

Supporting Information

Title

Diaza-18-crown-6 hydroxyquinoline derivatives as flexible tools for the assessment and imaging of total intracellular magnesium

Author List

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1. General Information and Instrumentation

General Spectroscopic Methods. All reagents were Ultrapure grade. The fluorescent probes were synthesized as reported in ¹ and were dissolved in dimethyl sulfoxide (DMSO) to a final concentration of 1 mg/mL (DCHQ1 1.7 mM; DCHQ3 1.4 mM; DCHQ4 1.4 mM; DCHQ5 1.4 mM; DCHQ6 1.3 mM). Aliquots were kept at 4°C in the dark.

Photophysical measurements. Absorption spectra were recorded on a Perkin-Elmer Lambda 45 spectrophotometer. For the fluorescence spectroscopy measurements, uncorrected emission and corrected excitation spectra were obtained with a Perkin-Elmer LS 55 spectrofluorimeter.

Cell Culture. HL60 (Human Promyelocytic Leukemia) cells were grown at 37°C and 5% CO₂ in RPMI 1640 medium supplemented with 10% heat-inactivated fetal calf serum (FCS), 2 mM L-glutamine, 1000 units/mL penicillin and 1 mg/mL streptomycin. Differentiation of HL60 cells was induced by treatment with 1.3 % DMSO for at least 3 days, as reported in ².

Flow Cytometric Assay of DNA content in DMSO-treated HL60 cells. Control and DMSO-differentiated HL60 cells were collected following the Nüsse protocol for the assessment of total DNA content.³ Briefly, cell suspensions were washed and resuspended at a final concentration of 2x10⁶ cells/mL in Nüsse-1 Solution (NaCl 0.584 g/L, trisodic citrate 1.139 g/L, DNA free RNAsi 10 mg/L, Nonidet P-40 0.03 % v/v).

After 30 minutes of incubation at 4°C, an equal volume of Nusse-2 Solution was added (sucrose 85.5 g/L, citric acid 15 g/L, propidium iodide 100 mg/L). Samples were analyzed on an Epics-XL cytometer (Beckman Coulter, USA) with excitation band centered on 488 nm and emission band on 605 nm in linear scale.

2. Photophysical properties of DCHQ1

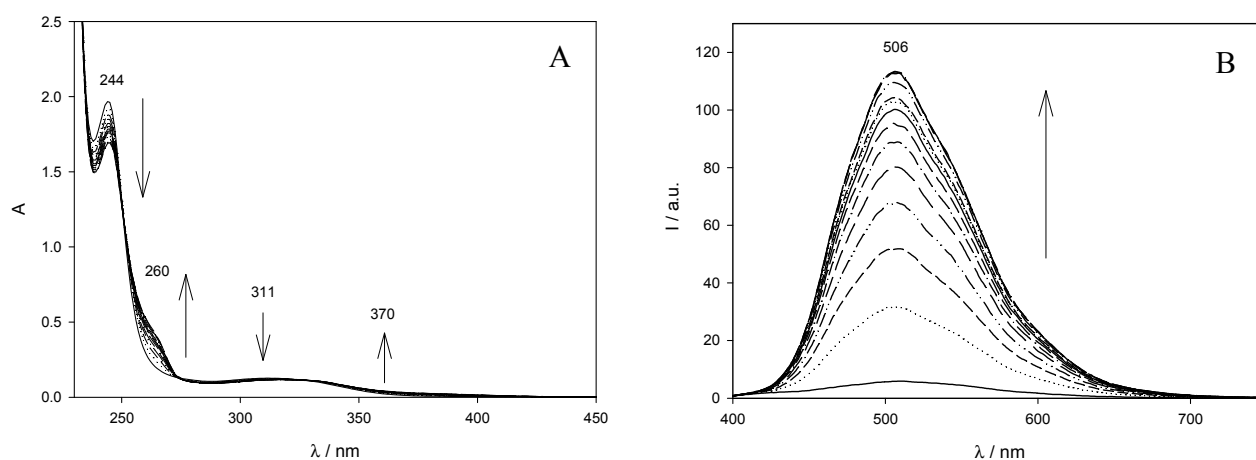


Figure 1 Titration with Mg^{2+} . Absorption (panel A) and emission ($\lambda_{exc} = 324$ nm, panel B) spectra of DCHQ1 (25 μM) in 1:1 $CH_3OH:H_2O$ mixture buffered at pH=7.4 with MOPS at room temperature upon addition of increasing amounts (up to 2 equivalents) of $Mg(NO_3)_2 \cdot 6H_2O$ solution (2.66 mM).

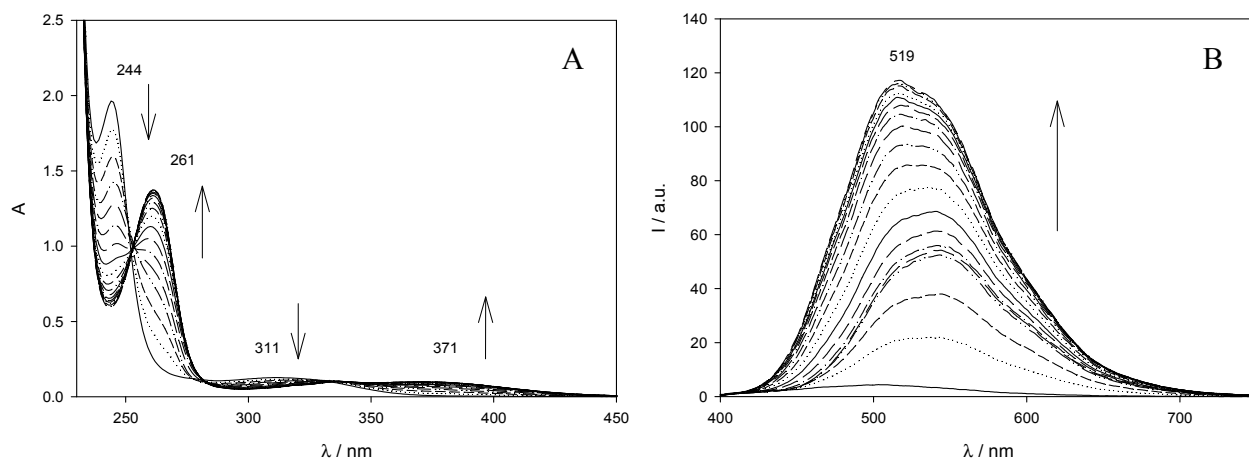


Figure 2 Titration with Zn^{2+} . Absorption (panel A) and emission ($\lambda_{exc} = 324$ nm, panel B) spectra of DCHQ1 (25 μM) in 1:1 $CH_3OH:H_2O$ mixture buffered at pH=7.4 with MOPS at room temperature upon addition of increasing amounts (up to 3 equivalents) of $Zn(NO_3)_2 \cdot 6H_2O$ solution (1.89 mM).

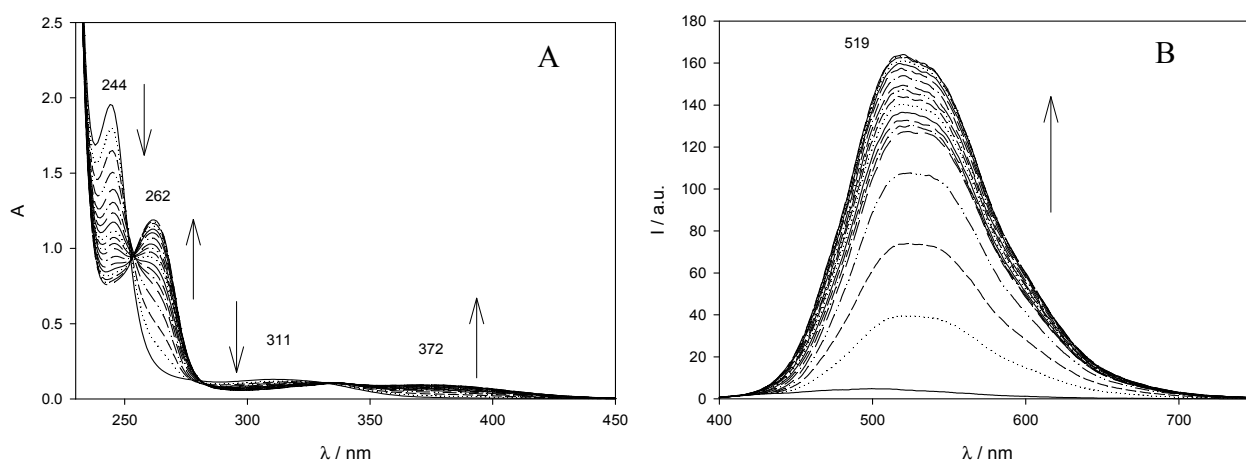


Figure 3 Titration with Cd²⁺. Absorption (panel A) and emission ($\lambda_{\text{exc}} = 330$ nm, panel B) spectra of DCHQ1 (25 μM) in 1:1 CH₃OH:H₂O mixture buffered at pH=7.4 with MOPS at room temperature upon addition of increasing amounts (up to 2.5 equivalents) of Cd(NO₃)₂·4H₂O solution (1.60 mM).

3. Photophysical properties of DCHQ3

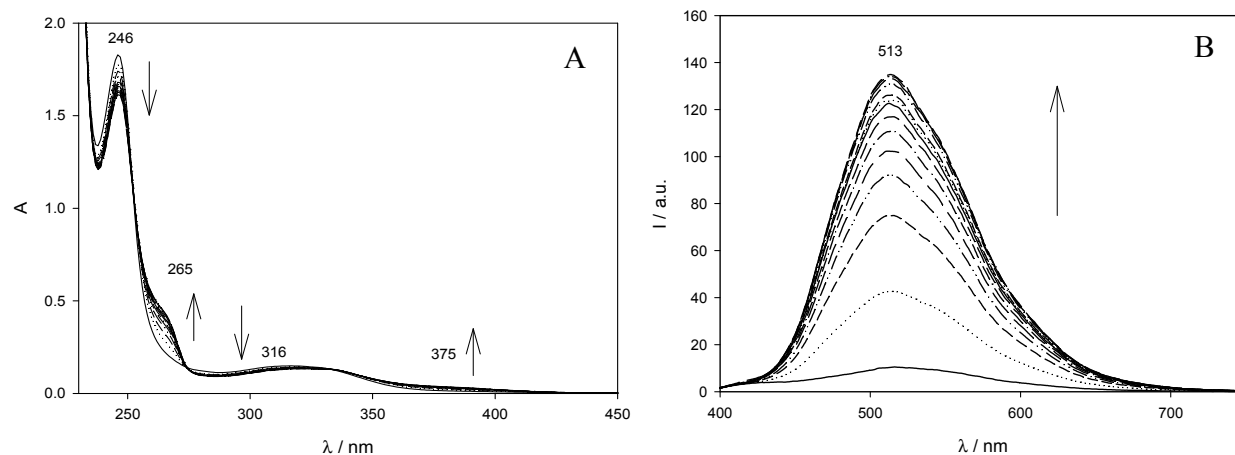


Figure 4 Titration with Mg^{2+} . Absorption (panel A) and emission ($\lambda_{exc} = 336$ nm, panel B) spectra of DCHQ4 (25 μM) in 1:1 $CH_3OH:H_2O$ mixture buffered at pH=7.4 with MOPS at room temperature upon addition of increasing amounts (up to 1.5 equivalents) of $Mg(NO_3)_2 \cdot 6H_2O$ solution (2.50 mM).

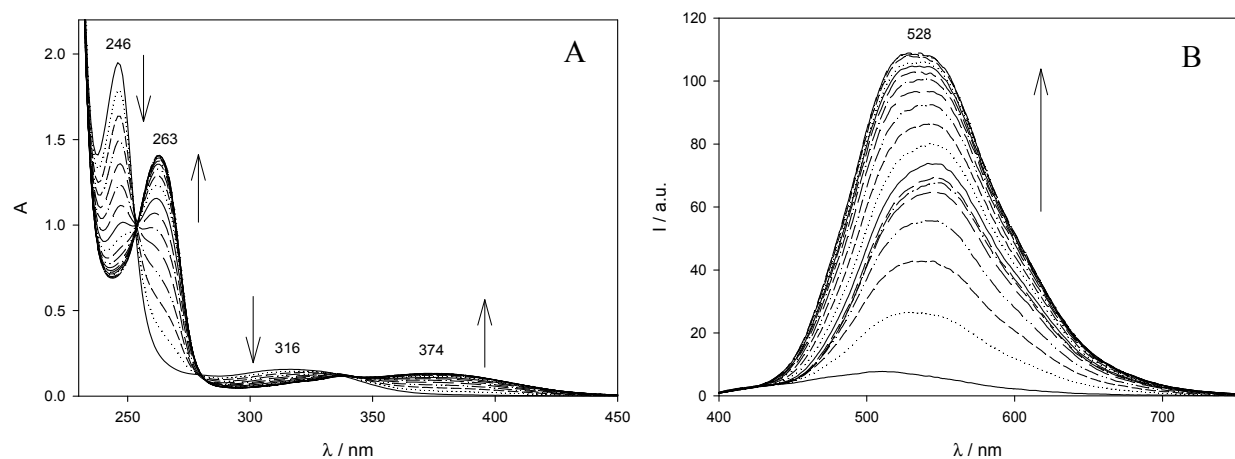


Figure 5 Titration with Zn^{2+} . Absorption (panel A) and emission ($\lambda_{exc} = 336$ nm, panel B) spectra of DCHQ4 (25 μM) in 1:1 $CH_3OH:H_2O$ mixture buffered at pH=7.4 with MOPS at room temperature upon addition of increasing amounts (up to 2.5 equivalents) of $Zn(NO_3)_2 \cdot 6H_2O$ solution (2.50 mM).

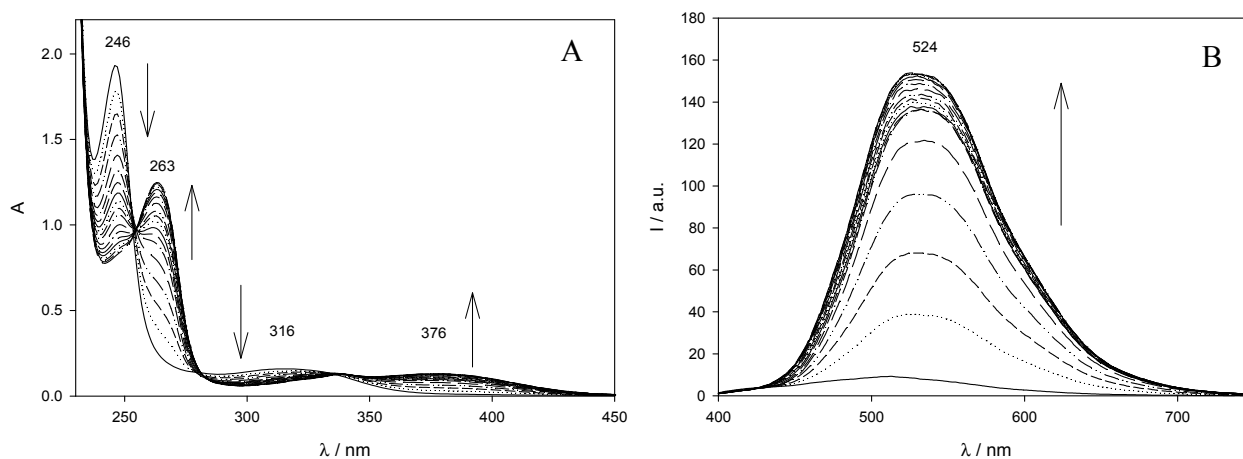


Figure 6 Titration with Cd²⁺. Absorption (panel A) and emission ($\lambda_{\text{exc}} = 336$ nm, panel B) spectra of DCHQ4 (25 μM) in 1:1 CH₃OH:H₂O mixture buffered at pH=7.4 with MOPS at room temperature upon addition of increasing amounts (up to 3 equivalents) of Cd(NO₃)₂·4H₂O solution (1.60 mM).

4. Photophysical properties of DCHQ4

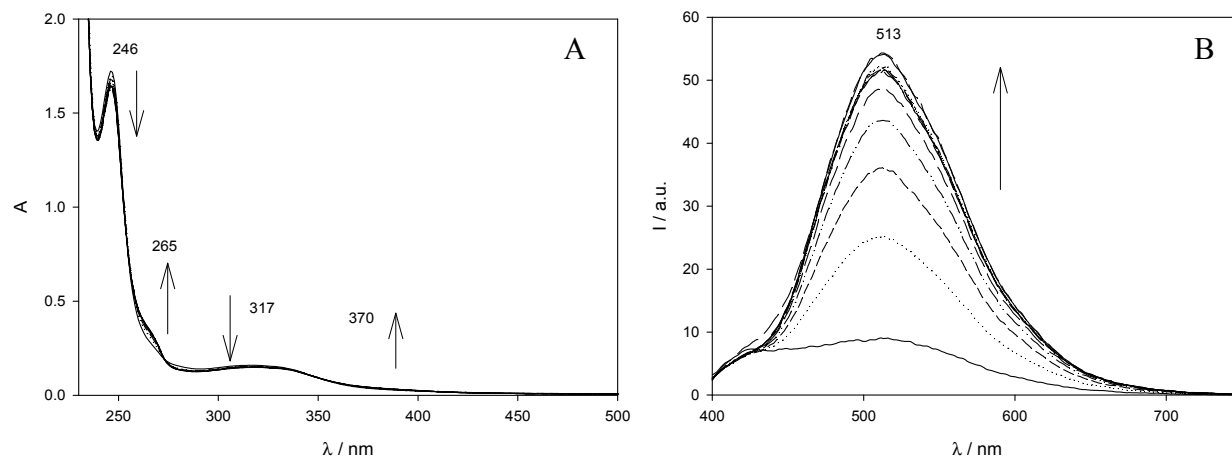


Figure 7 Titration with Mg²⁺. Absorption (panel A) and emission ($\lambda_{\text{exc}} = 330$ nm, panel B) spectra of DCHQ4 (25 μM) in 1:1 CH₃OH:H₂O mixture buffered at pH=7.4 with MOPS at room temperature upon addition of increasing amounts (up to 1.5 equivalents) of Mg(NO₃)₂·6H₂O solution (2.50 mM).

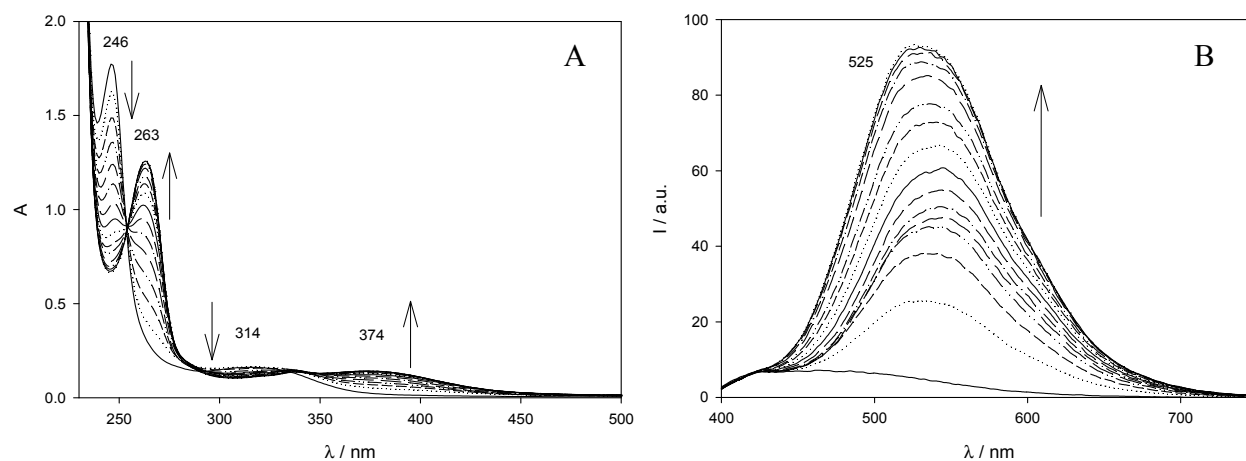


Figure 8 Titration with Zn²⁺. Absorption (panel A) and emission ($\lambda_{\text{exc}} = 330$ nm, panel B) spectra of DCHQ4 (25 μM) in 1:1 CH₃OH:H₂O mixture buffered at pH=7.4 with MOPS at room temperature upon addition of increasing amounts (up to 2 equivalents) of Zn(NO₃)₂·6H₂O solution (2.50 mM).

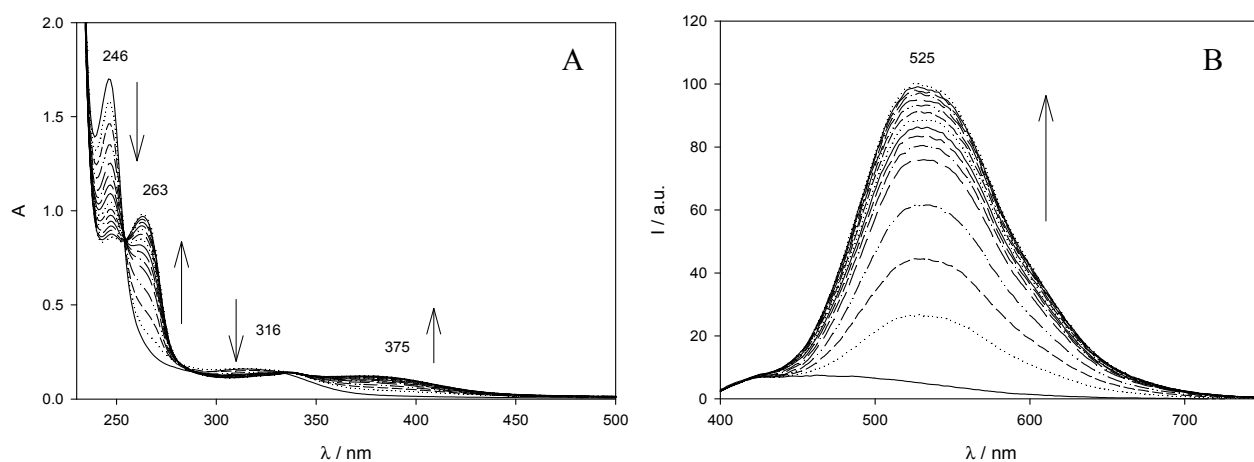


Figure 9 Titration with Cd^{2+} . Absorption (panel A) and emission ($\lambda_{\text{exc}} = 330$ nm, panel B) spectra of DCHQ4 (25 μM) in 1:1 $\text{CH}_3\text{OH}:\text{H}_2\text{O}$ mixture buffered at $\text{pH}=7.4$ with MOPS at room temperature upon addition of increasing amounts (up to 2.5 equivalents) of $\text{Cd}(\text{NO}_3)_2 \cdot 4\text{H}_2\text{O}$ solution (1.60 mM).

5. Photophysical properties of DCHQ5

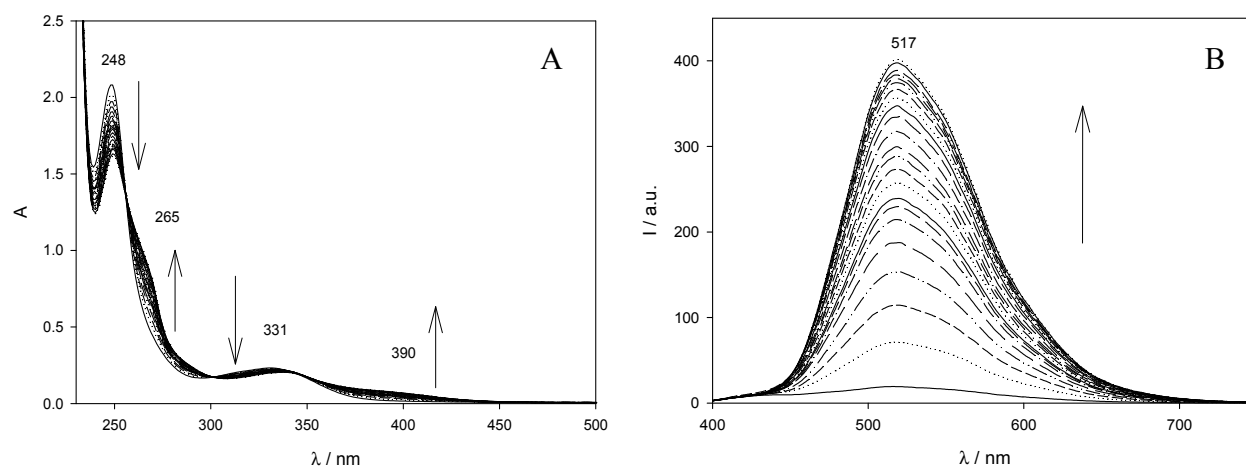


Figure 10 Titration with Mg^{2+} . Absorption (panel A) and emission ($\lambda_{exc} = 344$ nm, panel B) spectra of DCHQ5 (25 μM) in 1:1 $CH_3OH:H_2O$ mixture buffered at pH=7.4 with MOPS at room temperature upon addition of increasing amounts (up to 4 equivalents) of $Mg(NO_3)_2 \cdot 6H_2O$ solution (2.50 mM).

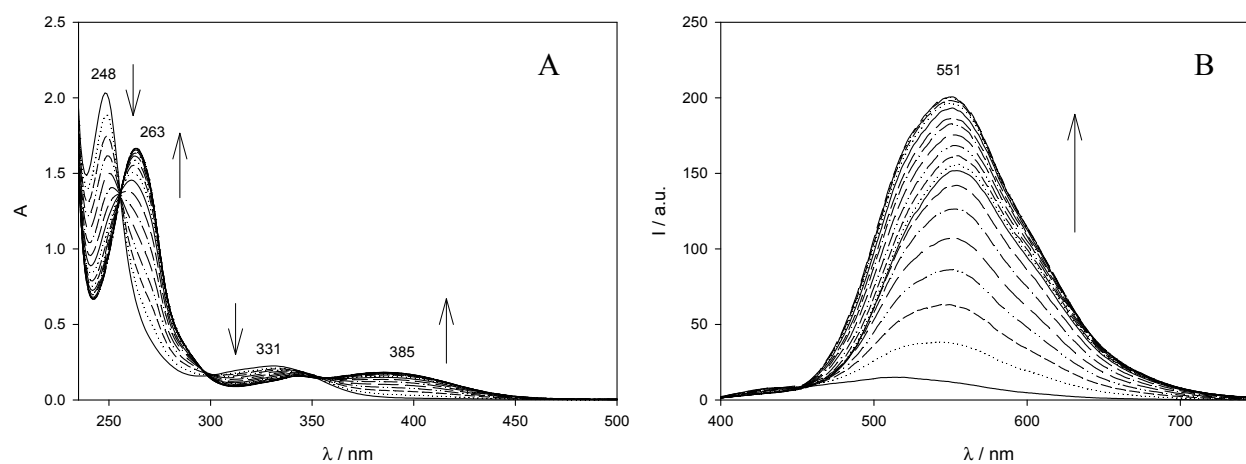


Figure 11 Titration with Zn^{2+} . Absorption (panel A) and emission ($\lambda_{exc} = 344$ nm, panel B) spectra of DCHQ5 (25 μM) in 1:1 $CH_3OH:H_2O$ mixture buffered at pH=7.4 with MOPS at room temperature upon addition of increasing amounts (up to 2 equivalents) of $Zn(NO_3)_2 \cdot 6H_2O$ solution (2.50 mM).

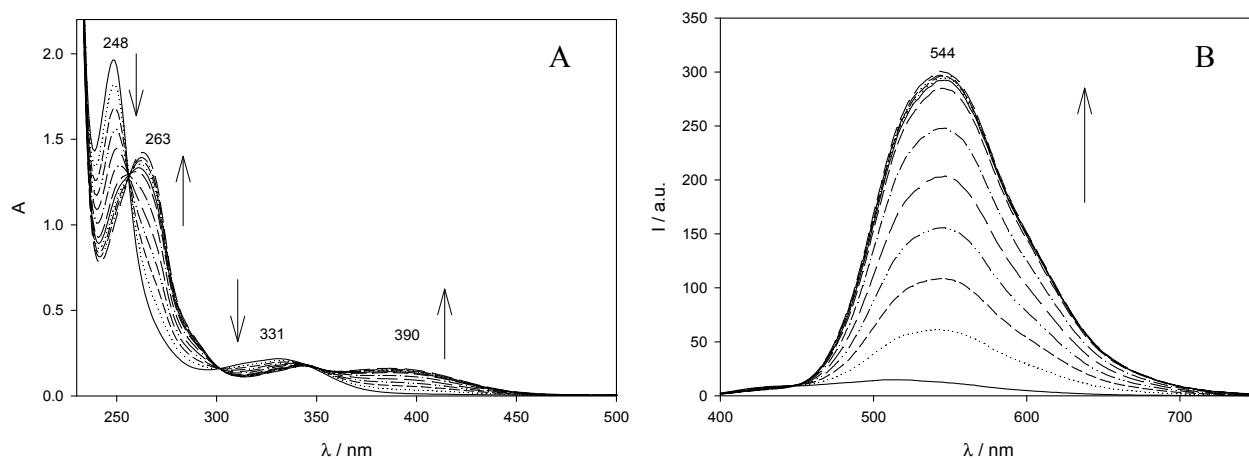


Figure 12 Titration with Cd²⁺. Absorption (panel A) and emission ($\lambda_{\text{exc}} = 344$ nm, panel B) spectra of DCHQ5 (25 μM) in 1:1 CH₃OH:H₂O mixture buffered at pH=7.4 with MOPS at room temperature upon addition of increasing amounts (up to 1.5 equivalents) of Cd(NO₃)₂·4H₂O solution (1.60 mM).

6. Photophysical properties of DCHQ6

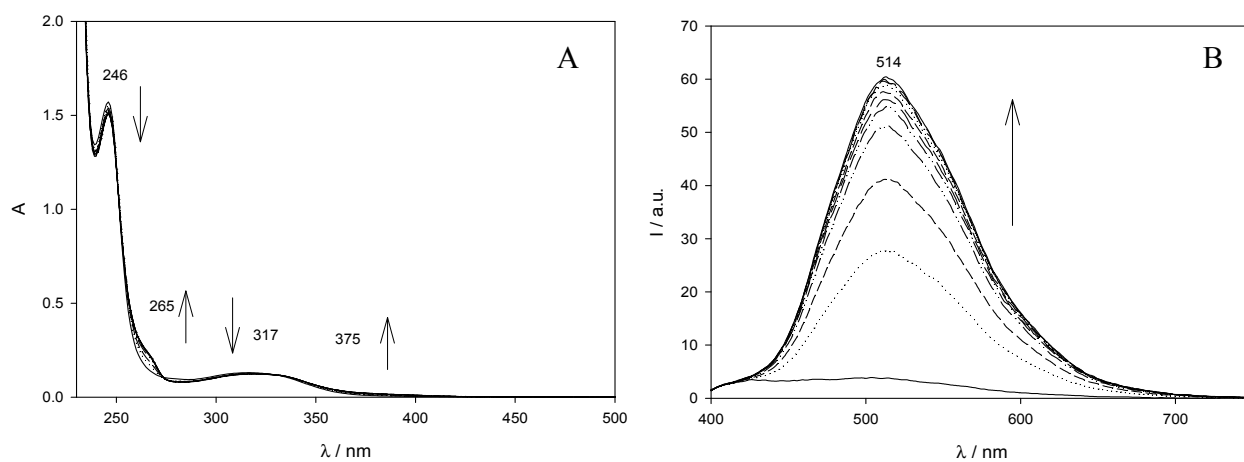


Figure 13 Titration with Mg^{2+} . Absorption (panel A) and emission ($\lambda_{\text{exc}} = 337$ nm, panel B) spectra of DCHQ6 (18 μM) in 1:1 $\text{CH}_3\text{OH}:\text{H}_2\text{O}$ mixture buffered at $\text{pH}=7.4$ with MOPS at room temperature upon addition of increasing amounts (up to 1 equivalent) of $\text{Mg}(\text{NO}_3)_2 \cdot 6\text{H}_2\text{O}$ solution (2.50 mM).

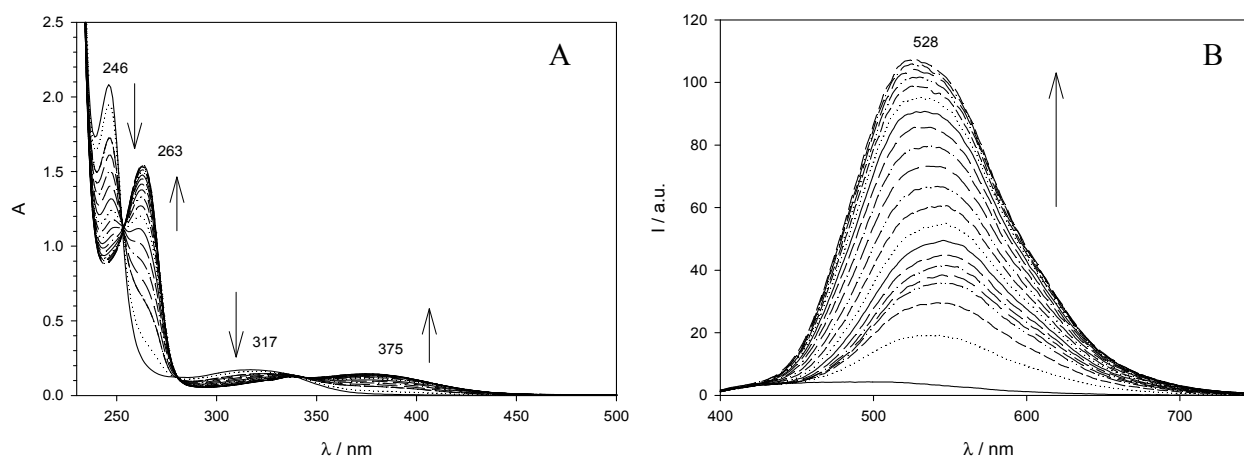


Figure 14 Titration with Zn^{2+} . Absorption (panel A) and emission ($\lambda_{\text{exc}} = 337$ nm, panel B) spectra of DCHQ6 (25 μM) in 1:1 $\text{CH}_3\text{OH}:\text{H}_2\text{O}$ mixture buffered at $\text{pH}=7.4$ with MOPS at room temperature upon addition of increasing amounts (up to 2 equivalents) of $\text{Zn}(\text{NO}_3)_2 \cdot 6\text{H}_2\text{O}$ solution (2.50 mM).

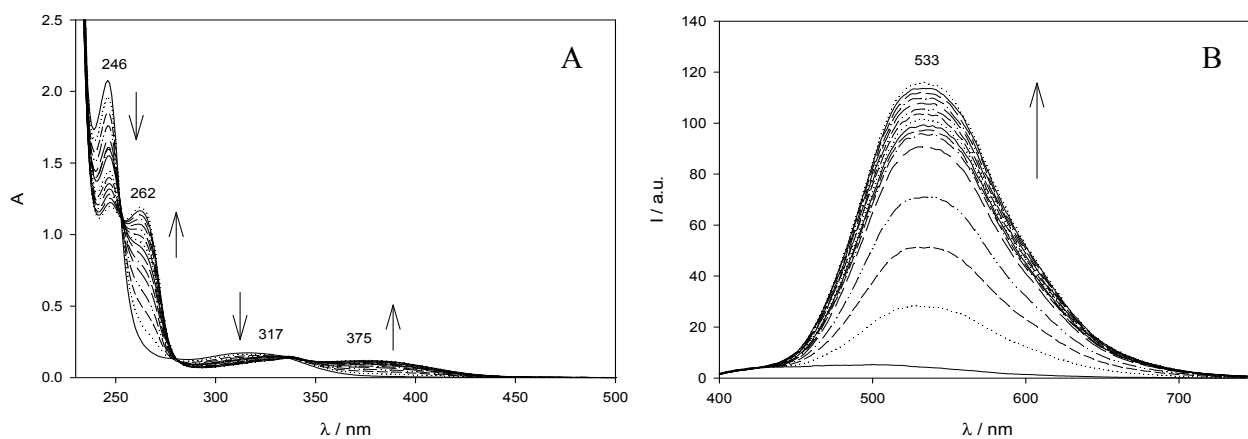


Figure 15 Titration with Cd^{2+} . Absorption (panel A) and emission ($\lambda_{\text{exc}} = 335 \text{ nm}$, panel B) spectra of DCHQ6 ($25 \mu\text{M}$) in 1:1 $\text{CH}_3\text{OH}:\text{H}_2\text{O}$ mixture buffered at $\text{pH}=7.4$ with MOPS at room temperature upon addition of increasing amounts (up to 1.5 equivalents) of $\text{Cd}(\text{NO}_3)_2 \cdot 4\text{H}_2\text{O}$ solution (1.60 mM).

7. Flow Cytometric Assay of DNA content of HL60 treated with DMSO

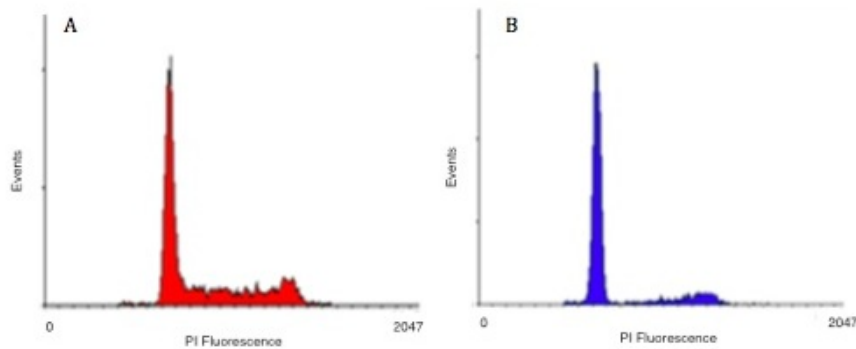


Figure 16 Assessment of the total DNA content of control (panel A) and DMSO-differentiated (panel B) HL60 cells. Histograms show the PI fluorescence (expressed in fluorescence channels), that is directly proportional to the DNA content, with respect to the number of events acquired. Results from a typical experiment.

8. References

¹A. Sanz-Medel, R. Fernandez de la Campa and J. I. Garcia-Alonso. *Analyst*, 1987, 112:493–497.

²F. I. Wolf, V. Covacci, N. Bruzzese, A. Di Francesco, A. Sacchetti, D. Corda and A. Cittadini. *J. Cell. Biochem.*, 1998, 71:441–448.

³M. Nusse and J. Kramer. *Cytometry*, 1984, 5:20-5.