Electronic Supplementary Information (ESI)

A Highly Sensitive and Selective "Turn-on" Fluorescent Probe for Hypochlorous Acid Monitoring

Ye-Xin Liao, Mao-Die Wang, Kun Li, * Zhao-Xuan Yang, Ji-Ting Hou, Ming-Yu Wu, Yan-Hong Liu, and Xiao-Qi Yu*

Key Laboratory of Green Chemistry and Technology (Ministry of Education), College of Chemistry, Sichuan University, Chengdu, 610064, P.R. China.

Contents

- 1. General experimental section
- 2. Synthetic procedures
- 3. Effect of different solvents
- 4. UV-visible Absorbance Measurements
- 5. High resolution mass spectrum (HRMS) of reaction system between 3 and

HCIO/CIO-

- 6. Effect of pH value
- 7. Detection limit of 3 to HClO/ClO-
- 8. Kinetic property of the reaction between 3 and ClO-
- 9. Fluorescence quantum yields
- **10.** Cells culture and imaging

1. General experimental section

1.1 Materials:

Unless otherwise indicated, all reagents and solvents were obtained from commercial suppliers and used without further purification. Column chromatography was performed on silica gel (Qingdao haiyang) 200-300 mesh or basic aluminum oxide (for chromatography use). All solvents used in spectra test systems were chromatographically pure. Ultrapure water was used throughout. Stock solutions of compound **3** and **1** were prepared by dissolving the corresponding compound in chromatographically pure DMSO. H₂O₂ stock solution was prepared by diluting 30% H₂O₂ solution, and the concentration was determined from absorption at λ =240 nm (ϵ =43.6 M⁻¹·cm⁻¹). BuOOH was prepared by diluting 70% 'BuOOH solution. OH was generated by Fenton reaction between FeSO₄ and H₂O₂, and concentration of OH was determined by Fe²⁺. 'BuOO' was generated by reaction between FeSO₄ and 'BuOOH, and concentration of 'BuOO' was determined by Fe²⁺. $^{1}O_{2}$ was generated by the reaction of H₂O₂ with Na₂MoO₄¹. O₂⁻ was generated from KO₂ solid diluted in DMSO. ONOO- was generated by the reaction of H₂O₂ and NaNO₂² and stocked at -20 °C, the concentration was determined from absorption at λ =302 nm (ϵ =1670 M⁻¹·cm⁻¹) in 0.1 M NaOH solution³.ClO⁻ was prepared by diluting 5% NaClO aqueous solution, and the concentration was determined from absorption at λ =292 nm (ϵ =350 M⁻¹·cm⁻¹)³. The fluorescence quantum yields in solution were calculated using Fluorescein as a standard material.

1.2 Instruments

¹H NMR and ¹³C NMR spectra were recorded on a Bruker AMX-400 with chemical shifts expressed in parts per million (in deuteriochloroform or DMSO-d₆, Me₄Si as internal standard). Fluorescence spectra were determined using a FluoroMax-4 Spectrofluoro photometer (HORIBA Jobin Yvon). UV/Vis absorption spectra were determined by a Hitachi PharmaSpec UV-1900 UV-Vis spectrophotometer. Mass spectral data were recorded on a Finnigan LCQDECA and a Bruker Daltonics Bio TOF mass spectrometer. High performance liquid chromatography (HPLC) were perform on a Waters e2695 Separatins Module using Waters 2998 PDA detector equipped with an Symmetry C18 column(4.6 X 150 mm, 5 μ m), CH₃OH:H₂O=95:5(v/v) were used as eluents with a flow rate of 1 ml/min. 280nm was used as wavelength. TLC analyses were performed on silica gel GF 254. The pH values were determined by a Leici pH3c (digital display) pH meter.

2. Synthetic procedures

2.1 Preparation of 1-(benzo[d]thiazol-2-yl)naphthalen-2-ol (1)

2-aminothiophenol (15.6 mmol, 1.95 g) and 2-hydroxy-1-naphthaldehyde (15.8 mmol, 2.72 g) were dissolved in DMF (50 mL), then sodium metabisulfite (16.0 mmol, 3.05 g) was added and the reaction mixture was reflux for 2 h. After cooling to room temperature, the mixture was poured into 100 mL water, and then extracted with ethyl acetate (EA) for three times. EA layer was collected and dried by anhydrous Na₂SO₄, solvent was removed under reduced pressure to give the crude product. Then crude product was purified by column chromatography using silica gel (200-300 mesh) and DCM/petroleum ether as a fluent to give a yellow green solid (0.80 g, yield 18.5%). ¹H NMR (400 MHz, DMSO-d₆) δ 11.04 (s, 1 H), 8.27-8.25 (d, *J*=8.56 Hz, 1 H), 8.19-8.14 (m, 2 H), 8.01-7.98 (d, *J*=8.96 Hz, 1 H), 7.92-7.90 (d, *J*=8.08 Hz, 1 H), 7.60-7.57 (m, 1 H), 7.53-7.48 (m, 2 H), 7.40-7.34 (m, 2 H).¹³C NMR (100 MHz, DMSO-d₆) δ 163.75, 154.83, 152.22, 135.37, 132.41, 132.35, 128.35, 127.86, 127.63, 126.06, 125.17, 123.91, 123.38, 122.64,

121.79, 118.28, 112.09. HRMS (ESI) calcd for $C_{17}H_{12}NOS^+$, $[M+H]^+$, m/z 278.0634 ; found, m/z 278.0617.

2.2 Preparation of 2-(2-(4-nitrophenoxy)naphthalen-1-yl)benzo[d]thiazole (2)

60% NaH (220 mg) was added in portion into a solution of **1** (5 mmol, 1.14 g) in anhydrous DMF (2 mL) in an ice-bath. And the resulting solution was kept and stirred for 30 min. Then 4-chloronitrobenzene (5 mmol, 0.79 g) was added and the mixture was heated to 120 °C and stirred overnight. After cooling to room temperature, 50 mL water was poured into the reaction mixture and extracted by DCM thrice. Organic layer was collected, washed with saturated brine and then dried by anhydrous Na₂SO₄. Solvent was removed under reduced pressure. The residue was then purified by column chromatography using silica gel (200-300 mesh) using DCM/petroleum ether as the fluent and finally yielded a yellow solid (0.26 g, yield 12.6%). ¹H NMR (400 MHz, DMSO-d₆) δ 8.24-8.21 (d, *J*=8.88 Hz, 1 H), 8.24-8.13 (m, 5 H), 8.02-8.00 (d, *J*=7.44 Hz, 1 H), 7.66-7.50 (m, 5 H), 7.20-7.18 (d, *J*=8.92 Hz, 2 H). ¹³C NMR (100 MHz, DMSO-d₆) δ 162.41, 161.04, 152.67, 150.08, 142.56, 135.55, 133.39, 132.15, 131.06, 128.54, 128.33, 126.44, 126.15, 125.78, 125.02, 123.19, 122.19, 121.63, 120.30, 117.53. HRMS (ESI) calcd for C₂₃H₁₅N₂O₃S⁺, [M+H]⁺, m/z 399.0798; found, m/z 399.0796.

2.3 Preparation of 4-((1-(benzo[d]thiazol-2-yl)naphthalen-2-yl)oxy)aniline (3)

Compound **2** (398 mg, 1 mmol) and 20% (w/w) Pd/C powder (Pd, 10 wt% on carbon powder) was dissolved into 20 mL CH₃OH. The resulting solution was stirred at ambient temperature overnight in hydrogen atmosphere. After the reaction finished, Pd/C was removed by using a pad of Celite, and the filtrate was concentrated in vacuo to afford crude product. The crude product was further purified using basic aluminum oxide with DCM/petroleum ether as the fluent and finally yielded a pale yellow solid (327 mg, yield 89%).¹H NMR (400 MHz, DMSO-d₆): δ 8.20-8.17 (m, 2 H), 8.11-8.09 (d, *J*=9.12 Hz, 1 H), 8.06-8.04 (d, *J*=8.40 Hz, 1 H), 8.01-7.99 (d, *J*=7.40 Hz, 1 H), 7.62-7.51 (m, 4 H), 7.18-7.16 (d, *J*=9.08 Hz, 1 H), 6.82-6.80 (m, 2 H), 6.61-6.58 (m, 2 H), 5.03 (s, 2 H). ¹³C NMR (100 MHz, DMSO-d₆): δ 164.48, 154.32, 152.67, 146.05, 145.61, 135.68, 132.27, 132.12, 129.34, 128.33, 127.95, 126.18, 125.48, 124.98, 124.49, 122.99, 121.99, 120.25, 117.75, 117.63, 114.85. HRMS (ESI) calcd for C₂₃H₁₇N₂OS⁺, [M+H]⁺, m/z 369.1056; found, m/z 369.1059.

3. Effect of different solvents

Different solvents including water, THF, CH₃CN, CH₃OH, DMF, DMSO and water with 1 mM CTAB were used as the test solvents and fluorescence emission intensity at 465 nm were recorded.



Figure S1 Flourescence responds of the reaction between 3 (5 μ M) and ClO⁻ (50 μ M) in HEPES buffer (0.5% DMSO, 20 mM, pH 7.40) at ambient temperature for 30 min. Fluorescence intensity was collected at 465 nm. λ_{ex} =350 nm, slits: 5/5 nm.

4. UV-visible Absorbance and Fluorescence Emission Measurements

UV spectra (Figure 2a) of 1 (20 μ M), 3 (20 μ M) and the reaction solution of 3 (20 μ M) and ClO⁻ (200 μ M) were recorded in DMSO/water (2/98, v/v) buffered by 20 mM HEPES at pH 7.40 (Figure S2a). Fluorescence emission spectra of 1 (5 μ M) and the reaction solution of 3 (5 μ M) and ClO⁻ (50 μ M) were collected in DMSO/water (1/99, v/v) buffered by 20 mM HEPES at pH 7.40 (Figure S2b). All the reaction solution was placed at room temperature for 30 min.



Figure S2 a) Absorption spectra of 1 (blue line), 3 (black line) and reaction between 3 and HCIO (red line) at ambient temperature for 30 min. b) Fluorescence emission spectra of compound 1 (blue line) and the reaction of 3 and HCIO (red line).

5. High resolution mass spectrum (HRMS) of reaction system between 3 and HClO/ClO-

25 mL PBS solution (with 2% DMSO, v/v) with probe **3** (20 μ M), HClO/ClO⁻ (200 μ M) and CTAB (1 mM) was prepare and kept at ambient temperature for 30 min. Then the reaction solution was extracted by DCM for thrice. DCM layer was collected and solvent was removed. Residue was resolved by DCM: MeOH= 10: 1 (v/v) and flowed on a pad of silica gel (200-300 mesh), and the effluent was collected. Solvent was removed and the residue was analyzed directly by HRMS. The ES- result was shown as follow:



Effect of pH value

Various pH solutions were prepared by adjusting the pH values of HEPES buffer (20 mM) using 50% NaOH solution and 20% HCl solution. The solutions of **1**, **3** and the reaction between **3** and ClO⁻ in different pH conditions (including: 3.02, 4.02, 5.26, 6.01, 6.52, 6.99, 7.40, 8.01, 8.53, 9.01, 10.15, 10.91 and 12.03) were prepared and kept at ambient temperature for 30 min, and spectra were recorded then by excited at 350 nm, using 5/5 nm as the excitation and emission slits.



Figure S4 Effect of pH values to the relative fluorescence intensity of 5 μ M 1 (•), 5 μ M 3 (\blacktriangle) and reaction between 5 μ M 3 and 50 μ M ClO- in water (0.5% DMSO in volum) buffered by 20 mM HEPES at pH 7.40 (\blacksquare) for 30 min at ambient temperature. (The investigated pH values include 3.02, 4.02, 5.26, 6.01, 6.52, 6.99, 7.40, 8.01, 8.53, 9.01, 10.15, 10.91 and 12.03).

6. Detection limit of 3 to HClO/ClO-

The detection limit (D) of **3** to HClO/ClO⁻ was calculated base on the equation as follow: $D=3\sigma/k$, σ stands for the standard deviation of the blank measurement of **3**'s fluorescence intensity at 465



nm within 10 times; And k stands for a slope between fluorescence intensity versus HClO/ClO⁻ concentration in the fluorescence titration curve of **3** in the present of HClO/ClO⁻ (0-20 μ M).

Figure S5 Fluorescence changes with various concentrations of HCIO/CIO⁻ at 465 nm in the reaction between **3** and HCIO/CIO⁻ at ambient temperature.

7. Kinetic property of the reaction between 3 and ClO-

Kinetic property of the reaction between **3** (5 μ M) and ClO⁻ (50 μ M) was studied. Intensity was collected at 465 nm while the excitation wavelength λ_{ex} =350 nm, slits: 5/5 nm. Solution was kept at ambient temperature.



Figure S6. Time-dependent fluorescence changes of 3 (5 μ M) in present of HCIO (50 μ M) at ambient temperature. λ_{ex} =350 nm, slits: 5/5 nm, emission intensity was collected at λ_{em} =465 nm.

8. Fluorescence quantum yields

Fluorescein in 0.1 N NaOH aqueous solution was used as the standard (quantum yield Φ_s =0.85). The quantum yields of **3** and the reaction solution between **3** and HClO/ClO⁻ in water (HEPES buffer, 20 mM, pH 7.40, 0.5% DMSO) was calculated based on the equation followed:

$$\Phi_{\rm u} = \Phi_{\rm s} * \frac{F_{\rm u}}{F_{\rm s}} * \frac{A_{\rm s}}{A_{\rm u}}$$

Where Φ is fluorescence quantum yield; F is integrated area under the corrected emission spectra; *A* is the absorbance at the excitation wavelength; the subscript s stands for the standard and u

stands for unknown.

9. Cells culture and imaging

HeLa cells were cultured in Dulbecco's modified Eagle medium (DMEM) containing 10% fetal bovine serum and 1% Antibiotic-Antimycotic at 37 °C in a 5% CO2/95% air incubator. For fluorescence imaging, cells (4×10^3 /well) were passed on a culture dish and incubated for 24h. Cells were washed twice with PBS, and then incubated with 5 μ M 3 (PBS buffer, 1 mM CTAB) for 30 min at 37 °C. After washed twice with PBS, cells were incubated with 50 µM NaClO (PBS buffer as solvent) for another 20 min. Finally cells were washed three times and the confocal fluorescent images were captured, excitation wavelength: 405 nm, fluorescence emission was collected from 440 nm to 500 nm.

References

- 1 H. Zhu, J. L. Fan, J. Y. Wang, H. Y. Mu and X. J. Peng, J. Am. Chem. Soc., 2014, 136, 12820-12823.
- 2 J. S. Beckman, J. Chen, H. Ischirpoulos, J. P. Crow, *Methods enzymol.*, 1994, 233, 229-240.
 3 M. Abo, Y. Urano, K.Hanaoka, T. Terai, T. Komatsu, T. Nagano, *J. Am. Chem. Soc.*, 2011, 133, 10629-10637.



Figure S8. ¹³C NMR spectrum of **1** in DMSO-d₆.



Figure S10. ¹³C NMR spectrum of **2** in DMSO-d₆.



Figure S11. ¹H NMR spectrum of **3** in DMSO-d₆.



210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 f1 (ppm)



