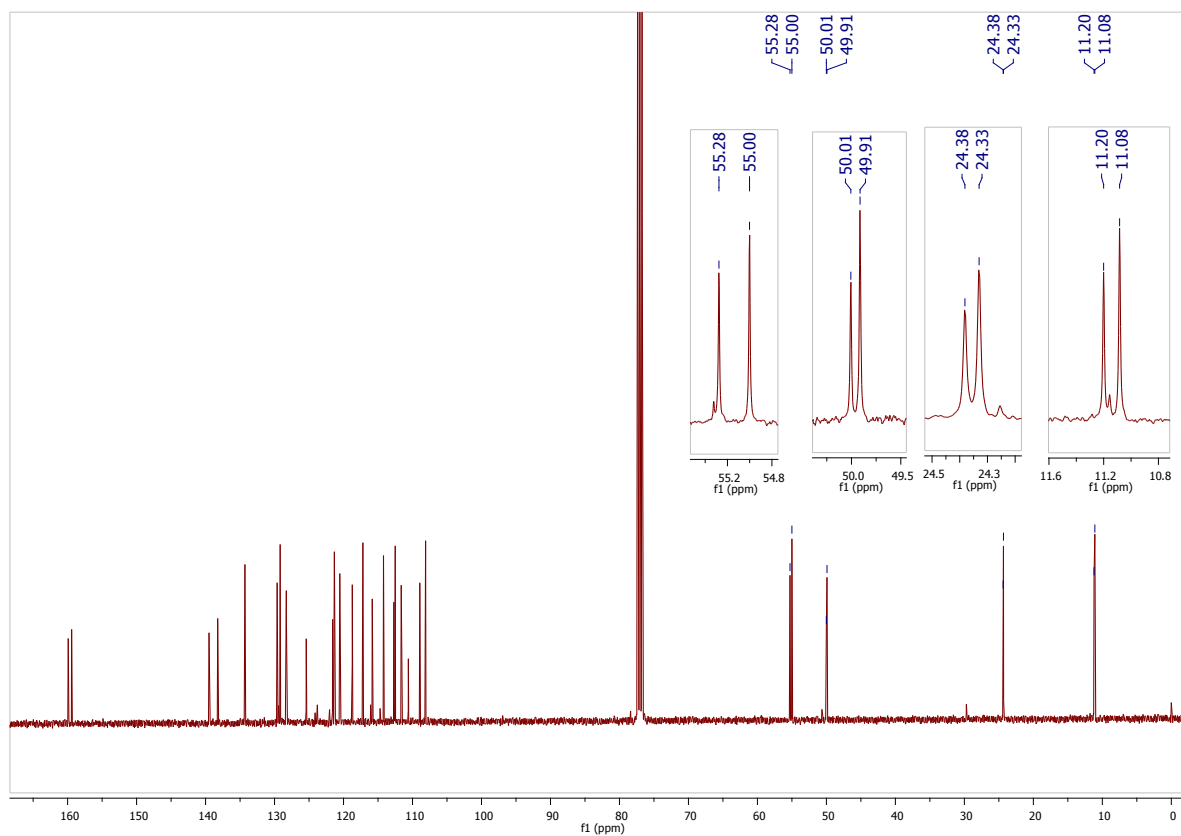
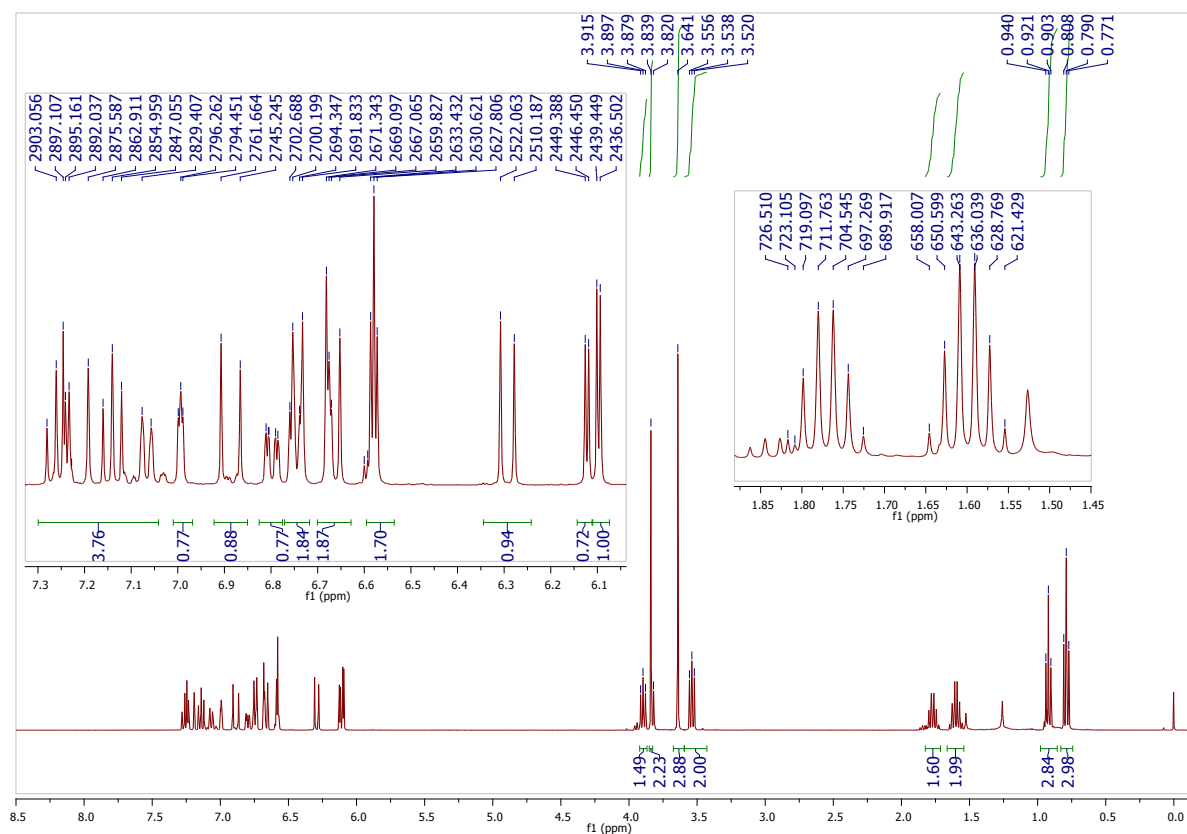


Figure 32. ^1H (400 MHz) and ^{13}C (100 MHz) NMR spectra of **6d** in CDCl_3 .

(*E/Z*)-3-Chloro-2-(3-methoxystyryl)-1-propyl-1*H*-pyrrole (6e**)**





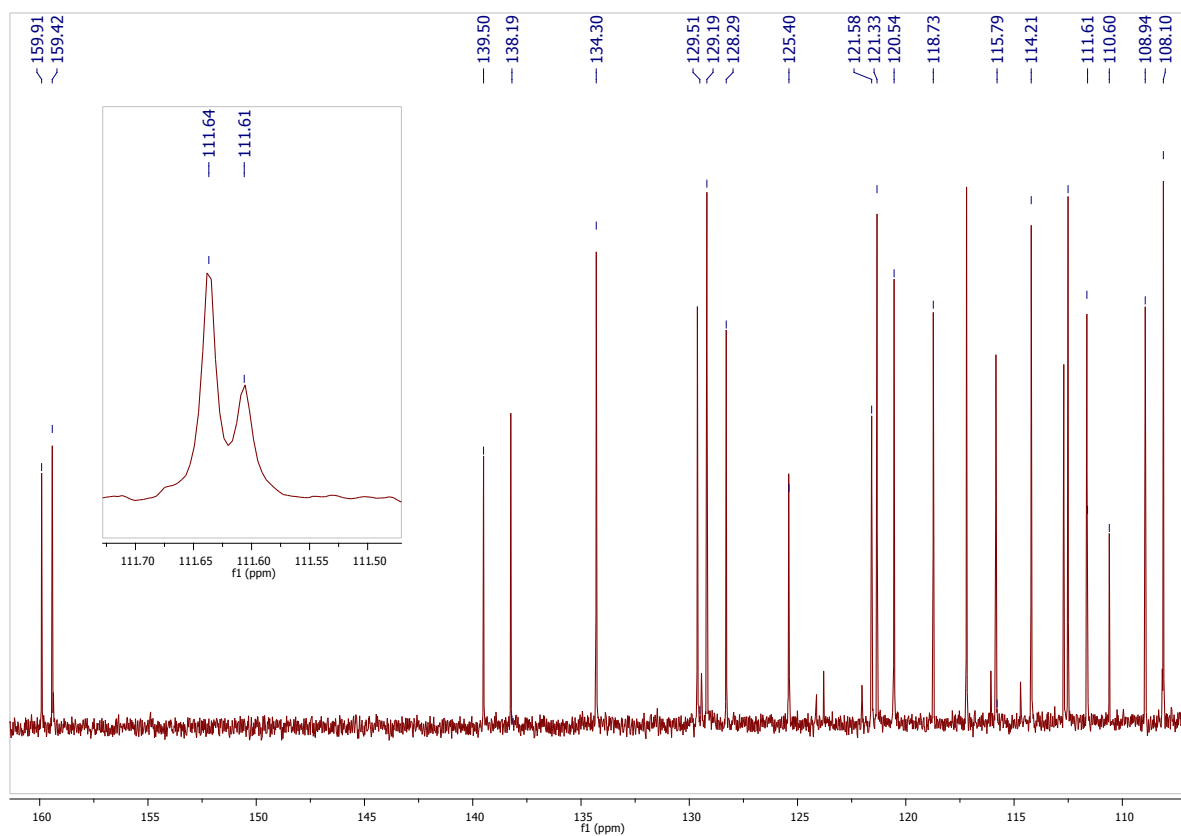
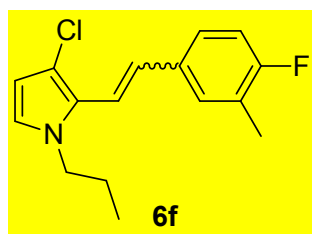
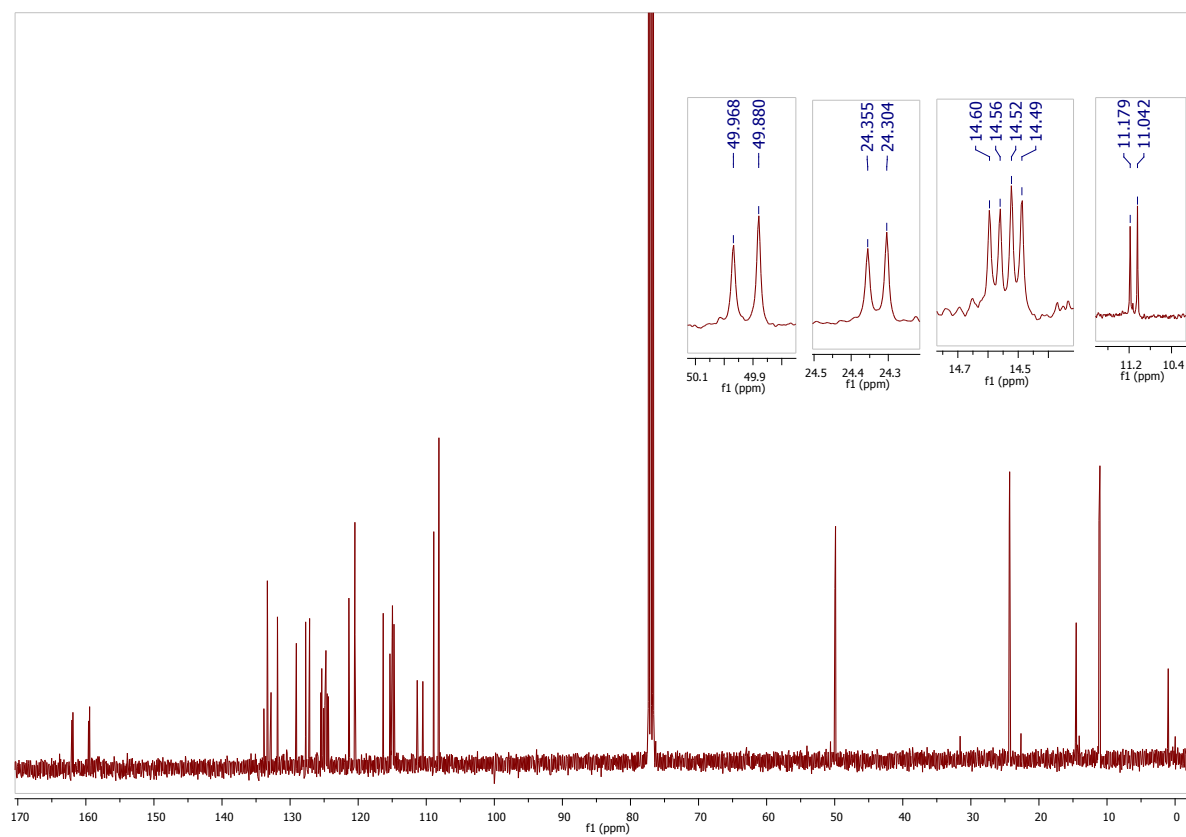
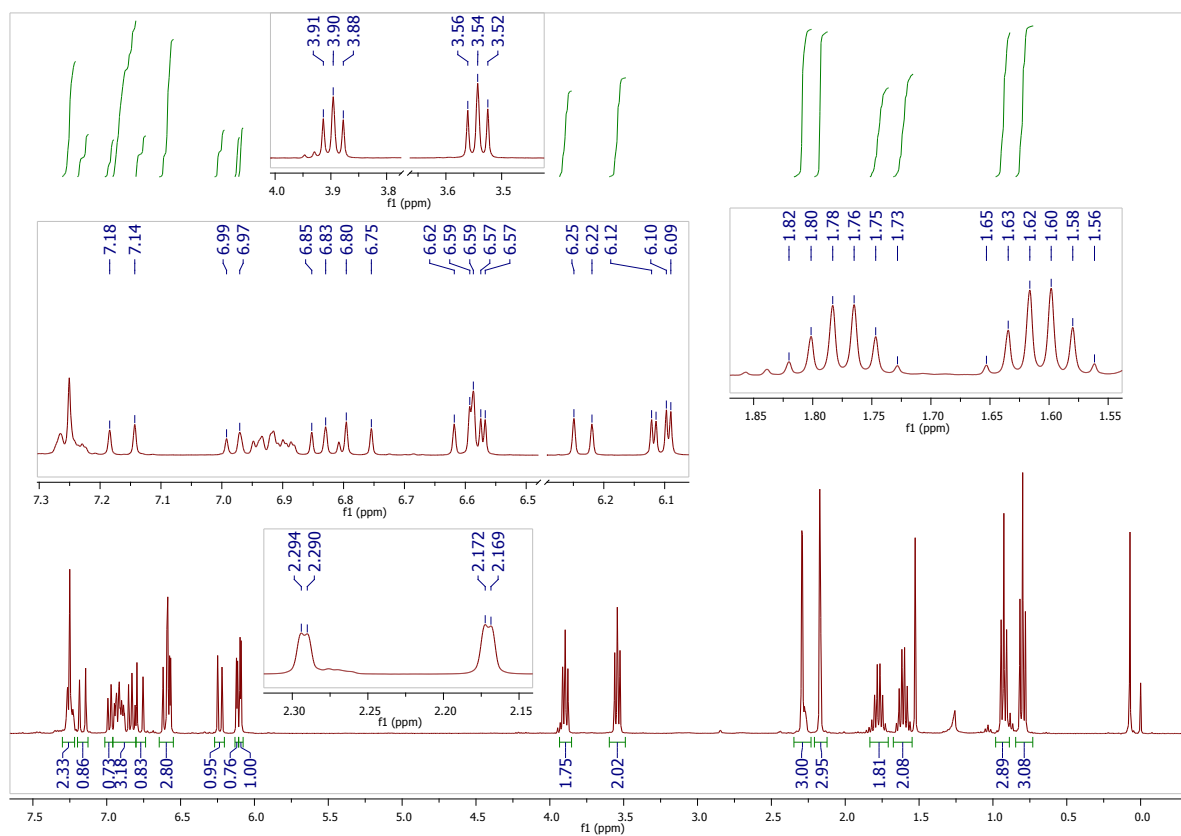


Figure 33. ^1H (400 MHz) and ^{13}C (100 MHz) NMR spectra of **6e** in CDCl_3 .

***E/Z*-(3-Chloro-2-(4-fluoro-3-methylstyryl)-1-propyl-1*H*-pyrrole (6f)**





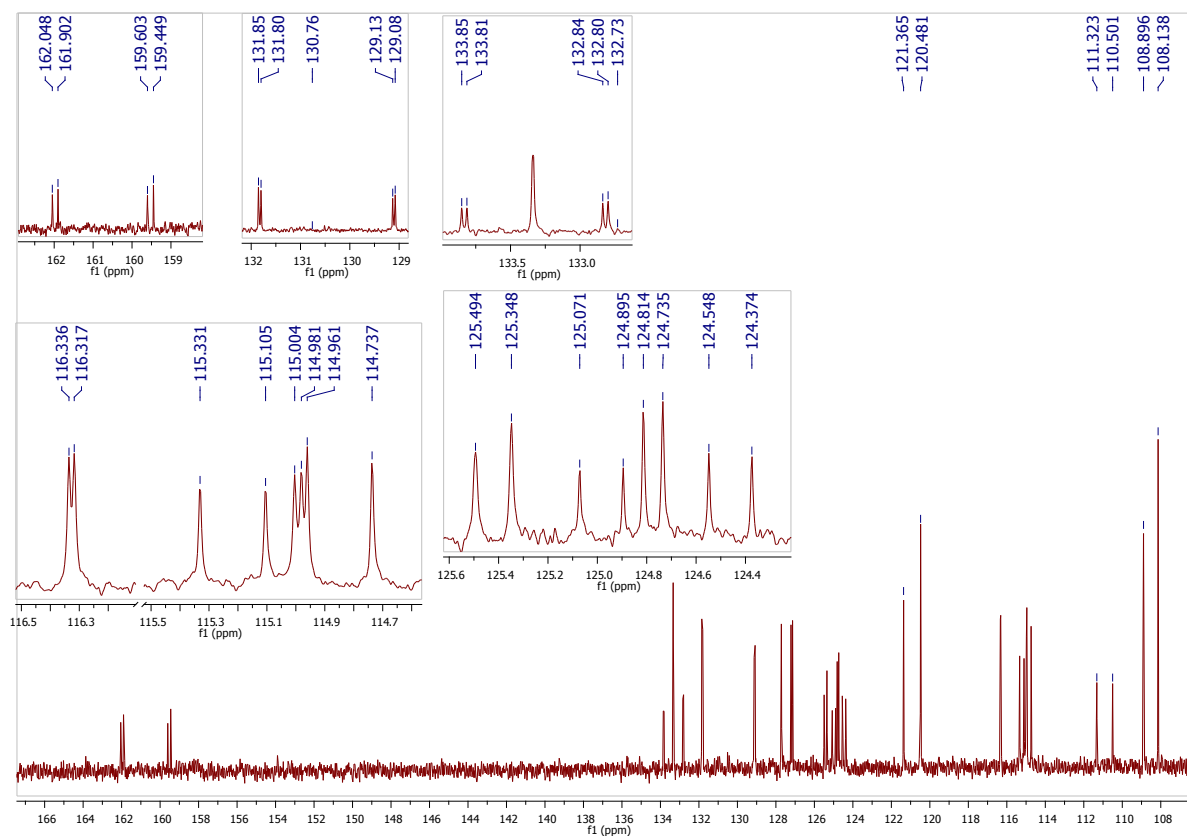
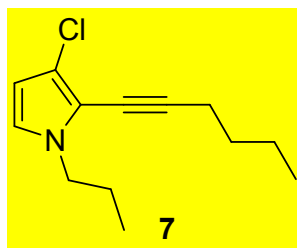


Figure 34. ¹H (400 MHz) and ¹³C (100 MHz) NMR spectra of **6f** in CDCl₃.

3-Chloro-2-(hex-1-ynyl)-1-propyl-1*H*-pyrrole (**7**)



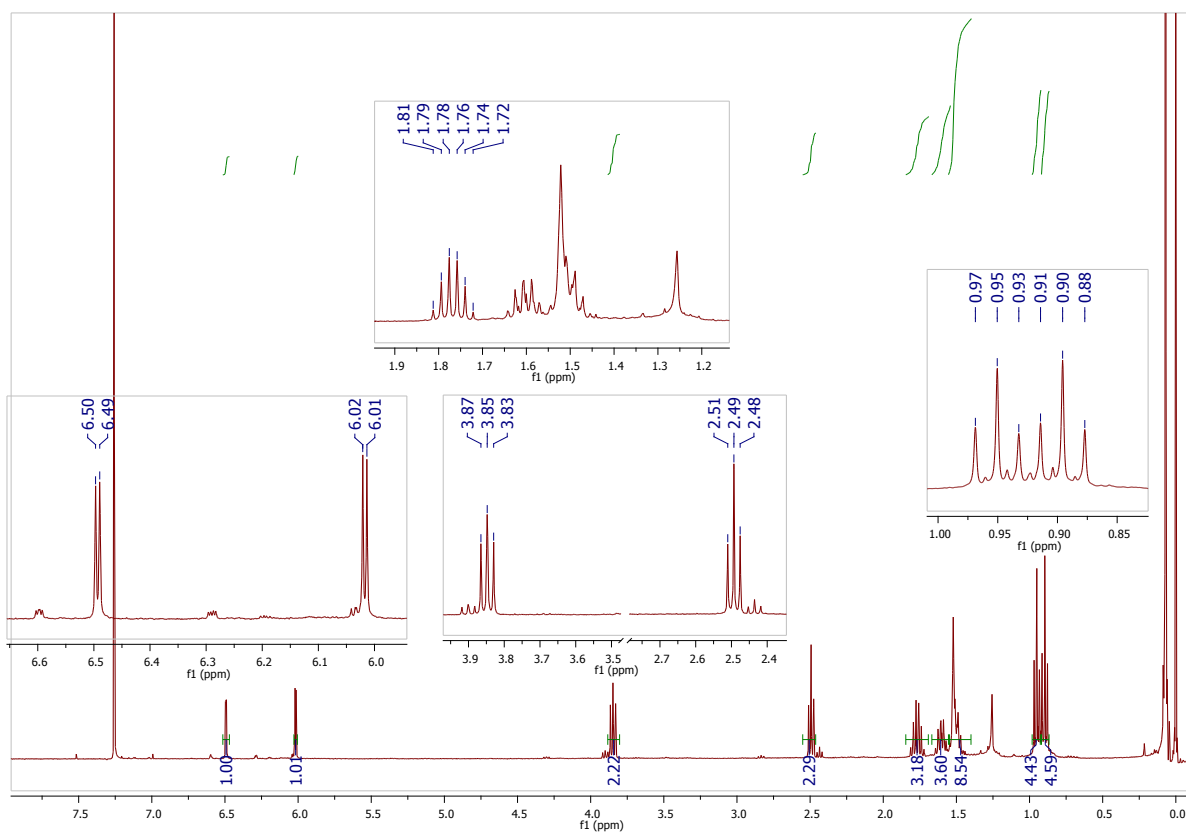
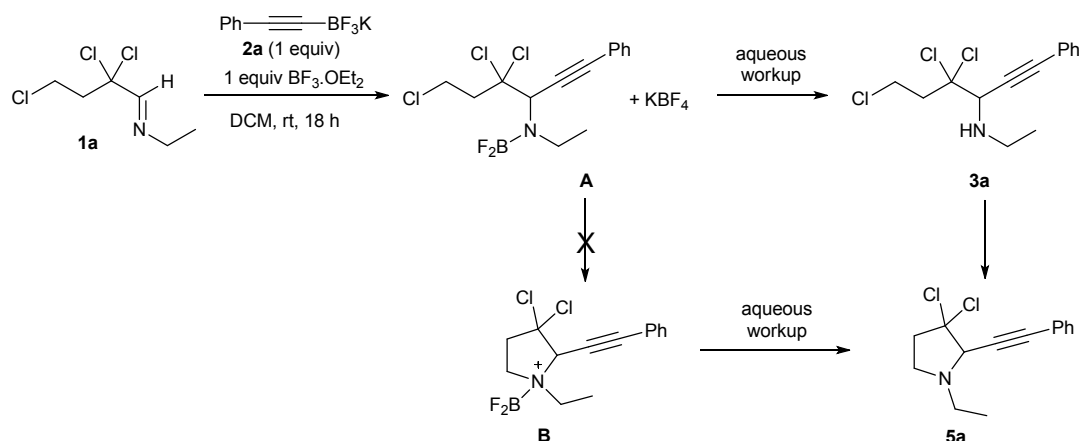


Figure 35. ^1H (400 MHz) NMR spectrum of **7** in CDCl_3 .

5. Mechanistic study for the borontrifluoride mediated reaction of imine with potassium alkynyltrifluoroborate



Scheme 1. Reaction mechanism of the Lewis acid promoted Petasis type reaction of α,α,ω -trichloroaldimines with organotrifluoroborates.

In a previous paper from our lab¹ it was established that the reaction of α,α -dichloroaldimines with potassiumtrifluoroborates results in an aminodifluoroborane intermediate which upon aqueous workup gives the β,β -dichloroamine. Because a ring closure takes place in the Petasis reaction of α,α,ω -trichloroaldimines with trifluoroborates, it would be valuable to establish if the ring closure either occurs during the reaction itself or during the aqueous workup. Since the aminodifluoroborane nitrogen atom is less nucleophilic it is expected that ring closure cannot take place after the addition of the alkyne moiety to the imino carbon. As a consequence the ring closure must occur during the aqueous workup. In order to prove the existence of an intermediate aminodifluoroborane, the reaction mixture obtained after Petasis reaction of *N*-(2,2,4-trichloro-1-butylidene)ethylamine (**1a**), potassium phenylethynyltrifluoroborate (**2a**) and $\text{BF}_3 \cdot \text{OEt}_2$, was filtered, evaporated and analysed by ^1H , ^{19}F and ^{11}B NMR. The solid, which was filtered off prior to evaporation was shown to be KBF_4 by comparison with a commercially available reference sample. The signals in the ^1H NMR (CDCl_3) of the filtrate after evaporation were shifted downfield compared to the signals of 3,3-dichloro-2-phenylethynylpyrrolidine (**5a**), pointing to the existence of an aminodifluoroborane intermediate **A**. Especially, the signal at 3.78 ppm is characteristic for a ClCH_2 and not a NCH_2 , indicating the presence of non-ring closed Mannich product. These results confirm the earlier stated assumption that first aminodifluoroborane **A** is formed and that ring closure takes only place during aqueous workup. A second experiment was executed to confirm the obtained results. The NMR sample of the filtrate was treated with a few drops

of water and the ^1H NMR was recorded again to check if trichlorinated Mannich product **3a** was present. In this case the formation of pyrrolidine **5a** (18%) and **3a** (82%) was observed. The presence of the latter was interpreted by an upfield shift of all the NMR-signals compared to the signals of aminodifluoroborane **A** (striking shift for CCl_2CHN from 5.00 ppm (**A**) to 4.80 ppm (**3a**)). Scheme 1 depicts the reaction mechanism of the Lewis acid mediated borono Mannich type reaction of α,α,ω -trichloroaldehydes **1** with organotrifluoroborates **2**. First, aminodifluoroborane **A** is formed which after aqueous workup is turned into the trichlorinated propargylic amine **3a**. Subsequently, an intramolecular nucleophilic substitution of the ω -chloro atom by the amine nitrogen atom of **3a** leads to pyrrolidine **5a**.

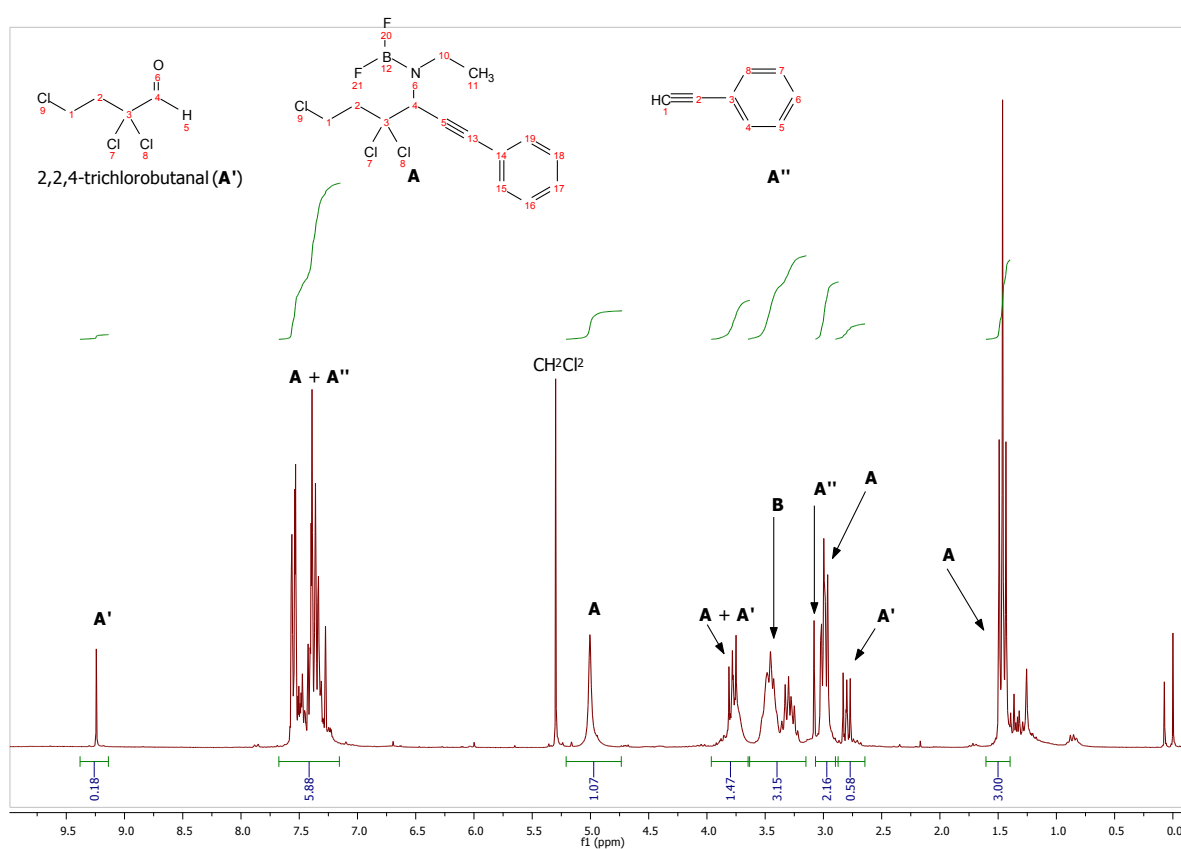


Figure 36. ^1H (400 MHz) NMR spectra of intermediate **A** in CDCl_3 .

6. Mechanistic study for the $\text{In}(\text{OTf})_3$ catalyzed reaction of imine with phenylacetylene

To prove that ring closure takes place during the alkynylation, a reaction was carried out between imine (**1b**) and phenylacetylene (2 equiv) in the presence of $\text{In}(\text{OTf})_3$ (0.25 equiv) in CH_2Cl_2 at 50 °C for 18 h. This reaction mixture was evaporated *in vacuo* and its ^1H NMR was measured. The mass spectrum of the reaction mixture showed a $[\text{M}+\text{H}^+]$ peak at 282 indicating the formation of the pyrrolidine ring during the reaction, while the absence of a

peak at 318 excludes the trichlorinated open chain intermediate (Figure 37). The ^1H NMR spectrum (Figure 38) also suggests the formation of a pyrrolidine ring, either complexed with the Lewis acid or in the form of a hydrochloride salt ($\text{X} = \text{H}$ or $\text{In}(\text{OTf})_3$; Figure 39, triplet at 1.03, multiplet at 1.94-1.83, multiplets at 2.96-3.13 & 3.45 ppm). A very broad peak around 5 ppm accounts for the NCHCCl_2 proton and this downfield shift, compared to the NCHCCl_2 proton (4.11 ppm) of **5b** (Figure 10) is indicative for a positively charged nitrogen atom. A comparison was made of this spectrum with the ^1H NMR of an independently prepared sample of the hydrochloride of pyrrolidine **5b** (Figure 39). Since both spectra are not the same, it can be concluded that most probably pyrrolidine **5b** formed a complex with $\text{In}(\text{OTf})_3$ during the reaction.

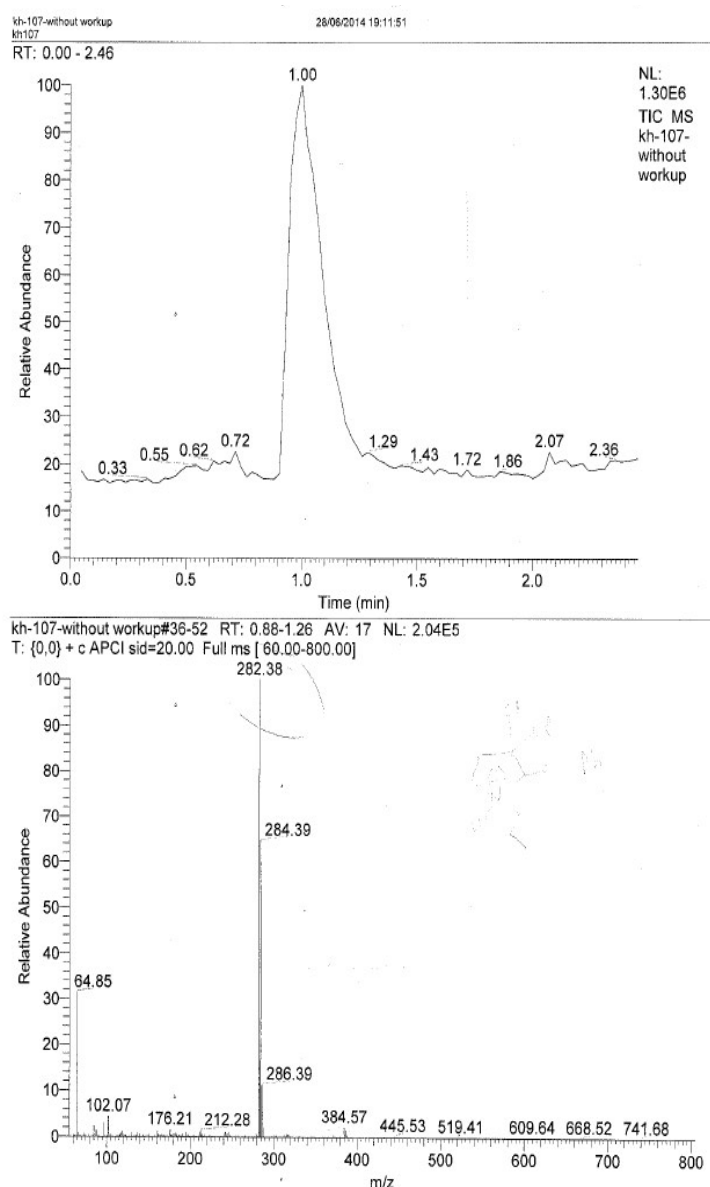


Figure 37. Mass spectrum of reaction mixture.

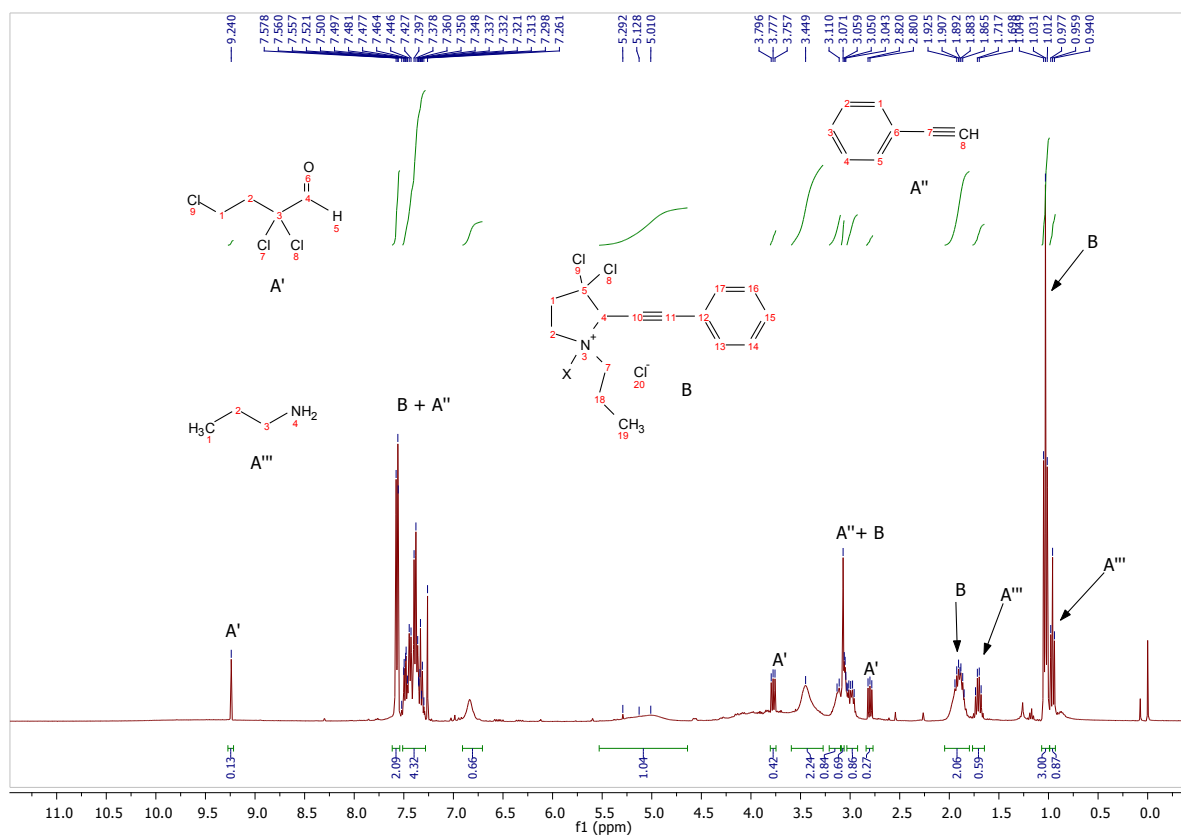


Figure 38. ^1H (400 MHz) NMR spectra of reaction mixture in CDCl_3 .

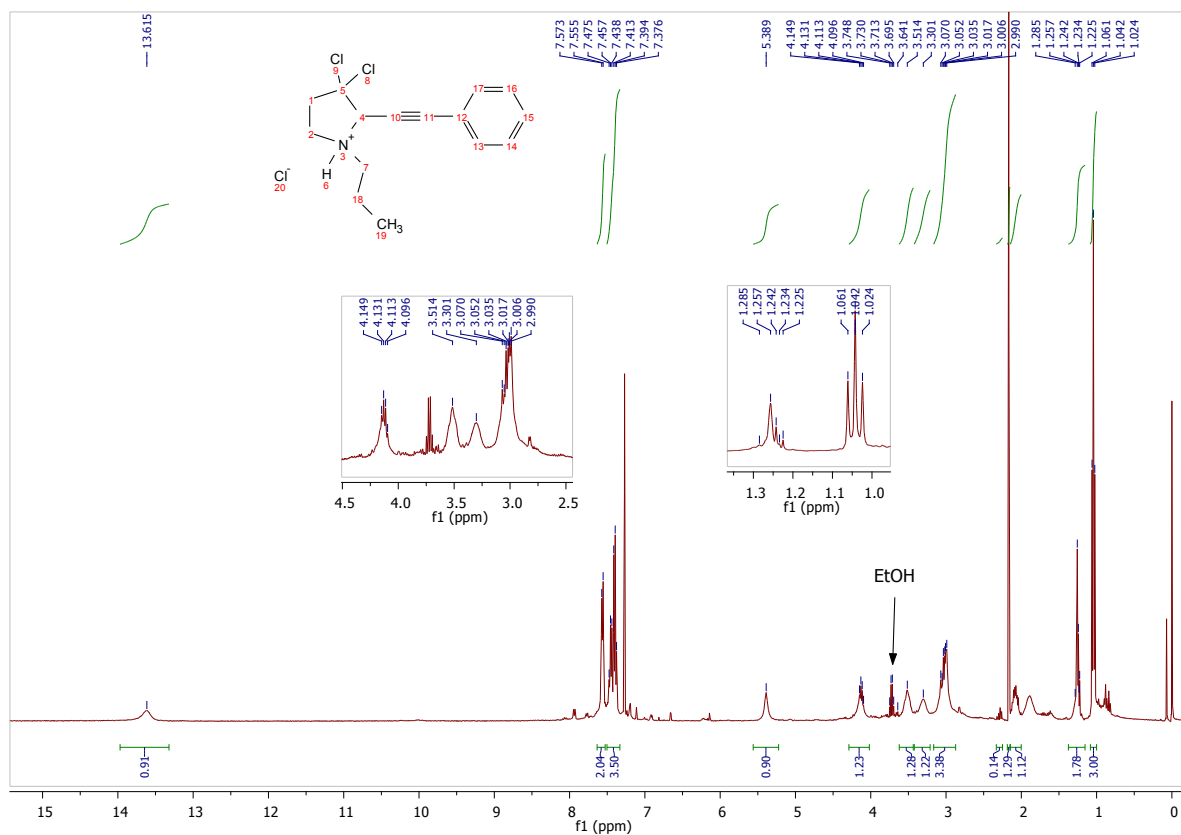
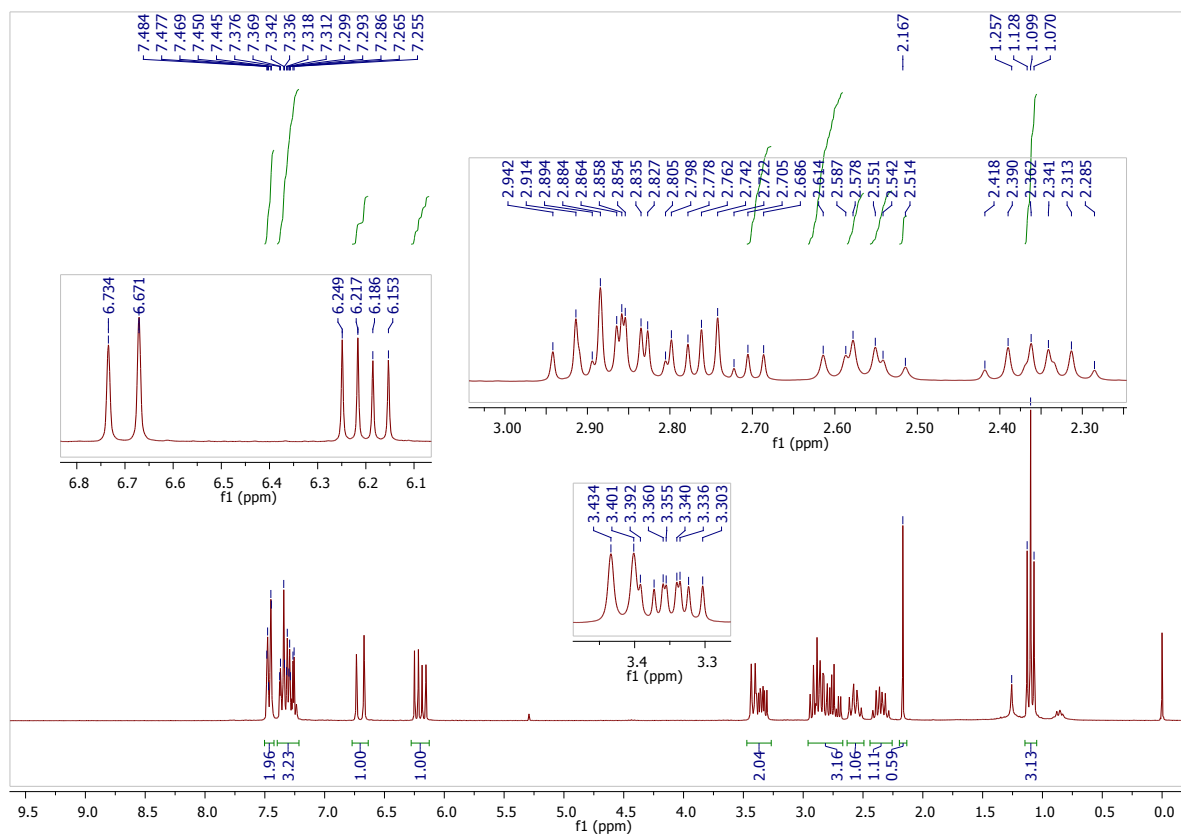
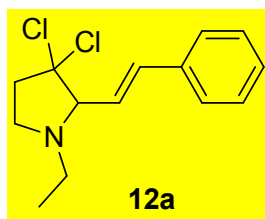


Figure 39. ^1H (400 MHz) NMR spectra of hydrochloride salt of pyrrolidine **5b** in CDCl_3 .

7. Copies of ^1H and ^{13}C spectra of 3,3-dichloro-1-ethyl-2-(2-phenylvinyl)pyrrolidine (12a)



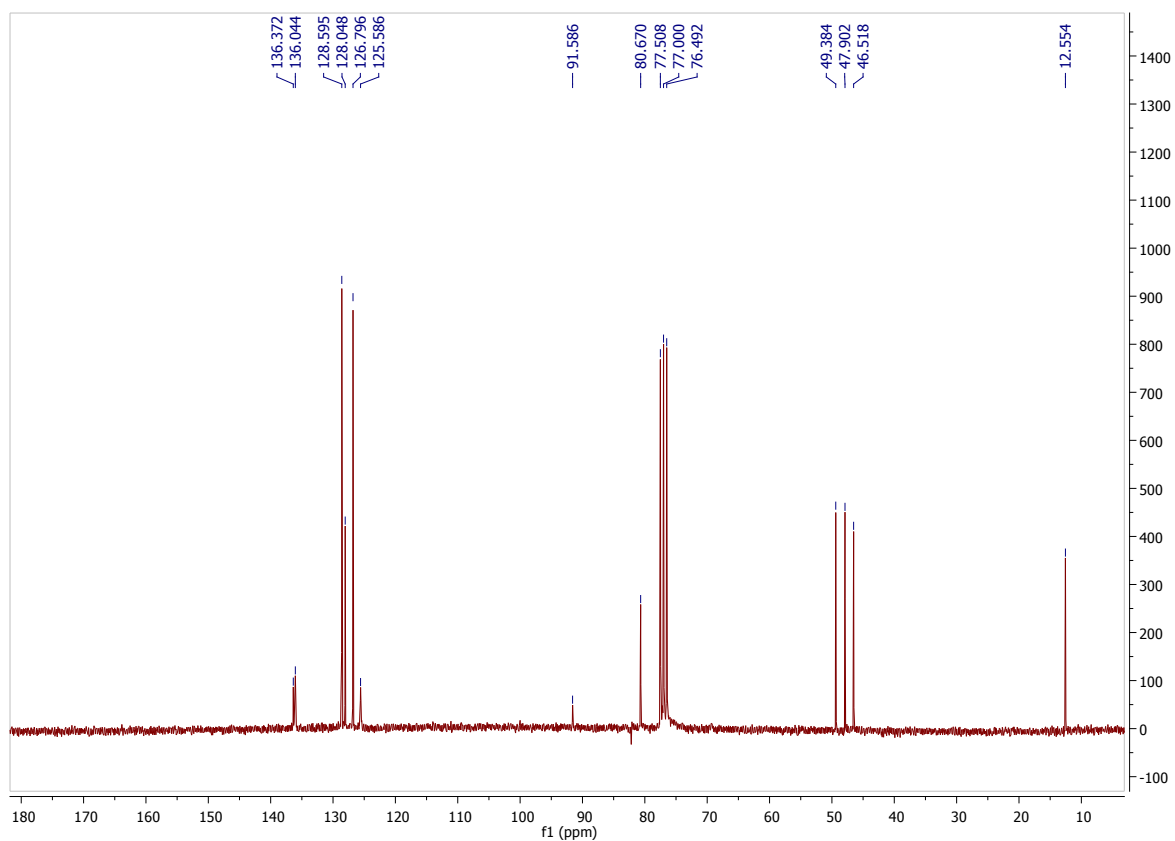
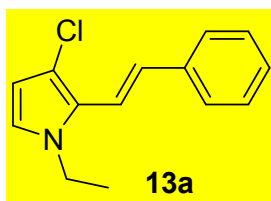


Figure 40. ^1H (250 MHz) and ^{13}C (63 MHz) NMR spectra of **12a** in CDCl_3 .

8. Copies of ^1H and ^{13}C Spectra of **13a** and **14a**

3-Chloro-1-ethyl-2-(2-phenylvinyl)-1*H*-pyrrole (**13a**)



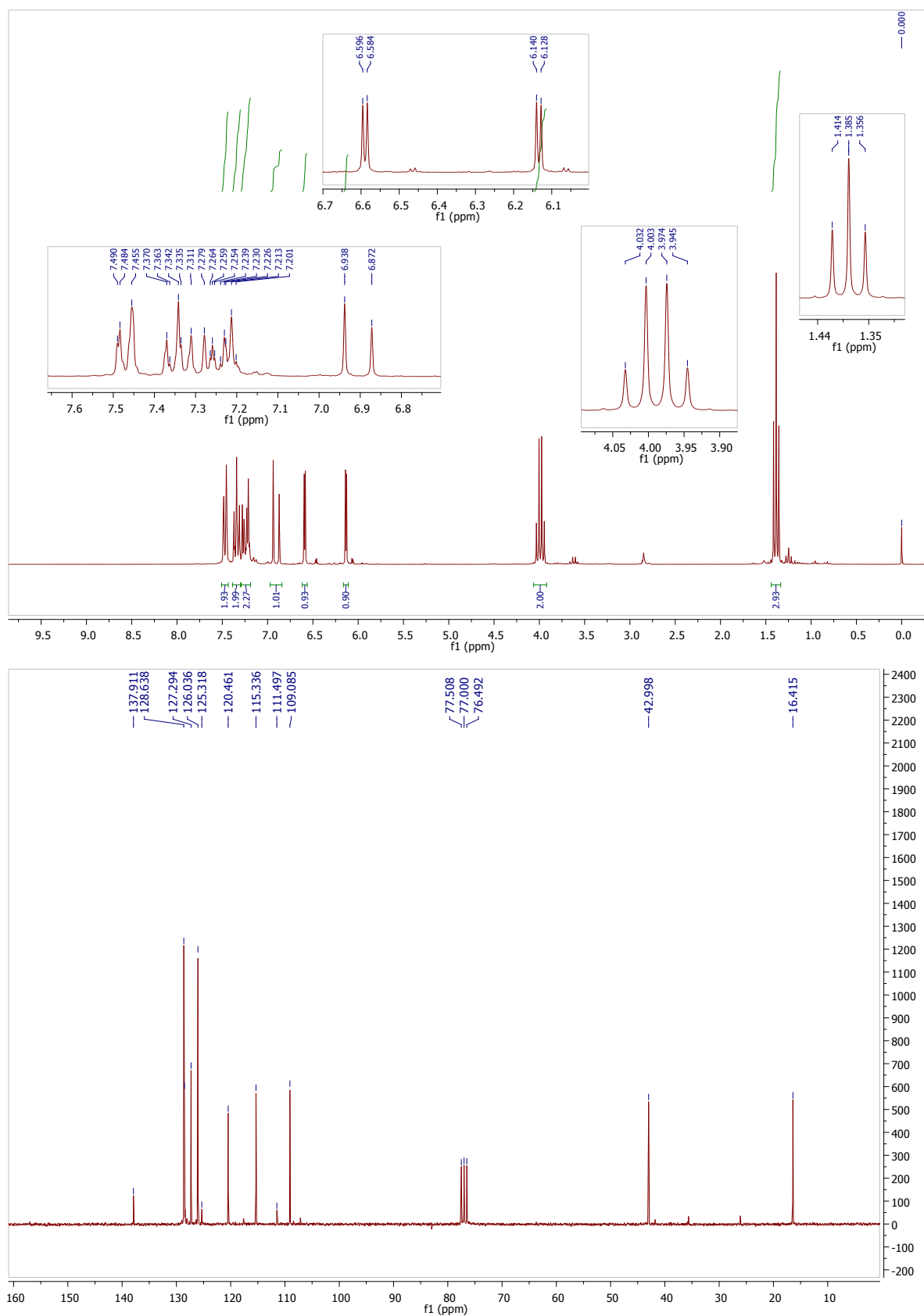
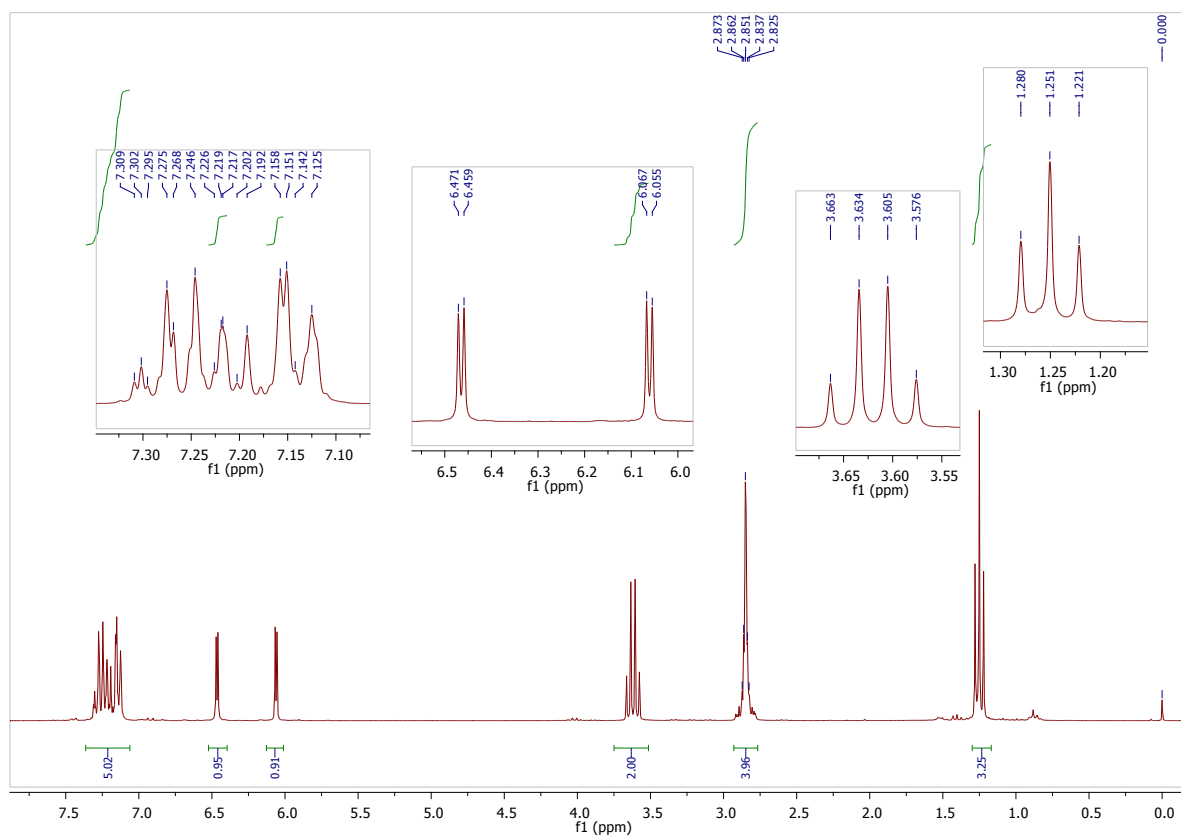
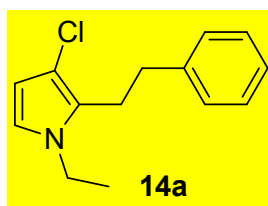


Figure 41. ¹H (250 MHz) and ¹³C (63 MHz) NMR spectra of **13a** in CDCl₃.

3-Chloro-1-ethyl-2-(2-phenylethyl)-1*H*-pyrrole (14a)



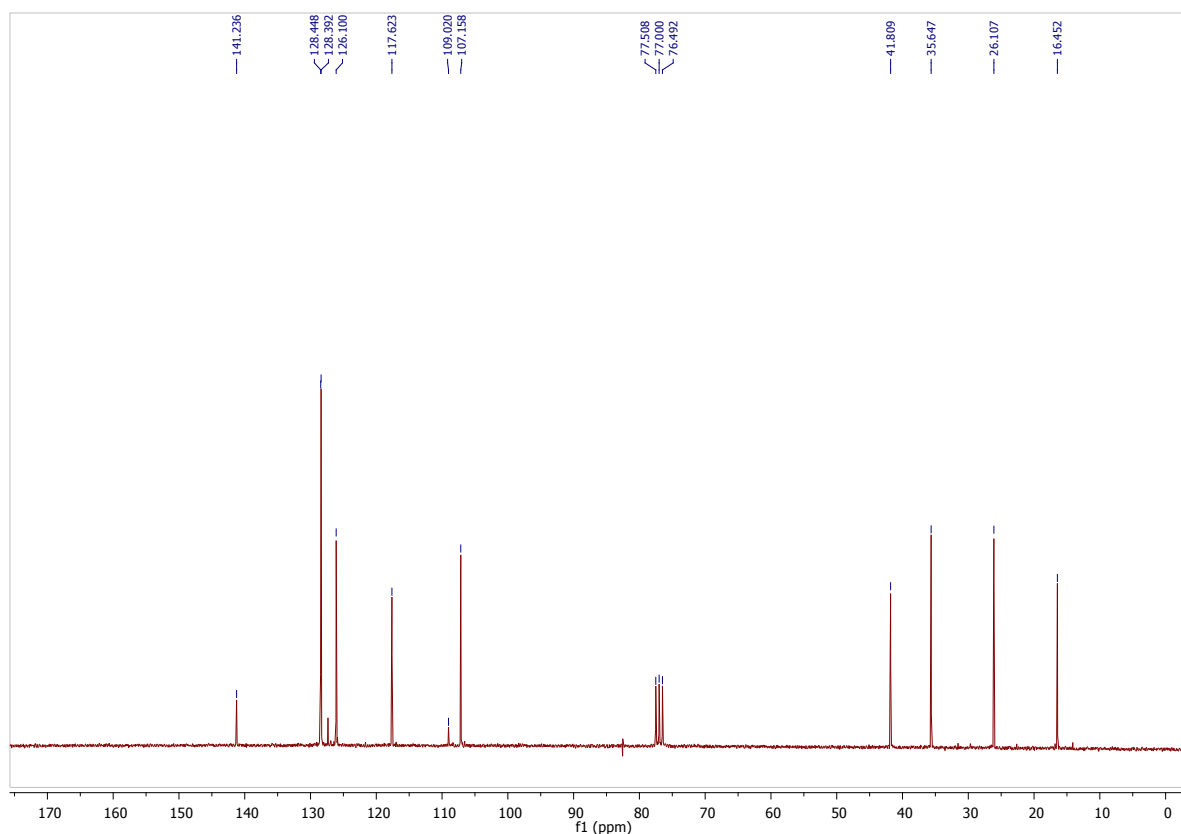


Figure 42. ^1H (250 MHz) and ^{13}C (63 MHz) NMR spectra of **14a** in CDCl_3 .

9. References

1. (a) K. K. Nanda and B. W. Trotter, *Tetrahedron Lett.*, 2005, **46**, 2025; (b) C. Cativiella, J. I. García, J. A. Mayoral and L. Salvatella, *Can. J. Chem.*, 1994, **72**, 308. (c) H. C. Brown, E. J. Mead and C. J. Shoaf, *J. Am. Chem. Soc.*, 1956, **78**, 3613; (d) D. Vuluga, J. Legros, B. Crousse, A. M. Z. Slawin, C. Laurence, P. Nicolet and D. Bonnet-Delpon, *J. Org. Chem.*, 2011, **76**, 1126-1133; (e) C. C. Malakar, S. Stas, W. Herrebout and K. Abbaspour Tehrani, *Chem. Eur. J.*, 2013, **19**, 14263; (f) G. A. Price, A. K. Brisdon, K. R. Flower, R. G. Pritchard and P. Quayle, *Tetrahedron Lett.*, 2014, **55**, 151.
2. C. C. Malakar, B. U. W. Maes and K. Abbaspour Tehrani, *Adv. Synth. Catal.*, 2012, **354**, 3461.