An in silico and chemical approach towards small protein production and

application in phosphoproteomics

Ana M.G.C. Dias ^{a,b}, Olga Iranzo ^{b*} and Ana C.A. Roque ^{a*}

Received (in XXX, XXX) Xth XXXXXXXX 20XX, Accepted Xth XXXXXXXX 20XX DOI: 10.1039/b000000x

^a UCIBIO, REQUIMTE, Departamento de Química, Faculdade de Ciências e Tecnologia, Universidade Nova de Lisboa, Campus Caparica, 2829-516 Caparica, Portugal, Fax: (+351) 21 294 8550; E-mail: <u>cecilia.roque@fct.unl.pt</u>

^b Aix Marseille Université, Centrale Marseille, CNRS, iSm2 UMR 7313, 13397 Marseille, France; E-mail: olga.iranzo@univ-amu.fr, Previous address: Instituto de Tecnologia Química e Biológica António Xavier, Universidade Nova de Lisboa, 2780-157 Oeiras, Portugal

Supplementary Information:

Content List:

	Pages
S1 - In silico studies	
S1.1 Molecular Dynamics Simulations - Stability studies - hPin1_WWdomain structures	2
S1.2 Molecular Docking - Affinity Studies - hPin1_WWmutated	3
S2 Peptide Solid-phase Chemical Synthesis and Characterization	
S2.1 Conditions for the standard and pseudoproline units protocol for peptide chemical	
synthesis	4
S2.2 Reverse-Phase HPLC hPin1_WWmutated	5
S2.3 - Electrospray Ionization-Mass spectra (ESI-MS)	6
ESI-MS hPin1_WWmutated	6

S1 - In silico Studies:

S1.1. - Molecular Dynamics simulations - Stability studies - hPin1_WW domain structures:



The protocols for preparation of the structure for in silico studies can be found in the main text.

Figure S1. Molecular Dynamics results analysis for the 3 structures: hPin1_WWnative, hPin1_WWnativeFL and hPin1_WWmutated: A) RMSD (Root Mean Square Deviation); B) H-Bond interactions, only shown the most frequently observed over the time of the simulation; C) Identification of the most frequent H-Bond interactions observed in B (in hPin1_WWmutated structure). (Figure was produced using PyMol software).

S1.2. - Molecular Docking - Affinity studies - hPin1_WWmutated:

Ligand	Docking Binding Energy (Kcal/mol)	Estimated Affinity Constant (M ⁻¹)	Literature Affinity Constant (M ⁻¹)	Source			
РРРРҮР	-10.09	2.49 x 10 ⁷	n.d.	[a]			
VPR pT PV	-7.31	2.28 x 10 ⁵	2.04 x 10 ⁵	Native ligand [b]			
VPR T PV	-5.94	2.26 x 10 ⁴	n.d.	-			
[a] Native ligand shorter version of YAP65 WW domain (Group I) ¹							

Table SI. Molecular Docking Results: hPin WWmutated and different peptides.

[b] Ligand described by Verdecia, M *et al* 2 with highest affinity constant with hPin1_WW. n.d. - non determined



Figure S2. Molecular Docking results analysis, where hPin1_WWmutated was used as receptor and peptides, PPPPYP, VPRpTPV and VPRTPV were used as ligands. These conformations correspond to the values estimated in Table S1. In yellow the Tyr-27 and Trp-38 that form the "X-P-groove region". A) Best docking conformation of hPin1_WWmutated against PPPPYP, B) Best docking conformation of hPin1_WWmutated against VPRpTPV; and C) Best docking conformation of hPin1_WWmutated against VPRpTPV. (Figures were produced using PyMol software).

S2. Peptide Solid-phase Chemical Synthesis and Characterization

S2.1 - Conditions for the standard and pseudoproline units protocol for peptide chemical synthesis

Method		Reagents	Power (watts)	Temperature (°C)	Reaction Time (sec)	
Standard	Resin	Rink Amide MBHA resin 100-200 mesh	-	-	-	
	Deprotection	20% piperidine/DMF	35	75	217	
	Coupling	HOBT/HBTU/DIEA/DMF	25	75	304	
	Coupling Arg (double coupling)	HOBT/HBTU/DIEA/DMF	$0^{a)}$ and $25^{b)}$	50	$1805^{a)}$ and $305^{b)}$	
	Coupling His, Cys, Asp	HOBT/HBTU/DIEA/DMF	0	50	733	
	Capping	10% Acetic Anhydrid	1	75	126	
Pseudoproline Units	Resin	NovaPEG Rink amide resin	-	-	-	
	Deprotection	20% piperidine/DMF	35	75	217	
	Deprotection after 23 residue	20% piperidine/DMF	35	75	341	
	Coupling	HBTU/DIEA/DMF	25	75	304	
	Coupling Arg (double coupling)	HBTU/DIEA/DMF	${0^{\mathrm{a})} \operatorname{and} \atop 25^{\mathrm{b})}}$	50	$1950^{a)}$ and $400^{b)}$	
	Coupling after 23 residue	HBTU/DIEA/DMF	25	75	605	
	Capping after 23 residue	10% Acetic Anhydrid	1	75	126	
	Note: Arginine residue requires double coupling: a) 1st coupling and b) 2nd coupling.					

Table SII. Microwave Synthesis conditions for hPin1_WWmutated

S2.2 Reverse-Phase HPLC hPin1_WWmutated:



Figure S3. Reverse-Phase HPLC chromatograms: A) Preparative HPLC of the crude product of hPin1_WWmutated obtained by standard SPPS (without pseudoproline); B) Preparative HPLC of crude product of hPin1_WWmutated obtained by SPPS with pseudoproline strategy; C) Analytical HPLC chromatogram of purified hPin1_WWmutant. HPLC conditions used are described in the main text.

S2.3 - Electrospray Ionization-Mass spectrometry (ESI-MS)

The ESI-MS spectra of hPin1_WWmutated (Figure S3) were acquired in positive mode on a mass spectrometer API-linear Ion trap (PO03MS), model LTQ (Thermo-Finnigan) by direct infusion at a flow rate of 5 μ L/min. All the spectra were obtained by the Mass Spectrometry Laboratory of the Analytical Services Unit at the Instituto de Tecnologia Química e Biológica António Xavier, Universidade Nova de Lisboa.





Figure S4. ESI-MS spectrum of hPin1_WWmutated (Acetyl - CADEEKLPPGWEKRMSRSSGRVYYFNHITNASQWE RPG-NH₂, Molecular Formula: $C_{198}H_{296}N_{60}O_{59}S_2$) in positive mode: m/z calcd for [hPin1_WWmutated + 6H]⁺⁶ = 755.18 Da, m/z found: 755.2; m/z calcd for [hPin1_WWmutated + 5H]⁺⁵ = 906.01 Da, m/z found: 906.0 and m/z calcd for [hPin1_WWmutated + 4H]⁺⁴ = 1132.26 Da, m/z found: 1132.3.

References:

1. E. K. Koepf, H. M. Petrassi, M. Sudol and J. W. Kelly, Protein Science, 1999, 8, 841-853.

2. M. A. Verdecia, M. E. Bowman, K. P. Lu, T. Hunter and J. P. Noel, Nat Struct Mol Biol, 2000, 7, 639-643.