

Supporting Information for:

A Highly Selective Red-emitting FRET Fluorescent Molecular Probe Derived From BODIPY for the Detection of Cysteine and Homocysteine: An Experimental and Theoretical Study

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Experimental Section

General

NMR spectra were taken on a 400 MHz Varian Unity Inova spectrophotometer. Mass spectra were recorded with a Q-TOF Micro MS spectrometer. UV-Vis spectra were taken on a HP8453 UV-visible spectrophotometer. Fluorescence spectra were recorded on a JASCO FP-6500 or a RF5301 PC (Shimadzu) spectrofluorometer. Luminescence quantum yields were measured with quinine bisulfate in 0.05 M H₂SO₄ (F = 54.6%). Luminescence lifetimes were measured on a Horiba Jobin Yvon Fluoro Max-4 (TCSPC) instrument. The cells luminescence images were obtained using a Nikon ECLIPSE-Ti confocal laser scanning microscopy.

The structures of complexes were optimized using density functional theory (DFT) with B3LYP functional and 6-31G(d) basis set. The excited state related calculations were carried out with the time dependent DFT (TD-DFT) with the ground state geometry. The 6-31G(d) basis set was employed for C, H, N, O, S. There are no imaginary frequencies for all optimized structures. All the DFT/TDDFT calculations were performed using Gaussian 09W.¹

Reference:

1. Gaussian 09W, Revision A.1, M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, G. Scalmani, V. Barone, B. Mennucci, G. A. Petersson, H. Nakatsuji, M. Caricato, X. Li, H. P. Hratchian, A. F. Izmaylov, J. Bloino, G. Zheng, J. L. Sonnenberg, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, T. Vreven, J. A. Montgomery, Jr., J. E. Peralta, F. Ogliaro, M. Bearpark, J. J. Heyd, E. Brothers, K. N. Kudin, V. N. Staroverov, R. Kobayashi, J. Normand, K. Raghavachari, A. Rendell, J. C. Burant, S. S. Iyengar, J. Tomasi, M. Cossi, N. Rega, J. M. Millam, M. Klene, J. E. Knox, J. B. Cross, V. Bakken, C. Adamo, J. Jaramillo, R. Gomperts, R. E. Stratmann, O. Yazyev, A. J. Austin, R. Cammi, C. Pomelli, J. W. Ochterski, R. L. Martin, K. Morokuma, V. G. Zakrzewski, G. A. Voth, P. Salvador, J. J. Dannenberg, S. Dapprich, A. D. Daniels, Ö. Farkas, J. B. Foresman, J. V. Ortiz, J. Cioslowski, and D. J. Fox, Gaussian, Inc., Wallingford CT, **2009**.

CHYH

11042604 70 (1.787) AM (Cen,6, 80.00, Ar,5000.0,475.28,0.70,LS 10); Sm (SG, 2x3.00); Sb (1,40.00); Cl
4.24e3

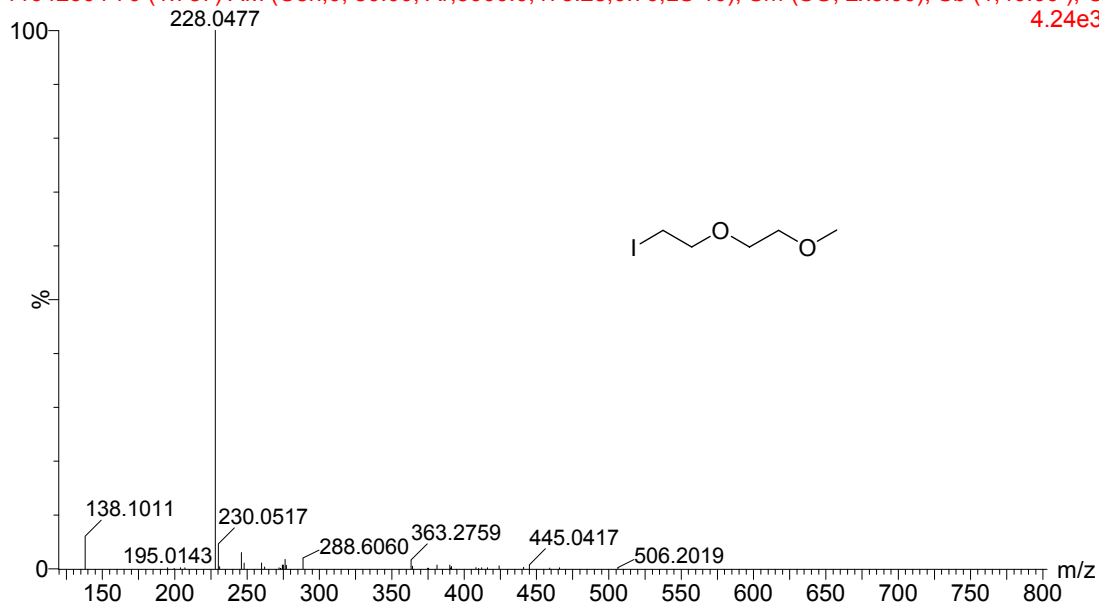


Figure S1. TOF MS EI of 1.

CHYH-1

11042516 73 (1.886) AM (Cen,6, 80.00, Ar,5000.0,475.27,0.70,LS 10); Sm (SG, 2x3.00); Sb (1,40.00); Cl
116

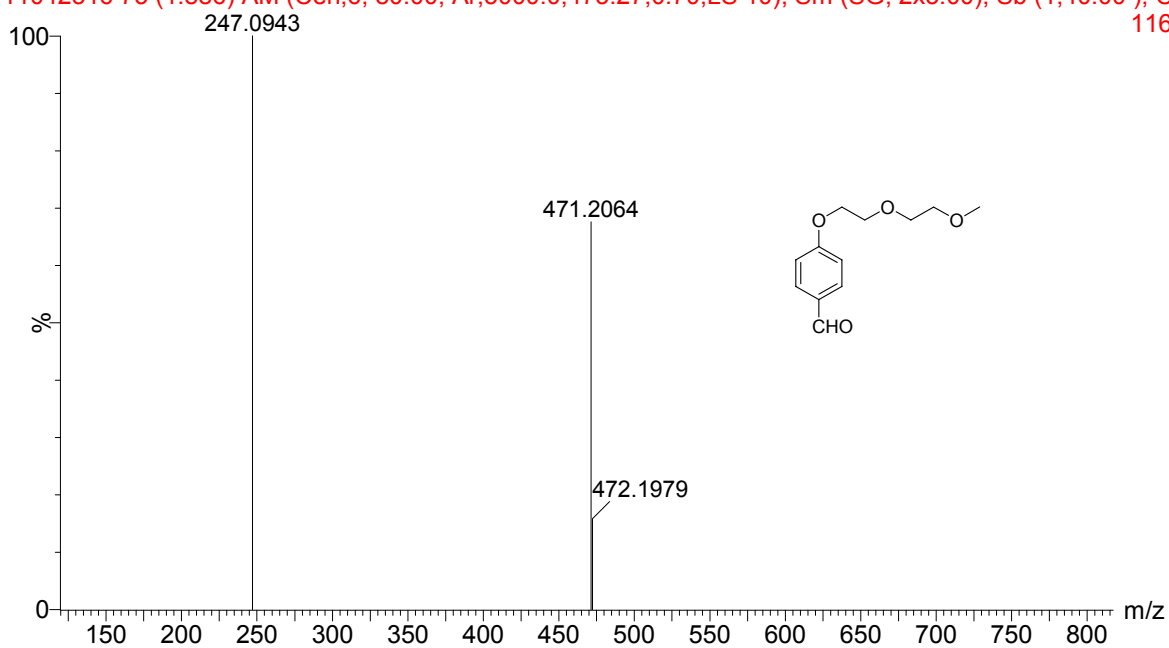


Figure S2. TOF MS EI of 2.

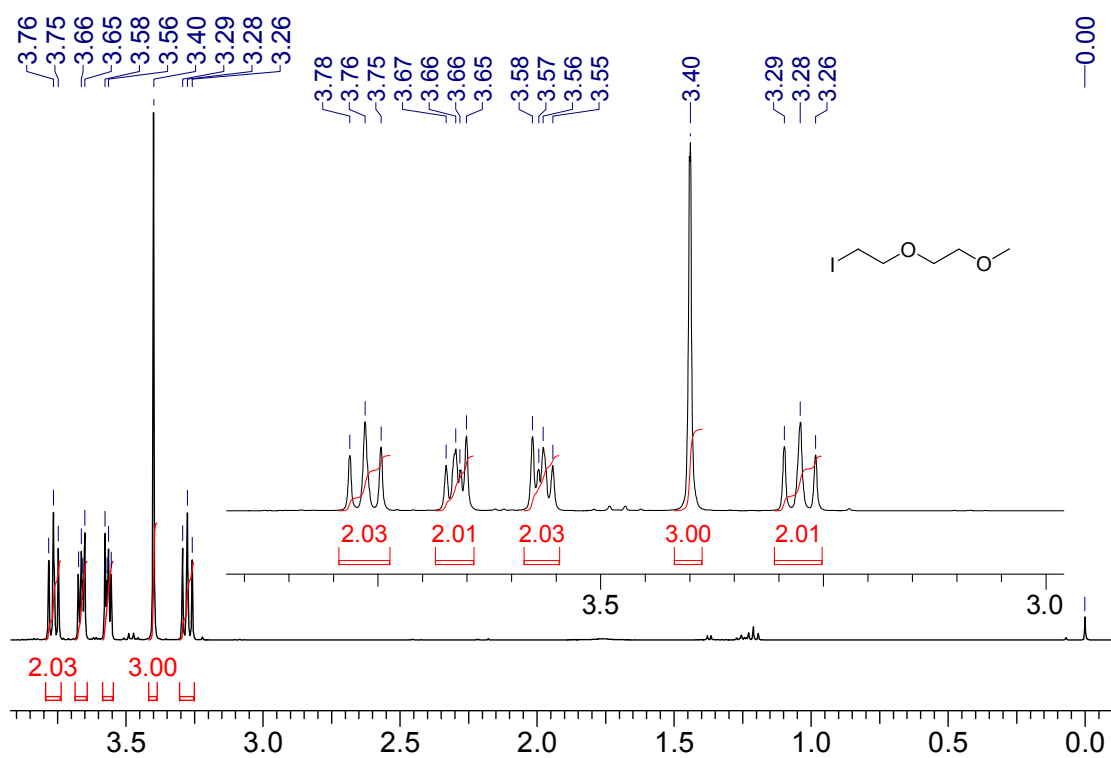


Figure S3. ¹H NMR of **1** (CDCl₃, 400 MHz).

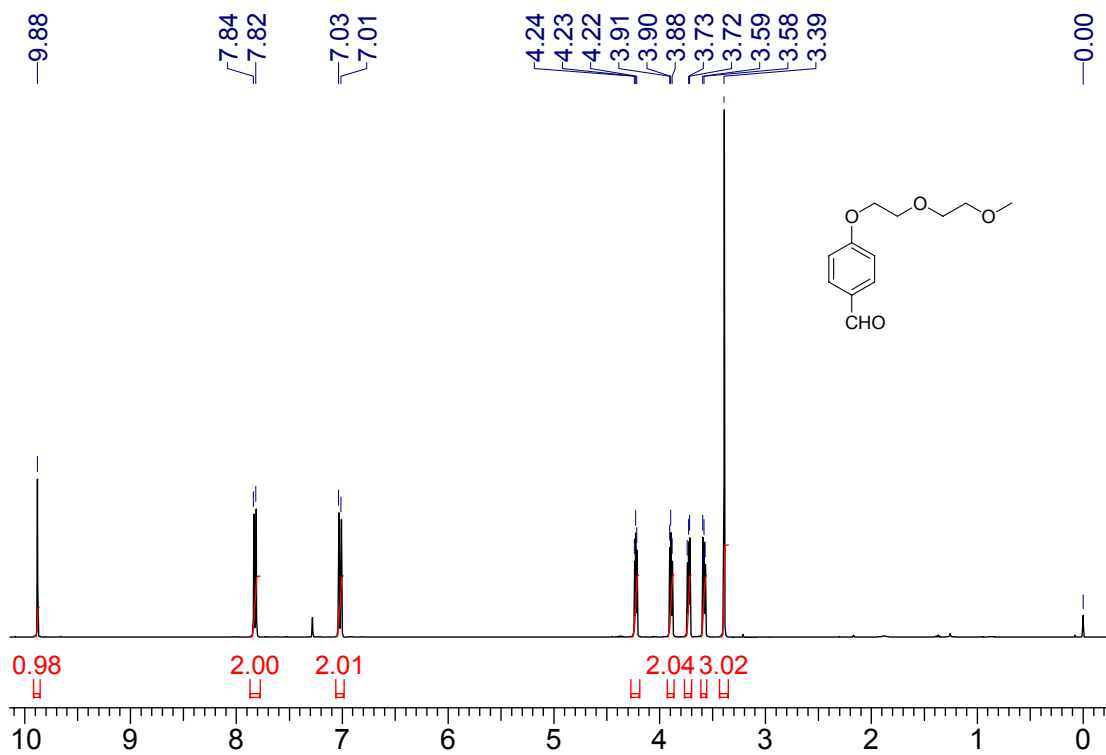


Figure S4. ¹H NMR of **2** (CDCl₃, 400 MHz).

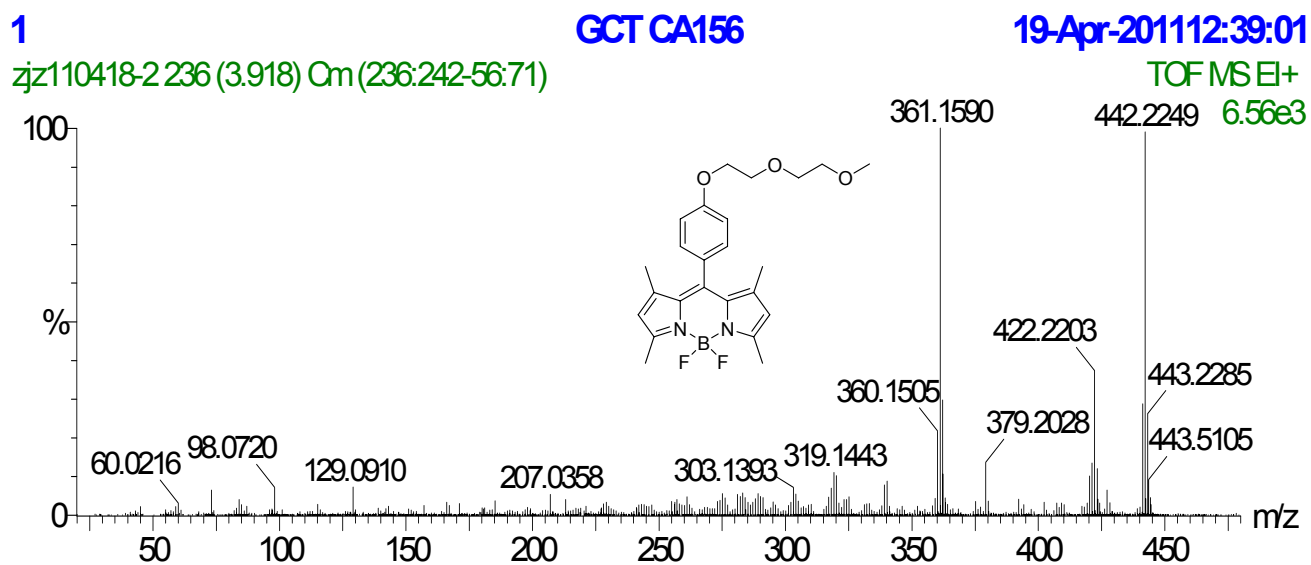


Figure S5. TOF MS EI of **3**.

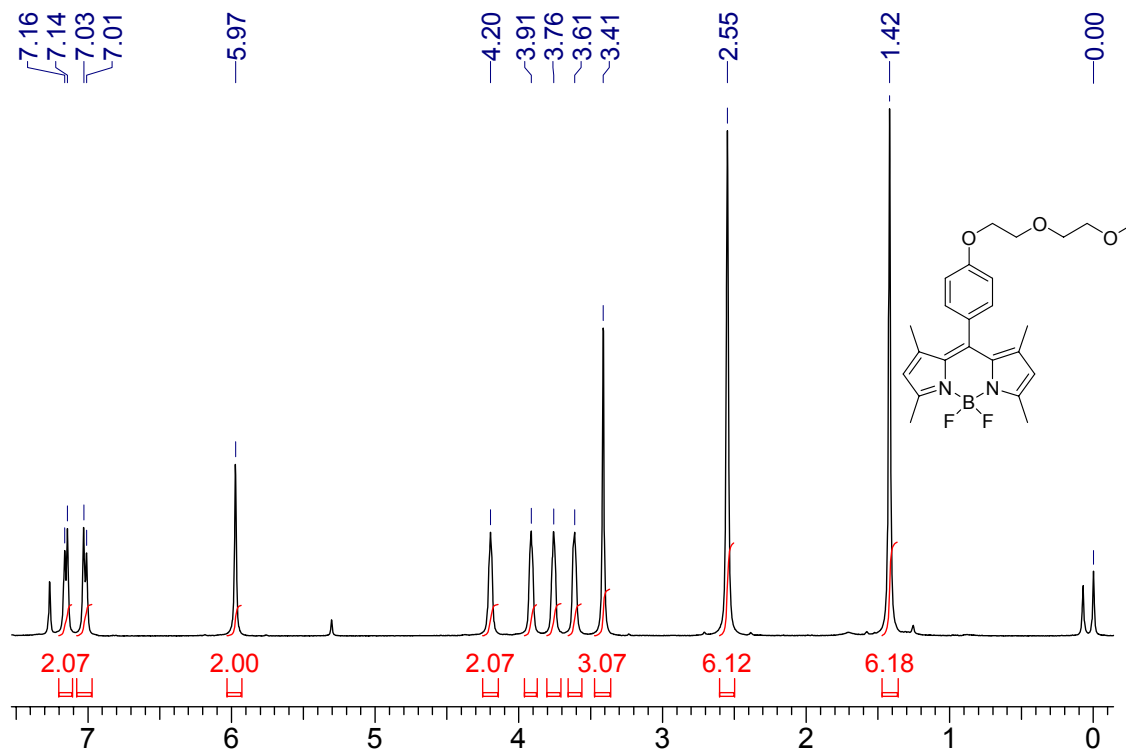


Figure S6. ^1H NMR of **3** (CDCl_3 , 400 MHz).

SJY

11041805 12 (0.399) Cn (Cen,4, 50.00, Ht); Sm (SG, 2x3.00); Sb (15,10.00); Cm (1:16) TOF LD+
1.95e3

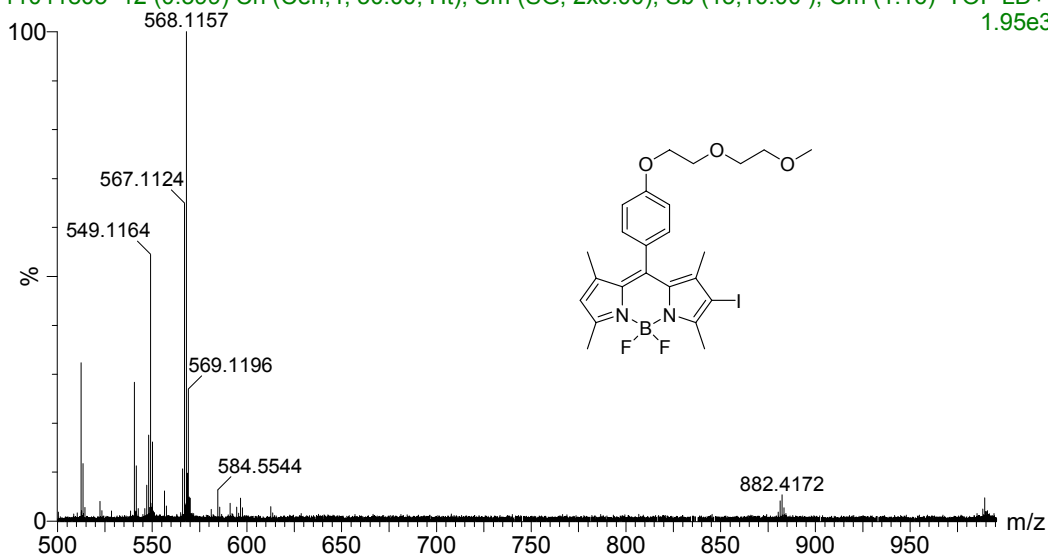


Figure S7. TOF MS EI of 4.

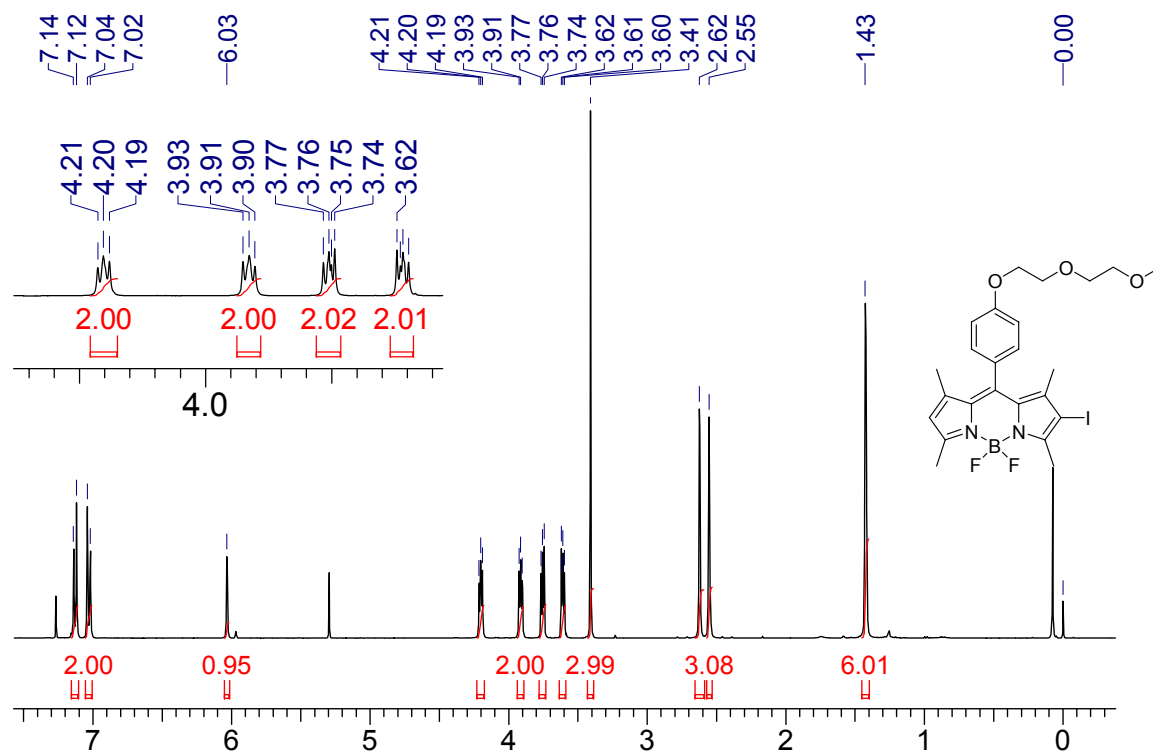


Figure S8. ^1H NMR of 4 (CDCl_3 , 400 MHz).

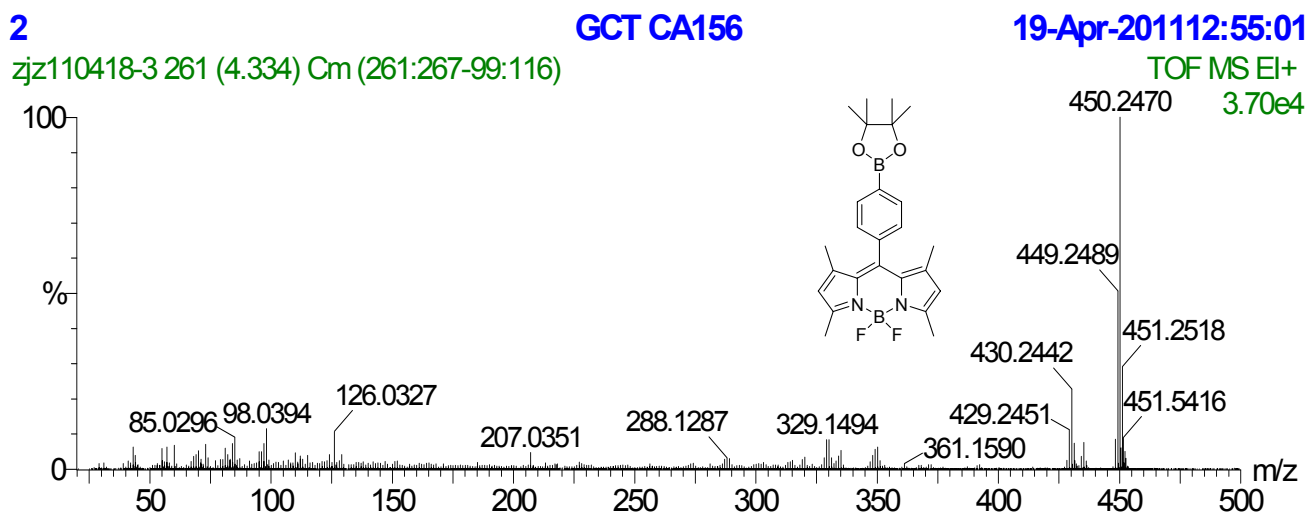


Figure S9. TOF MS EI of **5**.

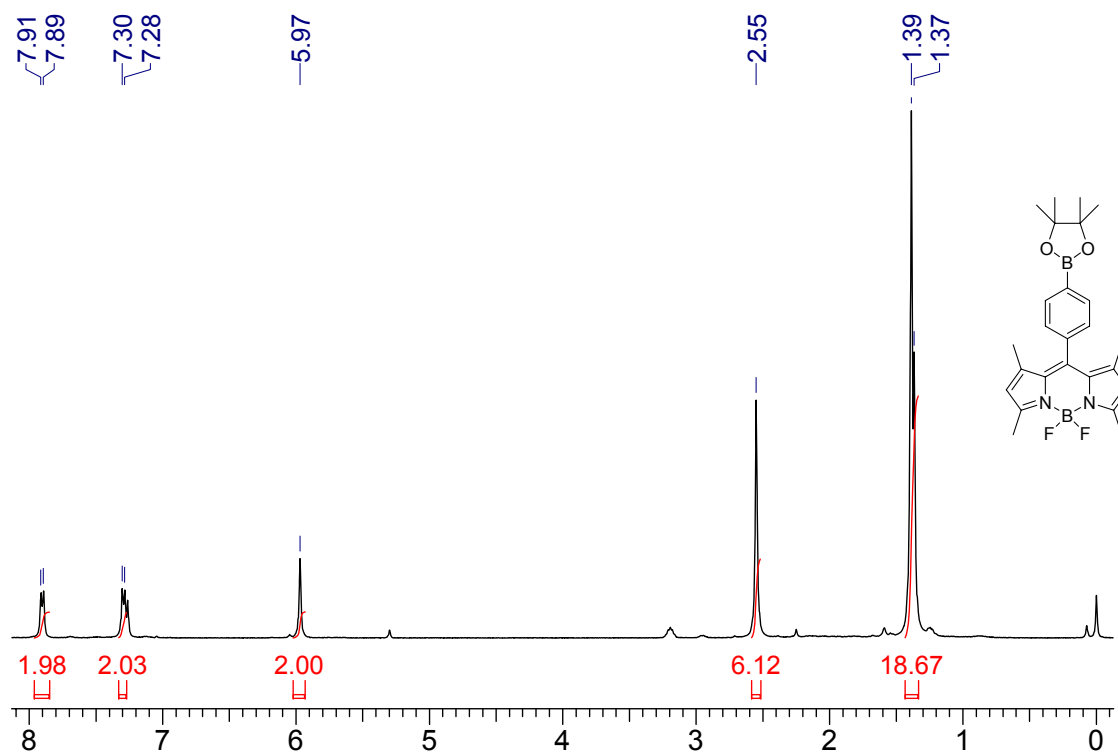


Figure S10. ^1H NMR of **5** (CDCl_3 , 400 MHz).

SJY

11011908 21 (0.700) Cn (Cen,4, 50.00, Ht); Sm (SG, 2x3.00); Sb (15,10.00); Cm (17:35)

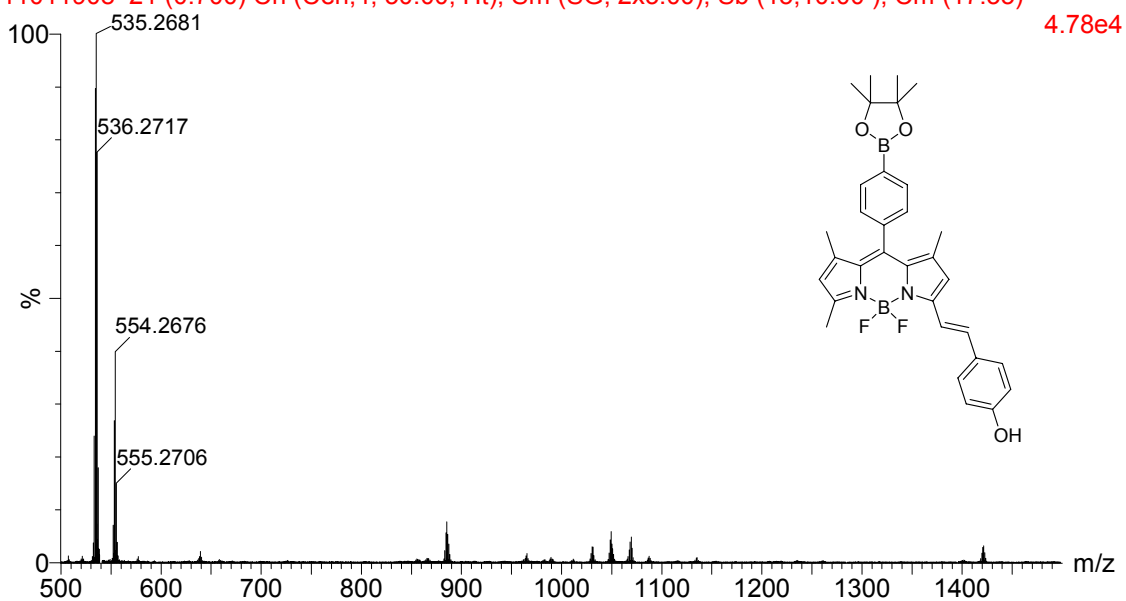


Figure S11. TOF LD of 6.

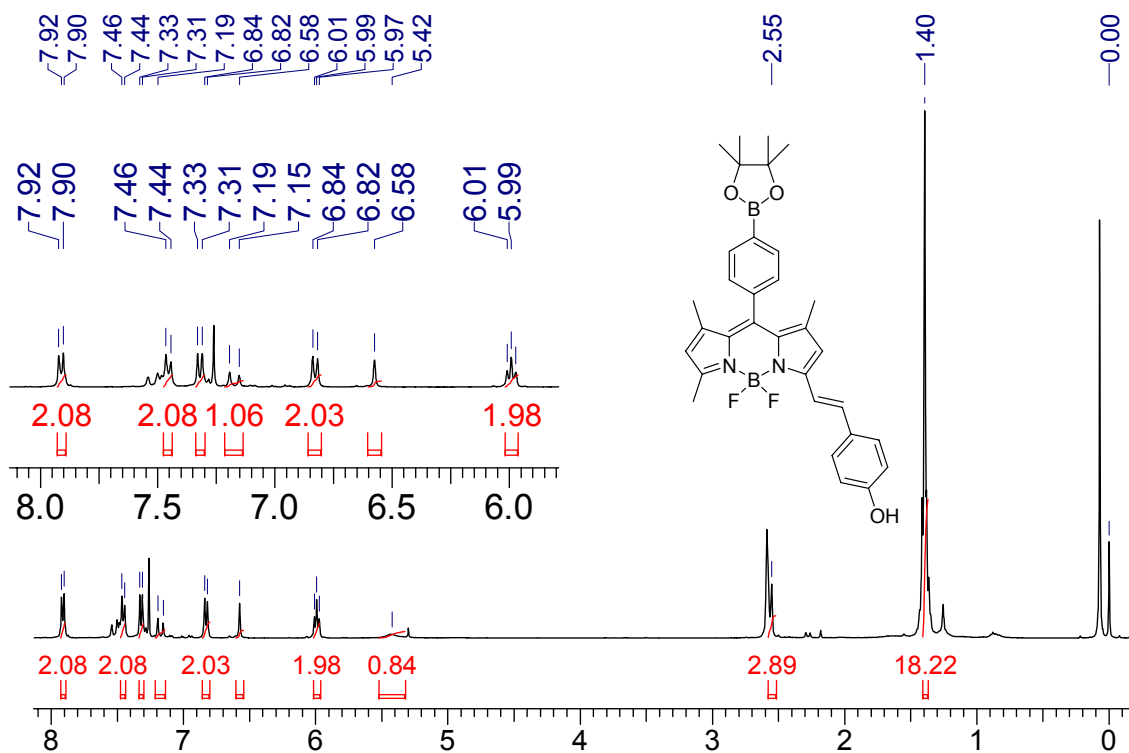


Figure S12. ^1H NMR of 6 (CDCl_3 , 400 MHz).

SJY

11022520 35 (3.006) Cn (Cen,4, 50.00, Ht); Sm (SG, 2x3.00); Sb (15,10.00); Cm (1:35)TOF LD+
7.43e3

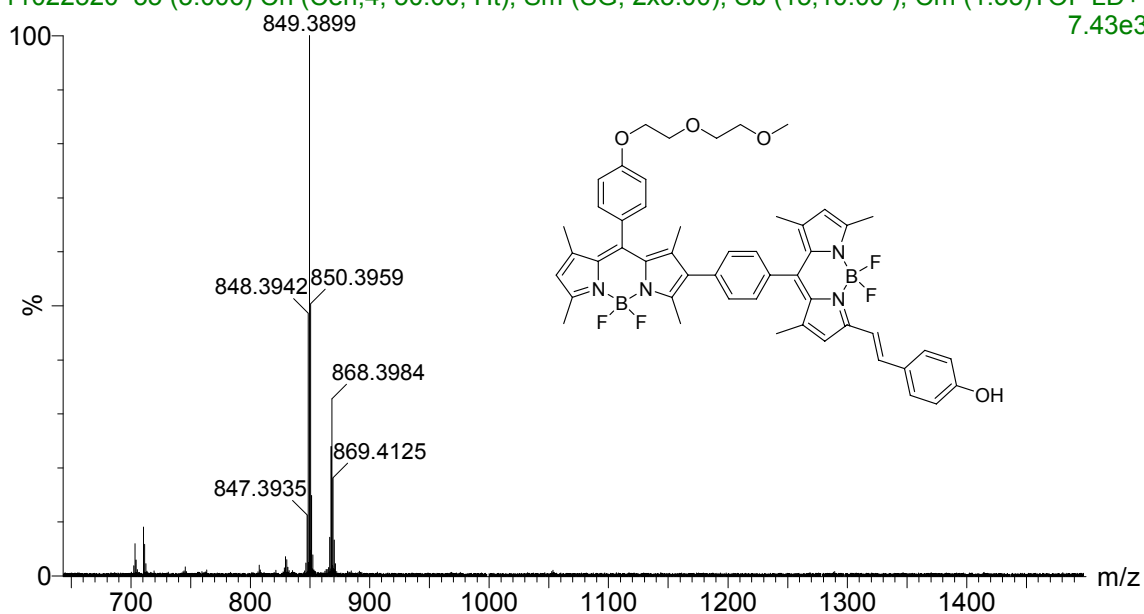


Figure S13. TOF LD of 7.

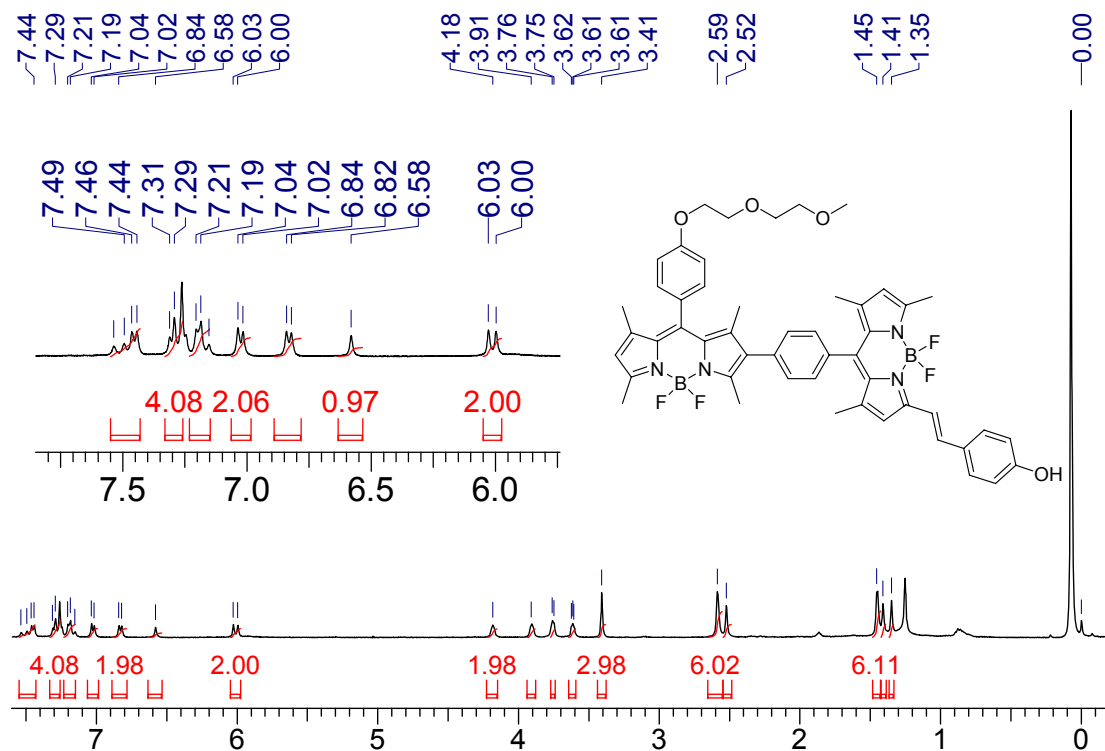


Figure S14. ^1H NMR of 7 (CDCl_3 , 400 MHz).

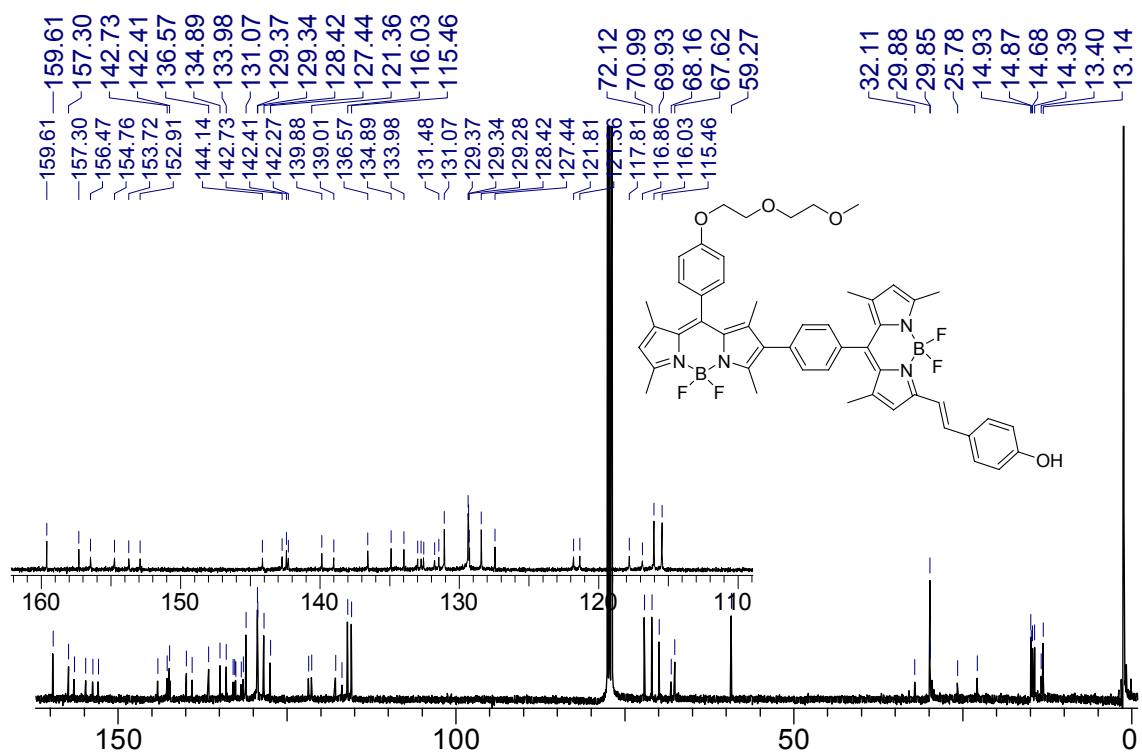


Figure S15. ^{13}C NMR of 7 (CDCl₃, 100 MHz).

SHJY

11032516 15 (0.388) AM (Cen,6, 80.00, Ar,5000.0,475.27,0.70,LS 10); Sm (SG, 2x3.00); Sb (1,40.00); C 2.81e3

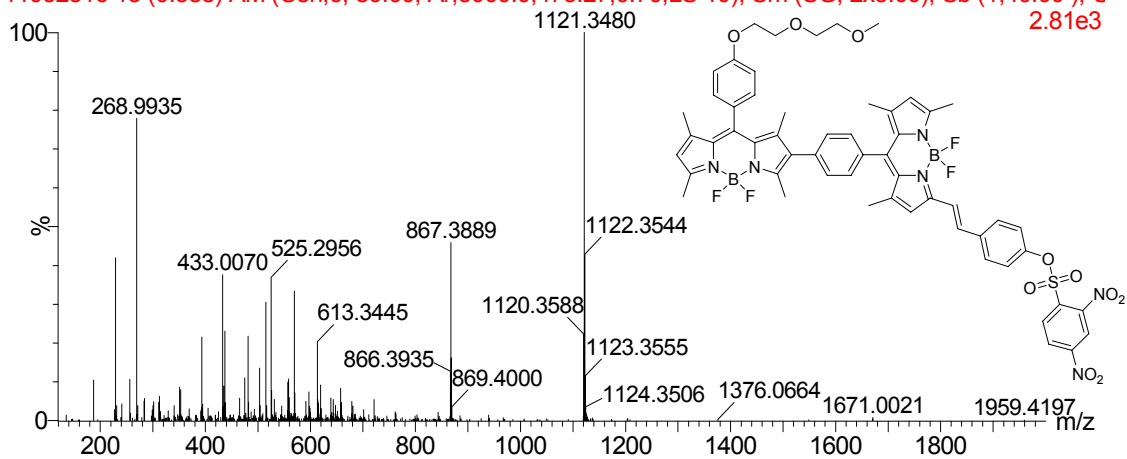


Figure S16. TOF MS EI of Probe 1.

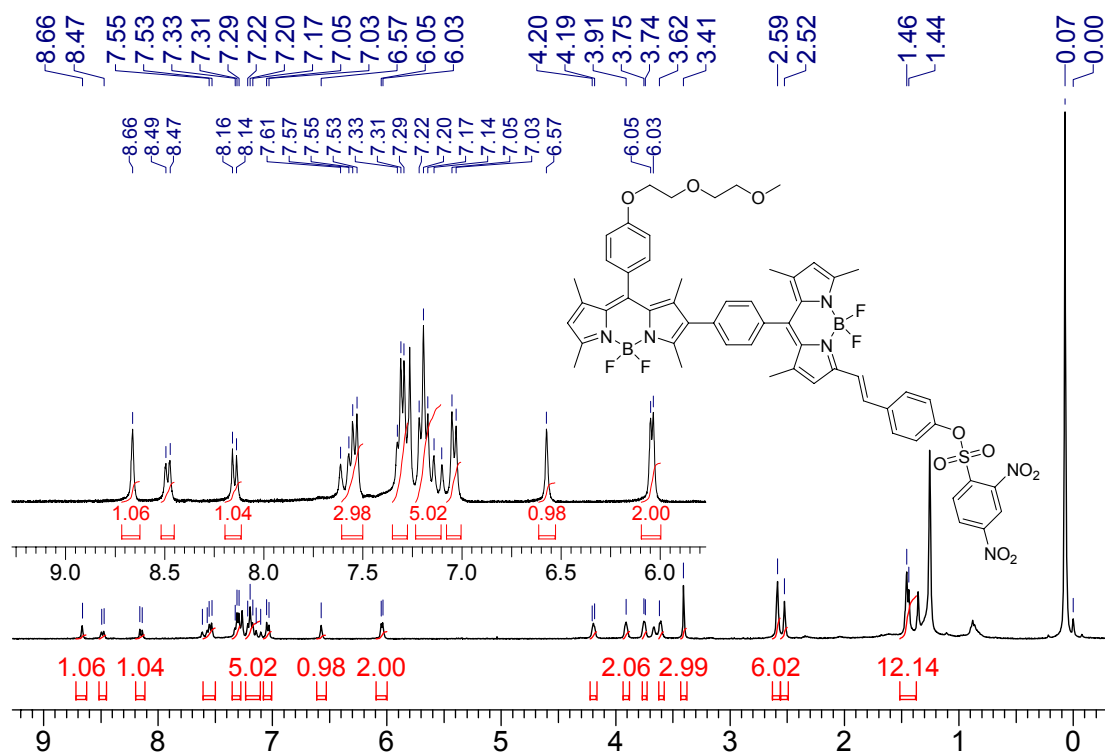


Figure S17. ^1H NMR of Probe 1. (CDCl_3 , 400 MHz).

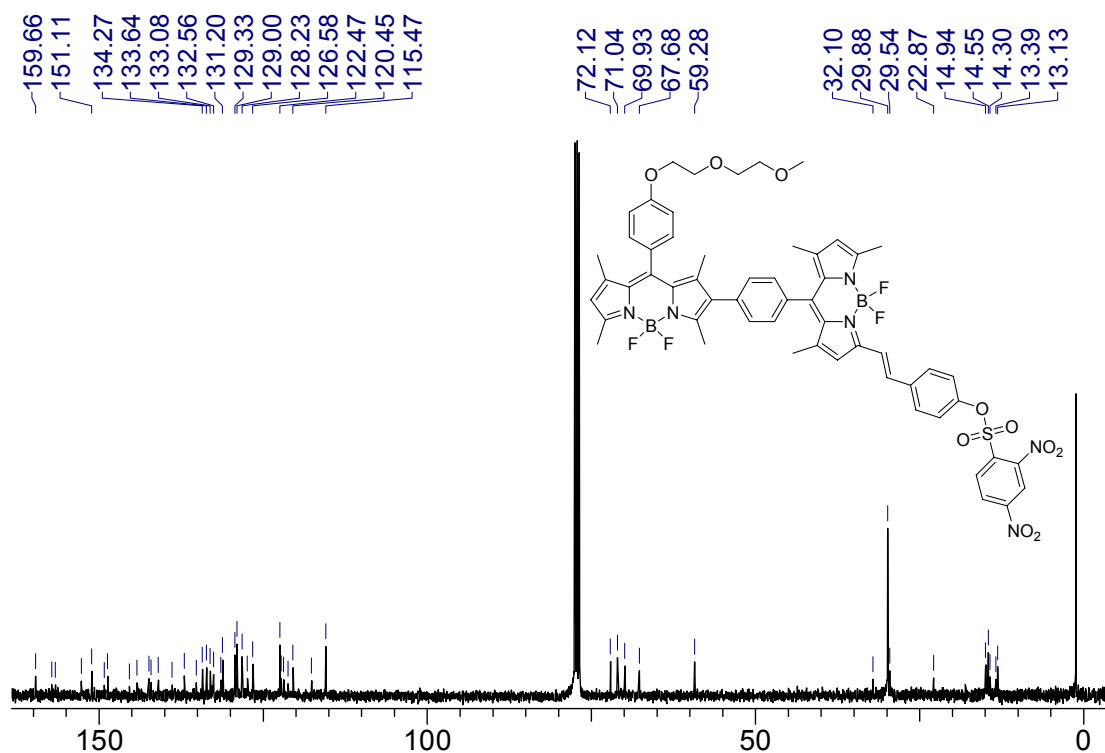


Figure S18. ^{13}C NMR of Probe 1 (CDCl_3 , 100 MHz).

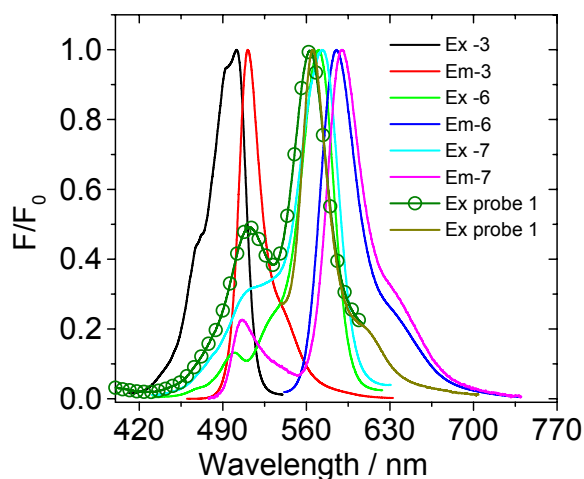


Figure S19. Relative fluorescence emission and excitation spectra of **3**, **6**, **7** and **Probe 1** in a mixture of MeOH/H₂O (V:V, 4:1). 25 °C.

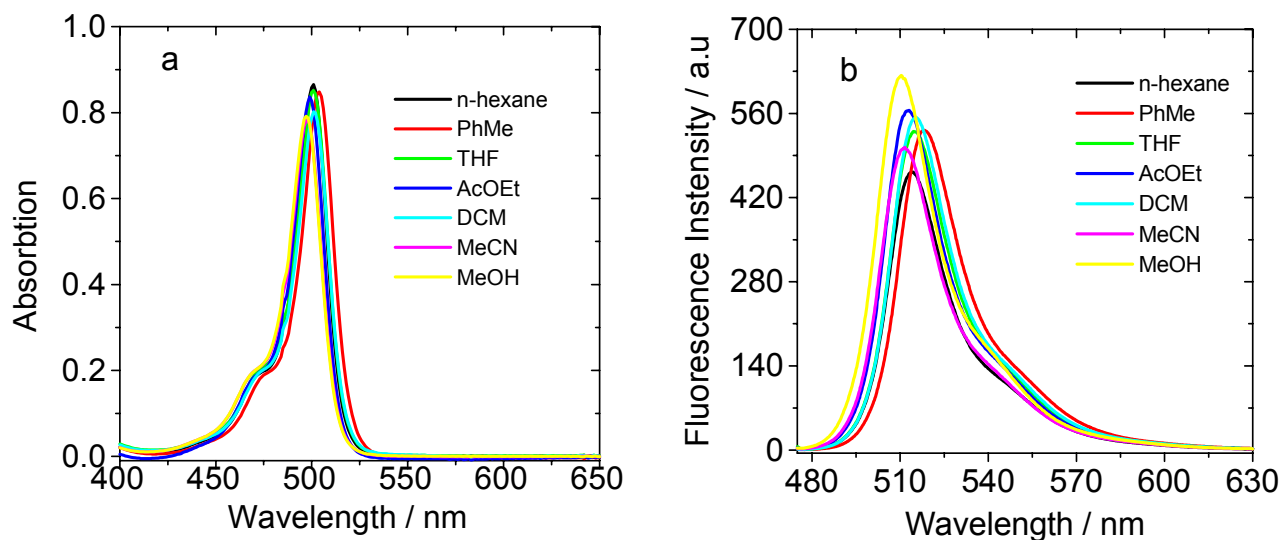


Figure S20. (a) The UV-Vis absorption of **5** in different solvents ($c = 1.0 \times 10^{-5}$ mol/L), 20 °C. (b) Fluorescence emission spectra of **5** in different solvents ($c=1.0 \times 10^{-5}$ mol/L, $\lambda_{ex} = 470$ nm), 20 °C.

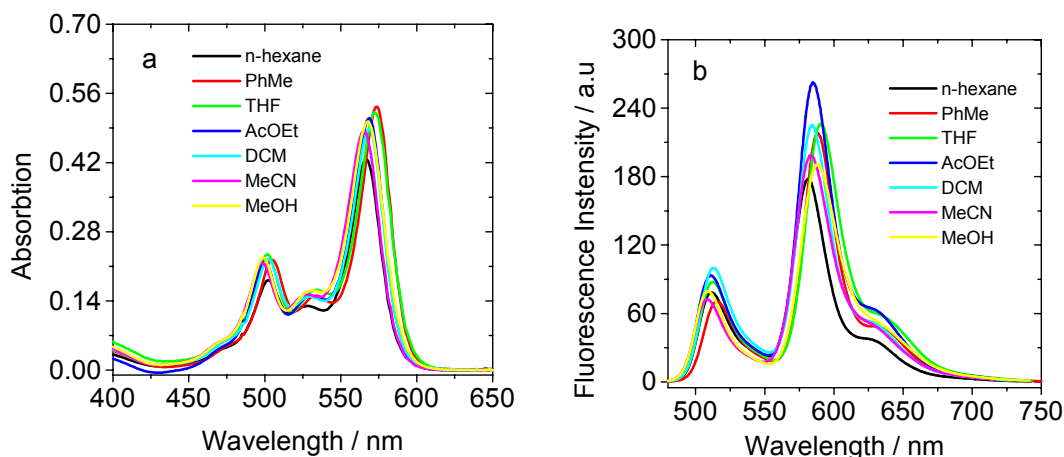


Figure S21. (a) The UV-Vis absorption of **6** in different solvents ($c = 1.0 \times 10^{-5}$ mol/L), 20 °C. (b) Fluorescence emission spectra of **6** in different solvents ($c=1.0 \times 10^{-5}$ mol/L, $\lambda_{ex} = 470$ nm), 20 °C.

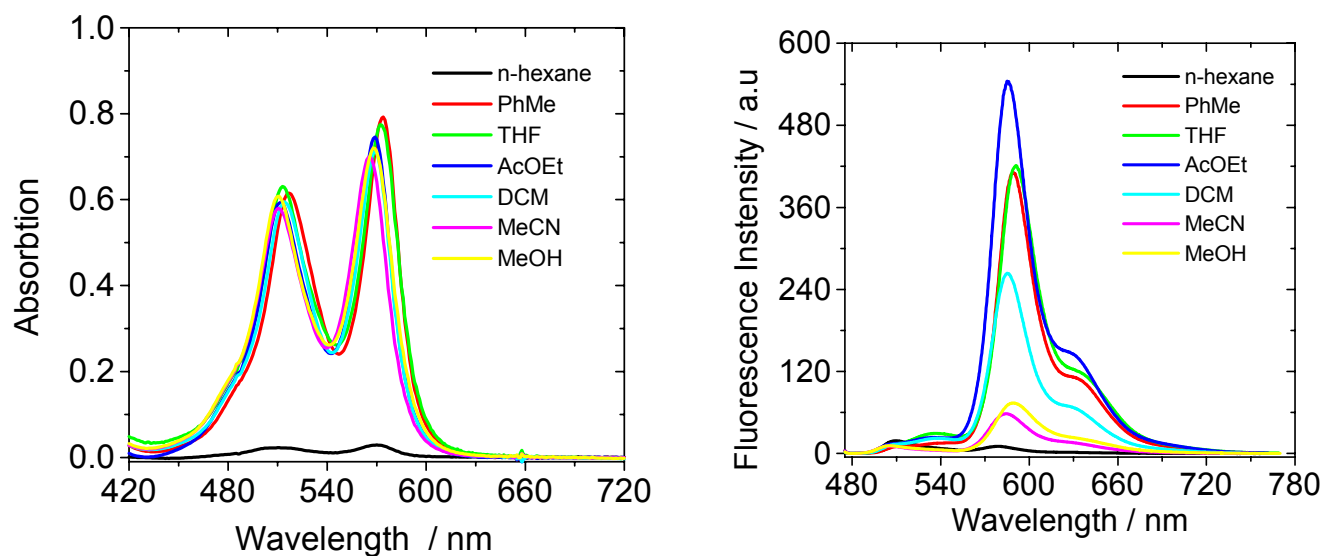


Figure S22. (a) The UV-Vis absorption of **7** in different solvents ($c = 1.0 \times 10^{-5}$ mol/L), 20 °C. (b) Fluorescence emission spectra of **7** in different solvents ($c=1.0 \times 10^{-5}$ mol/L, $\lambda_{\text{ex}} = 470$ nm), 20 °C.

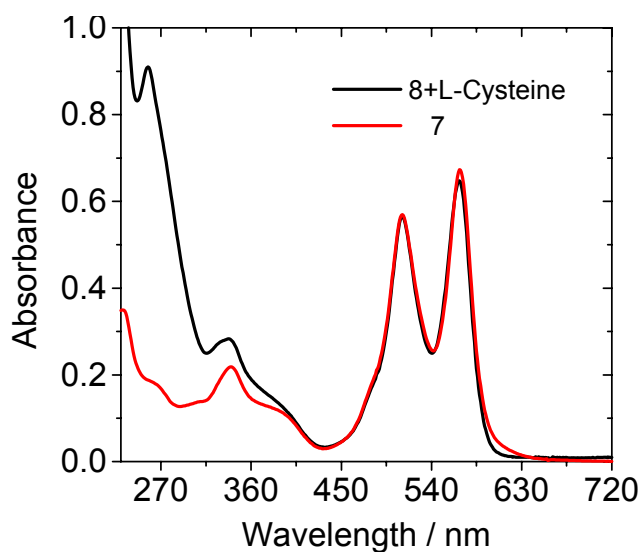


Figure S23. UV-Vis absorption of **7** and **Probe 1** after addition of L-cysteine. (MeOH/H₂O (4:1, v/v). $c_{\text{Probe}} = 1.0 \times 10^{-5}$ mol/L, $c_{\text{(L-cysteine)}} = 3.0 \times 10^{-3}$ mol/L. 37 °C)

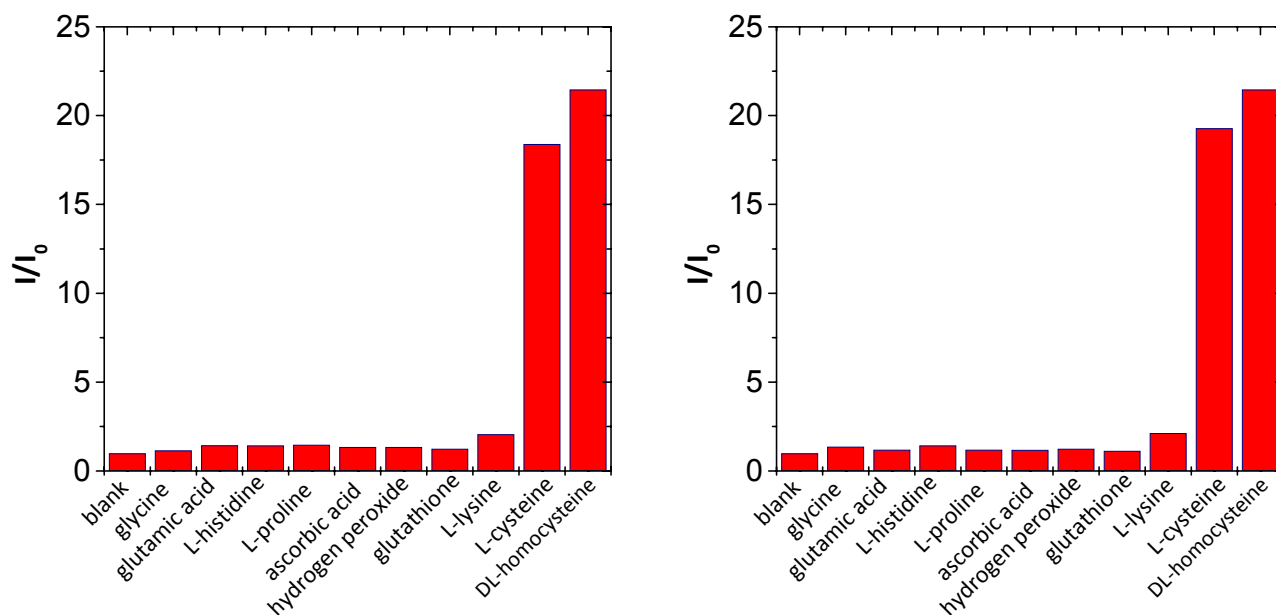


Figure S24. Response of **Probe 1** to different analytes. Relative fluorescence intensity of 10 μM probe at 590 nm ($\lambda_{\text{ex}} = 505$ nm) before and after incubation in the presence of 3.0×10^{-3} mol/L analyte at 37 °C. pH 7.4, MeOH : H₂O (V/V = 4/1).

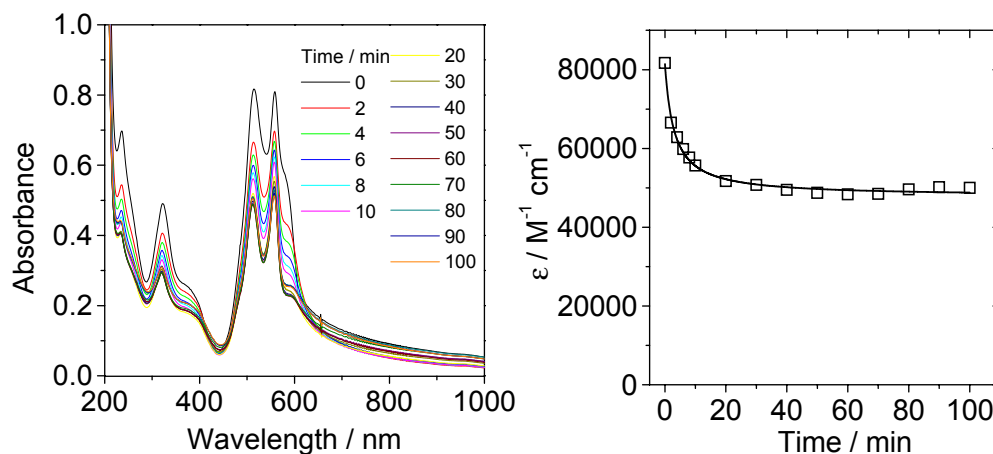


Figure S25. Photostability of the probe **1**, measured by (a) UV-vis spectral changes of probe **1** under illumination with 30 W Xenon lamp (Power density at the solution position: 57 w/m^2). (b) the evaluation of the molar extinction coefficients at 513 nm. In MeOH/H₂O (4:1, v:v). 25 °C.

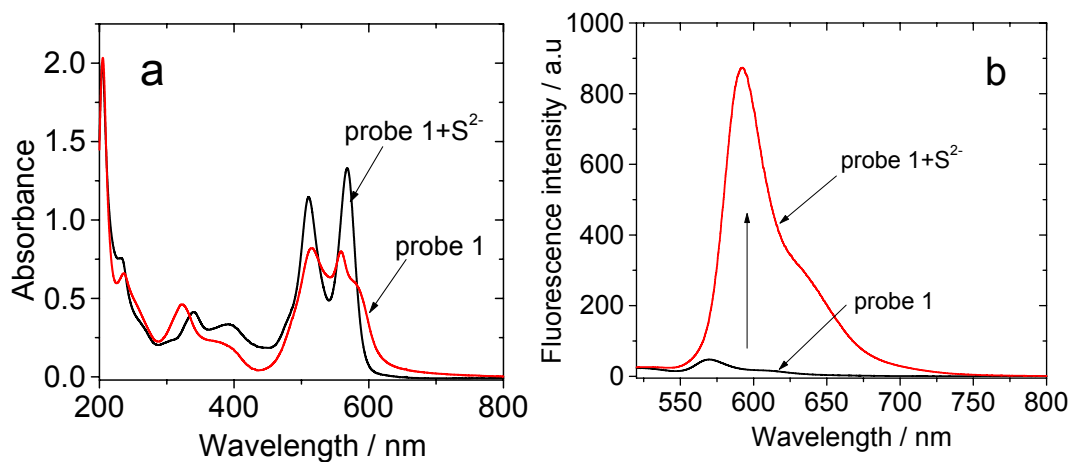


Figure S26. Response of probe **1** to Na₂S. (a) Uv-vis absorption spectra of probe **1** (10.0 μM) upon the addition of S²⁻ (10 equiv). (b) Emission spectra of probe **1** (10.0 μM) upon the addition of S²⁻ (10 equiv). The S²⁻ was added as Na₂S. In MeOH/H₂O (4:1, v:v). 25 °C.

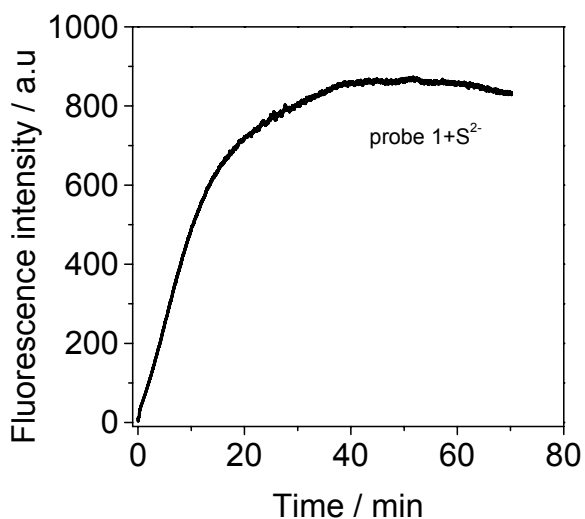


Figure S27. The sensing kinetics of probe **1** toward S²⁻. In a mixture of MeOH/H₂O (4:1, v:v). $c_{[\text{probe } 1]} = 1.0 \times 10^{-5}$ mol/L, $c[\text{S}^{2-}] = 1.0 \times 10^{-4}$ mol/L. 25 °C.

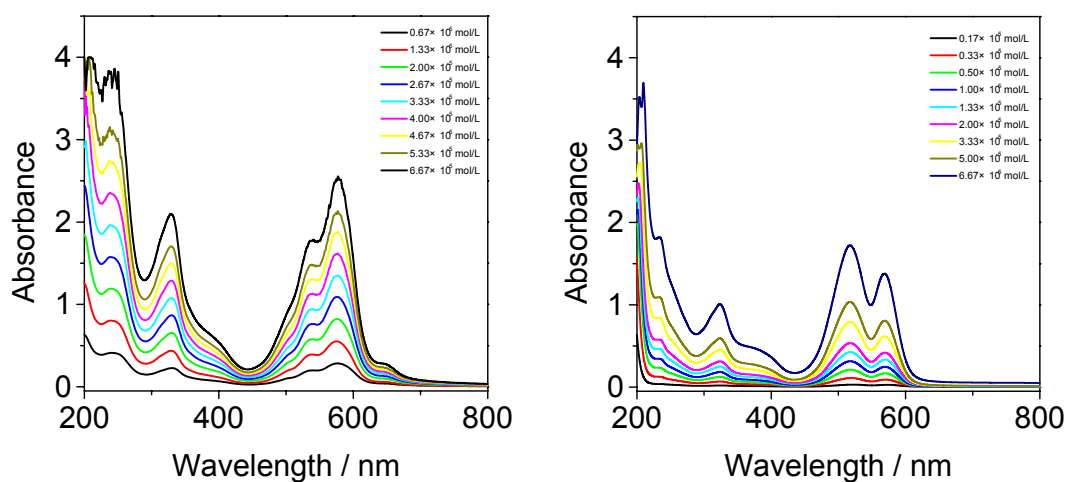
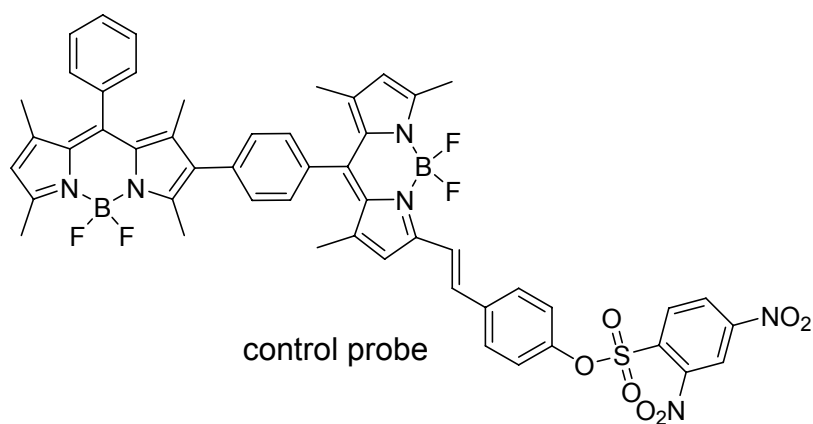


Figure S28. Solubility of the probes **1** and a control probe. The UV-vis spectral changes of the different concentration of probe **1** and the control probe in MeOH/H₂O (1:1, V:V). 25 °C.



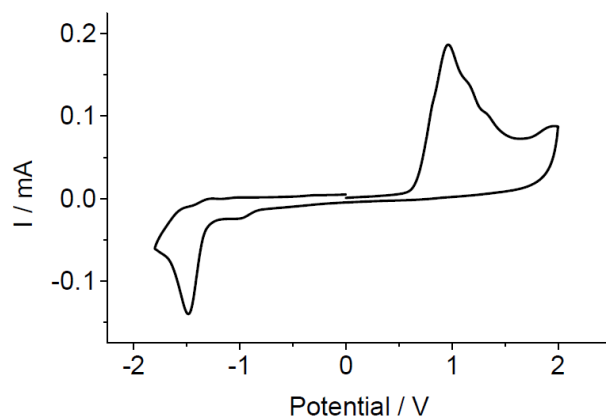


Figure S29. Cyclic voltammograms of probe **1** in CH₃CN at 20 °C using 0.1 M ⁿBu₄BF₄ as supporting electrolyte at a scan rate of 200 mV/S. Ferrocene is used as internal standard (half-wave potential = 0.06 V vs Ag/AgNO₃).

	$E^{i0}_{ox}, V (\Delta E, mV)$	$E^{i0}_{red}, V (\Delta E, mV)$	$E_{1/2}(F_c/F_c^+)$
1	0.97(irrev)	-1.50(irrev)	0.06

