Supplementary Information for:

Palladium Nanoparticles Embedded on Thiourea-Modified Chitosan: A Green and Sustainable Heterogeneous Catalyst for Suzuki Reaction in Water

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General Methods

Absorbance spectroscopy was carried out using a UV-Vis., JASCO V-550 double beam spectrophotometer. Readings were taken at a wavelength range of 250–600 nm. FT-IR analyses were performed using shimandzu FT-IR-410 spectrometer in the range of 4000–400 cm⁻¹. The solid samples were dried and mixed with KBr to form pellets. The XRD pattern of the catalyst samples is measured using a Cu Kα radiation at room temperature. HR-TEM images were taken by JEOL-JEM-2100, High Resolution Transmission Electron Microscopy (HR-TEM). NMR spectra are registered on NMR spectrometer operating at 300 MHz for ¹H and 75 MHz for ¹³C. All ¹H-NMR and ¹³C-NMR spectra are measured in CDCl₃ with TMS as the internal standard. Chemical shifts are expressed in ppm. Purifications of biaryl products by column chromatography are performed on silica gel 60-120 mesh.

Experimental details

a) Preparation of thiourea modified chitosan (TMC)

A mixture of 16.1 g (0.1 mol) chitosan powder, 15.2 g (0.2 mol) ammonium thiocyanate and 150 mL ethanol in a three-neck flask with a magnetic stirring was refluxed for 12 h. After cooling down to room temperature, the precipitate was collected by filtration and repeatedly washed with ethanol, and then was dissolved in 500 mL of 1% (v/v) acetic acid solution. By adding 10% (w/v) NaOH solution into the solution and filtrating, the precipitate was collected and successfully washed with water and finally dried to give 16.5 g of thiourea modified chitosan. FT-IR spectra of chitosan and TMC are shown in figure S3 in SI.

b) Green synthesis of PdNPs supported on TMC

Increasing the concentrations of EA, from 0.1 eq to 0.5 eq (0.1 mmol to 0.5 mmol) in distilled water at pH=7 were prepared and immediately added with 1eq (1 mmol) of palladium (II) acetate solution. The samples were allowed to heat at 50 °C for 3 h, occurrence of color changed from orange red to brown black then UV-Vis., absorption spectroscopy was

recorded. This is the preliminary observations to conform the formation of Pd(0) in water which shown in figure 1. TMC was prepare as reported in literature available and characterized by FT-IR spectroscopy we took 1 g TMC with 100 mL of 0.1 % (v/v) acetic acid and palladium(II) acetate (1 mmol) stirred well for 5 min to the formation of Pd(II)/TMC complex at RT then added 0.5 mmol of fixed concentration of EA in distilled water at 50 °C for 3 h with constant stirring brown black color precipitate were formed contains PdNPs/TMC solid, at which they were filtered, and washed prior to characterized as well as catalyzed for Suzuki reaction in water.

c) Typical procedures for Suzuki cross-coupling reaction

PdNPs/TMC (20 mg), aryl halide (0.5 mmol), phenylboronic acid (0.6 mmol) K_2CO_3 (3.0 mmol), water (3mL) are taken in reaction tube. The tube is heated on a preheated oil bath at a given temperature and magnetically stirred under atmospheric pressure. After the reaction was completed, the mixture was cooled to room temperature. Subsequently, the product and catalyst was filtered from the reaction mixture and washed three times with water (3 x 10 mL). Then the solid biaryl product was washed with diethyl ether and the organic phase is dried over Na₂SO₄. After evaporation of solvent the compound was analyzed by ¹H & ¹³C-NMR.

The reusability of PdNPs/TMC for Suzuki reaction was carried out by taking, iodobenzene (0.06 mL, 0.5 mmol), phenylboronic acid (0.06 g, 0.5 mmol) and K₂CO₃ (0.27 g, 2 mmol) in a reaction tube containing the PdNPs/TMC heterogeneous catayst (0.02 g) in water (2 mL) at 80 °C. After completion of the reaction, the reaction mixture was cooled to room temperature to give a dark solid mass of the catalyst. The resulting mass compound was washed with diethyl ether. The ethereal phase was separated and the resulting dark gray solid catalyst mass was used for another batch of the reaction.



Figure S1 UV-Vis., absorption spectra of PdNPs using EA as a green reducing agent. a) EA (0.5 mmol, 0.151 g); b) Pd(OAc)₂ (1 mmol, 0.224 g); 'c to g' increasing concentration of EA from (0.1-0.5 equiv.) in water at 50 °C.



Figure S2 FT-IR spectra of a) pure EA b) TMC c) PdNPs/TMC.



Figure S3 FT-IR spectra of pure a) chitosan b) TMC c) TMC/PdNPs heterogeneous catalyst.



Figure S4 XRD pattern of PdNPs/TMC catalyst



Figure S5 a) TEM image of cubical and spherical shaped PdNPs observed on TMC solid surface (scale 20 nm).



Figure S6 a) HR-TEM image of single cube (scale 10 nm). c) EDX spectra of the freshly prepared PdNPs/TMC. d) EDX spectra of PdNPs/TMC catalyst after reused.

¹H and ¹³C-NMR Characterisation data



	a = a = a = 1 and $a = a = a = b = a = a = b = a = a = b = a = a$
	general procedure, white solid (91% yield). 'H-NMR (300
	MHz, CDCl ₃ , 25°C, TMS, δ ppm): 3.07 (s, 3H), 8.24 (s, 1H),
	8.14 (d, 1H), 8.00(m, 1H), 7.8 (m, 1H), 7.5 (m, 5H). ¹³ C-
	NMR (75 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 44.50, 125.7,
	127.2, 128.4, 129.8, 132.2, 130.2, 138.9, 142.6.
0	Characterization of N-([1,1'-biphenyl]-3-
	yl)isobutyramide, (Table 2, entry 11) Compound 3e is
	prepared according to the general procedure, white solid
	(94% yield). ¹ H-NMR (300 MHz, CDCl ₃ , 25°C, TMS, δ
	ppm): 3.13(sep, 1H), 1.5 (d, 6H), 8.2 (s, 1H), 7.15(d, 1H),
	7.21 (d. 1H), 7.55 (d. 1H), 7.3-7.4 (m. 5H), ¹³ C-NMR (75
3e, (94%)	MHz CDCl ₂ 25°C TMS δ ppm): 21.5 38.2 129.2 127.1
	127 0 128 6 138 2 141 2 11 3 160 0
	127.9, 120.0, 130.2, 141.2, 11.3, 109.9.
ÇI	Characterization of 5-chloro-2-methyl-1,1'-biphenyl,
	(Table 2, entry 13) Compound 3f is prepared according to
	the general procedure, white solid (80% yield). ¹ H-NMR
	(300 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 2.1(s, 3H), 7.14 (s,
CH ₃	1H) $7.25(d$ 1H) $7.10(d$ 1H) $7.31-7.6$ (m 5H) ^{13}C -NMR
31, (80%)	(75 MHz CDC1, 25°C TMS & ppm): 21.7 126.0 126.5
31, (80%)	(75 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 21.7, 126.0, 126.5,
31, (80%)	(75 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 21.7, 126.0, 126.5, 126.9, 127.3, 127.8, 128.17, 128.37, 128.68, 137.32.
3t, (80%)	 (75 MHz, CDCl₃, 25°C, TMS, δ ppm): 21.7, 126.0, 126.5, 126.9, 127.3, 127.8, 128.17, 128.37, 128.68, 137.32. Characterization of 5-phenylbenzo[d][1,3]dioxole, (Table
3t, (80%)	 (75 MHz, CDCl₃, 25°C, TMS, δ ppm): 21.7, 126.0, 126.5, 126.9, 127.3, 127.8, 128.17, 128.37, 128.68, 137.32. Characterization of 5-phenylbenzo[d][1,3]dioxole, (Table 2, entry 14) Compound 3g is prepared according to the
3t, (80%)	 (75 MHz, CDCl₃, 25°C, TMS, δ ppm): 21.7, 126.0, 126.5, 126.9, 127.3, 127.8, 128.17, 128.37, 128.68, 137.32. Characterization of 5-phenylbenzo[d][1,3]dioxole, (Table 2, entry 14) Compound 3g is prepared according to the general procedure, white solid (90% yield). ¹H-NMR (300
	 (75 MHz, CDCl₃, 25°C, TMS, δ ppm): 21.7, 126.0, 126.5, 126.9, 127.3, 127.8, 128.17, 128.37, 128.68, 137.32. Characterization of 5-phenylbenzo[d][1,3]dioxole, (Table 2, entry 14) Compound 3g is prepared according to the general procedure, white solid (90% yield). ¹H-NMR (300 MHz, CDCl₂, 25°C, TMS δ ppm): 5.94(s, 2H), 6.92 (d, 1H).
3t, (80%)	 (75 MHz, CDCl₃, 25°C, TMS, δ ppm): 21.7, 126.0, 126.5, 126.9, 127.3, 127.8, 128.17, 128.37, 128.68, 137.32. Characterization of 5-phenylbenzo[d][1,3]dioxole, (Table 2, entry 14) Compound 3g is prepared according to the general procedure, white solid (90% yield). ¹H-NMR (300 MHz, CDCl₃, 25°C, TMS, δ ppm): 5.94(s, 2H), 6.92 (d, 1H), 7, 10(d, 1H), 7, 15 (s, 1H), 7, 3-7,5 (m, 5H), ¹³C-NMR (75)
3t, (80%)	 (75 MHz, CDCl₃, 25°C, TMS, δ ppm): 21.7, 126.0, 126.5, 126.9, 127.3, 127.8, 128.17, 128.37, 128.68, 137.32. Characterization of 5-phenylbenzo[d][1,3]dioxole, (Table 2, entry 14) Compound 3g is prepared according to the general procedure, white solid (90% yield). ¹H-NMR (300 MHz, CDCl₃, 25°C, TMS, δ ppm): 5.94(s, 2H), 6.92 (d, 1H), 7.10(d, 1H), 7.15 (s, 1H), 7.3-7.5 (m, 5H). ¹³C-NMR (75 MHz, CDCl₂, 25°C, TMS, δ ppm): 101.0, 107.58, 108.47
3t, (80%)	 (75 MHz, CDCl₃, 25°C, TMS, δ ppm): 21.7, 126.0, 126.5, 126.9, 127.3, 127.8, 128.17, 128.37, 128.68, 137.32. Characterization of 5-phenylbenzo[d][1,3]dioxole, (Table 2, entry 14) Compound 3g is prepared according to the general procedure, white solid (90% yield). ¹H-NMR (300 MHz, CDCl₃, 25°C, TMS, δ ppm): 5.94(s, 2H), 6.92 (d, 1H), 7.10(d, 1H), 7.15 (s, 1H), 7.3-7.5 (m, 5H). ¹³C-NMR (75 MHz, CDCl₃, 25°C, TMS, δ ppm): 101.0, 107.58, 108.47, 127.2, 128.1, 128.7, 120.57, 121.2, 121.5, 122.78, 140.72
3t, (80%)	 (75 MHz, CDCl₃, 25°C, TMS, δ ppm): 21.7, 126.0, 126.5, 126.9, 127.3, 127.8, 128.17, 128.37, 128.68, 137.32. Characterization of 5-phenylbenzo[d][1,3]dioxole, (Table 2, entry 14) Compound 3g is prepared according to the general procedure, white solid (90% yield). ¹H-NMR (300 MHz, CDCl₃, 25°C, TMS, δ ppm): 5.94(s, 2H), 6.92 (d, 1H), 7.10(d, 1H), 7.15 (s, 1H), 7.3-7.5 (m, 5H). ¹³C-NMR (75 MHz, CDCl₃, 25°C, TMS, δ ppm): 101.0, 107.58, 108.47, 127.2, 128.1, 128.7, 129.57, 131.3, 131.5, 133.78, 140.72, 142.52
31, (80%)	 (75 MHz, CDCl₃, 25°C, TMS, δ ppm): 21.7, 126.0, 126.5, 126.9, 127.3, 127.8, 128.17, 128.37, 128.68, 137.32. Characterization of 5-phenylbenzo[d][1,3]dioxole, (Table 2, entry 14) Compound 3g is prepared according to the general procedure, white solid (90% yield). ¹H-NMR (300 MHz, CDCl₃, 25°C, TMS, δ ppm): 5.94(s, 2H), 6.92 (d, 1H), 7.10(d, 1H), 7.15 (s, 1H), 7.3-7.5 (m, 5H). ¹³C-NMR (75 MHz, CDCl₃, 25°C, TMS, δ ppm): 101.0, 107.58, 108.47, 127.2, 128.1, 128.7, 129.57, 131.3, 131.5, 133.78, 140.72, 143.52.
3t, (80%)	 (75 MHz, CDCl₃, 25°C, TMS, δ ppm): 21.7, 126.0, 126.5, 126.9, 127.3, 127.8, 128.17, 128.37, 128.68, 137.32. Characterization of 5-phenylbenzo[d][1,3]dioxole, (Table 2, entry 14) Compound 3g is prepared according to the general procedure, white solid (90% yield). ¹H-NMR (300 MHz, CDCl₃, 25°C, TMS, δ ppm): 5.94(s, 2H), 6.92 (d, 1H), 7.10(d, 1H), 7.15 (s, 1H), 7.3-7.5 (m, 5H). ¹³C-NMR (75 MHz, CDCl₃, 25°C, TMS, δ ppm): 101.0, 107.58, 108.47, 127.2, 128.1, 128.7, 129.57, 131.3, 131.5, 133.78, 140.72, 143.52. Characterization of 2-methoxy-5-phenylpyridine, (Table 2, 128.1, 128.7, 129.57, 131.3, 131.5, 133.78, 140.72, 143.52.
3t, (80%)	 (75 MHz, CDCl₃, 25°C, TMS, δ ppm): 21.7, 126.0, 126.5, 126.9, 127.3, 127.8, 128.17, 128.37, 128.68, 137.32. Characterization of 5-phenylbenzo[d][1,3]dioxole, (Table 2, entry 14) Compound 3g is prepared according to the general procedure, white solid (90% yield). ¹H-NMR (300 MHz, CDCl₃, 25°C, TMS, δ ppm): 5.94(s, 2H), 6.92 (d, 1H), 7.10(d, 1H), 7.15 (s, 1H), 7.3-7.5 (m, 5H). ¹³C-NMR (75 MHz, CDCl₃, 25°C, TMS, δ ppm): 101.0, 107.58, 108.47, 127.2, 128.1, 128.7, 129.57, 131.3, 131.5, 133.78, 140.72, 143.52. Characterization of 2-methoxy-5-phenylpyridine, (Table 2, entry 15) Compound 3h is prepared according to the
3t, (80%)	 (75 MHz, CDCl₃, 25°C, TMS, δ ppm): 21.7, 126.0, 126.5, 126.9, 127.3, 127.8, 128.17, 128.37, 128.68, 137.32. Characterization of 5-phenylbenzo[d][1,3]dioxole, (Table 2, entry 14) Compound 3g is prepared according to the general procedure, white solid (90% yield). ¹H-NMR (300 MHz, CDCl₃, 25°C, TMS, δ ppm): 5.94(s, 2H), 6.92 (d, 1H), 7.10(d, 1H), 7.15 (s, 1H), 7.3-7.5 (m, 5H). ¹³C-NMR (75 MHz, CDCl₃, 25°C, TMS, δ ppm): 101.0, 107.58, 108.47, 127.2, 128.1, 128.7, 129.57, 131.3, 131.5, 133.78, 140.72, 143.52. Characterization of 2-methoxy-5-phenylpyridine, (Table 2, entry 15) Compound 3h is prepared according to the general procedure, white solid (87% vield). ¹H-NMR (300
3t, (80%)	 (75 MHz, CDCl₃, 25°C, TMS, δ ppm): 21.7, 126.0, 126.5, 126.9, 127.3, 127.8, 128.17, 128.37, 128.68, 137.32. Characterization of 5-phenylbenzo[d][1,3]dioxole, (Table 2, entry 14) Compound 3g is prepared according to the general procedure, white solid (90% yield). ¹H-NMR (300 MHz, CDCl₃, 25°C, TMS, δ ppm): 5.94(s, 2H), 6.92 (d, 1H), 7.10(d, 1H), 7.15 (s, 1H), 7.3-7.5 (m, 5H). ¹³C-NMR (75 MHz, CDCl₃, 25°C, TMS, δ ppm): 101.0, 107.58, 108.47, 127.2, 128.1, 128.7, 129.57, 131.3, 131.5, 133.78, 140.72, 143.52. Characterization of 2-methoxy-5-phenylpyridine, (Table 2, entry 15) Compound 3h is prepared according to the general procedure, white solid (87% yield). ¹H-NMR (300 8
31, (80%)	 (75 MHz, CDCl₃, 25°C, TMS, δ ppm): 21.7, 126.0, 126.5, 126.9, 127.3, 127.8, 128.17, 128.37, 128.68, 137.32. Characterization of 5-phenylbenzo[d][1,3]dioxole, (Table 2, entry 14) Compound 3g is prepared according to the general procedure, white solid (90% yield). ¹H-NMR (300 MHz, CDCl₃, 25°C, TMS, δ ppm): 5.94(s, 2H), 6.92 (d, 1H), 7.10(d, 1H), 7.15 (s, 1H), 7.3-7.5 (m, 5H). ¹³C-NMR (75 MHz, CDCl₃, 25°C, TMS, δ ppm): 101.0, 107.58, 108.47, 127.2, 128.1, 128.7, 129.57, 131.3, 131.5, 133.78, 140.72, 143.52. Characterization of 2-methoxy-5-phenylpyridine, (Table 2, entry 15) Compound 3h is prepared according to the general procedure, white solid (87% yield). ¹H-NMR (300 8

	MHz CDC1, 25°C TMS & nnm): 3.85(g. 3H) 5.04 (g. 1H)
	MHZ, CDC1 ₃ , 25 C, 1MS, 0 ppill). 5.85(8, 5H), 5.94 (8, 1H),
	6.85(d, 1H), 7.0 (d, 1H), 7.20-7.56 (m, 5H). ¹³ C-NMR (100
	MHz, CDCl ₃ , 25°C, TMS, δ ppm): 55.83, 101.03, 107.5,
	120.5, 126.7, 127.16, 128.64, 135.52, 146.99, 140.05.
	Characterization of 1-phenylnaphthalene, (Table 2, entry
	16) Compound 3i is prepared according to the general
	procedure, white solid (97% yield). ¹ H-NMR (400 MHz,
	CDCl ₃ , 25°C, TMS, δ ppm): 7.07 (t, 1H), 7.80(d, 1H), 7.85
3i, (97%)	(d, 1H), 8.20 (d, 2H), 7.28 (t, 1H), 8.30 (d, 1H), 7.78 (t, 1H),
	7.50 (m, 5H). ¹³ C-NMR (100 MHz, CDCl ₃ , 25°C, TMS, δ
	ppm): 126.34, 126.73, 127.35, 127.42, 127.97, 128.59,
	128.96, 131.55, 132.55, 133.39, 134.79, 149.48.
0	Characterization of 3-phenylfuran, (Table 2, entry 19)
	Compound $3k$ is prepared according to the general procedure,
	Compound 3k is prepared according to the general procedure, white solid (84% yield). ¹ H-NMR (300 MHz, CDCl ₃ , 25°C,
	Compound 3k is prepared according to the general procedure, white solid (84% yield). ¹ H-NMR (300 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 6.61 (d, 1H), 7.15 (d, 1H), 7.25 (d, 1H), 7.34
3k, (84%)	Compound 3k is prepared according to the general procedure, white solid (84% yield). ¹ H-NMR (300 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 6.61 (d, 1H), 7.15 (d, 1H), 7.25 (d, 1H), 7.34 (m, 5H), 8.12 (s, 1H). ¹³ C-NMR (100 MHz, CDCl ₃ , 25°C,
3k, (84%)	Compound 3k is prepared according to the general procedure, white solid (84% yield). ¹ H-NMR (300 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 6.61 (d, 1H), 7.15 (d, 1H), 7.25 (d, 1H), 7.34 (m, 5H), 8.12 (s, 1H). ¹³ C-NMR (100 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 120.6, 124.8, 126.2, 127.60, 128.37, 128.61,
3k, (84%)	Compound 3k is prepared according to the general procedure, white solid (84% yield). ¹ H-NMR (300 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 6.61 (d, 1H), 7.15 (d, 1H), 7.25 (d, 1H), 7.34 (m, 5H), 8.12 (s, 1H). ¹³ C-NMR (100 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 120.6, 124.8, 126.2, 127.60, 128.37, 128.61, 129.25, 142.53.
3k, (84%)	Compound 3k is prepared according to the general procedure, white solid (84% yield). ¹ H-NMR (300 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 6.61 (d, 1H), 7.15 (d, 1H), 7.25 (d, 1H), 7.34 (m, 5H), 8.12 (s, 1H). ¹³ C-NMR (100 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 120.6, 124.8, 126.2, 127.60, 128.37, 128.61, 129.25, 142.53.
3k, (84%)	Compound 3k is prepared according to the general procedure, white solid (84% yield). ¹ H-NMR (300 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 6.61 (d, 1H), 7.15 (d, 1H), 7.25 (d, 1H), 7.34 (m, 5H), 8.12 (s, 1H). ¹³ C-NMR (100 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 120.6, 124.8, 126.2, 127.60, 128.37, 128.61, 129.25, 142.53. Characterization of 3-phenylthiophene, (Table 2, entry
3k, (84%)	Compound 3k is prepared according to the general procedure, white solid (84% yield). ¹ H-NMR (300 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 6.61 (d, 1H), 7.15 (d, 1H), 7.25 (d, 1H), 7.34 (m, 5H), 8.12 (s, 1H). ¹³ C-NMR (100 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 120.6, 124.8, 126.2, 127.60, 128.37, 128.61, 129.25, 142.53. Characterization of 3-phenylthiophene, (Table 2, entry 21) Compound 31 is prepared according to the general
3k, (84%)	Compound 3k is prepared according to the general procedure, white solid (84% yield). ¹ H-NMR (300 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 6.61 (d, 1H), 7.15 (d, 1H), 7.25 (d, 1H), 7.34 (m, 5H), 8.12 (s, 1H). ¹³ C-NMR (100 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 120.6, 124.8, 126.2, 127.60, 128.37, 128.61, 129.25, 142.53. Characterization of 3-phenylthiophene, (Table 2, entry 21) Compound 3I is prepared according to the general procedure, white solid (92% yield), ¹ H-NMR (300 MHz,
3k, (84%)	Compound 3k is prepared according to the general procedure, white solid (84% yield). ¹ H-NMR (300 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 6.61 (d, 1H), 7.15 (d, 1H), 7.25 (d, 1H), 7.34 (m, 5H), 8.12 (s, 1H). ¹³ C-NMR (100 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 120.6, 124.8, 126.2, 127.60, 128.37, 128.61, 129.25, 142.53. Characterization of 3-phenylthiophene, (Table 2, entry 21) Compound 31 is prepared according to the general procedure, white solid (92% yield). ¹ H-NMR (300 MHz, CDCl ₂ 25°C, TMS δ ppm): 7.59 (s, 1H), 7.15 (d, 1H), 7.21
3k, (84%)	Compound 3k is prepared according to the general procedure, white solid (84% yield). ¹ H-NMR (300 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 6.61 (d, 1H), 7.15 (d, 1H), 7.25 (d, 1H), 7.34 (m, 5H), 8.12 (s, 1H). ¹³ C-NMR (100 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 120.6, 124.8, 126.2, 127.60, 128.37, 128.61, 129.25, 142.53. Characterization of 3-phenylthiophene, (Table 2, entry 21) Compound 31 is prepared according to the general procedure, white solid (92% yield). ¹ H-NMR (300 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 7.59 (s, 1H), 7.15 (d, 1H), 7.21 (d, 1H), 7.32 (m, 5H), ¹³ C NMP (100 MHz, CDCl, 25°C
3k, (84%)	Compound 3k is prepared according to the general procedure, white solid (84% yield). ¹ H-NMR (300 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 6.61 (d, 1H), 7.15 (d, 1H), 7.25 (d, 1H), 7.34 (m, 5H), 8.12 (s, 1H). ¹³ C-NMR (100 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 120.6, 124.8, 126.2, 127.60, 128.37, 128.61, 129.25, 142.53. Characterization of 3-phenylthiophene, (Table 2, entry 21) Compound 3I is prepared according to the general procedure, white solid (92% yield). ¹ H-NMR (300 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 7.59 (s, 1H), 7.15 (d, 1H), 7.21 (d, 1H), 7.32 (m, 5H). ¹³ C-NMR (100 MHz, CDCl ₃ , 25°C,
3k, (84%)	Compound 3k is prepared according to the general procedure, white solid (84% yield). ¹ H-NMR (300 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 6.61 (d, 1H), 7.15 (d, 1H), 7.25 (d, 1H), 7.34 (m, 5H), 8.12 (s, 1H). ¹³ C-NMR (100 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 120.6, 124.8, 126.2, 127.60, 128.37, 128.61, 129.25, 142.53. Characterization of 3-phenylthiophene, (Table 2, entry 21) Compound 31 is prepared according to the general procedure, white solid (92% yield). ¹ H-NMR (300 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 7.59 (s, 1H), 7.15 (d, 1H), 7.21 (d, 1H), 7.32 (m, 5H). ¹³ C-NMR (100 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 110.36, 115.22, 123.1, 127.5, 128.7, 132.99,
3k, (84%)	Compound 3k is prepared according to the general procedure, white solid (84% yield). ¹ H-NMR (300 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 6.61 (d, 1H), 7.15 (d, 1H), 7.25 (d, 1H), 7.34 (m, 5H), 8.12 (s, 1H). ¹³ C-NMR (100 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 120.6, 124.8, 126.2, 127.60, 128.37, 128.61, 129.25, 142.53. Characterization of 3-phenylthiophene, (Table 2, entry 21) Compound 31 is prepared according to the general procedure, white solid (92% yield). ¹ H-NMR (300 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 7.59 (s, 1H), 7.15 (d, 1H), 7.21 (d, 1H), 7.32 (m, 5H). ¹³ C-NMR (100 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 110.36, 115.22, 123.1, 127.5, 128.7, 132.99, 137.05, 143.30.

	Characterization of 2-methyl-5-phenylthiophene, (Table
CH ₃	2, entry 22) Compound 3m is prepared according to the
	general procedure, white solid (91% yield). ¹ H-NMR (300
3m, (91%)	MHz, CDCl ₃ , 25°C, TMS, δ ppm): 1.25 (s, 3H), 7.10 (d, 1H),
	7.34 (d, 1H), 7.97 (t, 1H) 7.43 (t, 2H), 7.59 (d, 2H). ¹³ C-NMR
	(100 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 29.52, 110.36, 115.2,
	123.15, 126.95, 127.10, 128.3, 132.9, 137.9, 143.30.
_	Characterization of 4-ethoxy-3-fluoro-1 1'-hinhenvl
F CH ₂ CH ₃	
F CH ₂ CH ₃	(Table 2, entry 23) Compound 3n is prepared according to
F CH ₂ CH ₃	(Table 2, entry 23) Compound 3n is prepared according to the general procedure, white solid (99% yield). ¹ H-NMR
F CH ₂ CH ₃	(Table 2, entry 23) Compound 3n is prepared according to the general procedure, white solid (99% yield). ¹ H-NMR (300 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 4.13(q, 2H), 1.46 (t,
F CH ₂ CH ₃	(Table 2, entry 23) Compound 3n is prepared according to the general procedure, white solid (99% yield). ¹ H-NMR (300 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 4.13(q, 2H), 1.46 (t, 3H), 7.21 (s, 1H), 7.50 (d, 1H), 7.60 (d, 1H), 7.33 (m, 5H).
F CH ₂ CH ₃ O 3n, (99 %)	(Table 2, entry 23) Compound 3n is prepared according to the general procedure, white solid (99% yield). ¹ H-NMR (300 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 4.13(q, 2H), 1.46 (t, 3H), 7.21 (s, 1H), 7.50 (d, 1H), 7.60 (d, 1H), 7.33 (m, 5H). ¹³ C-NMR (100 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 14.56,
F CH ₂ CH ₃ 0 3n, (99 %)	(Table 2, entry 23) Compound 3n is prepared according to the general procedure, white solid (99% yield). ¹ H-NMR (300 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 4.13(q, 2H), 1.46 (t, 3H), 7.21 (s, 1H), 7.50 (d, 1H), 7.60 (d, 1H), 7.33 (m, 5H). ¹³ C-NMR (100 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 14.56, 64.64, 114.5, 122.3, 126.3, 127.06, 128.65, 133.97, 139.43,
F CH ₂ CH ₃ 0 3n, (99 %)	(Table 2, entry 23) Compound 3n is prepared according to the general procedure, white solid (99% yield). ¹ H-NMR (300 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 4.13(q, 2H), 1.46 (t, 3H), 7.21 (s, 1H), 7.50 (d, 1H), 7.60 (d, 1H), 7.33 (m, 5H). ¹³ C-NMR (100 MHz, CDCl ₃ , 25°C, TMS, δ ppm): 14.56, 64.64, 114.5, 122.3, 126.3, 127.06, 128.65, 133.97, 139.43, 146.15, 150.99, 154.24.

¹H & ¹³C-NMR spectra



Figure S6 ¹H-NMR spectrum of 1,1'-biphenyl 3 (Table 2)



Figure S7 ¹³C-NMR spectrum of 1,1'-biphenyl **3** (Table 2)



Figure S8 ¹H-NMR spectrum of 3,4-dimethoxy-1,1'-biphenyl **3a** (Table 2)



Figure S9 ¹³C-NMR spectrum of 3,4-dimethoxy-1,1'-biphenyl **3a** (Table 2)



Figure S10 ¹H-NMR spectrum of 3-nitro-1,1'-biphenyl 3b (Table 2)



Figure S11 ¹³C-NMR spectrum of 3-nitro-1,1'-biphenyl **3b** (Table 2)



Figure S11 ¹³C-NMR spectrum of 1,1':4',1"-terphenyl 3c (Table 2)



Figure S12 ¹³C-NMR spectrum of 1,1':4',1"-terphenyl 3c (Table 2)



Figure S13 ¹H-NMR spectrum of 3-(methylsulfonyl)-1,1'-biphenyl 3d (Table 2)



Figure S14 ¹³C-NMR spectrum of 3-(methylsulfonyl)-1,1'-biphenyl **3d** (Table 2)



Figure S15 ¹H-NMR spectrum of N-([1,1'-biphenyl]-3-yl)isobutyramide 3e (Table 2)



Figure S16 ¹³C-NMR spectrum of N-([1,1'-biphenyl]-3-yl)isobutyramide 3e (Table 2)



Figure S17 ¹H-NMR spectrum of 5-chloro-2-methyl-1,1'-biphenyl **3f** (Table 2)



Figure S18 ¹³C-NMR spectrum of 5-chloro-2-methyl-1,1'-biphenyl 3f (Table 2)



Figure S19 ¹H-NMR spectrum of 5-phenylbenzo[d][1,3]dioxole **3g** (Table 2)



Figure S20 ¹³C-NMR spectrum of 5-phenylbenzo[d][1,3]dioxole **3g** (Table 2)



Figure S21 ¹H-NMR spectrum of 2-methoxy-5-phenylpyridine 3h (Table 2)



Figure S22 ¹³C-NMR spectrum of 2-methoxy-5-phenylpyridine 3h (Table 2)



Figure S23 ¹³C-NMR spectrum of 1-phenylnaphthalene **3i** (Table 2)



Figure S24 ¹³C-NMR spectrum of 1-phenylnaphthalene 3i (Table 2)



Figure S25 ¹³C-NMR spectrum of 3-phenylfuran 3k (Table 2)



Figure S26 ¹³C-NMR spectrum of 3-phenylfuran 3k (Table 2)



Figure S27 ¹³C-NMR spectrum of 3-phenylthiophene 3l (Table 2)



Figure S28 ¹³C-NMR spectrum of 3-phenylthiophene 3l (Table 2)



Figure S29 ¹³C-NMR spectrum of 2-methyl-5-phenylthiophene **3m** (Table 2)

Figure S30 ¹³C-NMR spectrum of 2-methyl-5-phenylthiophene 3m (Table 2)



Figure S31 ¹H-NMR spectrum of 4-ethoxy-3-fluoro-1,1'-biphenyl 3n (Table 2)



Figure S32 ¹³C-NMR spectrum of 4-ethoxy-3-fluoro-1,1'-biphenyl 3n (Table 2)