Electronic Supplementary Information

Novel self-assembled dynamic [2]catenanes interlocked by the quadruple hydrogen bonding ureidopyrimidinone motif

Tangxin Xiao,^a Shao-Lu Li,^a Yajie Zhang,^a Chen Lin,^a Bingjie Hu,^b Xinchao Guan,^a Yihua Yu,^b Juli Jiang,^a and Leyong Wang^{*a}

- ^a Key Laboratory of Mesoscopic Chemistry of MOE, School of Chemistry and Chemical Engineering, Nanjing University, Nanjing 210093 (China)
- ^b Shanghai Key Laboratory of Magnetic Resonance, Department of Physics, East China Normal University, Shanghai 200062 (China)

Table of Contents

- 1. Materials and methods
- 2. Concentration-dependent ¹H NMR spectra of L1 and C1
- 3. Variable-temperature ¹H NMR spectra of C1
- 4. ESI-MS spectra of the [2] Catenane C1, C2, and C3.
- 5. Time-dependent ¹H NMR spectra of L1 and CBPQT⁴⁺
- 6. ¹*H* NMR spectra of C1 and L1 in DMSO- d_6
- 7. ${}^{1}H^{-1}H COSY and {}^{1}H^{-1}H NOESY of C1$
- 8. Synthetic procedures for the title [2] Catenanes

1. Materials and methods

All reactions were performed in atmosphere unless noted. The commercially available reagents and solvents were either employed as purchased or dried according to procedures described in the literature. All yields were given as isolated yields. NMR spectra were recorded on a Bruker DPX 300 MHz spectrometer with internal standard tetramethylsilane (TMS) and solvent signals as internal references, where CDCl₃ and CD₃CN were dried using neutral aluminium oxide. Low-resolution electrospray ionization mass spectra (LR-ESI-MS) were obtained on Finnigan Mat TSQ 7000 instruments. Elemental analyses were obtained on Perkin-Elmer 240 instruments. NOESY and DOSY experiments were performed on a Bruker DPX 500 MHz spectrometer. Viscosity measurements were carried out with Ubbelohde micro viscometers (Shanghai Liangjing Glass Instrument Factory, 0.40 mm and 0.71 mm inner diameter) at 298 K in chloroform/ acetonitrile (ν/ν , 1/1). Cyclobis(paraquat-*p*-phenylene) tetra(hexafluorophosphate)^{S1}, 1^{S2}, 2^{S3}, **M1**^{S4}, and **M2**^{S5} was prepared according to literature procedure.

2. Concentration-dependent ¹H NMR spectra of L1 and C1

In order to compare the assembly behavior of L1 and C1, we performed all the ¹H NMR experiments in the mixed solvents of acetonitrile and chloroform. The UPy N–H signals still showed a large downfield shift (between 10 and 13.5 ppm) in the mixed solvents, giving direct evidence for the UPy unit dimerization. According to literature, there are many previous examples showing the strong UPy dimerization in acetonitrile or acetonitrile/chloroform (1/1, v/v), which could promote the formation of pseudo[2]rotaxane dimer or supramolecular polymers.^{S6}



Figure S1 ¹H NMR spectra (300MHz, CDCl₃/CD₃CN = 1/1, v/v, 298K) of **L1** at different monomer concentrations: (a) 4, (b) 16, (c) 32, (d) 75, (e) 150, and (f) 200 mM. The asterisk symbol stands for minor tautomers arised from competitive intramolecular hydrogen bonding of the ether atoms of the oligo(ethylene oxide) (oligoEO) chain to the hydrogen bond donors of the **UPy** unit.^{S7}



Figure S2 ¹H NMR spectra (300MHz, CDCl₃/CD₃CN = 1/1, *v*/*v*, 298K) of **C1** at different monomer concentrations: (a) 2, (b) 10, (c) 50, (d) 100, (e) 150, and (f) 200 mM.

As can be seen from **Figure S2**, no chemical shift has been occurred upon increasing concentration, indicating no polymerization process happened during the procedure. As we know, substituents on the flexible chain have a large influence on the concentration and polymerizability of cyclic products formed. No doubt, the substituents in cyclic monomer exists less steric repulsion compared to polymer. Consequently, large or multiple substituents can decrease or even prevent polymerization. In our system, the entropy of the polypseudorotaxane should be largely decreased due to its rotational restriction by **CBPQT**⁴⁺, whereas the entropy of the [2]catenane is not altered much, as compared to **L1**.



3. Variable-temperature ¹H NMR spectra of C1

Figure S3(a) Partial ¹H NMR spectrum(500MHz, CDCl₃/CD₃CN = 1/1, ν/ν) of **C1** at different temperature: (a)

280, (b) 293, (c) 308, (d) 313, (e) 318, and (f) 328 K.



Figure S3(b) The pirouetting rotation model of the [2]catenane C1

The minor set of peaks were generated from the state of (B) and (D) (Figure S3(b)), which

were relatively less stable than (A) and (C). The disappearance of the smaller one at high temperature can be ascribed to the rapid pirouetting of the neutral component loop around **CBPQT**⁴⁺ ring.

4. ESI-MS spectra of the [2]Catenane C1 and C2.

- (a) ESI-MS of the tetrahexafluorophosphate salt C1: calcd for [C1-2PF₆⁻]²⁺ m/z = 851.35, found m/z = 851.85; calcd for [C1- HPF₆-2PF₆⁻]²⁺ m/z = 778.36, found m/z = 779.00; calcd for [C1- 2HPF₆-2PF₆⁻]²⁺ m/z = 705.38, found m/z = 706.20; calcd for [C1-3PF₆⁻]³⁺ m/z = 519.24, found m/z = 519.80.
- (b) ESI-MS of the tetrahexafluorophosphate salt **C2**: calcd for $[C2-2PF_6^-]^{2+} m/z = 826.81$, found m/z = 826.65; calcd for $[C2-HPF_6^-2PF_6^-]^{2+} m/z = 753.83$, found m/z = 753.65; calcd for $[C2-3PF_6^-]^{3+} m/z = 502.89$, found m/z = 502.55. Signals from the free neutral component is also found: calcd for $[L2+H]^+ m/z = 843.49$, found m/z = 843.45; calcd for $[L2+Na]^+ m/z = 865.48$, found m/z = 865.44.



Figure S4(a) ESI-MS spectrum of the [2]catenane C1



Figure S4(b) ESI-MS spectrum of the [2]catenane C2

5. Time-dependent ¹H NMR spectra of L1 mixed with CBPQT⁴⁺



Figure S5 Partial ¹H NMR spectrum (300MHz, CDCl₃/CD₃CN = 1/2, ν/ν , 298K) showing the formation of the [2]catenane C1 from L1 (8 mM) and CBPQT⁴⁺ (ca. 1.5 equiv.) by using the method of

"threading-followed-by-selective cyclization" with the reaction temperature at 50 °C. From bottom to top: time = (a) 0, (b) 15 min, (c) 1, (d) 12, and (e) 36 h. Designations c = complexed and uc = uncomplexed.

Further evidence showed that using excess **L1** could not only shorten the reaction time (within 24 h), but also greatly reduce the purification procedure, where excess **L1** in product can be removed only by rinsing with CHCl₃ easily.

6. ¹H NMR spectra of C1 and L1 in DMSO- d_6

The quadruple hydrogen bond was completely broken in DMSO due to its strong acceptor nature. So the [2]catenane C1 was dissociated into a pseudo[2]rotaxane (R1) and a certain amount of free L1 and CBPQT⁴⁺ in DMSO-*d*₆. The spectrum of C1 in DMSO-*d*₆ was complicated because of the overlapping peaks. The figure indicated that one of the N-*H* protons (H_c) was folded with H_{4/8} in naphthylene ring of L1 (Figure S6(a)), and these two peaks were folded again with H_β of R1 in DMSO-*d*₆ (Figure S6(b) and (c)). In contrast to L1 in DMSO-*d*₆, the N-*H* protons of R1 showed a minor downfield shift, suggesting that the UPy motif emanating from naphthalene ring by oligoEO chain was located at the unshielding area of the paraquat moieties. It was wholly different from C1 in CDCl₃ and CD₃CN mixed solvents.



Figure S6 Partial ¹H NMR spectra(300MHz, DMSO-d₆, 298K) of (a) L1; the [2]catenane C1 dissolved in

DMSO- d_6 (ca. 12 mM) after: (b) 0.5, (c) 120 min; and d) **CBPQT**⁴⁺. Designations α = **CBPQT**⁴⁺ N-CH=CH, β = **CBPQT**⁴⁺ N-CH=CH, Ar = **CBPQT**⁴⁺ Aril -H, Bz = **CBPQT**⁴⁺ benzyl-H. The chemical shift at 2.50 ppm and 3.33 ppm are solvent peaks for DMSO and H₂O, respectively. The asterisk symbol stands for residual CHCl₃. Red peaks represent complexed compound while blue peaks indicate free components.

7. ¹H-¹H COSY and ¹H-¹H NOESY spectrum of C1

The two-dimensional COSY NMR spectrum (**Figure S7**) of **C1** at 50 mM showed the proton H- α was correlated with the protons H- β on the paraquat groups. Noteworthy, similar correlations were observed between H_{2/6}, H_{3/7}, and H_{4/8} of the naphthalene protons. Hence, H_{4/8} (2.36 ppm) folded with *CH*(CH₂)₂ in the alkyl chain of **UPy** could be accurately assigned in the ¹H NMR spectra. The two-dimensional NOESY NMR spectrum of **C1** at 50 mM is displayed in **Figure S8**. The spectrum showed the protons H_{3/7} was correlated with H_{Ar}. Similar correlations were observed for oligo(ethylene oxide) (oligoEO) chain with *p*-xylene group of **CBPQT⁴⁺** (H_{EO} and H_{Ar}) and **UPy** motif (H_{EO} and H_c). From these information we could accurately assign the ¹H NMR spectrum of **C1**.



Scheme S1. Structure of the [2]catenane C1



Figure S7 Partial ¹H-¹H COSY (300MHz, CDCl₃/CD₃CN = 1/1, *v*/*v*, 298K) of **C1**



Figure S8 Partial ¹H-¹H NOESY(400MHz, CDCl₃/CD₃CN = 1/1, v/v, 298K) of C1



8. Synthetic procedures for the title [2]Catenanes

Scheme S2. Synthesis of the [2]catenane C1



Scheme S4. Synthesis of the [2]catenane C2

8.1 Synthesis of Compound L1

Imidazolide **M1** (0.38 g, 1.25 mmol) and **1** (0.21 g, 0.50 mmol) were dissolved in dry CHCl₃ (20 mL) and this solution was stirred for three hours under nitrogen at r.t. To the above reaction mixture, CHCl₃ (20 mL) was added and the organic layer was washed with 1N HCl (20 mL), saturated NaHCO₃ (20 mL), brine (20 mL), and dried over Na₂SO₄. After the solvent was removed, the resulting residue was subjected to column chromatography (CHCl₃/MeOH 150:1 (ν/ν)) to give **L1** (0.32 g, 71%) as a colorless viscous solid. ¹H NMR (300MHz, DMSO-*d₆*): δ 11.37 (*br* s, 2H, N*H*), 9.59 (*br* s, 2H, N*H*), 7.74-7.70 (m, 4H, 2H N*H* and 2H naphthalene *H*-4,8), 7.36 (t, *J* = 8.0 Hz, 2H, naphthalene *H*-3,7), 6.97 (d, *J* = 7.7 Hz, 2H, naphthalene *H*-2,6), 5.74 (s, 2H, alkylidene-*H*), 4.25 (m, 4H, OC*H*₂), 3.89 (m, 4H, OC*H*₂), 3.69 (m, 4H, OC*H*₂), 3.60 (m, 4H, OC*H*₂), 3.51 (m, 4H, OC*H*₂), 3.32 (m, 4H, C*H*₂N), 2.21 (m, 2H, C*H*(CH₂)₂), 1.49 (m, 8H, C*H*₂), 1.34-1.02 (m, 8H, C*H*₂), 0.82-0.71 (m, 12H, C*H*₃) ppm; ¹³C NMR (75 MHz, CDCl₃) δ 172.7,

156.9, 155.1, 154.6, 154.5, 126.8, 124.4, 114.6, 105.9, 104.7, 72.0, 70.8, 70.2, 69.9, 67.8, 45.3, 40.3, 32.9, 29.5, 26.6, 22.6, 14.1, 11.8 ppm; ESI-MS: m/z 893.17 [M + H]⁺ (100%); Elemental analysis calcd (%) for **L1** (C₄₆H₆₈N₈O₁₀): C 61.86, H 7.67, N 12.55; found C 61.93, H 7.81, N 12.29.



Figure S9(a) ¹H NMR spectrum (300MHz, DMSO- d_6 , 298K) of L1



Figure S9(b) ¹³C NMR spectrum (75MHz, CDCl₃, 298K) of L1



Figure S9(c) Electrospray ionization mass spectrum of L1

8.2 Synthesis of Compound C1

To a 50 mL three-necked flask, a solution of **CBPQT⁴⁺** (0.24 g, 0.22 mmol) in CH₃CN (20 mL) and a solution of **L1** (0.98 g, 1.10 mmol) in CHCl₃ (10 mL) were added under nitrogen atmosphere. The resulting mixture was stirred at 50 °C for 24 h. After the solvent was evaporated

by rotary evaporation at 30 °C, the residue was dispersed in CHCl₃ (20 mL) and stirred for 15 min. The obtained suspension was centrifuged at 3000 rpm for 30 min, followed by filtration. The solid was rinsed thoroughly with CHCl₃ to give the desired product **C1** (0.41 g, 93%) as a purple powder. ¹H NMR (300 MHz, CDCl₃/CD₃CN = 1/1, ν/ν): δ 12.89 (d, J = 12.0 Hz, 2H, NH-C=N), 10.32 (s, 2H, NH(C=O)NH-Ar), 9.47 (d, J = 6.2 Hz, 2H, α -H in CBPQT⁴⁺), 9.33-9.17 (m, 2H, NH (C=O)NH-Ar), 8.77 (d, J = 6.3 Hz, 2H, α -H in CBPQT⁴⁺), 8.69 (d, J = 6.5 Hz, 2H, α -H in CBPQT⁴⁺), 8.25-7.80 (m, 10H, α -H in CBPQT⁴⁺-2H and C₆H₄ in CBPQT⁴⁺-8H), 7.42 (d, J = 5.0 Hz, 2H, β -H in CBPQT⁴⁺), 7.33 (t, J = 5.4 Hz, 2H, β -H in CBPQT⁴⁺), 7.23 (d, J = 5.2 Hz, 2H, β -H in CBPQT⁴⁺), 7.09 (m, 2H, β -H in CBPQT⁴⁺), 6.21 (d, J = 7.8 Hz, 2H, naphthalene H-2,6), 5.93 (t, J = 8.0 Hz, 2H, naphthalene H-3,7), 5.81-5.54 (m, 10H, (C=O)CH=C-2H and CH₂ in CBPQT⁴⁺-8H), 4.61-2.80 (m, 24H, protons in EO chain), 2.49-2.29 (m, 4H, 2H in naphthalene H-4,8 and 2H in CH(CH₂)₂), 1.78-1.48 (m, 8H, CH₂), 1.49-1.08 (m, 8H, CH₂), 1.07-0.78 (m, 12H, CH₃) ppm; ESI-MS: m/z 519.24 [M–3PF₆]³⁺ (100%), 778.36 [M–HPF₆–2PF₆]²⁺ (50%), 851.85 [M–2PF₆]²⁺ (31%); Elemental analysis calcd (%) for **C1** (C₈₂H₁₀₀F₂₄N₁₂O₁₀P₄): C 49.40, H 5.06, N 8.43; found C 49.24, H 5.28, N 8.21.



Figure S10 ¹H NMR spectrum (300MHz, CD₃CN/CDCl₃ = 1/1, v/v, 298K) of **C1**, The asterisk symbol stands for another set of signals as a result of a pirouetting motion of the two rings (For detail, please see the variable temperature ¹H NMR).

8.3 Synthesis of Compound L2

Imidazolide **M1** (0.73 g, 2.42 mmol) and **2** (0.41 g, 1.10 mmol) were dissolved in dry CHCl₃ (20 mL) and this solution was stirred for three hours under nitrogen at 25 °C. To the above reaction mixture, CHCl₃ (20 mL) was added and the organic layer was washed with 1N HCl (20 mL), saturated NaHCO₃ (20 mL), brine (20 mL) and dried over Na₂SO₄. After the solvent was removed, the resulting residue was subjected to column chromatography with CHCl₃ as eluent to give **L2** (0.59 g, 64%) as a colorless viscous solid. ¹H NMR (300MHz, CDCl₃): δ 13.20 (br s, 2H, N*H*), 11.92 (br s, 2H, N*H*), 10.36 (br s, 2H, N*H*), 6.72 (s, 4H, Ar-*H*), 5.81 (s, 2H, alkylidene-*H*), 3.96-3.49(m, 24H, OC*H*₂), 2.28 (m, 2H, C*H*(CH₂)₂), 1.58 (m, 8H, C*H*₂), 1.25 (m, 8H, C*H*₂), 0.87(m, 12H, C*H*₃) ppm; ¹³C NMR (75MHz, DMSO-*d*₆): δ 170.7, 161.3, 154.5, 152.5, 151.6, 115.3, 105.2, 69.9, 69.7, 69.2, 69.1, 67.5, 47.9, 39.2, 32.9, 29.1, 26.5, 22.2, 13.8, 11.8 ppm; ESI-MS: *m*/z 843.33 [M+H]⁺ (53%), 865.33 [M+Na]⁺ (100%); Elemental analysis calcd (%) for **L2** (C₄₂H₆₆N₈O₁₀): C 59.84, H 7.89, N 13.29; found C 59.99, H 7.69, N 13.48.



Figure S11(a) ¹H NMR spectrum (300MHz, CDCl₃, 298K) of L2



Figure S11(b) ¹³C NMR spectrum (75MHz, DMSO-*d*₆, 298K) of L2



Figure S11(c) Electrospray ionization mass spectrum of L2

8.4 Synthesis of Compound C2

To a round bottom flask, a solution of **CBPQT**⁴⁺ (0.060 g, 0.055 mmol) in CH₃CN (10 mL) and a solution of **L2** (0.18 g, 0.22 mmol) in CHCl₃ (5 mL) were added under nitrogen atmosphere. The resulting mixture was stirred at 50 °C for 24 h. After the solvent was evaporated by rotary evaporation at 30 °C, the residue was dispersed in CHCl₃ (10 mL) and stirred for 15 min. The obtained suspension was centrifuged at 3000 rpm for 30 min, followed by filtration. The solid was rinsed thoroughly with CHCl₃ to give the desired product **C1** (0.10 g, 90%) as a magenta powder. ¹H NMR (300MHz, CD₃CN/CDCl₃ = 1/1, ν/ν , 298 K) of the major component: δ 12.88 (s, 2H, NH-C=N), 10.55(s, 2H,C-NH (C=O)NH-Ar), 9.54 (s, 2H, C-NH (C=O)NH-Ar), 9.11-8.60 (m, 8H, α -*H* in CBPQT⁴⁺), 7.95-7.69 (m, 16H, β -*H* and C₆H₄ in CBPQT⁴⁺), 5.66(s, 8H, CH₂ in CBPQT⁴⁺), 5.64 (s, 2H, (C=O)CH=C)), 4.20-3.44 (m, 26H, 22H in OCH₂ and 4H in C₆H₄), 2.92 (*br* s, 2H, OCH₂), 2.41 (s, 2H, CH(CH₂)₂), 1.91-1.52 (m, 8H, CH₂), 1.45-1.15 (m, 8H, CH₂), 1.08-0.76(m, 12H, CH₃) ppm; ESI-MS: *m*/z 826.65 [M-2PF₆]²⁺, 502.55 [M-3PF₆-]³⁺; Elemental analysis calcd (%) for **C2** (C₇₈H₉₈F₂₄N₁₂O₁₀P₄): C 48.20, H 5.08, N 8.65; found C 48.40, H 5.25, N 8.42.



• Signals from the free **CBPQT**⁴⁺macrocycle, * Signals from the free neutral component

References:

- S1. M. Asakawa, W. Dehaen, G. Labbe, S. Menzer, J. Nouwen, F. M. Raymo, J. F. Stoddart and D. J. Williams, *J. Org. Chem.*, 1996, **61**, 9591-9595.
- S2. S.-Y. Chang, J. S. Choi and K.-S. Jeong, Chem. -Eur. J. 2001, 7, 2687-2697.
- S3. Q. Z. Zhou, C. L. He, H. N. Gu, Q. M. Miao and C. X. Zhai, *Chin. Chem. Lett.*, 2008, 19, 911-914.
- S4. H. M. Keizer, R. P. Sijbesma and E. W. Meijer, Eur. J. Org. Chem. 2004, 2553-2555.
- S5. C.-H. Wong, H.-F. Chow, S.-K. Hui and K.-H. Sze, Org. Lett. 2006, 8, 1811-1814.
- S6. (a) Y. Tokunaga and T. Seo, *Chem. Commun.* 2002, 970-971.; (b) H. Hofmeier, A. El-ghayoury, A. P. H. J. Schenning and U. S. Schubert, *Chem. Commun.* 2004, 318-319.; (c) S.-L. Li, T. Xiao, Y. Wu, J. Jiang and L. Wang, *Chem. Commun.* 2011, 47, 6903-6905.
- S7. T. F. A. de Greef, M. M. L. Nieuwenhuizen, R. P. Sijbesma and E. W. Meijer, *J. Org. Chem.* 2010, **75**, 598-610.