

## Supporting Information

### Electrosynthesis of cyclic carbonates from CO<sub>2</sub> and epoxides on reusable copper nanoparticles cathode

Laxia Wu <sup>a, b</sup>, Hengpan Yang <sup>b</sup>, Huan Wang <sup>b</sup>, Jiaxing Lu <sup>\*b</sup>

<sup>a</sup> School of Chemistry and Chemical Engineering, Anqing Normal University, Anqing 246011, China

<sup>b</sup> Shanghai Key Laboratory of Green Chemistry and Chemical Processes, School of Chemistry and Molecular Engineering, East China Normal University, Shanghai 200062, China

#### List of Contents

<b>1. Materials and Instrument</b> .....	2
<b>2. General procedure</b> .....	2
<b>3. Characterization of Cu nanoparticles</b> .....	3
<b>4. Characterization Data for All Products</b> .....	3

## 1. Materials and Instruments

All reagents were used as received except for products in Table 2, entries 2-8.

Galvanostatic electrosynthesis was performed using a direct current-regulated power supply (HY3002D, HYelec®, China).

<sup>1</sup>H-NMR spectra were recorded on an AVANCE 500 (500 MHz, Bruker, Germany) or Ascend 400 (400 MHz, Bruker, Germany) spectrometer in CDCl<sub>3</sub> with Me<sub>4</sub>Si as an internal standard.

The product yield was determined by Gas Chromatography-Flame Ionization Detector (GC-FID) (SHIMADZU, GC-2014).

Microstructure and morphology of Cu NPs were analyzed using Hitachi S-4800 field emission Scanning Electron Microscope (FE-SEM) and TECNAI G<sup>2</sup>F30 Transmission Electron Microscope (TEM).

X-ray diffraction (XRD) patterns were recorded by a Ultima IV X-ray powder diffractometer using Cu K $\alpha$  radiation (k= 1.5406 Å).

N<sub>2</sub> adsorption was carried out at 77 K on a BELSORP-MAX instrument after outgassing the samples for 10 h under vacuum at 573 K.

## 2. General procedure

### 2.1 Prepare compacted Cu NPs electrode

**100 nm:** 5 g CuSO<sub>4</sub>·5H<sub>2</sub>O was dissolved in 100 mL deionized water. After 10 min stirring, 50 mL hydrazine hydrate solution (10%) was added, and the mixture was stirred at 25 °C for 4 h. Cu was precipitated immediately. The precipitate was filtered, washed with 10 mL water and 10 mL anhydrous ethanol for 4 times respectively, and dried for 12 h at 35 °C under vacuum. Cu NPs powder was pressed into coin and used as cathode for electrosynthesis of cyclic carbonates from CO<sub>2</sub> and epoxide.

**300 nm:** The same method except that 25 mL hydrazine hydrate solution (20%) was added.

**50 nm:** 10 mL 0.25 M copper acetate and urea aqueous solution were mixed and transferred to a Teflon-lined reactor. The reactor was kept in an oven at 130 °C for 1 h. The reaction solution was centrifuged. Dark precipitate was obtained after being washed three times with distilled water and drying at 60 °C for 12 h. The dark precipitate was reduced in H<sub>2</sub> atmosphere at 130 °C for 3 h.

**10 nm:** The method was the same as 50 nm except for 0.025 M copper acetate and urea aqueous

solution.

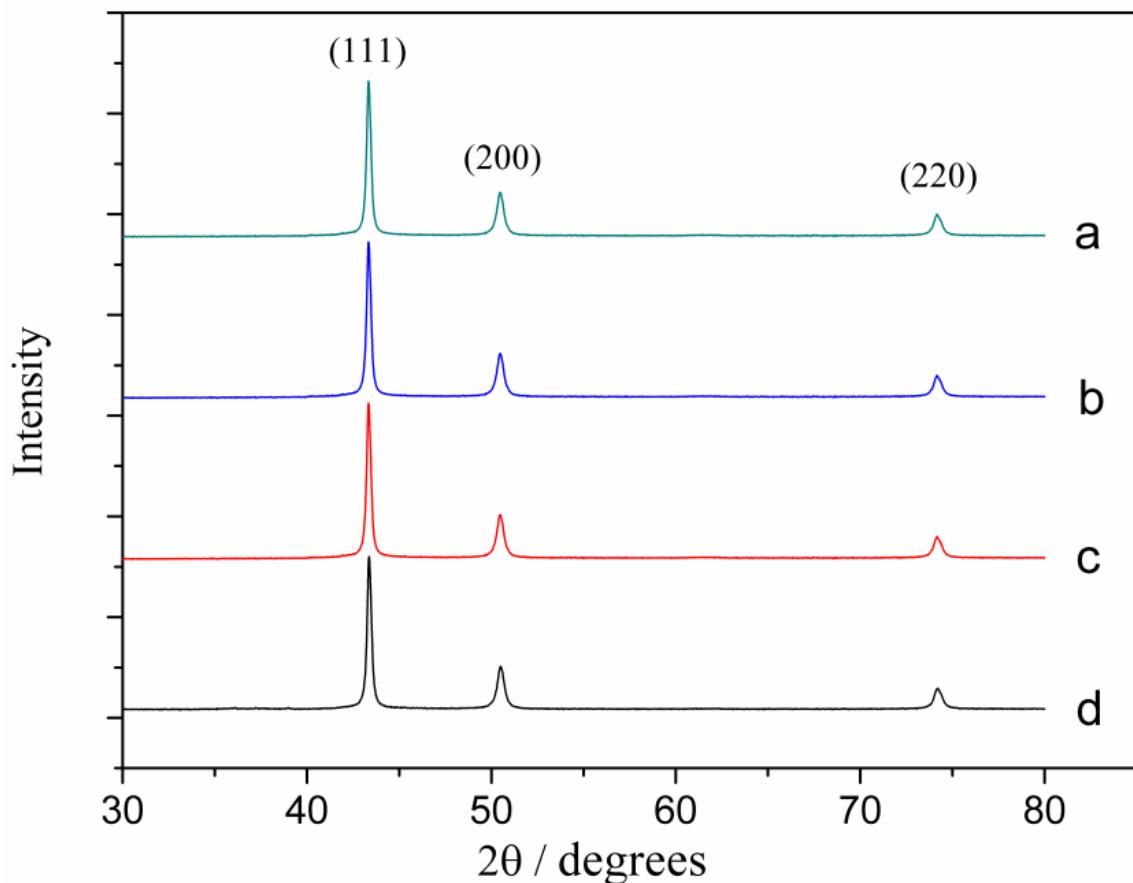


**Fig. S1** Cu NPs before (left) and after (right) press.

## 2.2 General electrosynthesis

A typical galvanostatic electrolysis was carried out in a mixture of propylene oxide (100 mM), supporting electrolyte TEAI (0.1 M) and 10 mL MeCN saturated with CO<sub>2</sub> (1 atm) in an undivided glass cell equipped with a sacrificial magnesium (Mg) rod anode and Cu NPs cathode (d = 2 cm). Continuous CO<sub>2</sub> flow was maintained throughout the duration of the whole electrolysis process. The pure products were isolated by column chromatography using petroleum ether/ethyl acetate mixture as an eluent.

## 3. Characterization of Cu nanoparticles



**Fig. S2** X-ray diffraction (XRD) patterns of Cu NPs in the size range of ~300 nm (a), 100 nm (b), 50 nm (c) and 10 nm (d).

#### 4. Characterization Data for All Products

**4-(Chloromethyl)-1,3-dioxolan-2-one 2b<sup>[1]</sup>** GC-MS (m/z, %) 132 (M<sup>+</sup>, 1), 87 (100), 64 (5), 43 (29), 28 (16); <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>): δ 3.72-3.78 (m, 2H), 4.43 (q, J = 8 Hz, 1H), 4.61(t, J = 8 Hz, 1H), 4.97-5.01 (m, 1H).

**4-Ethyl-1,3-dioxolan-2-one 2c<sup>[2]</sup>** GC-MS (m/z, %) 116 (M<sup>+</sup>, 4), 87 (64), 71 (4), 57 (15), 43 (100); <sup>1</sup>H-NMR (500 MHz, CDCl<sub>3</sub>): δ 1.04 (t, J = 8 Hz, 3H), 1.73-1.86 (m, 2H), 4.10 (t, J = 8 Hz, 1H), 4.54 (t, J = 8 Hz, 1H), 4.67 (q, J = 7 Hz, 1H).

**4-Propyl-1,3-dioxolan-2-one 2d** GC-MS (m/z, %) 129 (1), 102 (1), 87 (67), 71 (20), 57 (40), 43 (100); <sup>1</sup>H-NMR (500 MHz, CDCl<sub>3</sub>): δ 0.99 (t, J = 7 Hz, 3H), 1.42-1.44 (m, 1H), 1.50-1.53 (m, 1H), 1.66-1.69 (m, 1H), 1.80-1.82 (m, 1H), 4.08 (t, J = 8 Hz, 1H), 4.54 (t, J = 8 Hz, 1H), 4.70-4.76 (m, 1H).

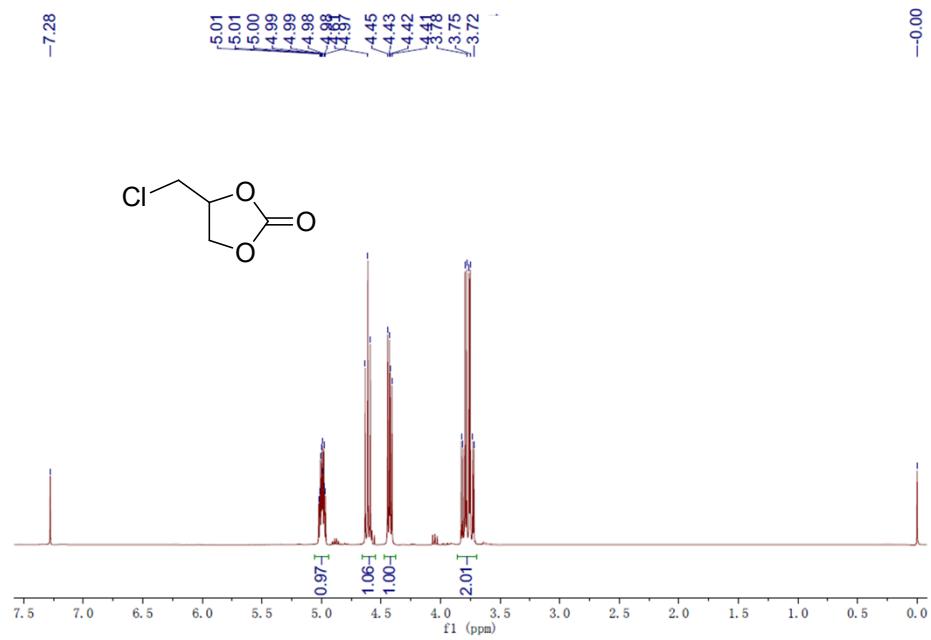
**4,5-Dimethyl-1,3-dioxolan-2-one 2e<sup>[3]</sup>** GC-MS (m/z, %) 116 (M<sup>+</sup>, 3), 101 (4), 86 (1), 73 (3), 57 (7), 43 (100), 38 (6), 28 (78); <sup>1</sup>H-NMR (500 MHz, CDCl<sub>3</sub>): δ 1.46 (q, J = 6 Hz, 6H), 4.32-4.35 (m, 2H).

**Tetrahydro-3aH-cyclopenta[d]-1,3-dioxol-2-one 2f** GC-MS (m/z, %) 113 (2), 97 (12), 83 (37), 69 (100), 55 (77), 41 (89), 28 (40); <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>): δ 1.41-1.47 (m, 2H), 1.59-1.66 (m, 2H), 1.88 (q, J = 4 Hz, 4H), 4.67-4.72 (m, 2H).

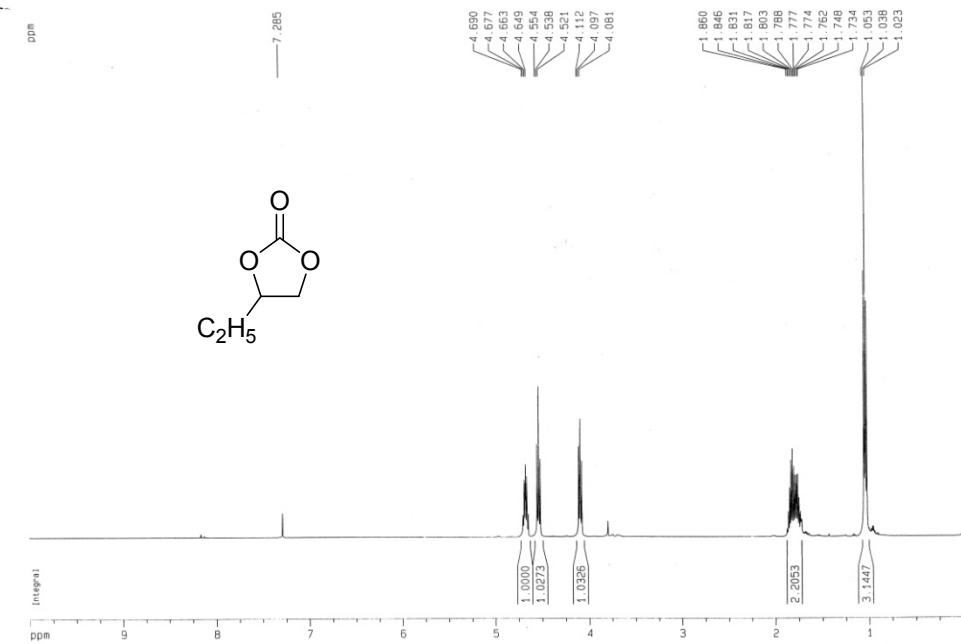
**Hexahydrobenzo[d]-1,3-dioxol-2-one 2g<sup>[4]</sup>** GC-MS (m/z, %) 128 (M<sup>+</sup>, 7), 99 (4), 83 (14), 69 (3), 55 (100), 41 (20), 28 (19); <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>): δ 1.68-1.82 (m, 4H), 2.15-2.18 (m, 2H), 5.10 (d, J = 4 Hz, 2H)

**4-Phenyl-1,3-dioxolan-2-one 2h<sup>[5]</sup>** GC-MS (m/z, %) 164 (M<sup>+</sup>, 66), 131 (1), 119 (15), 105 (32), 90 (100), 78 (71), 65 (19), 51 (25), 39 (13); <sup>1</sup>H-NMR (500 MHz, CDCl<sub>3</sub>): δ 4.36 (t, J = 9 Hz, 1H), 4.83 (t, J = 9 Hz, 1H), 5.69 (t, J = 8 Hz, 1H), 7.27-7.48 (m, 5H).

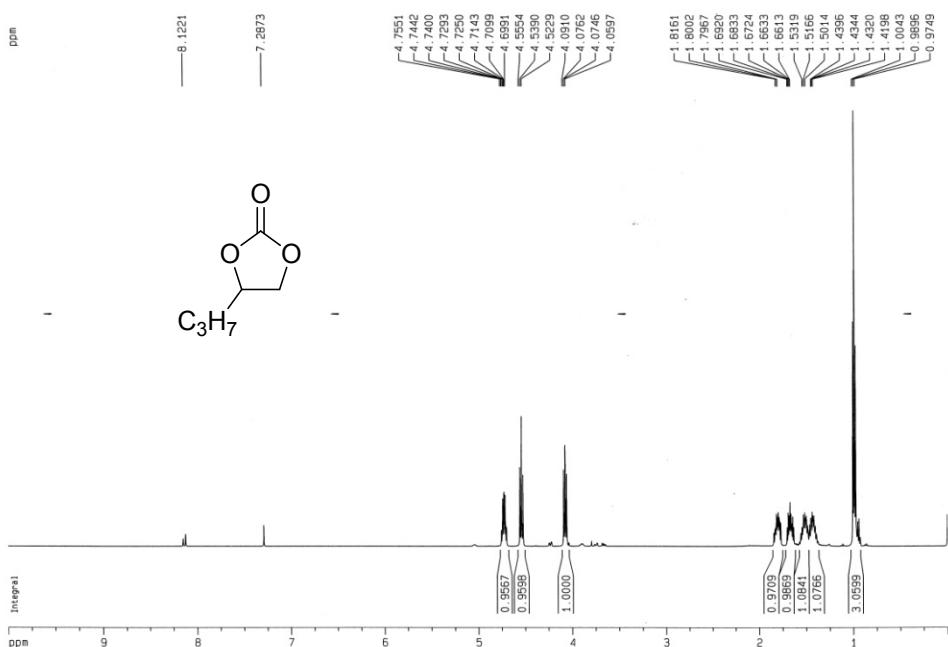
#### <sup>1</sup>H-NMR Spectra



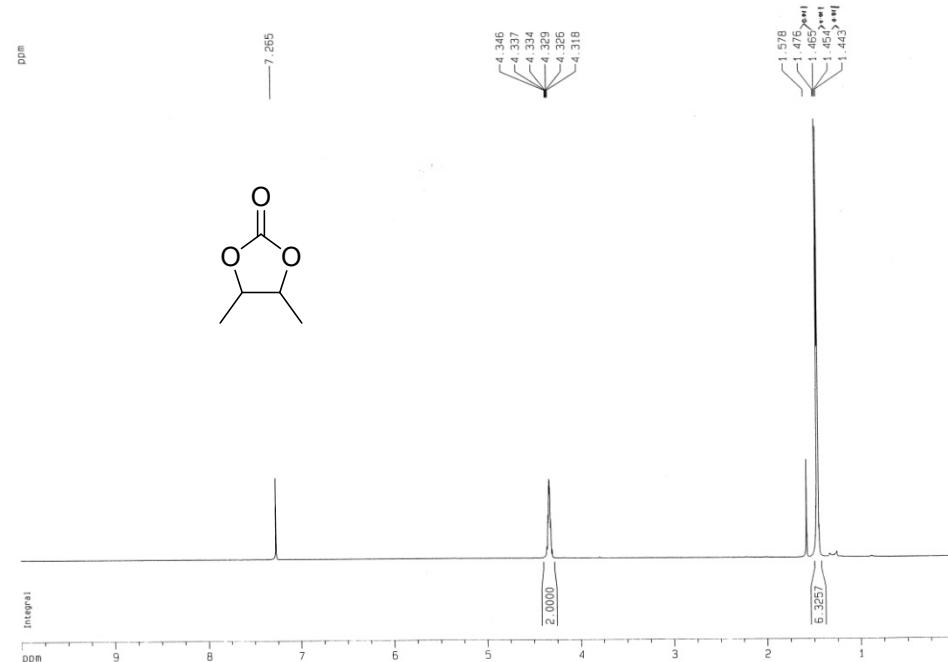
**Fig. S3.**  $^1\text{H}$ -NMR spectrum of **2b**.



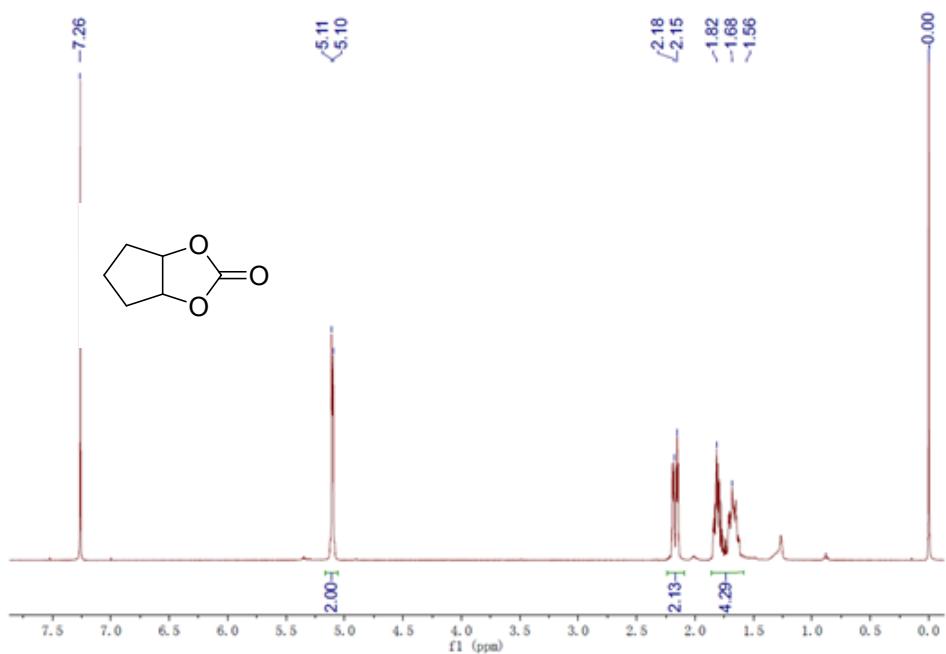
**Fig. S4.**  $^1\text{H}$ -NMR spectrum of **2c**.



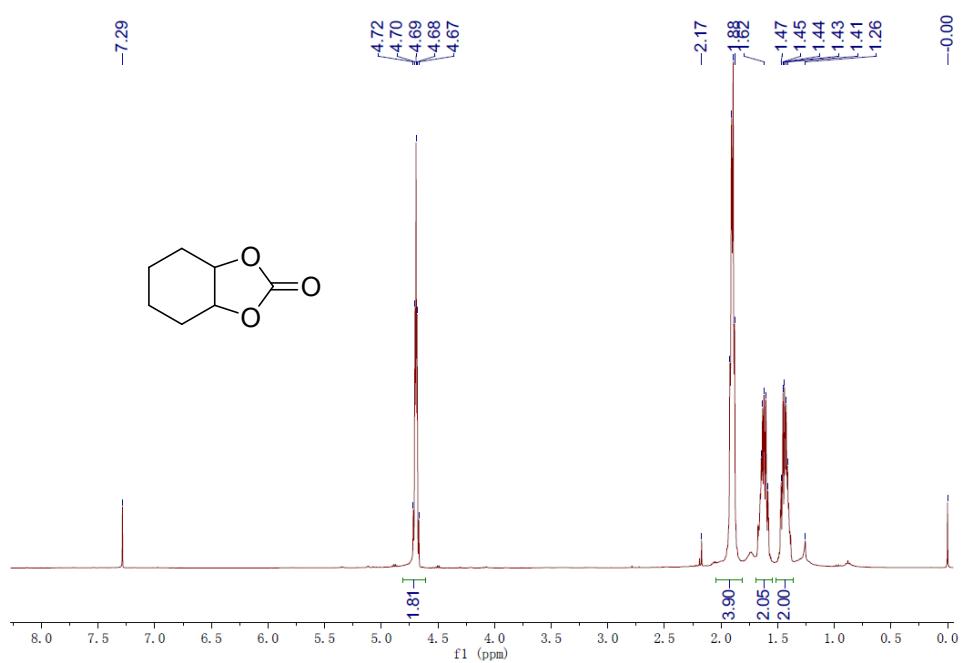
**Fig. S5.** <sup>1</sup>H-NMR spectrum of **2d**.



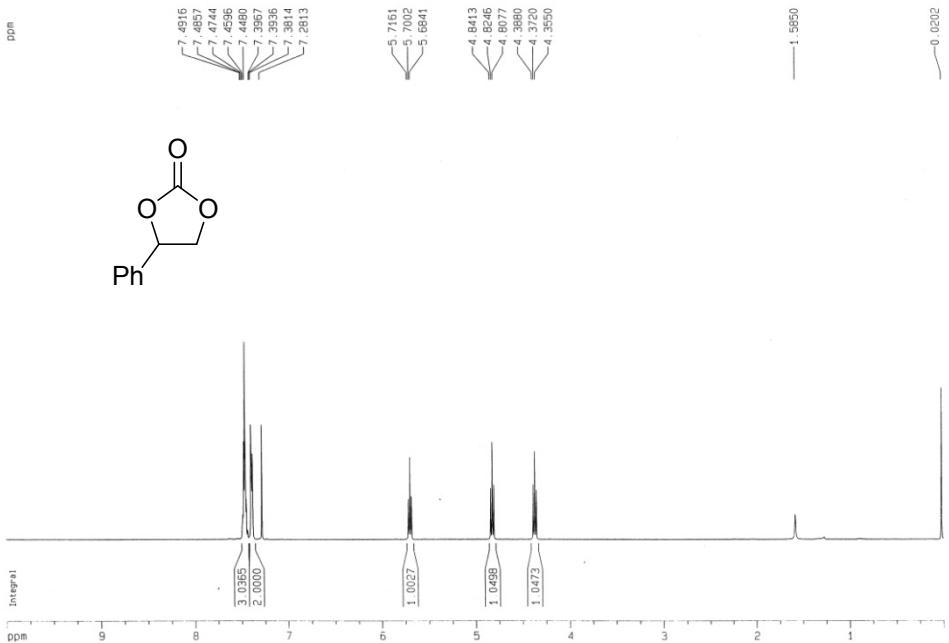
**Fig. S6.** <sup>1</sup>H-NMR spectrum of **2e**.



**Fig. S7.**  $^1\text{H}$ -NMR spectrum of **2f**.



**Fig. S8.**  $^1\text{H}$ -NMR spectrum of **2g**.



**Fig. S9.**  $^1\text{H}$ -NMR spectrum of **2h**.

## References

- [1] J. L. Song, Z. F. Zhang, S. Q. Hu, T. B. Wu, T. Jiang, B. X. Han, *Green Chem.* 2009, **11**, 1031-1036.
- [2] X. B. Lu, Y. J. Zhang, B. Liang, X. Li, H. Wang, *J. Mol. Catal. A: Chem.* 2004, **210**, 31-34.
- [3] M. A. Casadei, S. Cesa, M. Feroci, A. Inesi, *New J. Chem.*, 1999, **23**, 433-436.
- [4] Y. X. Zhou, S. Q. Hu, X. M. Ma, S. G. Liang, T. Jiang, B. X. Han, *J. Mol. Catal. A: Chem.* 2008, **284**, 52-57.
- [5] N. Eghbali, C. J. Li, *Green Chem.*, 2007, **9**, 213–215.