Chemical Science

A Redox-Active Reverse Donor-Acceptor Bistable [2]Rotaxane

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Electronic Supplementary Information

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Professor J Fraser Stoddart Department of Chemistry Northwestern University 2145 Sheridan Road Evanston, Illinois 60208-3113 Tel: (+1)-847-491-3793 Fax: (+1)-847-491-1009 *E-Mail: stoddart@northwestern.edu* General Methods and Materials: Starting materials and reagents were purchased from Aldrich or Fisher and used as received. Compound $6^{[S1]}$ and $9 \cdot PF_6^{[S2]}$ were prepared following procedures reported in the literature. All reactions were performed under an atmosphere of nitrogen and in dry solvents, unless otherwise noted. Analytical thin-layer chromatography (TLC) was performed on aluminum sheets, precoated with silica gel 60-F254 (Merck 5554). Flash chromatography was carried out using silica gel 60 (Silicycle) as the stationary phase. HPLC was performed on a preperative RP-HPLC instrument, using a C18 column. ¹H and ¹³C NMR spectra and Double Quantum Field Correlation Spectroscopy (DQF-COSY) were recorded on either a Bruker Avance 500 MHz, or a Bruker Avance 600 MHz spectrometer. Chemical shifts are reported in ppm relative to the signals corresponding to the residual non-deuterated solvents (CDCl₃: δ 7.26 ppm, CD₃CN: δ 1.94 ppm, d_7 -DMF: 8.02 ppm). Variable temperature ¹H NMR experiments were calibrated temperature calibrated using neat methanol. High resolution electrospray ionization (HR-ESI) mass spectra were measured on Agilent 6210 LC-TOF with Agilent 1200 HPLC introduction. Highresolution matrix-assisted laser desorption/ionization (MALDI) mass spectra were measured on a Bruker Autoflex III mass spectrometer. Electrochemical experiments were carried out at room temperature in argon-purged aqueous solutions, with a Gamry Reference 600 potentiostat interfaced to a PC. Cyclic voltammetry experiments were performed using a glassy carbon working electrode $(0.071 \text{ cm}^2, \text{Cypress Systems})$. Its surface was polished routinely with 0.05 µm alumina-water slurry on a felt surface immediately before use. The counter electrode was a Pt coil and the reference electrode was a standard Ag/AgCl electrode. The concentration of the sample and supporting electrolyte (TBAPF₆) were 1 x 10^{-3} mol L⁻¹ and 0.1 mol L⁻¹, respectively. SEC experiments were made in a BASi thin layer quartz glass cell with an optical pathlength of 1 mm using a Pt gauze working electrode, a Pt wire counter electrode, and a Ag/AgCl reference electrode.

Experimental Section

Compound 1: A Grignard reagent was prepared by mixing magnesium turnings (3.0 g, 123 mmol) and 4-ethylbromobenzene (21.7 g, 117 mmol) in dry THF (100 mL). Methyl-4-methoxybenzoate (8.4 g, 50 mmol) in dry THF (100 mL) was added slowly to this mixture over 20 min. The resulting reaction mixture was heated under reflux for 24 h before being cooled to RT, poured into 300 g ice-water containing conc. H₂SO₄. The aqueous solution was extracted with CHCl₃. The organic layer was washed with saturated NaHCO₃ (1 x 100 mL), dried (Na₂SO₄) and evaporated under vacuum to yield pure **1** (14 g, 80%). ¹H NMR (500 MHz, CDCl₃, 298 K): δ = 7.28–6.92 (m, 12H), 3.88 (s, 3H), 2.73 (q, *J* = 7.2 Hz, 4H), 1.32 (t, *J* = 7.2 Hz, 6H) ppm. ¹³C NMR (125 MHz, CDCl₃, 298 K): δ = 158.7, 144.8, 143.2, 139.7, 131.5, 129.8, 129.3, 127.9,127.5, 113.2, 81.6 55.4, 28.6, 15.6 ppm. ESI-MS calcd for *m/z* = 329.20 [*M* – OH₂]⁺, found *m/z* = 329.35.

Compound 2: A mixture of **1** (20 g, 58 mmol), phenol (108 g, 1.16 mol) was heated to 80°C before the addition of concentrated HCl (3 mL) under N₂ atmosphere. The reaction mixture was further heated at 100°C for 24 h. After the reaction completed, the mixture was diluted with PhMe, washed with 0.5 N NaOH (7 x 75 mL) and dried (Na₂SO₄). The solvent was evaporated in vacuo. The crude product was purified by column chromatography [SiO₂: EtOAc / Hexane (1:3)] to afford pure **2** (15.9 g, 65%). ¹H NMR (500 MHz, CDCl₃, 298 K): δ = 7.28–6.60 (m, 18H), 3.68 (s, 3H), 2.55 (q, *J* = 7.3 Hz, 4H), 1.12 (t, *J* = 7.3Hz, 6H) ppm. ¹³C NMR (125 MHz, CDCl₃, 298 K): δ = 157.3, 155.5, 153.3, 144.6, 141.5, 132.3, 130.9, 129.7, 126.7, 120.8, 115.3, 114.1, 112.6, 63.1, 28.2, 15.5 ppm. ESI-MS calcd for *m/z* = 406.22 [*M* – OH₂]⁺, found *m/z* = 407.25.

Compound 3: A 1 M solution of BBr₃ in CH₂Cl₂ (1.5 mL, 15.6 mmol) was added dropwise to a solution of **2** (3.0 g, 7.1 mmol) in CH₂Cl₂ (50 mL) at -78° C. The mixture was warmed up to the RT and stirred for 48 h under N₂. The reaction was quenched by addition of MeOH (5 mL) and H₂O

(150 mL). The organic layer was separated and the aqueous layer was washed with CHCl₃ (3 x 75 mL). The combined organic layer was dried (Na₂SO₄) and then evaporated in vacuo. The crude product was purified by column chromatography [SiO₂: EtOAc / Hexane (1:3)] to afford pure **3** (2.8 g, 99%). ¹H NMR (500 MHz, CDCl₃, 298 K): δ = 7.07–6.70 (m, 16H), 2.67 (q, *J* = 7.2 Hz, 4H), 1.28 (t, *J* = 7.2 Hz, 6H) ppm. ¹³C NMR (125 MHz, CDCl₃, 298 K): δ = 153.5, 144.6, 141.6, 139.8, 132.6, 131.1, 126.9, 114.3, 28.3, 15.3 ppm. ESI-MS calcd for *m*/*z* = 391.21 [*M* – OH₂]⁺, found *m*/*z* = 391.14.

Compound 5: Solid K₂CO₃ (6.4 g, 47 mmol), 5-bromopentanol (3.14 g, 18.8 mmol), 18C6 (100 mg) were added to a solution of diol **3** (1.9 g, 4.7 mmol) in dry MeCN (75 mL), and the solution was heated under reflux in an N₂ atmosphere for 12 h. The reaction was cooled to RT, and diluted with H₂O (50 mL), and then extracted with CH₂Cl₂. The organic layer was dried (Na₂SO₄) and filtered. The solvent was removed under reduced pressure to obtain the crude **4** product, which was subjected to tosylation without further purification. Et₃N (2 mL) and DMAP (50 mg) were added to a solution of the extended diol (1 g, 1.72 mmol) in dry CH₂Cl₂ (200 mL) and were followed by the dropwise addition of TsCl (297 mg, 1.56 mmol) in dry CH₂Cl₂ (20 mL) over 30 min at 0°C. The solution was stirred at 0°C and slowly warmed to RT overnight. The solvent was removed in vacuo and the resulting crude oil was purified by column chromatography [SiO₂: CH₂Cl₂ / MeOH (98:2)] to afford pure **5** as a colorless oil (1.5 g, 43%). ¹H NMR (500 MHz, CDCl₃, 298 K): δ = 7.82 (d, *J* = 8 Hz, 2H), 7.34 (d, *J* = 8 Hz, 2H), 7.15–6.74 (m, 16H), 4.08 (t, *J* = 6.5 Hz, 2H), 3.97 (t, *J* = 6.5 Hz, 2H), 3.90 (t, *J* = 6.5 Hz, 2H), 3.70 (t, *J* = 6.5 Hz, 2H), 2.65 (q, *J* = 7.6 Hz, 4H), 2.45 (s, 3H), 1.86–1.48 (m, 12 H), 1.26 (t, *J* = 7.6 Hz, 6H). ¹³C NMR (125 MHz, CDCl₃, 298 K): δ = 156.8, 156.7, 144.8, 144.7, 141.4, 139.6, 139.5, 133.0, 132.1, 132.0, 130.9, 129.9, 127.9, 126.8, 113.1, 113.0, 70.5, 67.6,

67.2, 62.9, 62.8, 32.5, 29.1, 28.7, 28.6, 28.3, 22.4, 2.2, 21.6, 15.4 ppm. ESI-MS calcd for $m/z = 757.35 [M + Na]^+$, found m/z = 757.31.

Compound 7: Et₃N (0.1 mL) and DMAP (10 mg) were added to a solution of compound **6** (100 mg, 0.17 mmol) in dry CH₂Cl₂ (25 mL) and were followed by the dropwise addition of TsCl (33 mg, 0.17 mmol) in dry CH₂Cl₂ (20 mL) over 30 min at RT. The solution was stirred overnight. The reaction mixture was diluted with H₂O (20 mL), and then extracted with CH₂Cl₂. The combined organic layers was dried (Na₂SO₄), and the solvent was removed under reduced pressure to obtain (98 mg, 78%) compound **7** in pure form as a tan solid. ¹H NMR (500 MHz, CDCl₃, 298 K): δ = 7.92 (d, *J* = 8.2 Hz, 2H), 7.32 (d, *J* = 8.2 Hz, 2H), 7.23 (d, *J* = 8.9 Hz, 6H), 7.08 (m, 8H), 6.7 (d, *J* = 9.9 Hz, 2H), 4.05 (t, *J* = 7.1 Hz, 2H), 3.87 (t, *J* = 6.1 Hz, 2H), 2.42 (s, 3H), 1.72 (m, 4H), 1.49 (m, 2H), 1.30 (s, 27H) ppm. ¹³C NMR (125 MHz, CDCl₃, 298 K): δ = 157.1, 148.8, 145.2, 144.6, 140.0, 133.6, 132.7, 131.2, 130.3, 128.3, 124.5, 113.2, 70.8, 67.7, 63.5, 34.8, 31.9, 29.12, 29.07, 22.6, 22.2 ppm.

Compound 8: The compound 7 (98 mg, 0.13 mmol) was dissolved in dry DMF (10 mL) and NaN₃ (85 mg, 1.3 mmol, 10 eq.) was added to this mixture. Thereafter, the solution was heated at 80°C overnight. The solvent was evaporated under vacuum and the crude product was diluted with CH₂Cl₂ and filtered. The filtrate was collected and the solvent was removed under reduced pressure to afford compound **8** (80 mg, 99%) as a tan solid. ¹H NMR (500 MHz, CDCl₃, 298 K): δ = 7.24 (d, *J* = 8.4 Hz, 6H), 7.09 (m, 8H), 6.76 (d, *J* = 9.5 Hz, 2H), 3.95, (t, *J* = 6.2 Hz, 2H), 3.31 (t, *J* = 7.0 Hz, 2H), 1.81 (m, 2H), 1.69 (m, 2H), 1.57 (m, 2H), 1.31 (s, 27H) ppm. ¹³C NMR (125 MHz, CDCl₃, 298 K): δ = 157.3, 148.8, 144.7, 140.1, 132.8, 131.3, 124.5, 113.4, 67.8, 63.6, 51.8, 34.8, 31.8, 29.4, 29.2, 23.9 ppm. MALDI-MS calcd for *m/z* = 638.41 [*M* + Na]⁺, found *m/z* = 638.16. **Compound 10-2PF**₆: A solution of **8** (139 mg, 0.23 mmol) in Me₂CO (20 mL) was added dropwise over 40 min to a solution of the dialkyne-functionalized compound **9-**2PF₆ (500 mg, 0.91 mmol), Cu(MeCN)₄PF₆ (42 mg, 0.11 mmol) and TBTA (60 mg, 0.11 mmol) in Me₂CO (20 mL). The resulting solution was stirred at RT overnight. The solvent was evaporated in vacuo. The crude product was purified by RP-HPLC (H₂O – MeCN / 0 – 100 % in 40 min λ = 254 nm) to afford pure **10-**2PF₆ (120 mg, 46%). ¹H NMR (500 MHz, Acetone-d₆, 298 K): δ = 9.50 (d, *J* = 6.5 Hz, 2H), 9.42 (d, *J* = 6.5 Hz, 2H), 8.90 (d, *J* = 6.5 Hz, 2H), 8.84 (d, *J* = 6.5 Hz, 2H), 7.89 (s, 1H), 7.34 (d, *J* = 8.45 Hz, 6H), 7.14 (d, *J* = 8.45 Hz, 2H), 7.09 (d, *J* = 8.75 Hz, 2H), 6.81 (d, *J* = 8.75 Hz, 2H), 5.15 (t, *J* = 6.9 Hz, 2H), 4.42 (t, *J* = 6.9 Hz, 2H), 3.96 (t, *J* = 6.9 Hz, 2H), 3.63 (t, *J* = 6.9 Hz, 2H), 3.20 (m, 2H), 2.73 (t, *J* = 2.1 Hz, 2H), 1.95 (m, 2H), 1.78 (m, 2H), 1.46 (m, 2H), 1.31 (s, 27H) ppm. ¹³C NMR (125 MHz, Acetone-d₆, 298 K): δ = 157.9, 151.42, 151.0, 149.2, 147.3, 147.2, 145.3, 142.4, 140.2, 132.8, 131.4, 128.1, 127.9, 125.1, 123.8, 114.0, 79.0, 75.1, 68.1, 63.9, 62.2, 61.0, 50.6, 34.9, 31.6, 30.7, 27.9, 23.8, 21.6 ppm. ESI-HRMS calcd for *m/z* = 1022.53 [*M* -PF₆]⁺, found *m/z* = 1022.49.

Compound 11: Compound **6** (563 mg, 0.95 mmol), **5** (700 mg, 0.95 mmol), and PPh₃ (498 mg, 1.9 mmol) were added at 0°C to a solution of 1,4,5,8 naphthalenetetracarboxydiimide (266 mg, 1 mmol) in dry THF (300 mL). Diethylazodicarboxylate (DEAD) (0.3 mL, 1.9 mmol) was added dropwise to this solution and the reaction mixture was stirred at RT for 48 h. The resulting suspension was filtered passed through celite, washed with CH₂Cl₂. The filtrate was concentrated under vacuum before being purified by column chromatography [SiO₂: EtOAc : Hexane (1 : 3)] to afford (250 mg) a white solid. ¹H NMR (500 MHz, CDCl₃, 298 K): δ = 8.78 (s, 4H), 7.80 (d, *J* = 8 Hz, 2H), 7.34 (d, *J* = 8 Hz, 2H), 7.25–6.70 (m, 32H), 4.25 (t, *J* = 6.5 Hz, 2H), 4.06 (t, *J* = 6.5 Hz, 2H), 3.98 (m, 4H), 3.88 (t, *J* = 6.5 Hz, 2H), 3.7 (t, *J* = 6.5 Hz, 2H), 2.62 (q, *J* = 7.6 Hz, 4H), 2.44 (s, 3H), 1.89–1.44 (m,

12 H), 1.24(t, J = 7.6 Hz, 6H) ppm. ¹³C NMR (125 MHz, CDCl₃, 298 K): $\delta = 162.9$, 156.9, 156.8, 156.7, 148.3, 144.8, 144.8, 144.1, 141.4, 139.6, 139.5, 139.4, 133.1, 132.2, 132.0, 131.0, 130.9, 130.7, 129.8, 129.4, 127.9, 126.8, 126.7, 126.6, 124.0, 114.4, 113.0, 112.9, 112.8, 70.4, 67.5, 67.2, 63.0, 62.9, 62.8, 40.8, 34.3, 32.5, 29.1, 29.0, 28.7, 28.6, 28.2, 27.9, 23.7, 22.4, 22.2, 21.6, 15.4 ppm. ESI-MS calcd for $m/z = 1577.77 [M + Na]^+$, found m/z = 1577.72. The product (235 mg, 0.15 mmol) was dissolved in dry DMF (10 mL) and NaN₃ (49 mg, 0.76 mmol, 5 eq.) was added to this mixture. Thereafter, the solution was heated at 80°C for 12 h. The solvent was evaporated under vacuum and the crude product was purified by column chromatography [SiO₂: CH₂Cl₂ / MeOH (98:2)] to afford compound 11 (212 mg, 35%) as a white solid. ¹H NMR (500 MHz, CDCl₃, 298 K): δ = 8.75 (s, 4H), 7.25–6.7 (m, 32H), 4.25 (m, 4H), 3.94 (m, 6H), 3.29 (t, J = 6.5 Hz, 2H), 2.62 (q, J = 7.6 Hz, 4H), 1.89–1.44 (m, 12 H), 1.22 (t, J = 7.6 Hz, 6H) ppm. ¹³C NMR (125 MHz, CDCl₃, 298 K): $\delta = 162.9$, 156.9, 156.8, 156.7, 148.3, 144.7, 144.2, 144.1, 139.5, 139.4, 132.2, 132.0, 132.0, 131.0, 130.9, 130.7, 126.8, 126.6, 124.1, 113.0, 112.9, 67.4, 67.3, 63.0, 62.9, 51.4, 40.8, 34.3, 29.7, 28.9, 28.7, 28.2, 27.8, 23.7, 23.4, 15.4 ppm. ESI-MS calcd for $m/z = 1448.76 [M + Na]^+$, found m/z = 1448.88. **Compound 12-2PF**₆: Compound 11 (48 mg, 0.034 mmol), 10-2PF₆ (40 mg, 0.034 mmol) dissolved in Me₂CO (1 mL), TBTA (9 mg), Cu(MeCN)₄PF₆ (6 mg) were added and the solution was allowed to stir at RT for 24 h. The solvent was evaporated under vacuum and the residue was purified by column chromatography (SiO₂ : 1% NH₄PF₆ in Me₂CO) to afford the dumbbell 12•2PF₆ as a reddish-white solid (50 mg, 57 %). ¹H NMR (CD₃CN, 500 MHz, 298 K): δ = 8.80 (d, J = 7.2 Hz, 4H), 8.66 (s, 4H), 8.23 (d, J = 7.2 Hz, 4H), 7.34–6.71 (m, 50H), 4.95 (bs, 4H), 4.32 (t, J = 7.0 Hz, 4H), 4.16 (t, J = 7.0 Hz, 4H), 3.94 (q, J = 6.7 Hz, 4H), 3.86 (q, J = 6.7 Hz, 4H), 3.42 (bs, 4H), 2.60 (q, J = 7.5 Hz, 4H), 1.87-1.55 (m, 24H), 1.29 (s, 27H), 1.27 (s, 27H), 1.19 (t, J = 7.5 Hz, 6H) ppm.¹³C NMR (CD₃CN, 125 MHz, 298 K): δ = 163.9, 157.8, 149.7, 146.8, 145.8, 145.5, 142.6, 132.7,

132.6, 131.5, 131.3, 131.1, 129.4, 127.9, 127.7, 125.5, 125.4, 114.2, 68.3, 63.9, 41.2, 34.9, 31.5, 29.5, 29.2, 28.7, 28.3, 24.1, 23.6, 15.9 ppm. ESI-HRMS calcd for $m/z = [M - 2PF_6]^{2+}$, found m/z = 1152.6735.

1. ¹H NMR Spectroscopic Characterization



Figure S1. 600 MHz ¹H-¹H gradient selected double quantum filtered phase sensitive COSY recorded in CD₃CN of **R**-2PF₆ at 298 K.



Figure S2. (a) ¹H NMR spectrum of **R**•2PF₆ in d_7 -DMF at 298 K. (b) ¹H-¹H double quantum filtered phase sensitive COSY of **R**•2PF₆ in d_7 -DMF at 298 K of the resonances arising from the H_a and H_β protons. (c) Variable temperature plot of the resonances of the H_a and H_β protons of **R**•2PF₆ in d_7 -DMF.

Using the resonances arising from H_{α} and H_{β} protons of **R**-2PF₆ as probes for the translational motion of the BDNP38C10 ring, the fact that only single resonances for the H_{α} and H_{β} protons are observed points to an exchange process between translational isomers which is fast on the ¹H NMR timescale at least in d_7 -DMF. This fast exchange persists all the way down to 221 K (Figure S2), an observation which indicates a significant decrease in the free energy barrier to translational isomerization compared to CD₃CN. We hypothesize that decrease in the free energy barrier is a consequence of the decreased affinity for the BIPY²⁺ and NpI stations in DMF.



Figure S3. ¹H NMR spectrum of 12-2PF₆ in CD₃CN at 298 K.





Determination of Rate (k) and Barrier of Relaxation (ΔG^{\ddagger}) from the ¹H NMR Spectroscopy

The kinetic data, in particular, the rate constant for the relaxation of the BDNP38C10 ring from the NpI to the $BIPY^{2+}$ station in accordance with its equilibrium ratio, along with its associated activation barrier, were determined based on the following equilibrium –

$$\mathbf{R}^{2+}_{\text{Npl}} \xrightarrow{k_1} \mathbf{R}^{2+}_{\text{BIPY}}$$
 [Eq. 1]

Our synthetic approach allowed us to populate (Figure S6a) only one translational isomer, in which the BDNP38C10 ring encircles exclusively the NpI station (NpI \subset BDNP38C10, \mathbf{R}^{2+}_{NpI}).

Equilibration to the other translational isomer, in which the BDNP38C10 ring encircles the BIPY²⁺ station (BIPY²⁺ \subset BDNP38C10, \mathbf{R}^{2+}_{BIPY}) begins to proceed following the synthesis of the rotaxane \mathbf{R}^{2+} . From the reaction shown above (Eq. 1), we can derive the following rate law:

$$d[\mathbf{R}^{2+}_{NpI}]/dt = -k_1[\mathbf{R}^{2+}_{NpI}] + k_2[\mathbf{R}^{2+}_{BIPY}]$$
[Eq. 2]

where k_1 is the rate constant for the forward reaction generating \mathbf{R}^{2+}_{BIPY} and k_2 is the backwards rate constant producing \mathbf{R}^{2+}_{NpI} .

The solution^[S3] to Eq. 2 is:

$$x_{t} = x_{eq} + (x_{o} - x_{eq})exp[-k_{1}t/(1 - x_{eq})]$$
[Eq. 3]

where \mathbf{x}_t is the mol fraction of $\mathbf{R}^{2+}{}_{NpI} \{ ([\mathbf{R}^{2+}{}_{NpI}] + [\mathbf{R}^{2+}{}_{BIPY}]) \}$ as a function of time, \mathbf{x}_o is the mol fraction of the initial amount of $\mathbf{R}^{2+}{}_{NpI}$ at t = 0, and \mathbf{x}_{eq} is the final mol fraction amount of $\mathbf{R}^{2+}{}_{NpI}$ at equilibrium when $t = \infty$. From ¹H NMR spectroscopy, \mathbf{x}_{eq} was measured to be 0.6, and we assume that \mathbf{x}_o is equal to 1. Fitting (Figure S6b) the data obtained from ¹H NMR spectroscopic experiments to this equation, we find the time constant, τ , and hence the rate constant, k_1 , which is equal to 2.2 ± 0.3 × 10⁻⁵ s⁻¹. Using the Eyring equation, this rate constant corresponds to a free energy of activation (ΔG^{\ddagger}) of 23.8 ± 0.1 kcal mol⁻¹ at room temperature in CD₃CN. Errors in the rate constant and activation barrier were determined from the error of the fit to the decay curve.



Figure S6. a) Reaction scheme illustrating the synthetic approach, which provides the ability to analyze the rate of shuttling from the BDNP38C10 ring from the NpI station to the BIPY station. Formation of the rotaxane **R** by a slippage mechanism proceeds first of all and prior to the reaction with **12**•2PF₆ containing the BIPY²⁺ station. Immediately following the click reaction with **12**•2PF₆, only a single isomer exist, wherein the BDNP38C10 ring encircles the NpI station. We can then follow the equilibration process wherein the BDNP38C10 ring shuttles over the central speed-bump to encircle the BIPY²⁺ station using ¹H NMR spectroscopy in CD₃CN at 298 K. b) First-order decay profile for the shuttling process of the BDNP38C10 ring from the NpI to BIPY²⁺ station obtained from ¹H NMR spectroscopy in CD₃CN at 298 K and the best-fit curve (red). From these data, we can determine the time constant, τ , and hence the rate constant k_1 according to equation 3.

Differential Pulse Voltammetery (DPV) Cyclic Voltammetry (CV) Measurements and Spectroelectrochemistry (SEC)

Figure S7 shows the DPV measurement of the [2]rotaxane \mathbf{R} -2PF₆ recorded in argon purged DMF (TBAPF₆ = 0.1 M). The observation of a shoulder peak accompanying the reduction process observed at approximately – 0.60 V, in addition to the observed broadness of the reduction processes occurring at approximately – 0.86 V support the existence of the overlapping reduction potentials.



Figure S7. DPV measurement of the rotaxane $\mathbf{R} \cdot 2\mathbf{PF}_6$ recorded in argon purged DMF solution at a 1 mM concentration with TBAPF₆ (0.1 M) as the supporting electrolyte at 298 K.



Figure S8. (a) Two sequential CV scans of the rotaxane \mathbf{R} -2PF₆ recorded in argon purged DMF at 298 K. The scans started at 0 V and were taken to the limit of -0.95 V. The concentration of the solution was 1 mM supported by 0.1 M TBAPF₆ electrolyte. (b) Graphical representation of the proposed scheme illustrating the inability of the rotaxane to switch upon generation of the NPI⁻ radical anion.

Figure S8a shows two scans of the CV of the rotaxane $\mathbf{R} \cdot 2PF_6$ starting from 0 V and scanning to the limit of – 0.95 V. No significant change in the relative intensity of the reduction process at – 0.46 V between the first and second scans were observed. This observation indicates that the NPI^{•-} radical anion is not able to induce switching of the BDNP38C10 macrocycle over the central speed bump to encircle the neutral BIPY⁰ on the timescale of the CV experiment illustrated by Figure S8b.



Figure S9. SEC of the rotaxane $\mathbf{R} \cdot 2\mathbf{PF}_6$ in DMF at 298 K at 1.2 mM concentration with 0.1 M TBAPF₆ electrolyte. (a) Resulting spectra that appeared over time after applying a voltage of -0.75 V starting from 0 V (red trace). (b) Resulting spetra that appeared over time following the application of a potential of -1.3 V.

SEC of the rotaxane **R**·2PF₆ results (Figure S9) in the observation of spectra consistent with the formation of the BIPY⁺⁺ radical cation and the NPI⁺⁻ radical anion after applying a potential of -0.75 V, followed by spectra that are consistent with the formation of neutral BIPY⁰ and NPI²⁻ dianion after the application of -1.3 V. The initial spectrum taken at 0 V (red trace) shows a charge transfer band ($\lambda_{max} = 481$ nm) which is consistent with encirclment of the BDNP38C10 ring over both NPI and BIPY²⁺ stations in approximately a 1:1 ratio.

UV/Vis Spectroscopy

Determination of Binding Affinities

The binding affinities for the BDNP38C10 macrocycle with the BIPY²⁺ and NpI stations were elucidated from UV/vis titration data based on model compounds. All titration studies were carried out in anhydrous DMF in a 2 mm cuvet, using a Shimadzu UV 3600 UV-Vis-NIR Spectrometer at 25°C. The calculations of binding constants for model compounds from absorbance measurements taken at 460 or 550 nm, which are close to the maxima of the charge transfer bands for the inclusion complexes formed were performed using DynaFit.

BIPY²⁺ Titration Study

A 10 mM stock solution of the BDNP38C10 macrocycle was prepared in DMF. Methyl viologen bishexafluorophosphate (MV) was dissolved with the BDNP38C10 host solution, creating a 1 M guest solution of MV doped with 10 mM concentration of the macrocycle. Additionally, a 200 mM MV solution was prepared by once again diluting further with the BDNP38C10 10 mM stock solution. During the titration 5 μ L aliquots of MV guest solution were sequentially added to 500 μ L of the 10 mM BDNP38C10 macrocycle stock solution, while monitoring by UV/vis spectroscopy (Figure S10). Fitting the titration data reveals a binding constant (K_a) equal to 29 ± 1 M⁻¹.

NpI Titration Study

A 10 mM host solution of BDNP38C10 macrocycle was prepared in DMF. Bis-diethylene glycol naphthalene diimide (DNpI) was dissolved using the 10 mM BDNP38C10 stock solution, to generate a 100 mM solution of DNpI doped with 10 mM of the BDNP38C10 macrocycle. The solution was heated to fully solubilize the DNpI in DMF. Aliquots of 10, 20 and 50 µL of the guest



Figure S10. Titration data and binding constant plots for the 1:1 complexes formed between the BDNP38C10 ring and either methyl viologen or an NPI derivative model compound in DMF at 298 K.

solution were added sequentially to 500 µL of the 10 mM stock solution of the BDNP38C10 while monitoring by UV/vis spectroscopy (Figure S10). Fitting the titration data reveals a binding constant (K_a) equal to 33 ± 1 M⁻¹.

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