

Supporting Information

**Combining Zn Ion Catalysis with Homogeneous Gold
Catalysis: An Efficient Annulation Approach to *N*-Protected
Indoles**

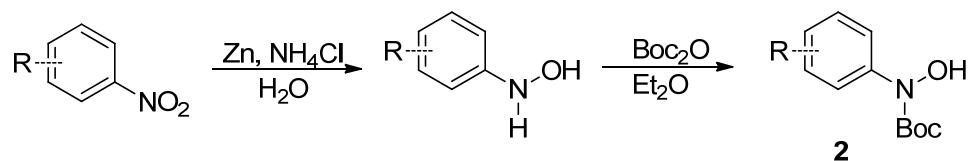
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California, 93117

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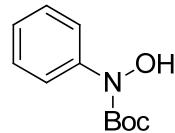
General. Ethyl acetate (ACS grade), hexanes (ACS grade), diethyl ether (ACS grade), were purchased from Fisher Scientific and used without further purification. Anhydrous toluene and 1,2-dichloroethane were bought from Acros and used directly. Commercially available reagents were used without further purification. Reactions were monitored by thin layer chromatography (TLC) using Silicycle precoated silica gel plates. Flash column chromatography was performed over Silicycle silica gel (230-400 mesh). ^1H NMR and ^{13}C NMR spectra were recorded on a Varian 500 MHz Unity plus spectrometer and a Varian 400 MHz spectrometer using residue solvent peaks as internal standards (CDCl_3 , ^1H : 7.26 ppm; ^{13}C : 77.00 ppm), (CD_3SOCD_3 , ^1H : 2.50 ppm; ^{13}C : 39.5 ppm). Infrared spectra were recorded with a Perkin Elmer FT-IR spectrum 2000 spectrometer and are reported in reciprocal centimeter (cm^{-1}). Mass spectra were recorded with Waters micromass ZQ detector using electrospray method.

General Procedure A: preparation of *tert*-butyl hydroxy(aryl)carbamate



The precursor *N*-phenylhydroxylamine was synthesized according to literature procedures.^[1] To a stirred mixture of nitrobenzene (21 mmol) and NH₄Cl (1.3 g, 24 mmol) in H₂O (40 mL) was slowly added zinc dust (90%, 3.08 g, 42 mmol) while maintaining the temperature below 60 °C. After 15 min's stirring, the reaction mixture was filtered while still warm and the solid was washed with hot water (10 mL). The filtrate was saturated with NaCl and cooled to 0 °C and the resulting solid was collected and dried. This crude *N*-phenylhydroxylamine was recrystallized from hexane-petroleum ether. To a solution of *N*-phenylhydroxylamine (10 mmol) in dry diethyl ether (50 mL) was added di-*t*-butyl dicarbonate (2.8 g, 10 mmol) at room temperature. After 12 h, the reaction was concentrated and the residue was purified through silica gel flash chromatography to give the desired product *tert*-butyl hydroxy(aryl)carbamate 2.

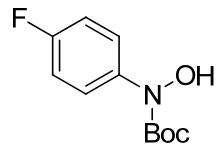
tert-Butyl hydroxy(phenyl)carbamate (2a)



2a

This compound was known and prepared in 70% yield according to the general procedure A (eluents: ethyl acetate: hexanes = 1:20). ^1H NMR (500 MHz, CDCl_3) δ 7.67 (bs, 1H), 7.47 (dt, 2H, J_1 = 8.5 Hz, J_2 = 1.5 Hz), 7.35 (t, 2H, J = 8 Hz), 7.16 (t, 1H, J = 7.5 Hz), 1.52 (s, 9H); ^{13}C NMR (125 MHz, CDCl_3) δ 154.3, 140.7, 128.3, 125.1, 121.4, 83.4, 28.2; IR (neat): 3218, 2979, 2933, 1682, 1596, 1495, 1368, 1125.

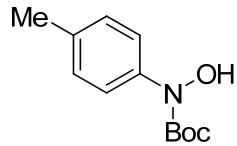
***tert*-Butyl 4-fluorophenyl(3hydroxyl)carbamate (2b)**



2b

This compound was prepared in 66% yield according to the general procedure A (eluents: ethyl acetate: hexanes = 1:15). ^1H NMR (400 MHz, CDCl_3) δ 8.26 (bs, 1H), 7.37 – 7.42 (m, 2H), 6.99 – 7.05 (m, 2H), 1.47 (s, 9H); ^{13}C NMR (125 MHz, CDCl_3) δ 160.2 (d, J = 243 Hz), 154.7, 137.1, 124.0 (d, J = 9 Hz), 115.1 (d, J = 23 Hz), 83.4, 28.2; IR (neat): 3238, 3232, 2980, 2934, 1695, 1683, 1507, 1369, 1230, 1126, 836; 250.09; HRMS (ES $^+$) calculated for $[\text{C}_{11}\text{H}_{14}\text{FNNaO}_3]^+$: 250.0855, found: 250.0853.

***tert*-Butyl hydroxy(p-tolyl)carbamate (2c)**

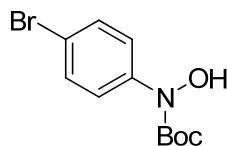


2c

This compound was known and prepared in 70% yield according to the general procedure A (eluents: ethyl acetate: hexanes = 1:15). ^1H NMR (400 MHz, CDCl_3) δ 7.82 (bs, 1H), 7.32 (d, 2H, J = 8.8 Hz), 7.14 (d, 2H, J = 8 Hz), 2.34 (s, 3H), 1.49 (s, 9H); ^{13}C NMR (100

MHz, CDCl₃) δ 154.5, 138.3, 135.1, 128.9, 121.9, 28.2, 20.9; IR (neat): 3194, 3130, 2975, 2925, 1694, 1667, 1509, 1367, 1161, 1122, 1106.

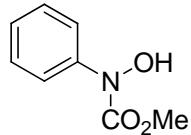
tert-Butyl 4-bromophenyl(hydroxy)carbamate (2d)



2d

This compound was prepared in 60% yield according to the general procedure A (eluents: ethyl acetate: hexanes = 1:25). ¹H NMR (400 MHz, CDCl₃) δ 7.92 (bs, 1H), 7.44 (dt, 2H, J₁ = 9.2 Hz, J₂ = 2.4 Hz), 7.34 (dt, 2H, J₁ = 9.2 Hz, J₂ = 2.4 Hz), 1.50 (s, 9H); ¹³C NMR (125 MHz, CDCl₃) δ 154.0, 139.9, 131.3, 122.7, 118.0, 83.9, 28.2; IR (neat): 3239, 2978, 2931, 1691, 1486, 1370, 1340, 1130; HRMS (ES+) calculated for [C₁₁H₁₄BrNNaO₃]⁺: 310.0055, found: 310.0046.

Methyl hydroxy(phenyl)carbamate (2e)

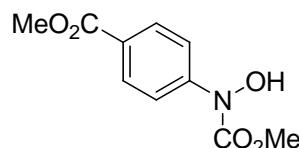


2e

This compound was known and prepared in 72% yield according to literature procedures.
^[2] ¹H NMR (500 MHz, CDCl₃) δ 7.85 (bs, 1H), 7.47 – 7.49 (m, 2H), 7.37 (tt, 2H, J₁ = 8 Hz, J₂ = 2 Hz), 7.21 (tt, 1H, J₁ = 7.5 Hz, J₂ = 1.3 Hz), 3.82 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 156.0, 140.5, 128.6, 126.0, 122.1, 53.8.

2f, 2g, 2h, 2i were prepared according to the literature procedures. ^[2] ^[3] ^[4]

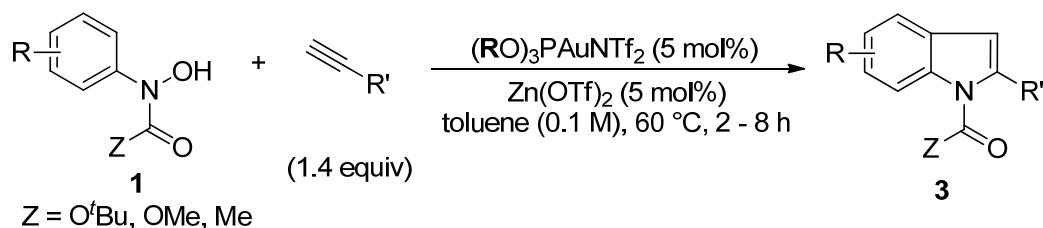
Methyl 4-(hydroxy(methoxycarbonyl)amino)benzoate (2g)



2g

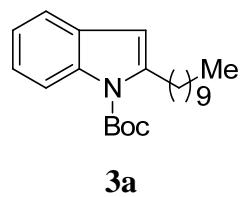
This compound was prepared in 70 % yield according to literature procedures.^[2] (eluents: ethyl acetate: hexanes = 1:8). ¹H NMR (400 MHz, CDCl₃) δ 7.99 (d, 2H, J = 8.4 Hz), 7.60 (d, 2H, J = 8.8 Hz), 3.90 (s, 3H), 3.88 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 166.8, 154.9, 144.3, 130.2, 126.0, 119.0, 54.1, 52.2; IR (neat): 3582, 3304, 3007, 2955, 2847, 1717, 1606, 1439, 1338, 1282, 1113; HRMS (ES+) calculated for [C₁₀H₁₁NNaO₅]⁺: 248.0535, found: 248.0523.

General procedure B: preparation of *N*-protected-2-alkyl indoles 3



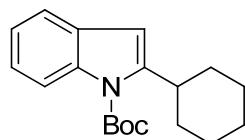
To a solution of *N*-protected hydroxylamine **2** (0.3 mmol, 1 equiv) in dry toluene (3 mL, 0.1 M) was added the corresponding alkyne (0.42 mmol, 1.4 equiv), Zn(OTf)₂ (0.015 mol%) and (ArO)₃PAuNTf₂ (Ar = 2,4-di-*tert*-butylphenyl, 0.015 mmol, 5 mol%) at room temperature. The reaction mixture was stirred at 60 °C for 2 – 8 h. Once the reaction finished by TLC, it was concentrated and the residue was purified through silica gel flash chromatography to give the desired product **3**.

***tert*-Butyl 2-decyl-1*H*-indole-1-carboxylate (3a)**



This compound was prepared in 90% yield according to the general procedure B (eluents: ethyl acetate: hexanes = 1:80). ^1H NMR (500 MHz, CDCl_3) δ 8.12 (d, 1H, J = 8.5 Hz), 8.46 – 8.48 (d, 1H, J = 8 Hz), 7.19 – 7.27 (m, 2H), 6.36 (d, 1H, J = 1 Hz), 3.02 (t, 2H, J = 8 Hz), 1.69 – 1.74 (m, 11 H), 1.41 – 1.48 (m, 2H), 1.30 – 1.39 (m, 12H), 0.91 (t, 3H, J = 7 Hz); ^{13}C NMR (125 MHz, CDCl_3) δ 150.5, 142.5, 136.6, 129.3, 123.0, 122.4, 119.5, 115.5, 106.9, 83.5, 31.9, 30.2, 29.7, 29.6, 29.5, 29.4, 29.0, 28.3, 22.7, 14.2; IR (neat): 3051, 2954, 2925, 2854, 1734, 1454, 1369, 1327, 1166, 1118, 1086; HRMS (ES+) calculated for $[\text{C}_{23}\text{H}_{35}\text{NNaO}_2]^+$: 380.2565, found: 380.2562.

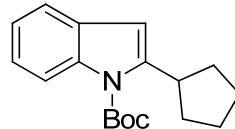
tert-Butyl 2-cyclohexyl-1*H*-indole-1-carboxylate (3b)



3b

This compound was prepared in 83% yield according to the general procedure B (eluents: ethyl acetate: hexanes = 1:100). ^1H NMR (400 MHz, CDCl_3) δ 8.09 (d, 1H, J = 8.0 Hz), 7.45 (dd, 1H, J_1 = 7.6 Hz, J_2 = 0.8 Hz), 7.15 – 7.23 (m, 2H), 6.37 (s, 1H), 3.35 (m, 1H, Hz), 2.10 (d, 1H, J = 12.8 Hz), 1.83 – 1.87 (m, 2H), 1.69 (s, 9H), 1.24 – 1.50 (m, 6H); ^{13}C NMR (100 MHz, CDCl_3) δ 150.5, 147.9, 136.7, 129.4, 123.1, 122.4, 119.7, 115.5, 104.8, 83.5, 37.4, 33.7, 28.3, 26.7, 26.4; IR (neat): 2977, 2929, 2852, 1730, 1454, 1369, 1324, 1158, 1113, 1084; HRMS (ES+) calculated for $[\text{C}_{19}\text{H}_{25}\text{NNaO}_2]^+$: 322.1783, found: 322.1774.

tert-Butyl 2-cyclopentyl-1*H*-indole-1-carboxylate (3c)

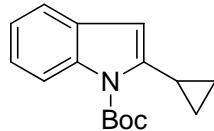


3c

This compound was prepared in 89% yield according to the general procedure B (eluents: ethyl acetate: hexanes = 1:100). ^1H NMR (400 MHz, CDCl_3) δ 8.07 (d, 1H, J = 8.0 Hz), 7.45 (dd, 1H, J_1 = 7.2 Hz, J_2 = 0.8 Hz), 7.16–7.24 (m, 2H), 6.37 (d, 1H, J = 0.8 Hz), 3.74

(m, 1H, $J = 7.6$ Hz), 2.13 – 2.17 (m, 2H), 1.71 – 1.77 (m, 2H), 1.56 – 1.70 (m, 13H); ^{13}C NMR (100 MHz, CDCl_3) δ 150.5, 146.5, 136.8, 129.2, 123.1, 122.4, 119.7, 115.4, 104.6, 83.5, 39.4, 33.0, 28.2, 24.8. IR (neat): 3050, 2959, 2869, 2852, 1732, 1454, 1369, 1327, 1159, 1115, 1089; HRMS (ES+) calculated for $[\text{C}_{18}\text{H}_{23}\text{NNaO}_2]^+$: 308.1626, found: 308.1616.

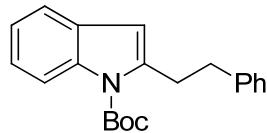
tert-Butyl 2-cyclopropyl-1*H*-indole-1-carboxylate (3d)



3d

This compound was prepared in 84% yield according to the general procedure B (eluent: ethyl acetate: hexanes = 1:100). ^1H NMR (400 MHz, CDCl_3) δ 8.13 (d, 1H, $J = 8.0$ Hz), 7.43 (dd, 1H, $J_1 = 7.6$ Hz, $J_2 = 0.8$ Hz), 7.17 – 7.24 (m, 2H), 6.24 (s, 1H), 2.34 – 2.40 (m, 1H), 1.70 (s, 9H), 0.94 – 1.00 (m, 2H), 0.72 – 0.78 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 150.6, 143.9, 136.8, 129.0, 123.3, 122.5, 119.8, 115.3, 105.6, 83.4, 28.3, 11.2, 7.6; IR (neat): 3050, 2979, 2930, 2851, 1730, 1454, 1369, 1335, 1165, 1118, 1096; HRMS (ES+) calculated for $[\text{C}_{16}\text{H}_{19}\text{NO}_2]^+$: 257.1416, found: 257.1425.

tert-Butyl 2-phenethyl-1*H*-indole-1-carboxylate (3e)

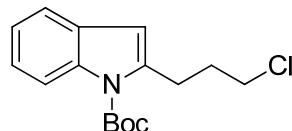


3e

This compound was prepared in 96% yield according to the general procedure B (eluent: ethyl acetate: hexanes = 1:100). ^1H NMR (400 MHz, CDCl_3) δ 8.10 (d, 1H, $J = 8.4$ Hz), 7.47 (dd, 1H, $J_1 = 7.2$ Hz, $J_2 = 0.8$ Hz), 7.21 – 7.34 (m, 7H), 6.38 (d, 1H, $J = 0.8$ Hz), 3.37 (d, 2H, $J = 7.2$ Hz), 3.05 (d, 2H, $J = 7.2$ Hz), 1.71 (s, 9H); ^{13}C NMR (100 MHz, CDCl_3) δ 150.6, 141.7, 141.5, 136.5, 129.3, 128.4, 128.3, 126.0, 123.3, 122.6, 119.8, 115.6, 107.4, 83.8, 35.3, 31.9, 28.4; IR (neat): 3051, 2977, 2925, 2851, 1732, 1460, 1369,

1328, 1158, 1115, 1086; HRMS (ES+) calculated for $[C_{21}H_{23}NNaO_2]^+$: 344.1626, found: 344.1623.

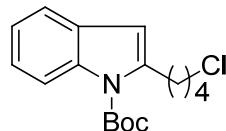
***tert*-Butyl 2-(3-chloropropyl)-1*H*-indole-1-carboxylate (3f)**



3f

This compound was prepared in 87% yield according to the general procedure B (eluent: ethyl acetate: hexanes = 1:100). 1H NMR (400 MHz, $CDCl_3$) δ 8.08 (d, 1H, J = 8.0 Hz), 7.46 (dd, 1H, J_1 = 7.8 Hz, J_2 = 1.2 Hz), 7.19 – 7.25 (m, 2H), 6.40 (s, 1H), 3.62 (t, 2H, J = 6.4 Hz), 3.20 (t, 2H, J = 7.2 Hz), 2.10 – 2.25 (m, 2H), 1.70 (s, 9H); ^{13}C NMR (100 MHz, $CDCl_3$) δ 150.4, 140.1, 136.6, 129.1, 123.5, 122.7, 119.8, 115.6, 107.9, 83.9, 44.4, 31.7, 31.5, 30.6, 28.3, 27.3; IR (neat): 3050, 2977, 2925, 2851, 1732, 1460, 1369, 1328, 1158, 1115, 1086; HRMS (ES+) calculated for $[C_{16}H_{20}ClNNaO_2]^+$: 316.1080, found: 316.1085.

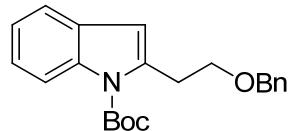
***tert*-Butyl 2-(4-chlorobutyl)-1*H*-indole-1-carboxylate (3g)**



3g

This compound was prepared in 95% yield according to the general procedure B (eluent: ethyl acetate: hexanes = 1:100). 1H NMR (400 MHz, $CDCl_3$) δ 8.07 (d, 1H, J = 8.4 Hz), 7.45 (dd, 1H, J_1 = 6.8 Hz, J_2 = 1.6 Hz), 7.18 – 7.23 (m, 2H), 6.37 (s, 1H), 3.60 (t, 2H, J = 6.4 Hz), 3.20 (t, 2H, J = 6.8 Hz), 1.86 – 1.90 (m, 2H), 1.70 (s, 9H); ^{13}C NMR (100 MHz, $CDCl_3$) δ 150.5, 141.5, 136.6, 129.2, 123.3, 122.6, 119.7, 115.6, 107.3, 83.8, 45.0, 32.3, 29.5, 28.3, 26.3; IR (neat): 3050, 2977, 2925, 2851, 1732, 1454, 1370, 1327, 1158, 1115, 1087; HRMS (ES+) calculated for $[C_{17}H_{22}ClNNaO_2]^+$: 330.1237, found: 330.1259.

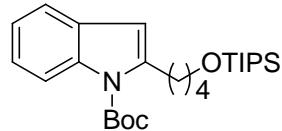
***tert*-Butyl 2-(2-(benzyloxy)ethyl)-1*H*-indole-1-carboxylate (3h)**



3h

This compound was prepared in 83% yield according to the general procedure B (eluents: ethyl acetate: hexanes = 1:100). ^1H NMR (400 MHz, CDCl_3) δ 8.12 (d, 1H, J = 8.2 Hz), 7.47 – 7.49 (m, 1H), 7.19 – 7.36 (m, 7H), 6.45 (d, 1H, J = 0.8 Hz), 4.59 (s, 2H), 3.83 (t, 2H, J = 7.2 Hz), 3.38 (td, 2H, J_1 = 6.4 Hz, J_2 = 0.8 Hz), 1.68 (s, 9H); ^{13}C NMR (100 MHz, CDCl_3) δ 150.4, 138.4, 138.2, 136.5, 129.2, 128.3, 127.7, 127.5, 123.4, 122.6, 119.8, 115.5, 108.3, 83.8, 72.9, 69.0, 30.6, 28.1; IR (neat): 3050, 2977, 2930, 2858, 1732, 1454, 1370, 1328, 1158, 1118, 1092; HRMS (ES+) calculated for $[\text{C}_{22}\text{H}_{25}\text{NNaO}_3]^+$: 374.1732, found: 374.1732.

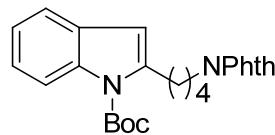
***tert*-Butyl 2-(4-((triisopropylsilyl)oxy)butyl)-1*H*-indole-1-carboxylate (3i)**



3i

This compound was prepared in 78% yield according to the general procedure B (eluents: ethyl acetate: hexanes = 1:200). ^1H NMR (400 MHz, CDCl_3) δ 8.02 (dd, 1H, J_1 = 8.8 Hz, J_2 = 0.8 Hz), 7.35 – 7.37 (m, 1H), 7.08 – 7.16 (m, 2H), 6.28 (d, 1H, J = 0.8 Hz), 3.67 (t, 2H, J = 6.0 Hz), 2.95 (t, 2H, J = 7.6 Hz), 1.69 – 1.74 (m, 2H), 1.57 – 1.62 (m, 12H), 0.95 – 0.99 (m, 18H); ^{13}C NMR (100 MHz, CDCl_3) δ 150.6, 142.3, 136.6, 129.4, 123.1, 122.5, 119.6, 115.5, 107.0, 83.6, 63.2, 32.7, 29.9, 28.2, 25.2, 18.0, 11.9; IR (neat): 3050, 2977, 2941, 2865, 1734, 1454, 1369, 1303, 1165, 1116, 1095; HRMS (ES+) calculated for $[\text{C}_{26}\text{H}_{43}\text{NNaSiO}_3]^+$: 468.2910, found: 468.2897.

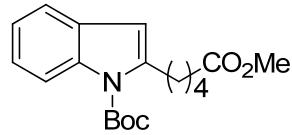
***tert*-Butyl 2-(4-(1,3-dioxoisooindolin-2-yl)butyl)-1*H*-indole-1-carboxylate (3j)**



3j

This compound was prepared in 93% yield according to the general procedure B (eluents: ethyl acetate: hexanes = 1:40). ^1H NMR (400 MHz, CDCl_3) δ 8.07 (d, 1H, J = 8.0 Hz), 7.83 – 7.85 (m, 2H), 7.69–7.2 (m, 2H), 7.42–7.44 (m, 2H), 7.16–7.21 (m, 2H), 6.34 (s, 1H), 3.75 (t, 2H, J = 7.2 Hz), 3.04 (t, 2H, J = 7.2 Hz), 1.75–1.82 (m, 4H), 1.66 (s, 9H); ^{13}C NMR (100 MHz, CDCl_3) δ 168.3, 150.4, 141.5, 136.6, 133.8, 132.1, 129.2, 123.2, 123.1, 122.5, 119.7, 115.6, 107.3, 83.7, 37.9, 29.8, 28.4, 28.3, 26.3; IR (neat): 3050, 2977, 2925, 2851, 1707, 1460, 1369, 1328, 1158, 1116, 1086; HRMS (ES+) calculated for $[\text{C}_{25}\text{H}_{26}\text{N}_2\text{NaO}_4]^+$: 441.1790, found: 441.1784.

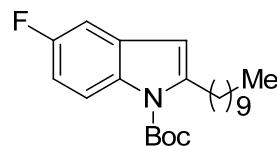
***tert*-Butyl 2-(5-methoxy-5-oxopentyl)-1*H*-indole-1-carboxylate (3k)**



3k

This compound was prepared in 82% yield according to the general procedure B (eluents: ethyl acetate: hexanes = 1:80). ^1H NMR (500 MHz, CDCl_3) δ 8.09 (d, 1H, J = 8.5 Hz), 7.17 – 7.23 (m, 2H), 6.36 (d, 1H, J = 0.5 Hz), 3.69 (s, 3H), 3.03 (t, 2H, J = 7.5 Hz), 2.40 (t, 2H, J = 7.5 Hz), 1.75 – 1.78 (m, 4H), 1.70 (s, 9H); ^{13}C NMR (125 MHz, CDCl_3) δ 173.9, 150.5, 141.7, 136.6, 129.3, 123.2, 122.6, 119.7, 115.5, 107.2, 83.7, 51.6, 34.0, 29.9, 28.4, 28.3, 24.7; IR (neat): 3050, 2977, 2925, 2851, 1732, 1455, 1369, 1328, 1161, 1115, 1086; HRMS (ES+) calculated for $[\text{C}_{19}\text{H}_{25}\text{NNaO}_4]^+$: 354.1681, found: 354.1684.

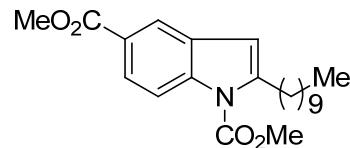
***tert*-Butyl 2-decyl-5-fluoro-1*H*-indole-1-carboxylate (3l)**



3l

This compound was prepared in 94% yield according to the general procedure B (eluent: ethyl acetate: hexanes = 1:80). ^1H NMR (400 MHz, CDCl_3) δ 8.04 (dd, 1H, $J_1 = 9.2$ Hz, $J_2 = 4.8$ Hz), 7.09 (dd, 1H, $J_1 = 8.8$ Hz, $J_2 = 2.8$ Hz), 6.93 (td, 1H, $J_1 = 8.8$ Hz, $J_2 = 2.4$ Hz), 6.30 (s, 1H), 2.97 (t, 2H, $J = 7.6$ Hz), 1.68 (s, 9H), 1.21 – 1.44 (m, 16H), 0.89 (t, 3H, $J = 7.2$ Hz); ^{13}C NMR (125 MHz, CDCl_3) δ 160.1, 158.2, 150.3, 144.2, 132.9, 130.2 (d, $J = 10.3$ Hz), 116.4 (d, $J = 9.0$ Hz), 110.6 (d, $J = 9.0$ Hz), 106.6 (d, $J = 3.4$ Hz), 105.0 (d, $J = 23.1$ Hz), 83.8, 32.0, 30.4, 29.7, 29.6, 29.5, 29.48, 29.41, 28.9, 28.3, 22.8, 14.2; IR (neat): 3050, 2977, 2925, 2851, 1732, 1460, 1369, 1327, 1158, 1115, 1086; HRMS (ES+) calculated for $[\text{C}_{23}\text{H}_{34}\text{NFO}_2]^+$: 375.2574, found: 375.2570.

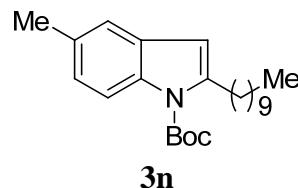
Dimethyl 2-decyl-1*H*-indole-1,5-dicarboxylate (3m)



3m

This compound was prepared in 80% yield according to the general procedure B (eluent: ethyl acetate: hexanes = 1:40). ^1H NMR (400 MHz, CDCl_3) δ 8.16 (d, 1H, $J = 1.6$ Hz), 8.09 (d, 1H, $J = 8.8$ Hz), 7.92 (dd, 1H, $J_1 = 8.8$ Hz, $J_2 = 1.6$ Hz), 7.42 (d, 1H, $J = 0.8$ Hz), 4.06 (s, 3H), 3.93 (s, 3H), 2.99 (t, 2H, $J = 7.6$ Hz), 1.67–1.71 (m, 2H), 1.27–1.42 (m, 14H), 0.88 (t, 3H, $J = 6.4$ Hz); ^{13}C NMR (100 MHz, CDCl_3) δ 167.5, 152.2, 144.0, 139.1, 129.3, 124.8, 124.7, 121.9, 115.2, 107.7, 53.7, 52.0, 31.9, 29.9, 29.6, 29.5, 29.3, 28.6, 22.7, 14.1; IR (neat): 3050, 2977, 2925, 2851, 1732, 1707, 1441, 1369, 1328, 1158, 1115, 1086; HRMS (ES+) calculated for $[\text{C}_{22}\text{H}_{31}\text{NNaO}_4]^+$: 396.2151, found: 396.2153.

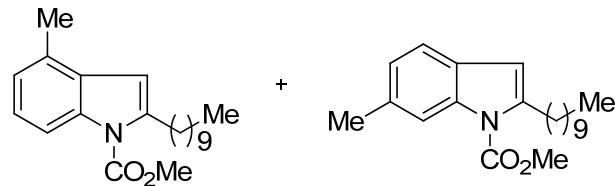
***tert*-Butyl 2-decyl-5-methyl-1*H*-indole-1-carboxylate (3n)**



This compound was prepared in 91% yield according to the general procedure B (eluents: ethyl acetate: hexanes = 1:100). ^1H NMR (400 MHz, CDCl_3) δ 7.96 (d, 1H, J = 8.8 Hz), 7.23-7.26 (m, 1H), 7.04 (d, 1H, J = 8.8 Hz), 6.27 (s, 1H), 2.98 (t, 2H, J = 7.6 Hz), 2.42 (s, 3H), 1.68-1.70 (m, 11H), 1.27-1.41 (m, 14H), 0.89 (t, 3H, J = 6.4 Hz); ^{13}C NMR (100 MHz, CDCl_3) δ 150.6, 142.6, 134.8, 131.9, 129.6, 124.3, 119.6, 115.2, 106.7, 83.4, 76.8, 43.9, 31.9, 30.3, 29.7, 29.6, 29.5, 29.48, 29.42, 29.0, 28.3, 22.8, 21.3, 14.2; IR (neat): 3050, 2977, 2925, 2851, 1732, 1460, 1369, 1327, 1158, 1116, 1086; HRMS (ES⁺) calculated for $[\text{C}_{24}\text{H}_{37}\text{NNaO}_2]^+$: 394.2722, found: 394.2713.

2-Decyl-4-methyl-indole-1-carboxylic acid methyl ester (3o) and

2-Decyl-6-methyl-indole-1-carboxylic acid methyl ester (3o')

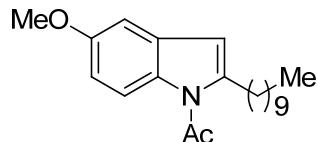


$$\mathbf{3o}:\mathbf{3o}' = 1.25:1$$

These two compounds were prepared as a pair of inseparable mixtures in 78% yield according to the general procedure B (eluents: ethyl acetate: hexanes = 1:100). The position of the methyl group is determined by ^1H NMR; ^1H NMR (400 MHz, CDCl_3) δ 7.90-7.92 (m, 1H), 7.32 (d, 0.44 H, J = 8.4 Hz), 7.12-7.16 (m, 0.63H), 7.00-7.12 (m, 1H), 6.40 (d, 0.54 H, J = 0.4 Hz), 6.40 (d, 0.43 H, J = 0.8 Hz), 4.04 (s, 3H), 2.97-3.02 (m, 2H), 2.47-2.49 (m, 3H), 1.68-1.71 (m, 2H), 12.8-14.6 (m, 16H), 0.87.-0.90 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 152.6, 142.0, 141.8, 136.8, 136.1, 133.2, 129.0, 128.9, 127.2, 124.2, 123.4, 123.3, 119.3, 115.8, 113.1, 107.4, 105.9, 53.34, 53.31, 43.8, 31.9, 29.99, 29.91, 29.84, 29.62, 29.53, 29.47, 29.39, 29.35, 29.30, 29.17, 28.89, 28.79, 23.84, 22.69, 21.

98, 18.39, 14.117; IR (neat): 3050, 2977, 2925, 2851, 1732, 1460, 1369, 1328, 1158, 1115, 1086; HRMS (ES+) calculated for $[C_{21}H_{31}NO_2]^+$: 329.2355, found: 329.2362.

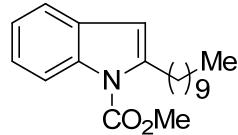
1-(2-Decyl-5-methoxy-1*H*-indol-1-yl)ethanone (3p)



3p

This compound was prepared in 80% yield according to the general procedure B (eluent: ethyl acetate: hexanes = 1:100). 1H NMR (400 MHz, $CDCl_3$) δ 7.64 (d, 1H, J = 8.8 Hz), 6.95 (d, 1H, J = 2.8 Hz), 6.84 (dd, 1H, J_1 = 9.2 Hz, J_2 = 2.4 Hz), 6.34 (s, 1H), 3.85 (s, 3H), 2.97 (t, 2H, J = 7.6 Hz), 2.72 (s, 3H), 1.66 – 1.74 (m, 2H), 1.20 – 1.43 (m, 14H), 0.88 (t, 3H, J = 6.4 Hz); ^{13}C NMR (100 MHz, $CDCl_3$) δ 169.9, 155.9, 143.7, 131.04, 130.96, 115.7, 150.5, 11.5, 108.2, 102.9, 5.6, 31.9, 30.6, 29.6, 29.5, 29.3, 28.8, 27.5, 22.7, 14.2; IR (neat): 2995, 2925, 2853, 1704, 1614, 1478, 1371, 1308, 1206; HRMS (ES+) calculated for $[C_{21}H_{31}NNaO_2]^+$: 352.2252, found: 352.2241.

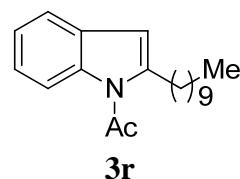
Methyl 2-decyl-1*H*-indole-1-carboxylate (3q)



3q

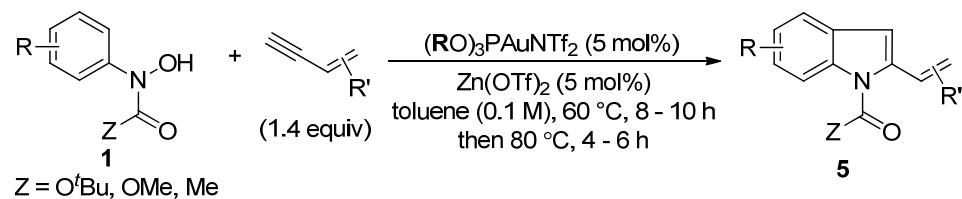
This compound was prepared in 81% yield according to the general procedure B (2 equiv of 1-dodecyne was used; reaction solvent was DCE; eluent: ethyl acetate: hexanes = 1:75). 1H NMR (400 MHz, $CDCl_3$) δ 8.08 (dd, 1H, J_1 = 8 Hz, J_2 = 0.8 Hz), 7.46 (dd, 1H, J_1 = 8 Hz, J_2 = 0.8 Hz), 7.19 – 7.27 (m, 2H), 6.38 (s, 1H), 4.05 (s, 3H), 3.00 (t, 2H, J = 7.6 Hz), 1.66 – 1.74 (m, 2H), 1.28 – 1.46 (m, 14H), 0.89 (t, 3H, J = 6.4 Hz); ^{13}C NMR (100 MHz, $CDCl_3$) δ 152.6, 142.6, 136.4, 129.5, 123.3, 122.9, 119.7, 115.5, 53.4, 31.9, 29.9, 29.6, 29.5, 29.3, 28.7, 22.7, 14.1; HRMS (ES+) calculated for $[C_{20}H_{29}NNaO_2]^+$: 338.2096, found: 338.2102.

1-(2-decyl-1*H*-indol-1-yl)ethanone (3r)



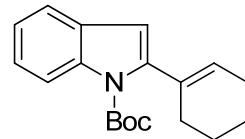
This compound was prepared in 90% yield according to the general procedure B (eluents: ethyl acetate: hexanes = 1:40). ^1H NMR (400 MHz, CDCl_3) δ 7.84 (d, 1H, J = 8.0 Hz), 7.47-7.50 (m, 1H), 7.21-7.26 (m, 2H), 6.41 (d, 1H, J = 0.8 Hz), 2.99 (t, 2H, J = 7.2 Hz), 2.76 (s, 3H), 1.68-1.74 (m, 2H), 1.20-1.43 (m, 14H), 0.89 (t, 3H, J = 6.8 Hz); ^{13}C NMR (100 MHz, CDCl_3) δ 170.3, 143.0, 136.3, 129.9, 123.4, 122.9, 120.1, 114.7, 108.1, 31.9, 30.5, 29.6, 29.5, 29.3, 28.9, 27.7, 22.7, 14.1; IR (neat): 3050, 2954, 2925, 2851, 1707, 1460, 1369, 1301, 1208; HRMS (ES+) calculated for $[\text{C}_{20}\text{H}_{29}\text{NNaO}]$ 322.2147, found: 322.2153.

General procedure C: preparation of *N*-protected-2-alkenyl indoles 5



To a solution of *N*-protected hydroxylamine **2** (0.3 mmol, 1 equiv) in dry toluene (3 mL, 0.1 M) was added the corresponding 1,3-ynye (0.42 mmol, 1.4 equiv), Zn(OTf)₂ (0.015 mmol, 5 mol%) and (ArO)₃PAuNTf₂ (Ar = 2,4-di-*tert*-butylphenyl, 0.015 mmol, 5 mol%) at room temperature. The reaction mixture was stirred at 60 °C for 8 – 10 h. Once TLC indicated there was no starting material left, the reaction was raised up to 80 °C, stirred for additional 4 - 8 h, and then concentrated. The residue was purified through silica gel flash chromatography to give the desired product **5**.

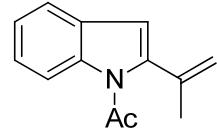
***tert*-Butyl 2-cyclohexenyl-1*H*-indole-1-carboxylate (**5a**)**



5a

This compound was prepared in 83 % yield according to the general procedure C (eluents: ethyl acetate: hexanes = 1:50). ^1H NMR (500 MHz, CDCl_3) δ 8.09 (dd, 1H, J_1 = 8.5 Hz, J_2 = 1 Hz), 7.50 (d, 1H, J = 7.5 Hz), 7.26 – 7.29 (m, 1H), 7.21 (td, 1H, J_1 = 7.5 Hz, J_2 = 1 Hz), 6.39 (s, 1H), 5.89 – 5.92 (m, 1H), 2.27 – 2.30 (m, 2H), 2.21 – 2.25 (m, 2H), 1.78 – 1.83 (m, 2H), 1.70 – 1.75 (m, 2H), 1.67 (s, 9H); ^{13}C NMR (125 MHz, CDCl_3) δ 150.3, 143.7, 136.9, 132.5, 129.3, 126.5, 123.6, 122.6, 120.1, 115.1, 107.8, 83.4, 29.1, 28.2, 25.4, 22.7, 21.9; IR (neat): 3051, 2979, 2932, 2859, 2834, 1731, 1453, 1369, 1326, 1161, 1126; HRMS (ES+) calculated for $[\text{C}_{19}\text{H}_{23}\text{NNaO}_2]^+$: 320.1626, found: 320.1627.

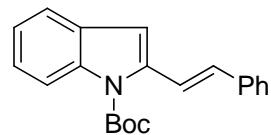
1-(2-(Prop-1-en-2-yl)-1*H*-indol-1-yl)ethanone (5b**)**



5b

This compound was prepared in 68% yield according to the general procedure C (2 equiv enyne was used; eluents: ethyl acetate: hexanes = 1:30). ^1H NMR (500 MHz, CDCl_3) δ 8.19 (ddd, 1H, J_1 = 8.5 Hz, J_2 = 2 Hz, J_3 = 1 Hz), 7.52 (ddd, 1H, J_1 = 8 Hz, J_2 = 2 Hz, J_3 = 1 Hz,), 7.31 – 7.34 (m, 1H), 7.24 – 7.27 (m, 1H), 6.56 (d, 1H, J = 1 Hz), 5.31 – 5.32 (m, 1H), 5.26 (s, 1H), 2.62 (s, 3H), 2.15 (dd, 3H, J_1 = 1.5 Hz, J_2 = 1 Hz); ^{13}C NMR (125 MHz, CDCl_3) δ 171.0, 141.6, 138.0, 137.3, 128.9, 124.8, 123.4, 120.4, 117.3, 115.3, 109.8, 26.8, 23.3; IR (neat): 3084, 3050, 2954, 2923, 2854, 1706, 1451, 1368, 1299, 1187; HRMS (ES+) calculated for $[\text{C}_{13}\text{H}_{13}\text{NNaO}]^+$: 222.0895, found: 222.0887.

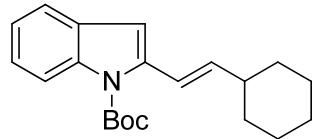
(E)-*tert*-Butyl 2-styryl-1*H*-indole-1-carboxylate (5c)



5c

This compound was prepared in 74% yield according to the general procedure C (eluent: ethyl acetate: hexanes = 1: 50). ^1H NMR (500 MHz, CDCl_3) δ 8.15 (d, 1H, J = 8.5 Hz), 7.78 (d, 1H, J = 16.5 Hz), 7.56 (d, 3H, J = 7.5 Hz), 7.39 (t, 2H, J = 7.5 Hz), 7.28 – 7.32 (m, 2H), 7.24 – 7.27 (m, 1H), 7.08 (d, 1H, J = 16.5 Hz), 6.88 (s, 1H), 1.73 (s, 9H); ^{13}C NMR (125 MHz, CDCl_3) δ 150.5, 139.5, 137.1, 136.8, 130.5, 129.3, 128.6, 127.7, 126.6, 124.1, 122.9, 120.7, 120.2, 115.7, 106.7, 84.1, 28.4; IR (neat): 3058, 3023, 2976, 2931, 1730, 1450, 1370, 1327, 1157; HRMS (ES+) calculated for $[\text{C}_{21}\text{H}_{21}\text{NNaO}_2]^+$: 342.1470, found: 342.1468.

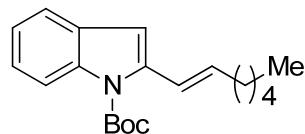
(E)-*tert*-Butyl 2-(2-cyclohexylvinyl)-1*H*-indole-1-carboxylate (5d)



5d

This compound was prepared in 90% yield according to the general procedure C (pent-2-en-4-yn-1-ylcyclohexane ($E/Z > 30: 1$) was used; eluent: ethyl acetate: hexanes = 1:50). ^1H NMR (500 MHz, CDCl_3) δ 8.11 (d, 1H, J = 8.5 Hz), 8.48 (d, 1H, J = 8 Hz), 7.25 (td, 1H, J_1 = 7.5 Hz, J_2 = 1.5 Hz), 7.20 (td, 1H, J_1 = 7.5 Hz, J_2 = 1.0 Hz), 6.91 (d, 1H, J = 16 Hz), 6.64 (s, 1H), 6.16 (dd, 1H, J_1 = 15.5 Hz, J_2 = 2 Hz), 2.18 – 2.21 (m, 1H), 1.87 (dd, 1H, J_1 = 14 Hz, J_2 = 2 Hz), 1.80 (dt, 2H, J_1 = 13 Hz, J_2 = 3.5 Hz), 1.71 (s, 9H), 1.31 – 1.39 (m, 3H), 1.19 – 1.27 (m, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 150.5, 140.0, 139.1, 136.5, 129.4, 123.5, 122.7, 119.87, 119.81, 115.5, 105.8, 83.8, 41.2, 32.8, 28.4, 26.2, 26.1; IR (neat): 2927, 2853, 1731, 1451, 1370, 1328, 1159; HRMS (ES+) calculated for $[\text{C}_{21}\text{H}_{27}\text{NO}_2]^+$: 325.2042, found: 325.2040.

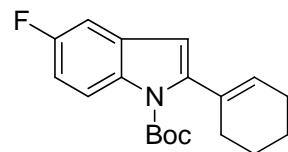
(E)-*tert*-Butyl 2-(hept-1-enyl)-1*H*-indole-1-carboxylate (5e)



5e

This compound was prepared in 86% yield (*E/Z* > 15: 1) according to the general procedure C (non-3-en-1-yne (*E/Z* = 9: 1) was used; eluents: ethyl acetate: hexanes = 1:50). ^1H NMR (500 MHz, CDCl_3) δ 8.11 (d, 1H, J = 8 Hz), 7.49 (d, 1H, J = 8 Hz), 7.19 – 7.27 (m, 2H), 6.64 (s, 1H), 6.20 (dt, 1H, J_1 = 16 Hz, J_2 = 7 Hz), 2.24 – 2.29 (m, 2H), 1.71 (s, 9H), 1.51 – 1.57 (m, 2H), 1.36 – 1.40 (m, 4H), 0.94 (t, 3H, J = 7 Hz); ^{13}C NMR (125 MHz, CDCl_3) δ 150.5, 139.8, 136.4, 133.7, 129.4, 123.5, 122.7, 122.1, 119.9, 115.5, 105.9, 83.8, 33.1, 31.5, 28.9, 28.3, 22.6, 14.1; IR (neat): 3051, 2957, 2856, 1731, 1451, 1327, 1159, 1116, 962; HRMS (ES+) calculated for $[\text{C}_{20}\text{H}_{27}\text{NO}_2]^+$: 313.2042, found: 325.2041.

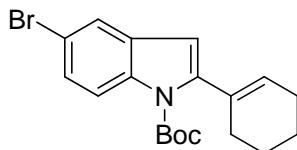
***tert*-Butyl 2-cyclohexenyl-5-fluoro-1*H*-indole-1-carboxylate (5f)**



5f

This compound was prepared in 75% yield according to the general procedure C (eluents: ethyl acetate: hexanes = 1:50). ^1H NMR (400 MHz, CDCl_3) δ 8.00 (dd, 1H, J_1 = 8.8 Hz, J_2 = 4.4 Hz), 7.12 (dd, 1H, J_1 = 8.8 Hz, J_2 = 2.8 Hz), 6.97 (td, 1H, J_1 = 8.8 Hz, J_2 = 2.8 Hz), 6.31 (s, 1H), 5.87 – 5.89 (m, 1H), 2.18 – 2.27 (m, 4H), 1.75 – 1.80 (m, 2H), 1.68 – 1.73 (m, 2H), 1.64 (s, 9H); ^{13}C NMR (100 MHz, CDCl_3) δ 159.2 (d, J = 236 Hz), 150.1, 145.3, 133.2, 132.3, 130.1 (d, J = 11 Hz), 127.0, 116.1 (d, J = 11 Hz), 111.2 (d, J = 24 Hz), 107.5, 105.5 (d, J = 24 Hz), 83.7, 29.1, 28.1, 25.3, 22.6, 21.9; IR (neat): 2979, 2970, 2933, 2860, 2834, 1732, 1471, 1370, 1314, 1115; HRMS (ES+) calculated for $[\text{C}_{19}\text{H}_{22}\text{NNaO}_2]^+$: 338.1532, found: 338.1523.

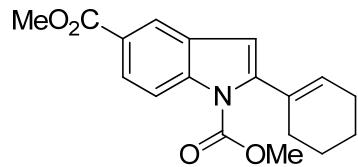
***tert*-Butyl 5-bromo-2-cyclohexenyl-1*H*-indole-1-carboxylate (5g)**



5g

This compound was prepared in 65% yield according to the general procedure C (eluents: ethyl acetate: hexanes = 1:50). ^1H NMR (500 MHz, CDCl_3) δ 7.93 (d, 1H, J = 9 Hz), 7.60 (d, 1H, J = 2 Hz), 7.33 (dd, 1H, J_1 = 9 Hz, J_2 = 2 Hz), 6.29 (s, 1H), 5.87 – 5.89 (m, 1H), 2.23 – 2.26 (m, 2H), 2.18 – 2.22 (m, 2H), 1.75 – 1.79 (m, 2H), 1.68 – 1.72 (m, 2H), 1.64 (s, 9H); ^{13}C NMR (125 MHz, CDCl_3) δ 149.9, 144.8, 135.6, 132.0, 131.0, 127.2, 126.3, 122.6, 116.6, 115.8, 106.9, 83.9, 29.1, 28.2, 25.4, 22.7, 21.9; IR (neat): 3111, 2965, 2932, 2859, 2834, 1733, 1446, 1348, 1155, 1013; HRMS (ES+) calculated for $[\text{C}_{19}\text{H}_{22}\text{BrNNaO}_2]^+$: 398.0732, found: 398.0738.

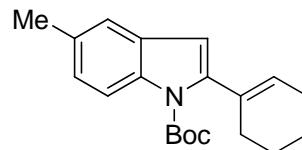
Dimethyl 2-cyclohexenyl-1*H*-indole-1,5-dicarboxylate (5h)



5h

This compound was prepared in 77% yield according to the general procedure C (reaction solvent: DCE; eluents: ethyl acetate: hexanes = 1:30). ^1H NMR (400 MHz, CDCl_3) δ 8.20 (d, 1H, J = 1.6 Hz), 8.10 (d, 1H, J = 8.8 Hz), 7.96 (dd, 1H, J_1 = 8.8 Hz, J_2 = 2 Hz), 6.43 (s, 1H), 5.87 – 5.89 (m, 1H), 4.02 (s, 3H), 3.93 (s, 3H), 2.19 – 2.22 (m, 4H), 1.68 – 1.77 (m, 4H); ^{13}C NMR (100 MHz, CDCl_3) δ 167.5, 151.8, 144.8, 139.2, 132.1, 129.3, 127.9, 125.2, 124.9, 122.4, 115.1, 108.5, 53.8, 52.0, 29.3, 25.4, 22.7, 21.9; IR (neat): 3022, 2933, 2856, 1741, 1722, 1444, 1338, 1210; HRMS (ES+) calculated for $[\text{C}_{18}\text{H}_{19}\text{NNaO}_4]^+$: 336.1212, found: 336.1205.

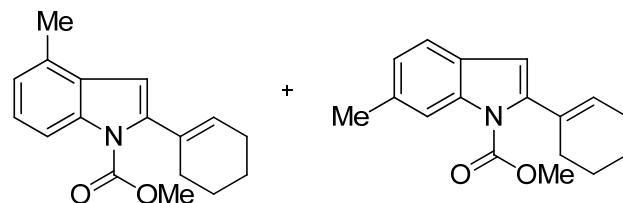
***tert*-Butyl 2-cyclohexenyl-5-methyl-1*H*-indole-1-carboxylate (**5i**)**



5i

This compound was prepared in 80% yield according to the general procedure C (eluents: ethyl acetate: hexanes = 1:50). ^1H NMR (400 MHz, CDCl_3) δ 7.93 (d, 1H, J = 8.4 Hz), 7.27 (s, 1H), 7.07 (d, 1H, J = 8.8 Hz), 6.29 (s, 1H), 5.87 – 5.89 (m, 1H), 2.42 (s, 3H), 2.23 – 2.26 (m, 2H), 2.19 – 2.21 (m, 2H), 1.75 – 1.81 (m, 2H), 1.67 – 1.73 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 150.3, 143.8, 135.1, 132.6, 132.0, 129.5, 126.3, 124.9, 120.1, 114.8, 107.6, 83.2, 29.1, 28.2, 25.3, 22.7, 21.9, 21.2; IR (neat): 2978, 2932, 2857, 2834, 1731, 1474, 1352, 1134; HRMS (ES $+$) calculated for $[\text{C}_{20}\text{H}_{25}\text{NNaO}_2]^+$: 334.1783, found: 334.1779.

Methyl 2-cyclohexenyl-4-methyl-1*H*-indole-1-carboxylate (5j**) and methyl 2-cyclohexenyl-6-methyl-1*H*-indole-1-carboxylate (**5j'**)**

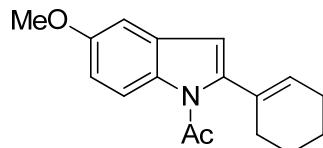


5j:5j' = (1.5 : 1)

These two compounds were prepared as a pair of inseparable mixtures in 71% yield according to the general procedure C (reaction solvent: DCE; eluents: ethyl acetate: hexanes = 1:40). ^1H NMR (400 MHz, CDCl_3) δ 7.92 – 7.94 (m, 0.75H), 7.37 (d, 0.39H, J = 8 Hz), 7.19 (t, 0.48H, J = 8 Hz), 7.03 – 7.07 (m, 0.83H), 6.45 (d, 0.47H, J = 0.8 Hz), 6.34 (d, 0.34H, J = 0.8 Hz), 5.86 – 5.91 (m, 0.86H), 4.01 (s, 2.70H), 2.51 (s, 1.5H), 2.50 (s, 1.2H), 2.21 – 2.25 (m, 4H), 1.60 – 1.79 (m, 4H); ^{13}C NMR (100 MHz, CDCl_3) δ 152.2, 152.1, 143.04, 142.95, 136.9, 136.2, 133.8, 132.7, 129.6, 129.0, 127.2, 127.0, 126.9, 124.3, 123.9, 123.4, 119.8, 115.6, 112.9, 108.2, 106.9, 53.50, 53.47, 29.42, 29.39, 25.4,

22.8, 22.1, 22.0, 18.4; IR (neat): 3025, 2931, 2856, 2834, 1734, 1441, 1327, 1129; HRMS (ES⁺) calculated for [C₁₇H₁₉NNaO₂]⁺: 292.1313, found: 292.1293.

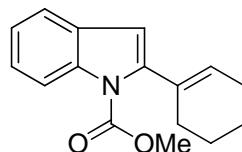
1-(2-(Cyclohex-1-en-1-yl)-5-methoxy-1*H*-indol-1-yl)ethanone (5k)



5k

This compound was prepared in 75% yield according to the general procedure C (eluent: ethyl acetate: hexanes = 1:60). ¹H NMR (400 MHz, CDCl₃) δ 8.16 (d, 1H, J = 8.8 Hz), 6.95 (d, 1H, J = 2.8 Hz), 6.89 (dd, 1H, J₁ = 9.2 Hz, J₂ = 2.8 Hz), 6.36 (s, 1H), 5.94 – 5.97 (m, 1H), 3.84 (s, 3H), 2.55 (s, 3H), 2.26 – 2.30 (m, 2H), 2.20 – 2.25 (m, 2H), 1.75 – 1.81 (m, 2H), 1.68 – 1.73 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 170.9, 156.3, 143.1, 132.0, 130.0, 129.9, 116.7, 112.7, 109.1, 102.9, 55.7, 30.1, 26.1, 25.6, 22.7, 21.7; IR (neat): 2996, 2927, 2856, 2832, 1697, 1609, 1472, 1368, 1305, 1212, 1141; HRMS (ES⁺) calculated for [C₁₇H₁₉NNaO₂]⁺: 292.1313, found: 292.1300.

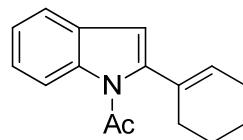
Methyl 2-cyclohexenyl-1*H*-indole-1-carboxylate (5l)



5l

This compound was prepared in 86% yield according to the general procedure C (reaction solvent: DCE, eluent: ethyl acetate: hexanes = 1:50). ¹H NMR (500 MHz, CDCl₃) δ 8.11 (d, 1H, J = 8.5 Hz), 7.50 (dt, 1H, J₁ = 7.5 Hz, J₂ = 1 Hz), 7.22 – 7.31 (m, 2H), 6.40 (s, 1H), 5.88 – 5.90 (m, 1H), 4.09 (s, 3H), 2.22 – 2.27 (m, 4H), 1.72 – 1.75 (m, 2H), 1.77 – 1.81 (m, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 125, 152.0, 143.5, 136.4, 132.5, 129.5, 127.1, 123.8, 122.9, 120.2, 115.4, 108.3, 53.5, 29.4, 25.5, 22.8, 22.1; IR (neat): 3027, 2932, 2856, 2834, 1744, 1455, 1324, 1213; HRMS (ES⁺) calculated for [C₁₆H₁₇NNaO₂]⁺: 278.1157, found: 278.1146.

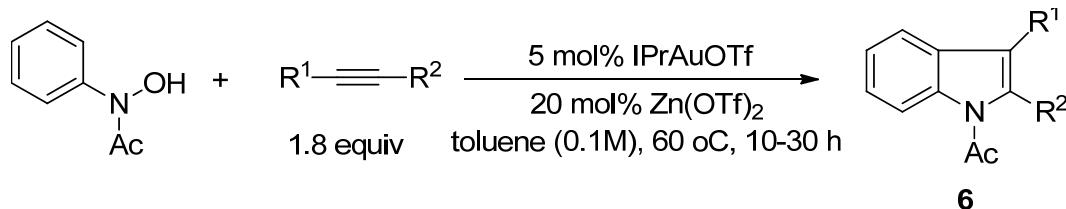
1-(2-Cyclohexenyl-1*H*-indol-1-yl)ethanone (5m**)**



5m

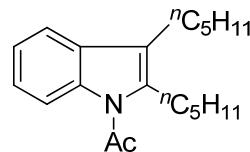
This compound was prepared in 92% yield according to the general procedure C (eluent: ethyl acetate: hexanes = 1:30). ^1H NMR (500 MHz, CDCl_3) δ 8.26 (d, 1H, J = 8.5 Hz), 7.50 (d, 1H, J = 7.5 Hz), 7.31 (td, 1H, J_1 = 7 Hz, J_2 = 1.5 Hz), 7.25 (td, 1H, J_1 = 7.5 Hz, J_2 = 1.5 Hz), 6.46 (s, 1H), 5.98 – 6.00 (m, 1H), 2.30 – 2.33 (m, 2H), 2.23 – 2.27 (m, 2H), 1.78 – 1.83 (m, 2H), 1.71 – 1.75 (m, 2H); ^{13}C NMR (125 MHz, CDCl_3) δ 171.2, 142.3, 137.1, 131.8, 129.8, 129.0, 124.4, 123.3, 120.0, 115.6, 109.0, 30.0, 26.4, 25.6, 22.7, 21.8; IR (neat): 3049, 2930, 2857, 2834, 1701, 1450, 1367, 1303; HRMS (ES+) calculated for $[\text{C}_{16}\text{H}_{17}\text{NNaO}]^+$: 262.1208, found: 262.1195.

General procedure D: preparation of *N*-acetyl-2,3-disubstituted indoles **6**



To a solution of **1c** (0.3 mmol, 1 equiv) in dry toluene (3 mL, 0.1 M) was added corresponding internal alkyne (0.54 mmol, 1.8 equiv), $\text{Zn}(\text{OTf})_2$ (0.06 mmol, 20 mol%) and IPrAuOTf (0.015 mmol, 5 mol%) at room temperature. The reaction mixture was stirred at 60 °C for 10 - 30 h. Once TLC indicated there was no starting material left, the reaction was concentrated. The residue was purified through silica gel flash chromatography to give the desired product **6**.

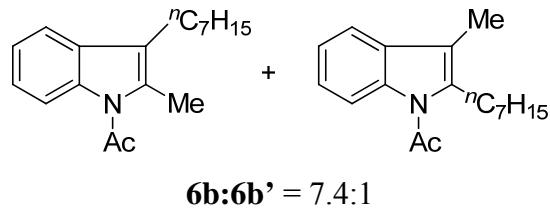
1-(2,3-dipentyl-1*H*-indol-1-yl)ethanone (6a**)**



6a

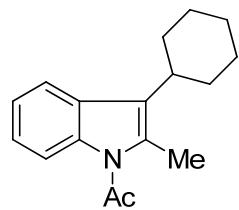
This compound was prepared in 74% yield according to the general procedure D. ^1H NMR (500 MHz, CDCl_3) δ 7.80 – 7.72 (m, 1H), 7.52 – 7.45 (m, 1H), 7.29 – 7.20 (m, 2H), 2.99 (t, J = 8.0 Hz, 2H), 2.77 (s, 3H), 2.65 (t, J = 8.0 Hz, 2H), 1.66 – 1.54 (m, 4H), 1.44 – 1.31 (m, 8H), 0.98 – 0.85 (m, 6H); ^{13}C NMR (125 MHz, CDCl_3) δ 170.00, 138.26, 135.60, 130.87, 123.27, 122.53, 120.10, 118.66, 114.45, 31.97, 31.91, 30.03, 29.91, 27.67, 27.01, 23.93, 22.56, 22.48, 14.04, 14.02; IR (neat): 2957, 2930, 2859, 1705, 1463, 1368, 1216; HRMS (ES+) calculated for $[\text{C}_{20}\text{H}_{29}\text{NO}]^+$: 299.2249, found: 299.2257.

1-(3-heptyl-2-methyl-1*H*-indol-1-yl)ethanone (**6b**)



Compound **6b** was prepared in 61% yield^[6] (regioselectivity 7.4: 1) according to the general procedure F and spectral data were in accordance with literature data^[7]. ^1H NMR (500 MHz, CDCl_3) δ 8.00 – 7.93 (m, 1H), 7.49 – 7.44 (m, 1H), 7.28 – 7.21 (m, 2H), 2.73 (s, 3H), 2.64 (t, J = 7.5 Hz, 2H), 2.56 (s, 3H), 1.63 – 1.53 (m, 2H), 1.41 – 1.23 (m, 8H), 0.89 (dd, J = 7.1, 6.5 Hz, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 170.19, 135.70, 132.49, 130.64, 123.50, 122.74, 120.38, 118.24, 115.00, 31.84, 29.94, 29.56, 29.20, 29.15, 27.55, 23.87, 22.64, 14.38, 14.07; IR (neat): 2956, 2929, 2856, 1701, 1458, 1368, 1311, 1216745; HRMS (ES+) calculated for $[\text{C}_{18}\text{H}_{25}\text{NO}]^+$: 271.1926, found: 271.1948.

1-(3-Cyclohexyl-2-methyl-1*H*-indol-1-yl)ethanone (**6c**)

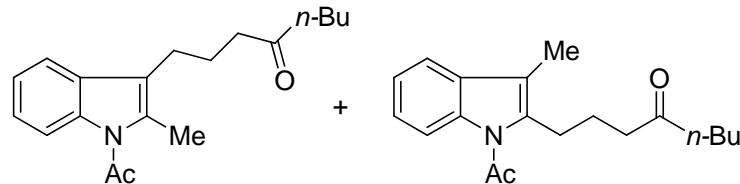


6c

This compound was prepared in 67% yield^[6] (regioselectivity > 19: 1) according to the general procedure D. ^1H NMR (500 MHz, CDCl_3) δ 7.99 – 7.93 (m, 1H), 7.75 – 7.69 (m, 1H), 7.26 – 7.19 (m, 2H), 2.80 (tt, J = 12.4, 3.7 Hz, 1H), 2.73 (s, 3H), 2.58 (s, 3H), 2.02 –

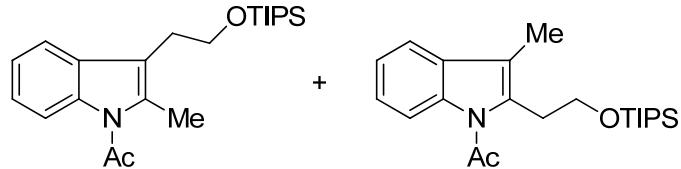
1.86 (m, 4H), 1.86 – 1.79 (m, 1H), 1.79 – 1.71 (m, 2H), 1.49 – 1.31 (m, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 170.40, 135.99, 131.49, 129.44, 124.37, 123.11, 122.27, 119.95, 114.90, 36.45, 31.80, 27.74, 27.12, 26.22, 14.42.; IR (neat): 2928, 2853, 1699, 1449, 1375, 1314, 1212, 1034, 756; HRMS (ES+) calculated for $[\text{C}_{17}\text{H}_{21}\text{NO}]^+$: 255.1623, found: 255.1628.

1-(1-Acetyl-2-methyl-1*H*-indol-3-yl)octan-4-one (**6d**)



Compound **6d** was prepared in 68% yield^[6] (regioselectivity 6.8:1) according to the general procedure D. **6d:** ^1H NMR (500 MHz, CDCl_3) δ 7.98 – 7.90 (m, 1H), 7.51 – 7.44 (m, 1H), 7.27 – 7.20 (m, 2H), 2.71 (s, 3H), 2.66 (t, J = 7.5 Hz, 2H), 2.55 (s, 3H), 2.44 (t, J = 7.0 Hz, 2H), 2.35 (t, J = 7.5 Hz, 2H), 1.90 – 1.81 (m, 2H), 1.57 – 1.48 (m, 3H), 1.32 – 1.23 (m, 2H), 0.91 – 0.86 (t, J = 7.5 Hz, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 210.87, 170.16, 135.61, 132.91, 130.40, 123.62, 122.84, 119.29, 118.20, 114.96, 42.51, 41.75, 27.51, 25.83, 23.60, 23.01, 22.27, 14.34, 13.79; IR (neat): 2958, 2933, 2873, 1707, 1458, 1369, 1311, 748; HRMS (ES+) calculated for $[\text{C}_{19}\text{H}_{25}\text{NNaO}_2]^+$: 322.1783, found: 322.1781.

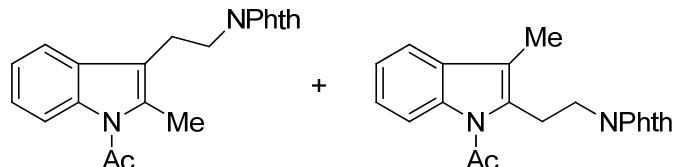
1-(2-Methyl-3-((triisopropylsilyl)oxy)ethyl)-1*H*-indol-1-yl)ethanone (**6e**)



Compound **6e** was prepared in 52% yield^[6] (regioselectivity 6:1) according to the general procedure D. **6e:** ^1H NMR (500 MHz, CDCl_3) δ 8.00 – 7.94 (m, 1H), 7.52 – 7.46 (m, 1H), 7.29 – 7.22 (m, 2H), 3.85 (t, J = 7.5 Hz, 2H), 2.95 (t, J = 7.3 Hz, 2H), 2.73 (s, 3H), 2.60 (s, 3H), 1.10 – 1.00 (m, 21H); ^{13}C NMR (125 MHz, CDCl_3) δ 170.22, 135.68,

133.78, 130.65, 123.62, 122.85, 118.17, 116.76, 115.01, 62.91, 27.94, 27.54, 17.98, 17.93, 14.53, 11.92; IR (neat): 2944, 2867, 1704, 1463, 1367, 1310, 1102, 742; HRMS (ES+) calculated for $[C_{22}H_{35}NNaO_2Si]^{+}$: 396.2349, found: 396.2342.

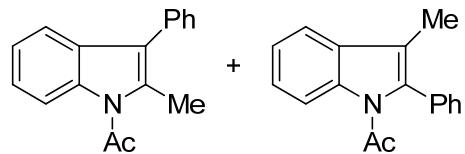
2-(2-(1-Acetyl-2-methyl-1*H*-indol-3-yl)ethyl)isoindoline-1,3-dione (6f**)**



6f:6f' = 16.6:1

Compound **6f** was prepared in 80% yield^[6] (regioselectivity 16.6:1) according to the general procedure D. **6f:** 1H NMR (600 MHz, $CDCl_3$) δ 7.96 – 7.89 (m, 1H), 7.82 (m, 2H), 7.73 – 7.67 (m, 2H), 7.66 – 7.60 (m, 1H), 7.27 – 7.21 (m, 2H), 3.85 (t, J = 7.8 Hz, 2H), 3.02 (t, J = 7.8 Hz, 2H), 2.71 (s, 3H), 2.59 (s, 3H); ^{13}C NMR (150 MHz, $CDCl_3$) δ 170.20, 168.15, 135.61, 134.03, 133.94, 132.04, 130.06, 123.87, 123.19, 123.11, 118.10, 115.90, 115.00, 37.29, 27.57, 23.19, 14.30; IR (neat): 3061, 2945, 1772, 1713, 1436, 1368, 1311, 1011, 722; HRMS (ES+) calculated for $[C_{21}H_{18}N_2O_3]^{+}$: 346.1317, found: 346.1321.

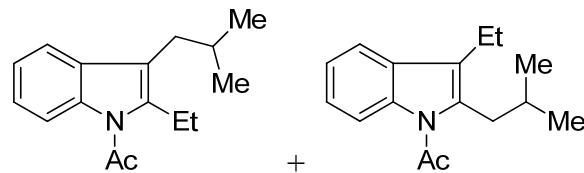
1-(2-Methyl-3-phenyl-1*H*-indol-1-yl)ethanone (6g**)**



6g:6g' = 8:1

Compound **6g** was prepared in 61% yield^[6] (regioselectivity 8:1) according to the general procedure D. **6g:** 1H NMR (500 MHz, $CDCl_3$) δ 8.06 (d, J = 8.3 Hz, 1H), 7.51 (m, 3H), 7.45 (m, 2H), 7.44 – 7.38 (m, 1H), 7.35 – 7.30 (m, 1H), 7.28 – 7.22 (m, 1H), 2.80 (s, 3H), 2.63 (s, 3H); ^{13}C NMR (125 MHz, $CDCl_3$) δ 170.47, 135.70, 133.45, 133.08, 130.11, 129.95, 128.53, 127.17, 124.07, 123.21, 122.49, 119.13, 115.04, 27.58, 15.25; IR (neat): 3052, 2929, 1701, 1367, 1309, 747; HRMS (ES+) calculated for $[C_{17}H_{15}NO]^{+}$: 249.1154, found: 249.1155.

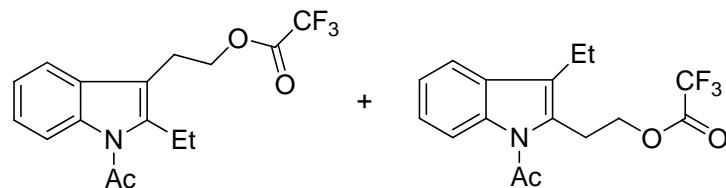
1-(2-Ethyl-3-isobutyl-1*H*-indol-1-yl)ethanone **6h + 1-(3-Ethyl-2-isobutyl-1*H*-indol-1-yl)ethanone (**6h'**)**



6h:6h' = 2.5:1

These compounds were prepared in 75 % overall yield (regioselectivity 2.5:1) according to the general procedure D. ¹H NMR (500 MHz, CDCl₃) δ 7.79 – 7.73 (m, 1H, 6m + 6m'), 7.55 – 7.51 (m, 1H, 6m'), 7.46 – 7.50 (m, 1H, 6m), 7.28 – 7.20 (m, 2H, 6m + 6m'), 3.05 (q, J = 7.3 Hz, 2H, 6m), 2.89 (d, J = 7.0 Hz, 2H, 6m'), 2.79 (s, 3H, 6m), 2.77 (s, 3H, 6m'), 2.70 (q, J = 7.5 Hz, 2H, 6m'), 2.54 (d, J = 7.4 Hz, 2H, 6m), 2.03 – 1.90 (m, 1H, 6m + 6m'), 1.26 – 1.20 (m, 3H, 6m + 6m'), 0.97 (d, J = 6.6 Hz, 6H, 6m), 0.93 (d, J = 6.7 Hz, 6H, 6m'); ¹³C NMR (125 MHz, CDCl₃) δ 170.20, 169.93, 140.20, 136.63, 135.82, 135.54, 131.29, 130.43, 123.35, 123.27, 122.51, 122.49, 122.39, 119.04, 118.75, 114.43, 114.34, 35.60, 33.07, 29.20, 28.95, 27.75, 27.68, 22.84, 22.44, 20.36, 17.40, 14.63, 14.47; IR (neat): 2958, 2933, 2870, 1705, 1463, 1367, 1312, 1204, 743; HRMS (ES+) calculated for [C₁₆H₂₁NO]⁺: 243.1623, found: 243.1631.

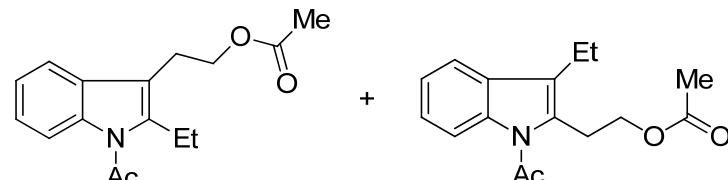
2-(1-Acetyl-2-ethyl-1*H*-indol-3-yl)ethyl 2,2,2-trifluoroacetate (6i**)**



6i:6i' = 10:1

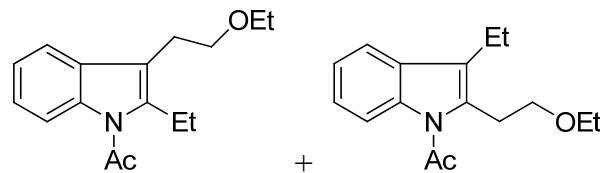
These compounds were prepared in 65% overall yield (regioselectivity 10:1) according to the general procedure D. **6i:** ¹H NMR (500 MHz, CDCl₃) δ 7.78 – 7.74 (m, 1H), 7.55 – 7.51 (m, 1H), 7.32 – 7.27 (m, 2H), 4.52 (t, J = 7.0 Hz, 2H), 3.15 (t, J = 7.0 Hz, 2H), 3.07 (q, J = 7.4 Hz, 2H), 2.80 (s, 3H), 1.25 (t, J = 7.4 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 169.91, 157.45, (q, J = 43.1 Hz) 141.68, 135.43, 129.96, 123.88, 122.95, 118.16, 114.51, 113.08, 67.04, 27.62, 23.00, 20.27, 14.82; IR (neat): 2968, 2930, 1785, 1707, 1464, 1352, 745; HRMS (ES+) calculated for [C₁₆H₁₆F₃NaNO₃]⁺: 350.0980, found: 350.0970.

2-(1-Acetyl-2-ethyl-1*H*-indol-3-yl)ethyl acetate (6j**)**



These compounds were prepared in 77 % overall yield (regioselectivity 4.6:1) according to the general procedure D. **6j:** ^1H NMR (500 MHz, CDCl_3) δ 7.77 – 7.71 (m, 1H), 7.56 – 7.51 (m, 1H), 7.29 – 7.24 (m, 3H), 4.25 (t, J = 7.0 Hz, 2H), 3.06 (q, J = 7.0 Hz, 2H), 3.01 (t, J = 7.0 Hz, 2H), 2.79 (s, 3H), 2.04 (s, 3H), 1.24 (t, J = 7.0 Hz, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 170.98, 169.92, 141.20, 135.45, 130.47, 123.65, 122.80, 118.51, 114.77, 114.45, 77.25, 77.00, 76.75, 63.74, 27.66, 23.49, 20.99, 20.30, 14.90; IR (neat): 2968, 2937, 1739, 1704, 1464, 1368, 1312, 1241, 1034, 745; HRMS (ES+) calculated for $[\text{C}_{16}\text{H}_{19}\text{NNaO}_3]^+$: 296.1263, found: 296.1257.

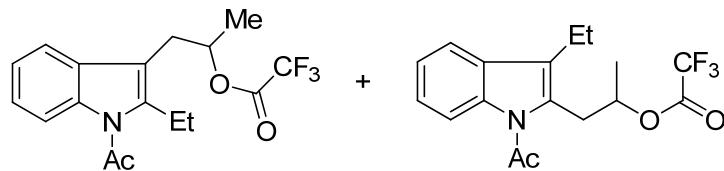
1-(3-(2-Ethoxyethyl)-2-ethyl-1*H*-indol-1-yl)ethanone (6k**) + 1-(2-(2-Ethoxyethyl)-3-ethyl-1*H*-indol-1-yl)ethanone (**6k'**)**



These compounds were prepared in 88 % overall yield (regioselectivity 2:1) according to the general procedure D. **6k:** ^1H NMR (600 MHz, CDCl_3) δ 7.77 – 7.72 (m, 1H), 7.54 – 7.49 (m, 1H), 7.28 – 7.21 (m, 2H), 3.58 (t, J = 7.6 Hz, 2H), 3.55 – 3.47 (q, J = 7.6 Hz, 2H), 3.05 (q, J = 7.3 Hz, 2H), 2.96 (t, J = 7.6 Hz, 2H), 2.77 (s, 3H), 1.23 (t, J = 7.6 Hz, 3H), 1.21 (t, J = 7.2 Hz, 3H); ^{13}C NMR (150 MHz, CDCl_3) δ 169.88, 140.79, 135.47, 130.70, 123.46, 122.64, 118.53, 115.75, 114.44, 70.11, 66.30, 27.60, 24.71, 20.29, 15.18, 14.91; **6k':** ^1H NMR (600 MHz, CDCl_3) δ 7.73 – 7.67 (m, 1H), 7.51 (m, 1H), 7.28 – 7.21 (m, 2H), 3.65 (t, J = 7.0 Hz, 2H), 3.49 (q, J = 7.0 Hz, 2H), 3.30 (t, J = 7.0 Hz, 2H), 2.77 (s, 3H), 2.71 (q, J = 7.6 Hz, 2H), 1.22 (t, J = 7.6 Hz, 3H), 1.18 (t, J = 7.0 Hz, 3H); ^{13}C NMR (151 MHz, CDCl_3) δ 170.13, 135.55, 134.15, 130.44, 123.55, 123.13, 122.60,

118.79, 114.36, 70.26, 66.21, 27.84, 27.83, 17.10, 15.23, 14.81. IR (neat): 2973, 1935, 2872, 1702, 1463, 1368, 1310, 1108, 943; HRMS (ES+) calculated for $[C_{16}H_{21}NO_2]^+$: 259.1572, found: 259.1574.

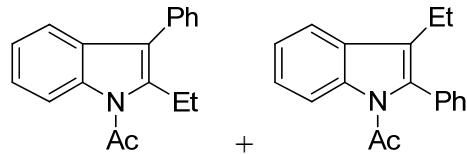
1-(1-Acetyl-2-ethyl-1*H*-indol-3-yl)propan-2-yl 2,2,2-trifluoroacetate (6l**)**



6l:6l' = 12:1

These compounds were prepared in 69 % overall yield (regioselectivity 12:1) according to the general procedure D. **6l:** 1H NMR (500 MHz, $CDCl_3$) δ 7.79 – 7.71 (m, 1H), 7.59 – 7.52 (m, 1H), 7.33 – 7.24 (m, 2H), 5.34 (h, J = 6.4 Hz, 1H), 3.14 (dd, J = 14.4, 6.6 Hz, 1H), 3.08 (ddd, J = 14.6, 7.3, 2.9 Hz, 2H), 2.99 – 2.91 (dd, J = 14.4, 6.6 Hz, 1H), 2.80 (s, 3H), 1.40 (t, J = 7.0 Hz, 3H), 1.26 – 1.21 (t, J = 7.0 Hz, 3H); ^{13}C NMR (125 MHz, $CDCl_3$) δ 169.95, 157.06 (q, J = 41.8 Hz), 141.73, 135.43, 130.24, 123.83, 122.92, 118.64, 114.47 (q, J = 41.8 Hz), 114.40, 113.52, 75.85, 30.00, 27.66, 20.32, 19.39, 14.59; IR (neat): 2985, 2943, 1783, 1717, 1383, 1338, 1221, 1167, 778; HRMS (ES+) calculated for $[C_{17}H_{18}F_3NNaO_3]^+$: 364.1136, found: 364.1121.

1-(2-Ethyl-3-phenyl-1*H*-indol-1-yl)ethanone (6m**) + 1-(3-Ethyl-2-phenyl-1*H*-indol-1-yl)ethanone (**6m'**)**

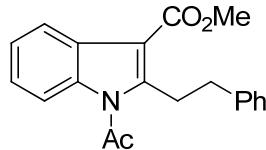


6m:6m' = 2.9:1

These compounds were prepared in 70 % overall yield (regioselectivity 2.9: 1) according to the general procedure D. **6m:** 1H NMR (600 MHz, $CDCl_3$) δ 7.83 (d, J = 8.4 Hz, 1H), 7.52 – 7.46 (m, 2H), 7.45 – 7.38 (m, 4H), 7.32 – 7.28 (m, 1H), 7.24 – 7.19 (m, 1H), 3.05 (q, J = 7.3 Hz, 2H), 2.85 (s, 3H), 1.23 (t, J = 7.3 Hz, 3H); ^{13}C NMR (150 MHz, $CDCl_3$) δ 170.27, 140.27, 135.50, 133.61, 130.53, 130.08, 128.56, 127.26, 123.89, 123.03, 121.98,

119.61, 114.45, 27.72, 20.76, 15.19; **6m'**: ^1H NMR (600 MHz, cdcl_3) δ 8.45 (d, $J = 8.3$ Hz, 1H), 7.58 (d, $J = 7.4$ Hz, 1H), 7.51 – 7.45 (m, 3H), 7.42 – 7.36 (m, 3H), 7.34 – 7.30 (m, 1H), 2.56 (q, $J = 7.6$ Hz, 2H), 1.95 (s, 3H), 1.21 – 1.17 (t, $J = 7.8$ Hz, 3H); ^{13}C NMR (150 MHz, CDCl_3) δ 171.07, 136.95, 134.39, 133.59, 130.22, 129.27, 128.69, 128.57, 125.18, 124.30, 123.38, 118.71, 116.52, 27.65, 17.60, 14.95. IR (neat): 3054, 2974, 2935, 2875, 1706, 1454, 1314, 1209, 1186, 747; HRMS (ES+) calculated for $[\text{C}_{18}\text{H}_{17}\text{NO}]^+$: 263.1310, found: 263.1313.

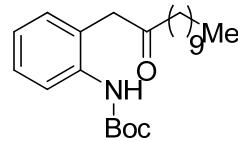
Methyl 1-acetyl-2-phenethyl-1*H*-indole-3-carboxylate (**6n**)



6n

This compound was prepared in 66 % yield^[6] (regioselectivity > 19: 1) according to the general procedure D. ^1H NMR (500 MHz, CDCl_3) δ 8.23 – 8.15 (m, 1H), 7.67 – 7.59 (m, 1H), 7.37 – 7.29 (m, 6H), 7.25 – 7.19 (m, 1H), 3.99 (s, 3H), 3.74 – 3.67 (t, 2H), 3.07 – 3.00 (t, 2H), 2.76 (s, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 170.87, 165.30, 149.20, 141.34, 134.65, 128.61, 128.38, 127.50, 126.05, 124.17, 123.71, 122.14, 113.47, 110.35, 51.30, 36.05, 29.74, 27.62.; IR (neat): 3027, 2951, 2861, 1726, 1706, 1560, 1435, 1280, 1194, 1088, 751; HRMS (ES+) calculated for $[\text{C}_{20}\text{H}_{19}\text{NnaO}_3]^+$: 344.1263, found: 344.1269.

tert-Butyl 2-(2-oxododecyl)phenylcarbamate **8**



8

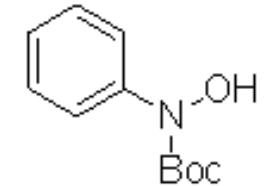
This compound was obtained by quenching the reaction to prepare **3a** right after 20 min according to the general procedure B (eluent: ethyl acetate: hexanes = 1:10). ^1H NMR (500 MHz, CDCl_3) δ 7.72 (d, 1H, $J = 8$ Hz), 7.51 (bs, 1H), 7.17 – 7.21 (m, 1H), 7.06 (dd, 1H, $J_1 = 7.5$ Hz, $J_2 = 1.5$ Hz), 6.97 (td, 1H, $J_1 = 7.5$ Hz, $J_2 = 1$ Hz), 3.68 (s, 2H), 2.55 (t, 2H, $J = 7.5$ Hz), 1.53 – 1.58 (m, 2H), 1.52 (s, 9H), 1.20 – 1.30 (m, 14H), 0.87 (t, 3H, $J =$

7 Hz); ^{13}C NMR (125 MHz, CDCl_3) δ 210.5, 153.6, 137.4, 130.5, 128.2, 125.4, 124.1, 123.4, 80.2, 46.9, 42.7, 31.9, 29.5, 29.4, 29.31, 29.27, 29.0, 28.3, 23.5, 22.7, 14.1; IR (neat): 3340, 2954, 2925, 2854, 1731, 1708, 1452, 1239, 1159; HRMS (ES+) calculated for $[\text{C}_{23}\text{H}_{37}\text{NNaO}_3]^+$: 398.2671, found: 398.2667.

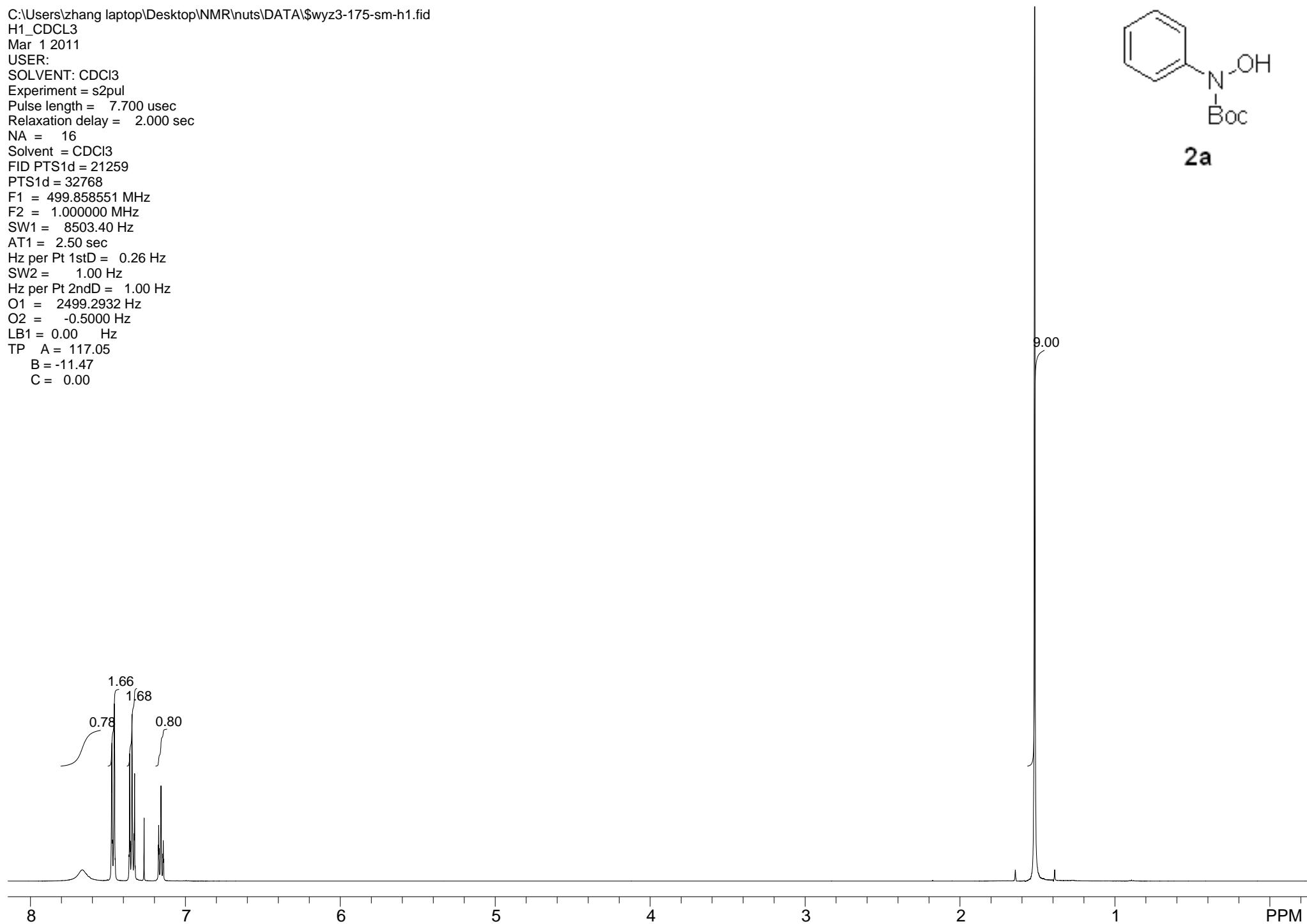
Reference:

1. D. A. Evans, H.-J. Song, K. R. Fandrick. *Org. Lett.* **2006**, *8*, 3351 – 3354.
2. A. Porzelle, N. C. O. Tomkinson, M. D. Woodrow. *Synlett*, **2009**, *5*, 798 – 802.
3. N. Boege, S. Krueger; M. Schroeder, C. Meier. *Synthesis*, **2007**, *24*, 3907 – 3914.
4. N. R. Ayyangar, K. C. Brahme, U. R. Kalkote, K.V. Srinivasan. *Synthesis*, **1984**, *11*, 938 – 941.
5. M. Novak, M. Pelecanou, L. Pollack. *J. Am. Chem. Soc.* **1986**, *108*, 112 – 120.
6. All the yields refer to major isomers if not specified.
7. Stuart, D. R.; Bertrand-Laperle, M.; Burgess, K. M. N.; Fagnou, K. *J. Am. Chem. Soc.* **2008**, *130*, 16474.

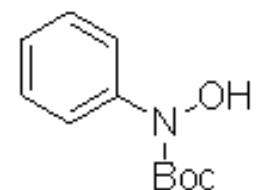
C:\Users\zhang laptop\Desktop\NMR\nuts\DATA\\$wyz3-175-sm-h1.fid
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Mar 1 2011
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SOLVENT: CDCl3
Experiment = s2pul
Pulse length = 7.700 usec
Relaxation delay = 2.000 sec
NA = 16
Solvent = CDCl3
FID PTS1d = 21259
PTS1d = 32768
F1 = 499.858551 MHz
F2 = 1.000000 MHz
SW1 = 8503.40 Hz
AT1 = 2.50 sec
Hz per Pt 1stD = 0.26 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 2499.2932 Hz
O2 = -0.5000 Hz
LB1 = 0.00 Hz
TP A = 117.05
B = -11.47
C = 0.00



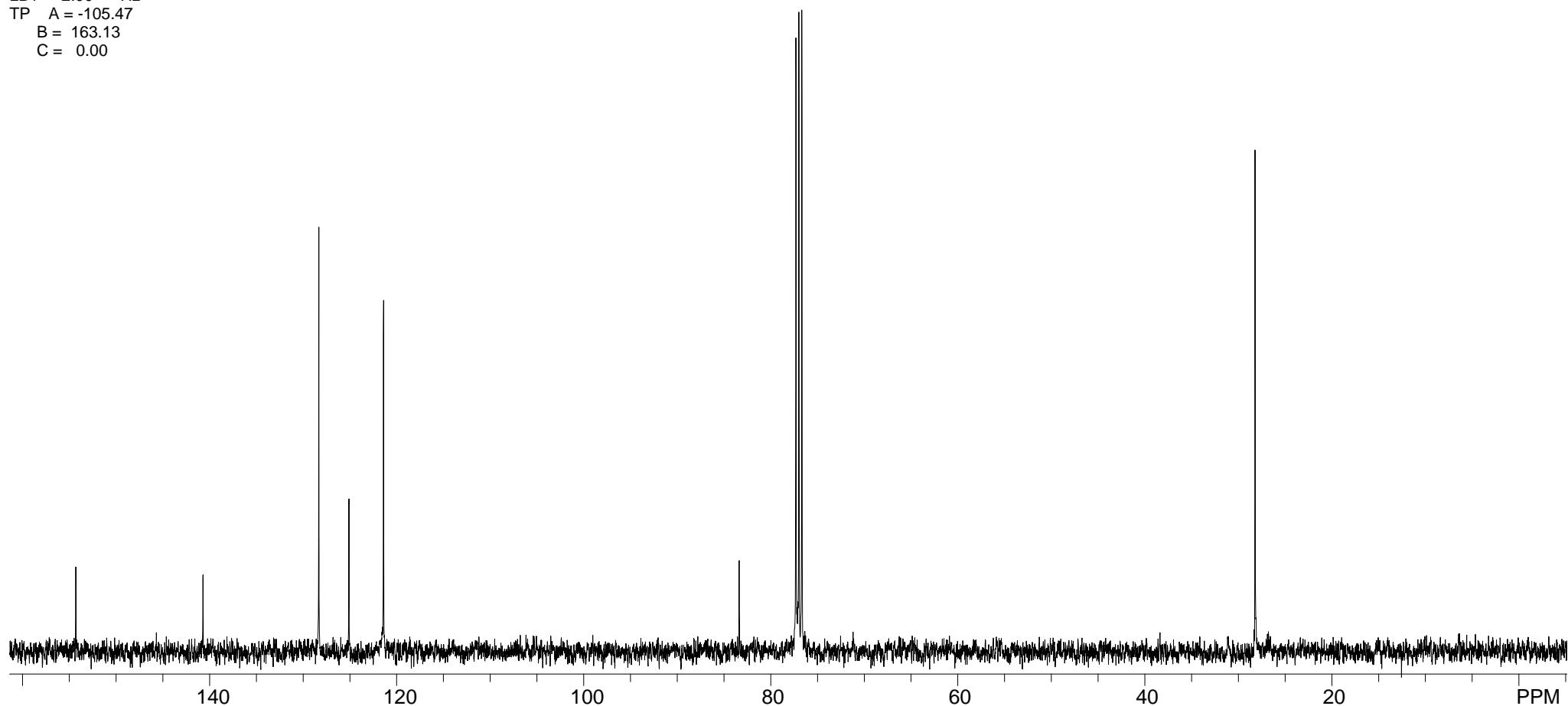
2a



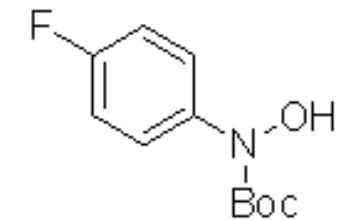
C:\Users\zhang laptop\Desktop\NMR\nuts\DATA\\$wyz3-175-sm-c13.fid
Standard c13 run using qnp probe
Mar 1 2011
USER:
SOLVENT: cdcl3
Experiment = s2pul
Pulse length = 9.500 usec
Relaxation delay = 5.000 sec
NA = 100
Solvent = cdcl3
FID PTS1d = 16000
PTS1d = 16384
F1 = 100.576706 MHz
F2 = 399.949585 MHz
SW1 = 25000.00 Hz
AT1 = 0.64 sec
Hz per Pt 1stD = 1.53 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 9529.8262 Hz
O2 = -0.5000 Hz
LB1 = 2.00 Hz
TP A = -105.47
B = 163.13
C = 0.00



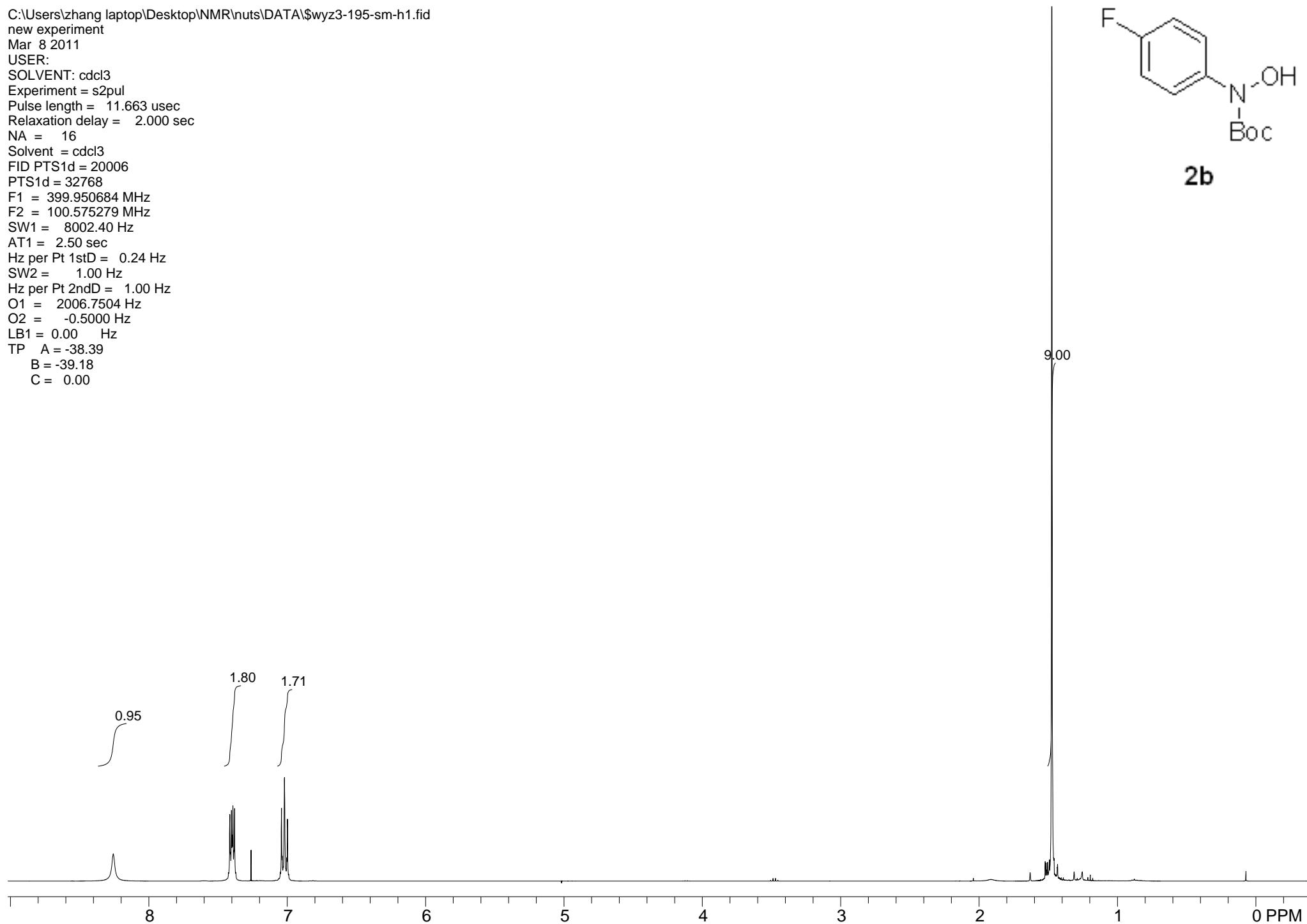
2a



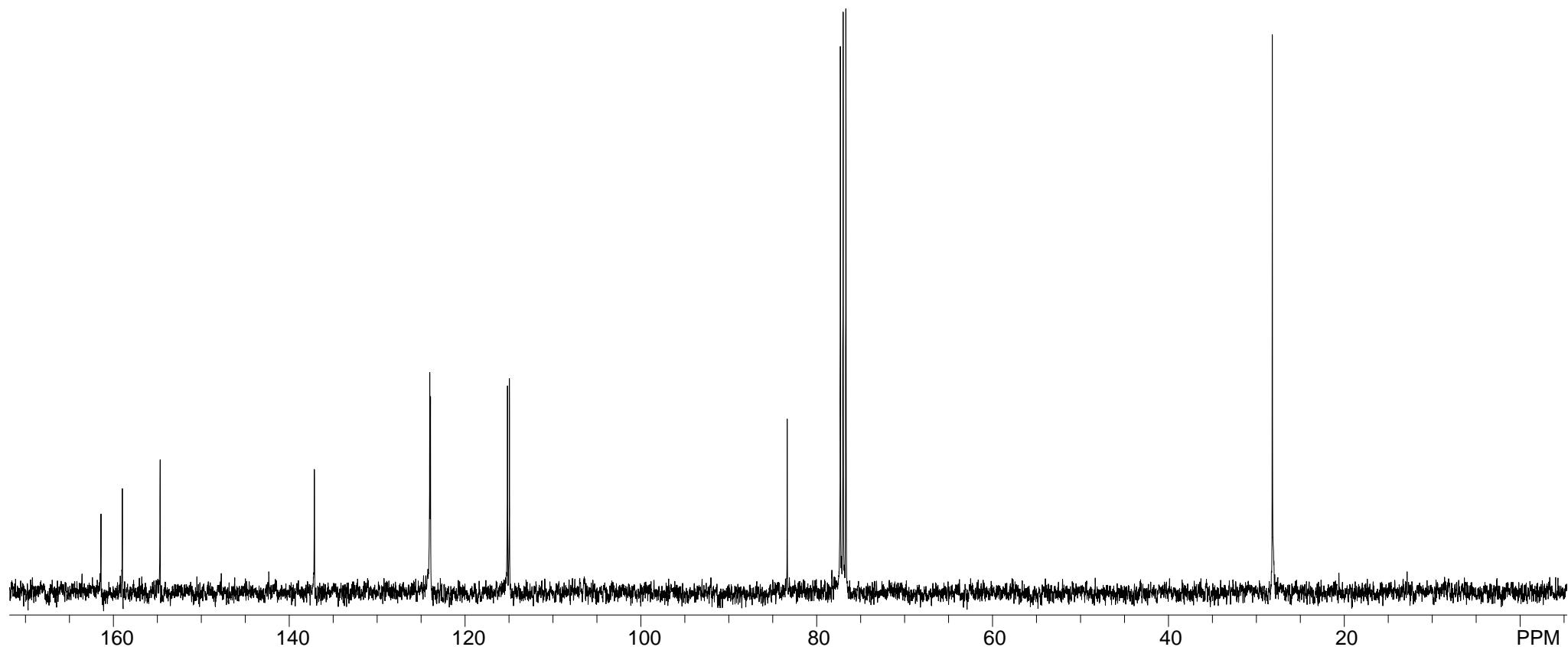
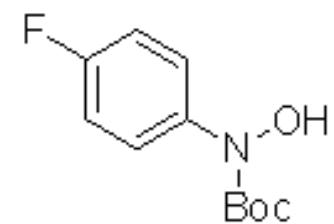
C:\Users\zhang laptop\Desktop\NMR\nuts\DATA\\$wyz3-195-sm-h1.fid
new experiment
Mar 8 2011
USER:
SOLVENT: cdcl3
Experiment = s2pul
Pulse length = 11.663 usec
Relaxation delay = 2.000 sec
NA = 16
Solvent = cdcl3
FID PTS1d = 20006
PTS1d = 32768
F1 = 399.950684 MHz
F2 = 100.575279 MHz
SW1 = 8002.40 Hz
AT1 = 2.50 sec
Hz per Pt 1stD = 0.24 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 2006.7504 Hz
O2 = -0.5000 Hz
LB1 = 0.00 Hz
TP A = -38.39
B = -39.18
C = 0.00



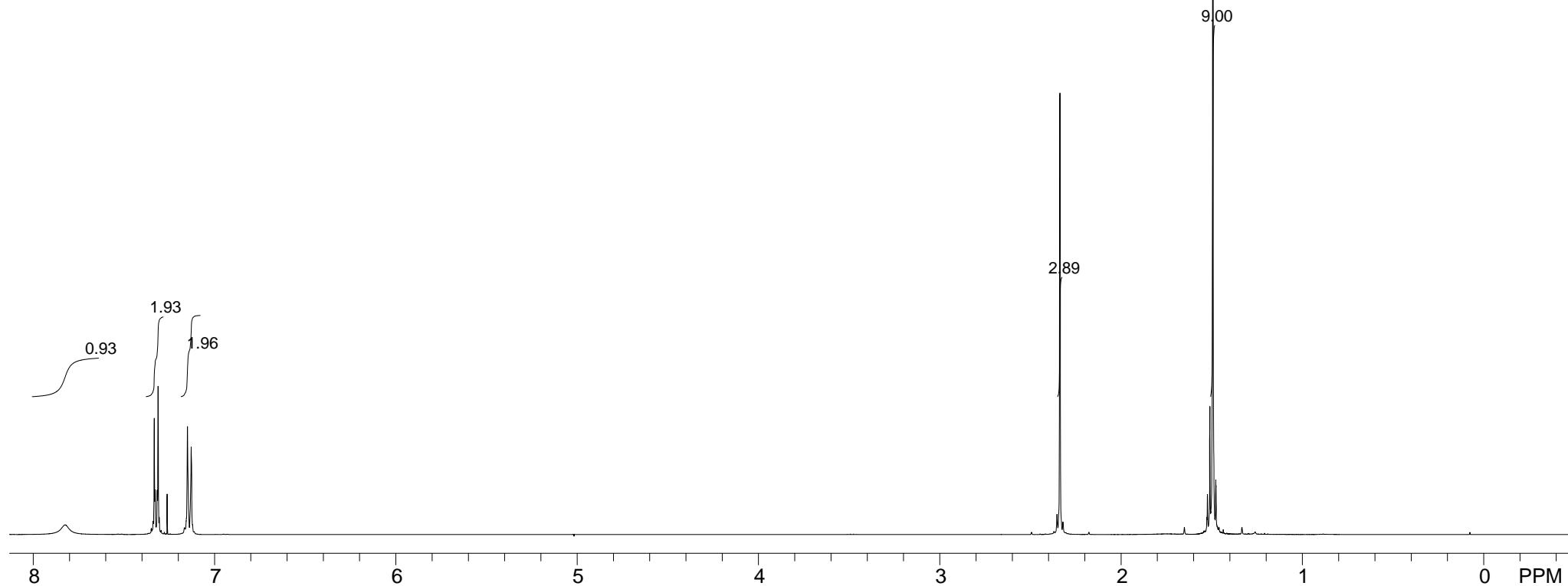
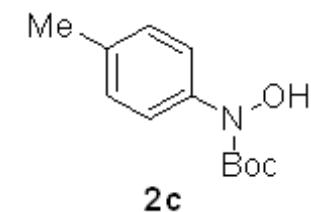
2b



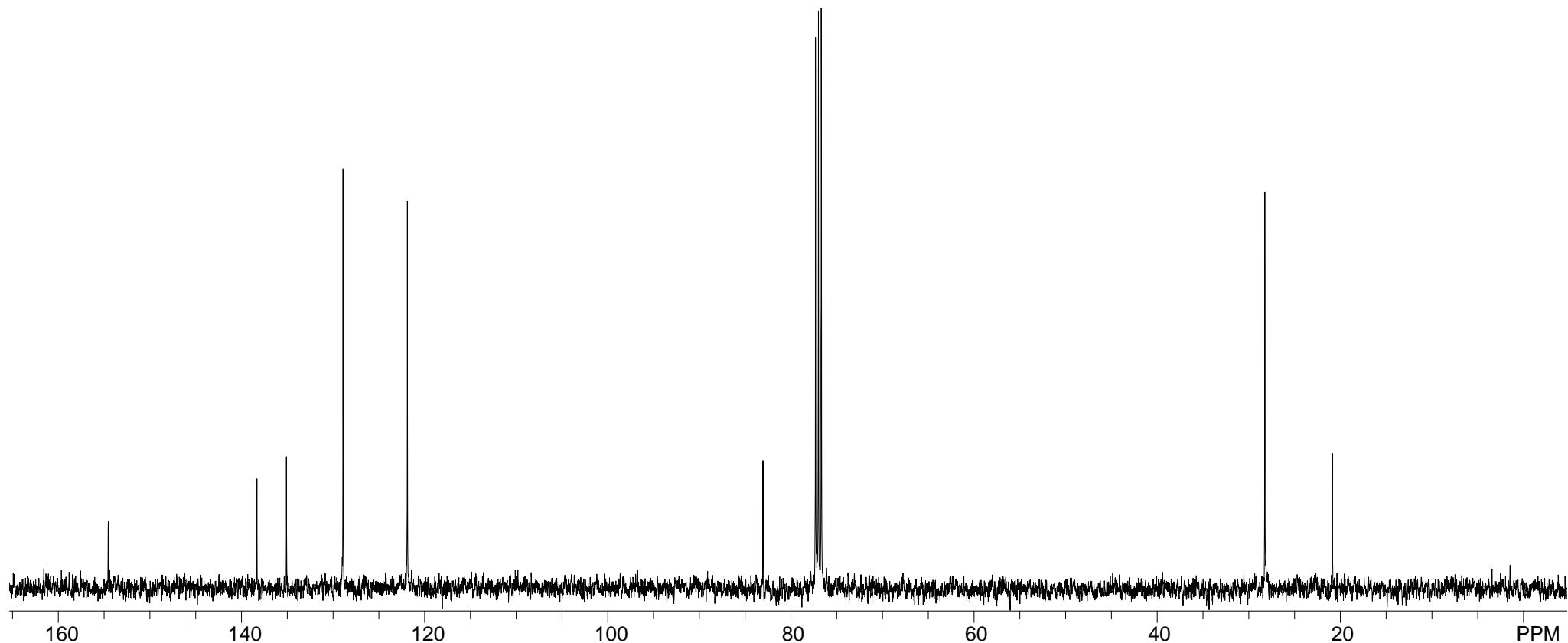
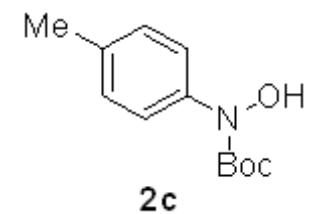
C:\Users\zhang laptop\Desktop\NMR\nuts\DATA\\$wyz3-195-sm-c13.fid
Standard c13 run using qnp probe
Mar 8 2011
USER:
SOLVENT: cdcl3
Experiment = s2pul
Pulse length = 9.500 usec
Relaxation delay = 5.000 sec
NA = 80
Solvent = cdcl3
FID PTS1d = 16000
PTS1d = 16384
F1 = 100.576706 MHz
F2 = 399.949585 MHz
SW1 = 25000.00 Hz
AT1 = 0.64 sec
Hz per Pt 1stD = 1.53 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 9529.8262 Hz
O2 = -0.5000 Hz
LB1 = 2.00 Hz
TP A = -97.03
B = 149.06
C = 0.00



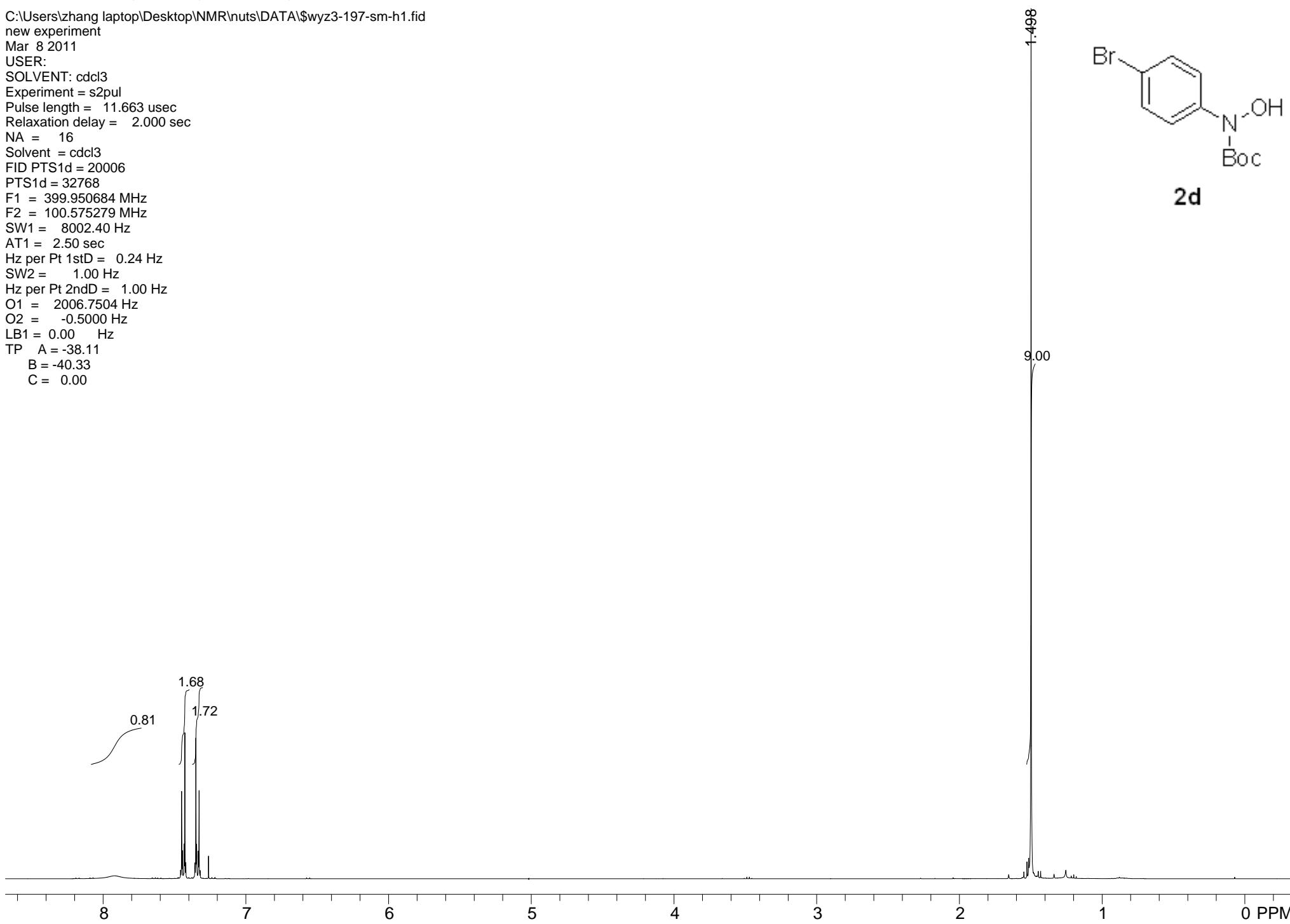
C:\Users\zhang laptop\Desktop\NMR\nuts\DATA\\$wyz3-196-sm-h1.fid
new experiment
Mar 8 2011
USER:
SOLVENT: cdcl3
Experiment = s2pul
Pulse length = 11.663 usec
Relaxation delay = 2.000 sec
NA = 16
Solvent = cdcl3
FID PTS1d = 20006
PTS1d = 32768
F1 = 399.950684 MHz
F2 = 100.575279 MHz
SW1 = 8002.40 Hz
AT1 = 2.50 sec
Hz per Pt 1stD = 0.24 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 2006.7504 Hz
O2 = -0.5000 Hz
LB1 = 0.00 Hz
TP A = -37.99
B = -40.04
C = 0.00



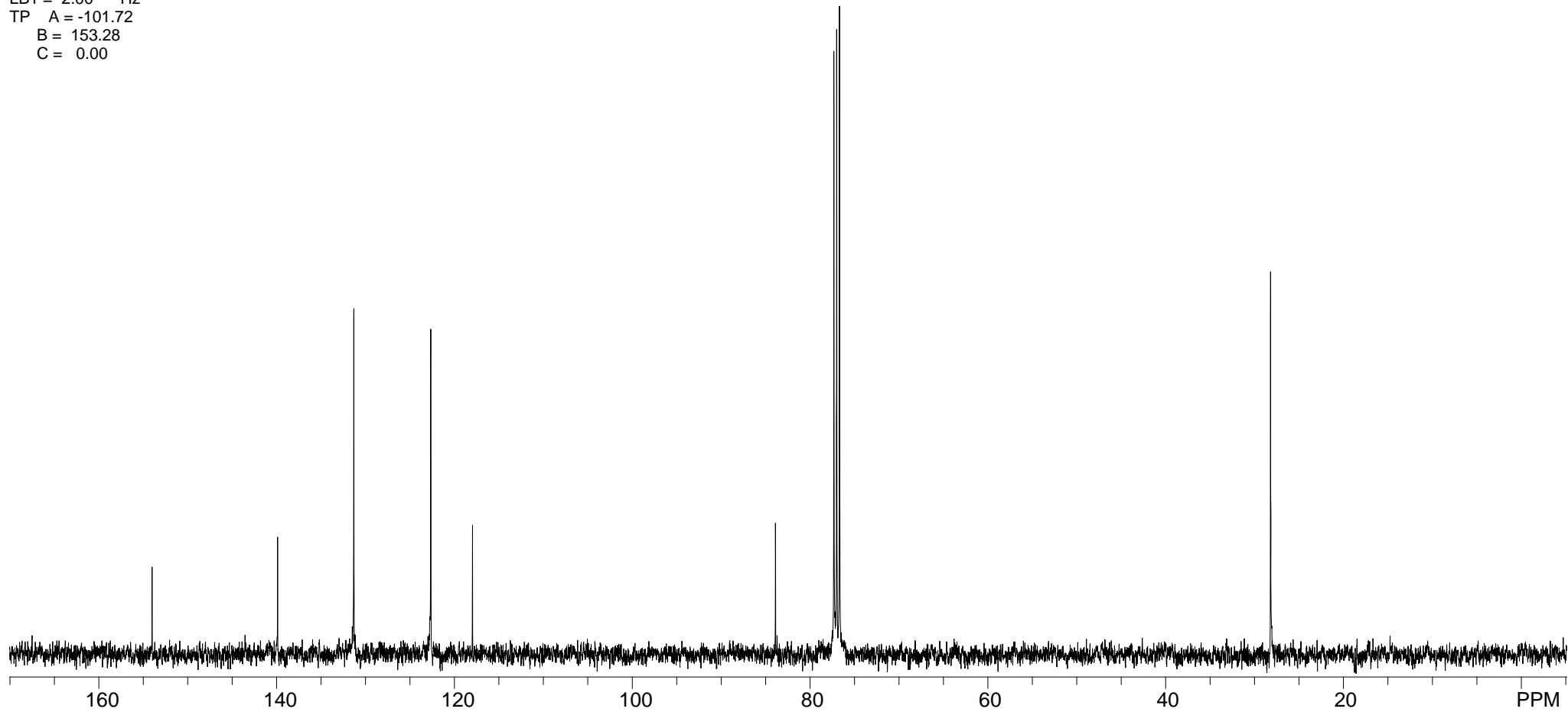
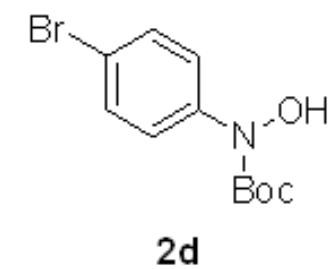
C:\Users\zhang laptop\Desktop\NMR\nuts\DATA\\$wyz3-196-sm-c13.fid
Standard c13 run using qnp probe
Mar 8 2011
USER:
SOLVENT: cdcl3
Experiment = s2pul
Pulse length = 9.500 usec
Relaxation delay = 5.000 sec
NA = 84
Solvent = cdcl3
FID PTS1d = 16000
PTS1d = 16384
F1 = 100.576706 MHz
F2 = 399.949585 MHz
SW1 = 25000.00 Hz
AT1 = 0.64 sec
Hz per Pt 1stD = 1.53 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 9529.8262 Hz
O2 = -0.5000 Hz
LB1 = 2.00 Hz
TP A = -126.09
B = 191.25
C = 0.00



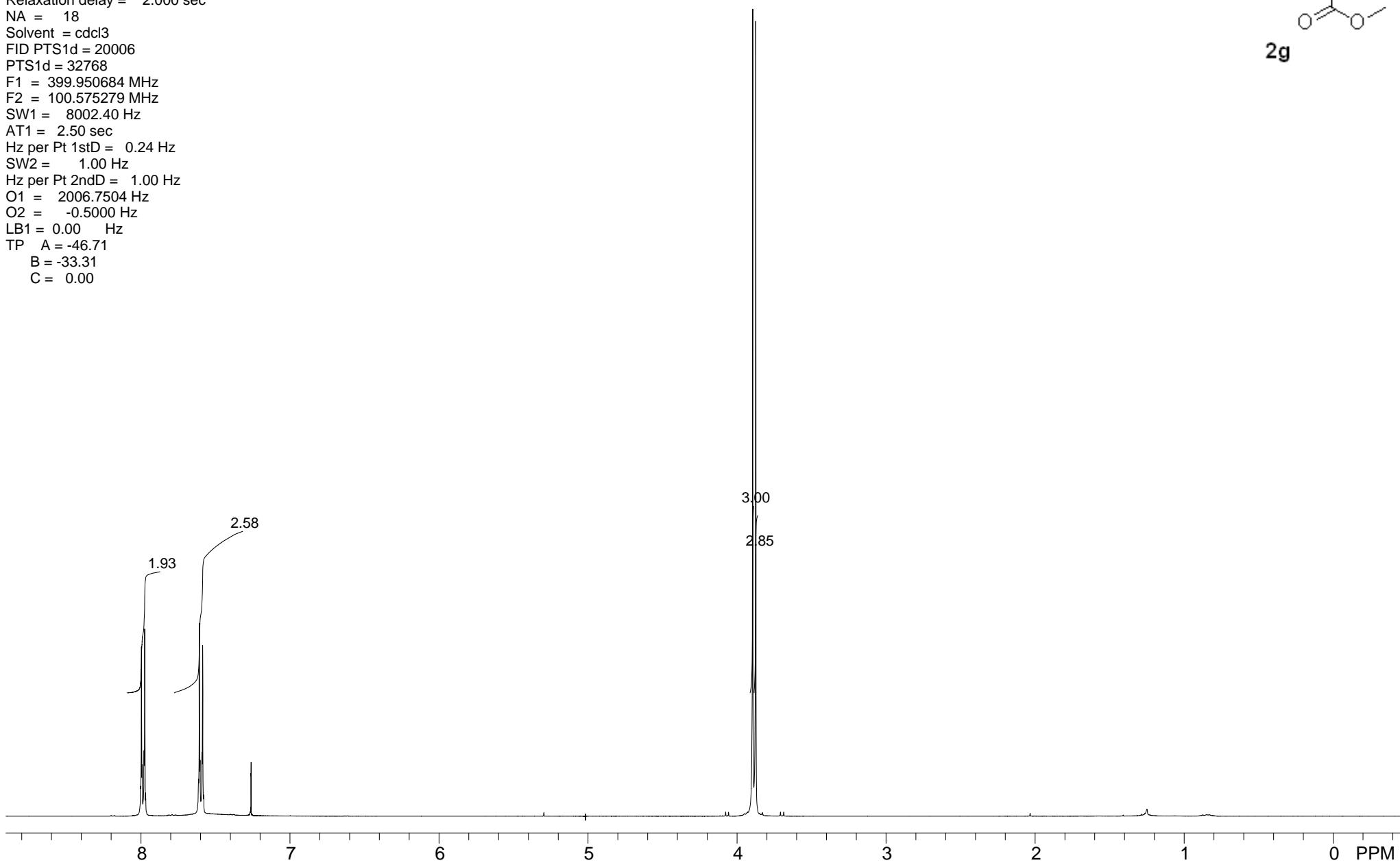
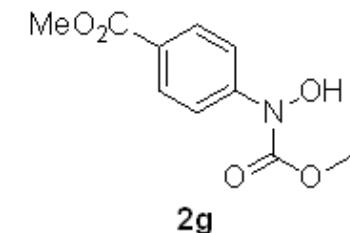
C:\Users\zhang laptop\Desktop\NMR\nuts\DATA\\$wyz3-197-sm-h1.fid
new experiment
Mar 8 2011
USER:
SOLVENT: cdcl3
Experiment = s2pul
Pulse length = 11.663 usec
Relaxation delay = 2.000 sec
NA = 16
Solvent = cdcl3
FID PTS1d = 20006
PTS1d = 32768
F1 = 399.950684 MHz
F2 = 100.575279 MHz
SW1 = 8002.40 Hz
AT1 = 2.50 sec
Hz per Pt 1stD = 0.24 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 2006.7504 Hz
O2 = -0.5000 Hz
LB1 = 0.00 Hz
TP A = -38.11
B = -40.33
C = 0.00



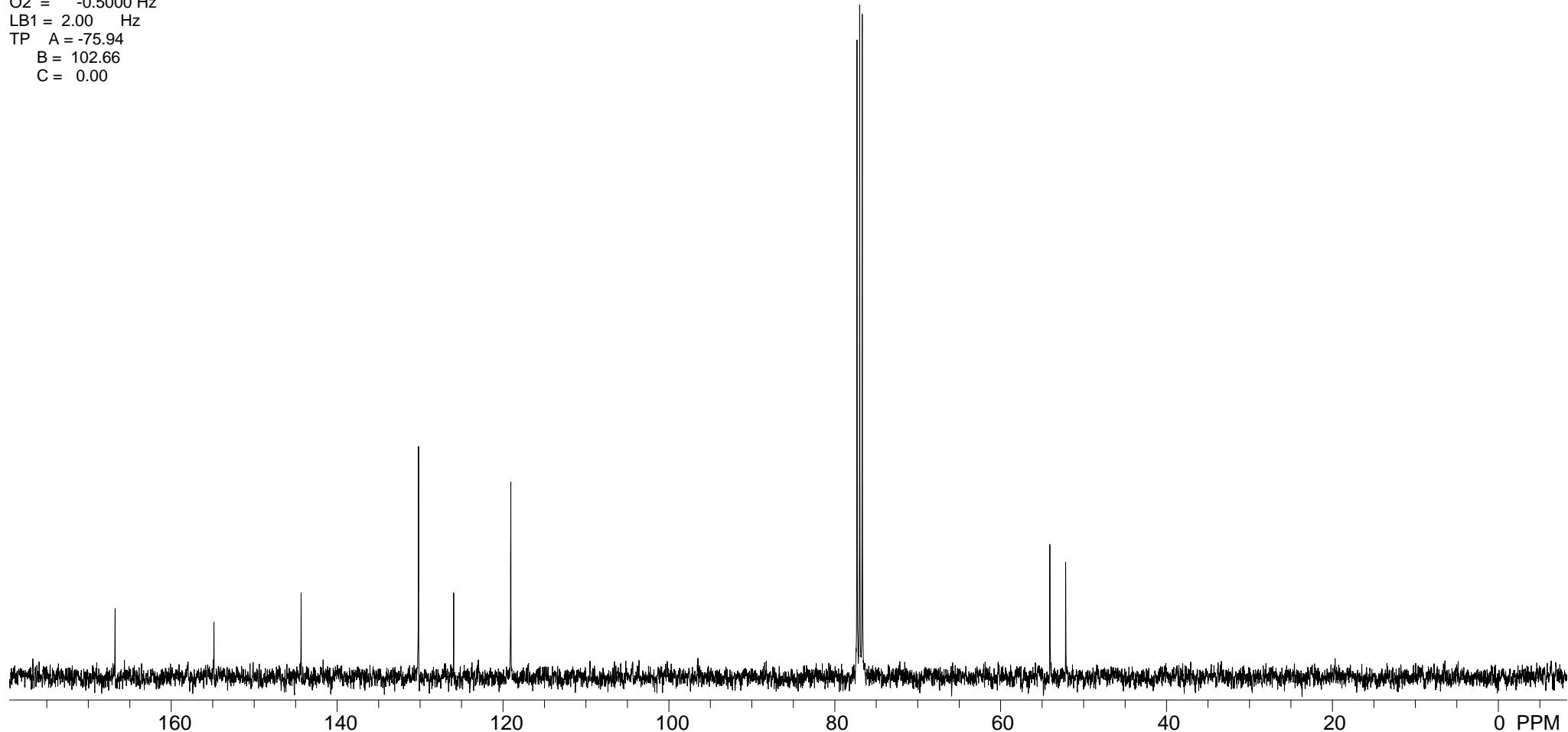
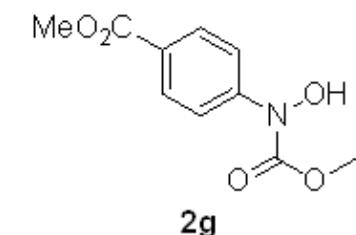
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Standard c13 run using qnp probe
Mar 8 2011
USER:
SOLVENT: cdcl3
Experiment = s2pul
Pulse length = 9.500 usec
Relaxation delay = 5.000 sec
NA = 88
Solvent = cdcl3
FID PTS1d = 16000
PTS1d = 16384
F1 = 100.576706 MHz
F2 = 399.949585 MHz
SW1 = 25000.00 Hz
AT1 = 0.64 sec
Hz per Pt 1stD = 1.53 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 9529.8223 Hz
O2 = -0.5000 Hz
LB1 = 2.00 Hz
TP A = -101.72
B = 153.28
C = 0.00



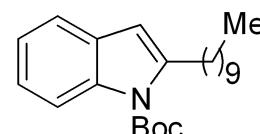
C:\Users\zhang laptop\Desktop\NMR\nuts\DATA\\$wyz3-216-sm-h1.fid
new experiment
Mar 21 2011
USER:
SOLVENT: cdcl3
Experiment = s2pul
Pulse length = 11.663 usec
Relaxation delay = 2.000 sec
NA = 18
Solvent = cdcl3
FID PTS1d = 20006
PTS1d = 32768
F1 = 399.950684 MHz
F2 = 100.575279 MHz
SW1 = 8002.40 Hz
AT1 = 2.50 sec
Hz per Pt 1stD = 0.24 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 2006.7504 Hz
O2 = -0.5000 Hz
LB1 = 0.00 Hz
TP A = -46.71
B = -33.31
C = 0.00



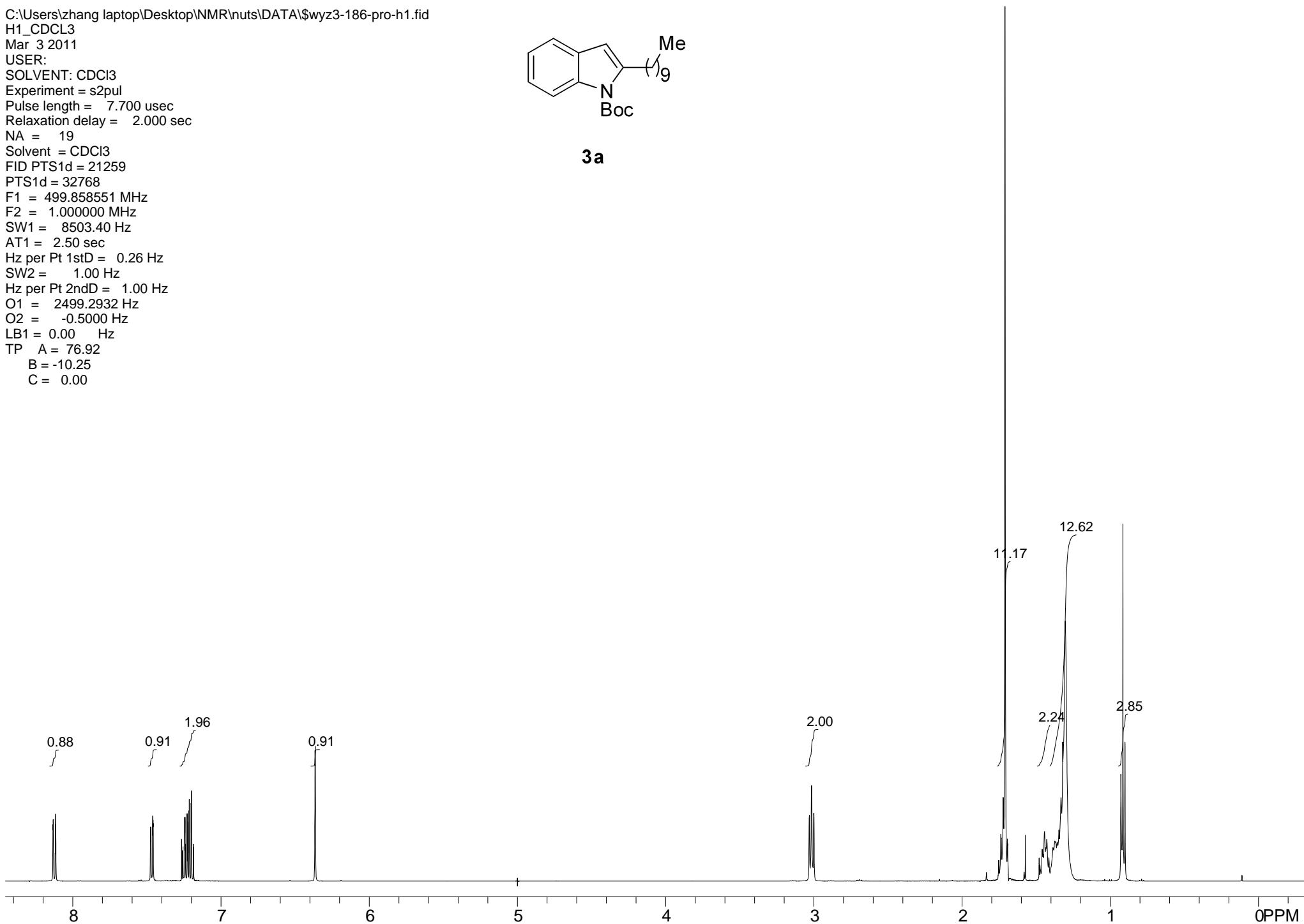
C:\Users\zhang laptop\Desktop\NMR\nuts\DATA\\$wyz3-216-sm-c13.fid
Standard c13 run using qnp probe
Mar 21 2011
USER:
SOLVENT: cdcl3
Experiment = s2pul
Pulse length = 9.500 usec
Relaxation delay = 5.000 sec
NA = 100
Solvent = cdcl3
FID PTS1d = 16000
PTS1d = 16384
F1 = 100.576706 MHz
F2 = 399.949585 MHz
SW1 = 25000.00 Hz
AT1 = 0.64 sec
Hz per Pt 1stD = 1.53 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 9529.8223 Hz
O2 = -0.5000 Hz
LB1 = 2.00 Hz
TP A = -75.94
B = 102.66
C = 0.00



C:\Users\zhang laptop\Desktop\NMR\nuts\DATA\\$wyz3-186-pro-h1.fid
H1_CDCL3
Mar. 3 2011
USER:
SOLVENT: CDCl3
Experiment = s2pul
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Relaxation delay = 2.000 sec
NA = 19
Solvent = CDCl3
FID PTS1d = 21259
PTS1d = 32768
F1 = 499.858551 MHz
F2 = 1.000000 MHz
SW1 = 8503.40 Hz
AT1 = 2.50 sec
Hz per Pt 1stD = 0.26 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 2499.2932 Hz
O2 = -0.5000 Hz
LB1 = 0.00 Hz
TP A = 76.92
B = -10.25
C = 0.00



3a



C:\Users\zhang laptop\Desktop\NMR\nuts\DATA\\$wyz3-186-pro-c13.fid

STANDARD CARBON PARAMETERS

Mar 3 2011

USER:

SOLVENT: cdcl3

Experiment = s2pul

Pulse length = 6.000 usec

Relaxation delay = 3.000 sec

NA = 60

Solvent = cdcl3

FID PTS1d = 36749

PTS1d = 65536

F1 = 125.701683 MHz

F2 = 499.858551 MHz

SW1 = 28258.57 Hz

AT1 = 1.30 sec

Hz per Pt 1stD = 0.43 Hz

SW2 = 1.00 Hz

Hz per Pt 2ndD = 1.00 Hz

O1 = 12769.5205 Hz

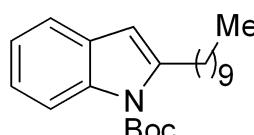
O2 = -0.5000 Hz

LB1 = 2.00 Hz

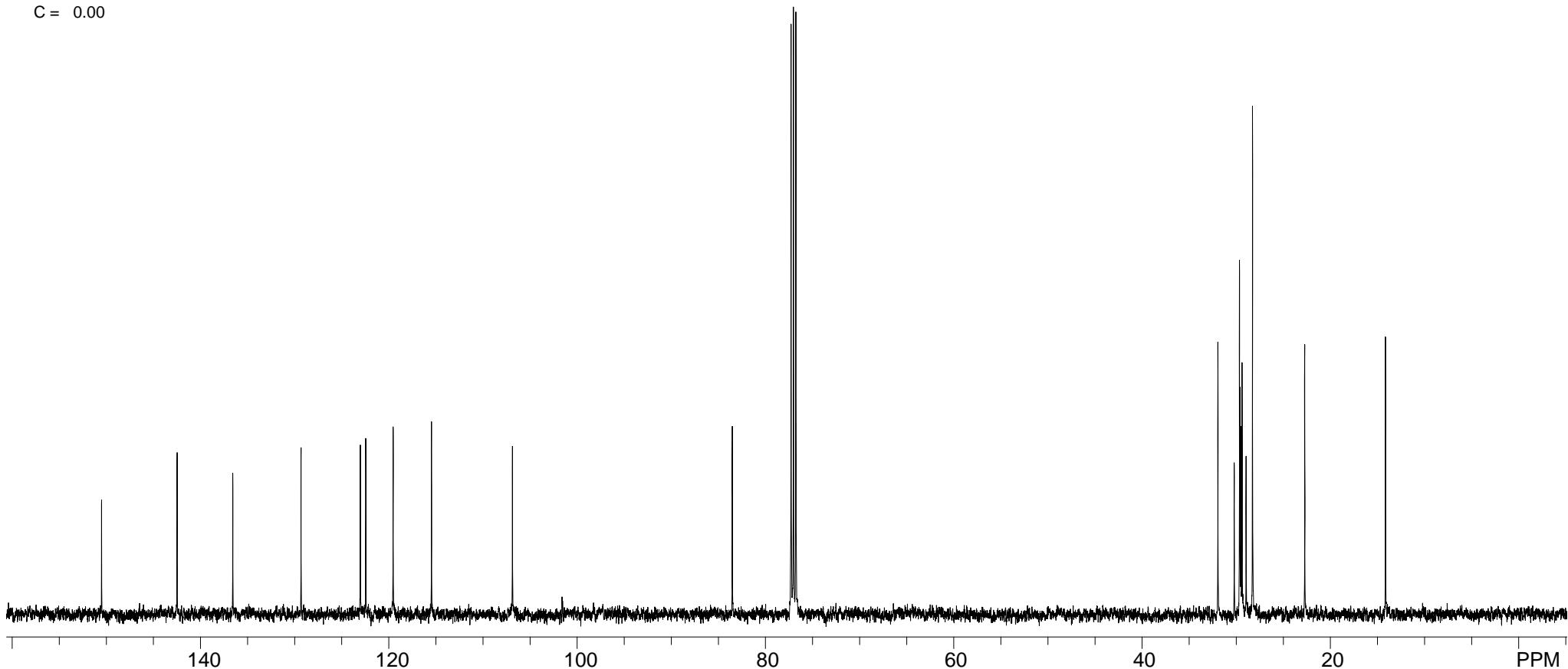
TP A = -245.63

B = 146.25

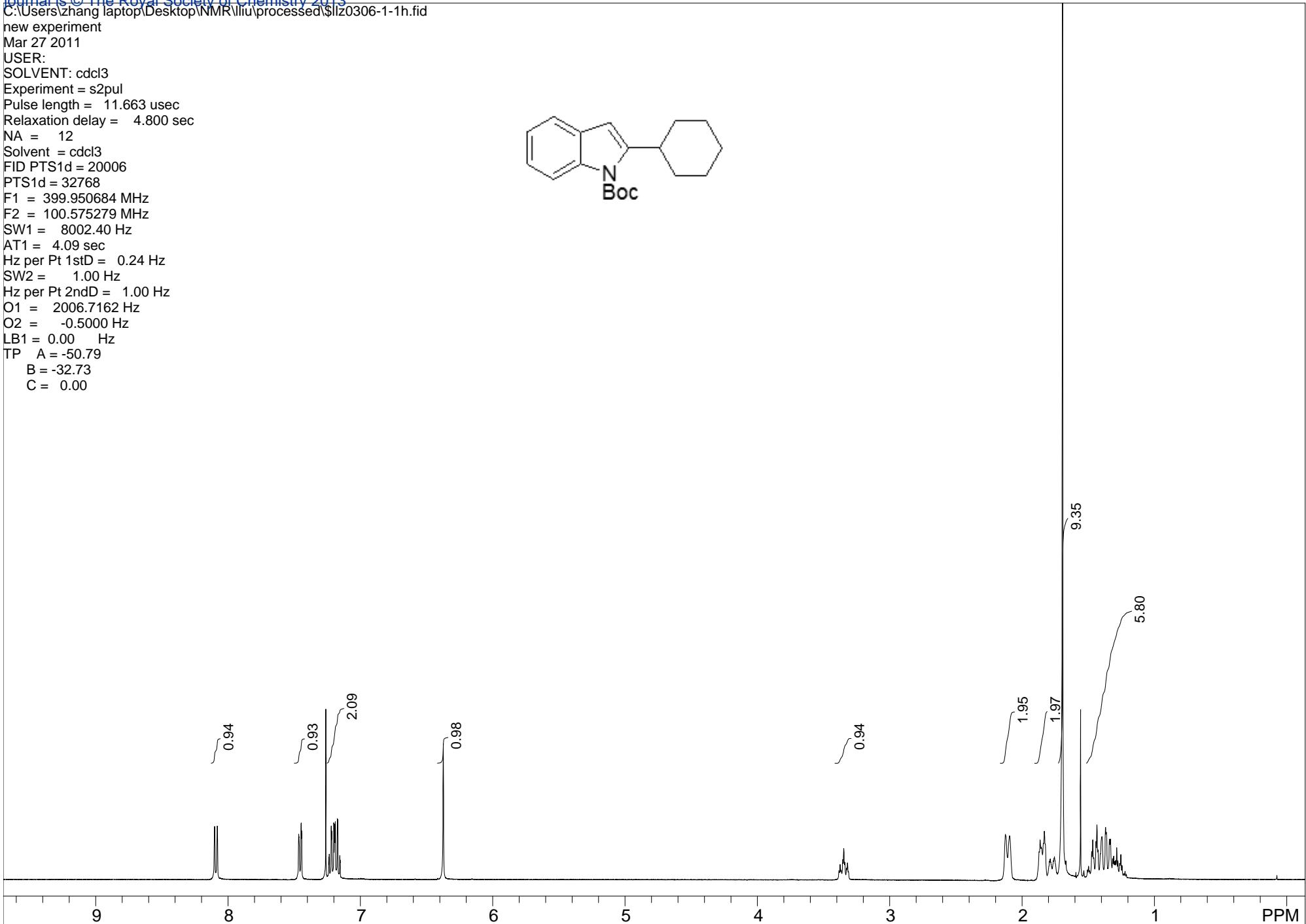
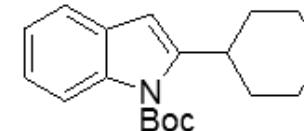
C = 0.00



3a



C:\Users\zhang\Desktop\NMR\lliu\processed\\$llz0306-1-1h.fid
 new experiment
 Mar 27 2011
 USER:
 SOLVENT: cdcl3
 Experiment = s2pul
 Pulse length = 11.663 usec
 Relaxation delay = 4.800 sec
 NA = 12
 Solvent = cdcl3
 FID PTS1d = 20006
 PTS1d = 32768
 F1 = 399.950684 MHz
 F2 = 100.575279 MHz
 SW1 = 8002.40 Hz
 AT1 = 4.09 sec
 Hz per Pt 1stD = 0.24 Hz
 SW2 = 1.00 Hz
 Hz per Pt 2ndD = 1.00 Hz
 O1 = 2006.7162 Hz
 O2 = -0.5000 Hz
 LB1 = 0.00 Hz
 TP A = -50.79
 B = -32.73
 C = 0.00



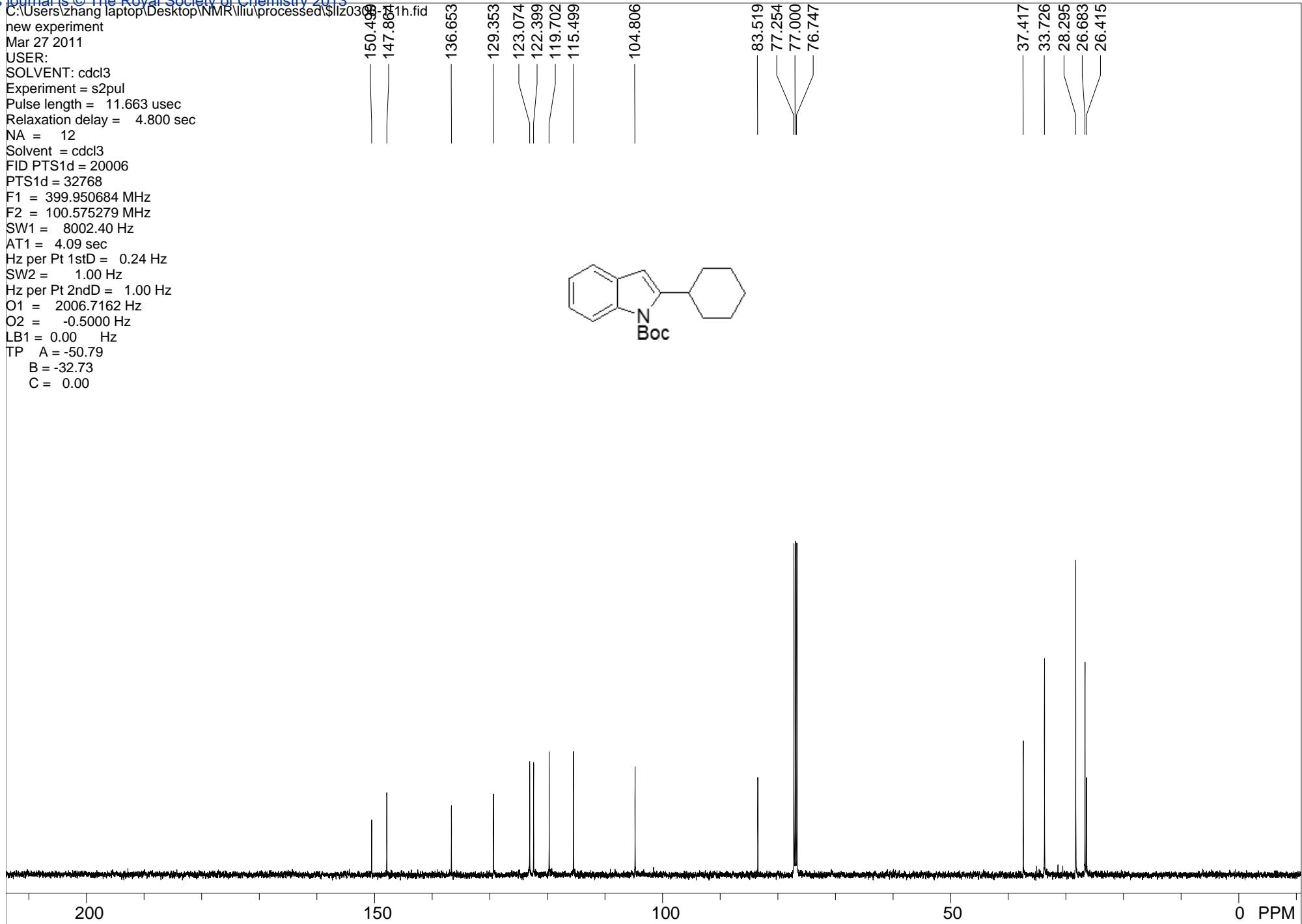
new experiment

USER: -- DATE: Mar 27 2011

F1: 399.951	F2: 100.575	SW1: 8002		OF1: 2006.7		PTS1d: 20006 , 32768	
EX: s2pul		PW: 11.7 us		PD: 4.8 sec		NA: 12	LB: 0.0

Nuts - \\$llz0306-1-1h.fid

C:\Users\zhang\Desktop\NMR\lliu\processed\\$llz0306-11h.fid
new experiment
Mar 27 2011
USER:
SOLVENT: cdcl3
Experiment = s2pul
Pulse length = 11.663 usec
Relaxation delay = 4.800 sec
NA = 12
Solvent = cdcl3
FID PTS1d = 20006
PTS1d = 32768
F1 = 399.950684 MHz
F2 = 100.575279 MHz
SW1 = 8002.40 Hz
AT1 = 4.09 sec
Hz per Pt 1stD = 0.24 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 2006.7162 Hz
O2 = -0.5000 Hz
LB1 = 0.00 Hz
TP A = -50.79
B = -32.73
C = 0.00



STANDARD CARBON PARAMETERS

F1: 125.702	F2: 499.859	SW1: 28259		OF1: 12768.3		PTS1d: 36749 , 65536	USER: -- DATE: Mar 6 2011
EX: s2pul		PW: 6.0 us	PD: 3.0 sec	NA: 69	LB: 2.0		Nuts - \\$llz20110306-1-13c

C:\Users\zhang\Desktop\NMR\lliu\processed\\$llz0306-1-1h.fid

new experiment

Mar 27 2011

USER:

SOLVENT: cdcl3

Experiment = s2pul

Pulse length = 11.663 usec

Relaxation delay = 4.800 sec

NA = 12

Solvent = cdcl3

FID PTS1d = 20006

PTS1d = 32768

F1 = 399.950684 MHz

F2 = 100.575279 MHz

SW1 = 8002.40 Hz

AT1 = 4.09 sec

Hz per Pt 1stD = 0.24 Hz

SW2 = 1.00 Hz

Hz per Pt 2ndD = 1.00 Hz

O1 = 2006.7162 Hz

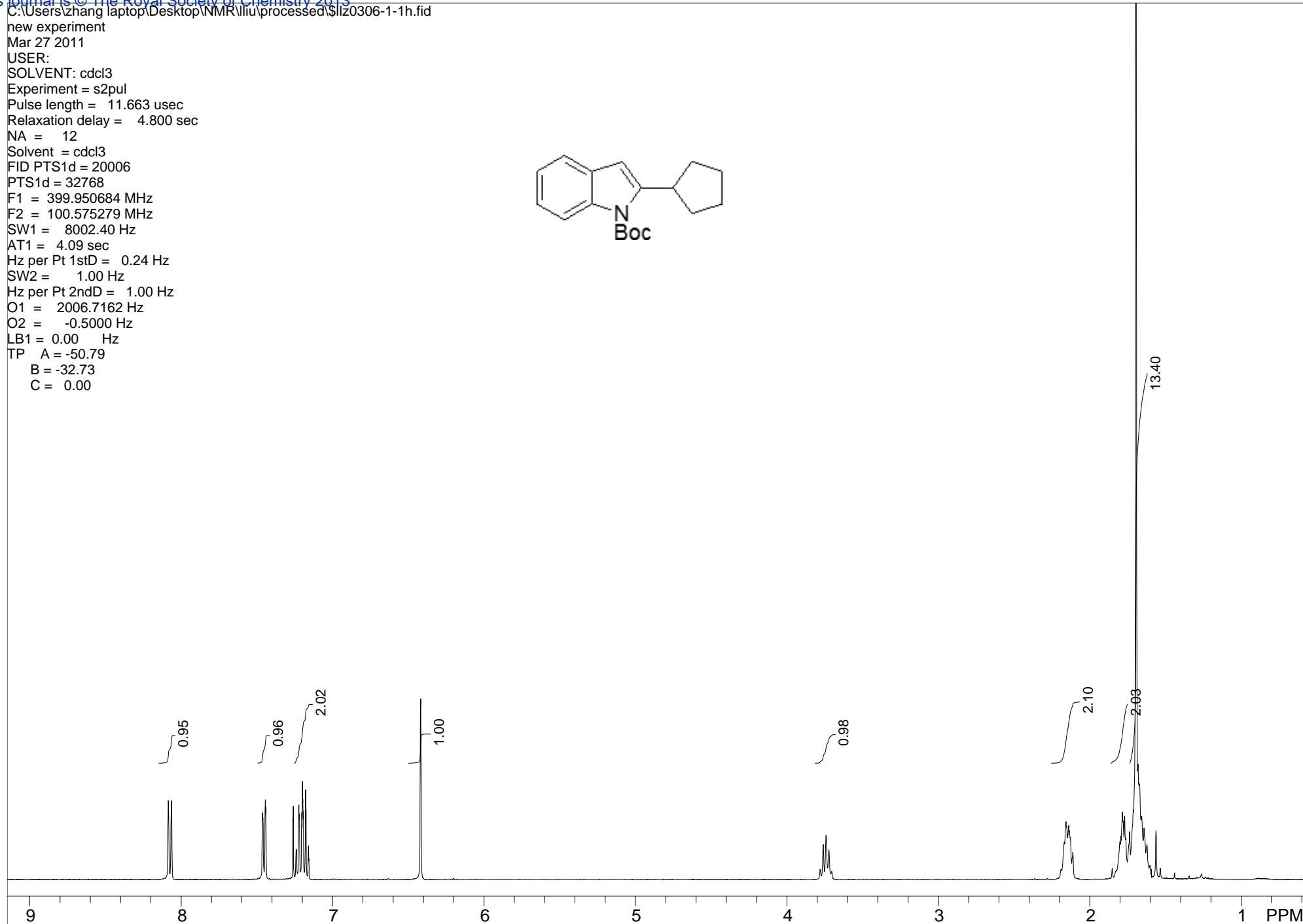
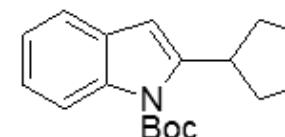
O2 = -0.5000 Hz

LB1 = 0.00 Hz

TP A = -50.79

B = -32.73

C = 0.00



new experiment

USER: -- DATE: Mar 27 2011

F1: 399.951	F2: 100.575	SW1: 8002		OF1: 2006.2		PTS1d: 20006 , 32768	
EX: s2pul		PW: 11.7 us		PD: 4.8 sec		NA: 16	LB: 0.0

Nuts - \\$llz-0306-2-1h.fid

C:\Users\zhang\Desktop\NMR\llz0306-2-13c

STANDARD CARBON PARAMETERS

Mar 6 2011

USER:

SOLVENT: cdcl3

Experiment = s2pul

Pulse length = 6.000 usec

Relaxation delay = 3.000 sec

NA = 109

Solvent = cdcl3

FID PTS1d = 36749

PTS1d = 65536

F1 = 125.701683 MHz

F2 = 499.858551 MHz

SW1 = 28258.57 Hz

AT1 = 2.32 sec

Hz per Pt 1stD = 0.43 Hz

SW2 = 1.00 Hz

Hz per Pt 2ndD = 1.00 Hz

O1 = 12770.9824 Hz

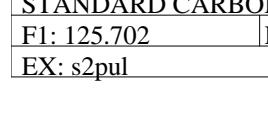
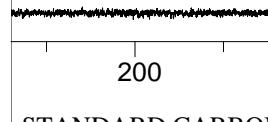
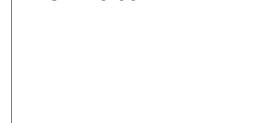
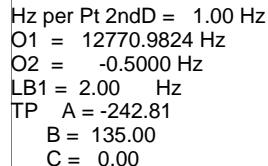
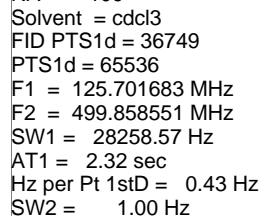
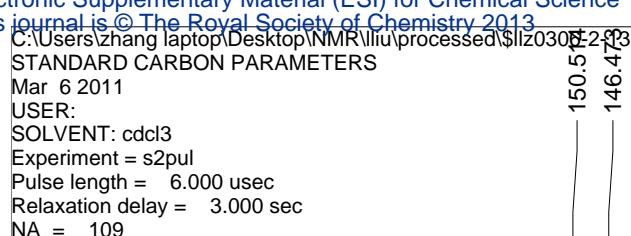
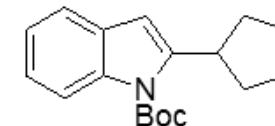
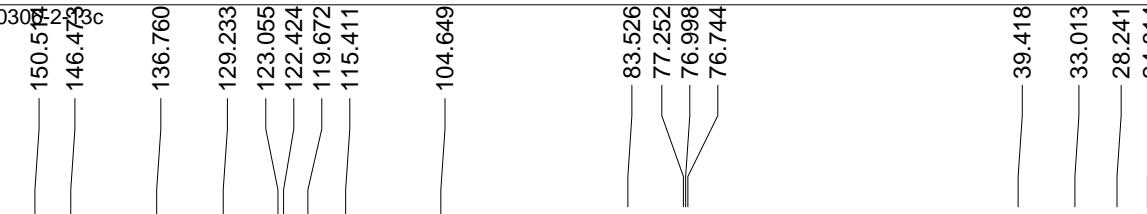
O2 = -0.5000 Hz

LB1 = 2.00 Hz

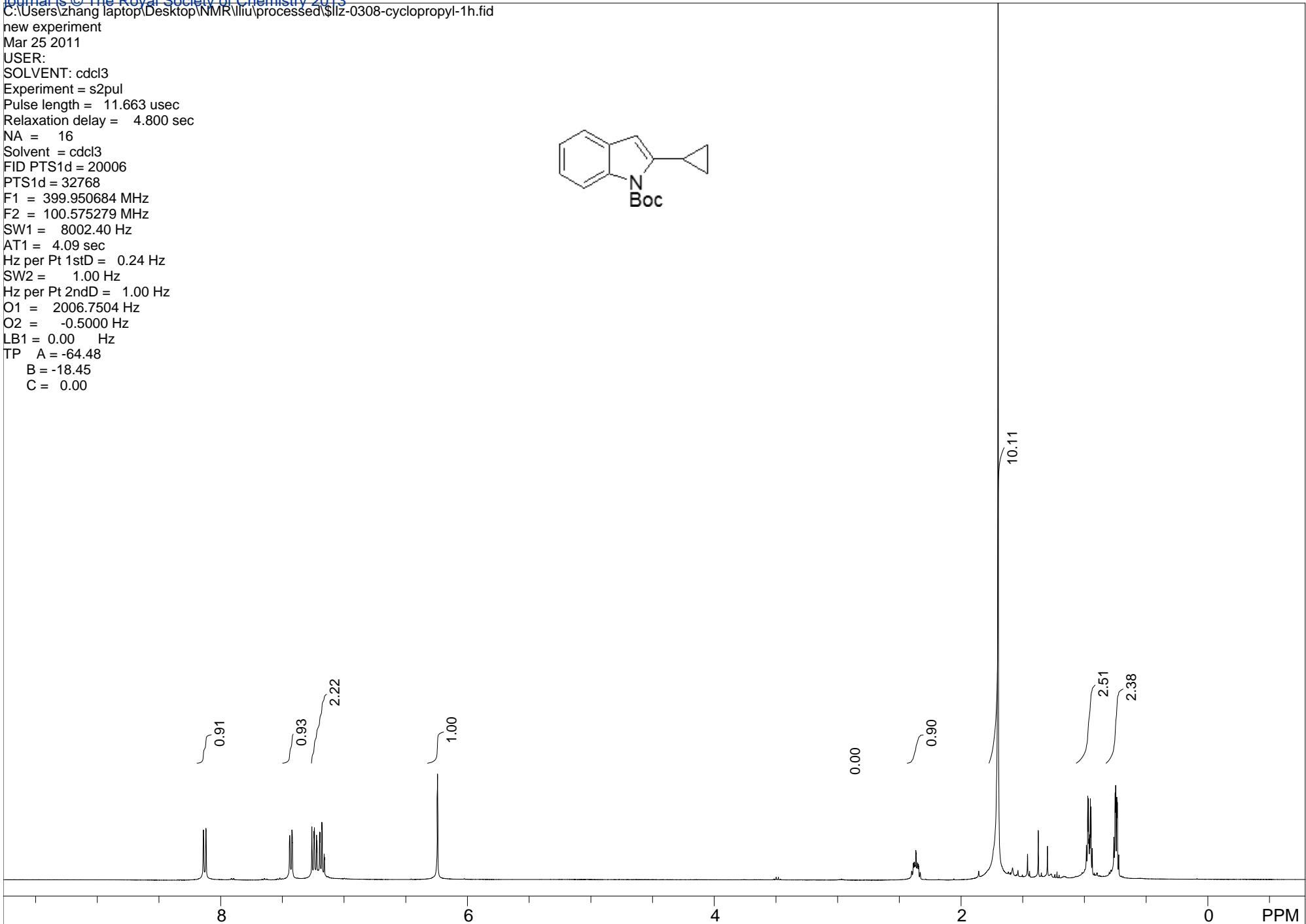
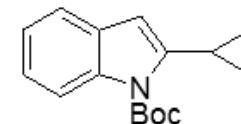
TP A = -242.81

B = 135.00

C = 0.00



C:\Users\zhang\Desktop\NMR\liu\processed\\$llz-0308-cyclopropyl-1h.fid
 new experiment
 Mar 25 2011
 USER:
 SOLVENT: cdcl3
 Experiment = s2pul
 Pulse length = 11.663 usec
 Relaxation delay = 4.800 sec
 NA = 16
 Solvent = cdcl3
 FID PTS1d = 20006
 PTS1d = 32768
 F1 = 399.950684 MHz
 F2 = 100.575279 MHz
 SW1 = 8002.40 Hz
 AT1 = 4.09 sec
 Hz per Pt 1stD = 0.24 Hz
 SW2 = 1.00 Hz
 Hz per Pt 2ndD = 1.00 Hz
 O1 = 2006.7504 Hz
 O2 = -0.5000 Hz
 LB1 = 0.00 Hz
 TP A = -64.48
 B = -18.45
 C = 0.00



new experiment

USER: -- DATE: Mar 25 2011

F1: 399.951	F2: 100.575	SW1: 8002		OF1: 2006.8		PTS1d: 20006 , 32768	
EX: s2pul		PW: 11.7 us		PD: 4.8 sec	NA: 16	LB: 0.0	Nuts - \\$llz-0308-cyclopropyl-1h.fid

C:\Users\zhang\Desktop\NMR\llz-0308-cyclopropyl-13c

STANDARD CARBON PARAMETERS

Mar 8 2011

USER:

SOLVENT: cdcl3

Experiment = s2pul

Pulse length = 6.000 usec

Relaxation delay = 3.000 sec

NA = 90

Solvent = cdcl3

FID PTS1d = 36749

PTS1d = 65536

F1 = 125.701683 MHz

F2 = 499.858551 MHz

SW1 = 28258.57 Hz

AT1 = 2.32 sec

Hz per Pt 1stD = 0.43 Hz

SW2 = 1.00 Hz

Hz per Pt 2ndD = 1.00 Hz

O1 = 12777.2832 Hz

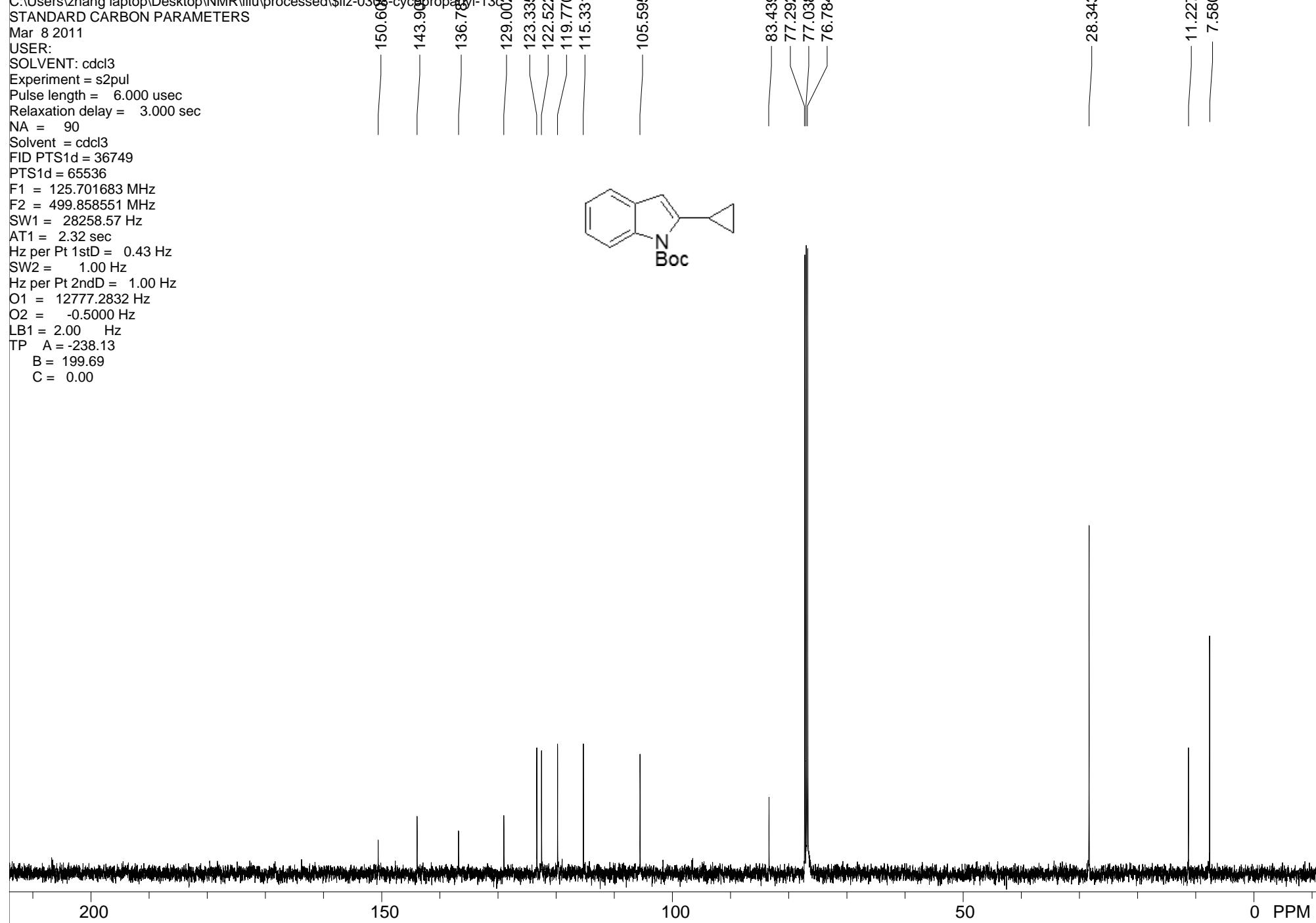
O2 = -0.5000 Hz

LB1 = 2.00 Hz

TP A = -238.13

B = 199.69

C = 0.00



STANDARD CARBON PARAMETERS

F1: 125.702

F2: 499.859

SW1: 28259

EX: s2pul

150

100

50

0 PPM

USER: -- DATE: Mar 8 2011

PW: 6.0 us

PD: 3.0 sec

NA: 90

LB: 2.0

PTS1d: 36749, 65536

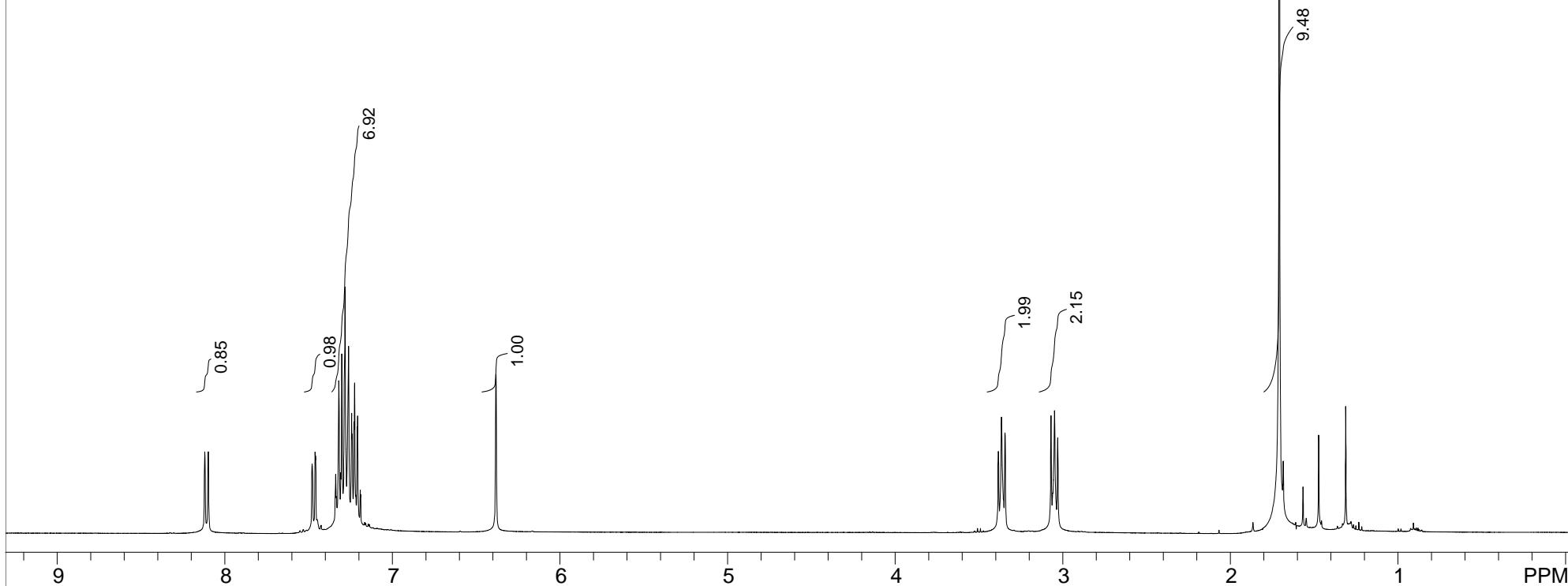
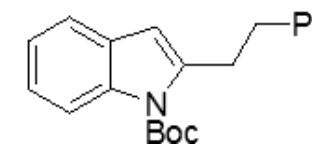
Nuts - \$llz-0308-cyclopropyl-13c

28.343

11.227

7.580

C:\Users\zhang\Desktop\NMR\liu\processed\\$llz-0308-CH2CH2Ph-1h.fid
 new experiment
 Mar 25 2011
 USER:
 SOLVENT: cdcl3
 Experiment = s2pul
 Pulse length = 11.663 usec
 Relaxation delay = 4.800 sec
 NA = 16
 Solvent = cdcl3
 FID PTS1d = 20006
 PTS1d = 32768
 F1 = 399.950684 MHz
 F2 = 100.575279 MHz
 SW1 = 8002.40 Hz
 AT1 = 4.09 sec
 Hz per Pt 1stD = 0.24 Hz
 SW2 = 1.00 Hz
 Hz per Pt 2ndD = 1.00 Hz
 O1 = 2006.7504 Hz
 O2 = -0.5000 Hz
 LB1 = 0.00 Hz
 TP A = -56.23
 B = -32.58
 C = 0.00



new experiment

USER: -- DATE: Mar 25 2011

F1: 399.951	F2: 100.575	SW1: 8002		OF1: 2006.8		PTS1d: 20006 , 32768
EX: s2pul		PW: 11.7 us	PD: 4.8 sec	NA: 16	LB: 0.0	Nuts - \\$llz-0308-CH2CH2Ph-1h.fid

C:\Users\zhang\Desktop\NMR\liu\processed\\$11z0308-phCH2CH2-13c

STANDARD CARBON PARAMETERS

Mar 8 2011

USER:

SOLVENT: cdcl3

Experiment = s2pul

Pulse length = 6.000 usec

Relaxation delay = 3.000 sec

NA = 46

Solvent = cdcl3

FID PTS1d = 36749

PTS1d = 65536

F1 = 125.701683 MHz

F2 = 499.858551 MHz

SW1 = 28258.57 Hz

AT1 = 2.32 sec

Hz per Pt 1stD = 0.43 Hz

SW2 = 1.00 Hz

Hz per Pt 2ndD = 1.00 Hz

O1 = 12777.3203 Hz

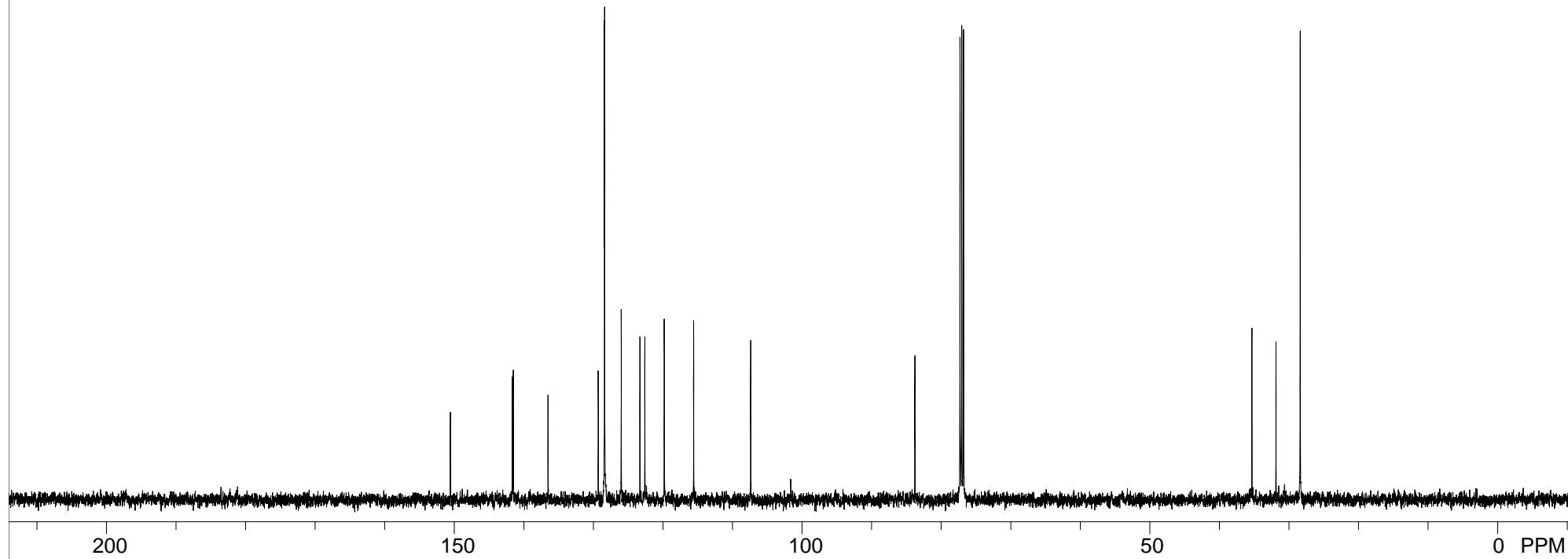
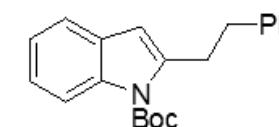
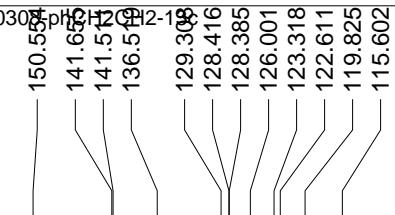
O2 = -0.5000 Hz

LB1 = 2.00 Hz

TP A = -217.97

B = 170.16

C = 0.00



STANDARD CARBON PARAMETERS

F1: 125.702

F2: 499.859

SW1: 28259

PW: 6.0 us

OF1: 12777.3

LB: 2.0

PTS1d: 36749, 65536

Nuts - \\$11z0308-phCH2CH2-13c

EX: s2pul

PD: 3.0 sec

NA: 46

User: -- DATE: Mar 8 2011

C:\Users\zhang\Desktop\NMR\llz0308-phCH2CH2-13c

STANDARD CARBON PARAMETERS

Mar 8 2011

USER:

SOLVENT: cdcl3

Experiment = s2pul

Pulse length = 6.000 usec

Relaxation delay = 3.000 sec

NA = 46

Solvent = cdcl3

FID PTS1d = 36749

PTS1d = 65536

F1 = 125.701683 MHz

F2 = 499.858551 MHz

SW1 = 28258.57 Hz

AT1 = 2.32 sec

Hz per Pt 1stD = 0.43 Hz

SW2 = 1.00 Hz

Hz per Pt 2ndD = 1.00 Hz

O1 = 12777.3203 Hz

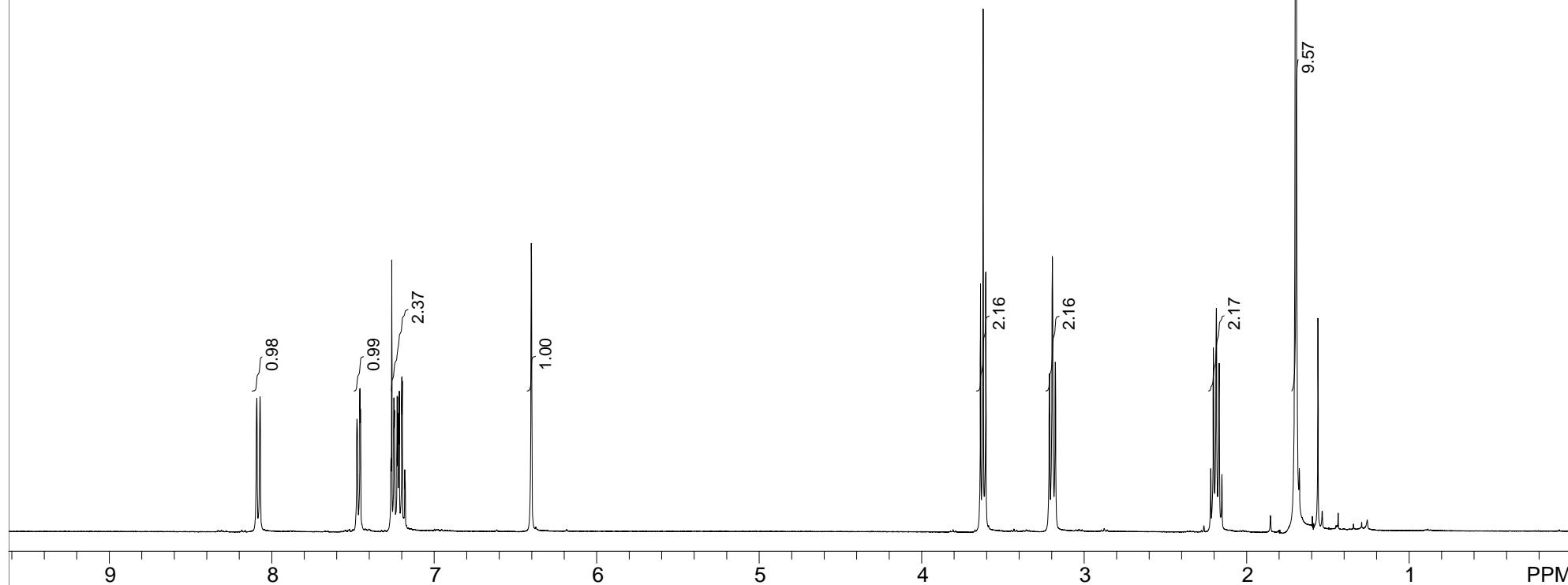
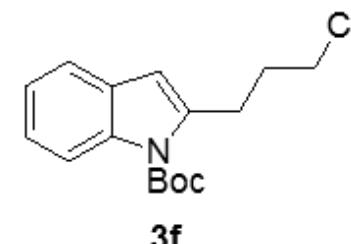
O2 = -0.5000 Hz

LB1 = 2.00 Hz

TP A = -217.97

B = 170.16

C = 0.00



new experiment

USER: -- DATE: Mar 27 2011

F1: 399.951	F2: 100.575	SW1: 8002		OF1: 2006.8		PTS1d: 20006 , 32768	
EX: s2pul		PW: 11.7 us	PD: 4.8 sec	NA: 16	LB: 0.0		Nuts - \$llz0307-C5-1h.fid

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STANDARD CARBON PARAMETERS

Mar 8 2011

USER:

SOLVENT: cdcl3

Experiment = s2pul

Pulse length = 6.000 usec

Relaxation delay = 3.000 sec

NA = 82

Solvent = cdcl3

FID PTS1d = 36749

PTS1d = 65536

F1 = 125.701683 MHz

F2 = 499.858551 MHz

SW1 = 28258.57 Hz

AT1 = 2.32 sec

Hz per Pt 1stD = 0.43 Hz

SW2 = 1.00 Hz

Hz per Pt 2ndD = 1.00 Hz

O1 = 12777.2832 Hz

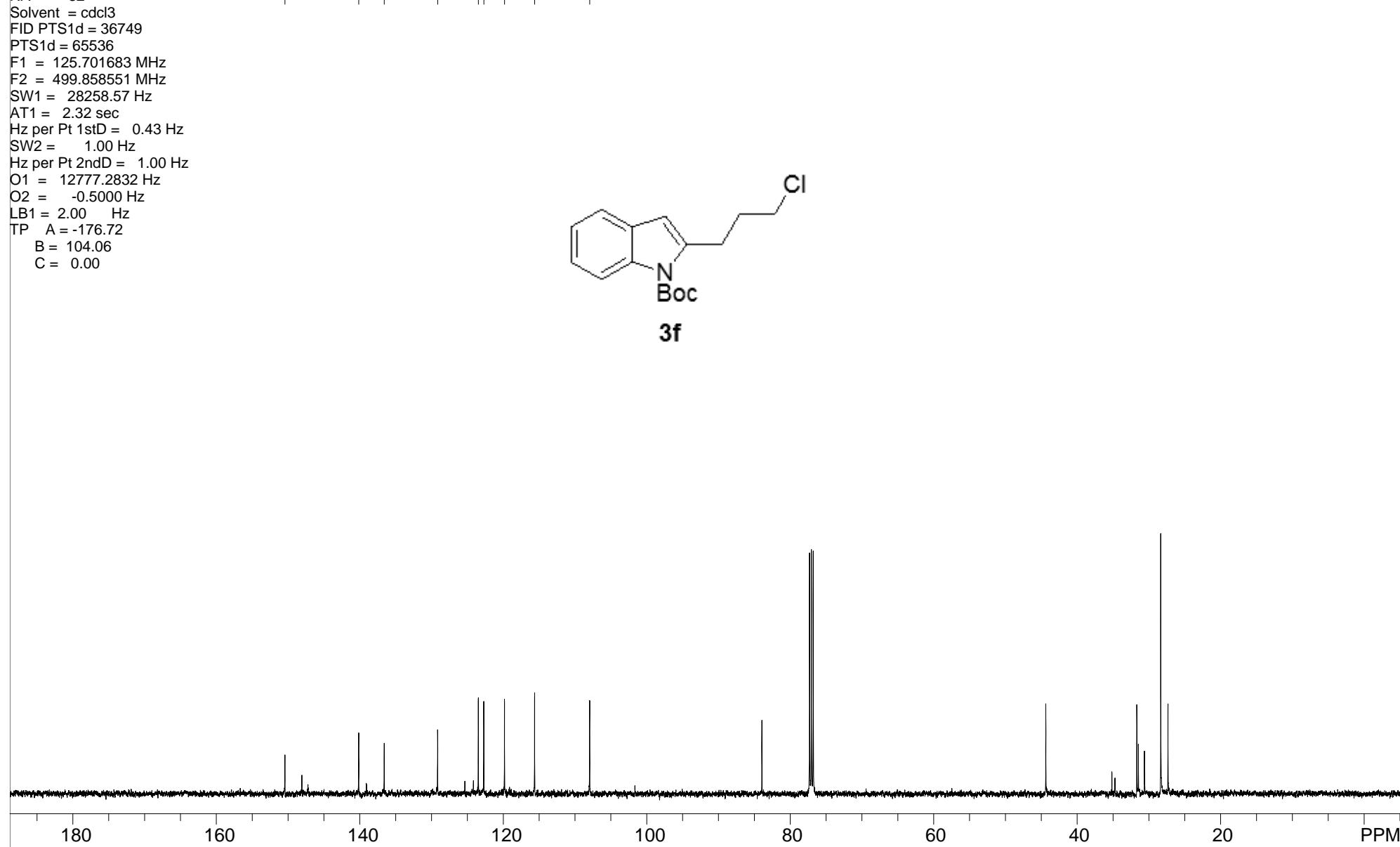
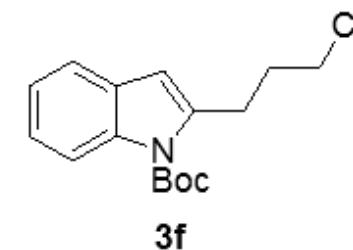
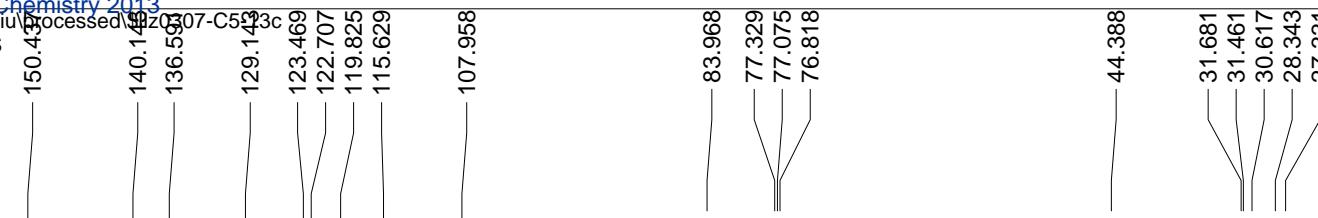
O2 = -0.5000 Hz

LB1 = 2.00 Hz

TP A = -176.72

B = 104.06

C = 0.00



STANDARD CARBON PARAMETERS

F1: 125.702

F2: 499.859

EX: s2pul

SW1: 28259

PW: 6.0 us

PD: 3.0 sec

OF1: 12777.3

NA: 82

LB: 2.0

USER: -- DATE: Mar 8 2011

PTS1d: 36749, 65536

Nuts - \\$llz0307-C5-13c

C:\Users\zhang\Desktop\NMR\lliu\processed\\$llz0307-C6-1h.fid

new experiment

Mar 27 2011

USER:

SOLVENT: cdcl3

Experiment = s2pul

Pulse length = 11.663 usec

Relaxation delay = 4.800 sec

NA = 16

Solvent = cdcl3

FID PTS1d = 20006

PTS1d = 32768

F1 = 399.950684 MHz

F2 = 100.575279 MHz

SW1 = 8002.40 Hz

AT1 = 4.09 sec

Hz per Pt 1stD = 0.24 Hz

SW2 = 1.00 Hz

Hz per Pt 2ndD = 1.00 Hz

O1 = 2006.7504 Hz

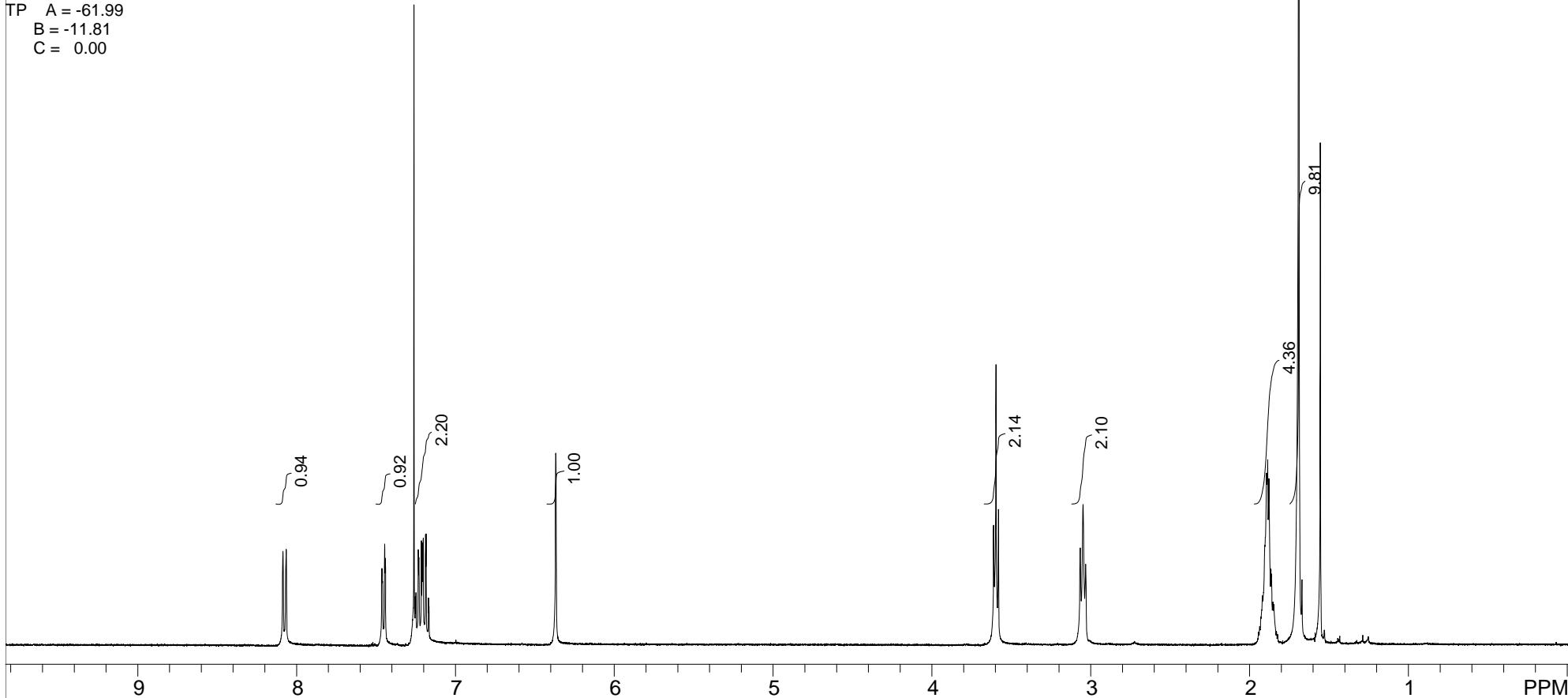
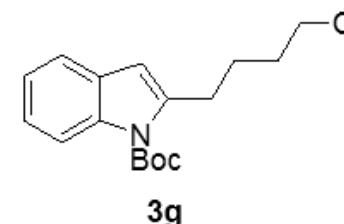
O2 = -0.5000 Hz

LB1 = 0.00 Hz

TP A = -61.99

B = -11.81

C = 0.00



new experiment

USER: -- DATE: Mar 27 2011

F1: 399.951	F2: 100.575	SW1: 8002		OF1: 2006.8		PTS1d: 20006 , 32768	Nuts - \\$llz0307-C6-1h.fid
EX: s2pul		PW: 11.7 us	PD: 4.8 sec	NA: 16	LB: 0.0		

C:\Users\zhang\Desktop\NMR\llz0307-C6-13c

STANDARD CARBON PARAMETERS

Mar 8 2011

USER:

SOLVENT: cdcl3

Experiment = s2pul

Pulse length = 6.000 usec

Relaxation delay = 3.000 sec

NA = 136

Solvent = cdcl3

FID PTS1d = 36749

PTS1d = 65536

F1 = 125.701683 MHz

F2 = 499.858551 MHz

SW1 = 28258.57 Hz

AT1 = 2.32 sec

Hz per Pt 1stD = 0.43 Hz

SW2 = 1.00 Hz

Hz per Pt 2ndD = 1.00 Hz

O1 = 12777.2832 Hz

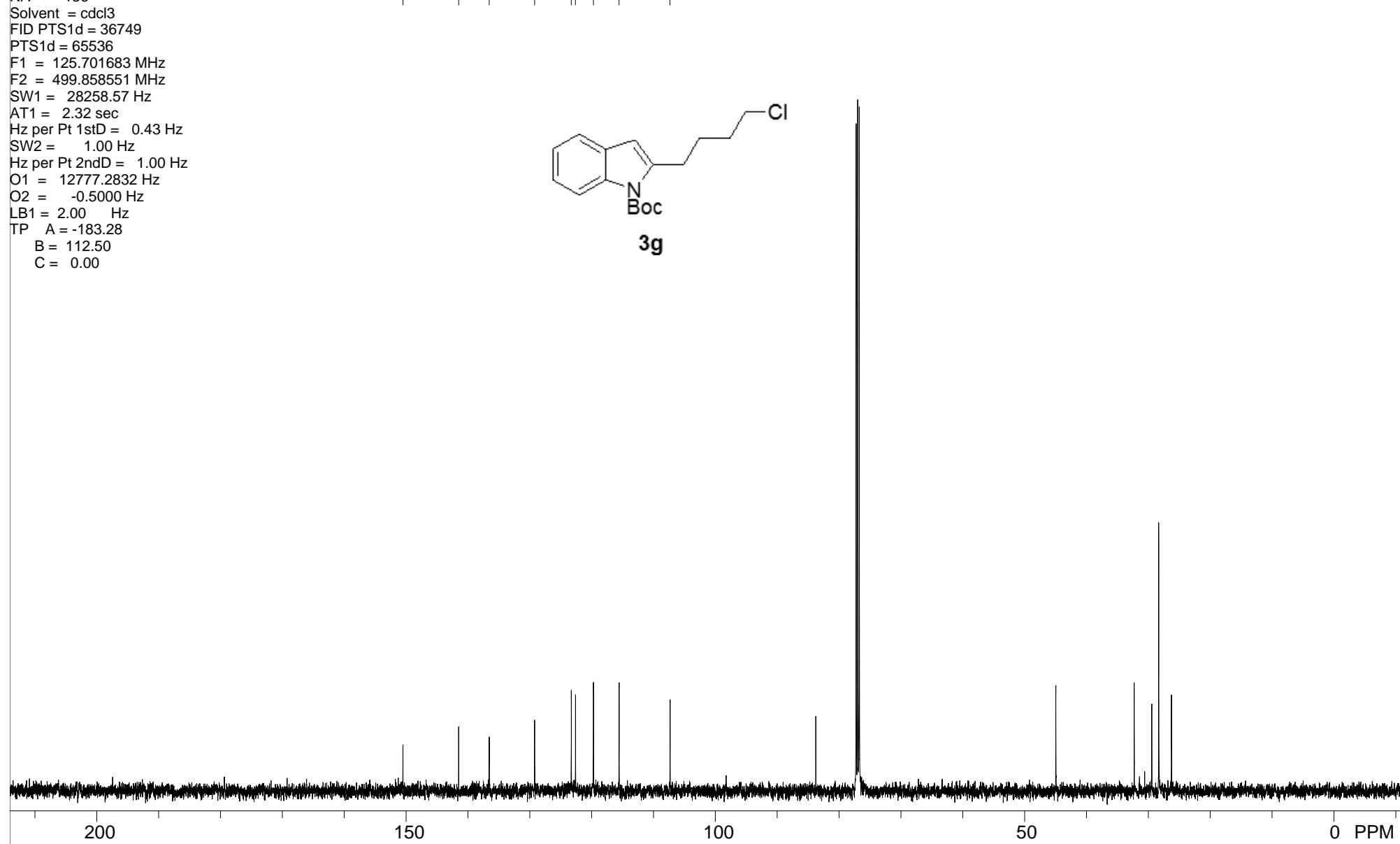
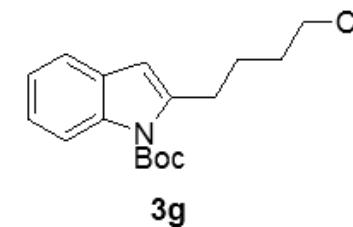
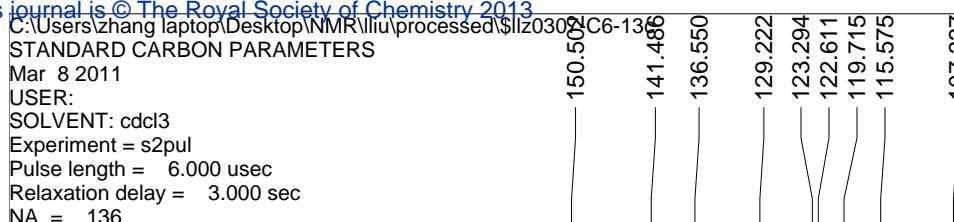
O2 = -0.5000 Hz

LB1 = 2.00 Hz

TP A = -183.28

B = 112.50

C = 0.00



STANDARD CARBON PARAMETERS

F1: 125.702	F2: 499.859	SW1: 28259		OF1: 12777.3		PTS1d: 36749 , 65536	USER: -- DATE: Mar 8 2011
EX: s2pul		PW: 6.0 us	PD: 3.0 sec	NA: 136	LB: 2.0		Nuts - \$llz0307-C6-13c

C:\Users\zhang\Desktop\NMR\liu\processed\\$llz-1-31-1h.fid

new experiment

Mar 10 2011

USER:

SOLVENT: cdcl3

Experiment = s2pul

Pulse length = 11.663 usec

Relaxation delay = 4.800 sec

NA = 12

Solvent = cdcl3

FID PTS1d = 20006

PTS1d = 32768

F1 = 399.950684 MHz

F2 = 100.575279 MHz

SW1 = 8002.40 Hz

AT1 = 4.09 sec

Hz per Pt 1stD = 0.24 Hz

SW2 = 1.00 Hz

Hz per Pt 2ndD = 1.00 Hz

O1 = 2006.7504 Hz

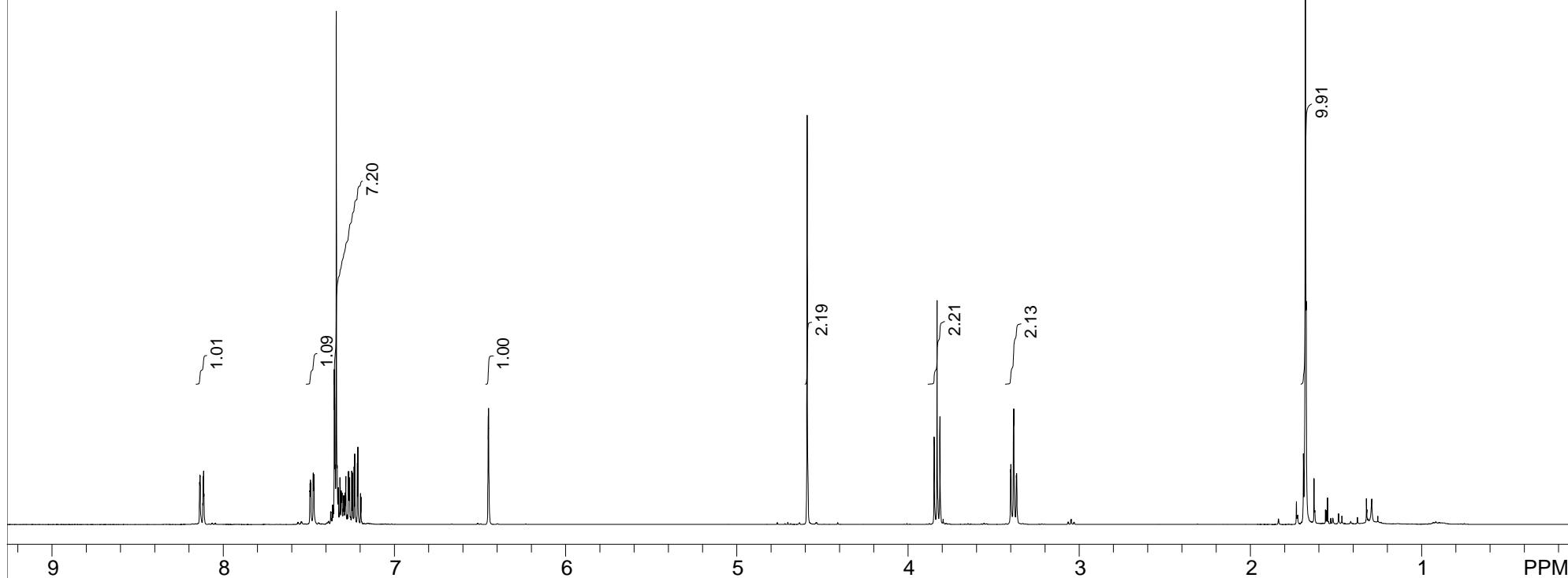
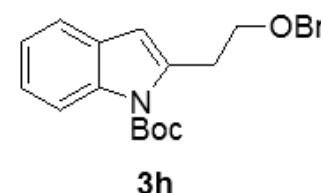
O2 = -0.5000 Hz

LB1 = 0.00 Hz

TP A = -45.30

B = -37.26

C = 0.00



new experiment

USER: -- DATE: Mar 10 2011

F1: 399.951	F2: 100.575	SW1: 8002		OF1: 2006.8		PTS1d: 20006 , 32768	
EX: s2pul		PW: 11.7 us		PD: 4.8 sec		NA: 12	LB: 0.0

Nuts - \\$llz-1-31-1h.fid

C:\Users\zhang\Desktop\NMR\lliu\processed\\$llz-131-13c.fid

Standard c13 run using qnp probe

Mar 10 2011

USER:

SOLVENT: cdcl3

Experiment = s2pul

Pulse length = 9.500 usec

Relaxation delay = 5.000 sec

NA = 100

Solvent = cdcl3

FID PTS1d = 16000

PTS1d = 16384

F1 = 100.576706 MHz

F2 = 399.949585 MHz

SW1 = 25000.00 Hz

AT1 = 0.66 sec

Hz per Pt 1stD = 1.53 Hz

SW2 = 1.00 Hz

Hz per Pt 2ndD = 1.00 Hz

O1 = 9525.1367 Hz

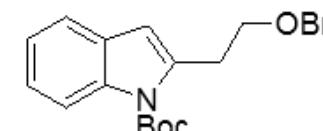
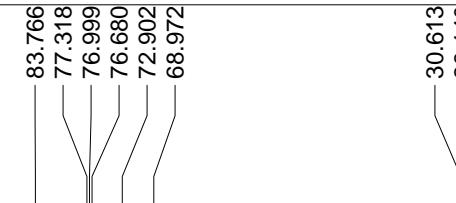
O2 = -0.5000 Hz

LB1 = 2.00 Hz

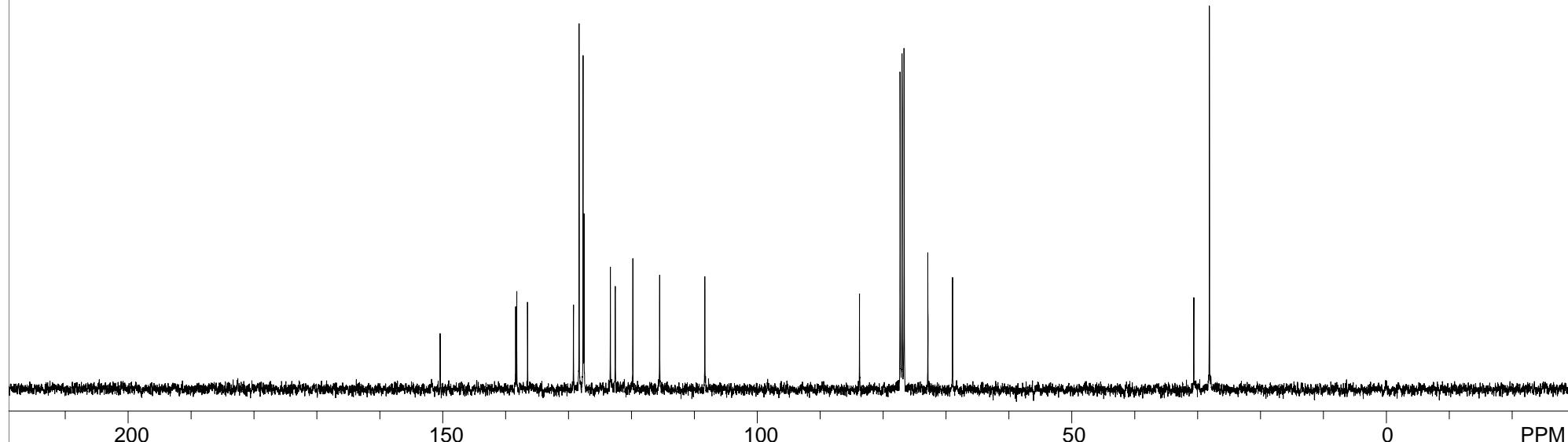
TP A = -134.06

B = 199.69

C = 0.00



3h



Standard c13 run using qnp probe

F1: 100.577 F2: 399.950

SW1: 25000

OF1: 9525.1

PTS1d: 16000 , 16384

EX: s2pul

PW: 9.5 us

PD:

5.0

sec

NA: 100

LB: 2.0

Nuts - \\$llz-1-31-13c.fid

USER: -- DATE: Mar 10 2011

C:\Users\zhang\Desktop\NMR\liu\processed\\$llz-1-28-1h.fid

new experiment

Mar 10 2011

USER:

SOLVENT: cdcl3

Experiment = s2pul

Pulse length = 11.663 usec

Relaxation delay = 4.800 sec

NA = 10

Solvent = cdcl3

FID PTS1d = 20006

PTS1d = 32768

F1 = 399.950684 MHz

F2 = 100.575279 MHz

SW1 = 8002.40 Hz

AT1 = 4.09 sec

Hz per Pt 1stD = 0.24 Hz

SW2 = 1.00 Hz

Hz per Pt 2ndD = 1.00 Hz

O1 = 1968.7904 Hz

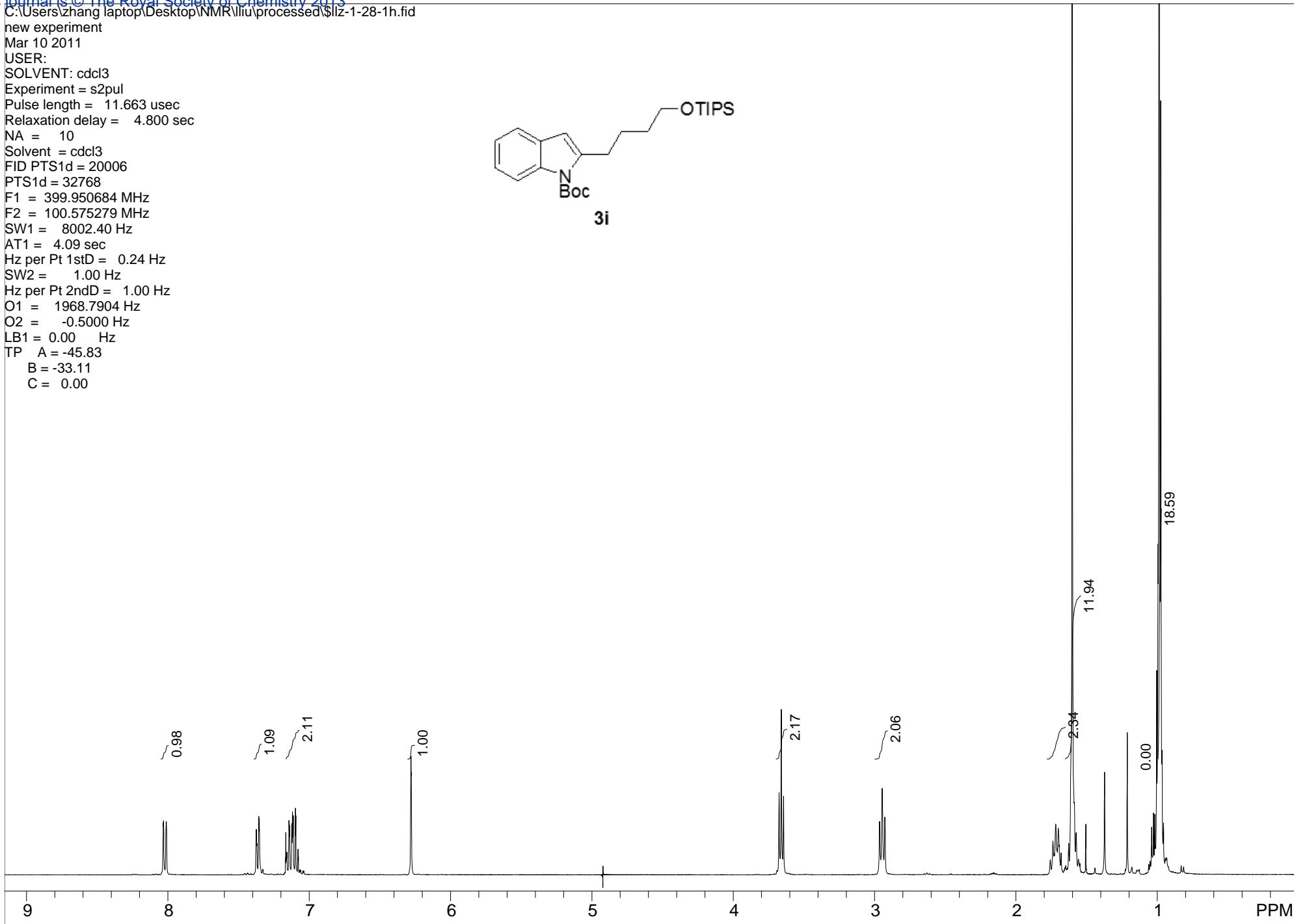
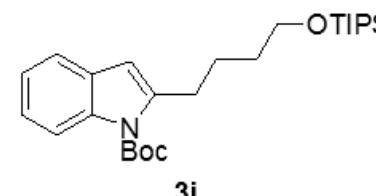
O2 = -0.5000 Hz

LB1 = 0.00 Hz

TP A = -45.83

B = -33.11

C = 0.00

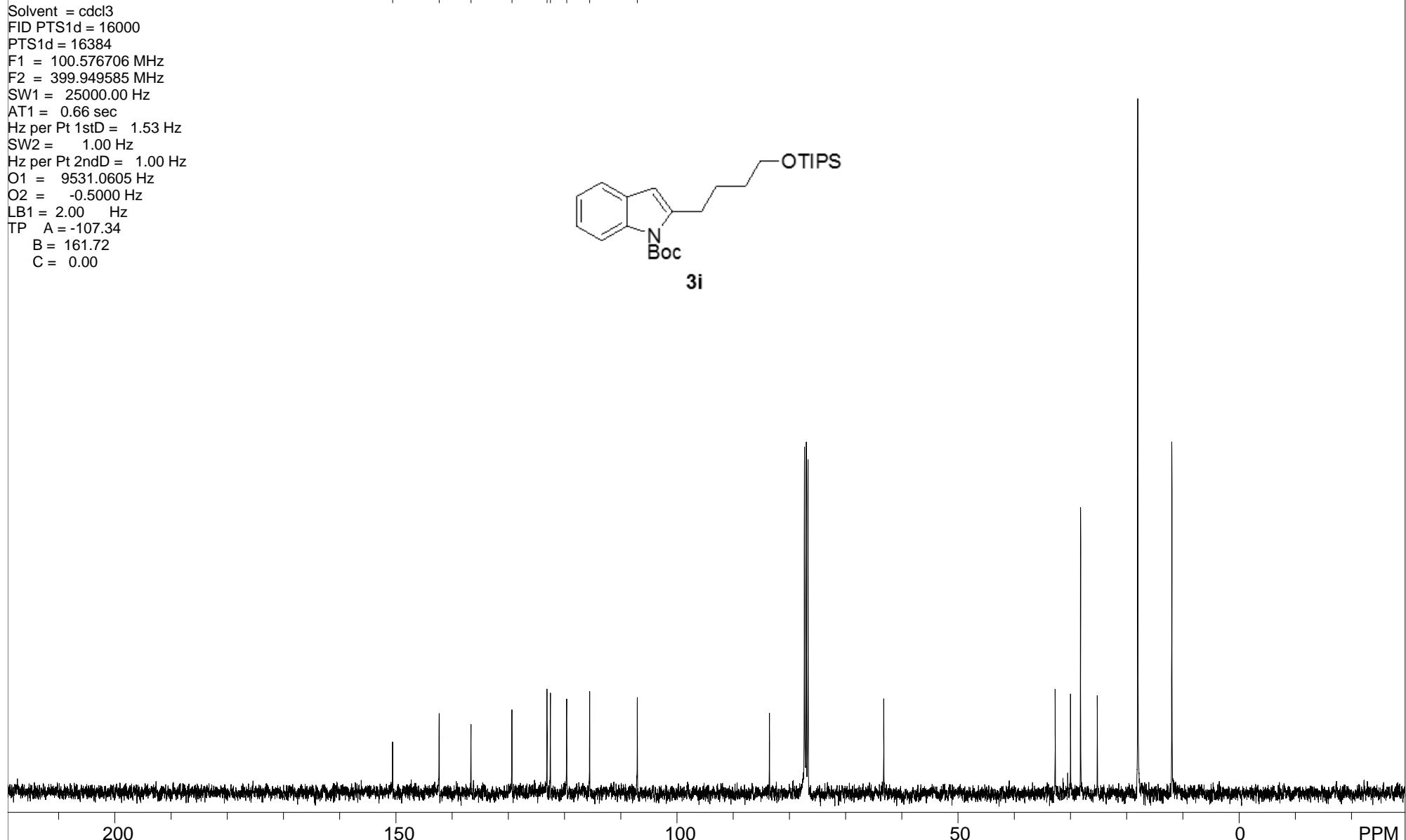
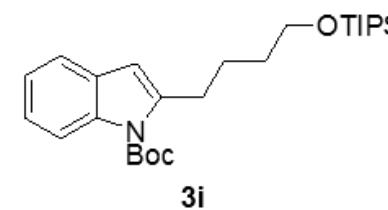


new experiment

USER: -- DATE: Mar 10 2011

F1: 399.951	F2: 100.575	SW1: 8002		OF1: 1968.8		PTS1d: 20006 , 32768		Nuts - \\$llz-1-28-1h.fid
EX: s2pul		PW: 11.7 us		PD: 4.8 sec		NA: 10		

C:\Users\zhang\Desktop\NMR\lliu\processed\\$llz-1-28-13c.fid
 Standard c13 run using qnp probe
 Mar 10 2011
 USER:
 SOLVENT: cdcl3
 Experiment = s2pul
 Pulse length = 9.500 usec
 Relaxation delay = 5.000 sec
 NA = 124
 Solvent = cdcl3
 FID PTS1d = 16000
 PTS1d = 16384
 F1 = 100.576706 MHz
 F2 = 399.949585 MHz
 SW1 = 25000.00 Hz
 AT1 = 0.66 sec
 Hz per Pt 1stD = 1.53 Hz
 SW2 = 1.00 Hz
 Hz per Pt 2ndD = 1.00 Hz
 O1 = 9531.0605 Hz
 O2 = -0.5000 Hz
 LB1 = 2.00 Hz
 TP A = -107.34
 B = 161.72
 C = 0.00



Standard c13 run using qnp probe

USER: -- DATE: Mar 10 2011

F1: 100.577	F2: 399.950	SW1: 25000		OF1: 9531.1		PTS1d: 16000 , 16384	
EX: s2pul		PW: 9.5 us	PD: 5.0 sec	NA: 124	LB: 2.0		Nuts - \\$llz-1-28-13c.fid

C:\Users\zhang\Desktop\NMR\liu\processed\\$llz-1-37-1h.fid

new experiment

Mar 27 2011

USER:

SOLVENT: cdcl3

Experiment = s2pul

Pulse length = 11.663 usec

Relaxation delay = 4.800 sec

NA = 16

Solvent = cdcl3

FID PTS1d = 20006

PTS1d = 32768

F1 = 399.950684 MHz

F2 = 100.575279 MHz

SW1 = 8002.40 Hz

AT1 = 4.09 sec

Hz per Pt 1stD = 0.24 Hz

SW2 = 1.00 Hz

Hz per Pt 2ndD = 1.00 Hz

O1 = 2006.7504 Hz

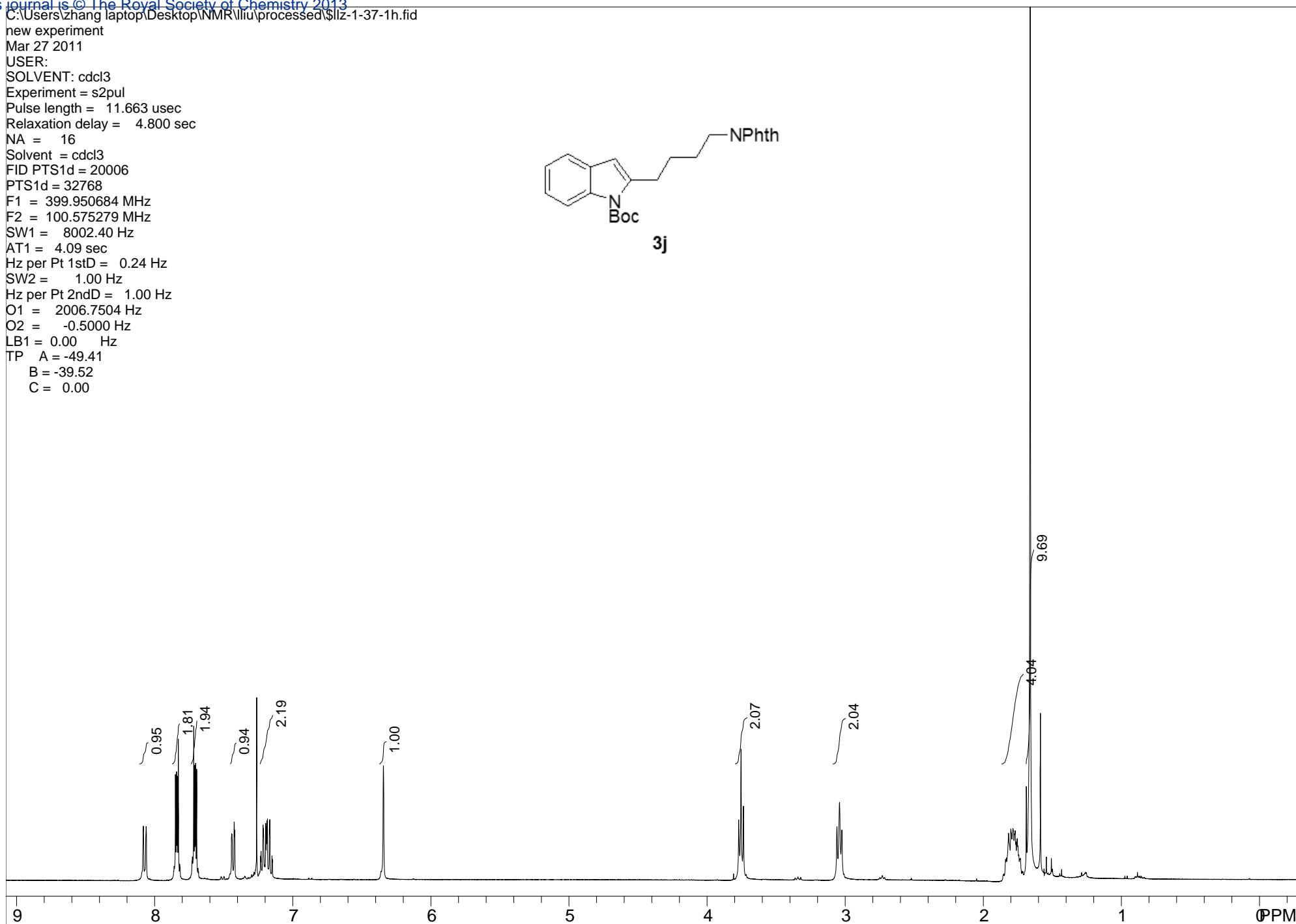
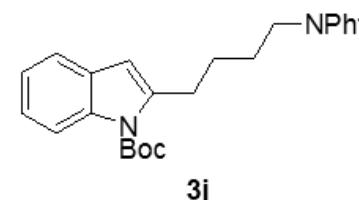
O2 = -0.5000 Hz

LB1 = 0.00 Hz

TP A = -49.41

B = -39.52

C = 0.00



new experiment

USER: -- DATE: Mar 27 2011

F1: 399.951	F2: 100.575	SW1: 8002		OF1: 2006.8		PTS1d: 20006 , 32768	
EX: s2pul		PW: 11.7 us		PD: 4.8 sec		NA: 16	Nuts - \\$llz-1-37-1h.fid

C:\Users\zhang\Desktop\NMR\llz-1-37-13c.fid

STANDARD CARBON PARAMETERS

Mar 14 2011

USER:

SOLVENT: cdcl3

Experiment = s2pul

Pulse length = 6.000 usec

Relaxation delay = 3.000 sec

NA = 64

Solvent = cdcl3

FID PTS1d = 36749

PTS1d = 65536

F1 = 125.701683 MHz

F2 = 499.858551 MHz

SW1 = 28258.57 Hz

AT1 = 2.32 sec

Hz per Pt 1stD = 0.43 Hz

SW2 = 1.00 Hz

Hz per Pt 2ndD = 1.00 Hz

O1 = 12777.2832 Hz

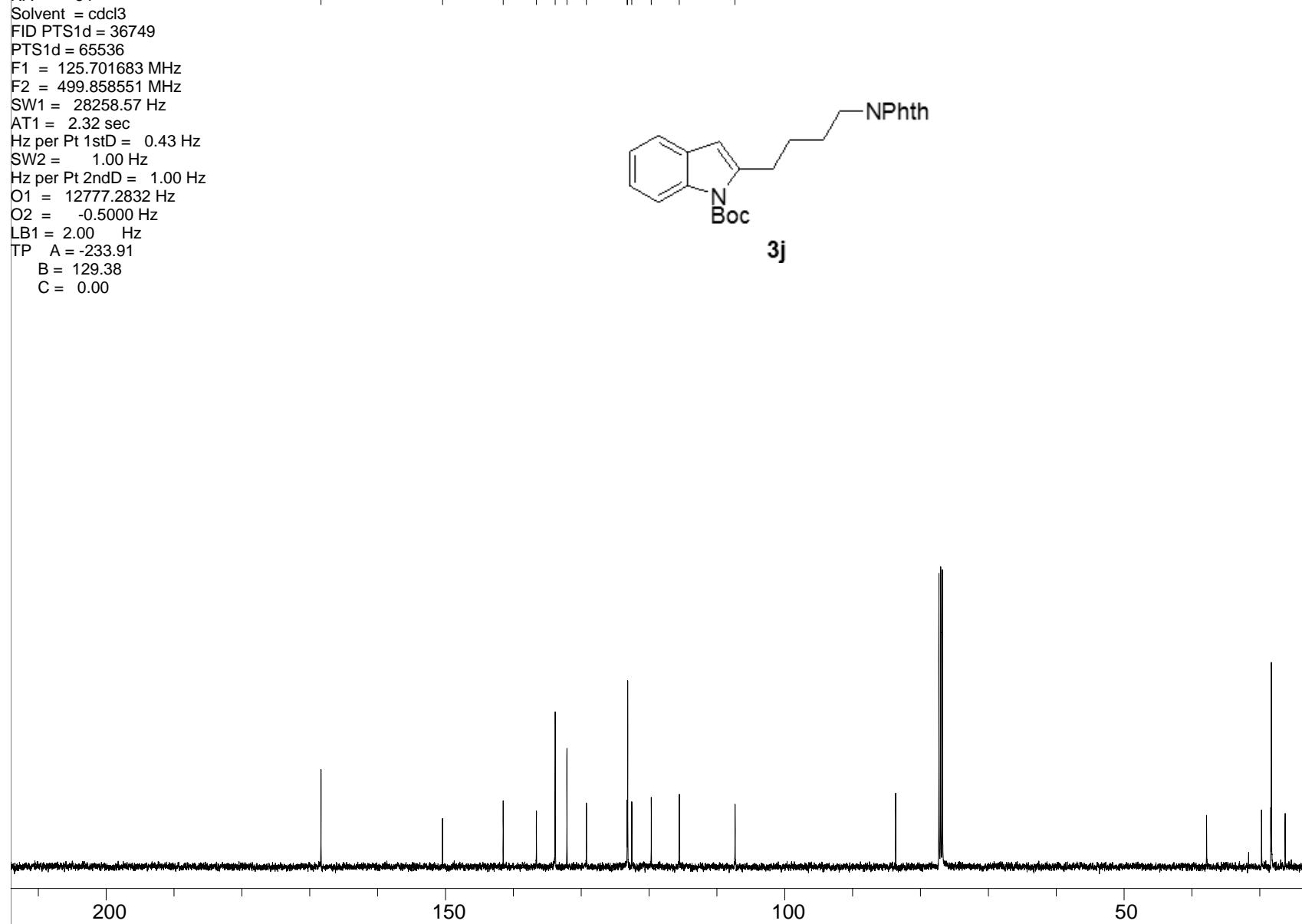
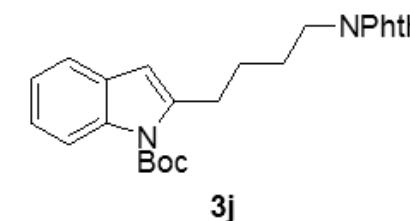
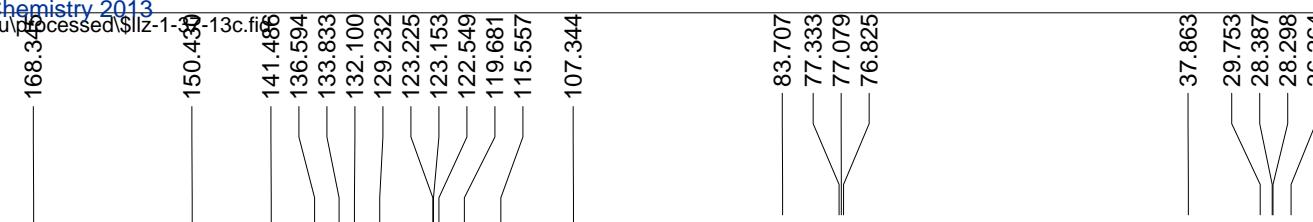
O2 = -0.5000 Hz

LB1 = 2.00 Hz

TP A = -233.91

B = 129.38

C = 0.00



STANDARD CARBON PARAMETERS

F1: 125.702

F2: 499.859

SW1: 28259

EX: s2pul

PW: 6.0 us

PD: 3.0 sec

NA: 64

OF1: 12777.3

LB: 2.0

PTS1d: 36749, 65536

USER: -- DATE: Mar 14 2011

Nuts - \$llz-1-37-13c.fid

C:\Users\zhang\Desktop\NMR\lliu\processed\\$llz-1-38-1h.fid

H1_CDCL3
Mar 13 2011

USER:

SOLVENT: CDCl3

Experiment = s2pul

Pulse length = 7.700 usec

Relaxation delay = 4.800 sec

NA = 7

Solvent = CDCl3

FID PTS1d = 21259

PTS1d = 32768

F1 = 499.858551 MHz

F2 = 1.000000 MHz

SW1 = 8503.40 Hz

AT1 = 3.85 sec

Hz per Pt 1stD = 0.26 Hz

SW2 = 1.00 Hz

Hz per Pt 2ndD = 1.00 Hz

O1 = 2499.2932 Hz

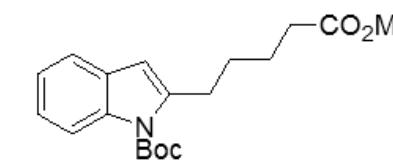
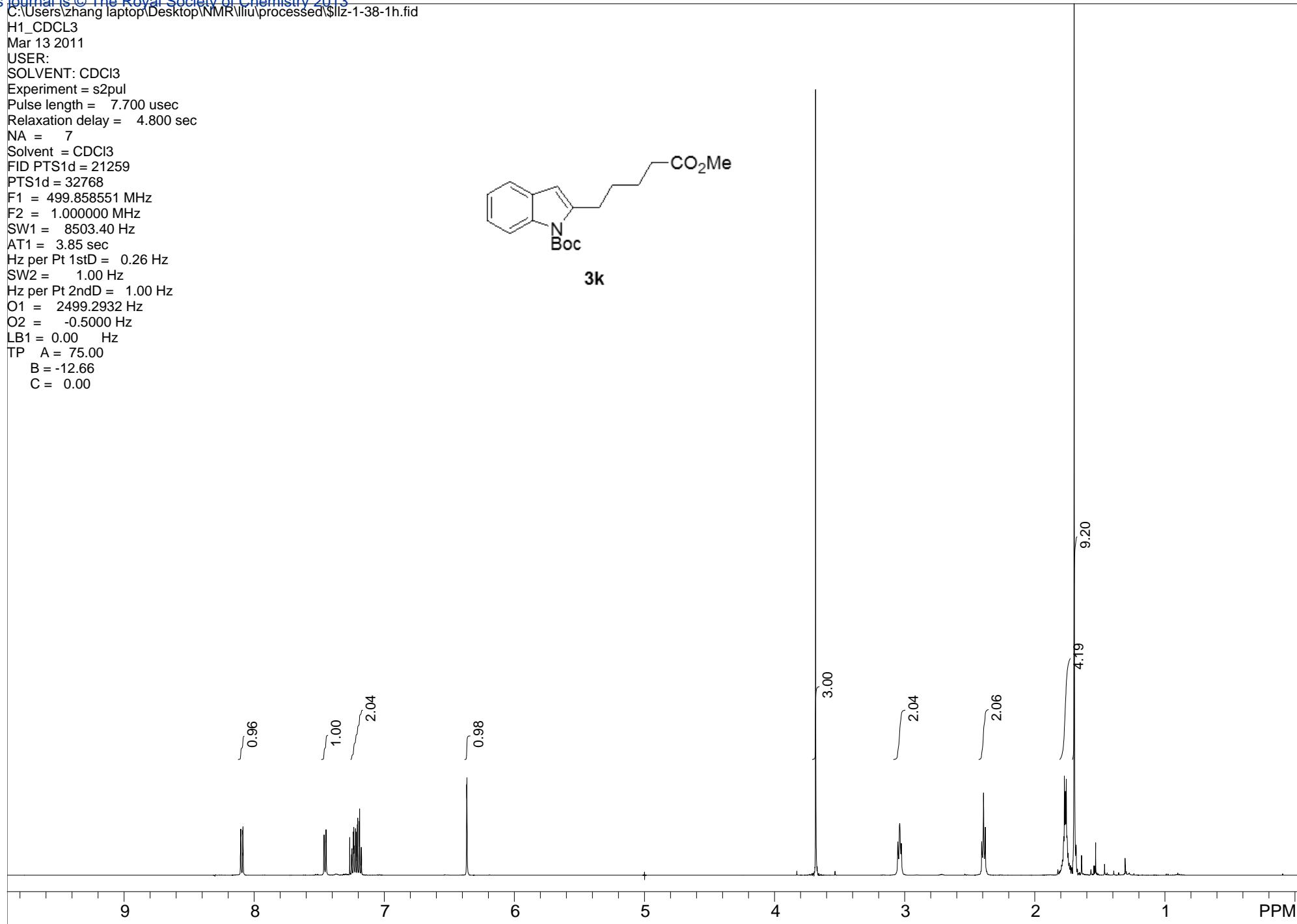
O2 = -0.5000 Hz

LB1 = 0.00 Hz

TP A = 75.00

B = -12.66

C = 0.00

**3k**

H1_CDCL3

USER: -- DATE: Mar 13 2011					
F1: 499.859	F2: 1.000	SW1: 8503		OF1: 2499.3	
EX: s2pul		PW: 7.7 us	PD: 4.8 sec	NA: 7	LB: 0

Nuts - \\$llz-1-38-1h.fid

C:\Users\zhang\Desktop\NMR\unprocessed\\$llz-1-38-13c.fid

STANDARD CARBON PARAMETERS

Mar 13 2011

USER:

SOLVENT: cdcl3

Experiment = s2pul

Pulse length = 6.000 usec

Relaxation delay = 3.000 sec

NA = 42

Solvent = cdcl3

FID PTS1d = 36749

PTS1d = 65536

F1 = 125.701683 MHz

F2 = 499.858551 MHz

SW1 = 28258.57 Hz

AT1 = 2.32 sec

Hz per Pt 1stD = 0.43 Hz

SW2 = 1.00 Hz

Hz per Pt 2ndD = 1.00 Hz

O1 = 12777.2832 Hz

O2 = -0.5000 Hz

LB1 = 2.00 Hz

TP A = -213.28

B = 105.47

C = 0.00

1732

150.498

141.720

136.570

129.273

123.215

122.566

119.678

115.547

107.217

83.710

77.322

77.069

76.815

51.555

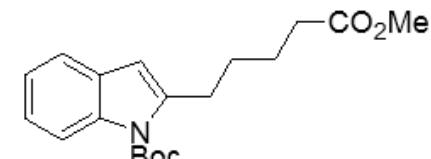
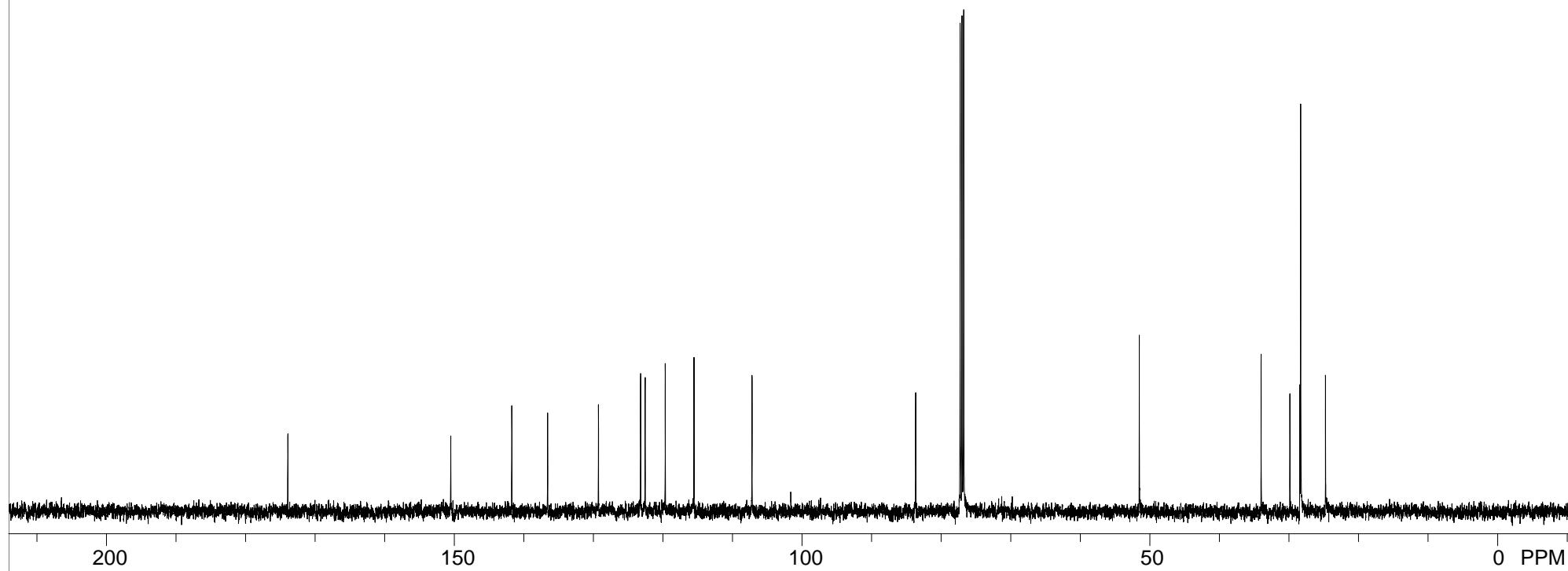
34.017

29.869

28.446

28.322

24.747

**3k**

STANDARD CARBON PARAMETERS

F1: 125.702

F2: 499.859

EX: s2pul

SW1: 28259

PW: 6.0 us

PD: 3.0 sec

OF1: 12777.3

NA: 42

LB: 2.0

PTS1d: 36749 , 65536

Nuts - \\$llz-1-38-13c.fid

USER: -- DATE: Mar 13 2011

C:\Users\zhang\Desktop\NMR\liu\processed\\$llz-1-34-1h.fid

new experiment

Mar 25 2011

USER:

SOLVENT: cdcl3

Experiment = s2pul

Pulse length = 11.663 usec

Relaxation delay = 4.800 sec

NA = 16

Solvent = cdcl3

FID PTS1d = 20006

PTS1d = 32768

F1 = 399.950684 MHz

F2 = 100.575279 MHz

SW1 = 8002.40 Hz

AT1 = 4.09 sec

Hz per Pt 1stD = 0.24 Hz

SW2 = 1.00 Hz

Hz per Pt 2ndD = 1.00 Hz

O1 = 2006.7162 Hz

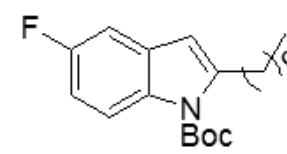
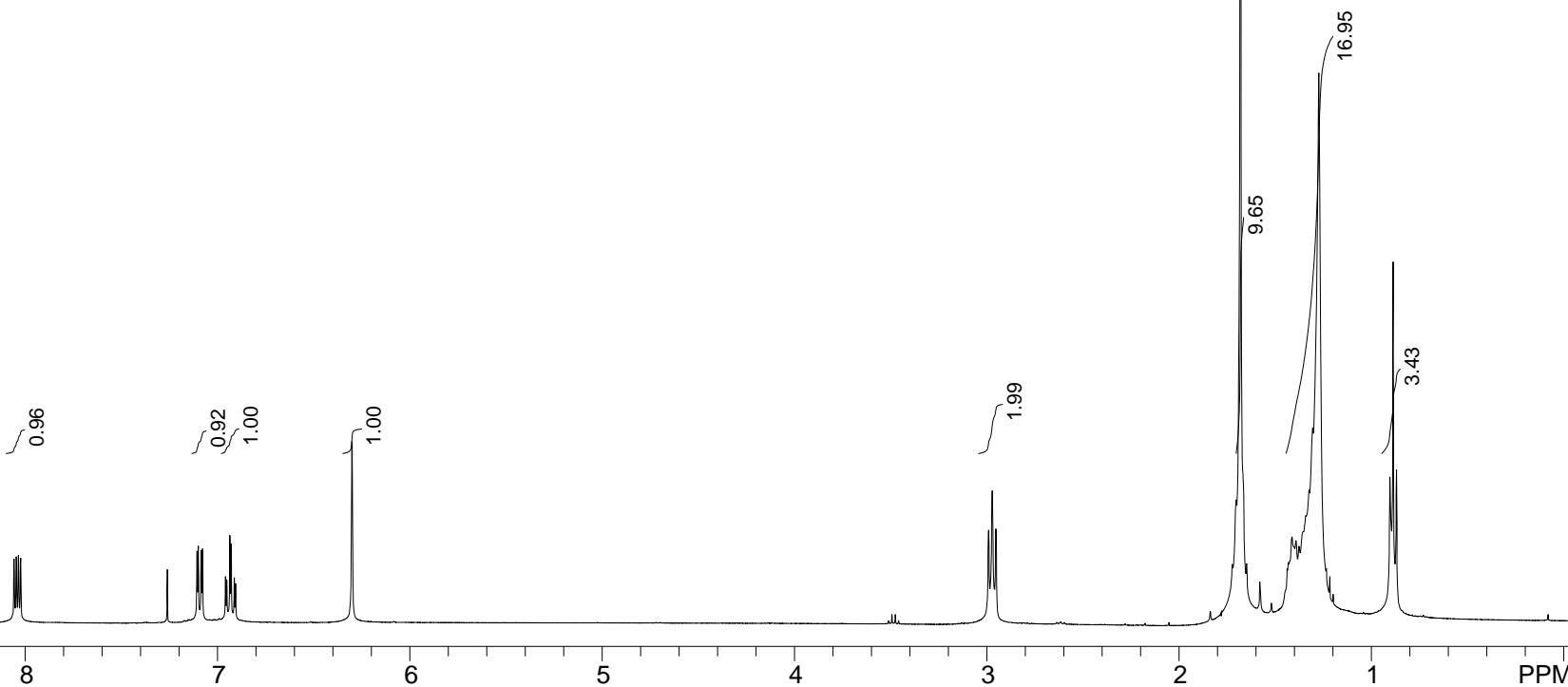
O2 = -0.5000 Hz

LB1 = 0.00 Hz

TP A = -60.25

B = -28.61

C = 0.00

**3l**

new experiment

USER: -- DATE: Mar 25 2011

F1: 399.951	F2: 100.575	SW1: 8002		OF1: 2006.7		PTS1d: 20006 , 32768	
EX: s2pul		PW: 11.7 us		PD: 4.8 sec		NA: 16	Nuts - \\$llz-1-34-1h.fid

C:\Users\zhang\Desktop\NMR\llz-1-34-13c.fid

STANDARD CARBON PARAMETERS

Mar 11 2011

USER:

SOLVENT: cdcl3

Experiment = s2pul

Pulse length = 6.000 usec

Relaxation delay = 3.000 sec

NA = 79

Solvent = cdcl3

FID PTS1d = 36749

PTS1d = 65536

F1 = 125.701683 MHz

F2 = 499.858551 MHz

SW1 = 28258.57 Hz

AT1 = 2.32 sec

Hz per Pt 1stD = 0.43 Hz

SW2 = 1.00 Hz

Hz per Pt 2ndD = 1.00 Hz

O1 = 12777.2832 Hz

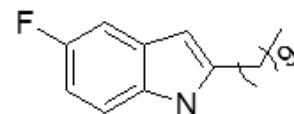
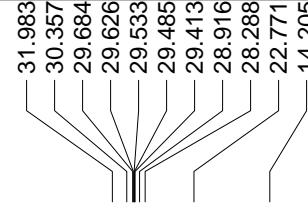
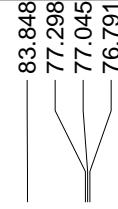
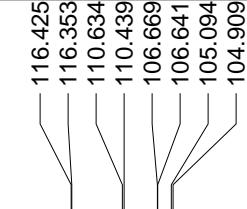
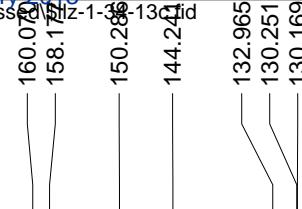
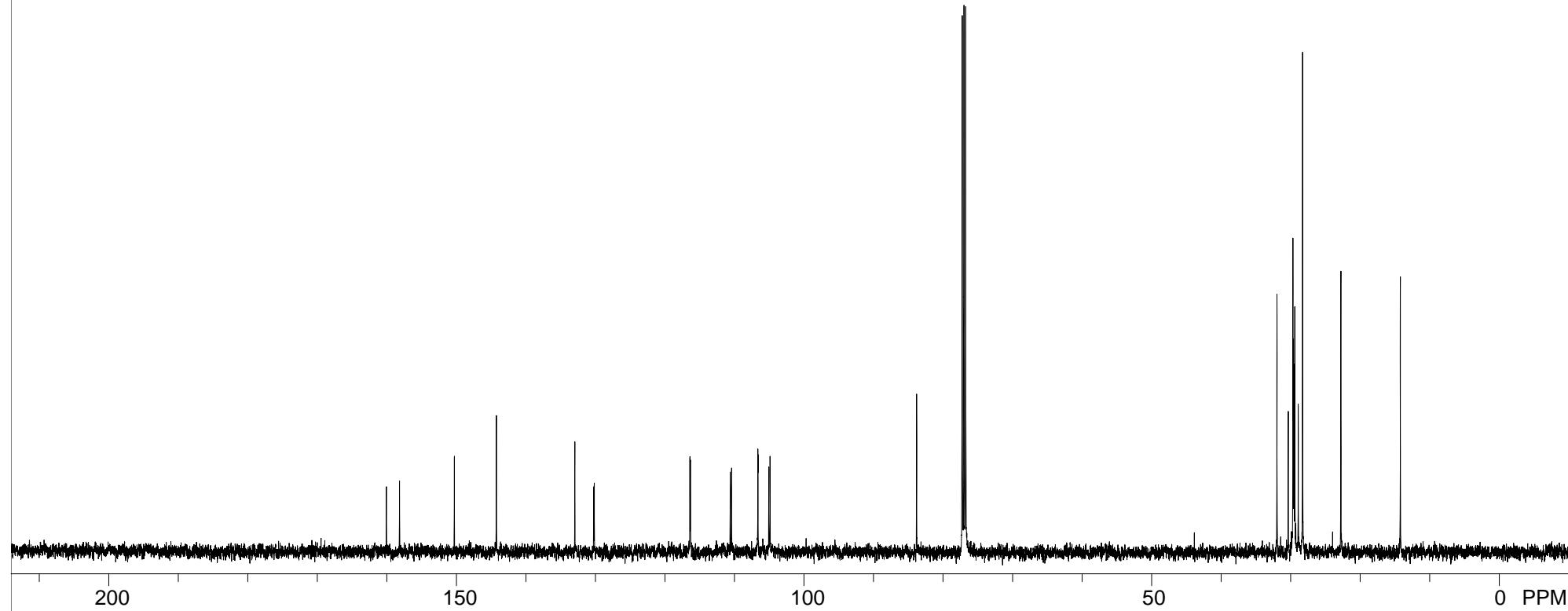
O2 = -0.5000 Hz

LB1 = 2.00 Hz

TP A = -255.00

B = 157.50

C = 0.00

**3l**

STANDARD CARBON PARAMETERS

F1: 125.702

F2: 499.859

SW1: 28259

EX: s2pul

PW: 6.0 us

PD: 3.0 sec

NA: 79

OF1: 12777.3

LB: 2.0

PTS1d: 36749

, 65536

USER: -- DATE: Mar 11 2011

Nuts - \$llz-1-34-13c.fid

C:\Users\zhang\Desktop\NMR\liu\processed\\$llz-1-58-1h.fid

new experimentU

Mar 22 2011

USER:

SOLVENT: cdcl3

Experiment = s2pul

Pulse length = 11.663 usec

Relaxation delay = 4.800 sec

NA = 16

Solvent = cdcl3

FID PTS1d = 20006

PTS1d = 32768

F1 = 399.950684 MHz

F2 = 100.575279 MHz

SW1 = 8002.40 Hz

AT1 = 2.50 sec

Hz per Pt 1stD = 0.24 Hz

SW2 = 1.00 Hz

Hz per Pt 2ndD = 1.00 Hz

O1 = 2006.2362 Hz

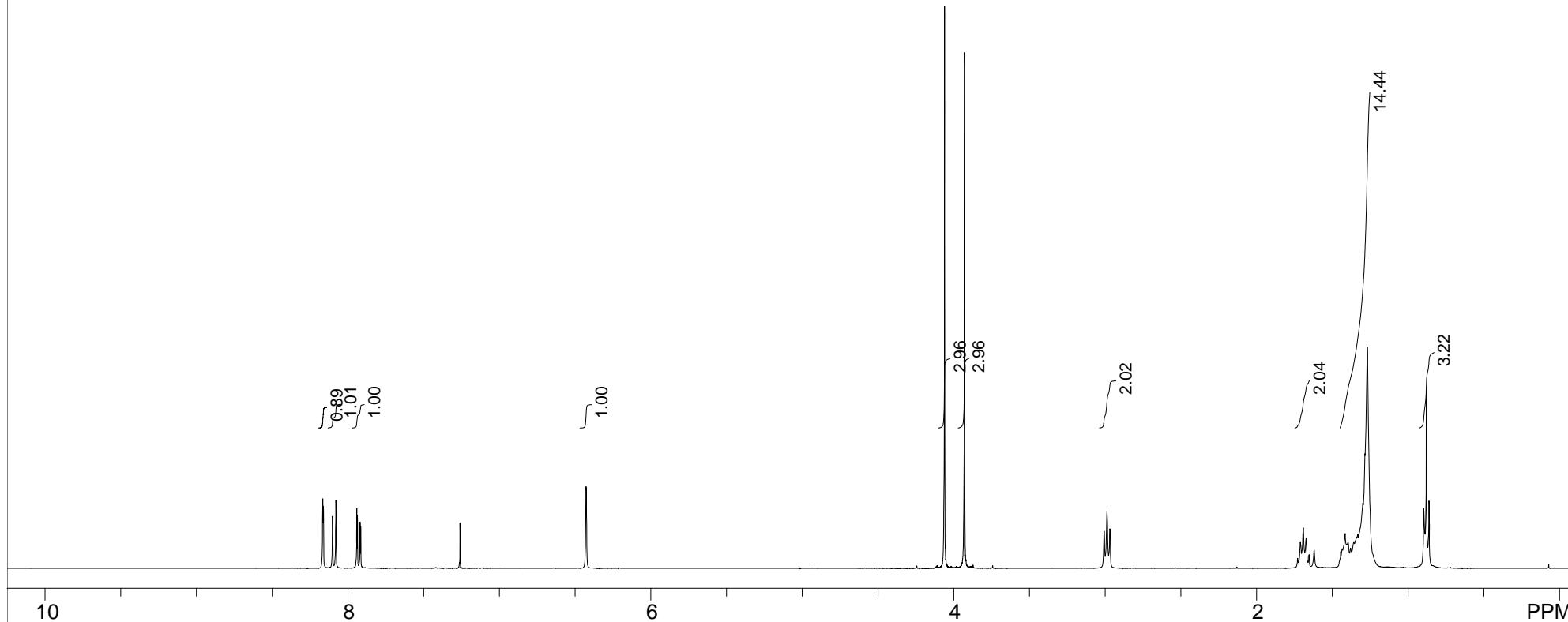
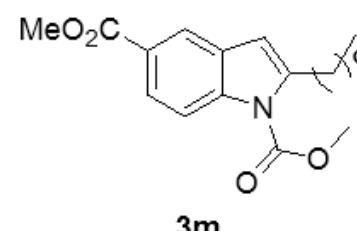
O2 = -0.5000 Hz

LB1 = 0.00 Hz

TP A = -48.69

B = -30.78

C = 0.00



new experimentU

USER: -- DATE: Mar 22 2011

F1: 399.951	F2: 100.575	SW1: 8002		OF1: 2006.2		PTS1d: 20006 , 32768	
EX: s2pul		PW: 11.7 us		PD: 4.8 sec	NA: 16	LB: 0.0	Nuts - \\$llz-1-58-1h.fid

C:\Users\zhang\Desktop\NMR\nuts\DATA\\$llz-1-58-13c.fid

Standard c13 run using qnp probe

Mar 22 2011

USER:

SOLVENT: cdcl3

Experiment = s2pul

Pulse length = 9.500 usec

Relaxation delay = 5.000 sec

NA = 100

Solvent = cdcl3

FID PTS1d = 16000

PTS1d = 16384

F1 = 100.576706 MHz

F2 = 399.949585 MHz

SW1 = 25000.00 Hz

AT1 = 0.64 sec

Hz per Pt 1stD = 1.53 Hz

SW2 = 1.00 Hz

Hz per Pt 2ndD = 1.00 Hz

O1 = 9531.0605 Hz

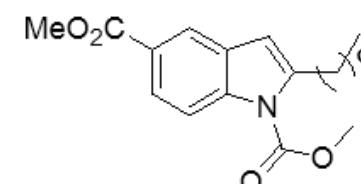
O2 = -0.5000 Hz

LB1 = 2.00 Hz

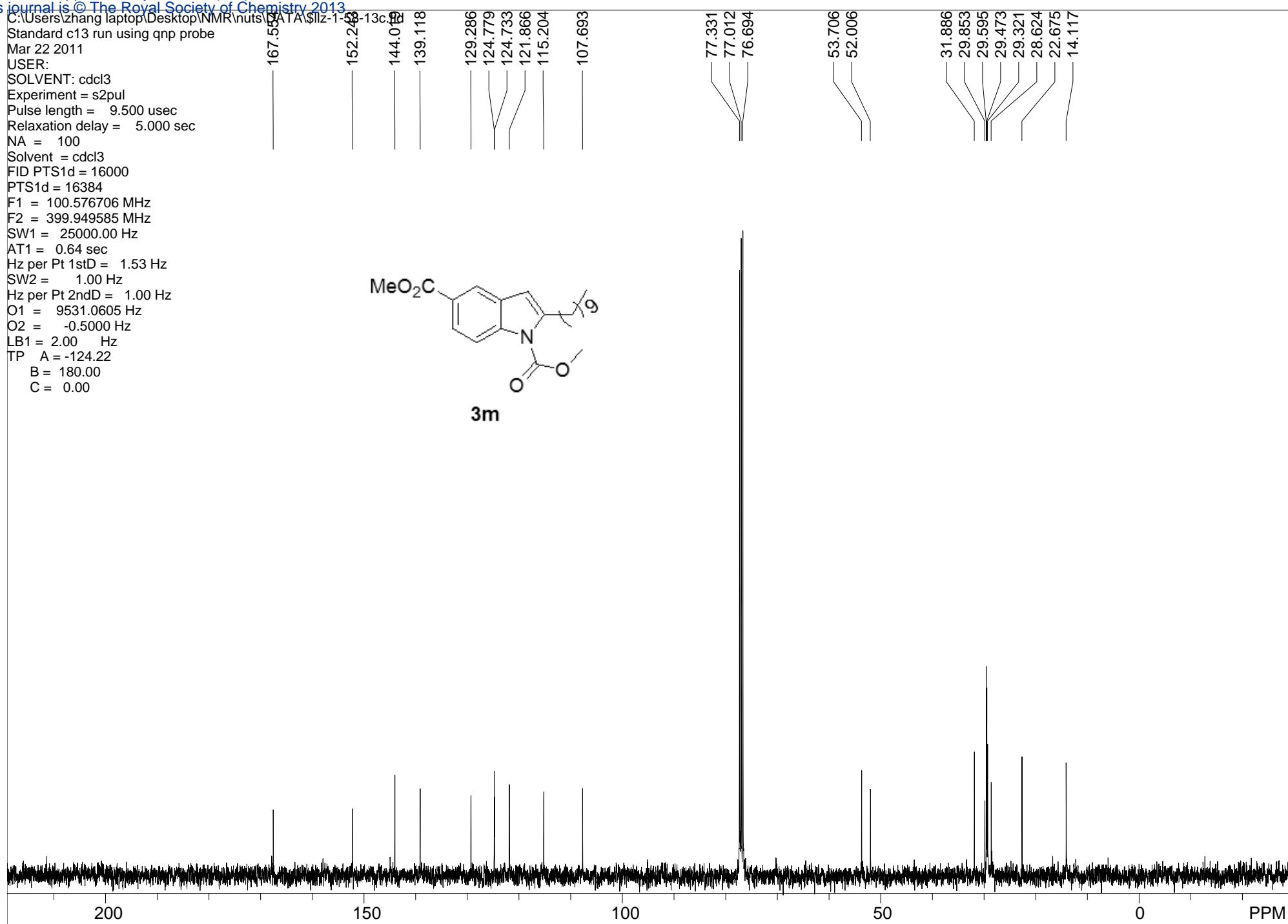
TP A = -124.22

B = 180.00

C = 0.00



3m



Standard c13 run using qnp probe

F1: 100.577

F2: 399.950

SW1: 25000

OF1: 9531.1

PTS1d: 16000 , 16384

EX: s2pul

PW: 9.5 us

PD: 5.0 sec

NA: 100

LB: 2.0

Nuts - \\$llz-1-58-13c.fid

USER: -- DATE: Mar 22 2011

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C:\Users\zhang laptop\Desktop\NMR\liu\processed\\$liz-1-33-1h.fid

new experiment

Mar 25 2011

USER:

SOLVENT: cdcl3

Experiment = s2pul

Pulse length = 11.663 usec

Relaxation delay = 4.800 sec

NA = 10

Solvent = cdcl3

FID PTS1d = 20006

PTS1d = 32768

F1 = 399.950684

F2 = 100.575279 MHz

SW1 = 8002.40 Hz

$$AT1 = 4.09 \text{ sec}$$

Hz per Pt 1stD =

SW2 = 1.00 H

Hz per Pt 2ndD = 1.00 Hz

O1 = 2006.750

O2 = -0.5000 Hz

LB1 = 0.00 Hz

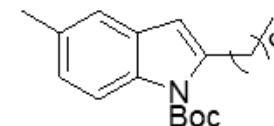
TP A = -45.56

B = -40.11

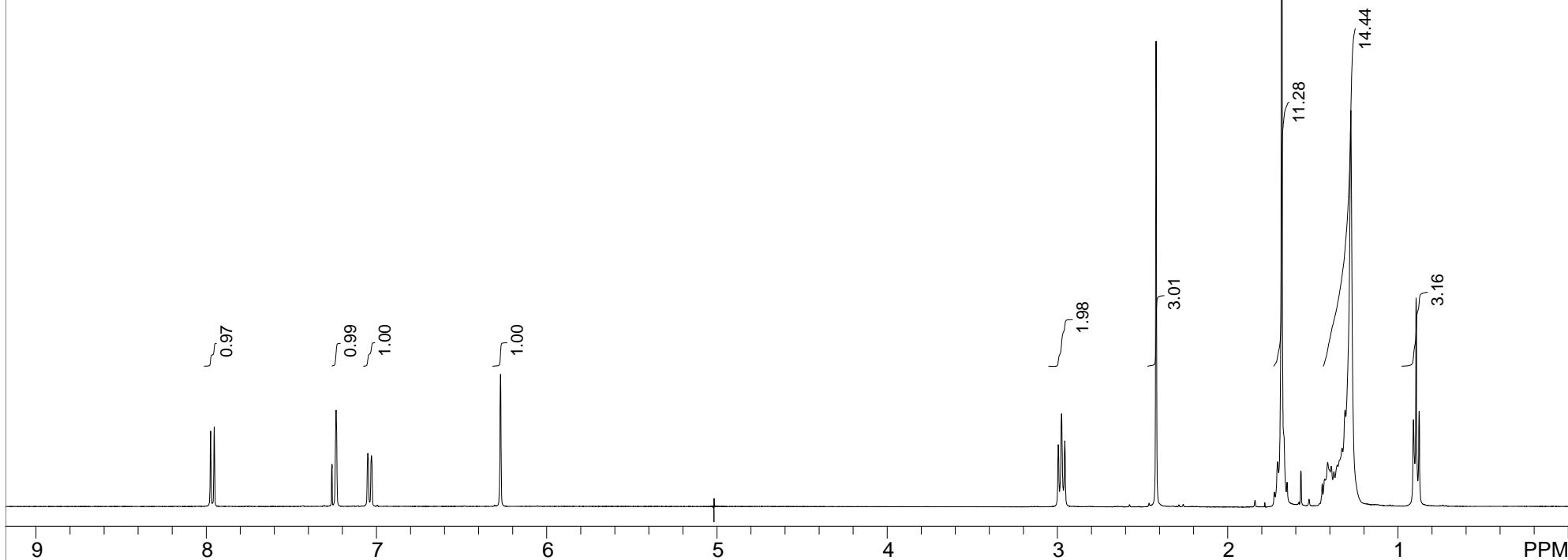
C = 0.00

8 - 8.00

THE INFLUENCE OF CULTURE ON PARENTING



3

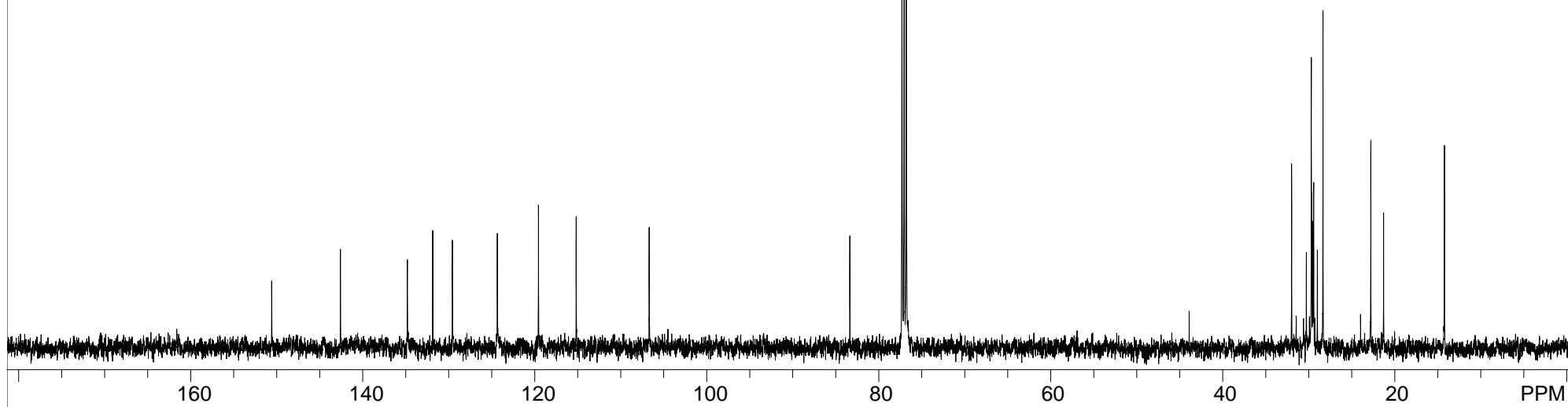


new experiment

USER: -- DATE: Mar 25 2011

F1: 399.951 F2: 100.575 SW1: 8002 OF1: 2006.8 PTS1d: 20006 , 32768
 EX: s2pul PW: 11.7 us PD: 4.8 sec NA: 10 LB: 0.0 Nuts - \$llz-1-33-1h.fid

C:\Users\zhang\Desktop\NMR\ll13c\processed\ll13c.fid
Mar 11 2011
USER:
SOLVENT: cdcl3
Experiment = s2pul
Pulse length = 6.000 usec
Relaxation delay = 3.000 sec
NA = 83
Solvent = cdcl3
FID PTS1d = 36749
PTS1d = 65536
F1 = 125.701683 MHz
F2 = 499.858551 MHz
SW1 = 28258.57 Hz
AT1 = 2.32 sec
Hz per Pt 1stD = 0.43 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 12777.2832 Hz
O2 = -0.5000 Hz
LB1 = 2.00 Hz
TP A = -271.41
B = 187.03
C = 0.00



USER: -- DATE: Mar 11 2011

F1: 125.702	F2: 499.859	SW1: 28259		OF1: 12777.3		PTS1d: 36749 , 65536	
EX: s2pul		PW: 6.0 us	PD: 3.0 sec	NA: 83	LB: 2.0		Nuts - \$llz-1-33-13c.fid

C:\Users\zhang\Desktop\NMR\nuts\DATA\\$llz-1-54-1h.fid

new experiment

Mar 21 2011

USER:

SOLVENT: cdcl3

Experiment = s2pul

Pulse length = 11.663 usec

Relaxation delay = 4.800 sec

NA = 32

Solvent = cdcl3

FID PTS1d = 20006

PTS1d = 32768

F1 = 399.950684 MHz

F2 = 100.575279 MHz

SW1 = 8002.40 Hz

AT1 = 2.50 sec

Hz per Pt 1stD = 0.24 Hz

SW2 = 1.00 Hz

Hz per Pt 2ndD = 1.00 Hz

O1 = 2006.3855 Hz

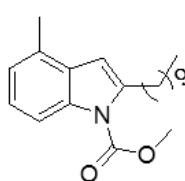
O2 = -0.5000 Hz

LB1 = 0.00 Hz

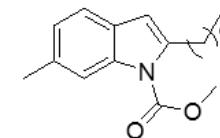
TP A = -48.63

B = -33.66

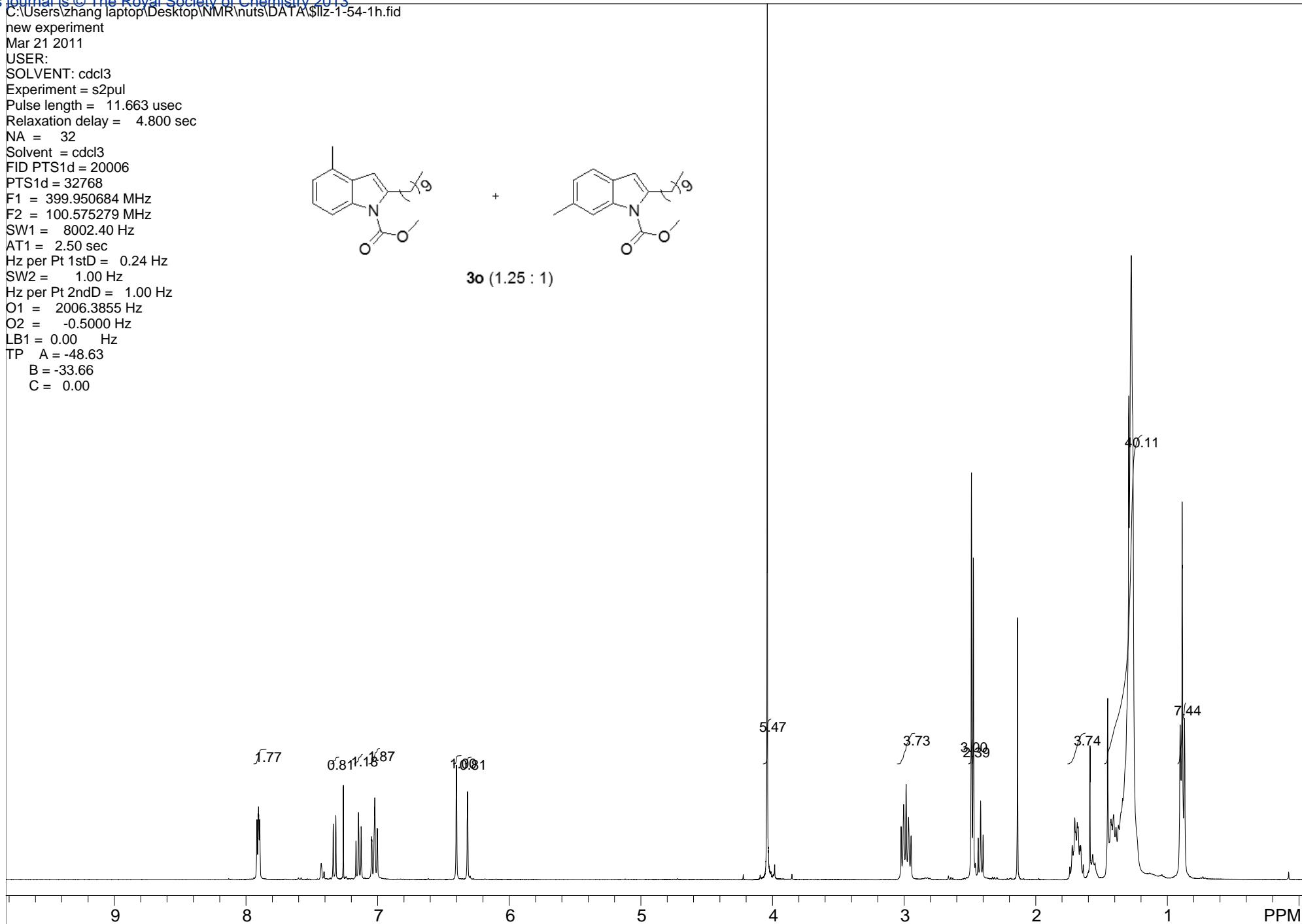
C = 0.00



+



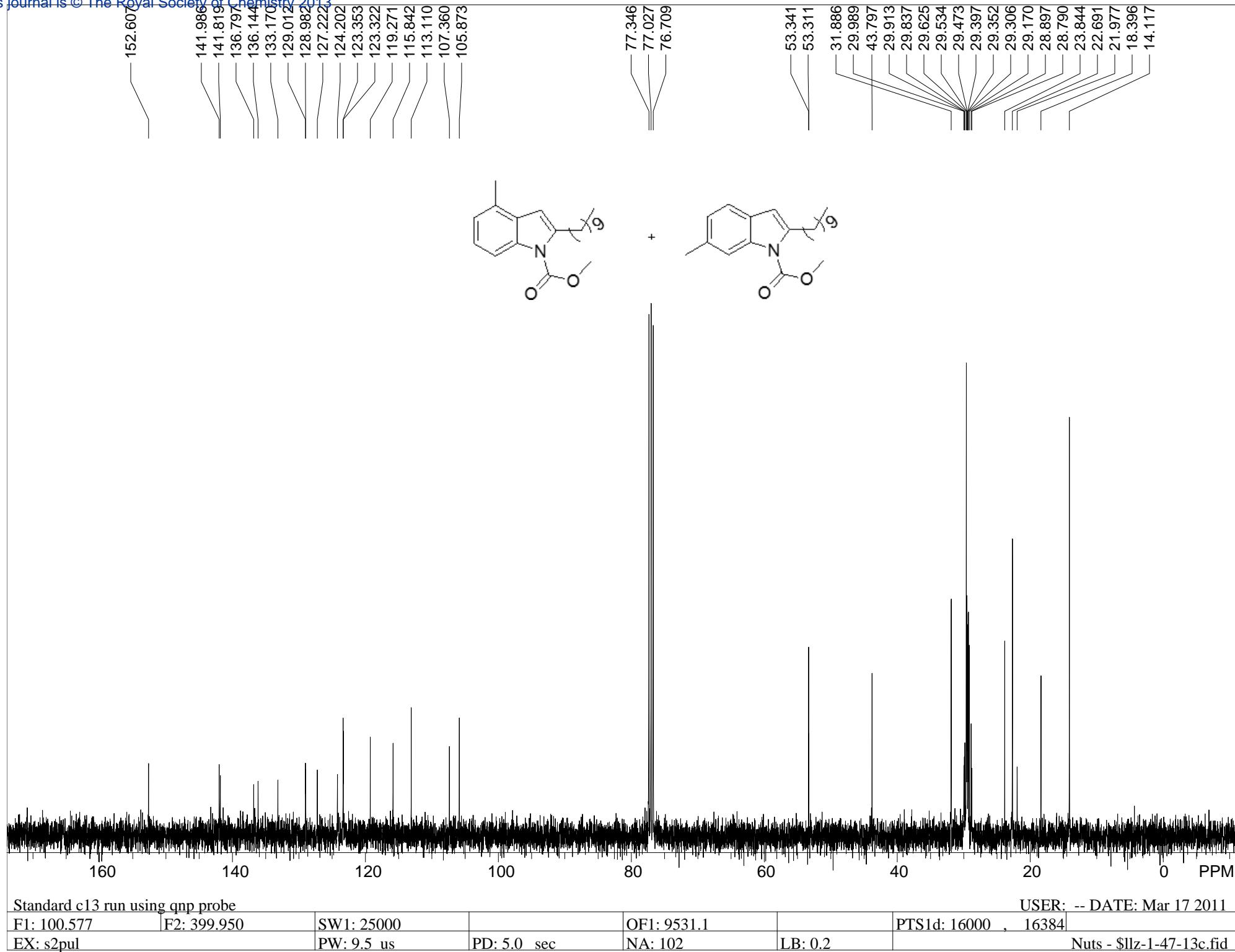
3o (1.25 : 1)



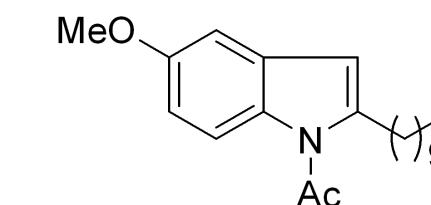
new experiment

USER: -- DATE: Mar 21 2011

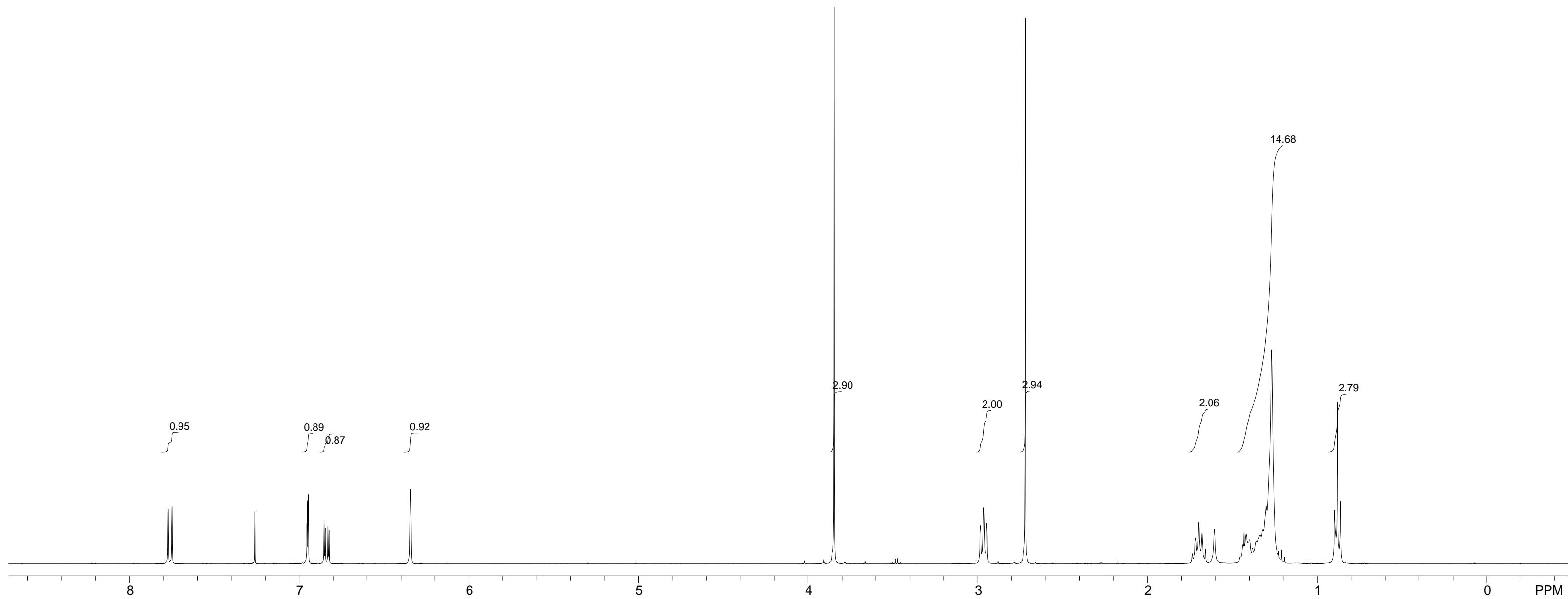
F1: 399.951	F2: 100.575	SW1: 8002		OF1: 2006.4		PTS1d: 20006 , 32768	
EX: s2pul		PW: 11.7 us		PD: 4.8 sec	NA: 32	LB: 0.0	Nuts - \\$llz-1-54-1h.fid



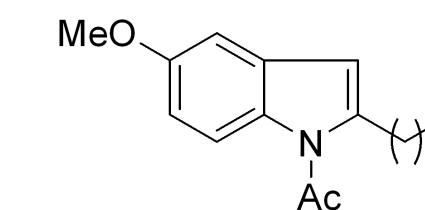
D:\Greenware\Ntus 20080731\DATA\\$wyz3-238-pro-h1.fid
new experiment
Apr 17 2011
USER:
SOLVENT: cdcl3
Experiment = s2pul
Pulse length = 11.663 usec
Relaxation delay = 2.000 sec
NA = 16
Solvent = cdcl3
FID PTS1d = 20006
PTS1d = 32768
F1 = 399.950684 MHz
F2 = 100.575279 MHz
SW1 = 8002.40 Hz
AT1 = 2.50 sec
Hz per Pt 1stD = 0.24 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 2006.2362 Hz
O2 = -0.5000 Hz
LB1 = 0.00 Hz
TP A = -41.51
B = -36.92
C = 0.00



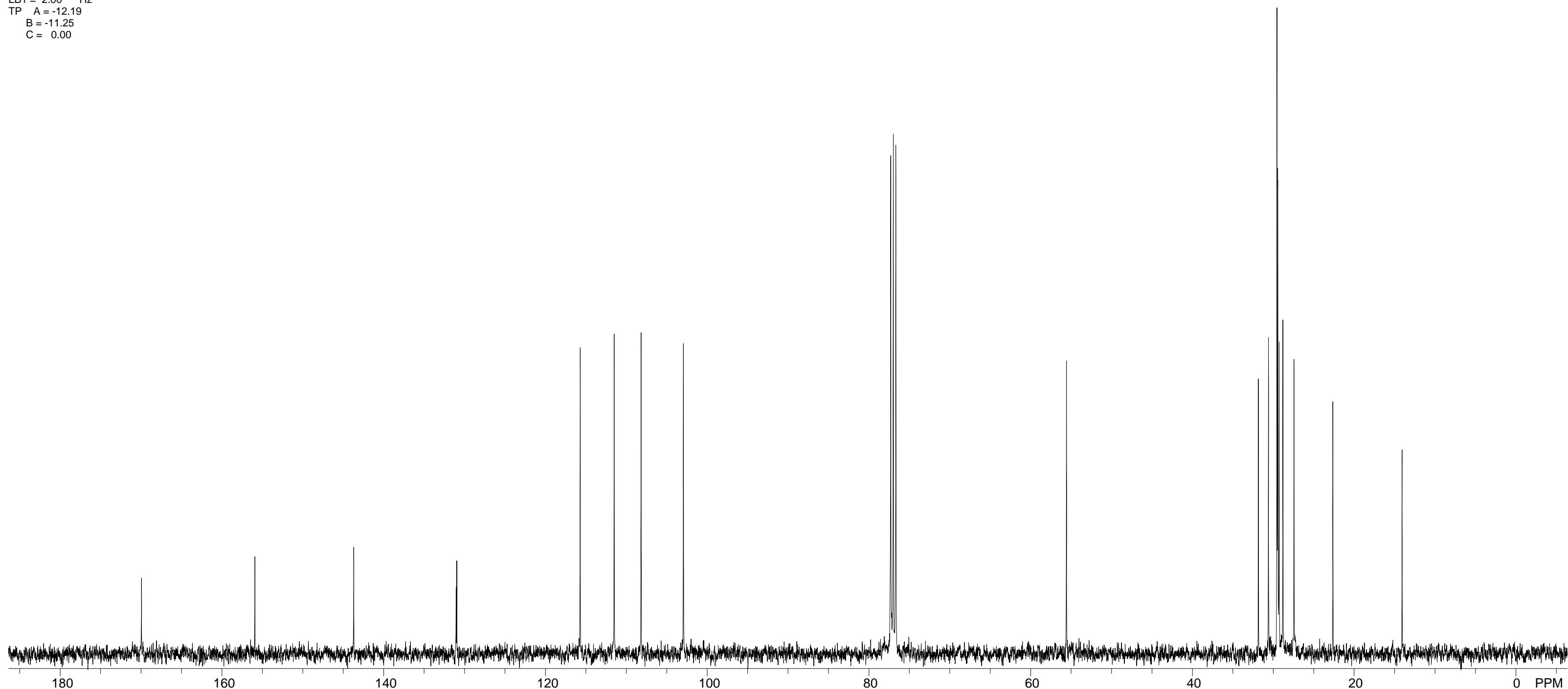
3



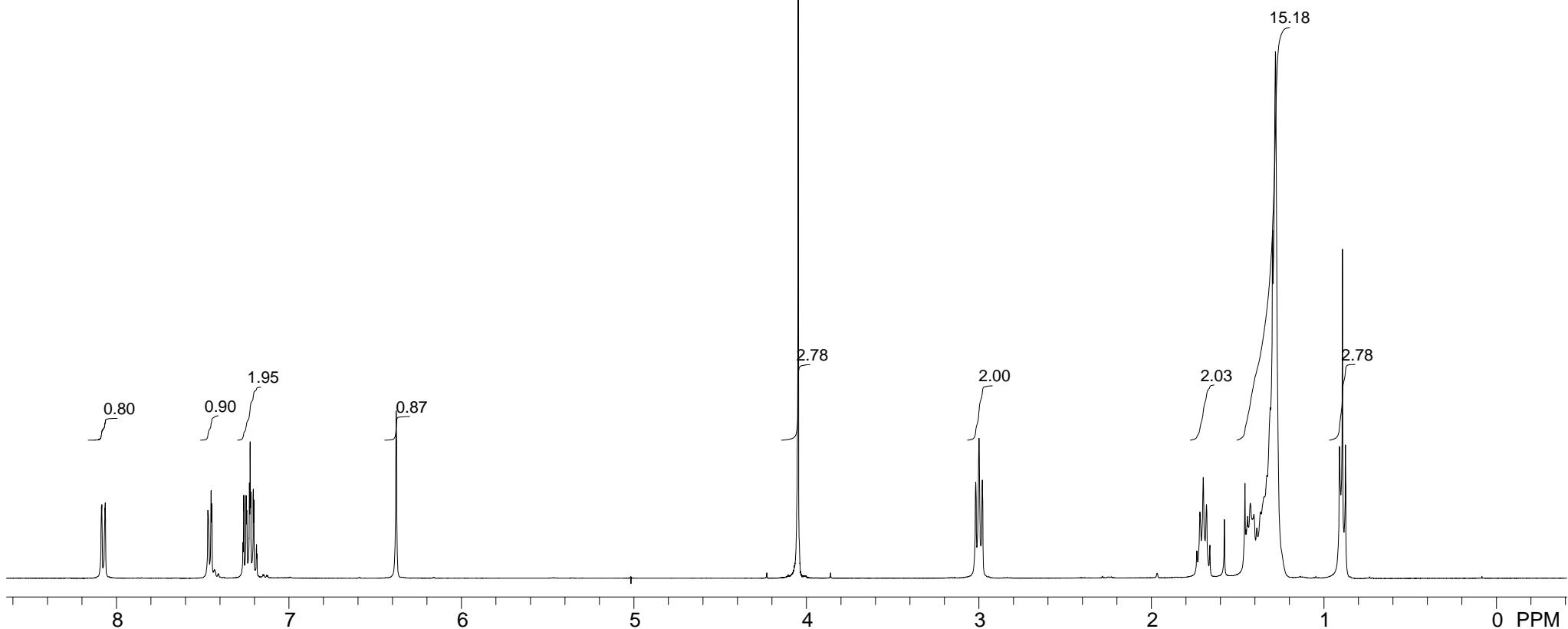
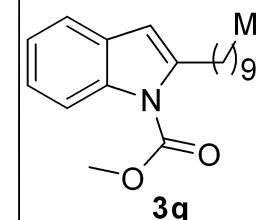
D:\Greenware\Ntus 20080731\DATA\\$wyz3-238-pro-c13.fid
Standard c13 run using qnp probe
Apr 17 2011
USER:
SOLVENT: cdcl3
Experiment = s2pul
Pulse length = 7.775 usec
Relaxation delay = 1.300 sec
NA = 560
Solvent = cdcl3
FID PTS1d = 28040
PTS1d = 32768
F1 = 100.577232 MHz
F2 = 399.950684 MHz
SW1 = 28040.66 Hz
AT1 = 1.00 sec
Hz per Pt 1stD = 0.86 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 10056.3604 Hz
O2 = -0.5000 Hz
LB1 = 2.00 Hz
TP A = -12.19
B = -11.25
C = 0.00



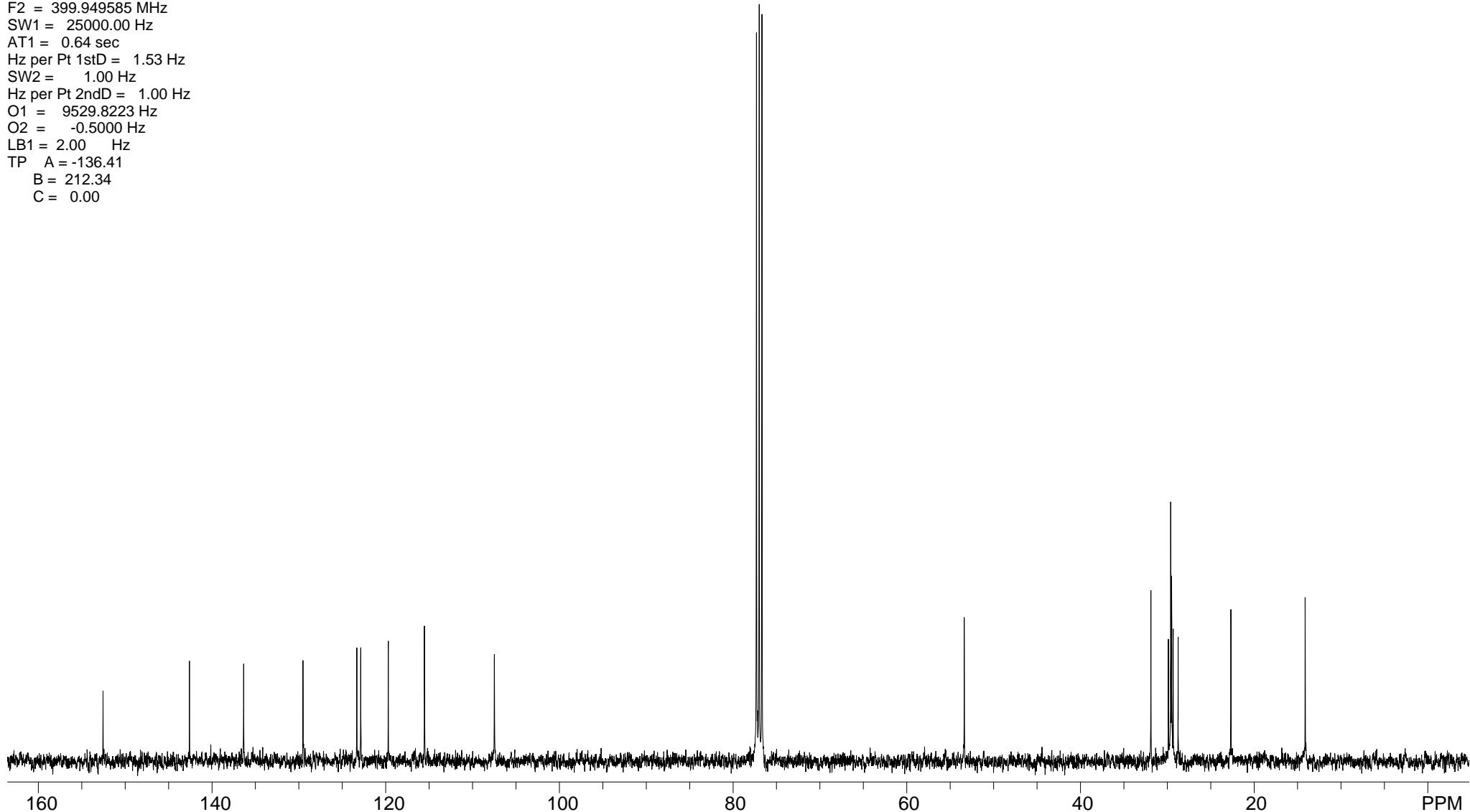
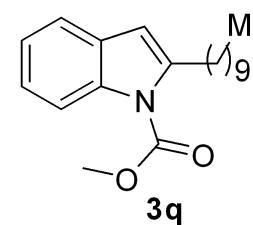
3p



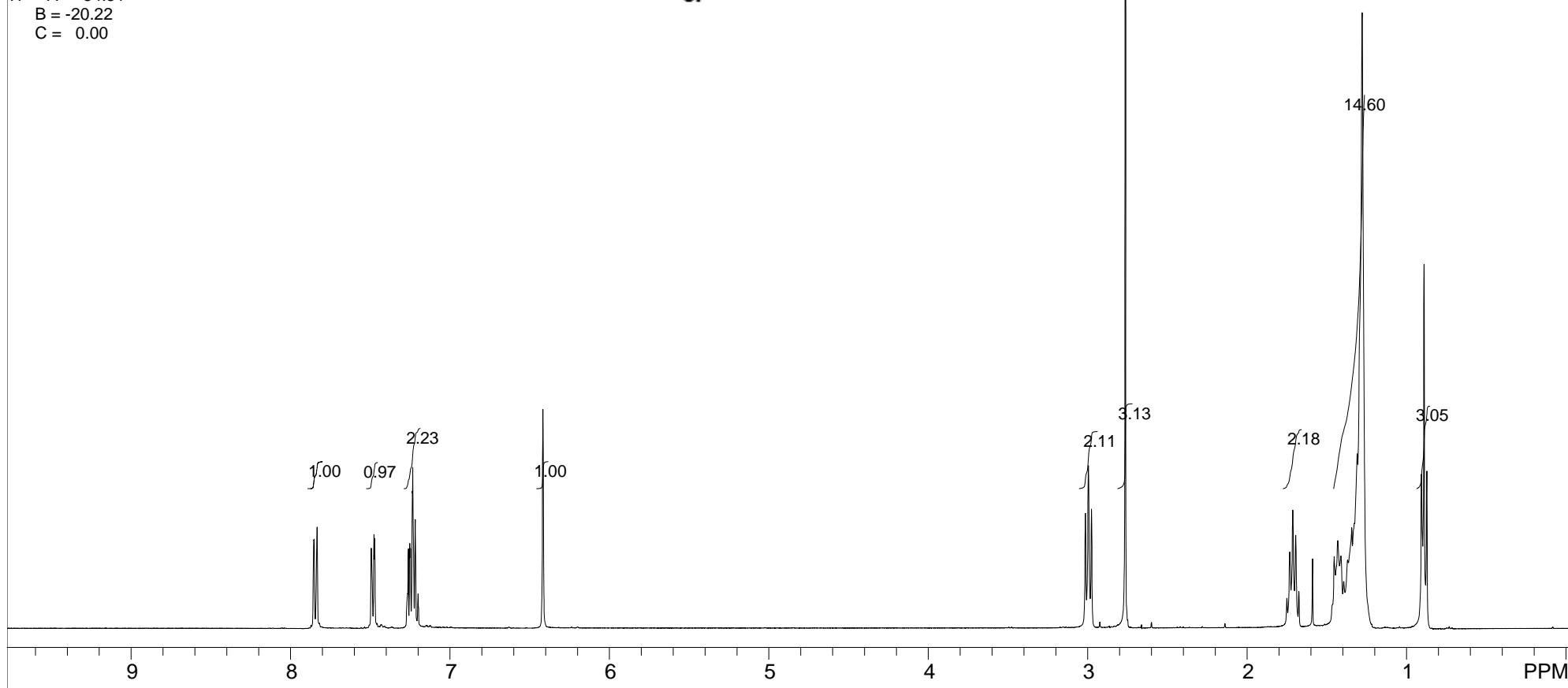
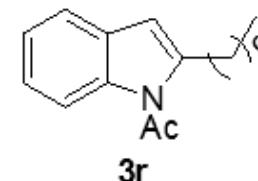
C:\Users\zhang laptop\Desktop\NMR\nuts\DATA\\$wyz3-213-pro-h1.fid
new experiment
Mar 22 2011
USER:
SOLVENT: cdcl3
Experiment = s2pul
Pulse length = 11.663 usec
Relaxation delay = 2.000 sec
NA = 20
Solvent = cdcl3
FID PTS1d = 20006
PTS1d = 32768
F1 = 399.950684 MHz
F2 = 100.575279 MHz
SW1 = 8002.40 Hz
AT1 = 2.50 sec
Hz per Pt 1stD = 0.24 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 2006.7504 Hz
O2 = -0.5000 Hz
LB1 = 0.00 Hz
TP A = -50.63
B = -26.20
C = 0.00



C:\Users\zhang laptop\Desktop\NMR\nuts\DATA\\$wyz3-213-pro-c13.fid
Standard c13 run using qnp probe
Mar 22 2011
USER:
SOLVENT: cdcl3
Experiment = s2pul
Pulse length = 9.500 usec
Relaxation delay = 5.000 sec
NA = 224
Solvent = cdcl3
FID PTS1d = 16000
PTS1d = 16384
F1 = 100.576706 MHz
F2 = 399.949585 MHz
SW1 = 25000.00 Hz
AT1 = 0.64 sec
Hz per Pt 1stD = 1.53 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 9529.8223 Hz
O2 = -0.5000 Hz
LB1 = 2.00 Hz
TP A = -136.41
B = 212.34
C = 0.00



C:\Users\zhang\Desktop\NMR\nuts\DATA\\$llz-1-51-1h.fid
 new experimentU
 Mar 27 2011
 USER:
 SOLVENT: cdcl3
 Experiment = s2pul
 Pulse length = 11.663 usec
 Relaxation delay = 4.800 sec
 NA = 16
 Solvent = cdcl3
 FID PTS1d = 20006
 PTS1d = 32768
 F1 = 399.950684 MHz
 F2 = 100.575279 MHz
 SW1 = 8002.40 Hz
 AT1 = 2.50 sec
 Hz per Pt 1stD = 0.24 Hz
 SW2 = 1.00 Hz
 Hz per Pt 2ndD = 1.00 Hz
 O1 = 2006.7504 Hz
 O2 = -0.5000 Hz
 LB1 = 0.00 Hz
 TP A = -54.51
 B = -20.22
 C = 0.00



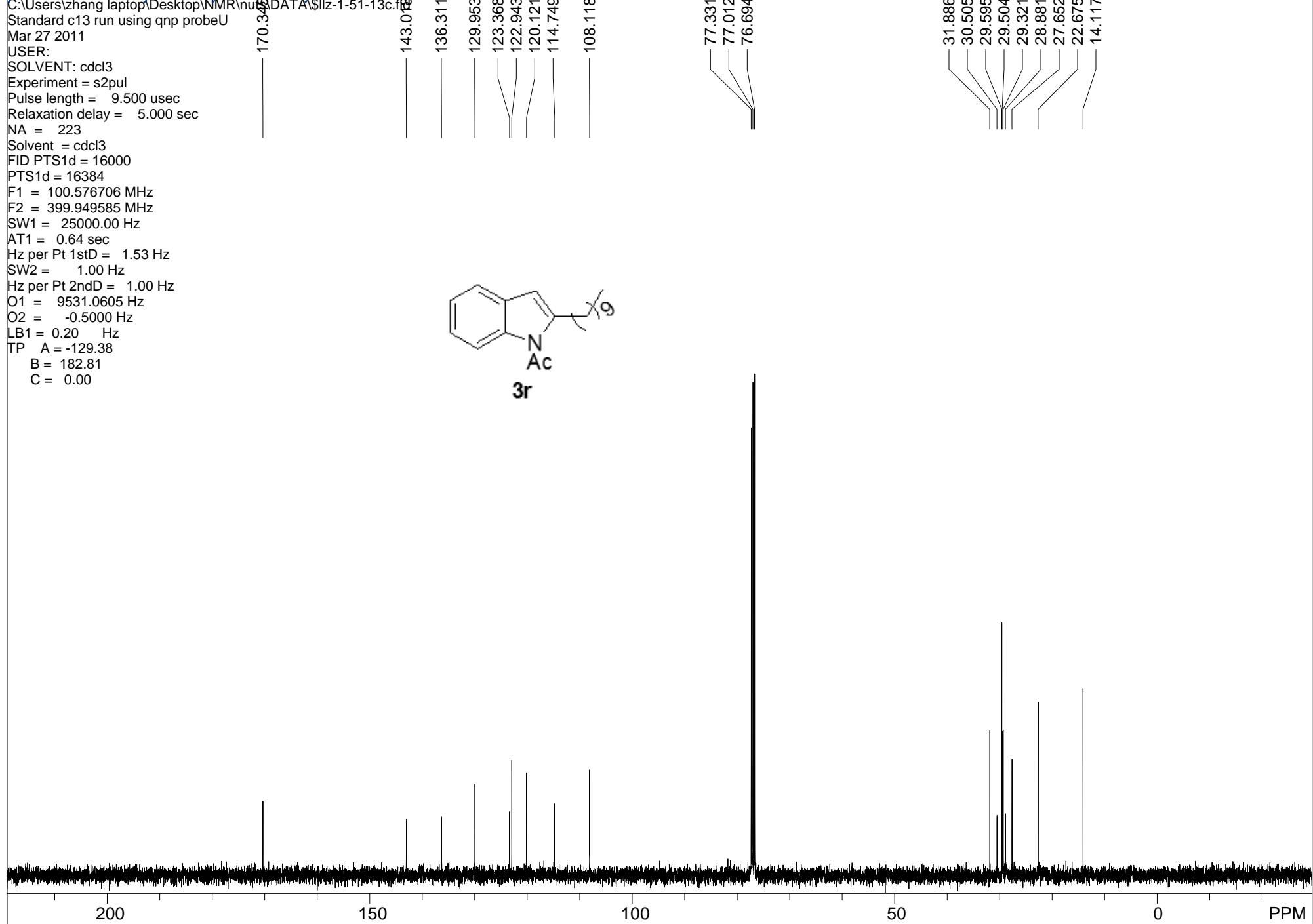
new experimentU

USER: -- DATE: Mar 27 2011

F1: 399.951	F2: 100.575	SW1: 8002		OF1: 2006.8		PTS1d: 20006 , 32768	
EX: s2pul		PW: 11.7 us		PD: 4.8 sec	NA: 16	LB: 0.0	Nuts - \\$llz-1-51-1h.fid

C:\Users\zhang\Desktop\NMR\nuts\DATA\\$llz-1-51-13c.fid

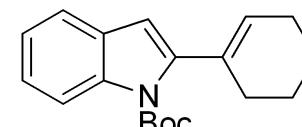
Standard c13 run using qnp probeU
Mar 27 2011
USER:
SOLVENT: cdcl3
Experiment = s2pul
Pulse length = 9.500 usec
Relaxation delay = 5.000 sec
NA = 223
Solvent = cdcl3
FID PTS1d = 16000
PTS1d = 16384
F1 = 100.576706 MHz
F2 = 399.949585 MHz
SW1 = 25000.00 Hz
AT1 = 0.64 sec
Hz per Pt 1stD = 1.53 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 9531.0605 Hz
O2 = -0.5000 Hz
LB1 = 0.20 Hz
TP A = -129.38
B = 182.81
C = 0.00



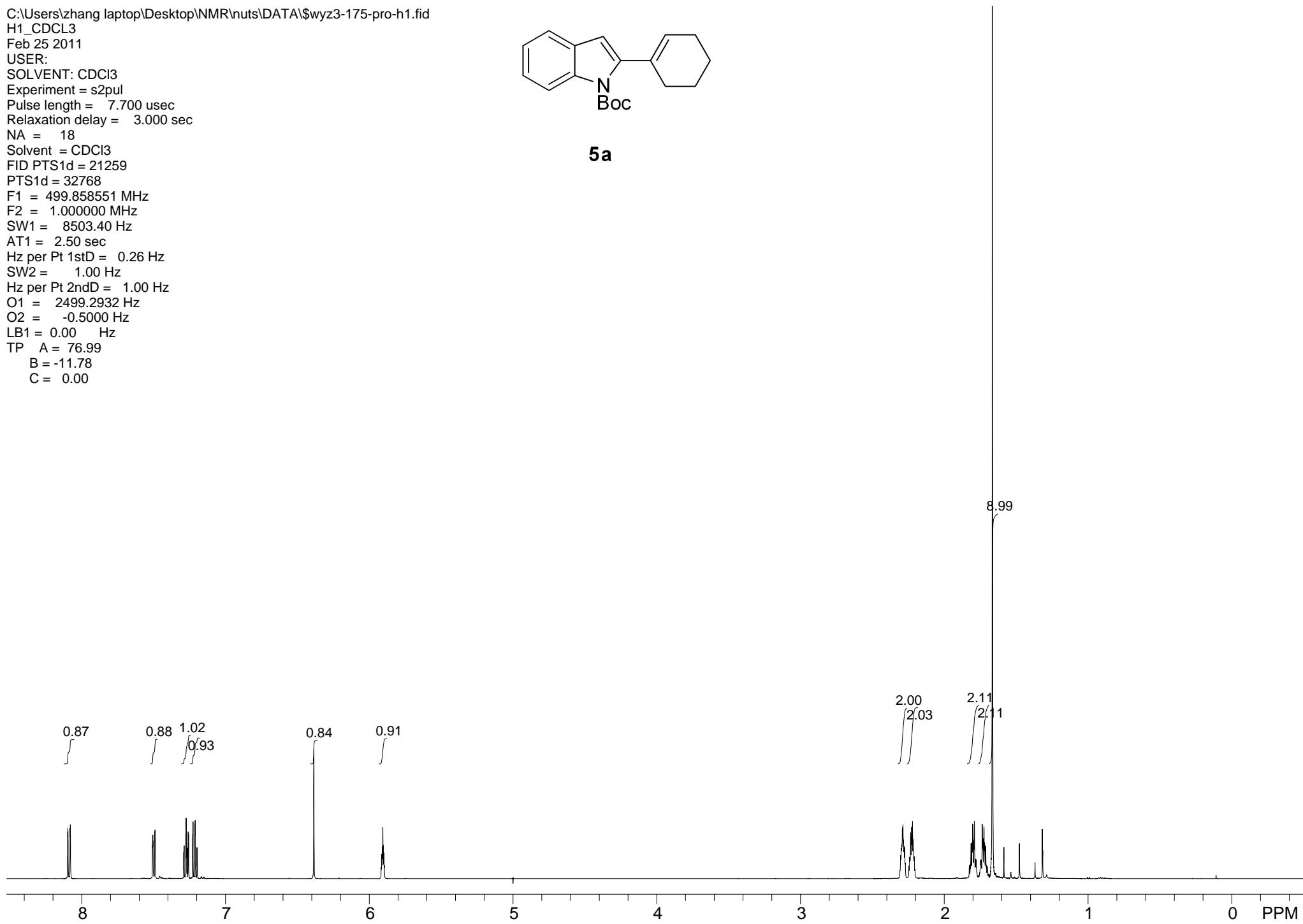
Standard c13 run using qnp probeU

F1: 100.577	F2: 399.950	SW1: 25000		OF1: 9531.1		PTS1d: 16000 , 16384	USER: -- DATE: Mar 27 2011
EX: s2pul		PW: 9.5 us	PD: 5.0 sec	NA: 223	LB: 0.2		Nuts - \\$llz-1-51-13c.fid

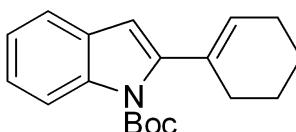
C:\Users\zhang laptop\Desktop\NMR\nuts\DATA\\$wyz3-175-pro-h1.fid
H1_CDCL3
Feb 25 2011
USER:
SOLVENT: CDCl3
Experiment = s2pul
Pulse length = 7.700 usec
Relaxation delay = 3.000 sec
NA = 18
Solvent = CDCl3
FID PTS1d = 21259
PTS1d = 32768
F1 = 499.858551 MHz
F2 = 1.000000 MHz
SW1 = 8503.40 Hz
AT1 = 2.50 sec
Hz per Pt 1stD = 0.26 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 2499.2932 Hz
O2 = -0.5000 Hz
LB1 = 0.00 Hz
TP A = 76.99
B = -11.78
C = 0.00



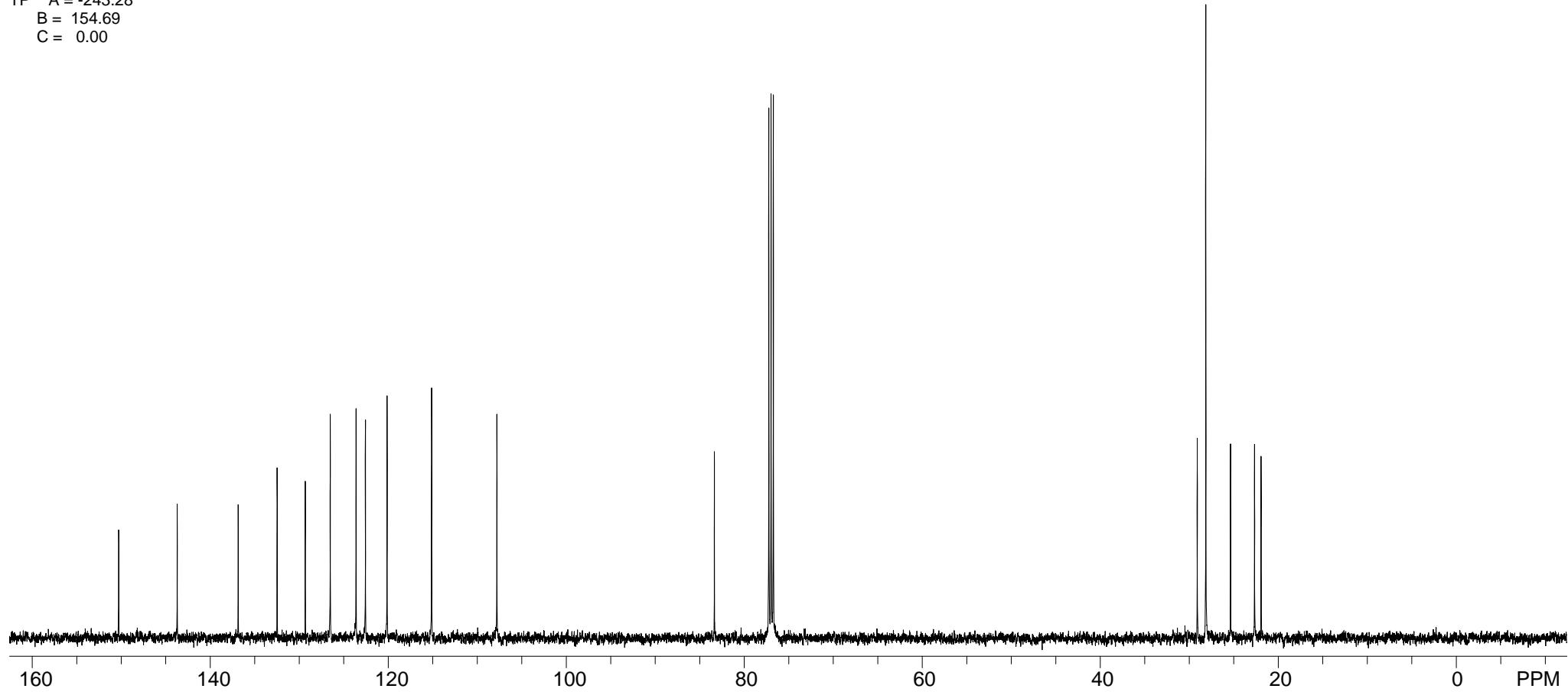
5a



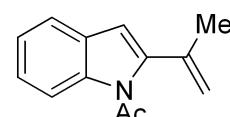
C:\Users\zhang laptop\Desktop\NMR\nuts\DATA\\$wyz3-175-pro-c13.fid
STANDARD CARBON PARAMETERS
Feb 25 2011
USER:
SOLVENT: cdcl3
Experiment = s2pul
Pulse length = 6.000 usec
Relaxation delay = 3.000 sec
NA = 96
Solvent = cdcl3
FID PTS1d = 42498
PTS1d = 65536
F1 = 125.702728 MHz
F2 = 499.858551 MHz
SW1 = 32679.74 Hz
AT1 = 1.30 sec
Hz per Pt 1stD = 0.50 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 13820.1719 Hz
O2 = -0.5000 Hz
LB1 = 2.00 Hz
TP A = -243.28
B = 154.69
C = 0.00



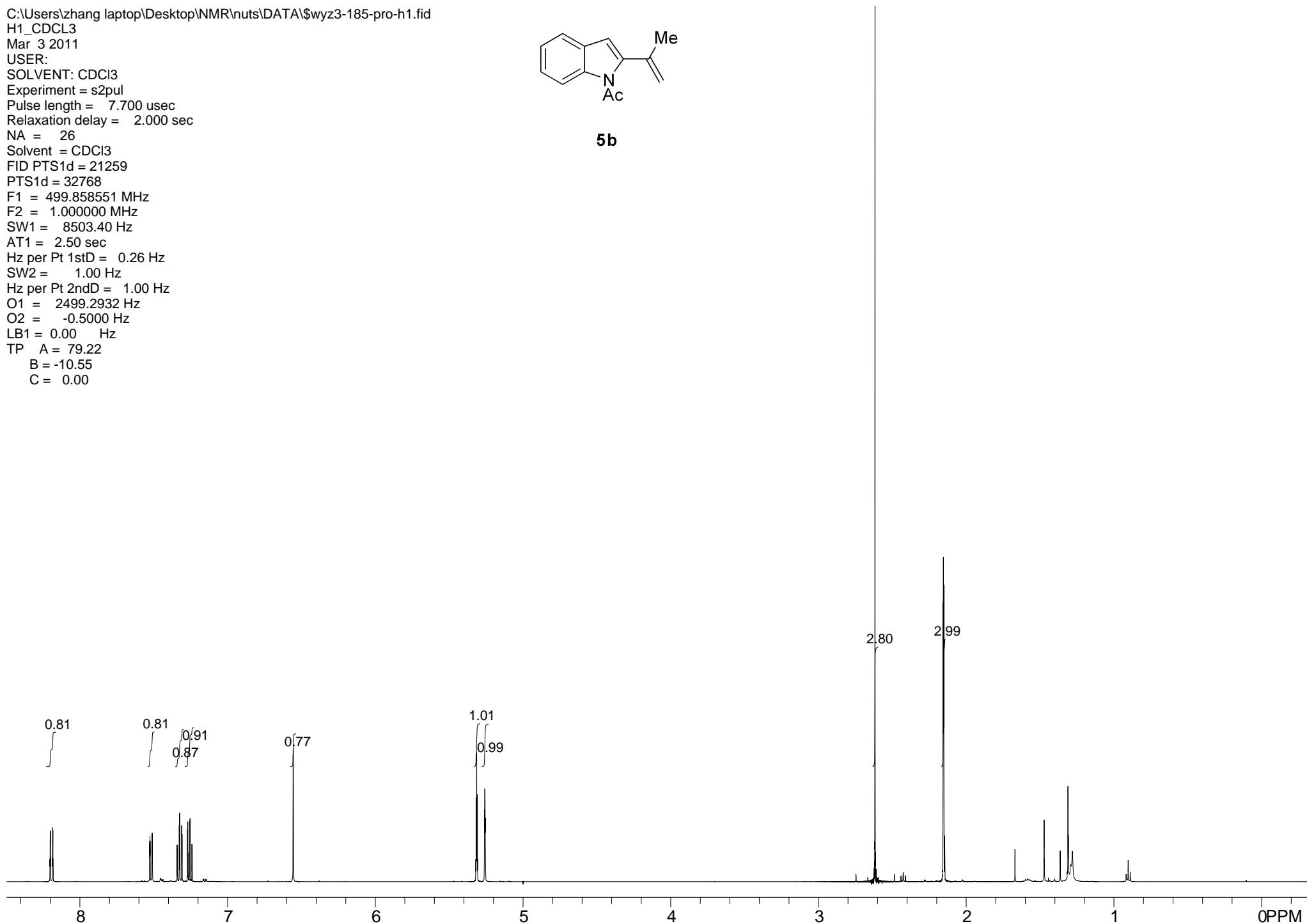
5a



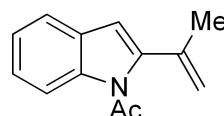
C:\Users\zhang laptop\Desktop\NMR\nuts\DATA\\$wyz3-185-pro-h1.fid
H1_CDCL3
Mar. 3 2011
USER:
SOLVENT: CDCl3
Experiment = s2pul
Pulse length = 7.700 usec
Relaxation delay = 2.000 sec
NA = 26
Solvent = CDCl3
FID PTS1d = 21259
PTS1d = 32768
F1 = 499.858551 MHz
F2 = 1.000000 MHz
SW1 = 8503.40 Hz
AT1 = 2.50 sec
Hz per Pt 1stD = 0.26 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 2499.2932 Hz
O2 = -0.5000 Hz
LB1 = 0.00 Hz
TP A = 79.22
B = -10.55
C = 0.00



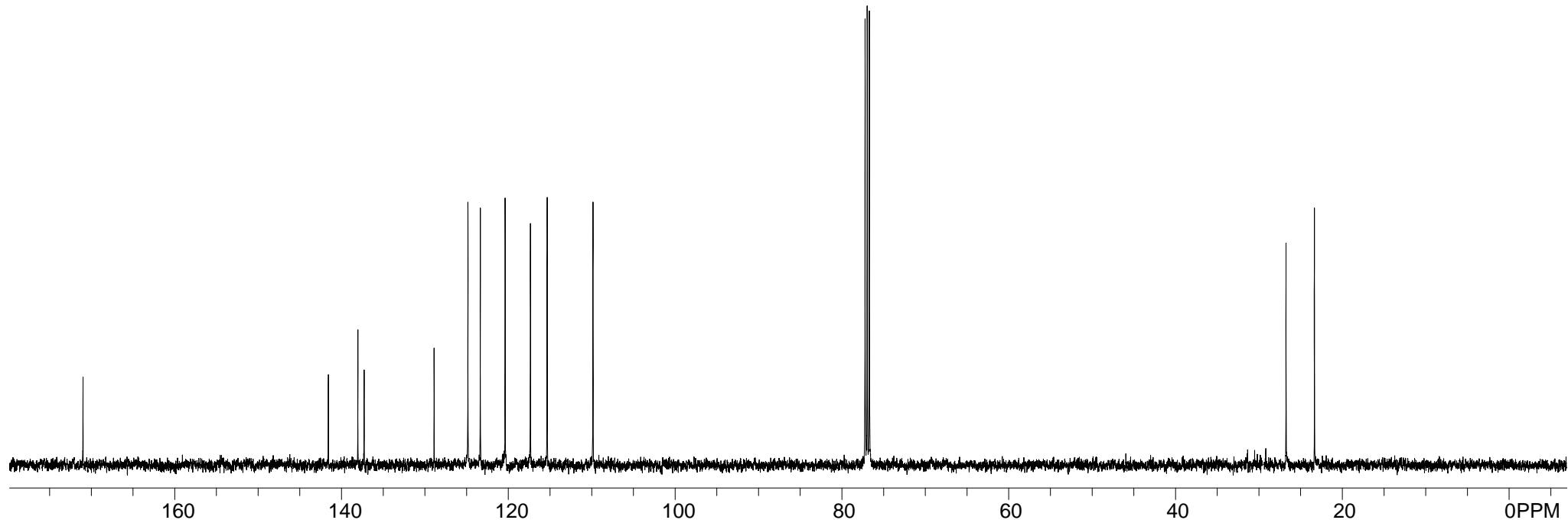
5b



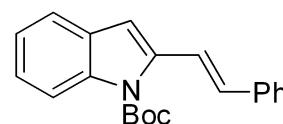
C:\Users\zhang laptop\Desktop\NMR\nuts\DATA\\$wyz3-185-pro-c13.fid
STANDARD CARBON PARAMETERS
Mar. 3 2011
USER:
SOLVENT: cdcl3
Experiment = s2pul
Pulse length = 6.000 usec
Relaxation delay = 3.000 sec
NA = 40
Solvent = cdcl3
FID PTS1d = 36749
PTS1d = 65536
F1 = 125.701683 MHz
F2 = 499.858551 MHz
SW1 = 28258.57 Hz
AT1 = 1.30 sec
Hz per Pt 1stD = 0.43 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 12766.0713 Hz
O2 = -0.5000 Hz
LB1 = 2.00 Hz
TP A = -227.81
B = 127.97
C = 0.00



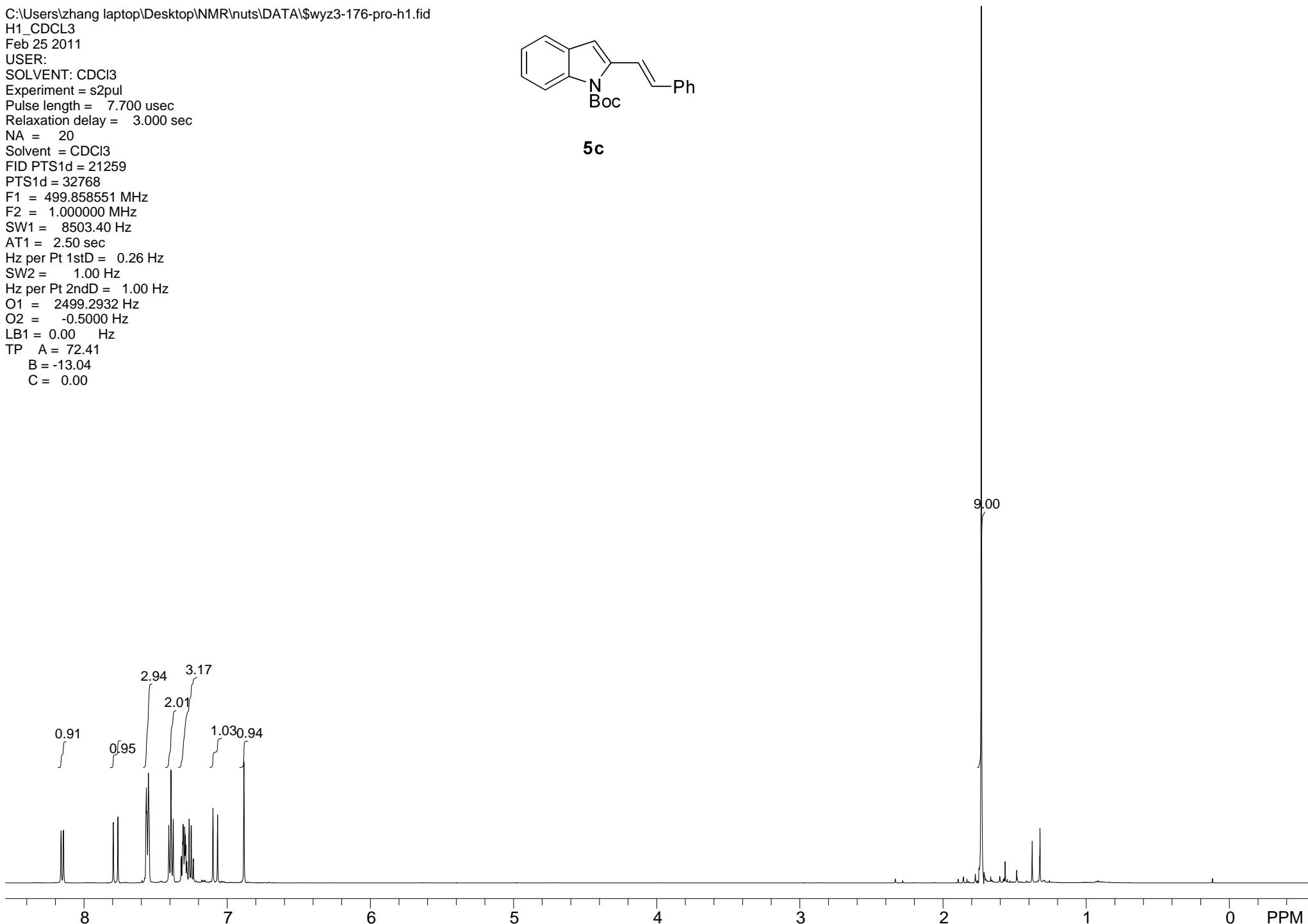
5b



C:\Users\zhang laptop\Desktop\NMR\nuts\DATA\\$wyz3-176-pro-h1.fid
H1_CDCL3
Feb 25 2011
USER:
SOLVENT: CDCl3
Experiment = s2pul
Pulse length = 7.700 usec
Relaxation delay = 3.000 sec
NA = 20
Solvent = CDCl3
FID PTS1d = 21259
PTS1d = 32768
F1 = 499.858551 MHz
F2 = 1.000000 MHz
SW1 = 8503.40 Hz
AT1 = 2.50 sec
Hz per Pt 1stD = 0.26 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 2499.2932 Hz
O2 = -0.5000 Hz
LB1 = 0.00 Hz
TP A = 72.41
B = -13.04
C = 0.00



5c



C:\Users\zhang laptop\Desktop\NMR\nuts\DATA\\$wyz3-176-pro-c13.fid
STANDARD CARBON PARAMETERS

Feb 25 2011

USER:

SOLVENT: cdcl3

Experiment = s2pul

Pulse length = 6.000 usec

Relaxation delay = 3.000 sec

NA = 60

Solvent = cdcl3

FID PTS1d = 36749

PTS1d = 65536

F1 = 125.701683 MHz

F2 = 499.858551 MHz

SW1 = 28258.57 Hz

AT1 = 1.30 sec

Hz per Pt 1stD = 0.43 Hz

SW2 = 1.00 Hz

Hz per Pt 2ndD = 1.00 Hz

O1 = 12768.2275 Hz

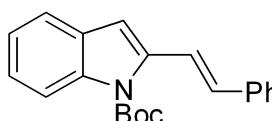
O2 = -0.5000 Hz

LB1 = 2.00 Hz

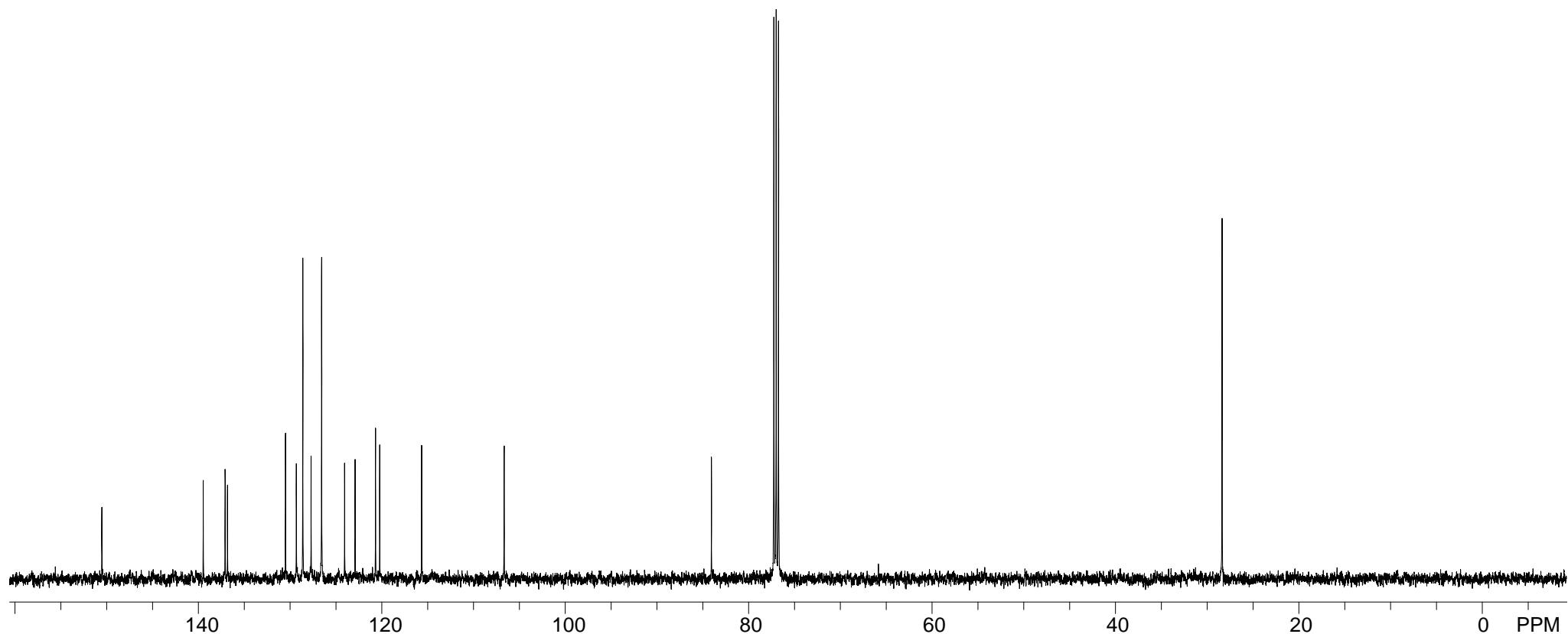
TP A = -249.93

B = 170.13

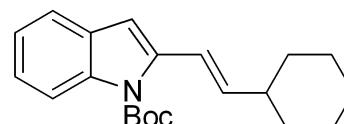
C = 0.00



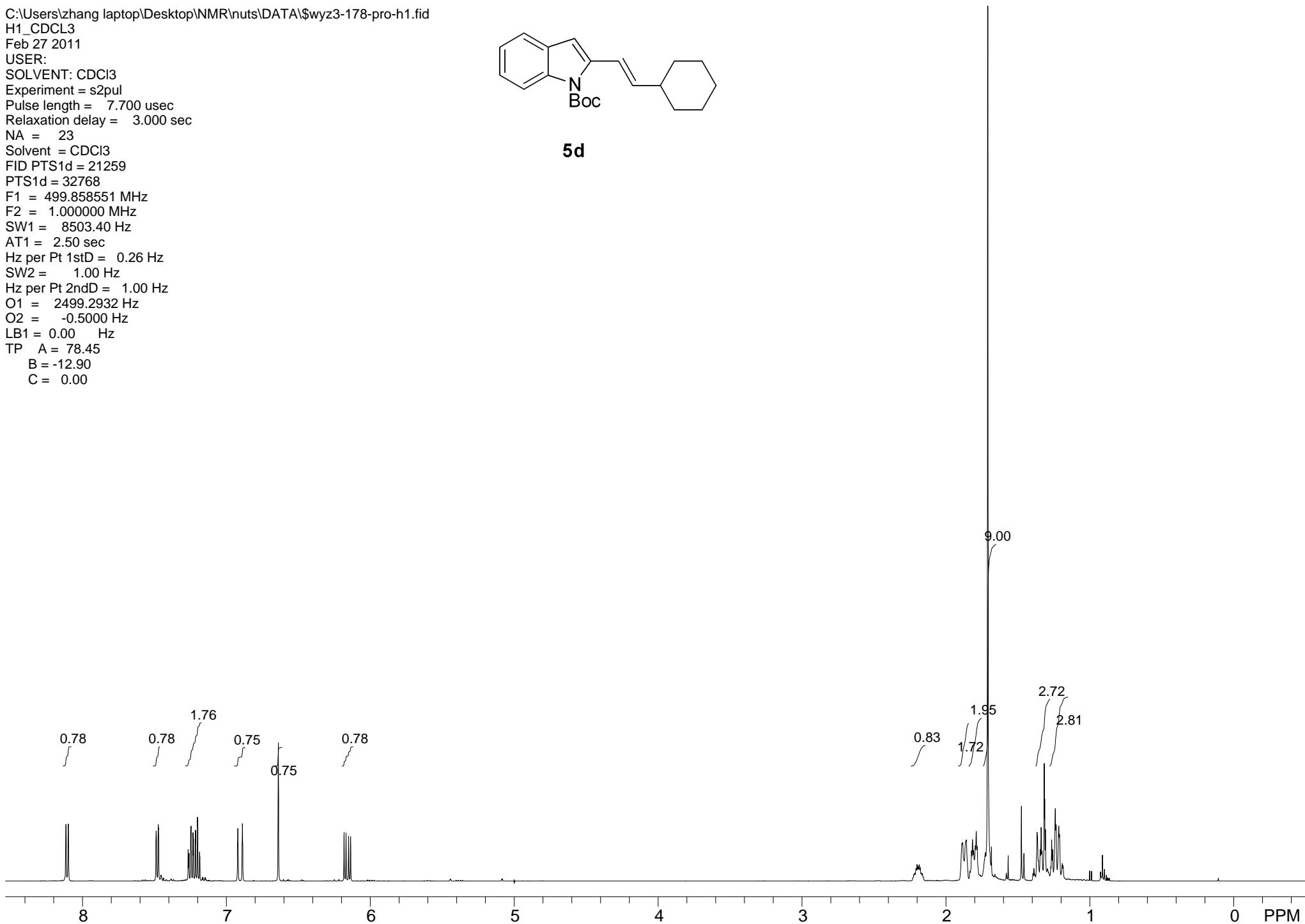
5c



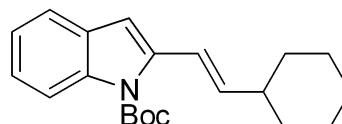
C:\Users\zhang laptop\Desktop\NMR\nuts\DATA\\$wyz3-178-pro-h1.fid
H1_CDCL3
Feb 27 2011
USER:
SOLVENT: CDCl3
Experiment = s2pul
Pulse length = 7.700 usec
Relaxation delay = 3.000 sec
NA = 23
Solvent = CDCl3
FID PTS1d = 21259
PTS1d = 32768
F1 = 499.858551 MHz
F2 = 1.000000 MHz
SW1 = 8503.40 Hz
AT1 = 2.50 sec
Hz per Pt 1stD = 0.26 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 2499.2932 Hz
O2 = -0.5000 Hz
LB1 = 0.00 Hz
TP A = 78.45
B = -12.90
C = 0.00



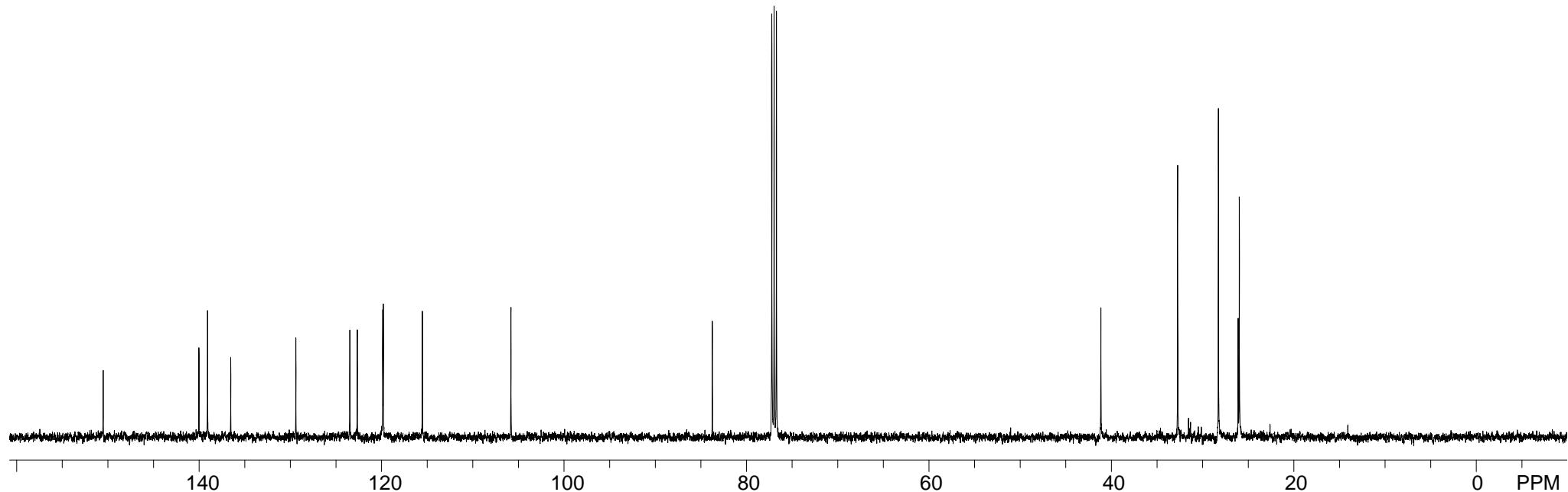
5d



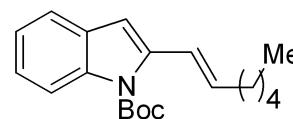
C:\Users\zhang laptop\Desktop\NMR\nuts\DATA\\$wyz3-178-pro-c13-1.fid
STANDARD CARBON PARAMETERS
Feb 27 2011
USER:
SOLVENT: cdcl3
Experiment = s2pul
Pulse length = 6.000 usec
Relaxation delay = 3.000 sec
NA = 52
Solvent = cdcl3
FID PTS1d = 36749
PTS1d = 65536
F1 = 125.701683 MHz
F2 = 499.858551 MHz
SW1 = 28258.57 Hz
AT1 = 1.30 sec
Hz per Pt 1stD = 0.43 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 12769.0908 Hz
O2 = -0.5000 Hz
LB1 = 2.00 Hz
TP A = -232.50
B = 135.00
C = 0.00



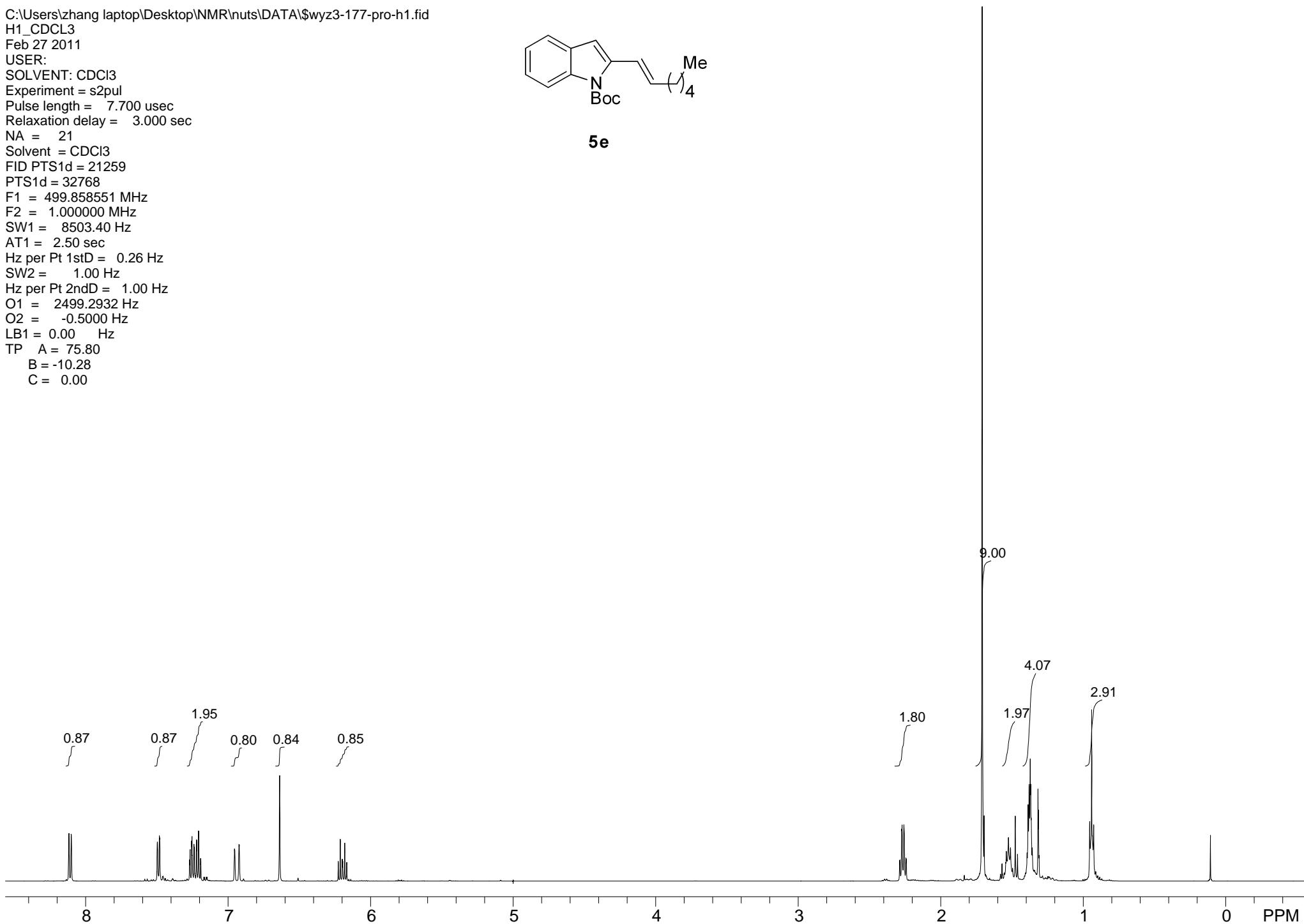
5d



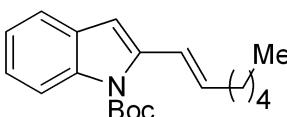
C:\Users\zhang laptop\Desktop\NMR\nuts\DATA\\$wyz3-177-pro-h1.fid
H1_CDCL3
Feb 27 2011
USER:
SOLVENT: CDCl3
Experiment = s2pul
Pulse length = 7.700 usec
Relaxation delay = 3.000 sec
NA = 21
Solvent = CDCl3
FID PTS1d = 21259
PTS1d = 32768
F1 = 499.858551 MHz
F2 = 1.000000 MHz
SW1 = 8503.40 Hz
AT1 = 2.50 sec
Hz per Pt 1stD = 0.26 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 2499.2932 Hz
O2 = -0.5000 Hz
LB1 = 0.00 Hz
TP A = 75.80
B = -10.28
C = 0.00



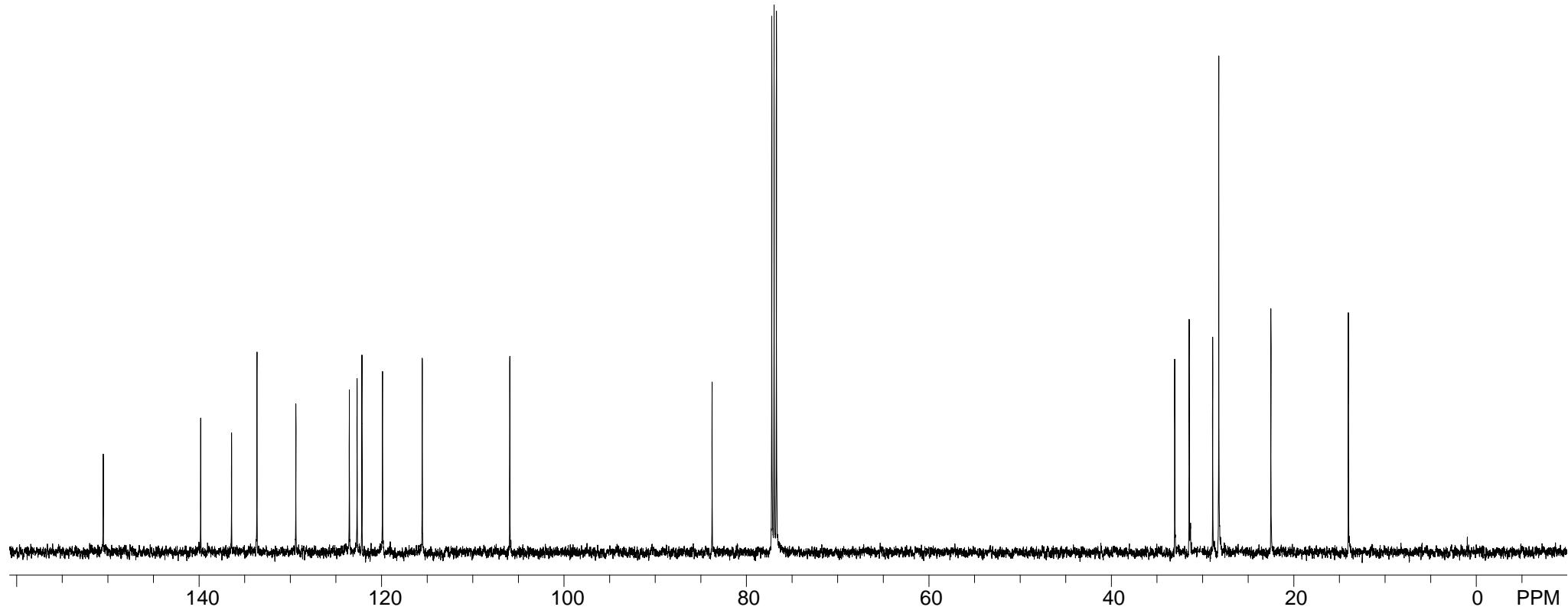
5e



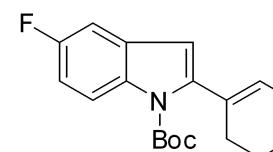
C:\Users\zhang laptop\Desktop\NMR\nuts\DATA\\$wyz3-177-pro-c13.fid
STANDARD CARBON PARAMETERS
Feb 27 2011
USER:
SOLVENT: cdcl3
Experiment = s2pul
Pulse length = 6.000 usec
Relaxation delay = 3.000 sec
NA = 88
Solvent = cdcl3
FID PTS1d = 36749
PTS1d = 65536
F1 = 125.701683 MHz
F2 = 499.858551 MHz
SW1 = 28258.57 Hz
AT1 = 1.30 sec
Hz per Pt 1stD = 0.43 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 12768.6592 Hz
O2 = -0.5000 Hz
LB1 = 2.00 Hz
TP A = -221.72
B = 123.75
C = 0.00



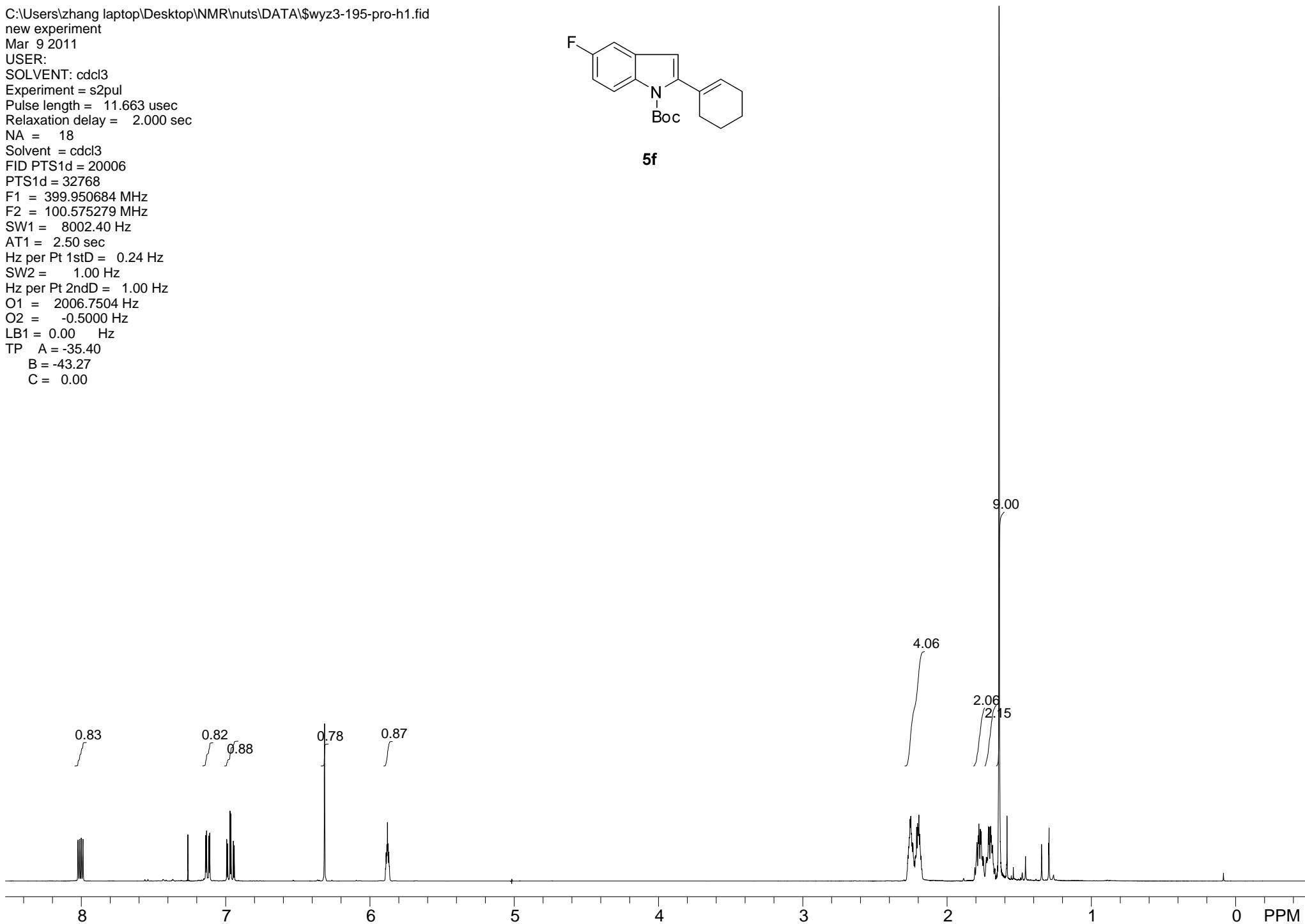
5e



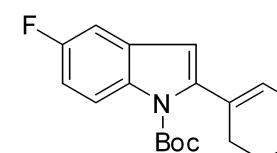
C:\Users\zhang laptop\Desktop\NMR\nuts\DATA\\$wyz3-195-pro-h1.fid
new experiment
Mar 9 2011
USER:
SOLVENT: cdcl3
Experiment = s2pul
Pulse length = 11.663 usec
Relaxation delay = 2.000 sec
NA = 18
Solvent = cdcl3
FID PTS1d = 20006
PTS1d = 32768
F1 = 399.950684 MHz
F2 = 100.575279 MHz
SW1 = 8002.40 Hz
AT1 = 2.50 sec
Hz per Pt 1stD = 0.24 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 2006.7504 Hz
O2 = -0.5000 Hz
LB1 = 0.00 Hz
TP A = -35.40
B = -43.27
C = 0.00



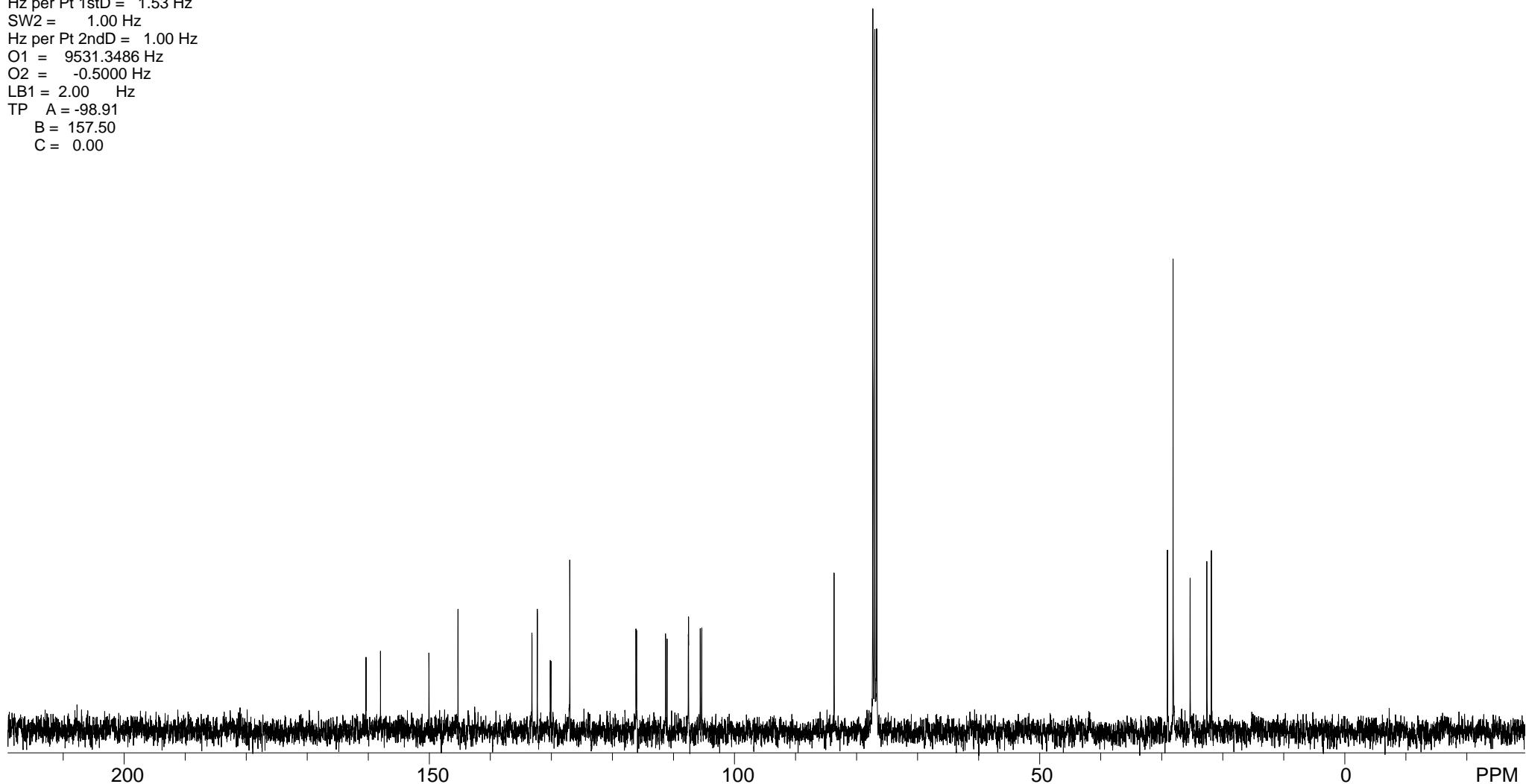
5f



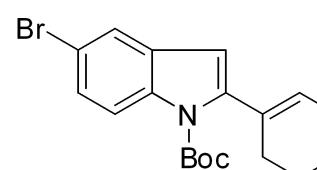
C:\Users\zhang laptop\Desktop\NMR\nuts\DATA\\$wyz3-195-pro-c13.fid
Standard c13 run using qnp probe
Mar. 9 2011
USER:
SOLVENT: cdcl3
Experiment = s2pul
Pulse length = 9.500 usec
Relaxation delay = 5.000 sec
NA = 88
Solvent = cdcl3
FID PTS1d = 16000
PTS1d = 16384
F1 = 100.576706 MHz
F2 = 399.949585 MHz
SW1 = 25000.00 Hz
AT1 = 0.64 sec
Hz per Pt 1stD = 1.53 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 9531.3486 Hz
O2 = -0.5000 Hz
LB1 = 2.00 Hz
TP A = -98.91
B = 157.50
C = 0.00



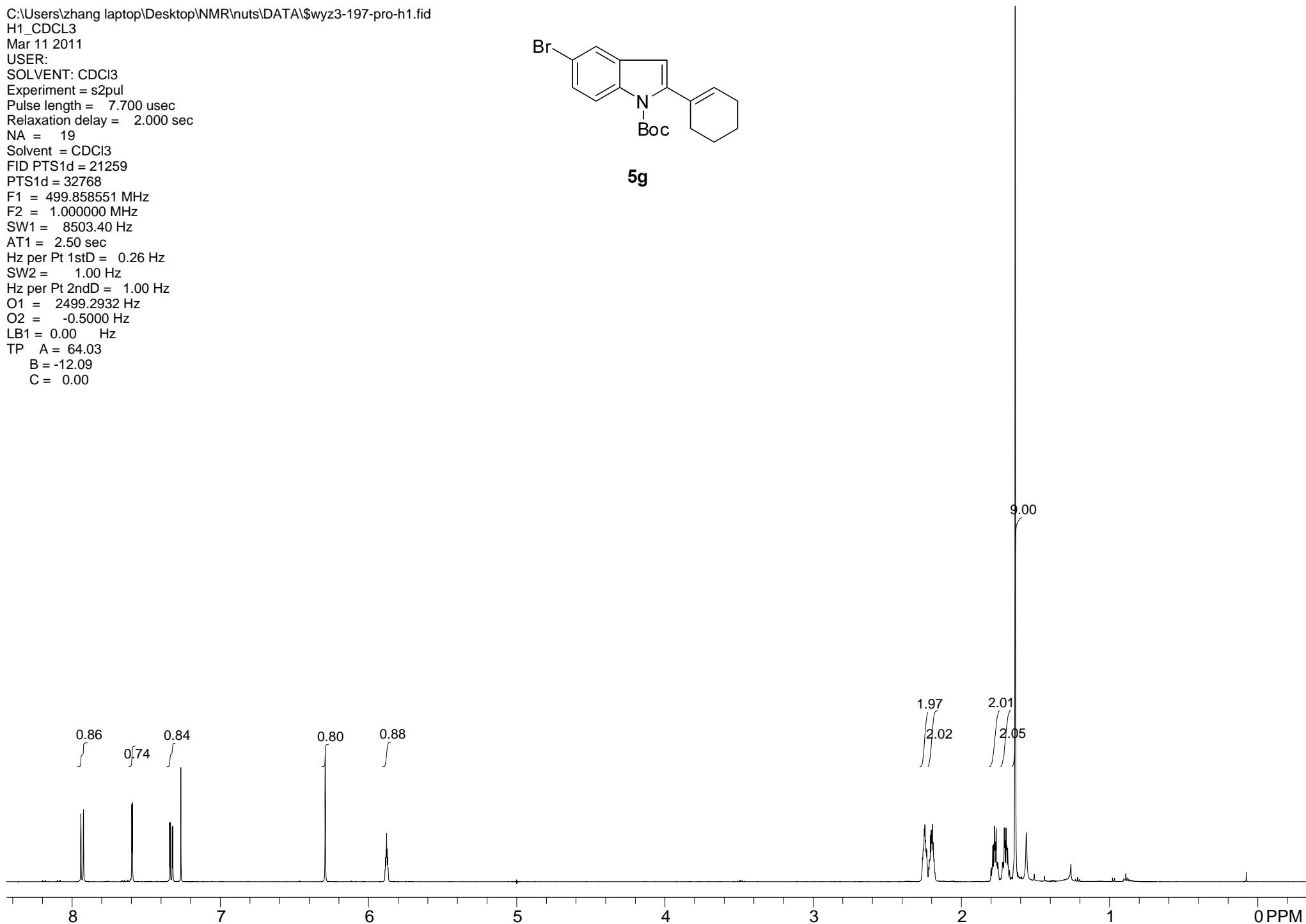
5f



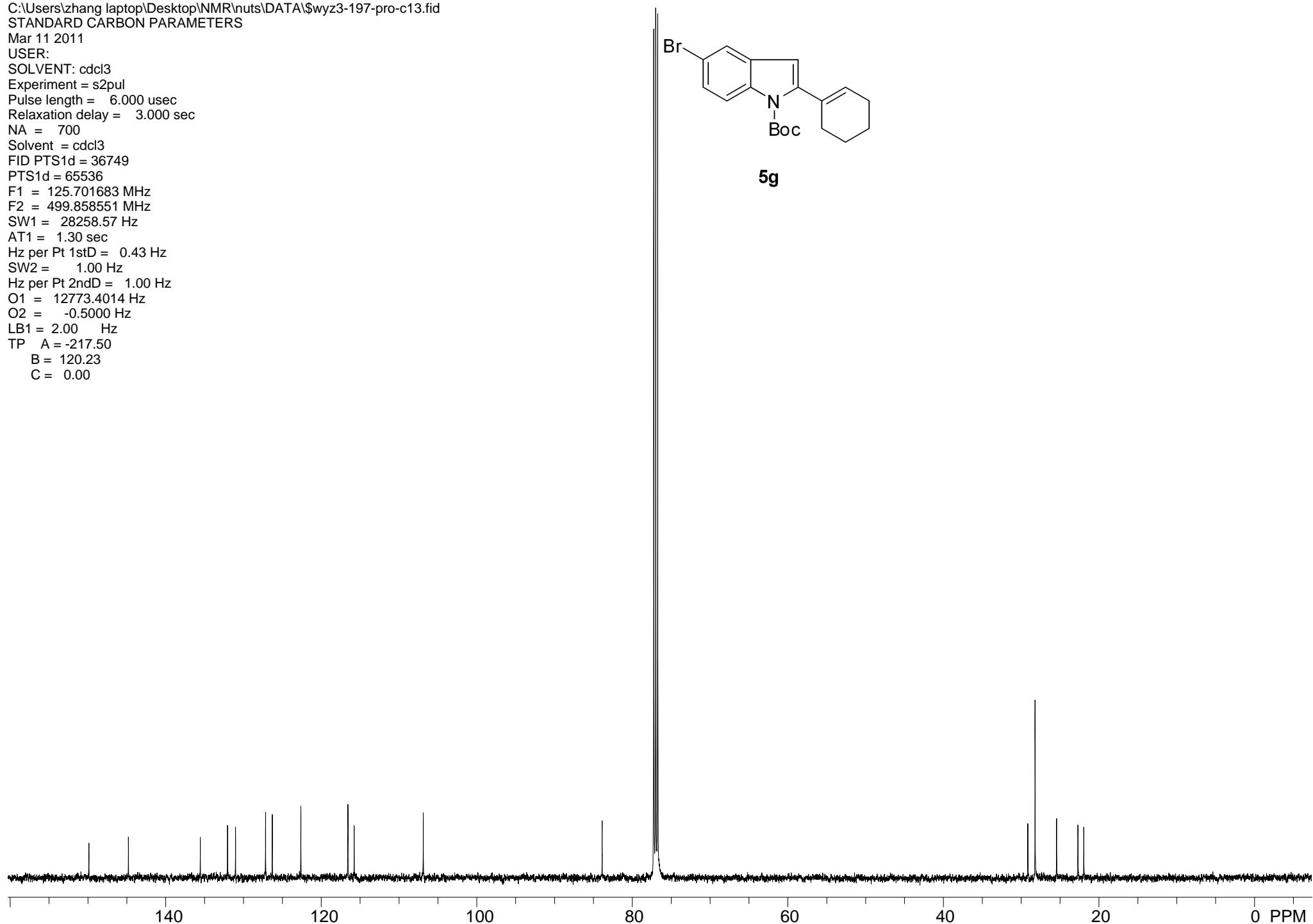
C:\Users\zhang laptop\Desktop\NMR\nuts\DATA\\$wyz3-197-pro-h1.fid
H1_CDCL3
Mar 11 2011
USER:
SOLVENT: CDCl3
Experiment = s2pul
Pulse length = 7.700 usec
Relaxation delay = 2.000 sec
NA = 19
Solvent = CDCl3
FID PTS1d = 21259
PTS1d = 32768
F1 = 499.858551 MHz
F2 = 1.000000 MHz
SW1 = 8503.40 Hz
AT1 = 2.50 sec
Hz per Pt 1stD = 0.26 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 2499.2932 Hz
O2 = -0.5000 Hz
LB1 = 0.00 Hz
TP A = 64.03
B = -12.09
C = 0.00



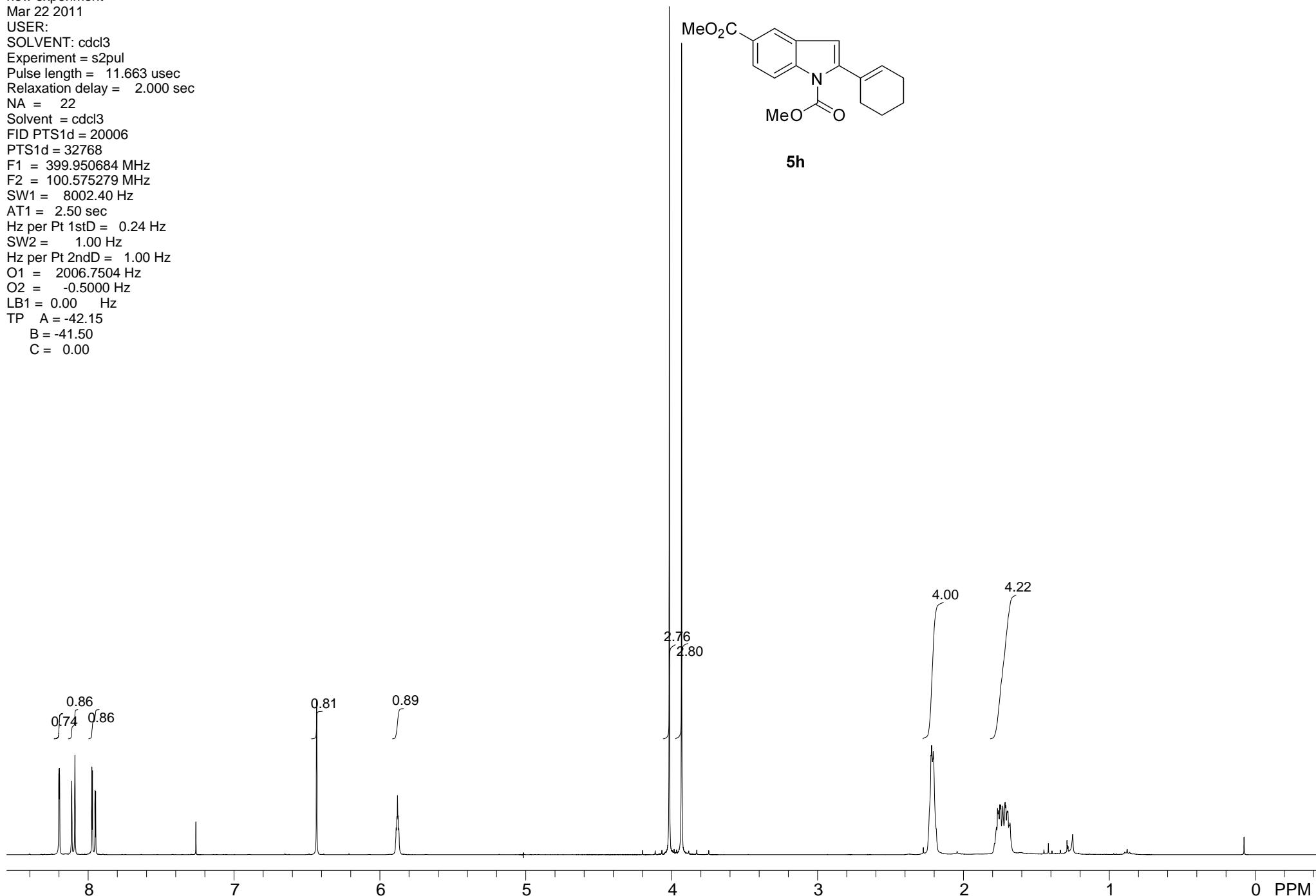
5g



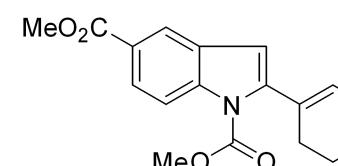
C:\Users\zhang laptop\Desktop\NMR\nuts\DATA\\$wyz3-197-pro-c13.fid
STANDARD CARBON PARAMETERS
Mar 11 2011
USER:
SOLVENT: cdcl3
Experiment = s2pul
Pulse length = 6.000 usec
Relaxation delay = 3.000 sec
NA = 700
Solvent = cdcl3
FID PTS1d = 36749
PTS1d = 65536
F1 = 125.701683 MHz
F2 = 499.858551 MHz
SW1 = 28258.57 Hz
AT1 = 1.30 sec
Hz per Pt 1stD = 0.43 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 12773.4014 Hz
O2 = -0.5000 Hz
LB1 = 2.00 Hz
TP A = -217.50
B = 120.23
C = 0.00



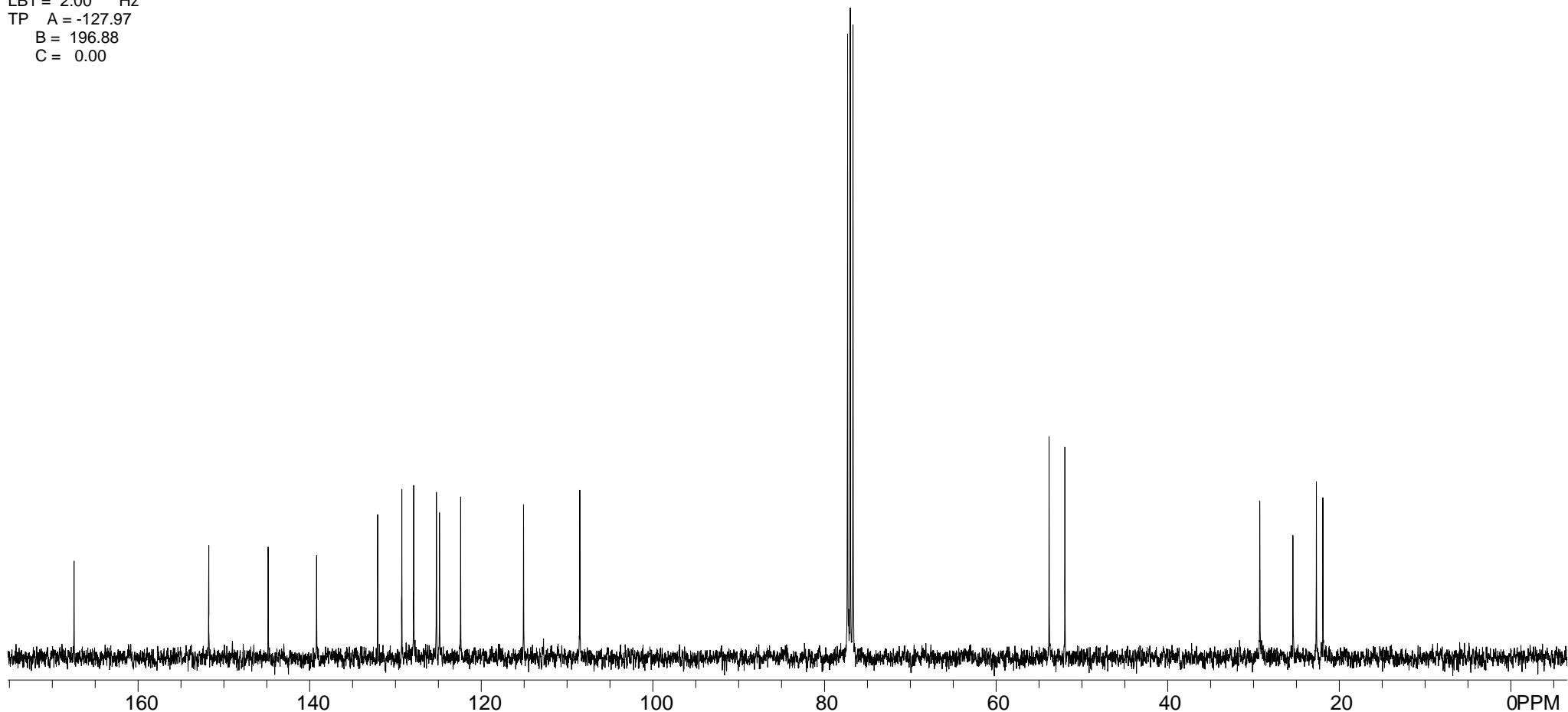
C:\Users\zhang laptop\Desktop\NMR\nuts\DATA\\$wyz3-216-pro-h1.fid
new experiment
Mar 22 2011
USER:
SOLVENT: cdcl3
Experiment = s2pul
Pulse length = 11.663 usec
Relaxation delay = 2.000 sec
NA = 22
Solvent = cdcl3
FID PTS1d = 20006
PTS1d = 32768
F1 = 399.950684 MHz
F2 = 100.575279 MHz
SW1 = 8002.40 Hz
AT1 = 2.50 sec
Hz per Pt 1stD = 0.24 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 2006.7504 Hz
O2 = -0.5000 Hz
LB1 = 0.00 Hz
TP A = -42.15
B = -41.50
C = 0.00



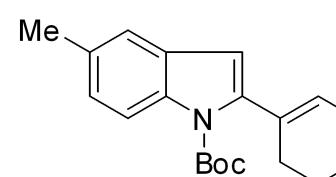
C:\Users\zhang laptop\Desktop\NMR\nuts\DATA\\$wyz3-216-pro-c13.fid
Standard c13 run using qnp probe
Mar 22 2011
USER:
SOLVENT: cdcl3
Experiment = s2pul
Pulse length = 9.500 usec
Relaxation delay = 5.000 sec
NA = 100
Solvent = cdcl3
FID PTS1d = 16000
PTS1d = 16384
F1 = 100.576706 MHz
F2 = 399.949585 MHz
SW1 = 25000.00 Hz
AT1 = 0.64 sec
Hz per Pt 1stD = 1.53 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 9528.2969 Hz
O2 = -0.5000 Hz
LB1 = 2.00 Hz
TP A = -127.97
B = 196.88
C = 0.00



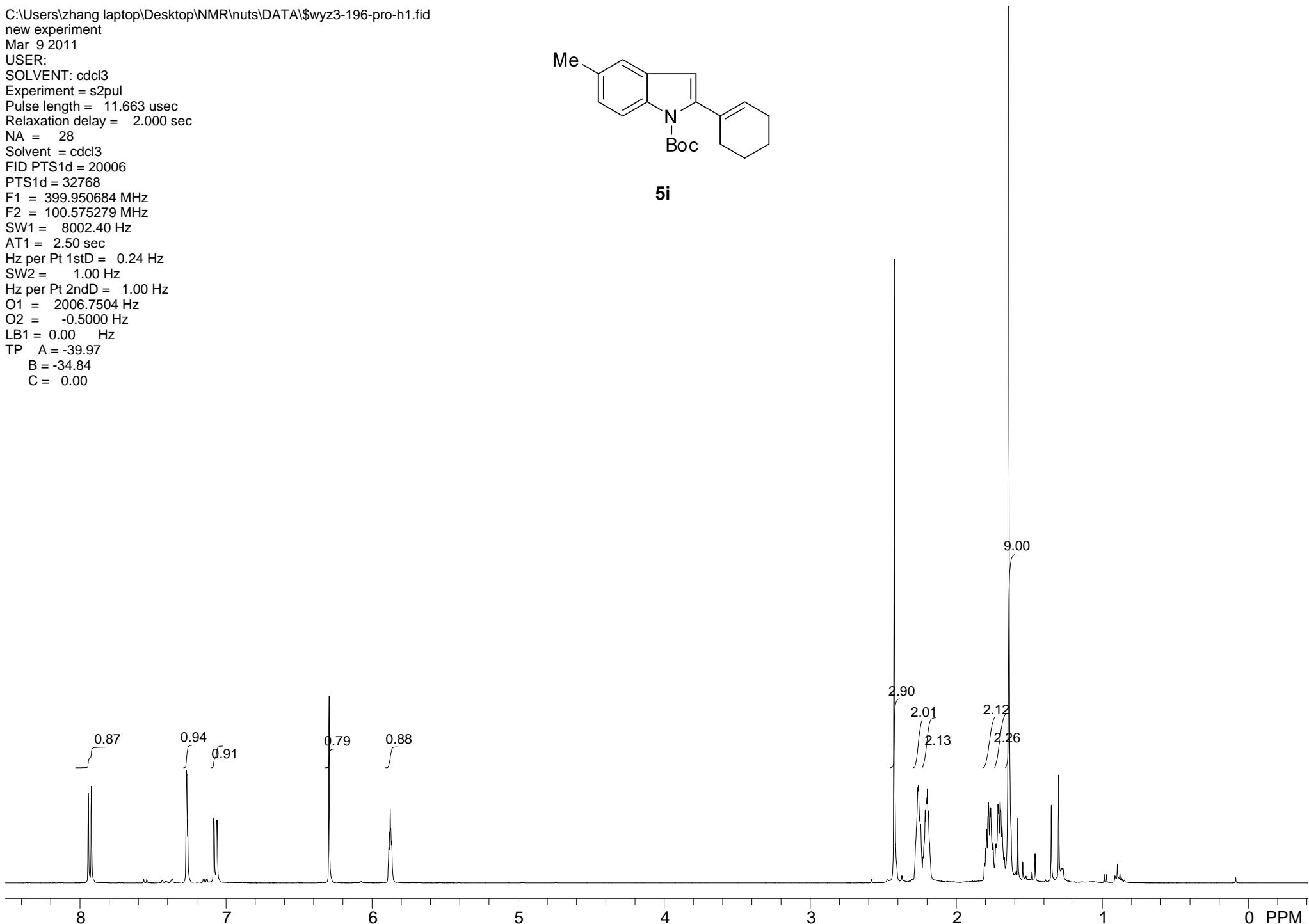
5h



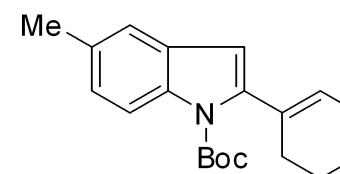
C:\Users\zhang laptop\Desktop\NMR\nuts\DATA\\$wyz3-196-pro-h1.fid
new experiment
Mar 9 2011
USER:
SOLVENT: cdcl3
Experiment = s2pul
Pulse length = 11.663 usec
Relaxation delay = 2.000 sec
NA = 28
Solvent = cdcl3
FID PTS1d = 20006
PTS1d = 32768
F1 = 399.950684 MHz
F2 = 100.575279 MHz
SW1 = 8002.40 Hz
AT1 = 2.50 sec
Hz per Pt 1stD = 0.24 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 2006.7504 Hz
O2 = -0.5000 Hz
LB1 = 0.00 Hz
TP A = -39.97
B = -34.84
C = 0.00



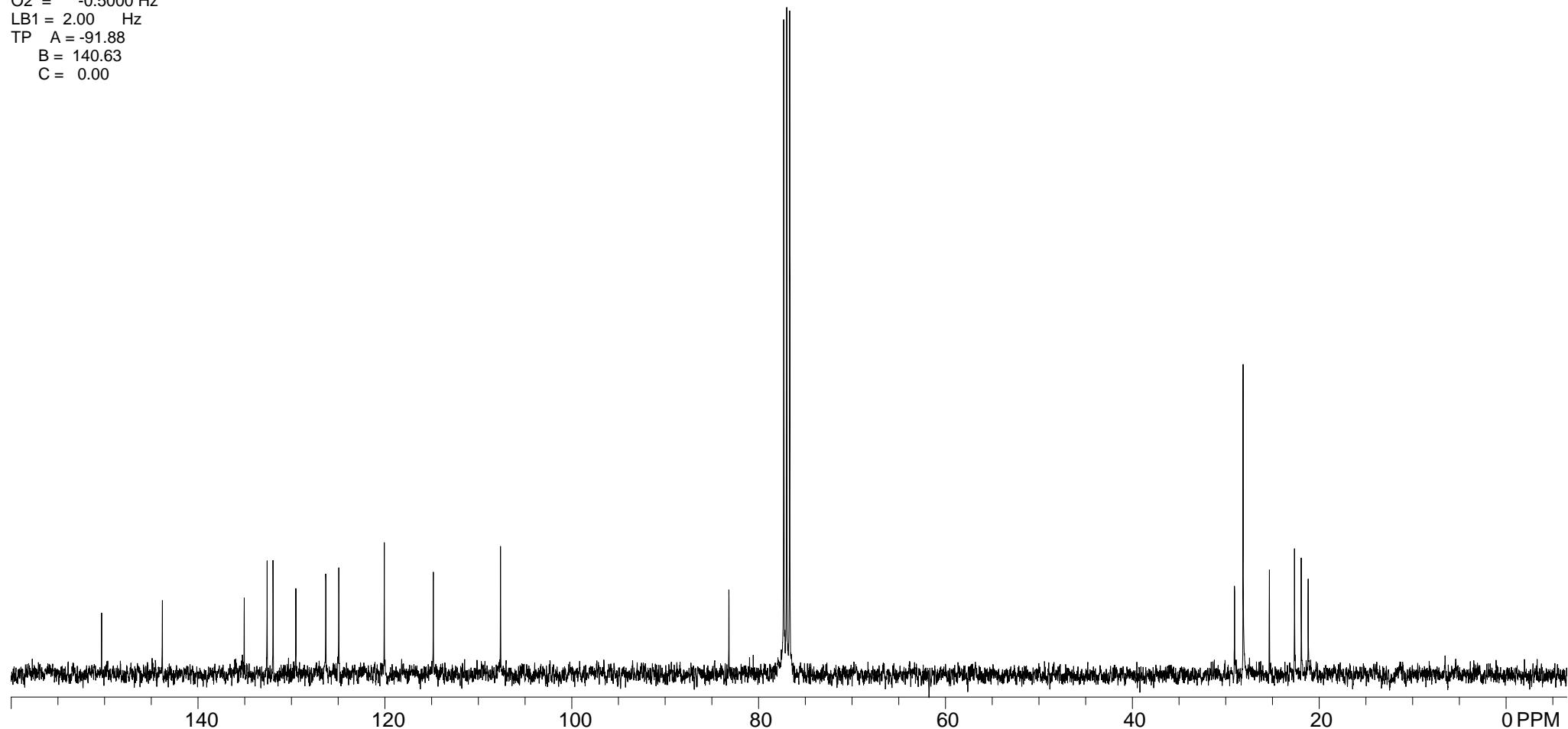
5i



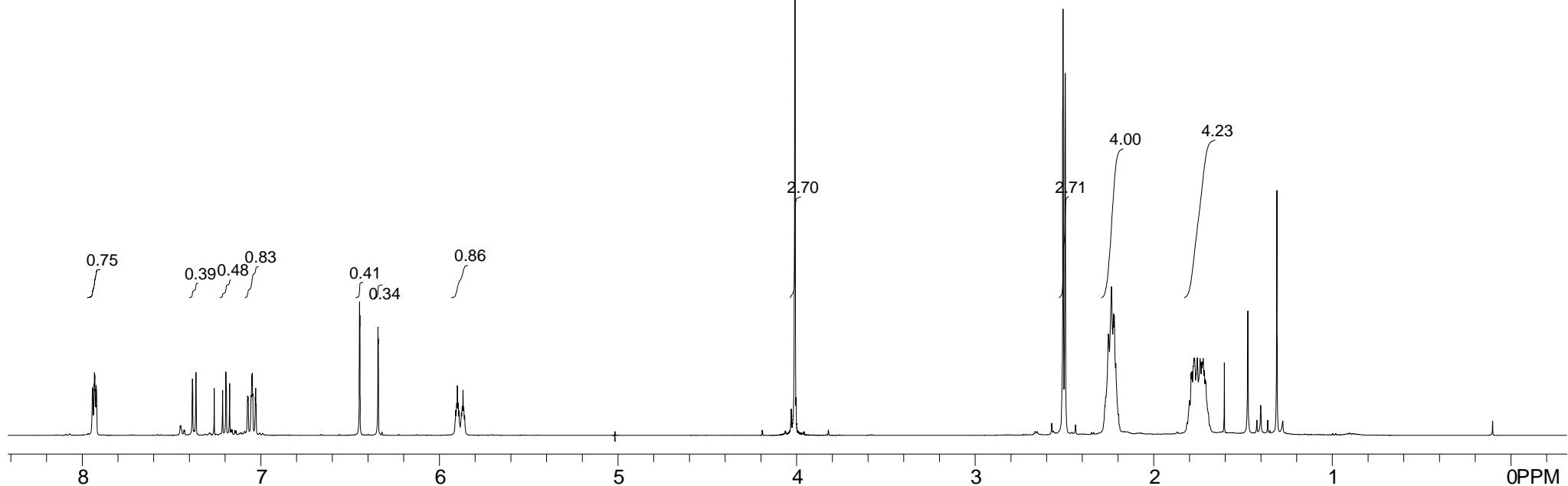
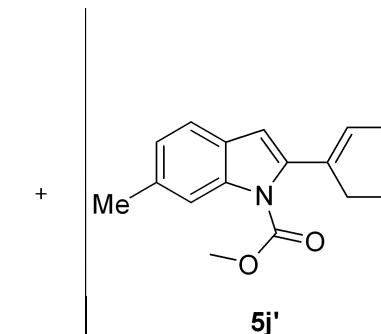
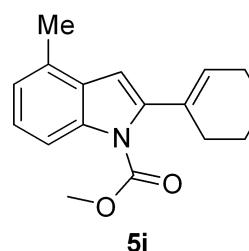
C:\Users\zhang laptop\Desktop\NMR\nuts\DATA\\$wyz3-196-pro-c13.fid
Standard c13 run using qnp probe
Mar. 9 2011
USER:
SOLVENT: cdcl3
Experiment = s2pul
Pulse length = 9.500 usec
Relaxation delay = 5.000 sec
NA = 128
Solvent = cdcl3
FID PTS1d = 16000
PTS1d = 16384
F1 = 100.576706 MHz
F2 = 399.949585 MHz
SW1 = 25000.00 Hz
AT1 = 0.64 sec
Hz per Pt 1stD = 1.53 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 9529.8262 Hz
O2 = -0.5000 Hz
LB1 = 2.00 Hz
TP A = -91.88
B = 140.63
C = 0.00



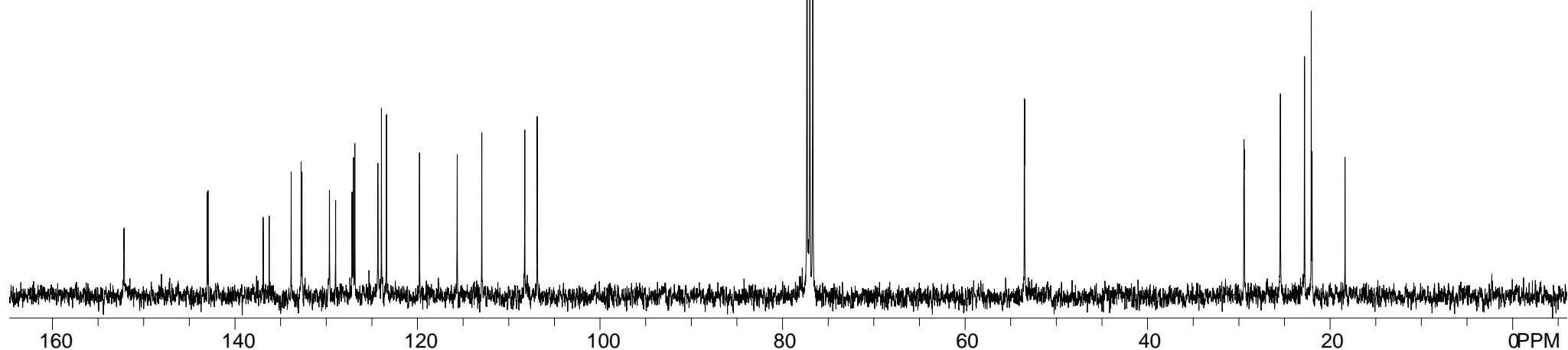
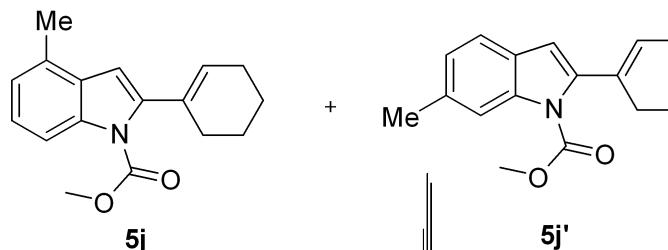
5i



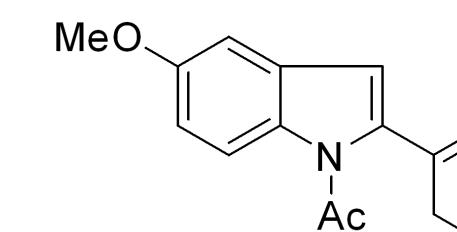
C:\Users\zhang laptop\Desktop\NMR\nuts\DATA\\$wyz3-214-pro-h1.fid
new experiment
Mar 22 2011
USER:
SOLVENT: cdcl3
Experiment = s2pul
Pulse length = 11.663 usec
Relaxation delay = 2.000 sec
NA = 18
Solvent = cdcl3
FID PTS1d = 20006
PTS1d = 32768
F1 = 399.950684 MHz
F2 = 100.575279 MHz
SW1 = 8002.40 Hz
AT1 = 2.50 sec
Hz per Pt 1stD = 0.24 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 2006.7504 Hz
O2 = -0.5000 Hz
LB1 = 0.00 Hz
TP A = -44.58
B = -37.03
C = 0.00



C:\Users\zhang laptop\Desktop\NMR\nuts\DATA\\$wyz3-214-pro-c13.fid
Standard c13 run using qnp probe
Mar 22 2011
USER:
SOLVENT: cdcl3
Experiment = s2pul
Pulse length = 9.500 usec
Relaxation delay = 5.000 sec
NA = 160
Solvent = cdcl3
FID PTS1d = 16000
PTS1d = 16384
F1 = 100.576706 MHz
F2 = 399.949585 MHz
SW1 = 25000.00 Hz
AT1 = 0.64 sec
Hz per Pt 1stD = 1.53 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 9526.7705 Hz
O2 = -0.5000 Hz
LB1 = 2.00 Hz
TP A = -133.13
B = 205.31
C = 0.00



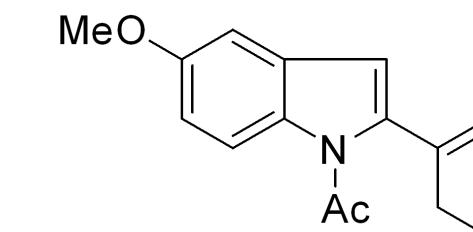
D:\Greenware\Ntus 20080731\DATA\\$wyz3-245-pro-h1.fid
new experiment
Apr 19 2011
USER:
SOLVENT: cdcl3
Experiment = s2pul
Pulse length = 11.663 usec
Relaxation delay = 2.000 sec
NA = 20
Solvent = cdcl3
FID PTS1d = 20006
PTS1d = 32768
F1 = 399.950684 MHz
F2 = 100.575279 MHz
SW1 = 8002.40 Hz
AT1 = 2.50 sec
Hz per Pt 1stD = 0.24 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 2006.2362 Hz
O2 = -0.5000 Hz
LB1 = 0.00 Hz
TP A = -45.83
B = -29.73
C = 0.00



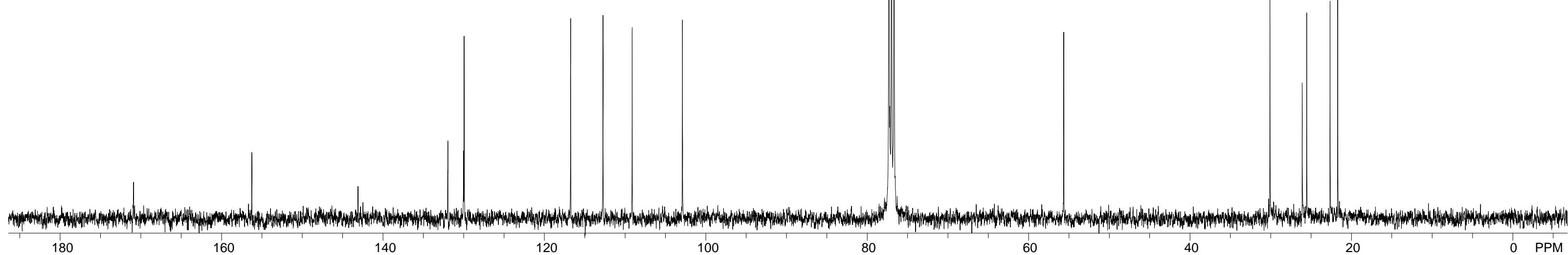
5k



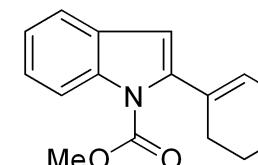
D:\Greenware\Ntus 20080731\DATA\\$wyz3-245-pro-c13.fid
Standard c13 run using qnp probe
Apr 19 2011
USER:
SOLVENT: cdcl3
Experiment = s2pul
Pulse length = 7.775 usec
Relaxation delay = 1.300 sec
NA = 3300
Solvent = cdcl3
FID PTS1d = 28040
PTS1d = 32768
F1 = 100.577232 MHz
F2 = 399.950684 MHz
SW1 = 28040.66 Hz
AT1 = 1.00 sec
Hz per Pt 1stD = 0.86 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 10057.2168 Hz
O2 = -0.5000 Hz
LB1 = 2.00 Hz
TP A = -8.44
B = -23.91
C = 0.00



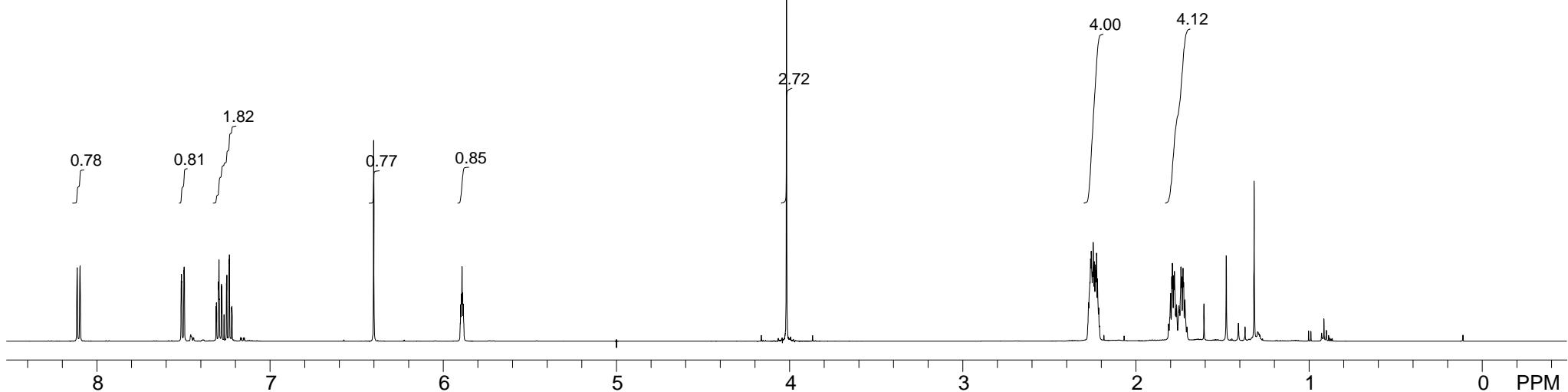
5k



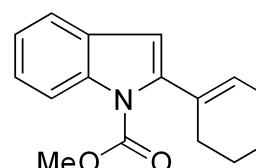
C:\Users\zhang laptop\Desktop\NMR\nuts\DATA\\$wyz3-182-pro-h1.fid
H1_CDCL3
Mar. 2 2011
USER:
SOLVENT: CDCl₃
Experiment = s2pul
Pulse length = 7.700 usec
Relaxation delay = 2.000 sec
NA = 18
Solvent = CDCl₃
FID PTS1d = 21259
PTS1d = 32768
F1 = 499.858551 MHz
F2 = 1.000000 MHz
SW1 = 8503.40 Hz
AT1 = 2.50 sec
Hz per Pt 1stD = 0.26 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 2499.2932 Hz
O2 = -0.5000 Hz
LB1 = 0.00 Hz
TP A = 71.41
B = -9.57
C = 0.00



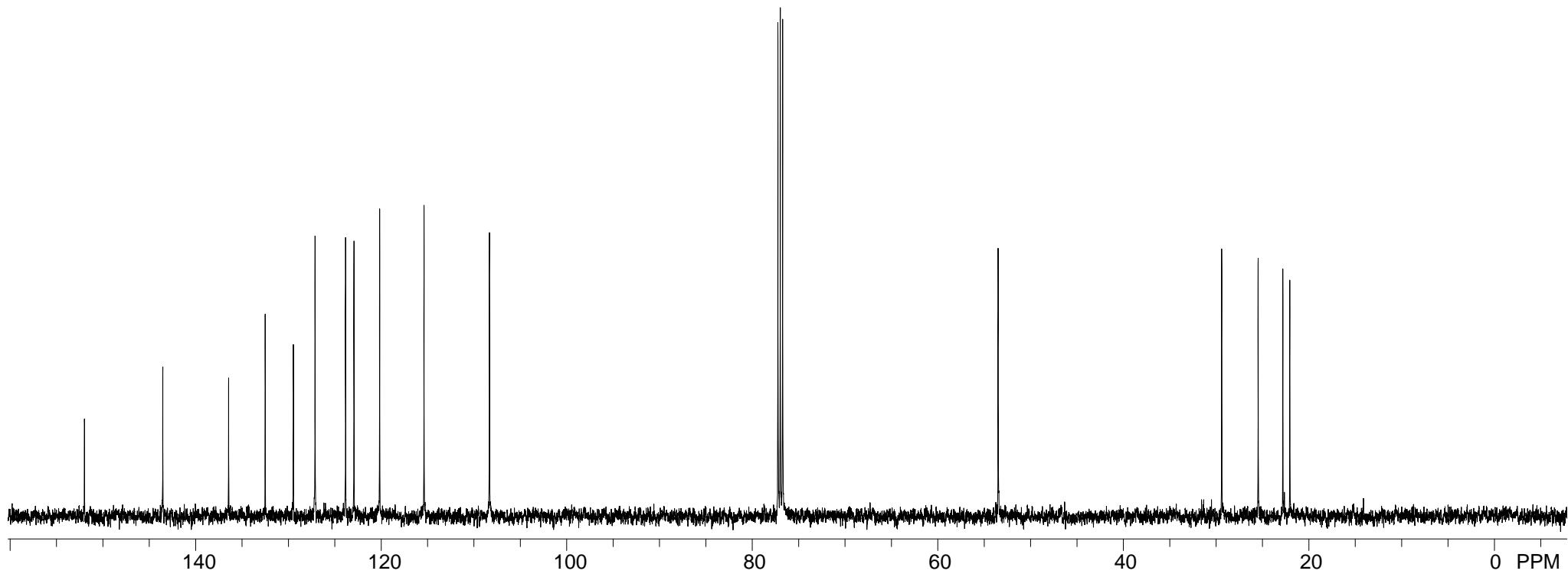
5l



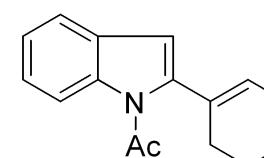
C:\Users\zhang laptop\Desktop\NMR\nuts\DATA\\$wyz3-182-pro-c13-1.fid
STANDARD CARBON PARAMETERS
Mar. 2 2011
USER:
SOLVENT: cdcl3
Experiment = s2pul
Pulse length = 6.000 usec
Relaxation delay = 3.000 sec
NA = 52
Solvent = cdcl3
FID PTS1d = 36749
PTS1d = 65536
F1 = 125.701683 MHz
F2 = 499.858551 MHz
SW1 = 28258.57 Hz
AT1 = 1.30 sec
Hz per Pt 1stD = 0.43 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 12766.5020 Hz
O2 = -0.5000 Hz
LB1 = 2.00 Hz
TP A = -190.31
B = 135.00
C = 0.00



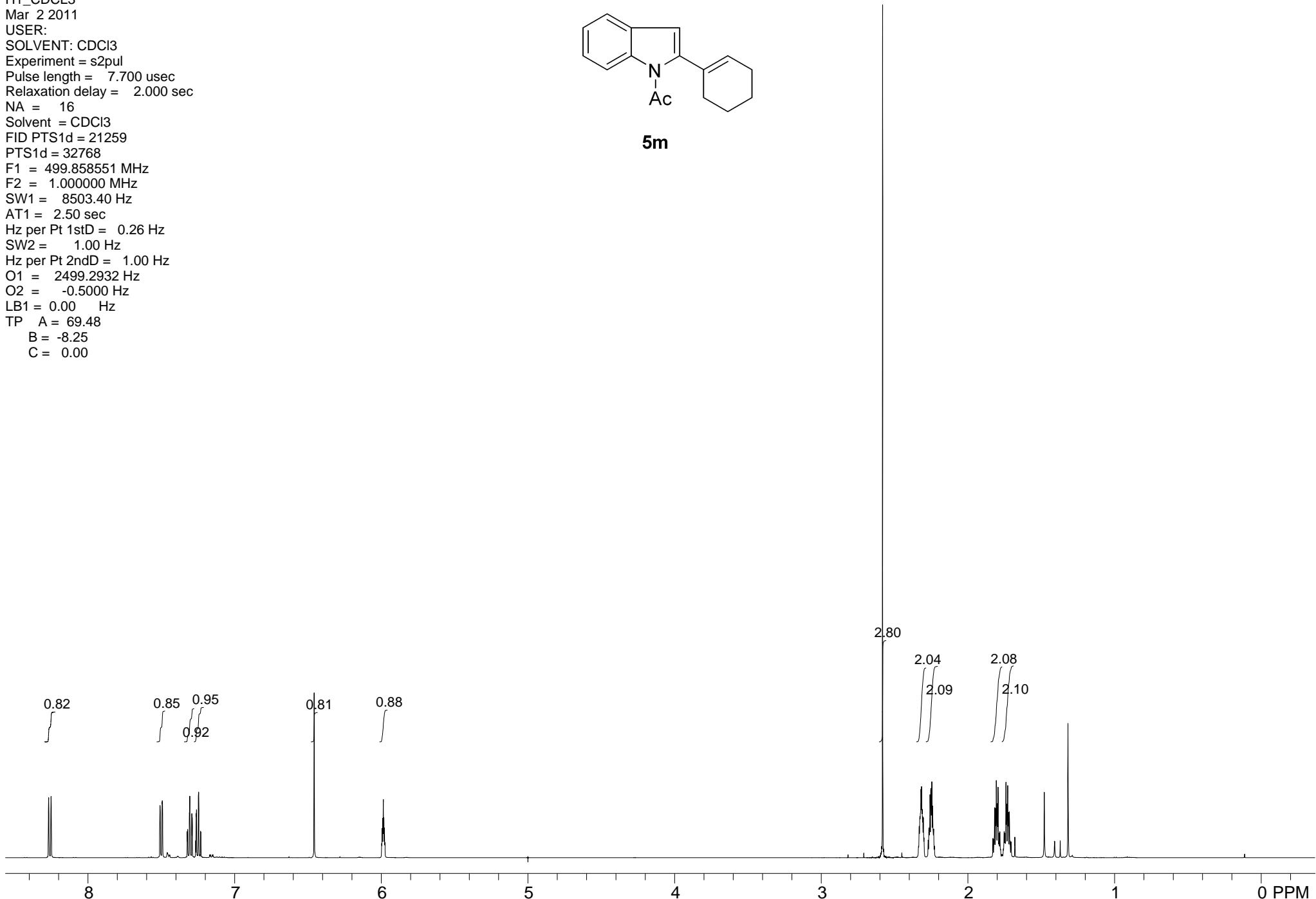
5l



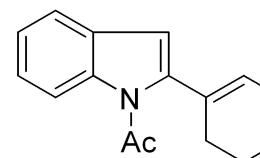
C:\Users\zhang laptop\Desktop\NMR\nuts\DATA\\$wyz3-183-pro-h1.fid
H1_CDCL3
Mar. 2 2011
USER:
SOLVENT: CDCl₃
Experiment = s2pul
Pulse length = 7.700 usec
Relaxation delay = 2.000 sec
NA = 16
Solvent = CDCl₃
FID PTS1d = 21259
PTS1d = 32768
F1 = 499.858551 MHz
F2 = 1.000000 MHz
SW1 = 8503.40 Hz
AT1 = 2.50 sec
Hz per Pt 1stD = 0.26 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 2499.2932 Hz
O2 = -0.5000 Hz
LB1 = 0.00 Hz
TP A = 69.48
B = -8.25
C = 0.00



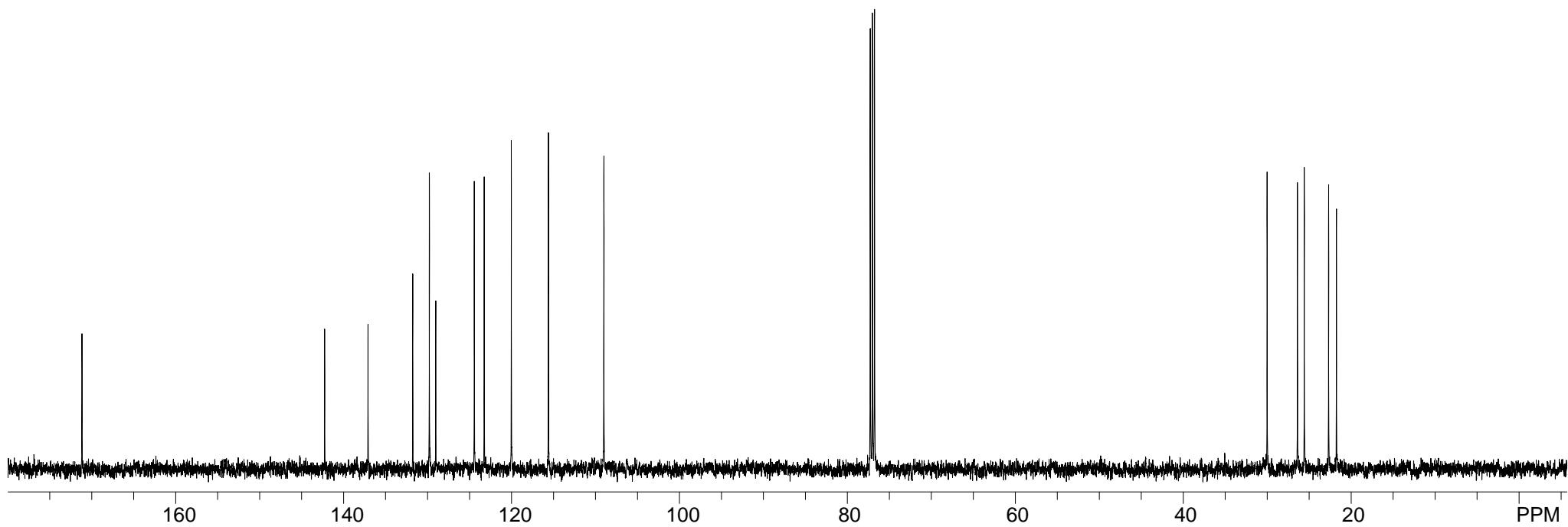
5m



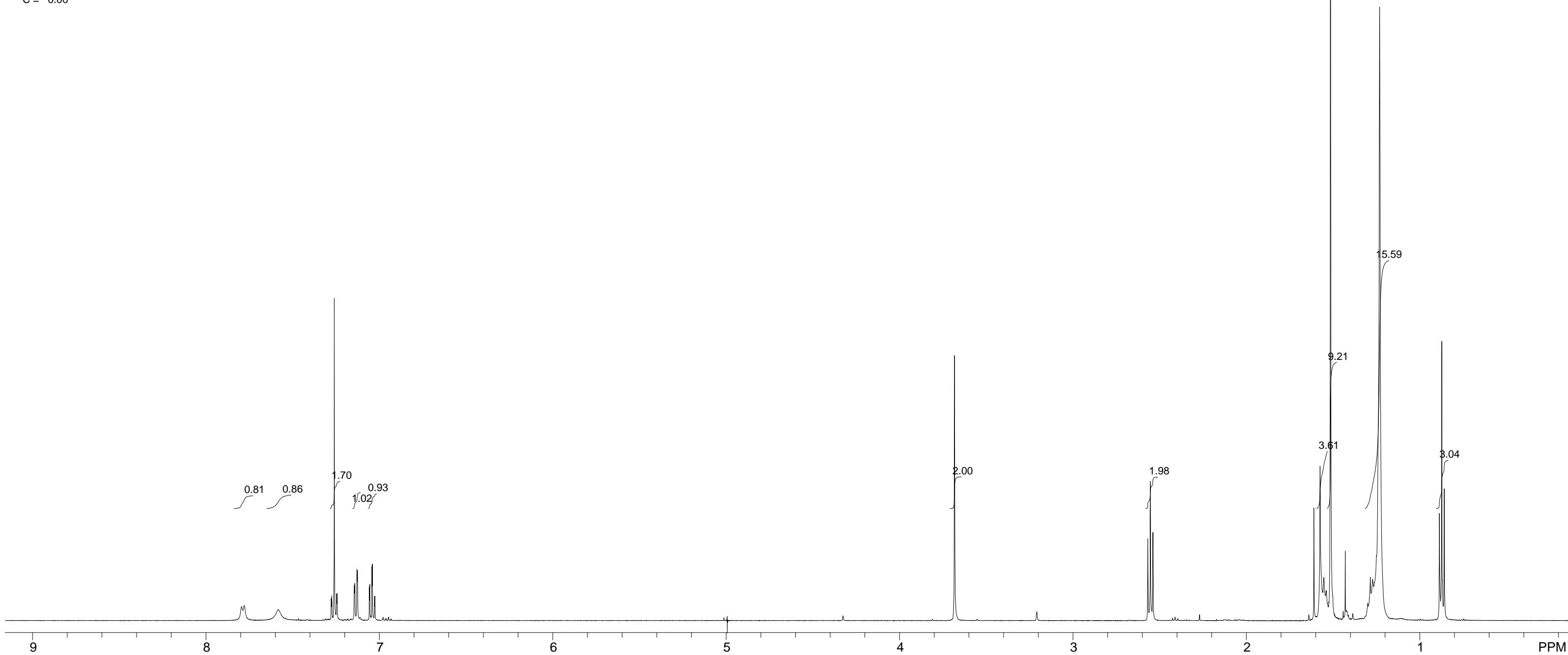
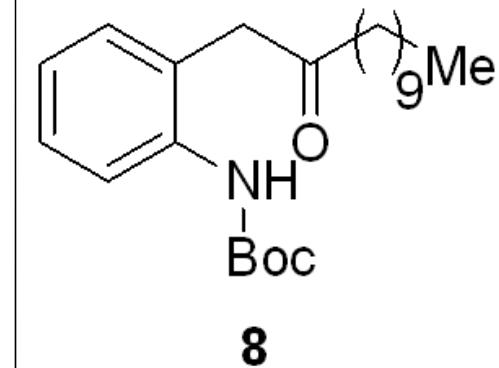
C:\Users\zhang laptop\Desktop\NMR\nuts\DATA\\$wyz3-183-pro-c13.fid
STANDARD CARBON PARAMETERS
Mar. 2 2011
USER:
SOLVENT: cdcl3
Experiment = s2pul
Pulse length = 6.000 usec
Relaxation delay = 3.000 sec
NA = 40
Solvent = cdcl3
FID PTS1d = 36749
PTS1d = 65536
F1 = 125.701683 MHz
F2 = 499.858551 MHz
SW1 = 28258.57 Hz
AT1 = 1.30 sec
Hz per Pt 1stD = 0.43 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 12764.3457 Hz
O2 = -0.5000 Hz
LB1 = 2.00 Hz
TP A = -202.50
B = 151.88
C = 0.00



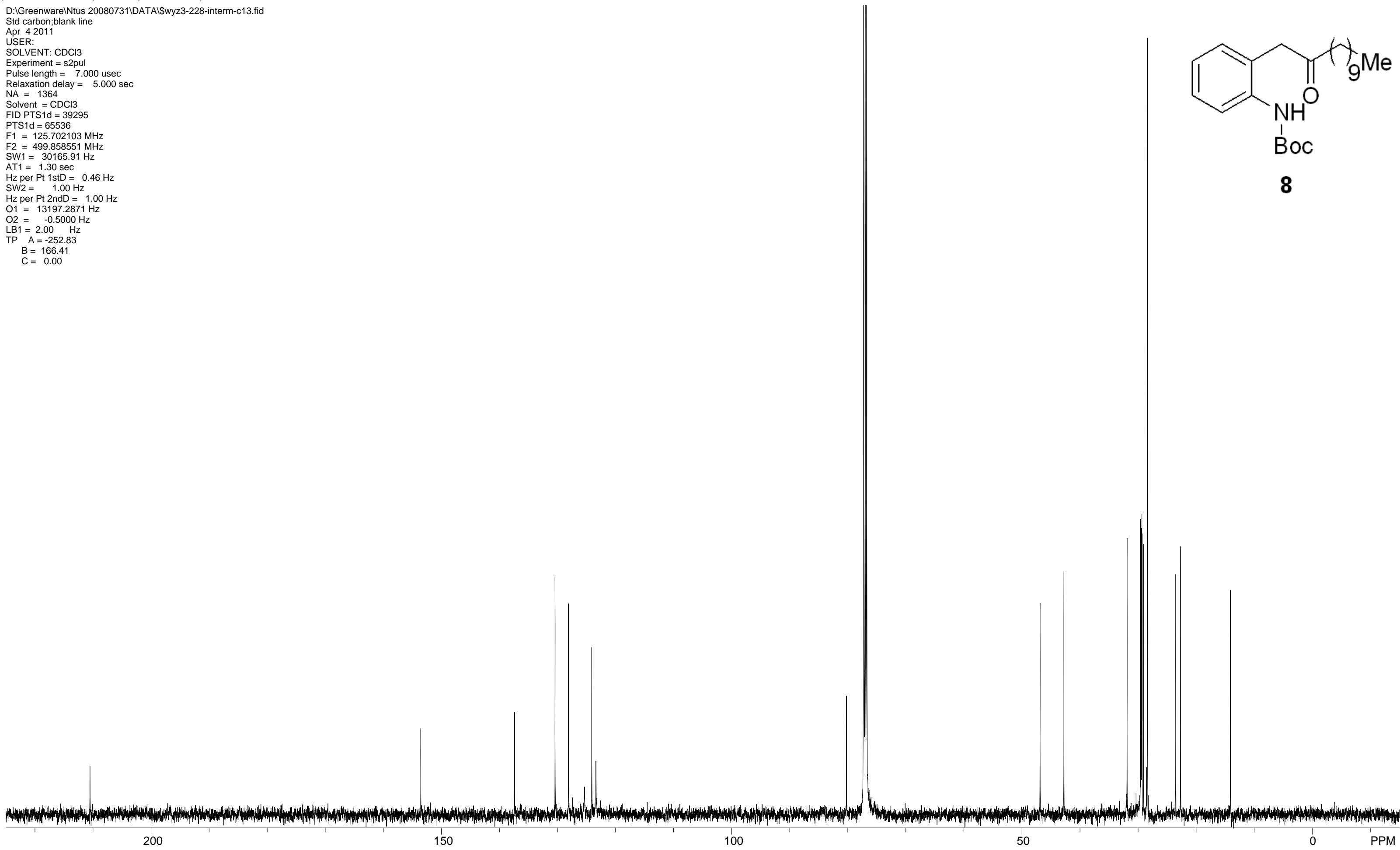
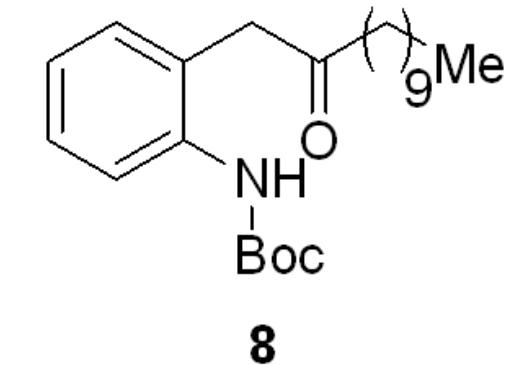
5m



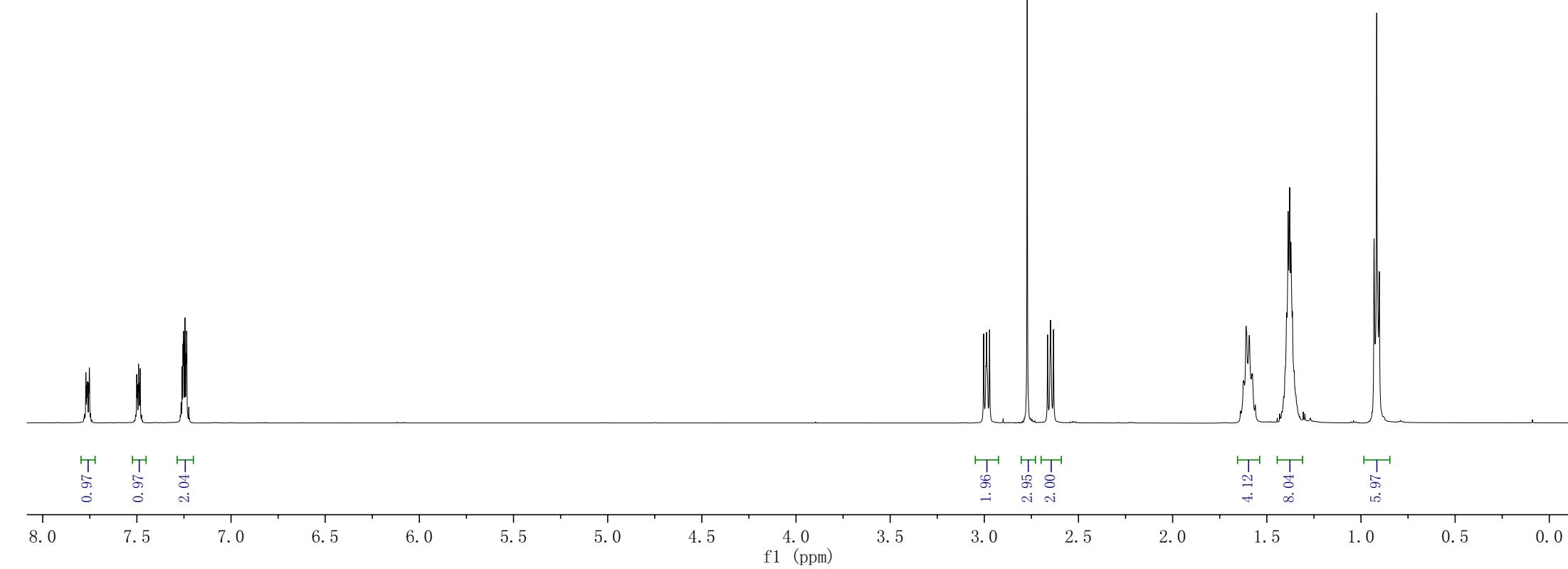
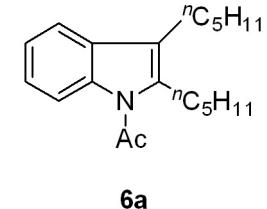
D:\Greenware\Ntus 20080731\DATA\\$wyz3-228-interm-h1.fid
H1_CDCL3
Apr 4 2011
USER:
SOLVENT: CDCl3
Experiment = s2pul
Pulse length = 7.700 usec
Relaxation delay = 2.000 sec
NA = 16
Solvent = CDCl3
FID PTS1d = 21259
PTS1d = 32768
F1 = 499.858551 MHz
F2 = 1.000000 MHz
SW1 = 8503.40 Hz
AT1 = 2.50 sec
Hz per Pt 1stD = 0.26 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 2496.3191 Hz
O2 = -0.5000 Hz
LB1 = 0.00 Hz
TP A = 64.22
B = -11.95
C = 0.00



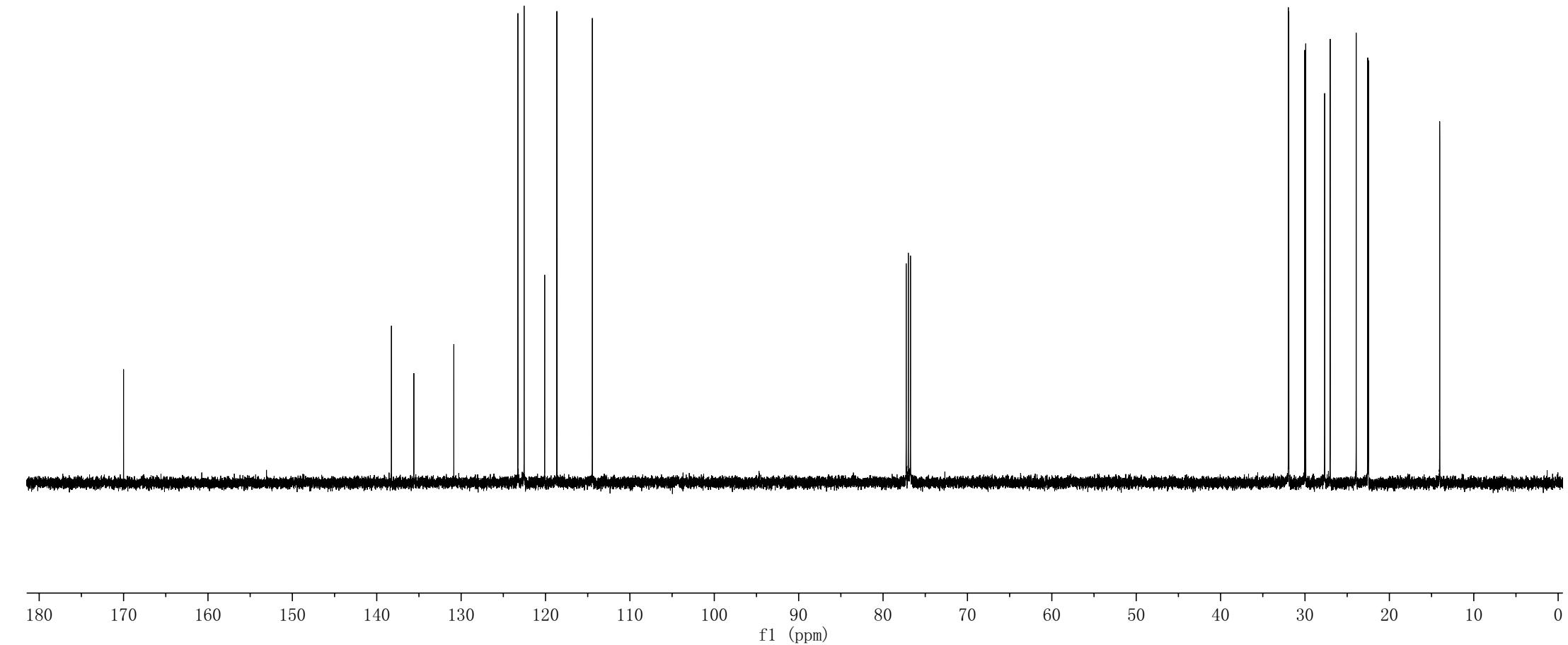
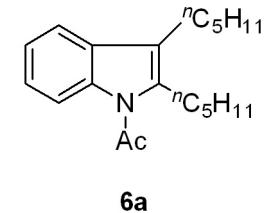
D:\Greenware\Ntus 20080731\DATA\\$wyz3-228-interm-c13.fid
Std carbon;blank line
Apr 4 2011
USER:
SOLVENT: CDCl₃
Experiment = s2pul
Pulse length = 7.000 usec
Relaxation delay = 5.000 sec
NA = 1364
Solvent = CDCl₃
FID PTS1d = 39295
PTS1d = 65536
F1 = 125.702103 MHz
F2 = 499.858551 MHz
SW1 = 30165.91 Hz
AT1 = 1.30 sec
Hz per Pt 1stD = 0.46 Hz
SW2 = 1.00 Hz
Hz per Pt 2ndD = 1.00 Hz
O1 = 13197.2871 Hz
O2 = -0.5000 Hz
LB1 = 2.00 Hz
TP A = -252.83
B = 166.41
C = 0.00



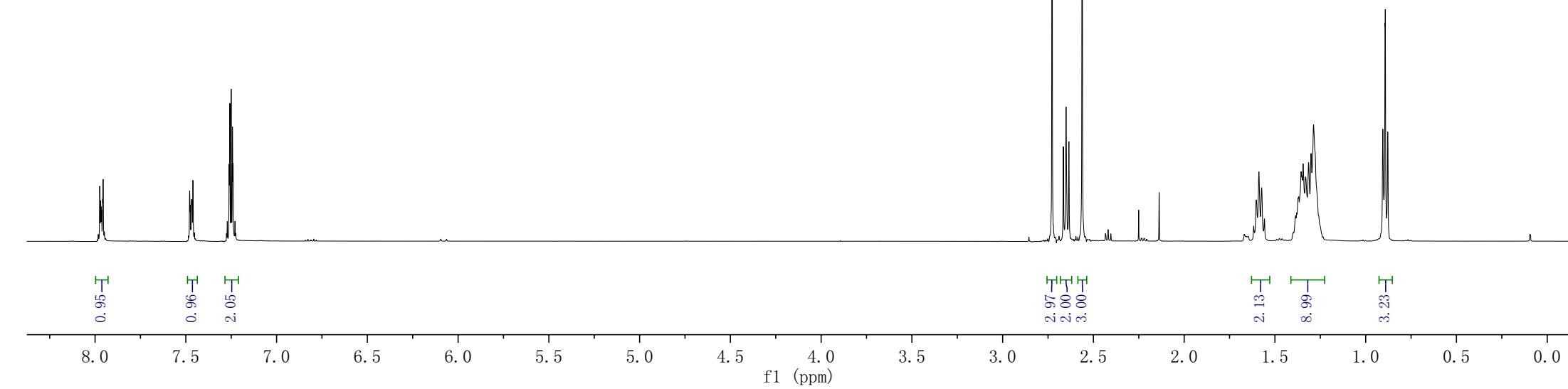
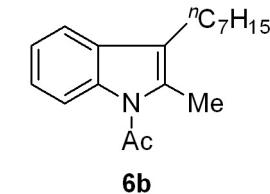
Parameter	Value
1 Data File Name	F:/ Yanzhao Wang Research/ Proj 2 Protected Indoles/ NMR Internal Indoles/ wyz5-96-pro-h1.fid/ fid
2 Title	wyz5-96-pro-h1
3 Solvent	CDCl ₃
4 Spectrometer Frequency	499.86
5 Nucleus	¹ H



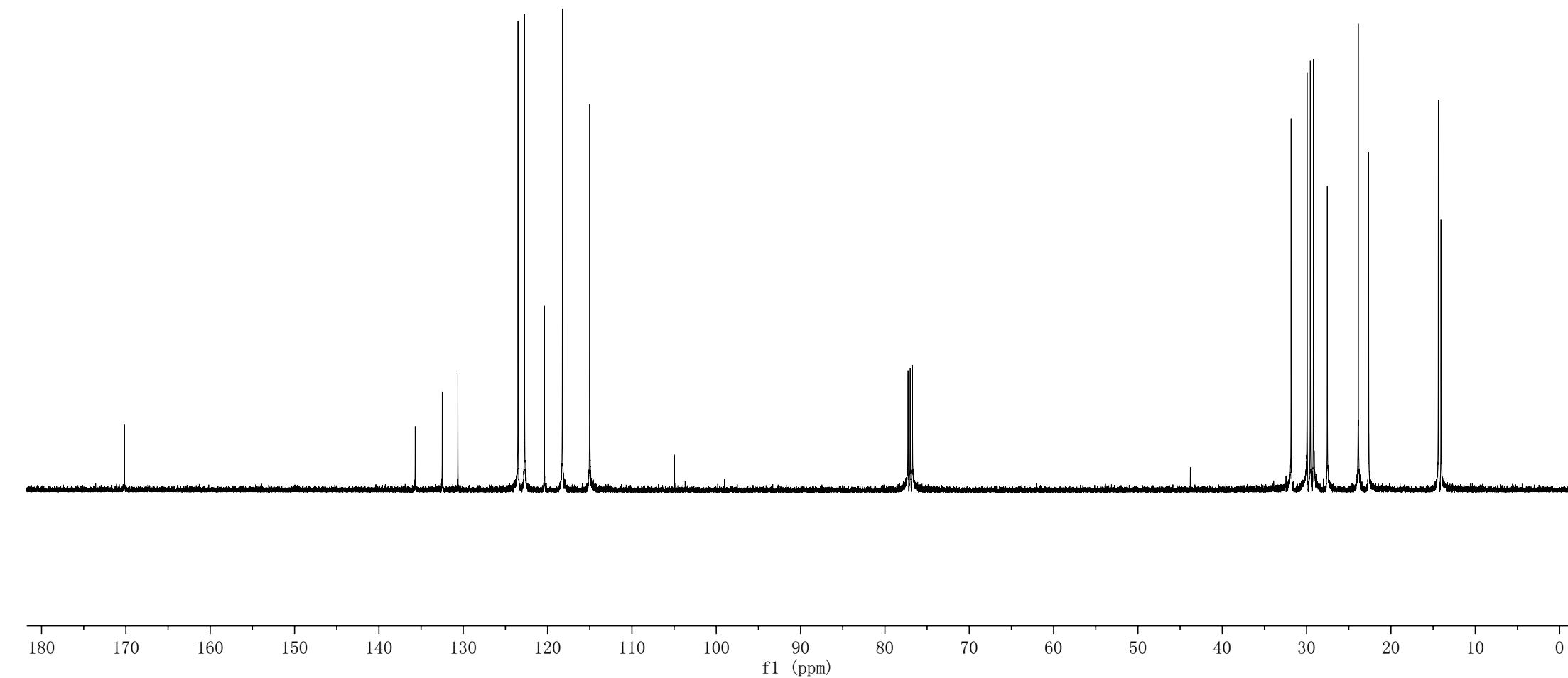
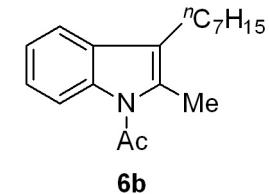
Parameter	Value
1 Data File Name	F:/ Yanzhao Wang Research/ Proj 2 Protected Indoles/ NMR Internal Indoles/ wyz5-96-pro-c13.fid/ fid
2 Title	wyz5-96-pro-c13
3 Solvent	CDCl ₃
4 Spectrometer Frequency	125.70
5 Nucleus	¹³ C



Parameter	Value
1 Data File Name	F:/ Yanzhao Wang Research/ Proj 2 Protected Indoles/ NMR Internal Indoles/ wyz5-103-pro1-h1.fid/ fid
2 Title	wyz5-103-pro1-h1
3 Solvent	CDCl ₃
4 Spectrometer Frequency	499.86
5 Nucleus	¹ H



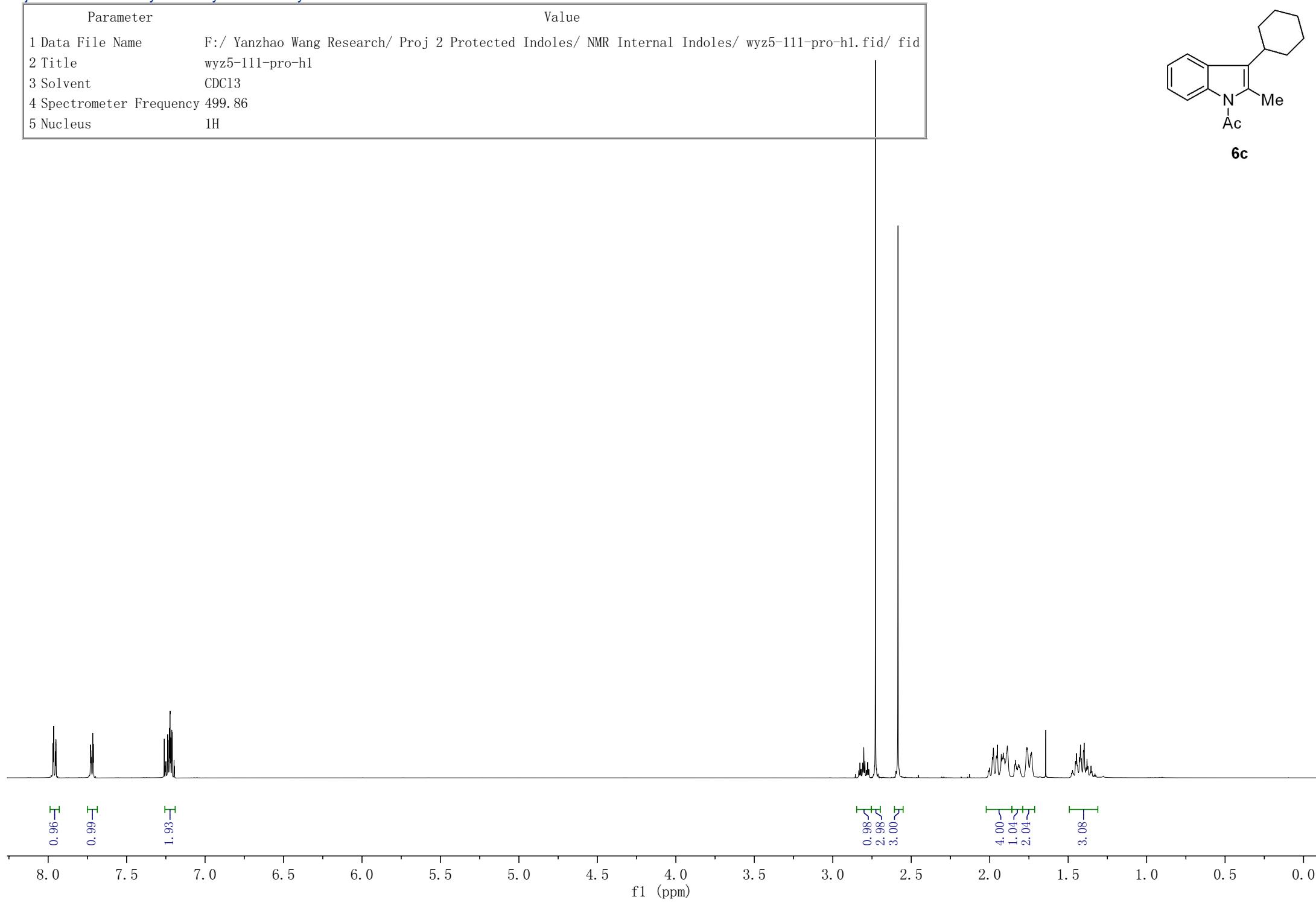
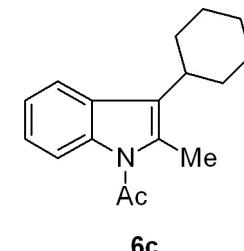
Parameter	Value
1 Data File Name	F:/ Yanzhao Wang Research/ Proj 2 Protected Indoles/ NMR Internal Indoles/ wyz5-103-pro1-c13.fid/ fid
2 Title	wyz5-103-pro1-c13
3 Solvent	CDCl ₃
4 Spectrometer Frequency	125.70
5 Nucleus	¹³ C



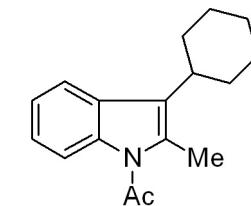
Parameter

Value

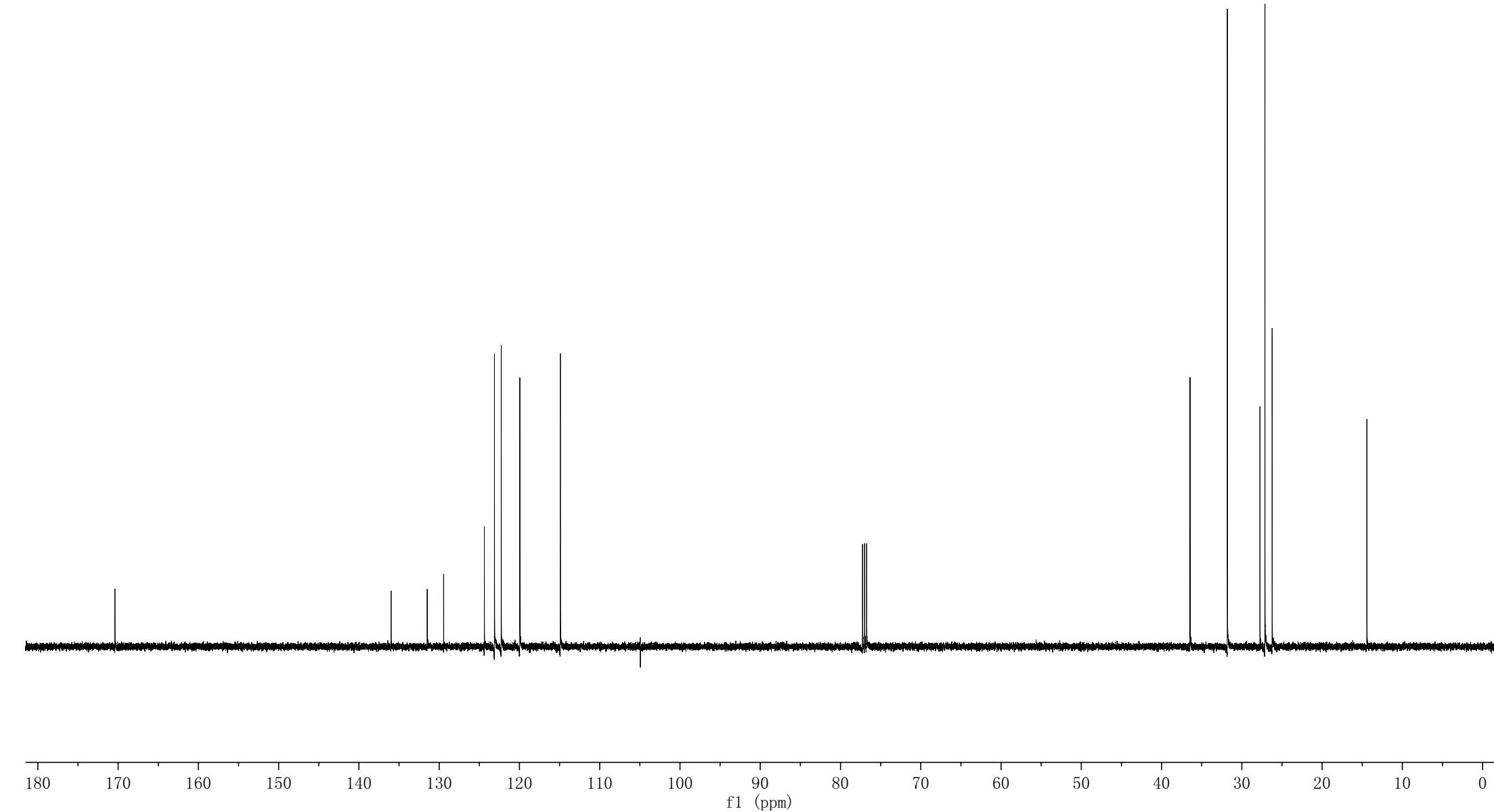
1 Data File Name	F:/ Yanzhao Wang Research/ Proj 2 Protected Indoles/ NMR Internal Indoles/ wyz5-111-pro-h1.fid/ fid
2 Title	wyz5-111-pro-h1
3 Solvent	CDCl ₃
4 Spectrometer Frequency	499.86
5 Nucleus	¹ H



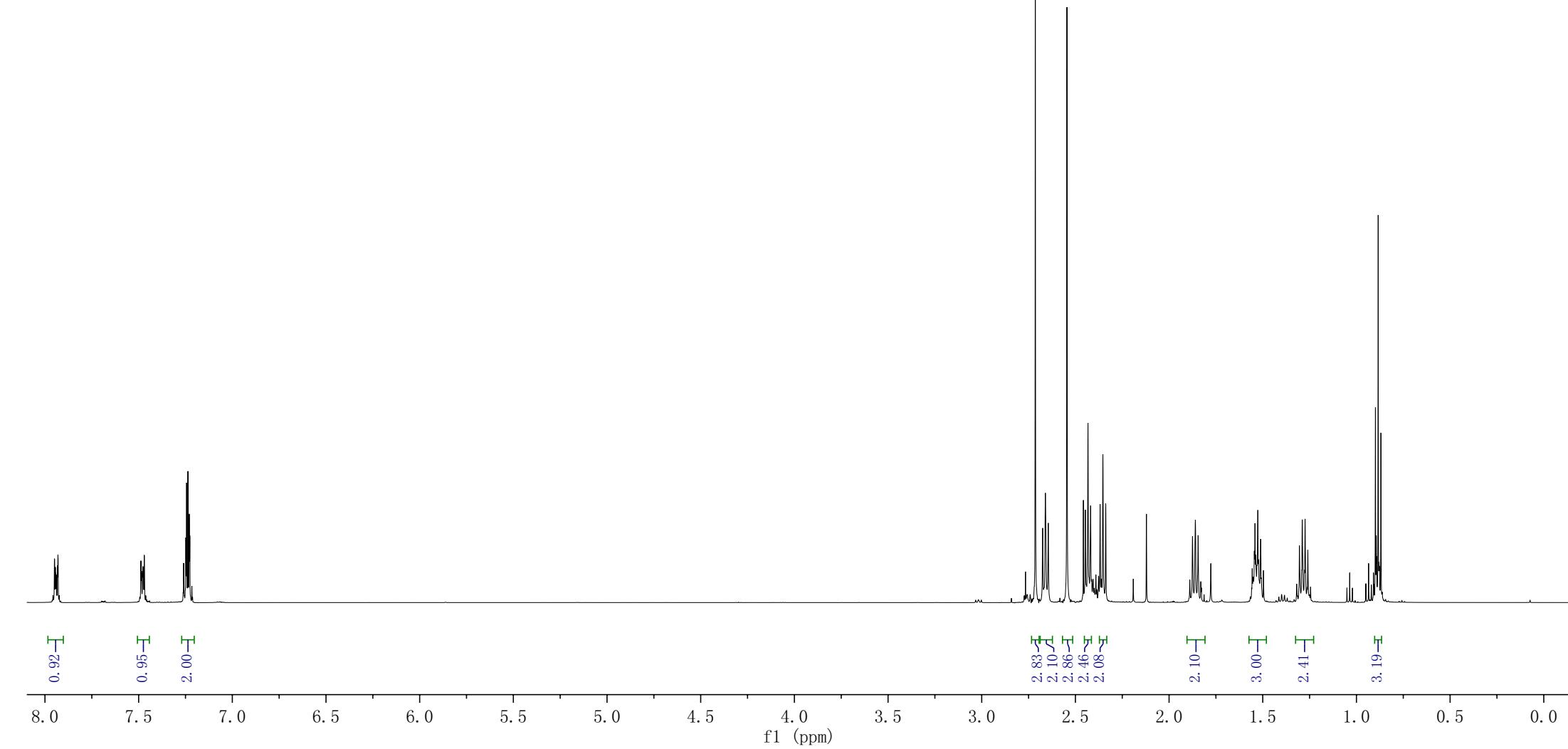
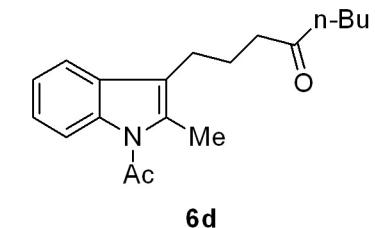
Parameter	Value
1 Data File Name	F:/ Yanzhao Wang Research/ Proj 2 Protected Indoles/ NMR Internal Indoles/ wyz5-111-pro-c13.fid/ fid
2 Title	wyz5-111-pro-c13
3 Solvent	CDCl ₃
4 Spectrometer Frequency	125.70
5 Nucleus	¹³ C



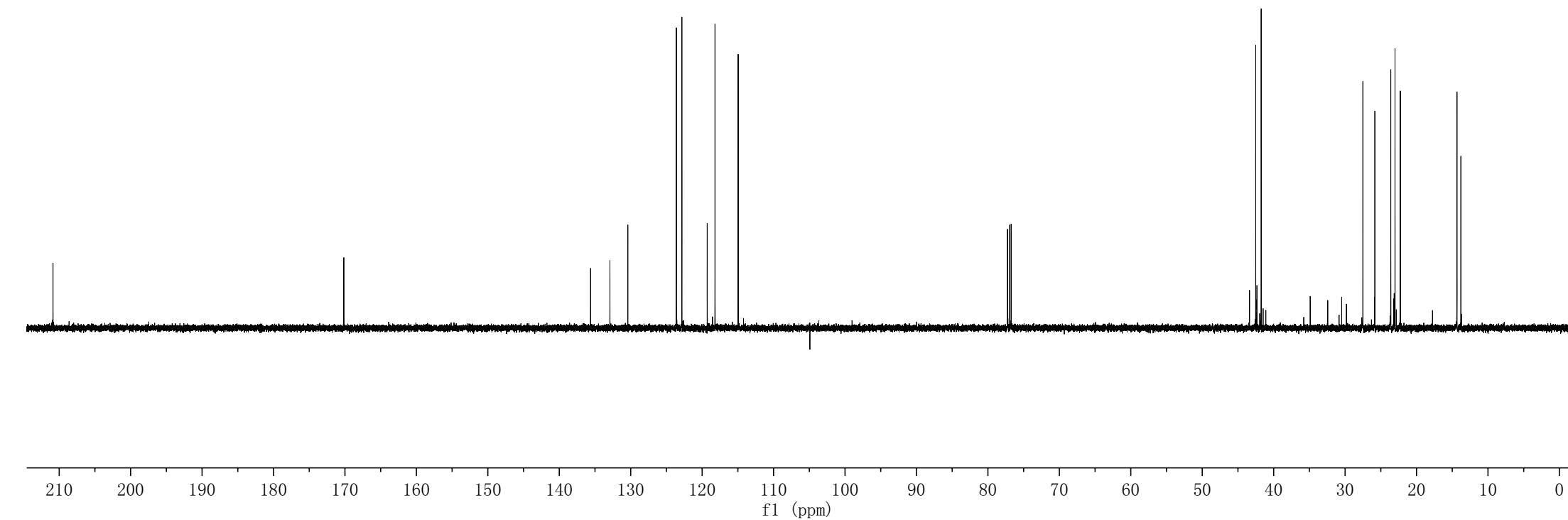
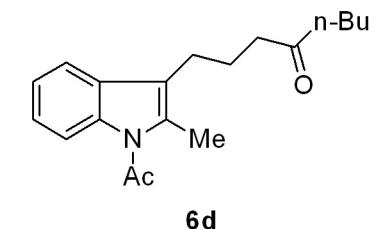
6c



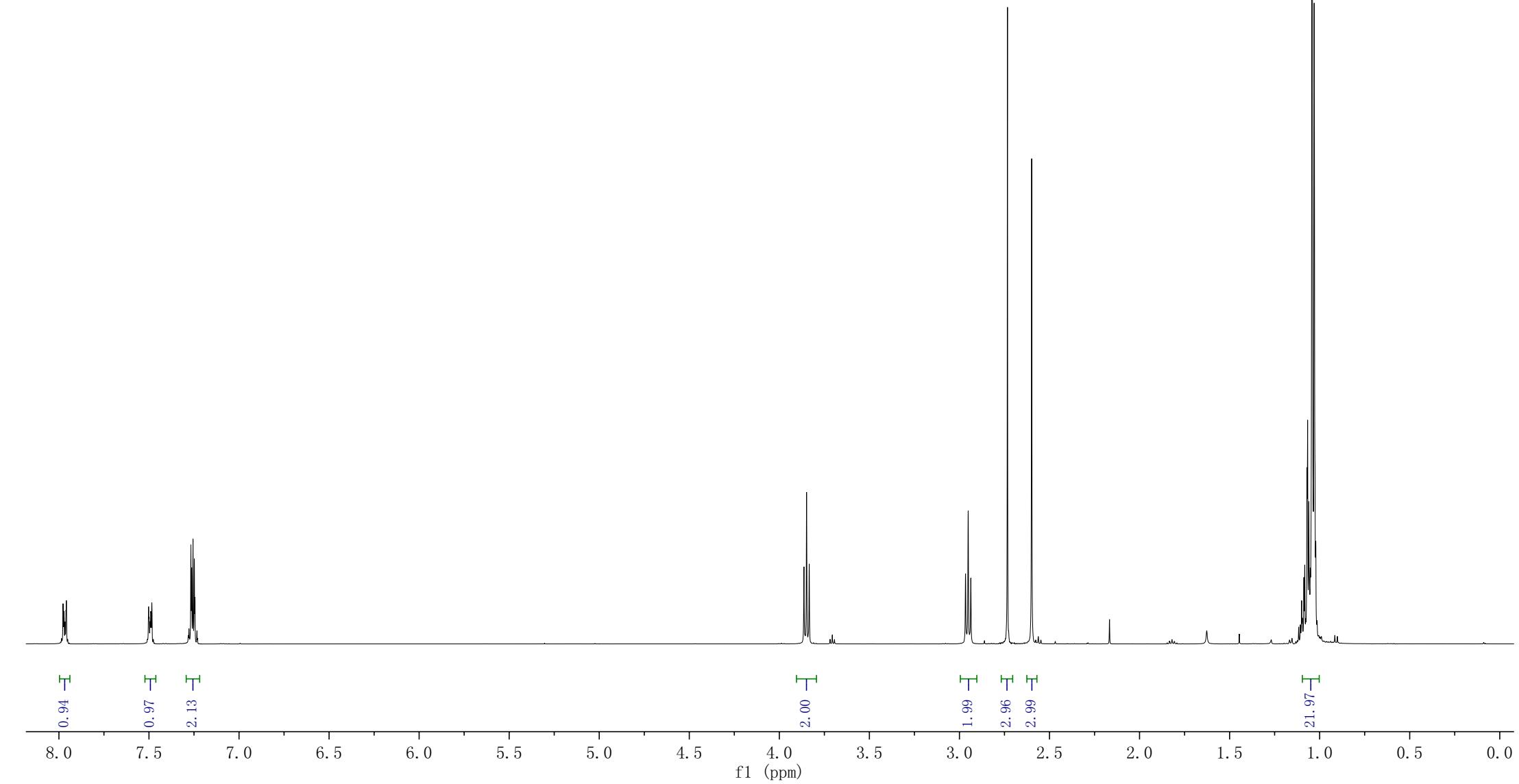
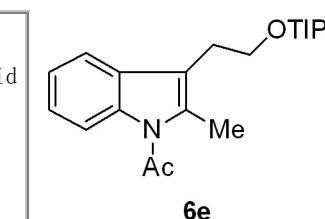
Parameter	Value
1 Data File Name	F:/ Yanzhao Wang Research/ Proj 2 Protected Indoles/ NMR Internal Indoles/ wyz5-104-pro1-h1.fid/ fid
2 Title	wyz5-104-pro1-h1
3 Solvent	CDCl ₃
4 Spectrometer Frequency	499.86
5 Nucleus	¹ H



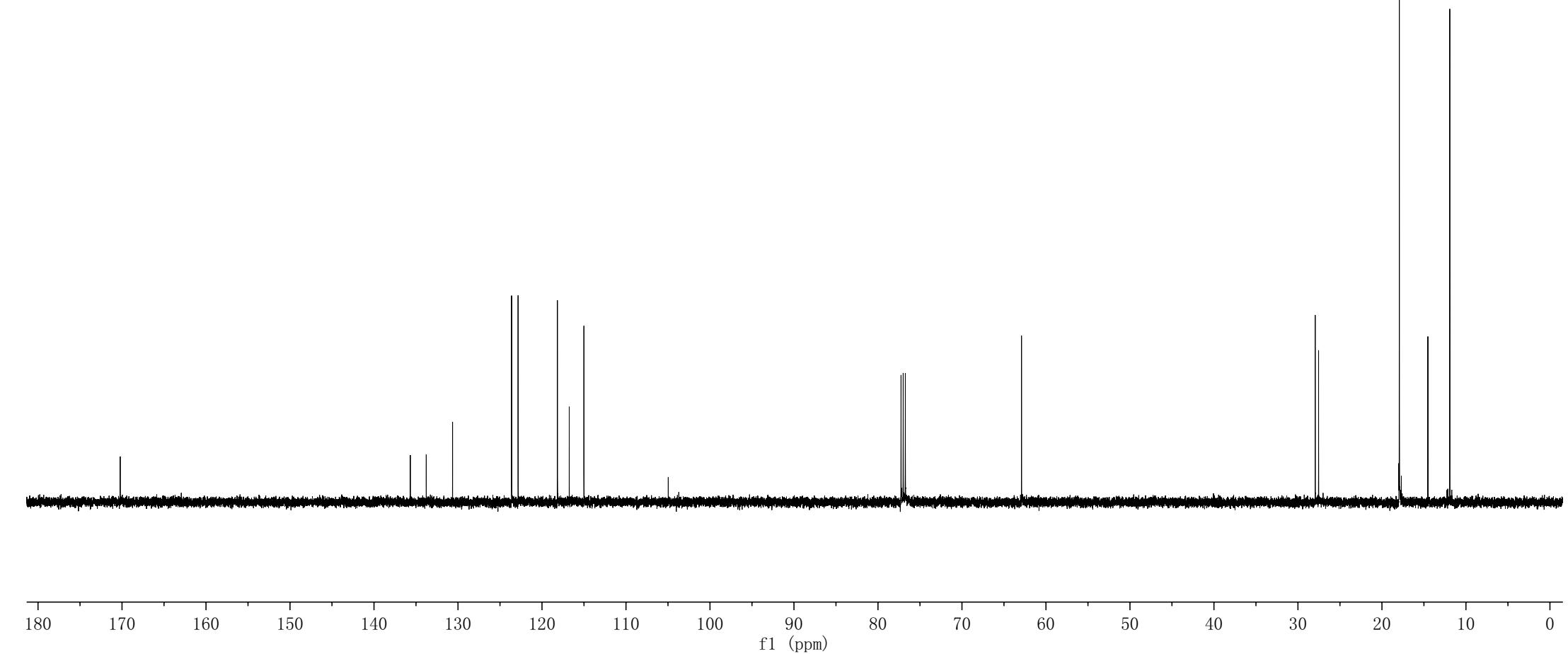
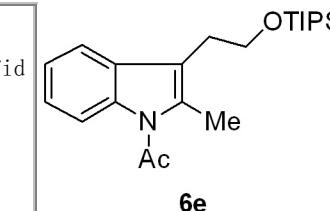
Parameter	Value
1 Data File Name	F:/ Yanzhao Wang Research/ Proj 2 Protected Indoles/ NMR Internal Indoles/ wyz5-104-pro1-c13.fid/ fid
2 Title	wyz5-104-pro1-c13
3 Solvent	CDCl ₃
4 Spectrometer Frequency	125.70
5 Nucleus	¹³ C



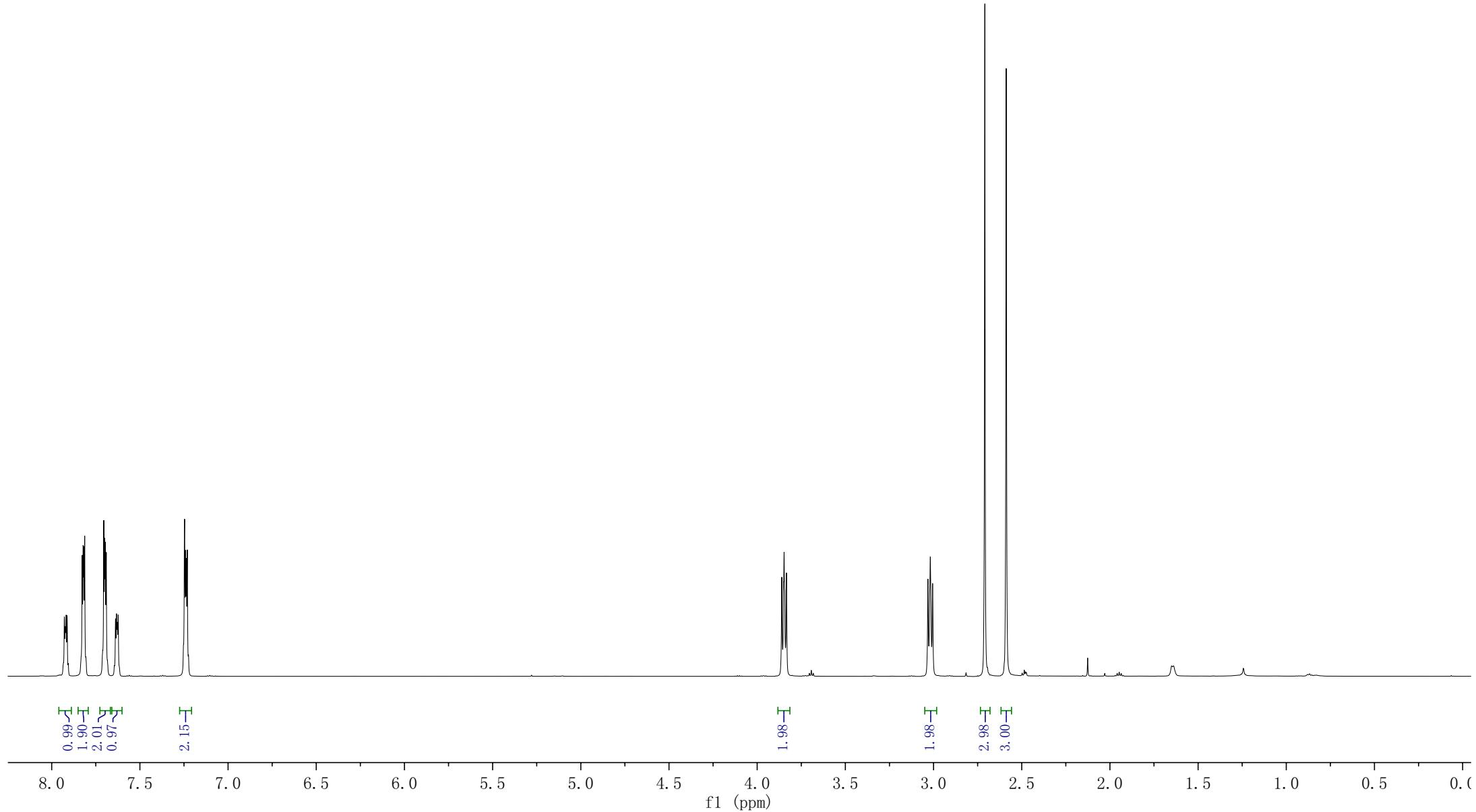
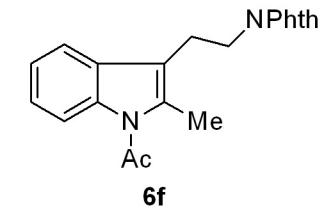
Parameter	Value
1 Data File Name	F:/ Yanzhao Wang Research/ Proj 2 Protected Indoles/ NMR Internal Indoles/ wyz5-107-pro1-h1.fid/ fid
2 Title	wyz5-107-pro1-h1
3 Solvent	CDCl ₃
4 Spectrometer Frequency	499.86
5 Nucleus	¹ H



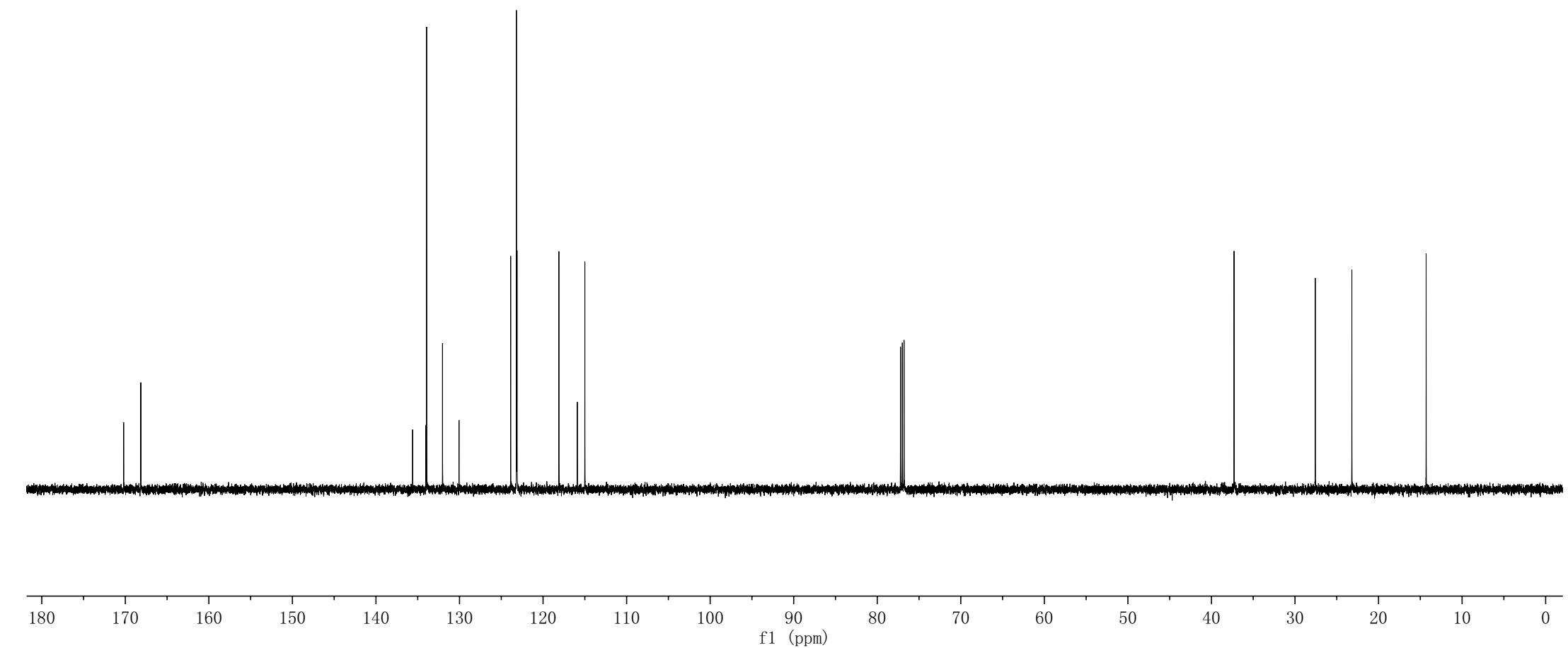
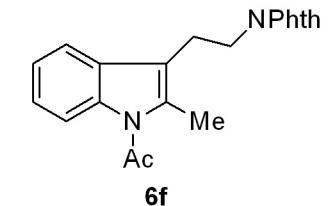
Parameter	Value
1 Data File Name	F:/ Yanzhao Wang Research/ Proj 2 Protected Indoles/ NMR Internal Indoles/ wyz5-107-pro1-c13.fid/ fid
2 Title	wyz5-107-pro1-c13
3 Solvent	CDCl ₃
4 Spectrometer Frequency	125.70
5 Nucleus	¹³ C



Parameter	Value
1 Data File Name	F:/ Yanzhao Wang Research/ Proj 2 Protected Indoles/ NMR Internal Indoles/ wyz5-106-pro-h1.fid/ fid
2 Title	wyz5-106-pro-h1
3 Solvent	cdcl3
4 Spectrometer Frequency	599. 63
5 Nucleus	1H



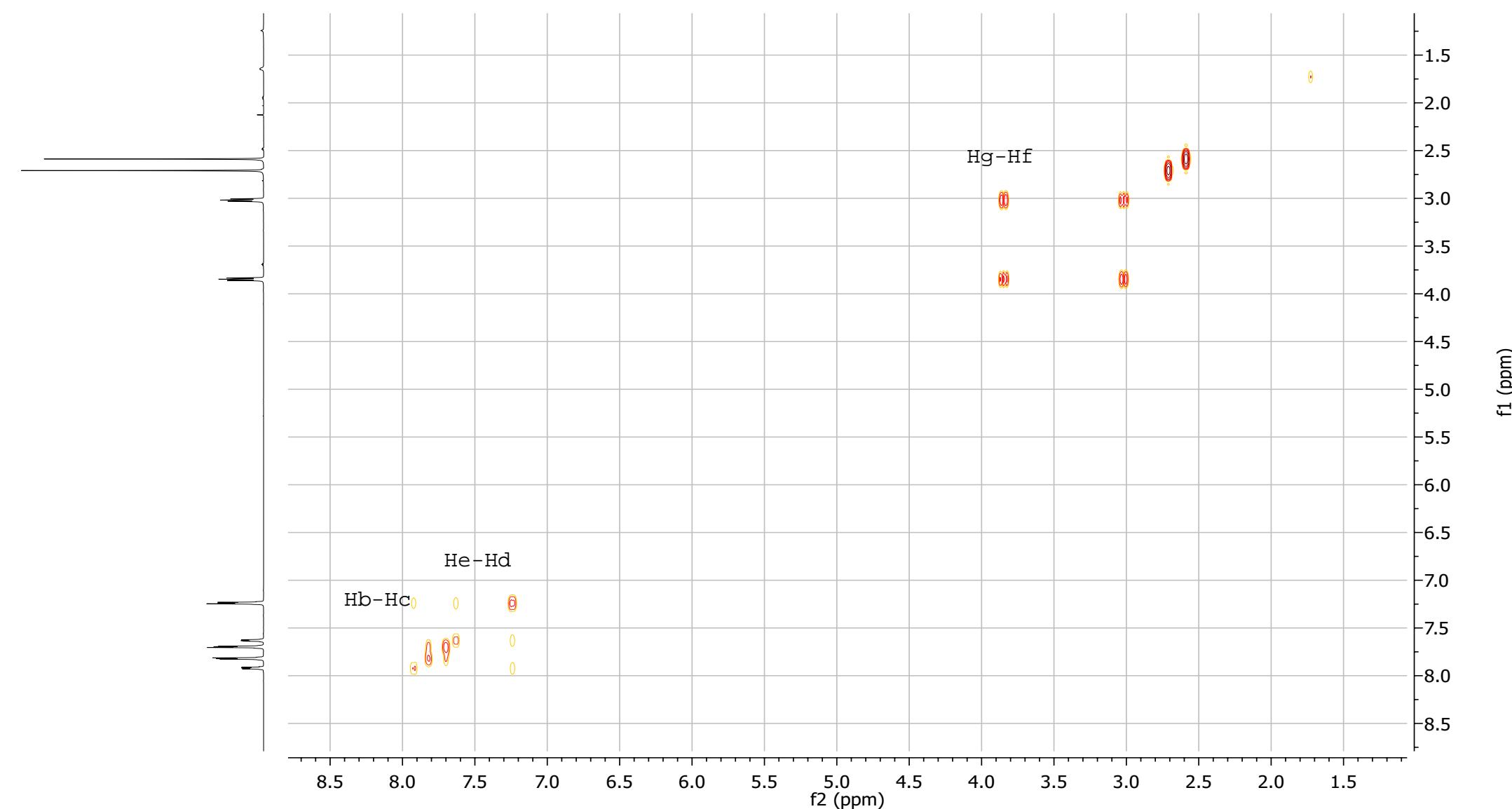
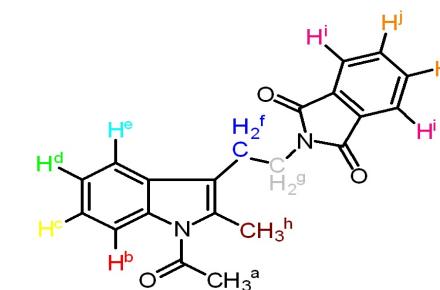
Parameter	Value
1 Data File Name	F:/ Yanzhao Wang Research/ Proj 2 Protected Indoles/ NMR Internal Indoles/ wyz5-106-pro-c13.fid/ fid
2 Title	wyz5-106-pro-c13
3 Solvent	cdcl3
4 Spectrometer Frequency	150.79
5 Nucleus	¹³ C



Parameter

Value (f2, f1)

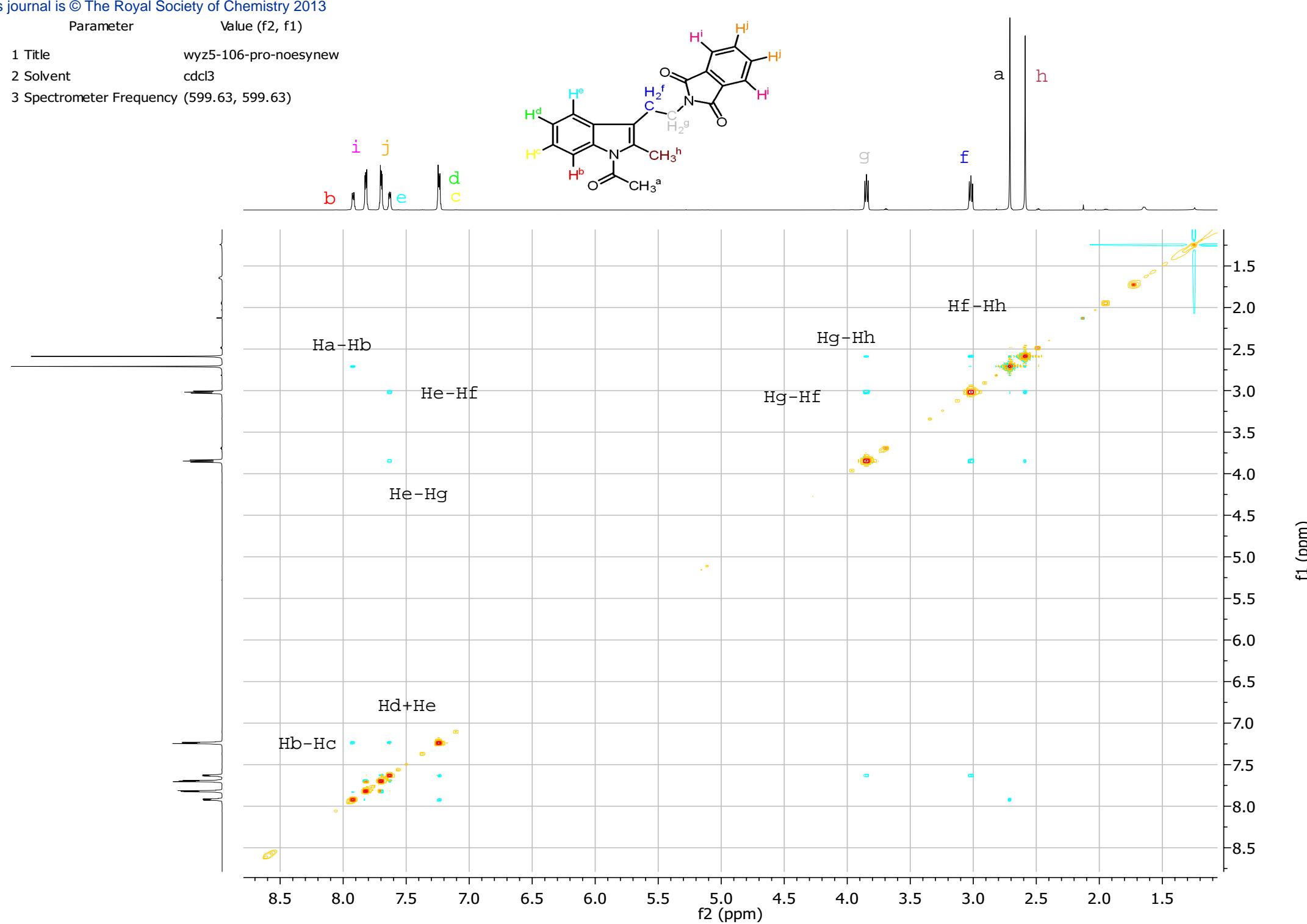
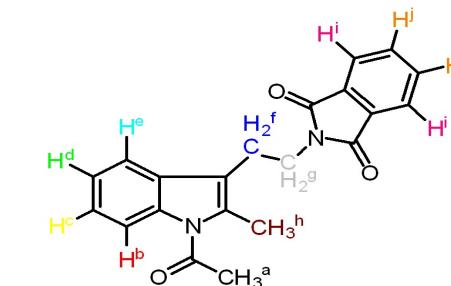
- 1 Title wyz5-106-pro-cosy
2 Solvent cdcl3
3 Spectrometer Frequency (599.63, 599.63)



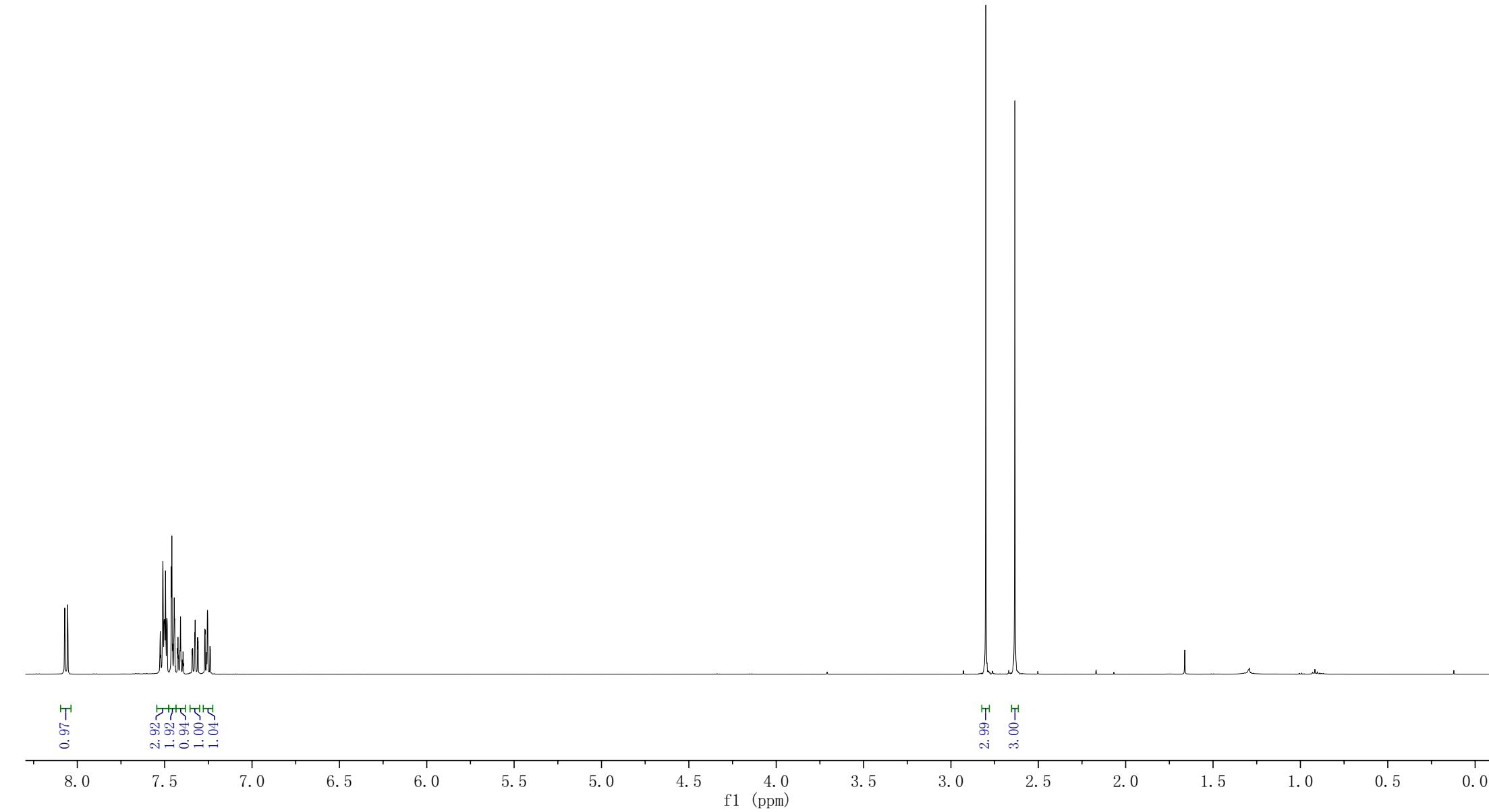
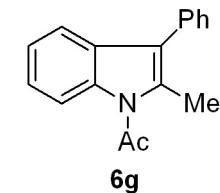
Parameter

Value (f2, f1)

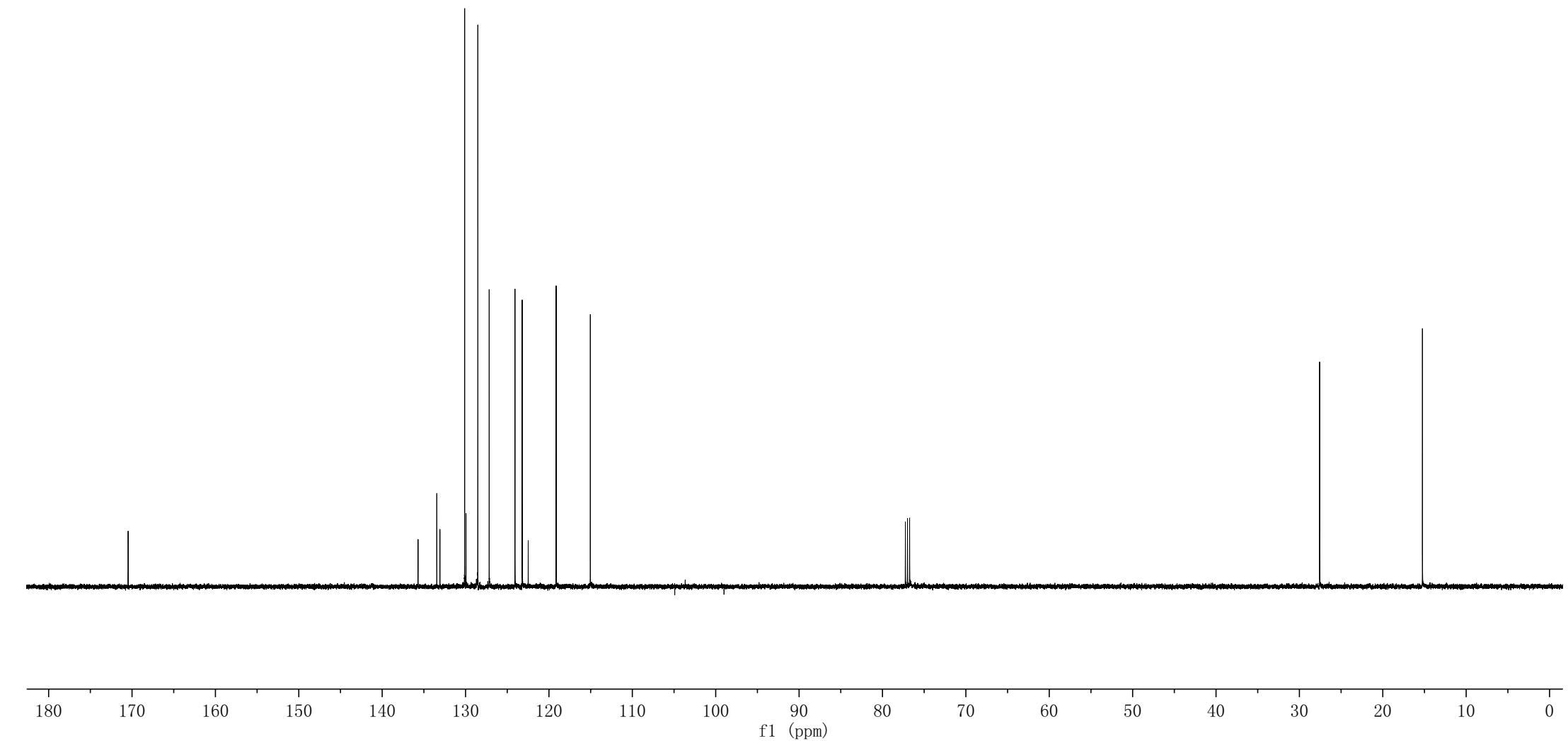
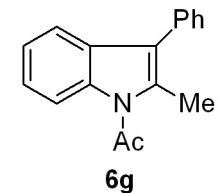
- 1 Title wyz5-106-pro-noesynew
2 Solvent cdcl3
3 Spectrometer Frequency (599.63, 599.63)



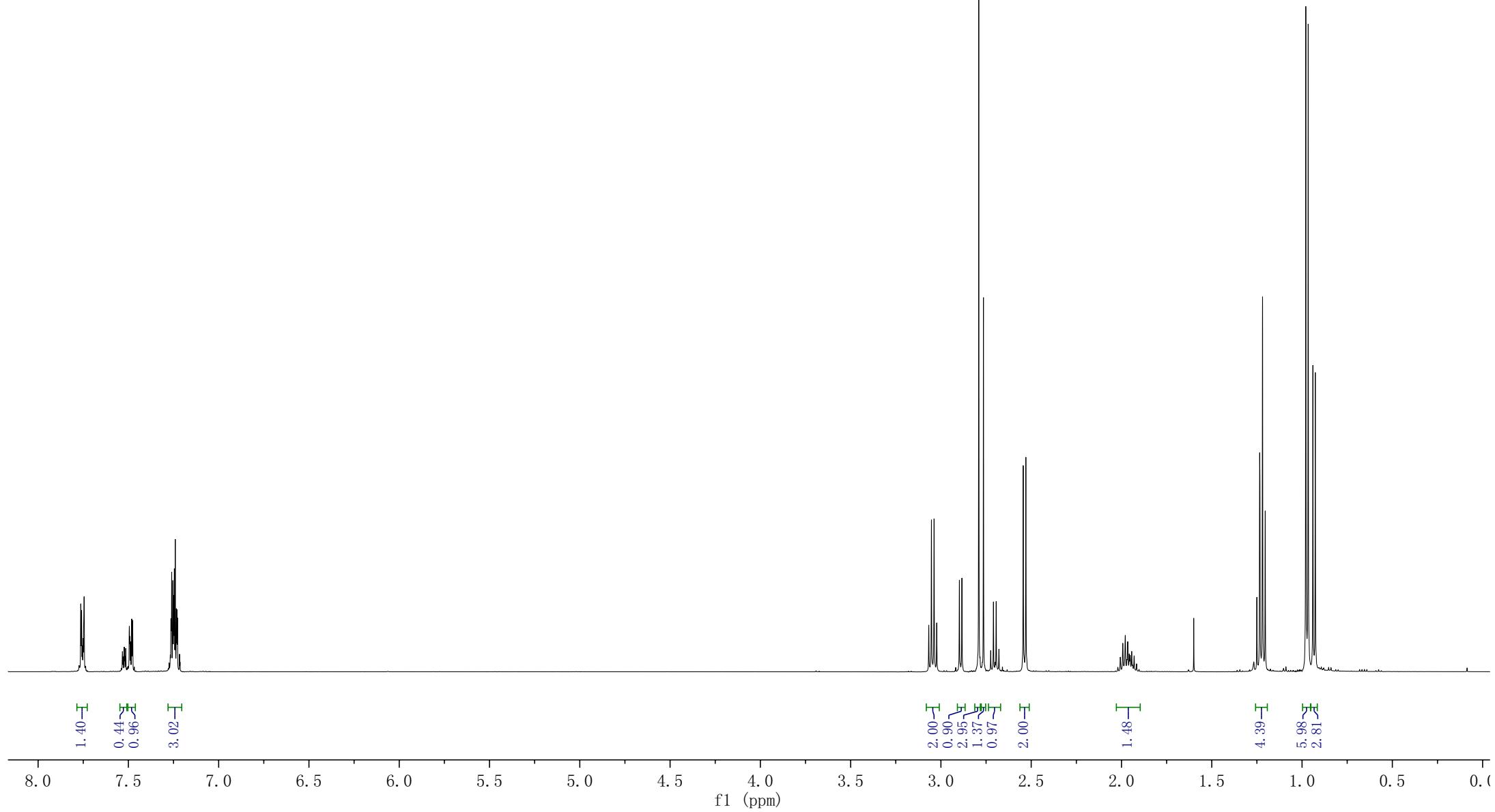
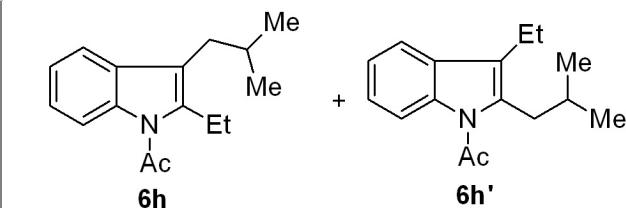
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2 Title	wyz5-101-pro1-h1
3 Solvent	CDCl ₃
4 Spectrometer Frequency	499.86
5 Nucleus	¹ H



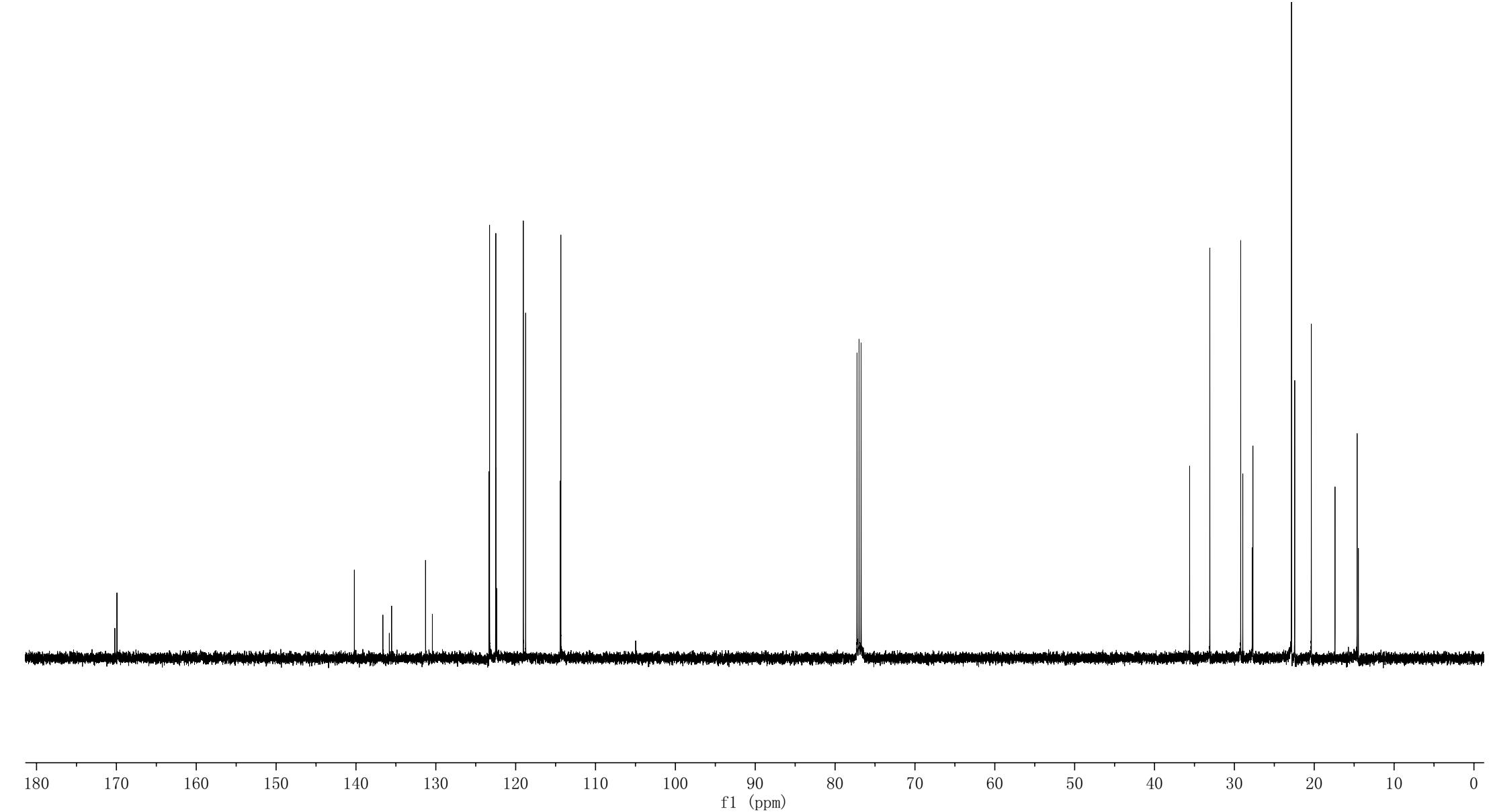
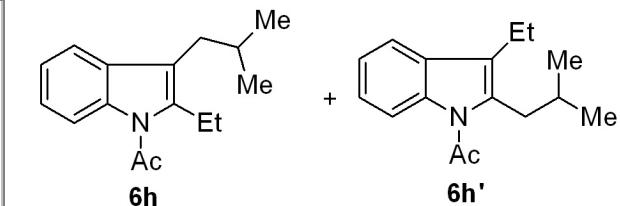
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1 Data File Name	F:/ Yanzhao Wang Research/ Proj 2 Protected Indoles/ NMR Internal Indoles/ wyz5-101-pro1-c13.fid/ fid
2 Title	wyz5-101-pro1-c13
3 Solvent	CDCl ₃
4 Spectrometer Frequency	125.70
5 Nucleus	¹³ C



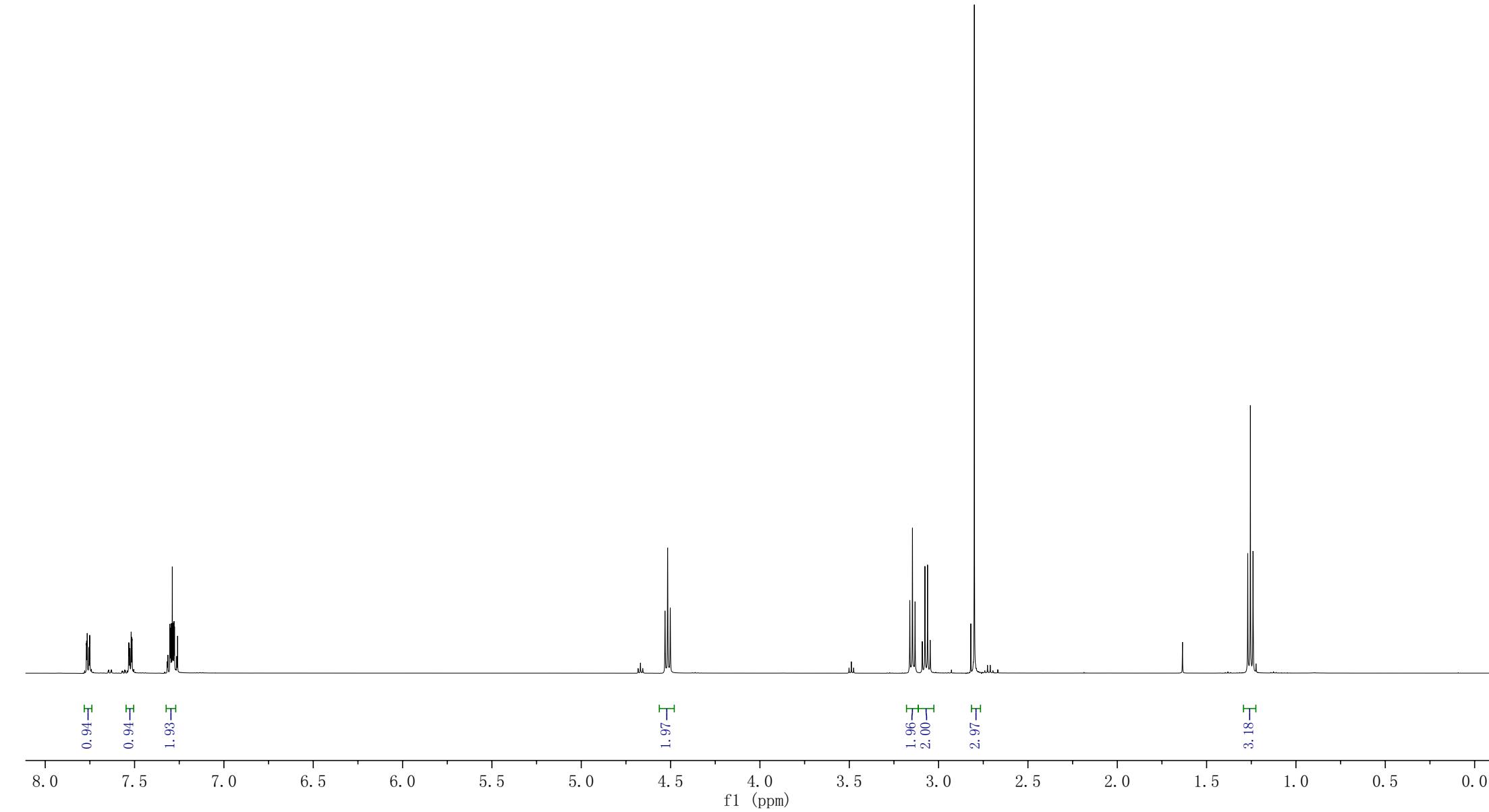
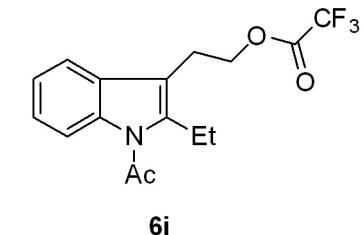
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3 Solvent	CDCl ₃
4 Spectrometer Frequency	499.86
5 Nucleus	¹ H



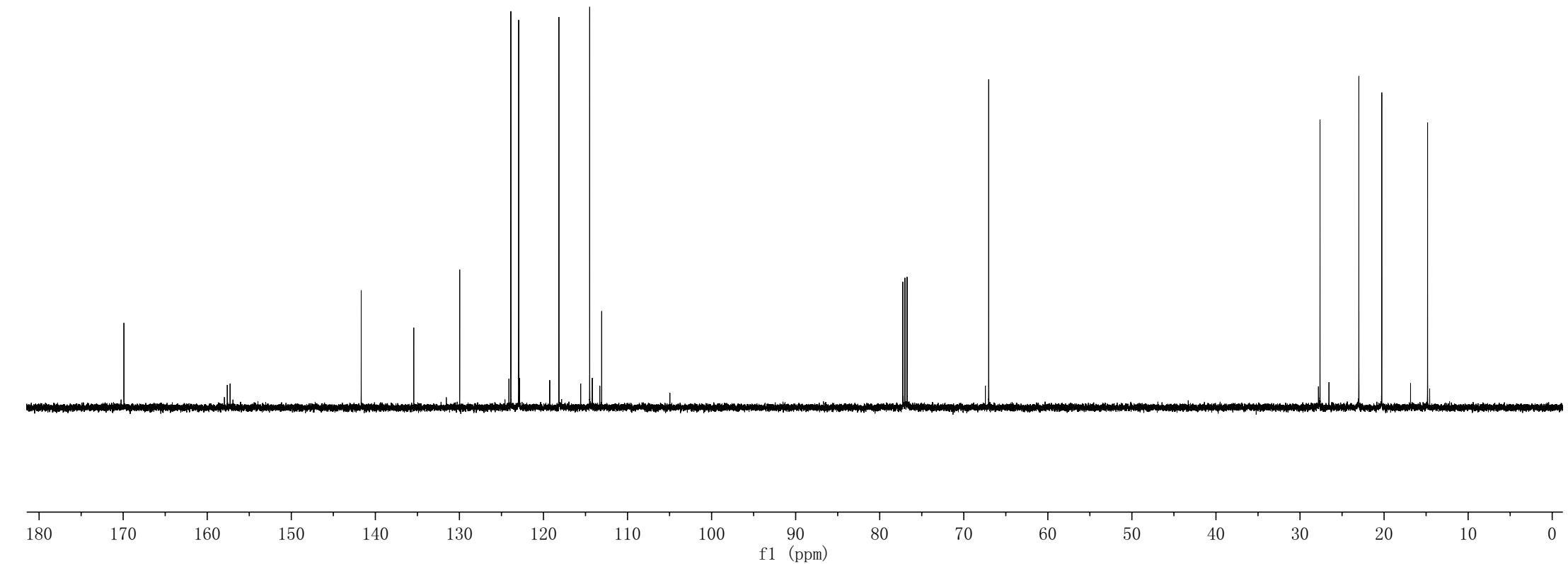
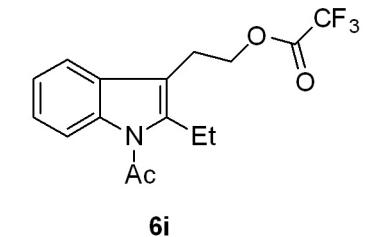
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1 Data File Name	F:/ Yanzhao Wang Research/ Proj 2 Protected Indoles/ NMR Internal Indoles/ wyz5-196-pro1-c13.fid/ fid
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3 Solvent	CDCl ₃
4 Spectrometer Frequency	125.70
5 Nucleus	¹³ C

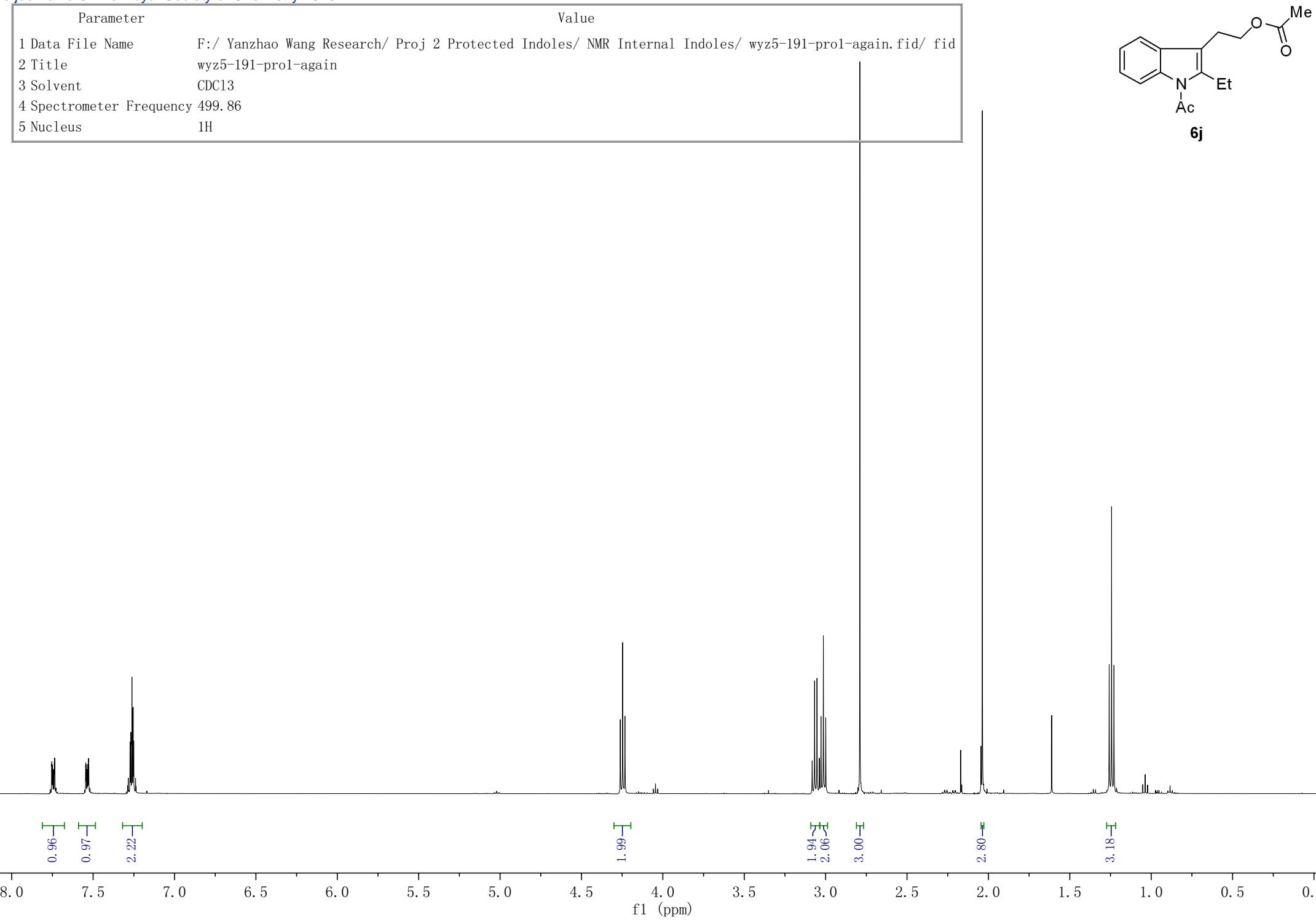


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2 Title	wyz5-123-pro-h1
3 Solvent	CDCl ₃
4 Spectrometer Frequency	499.86
5 Nucleus	¹ H

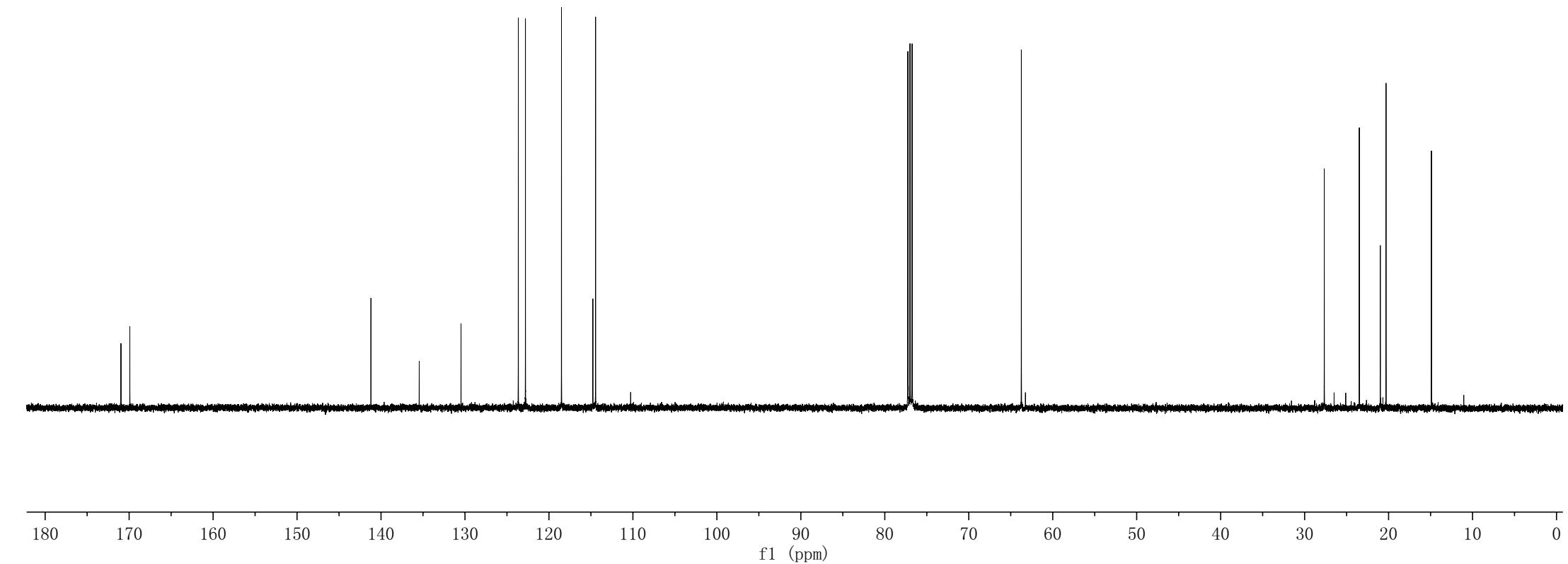
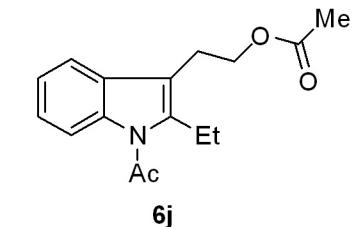


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2 Title	wyz5-123-pro-c13
3 Solvent	CDCl ₃
4 Spectrometer Frequency	125.70
5 Nucleus	¹³ C

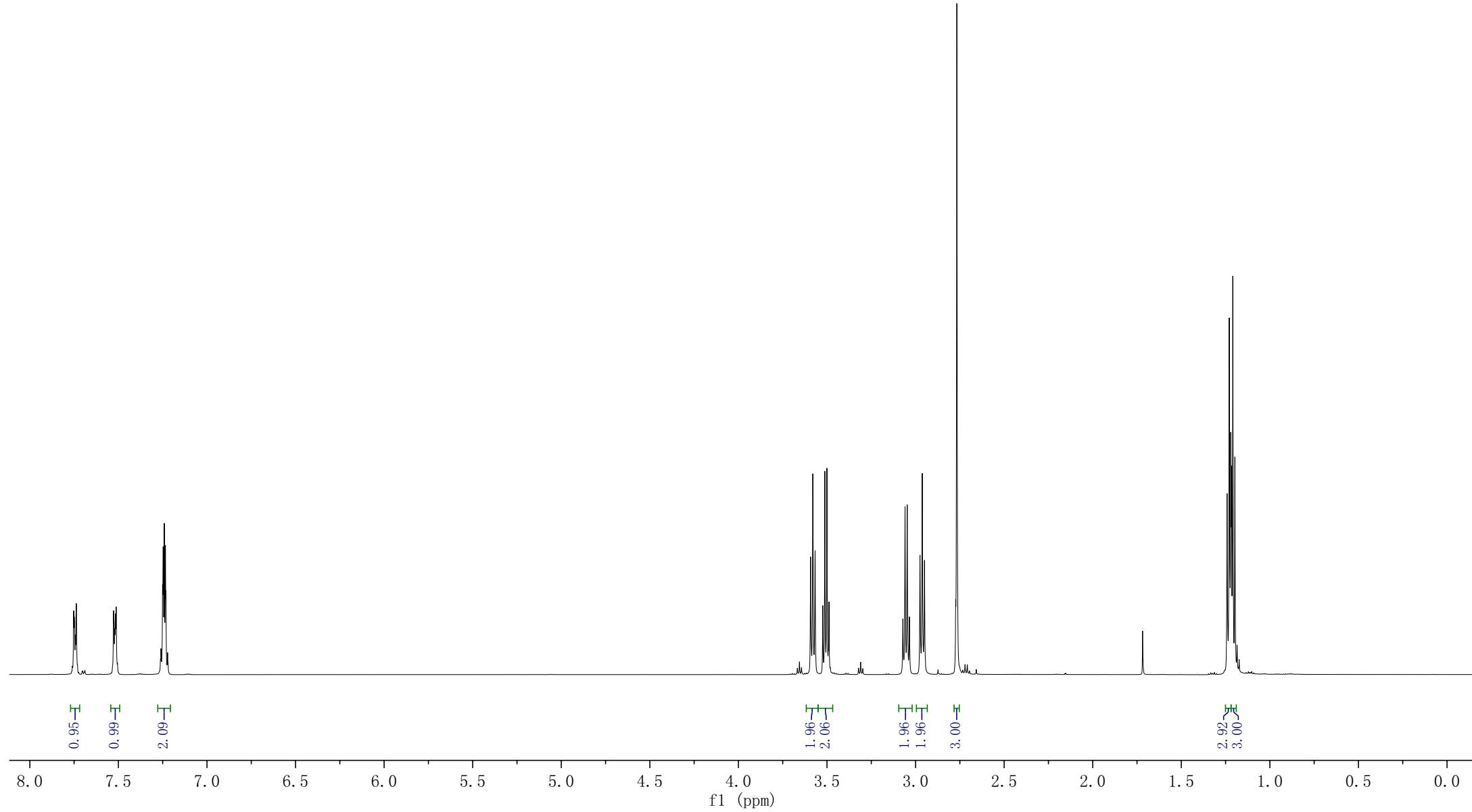
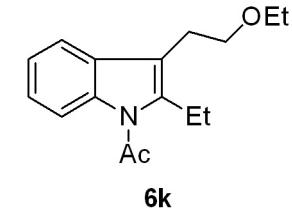




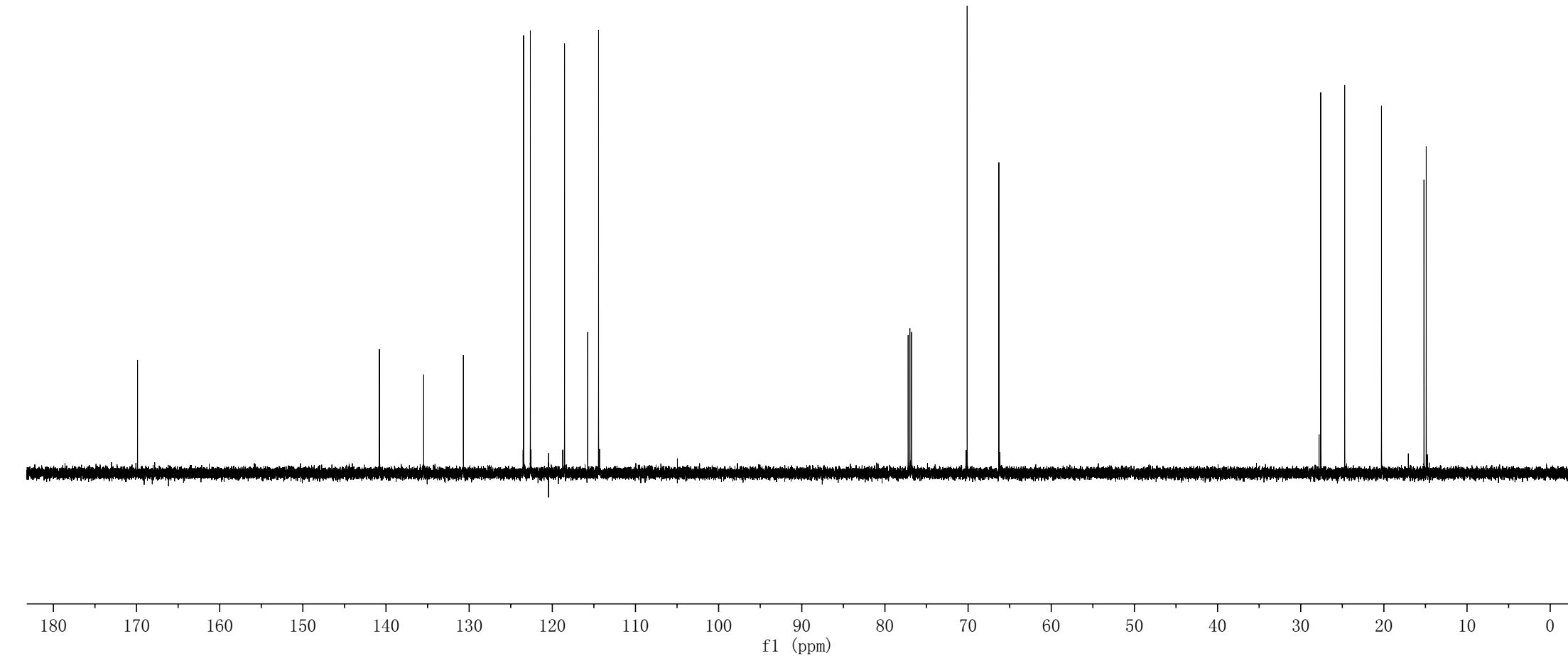
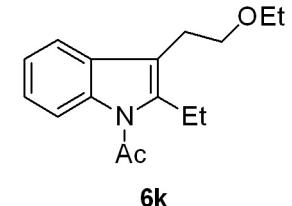
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3 Solvent	CDCl ₃
4 Spectrometer Frequency	125.70
5 Nucleus	¹³ C



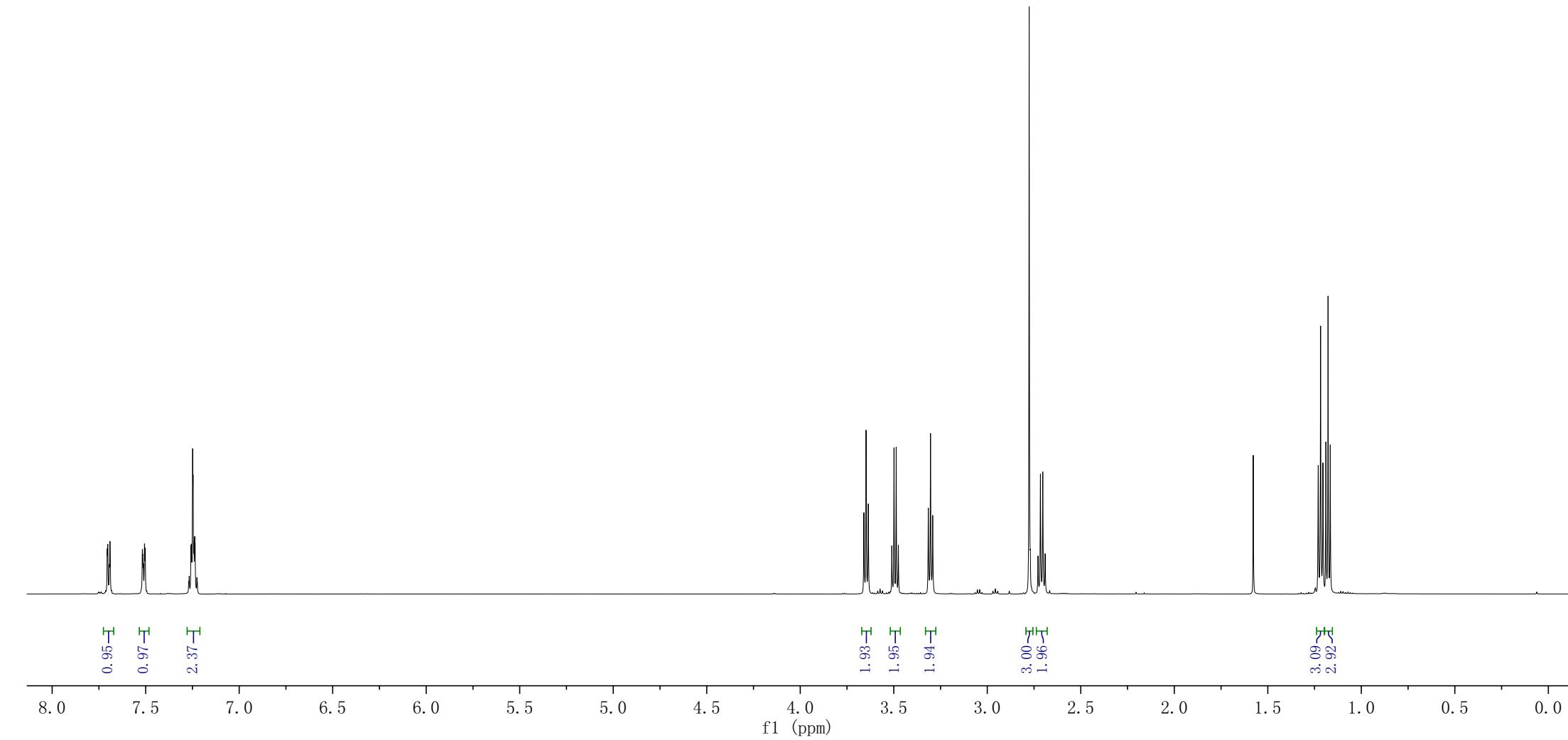
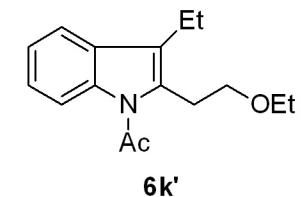
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4 Spectrometer Frequency	599. 63
5 Nucleus	1H



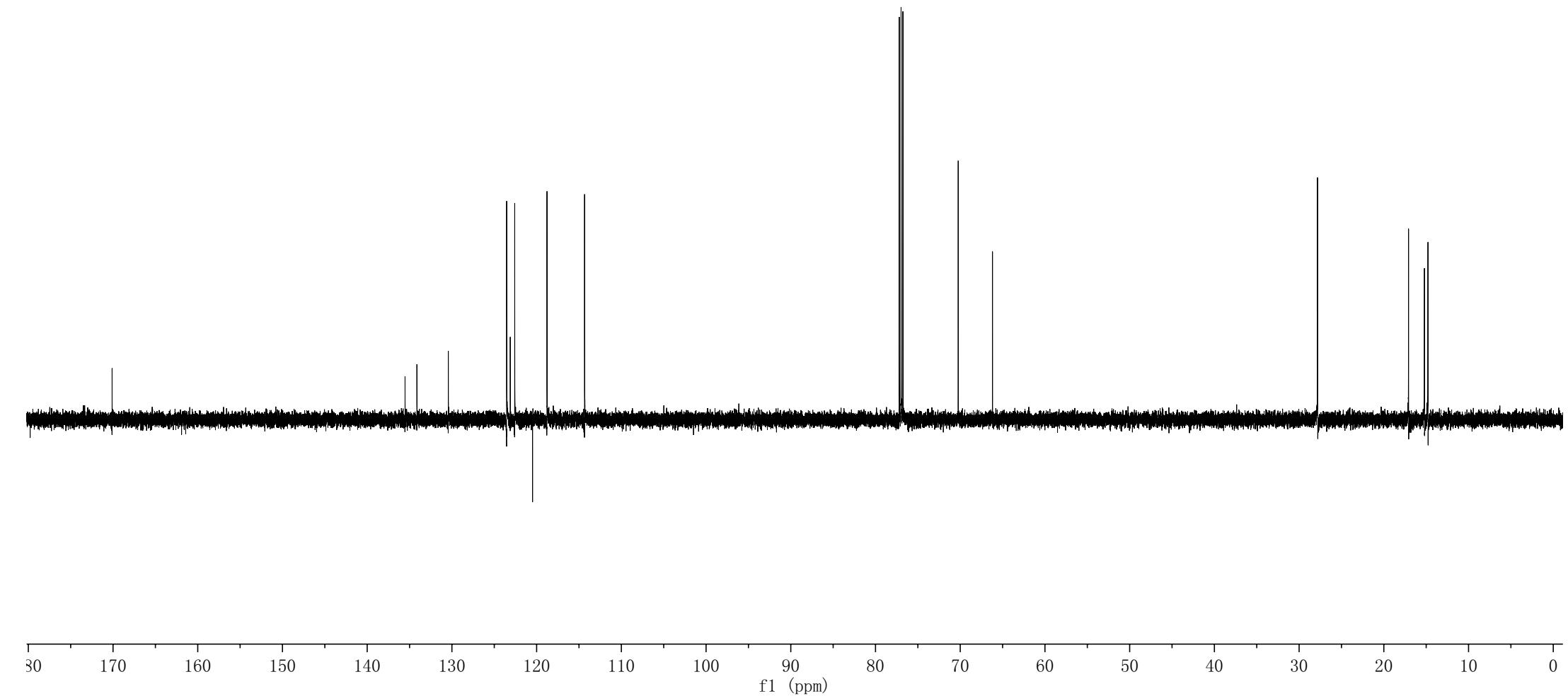
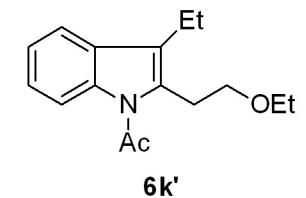
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3 Solvent	cdcl3
4 Spectrometer Frequency	150.79
5 Nucleus	¹³ C



Parameter	Value
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2 Title	wyz5-195-pro2-h1
3 Solvent	cdcl3
4 Spectrometer Frequency	599. 63
5 Nucleus	1H



Parameter	Value
1 Data File Name	F:/ Yanzhao Wang Research/ Proj 2 Protected Indoles/ NMR Internal Indoles/ wyz5-195-pro2-c13.fid/ fid
2 Title	wyz5-195-pro2-c13
3 Solvent	cdcl3
4 Spectrometer Frequency	150.79
5 Nucleus	¹³ C



Parameter

Value (f2, f1)

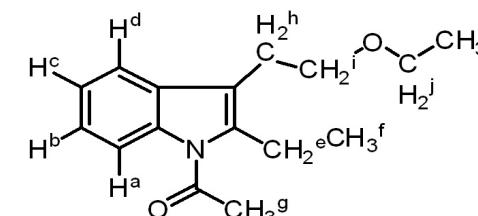
1 Title

wyz5-195-pro1-noesy

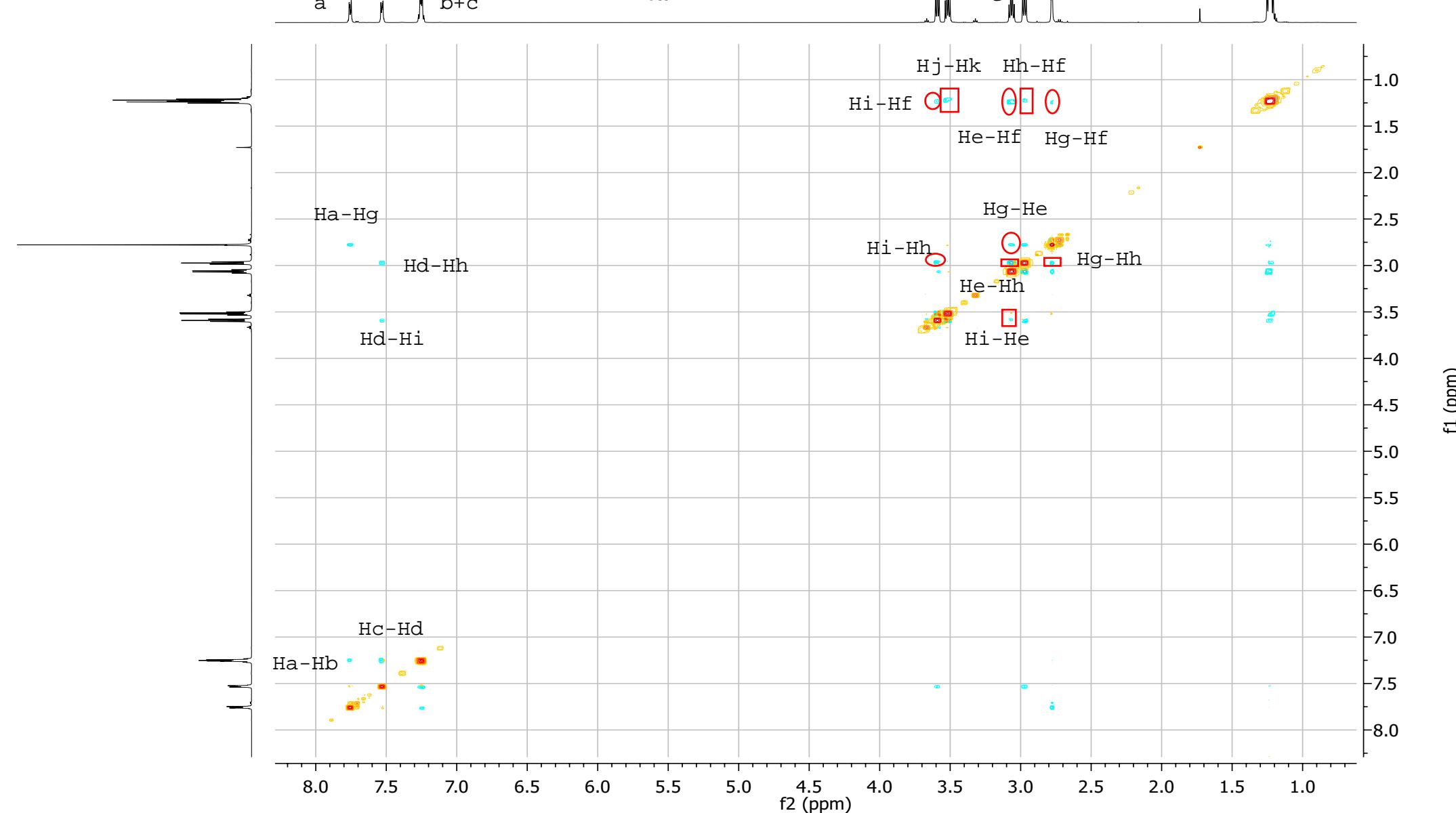
2 Solvent

cdcl3

3 Spectrometer Frequency (599.63, 599.63)



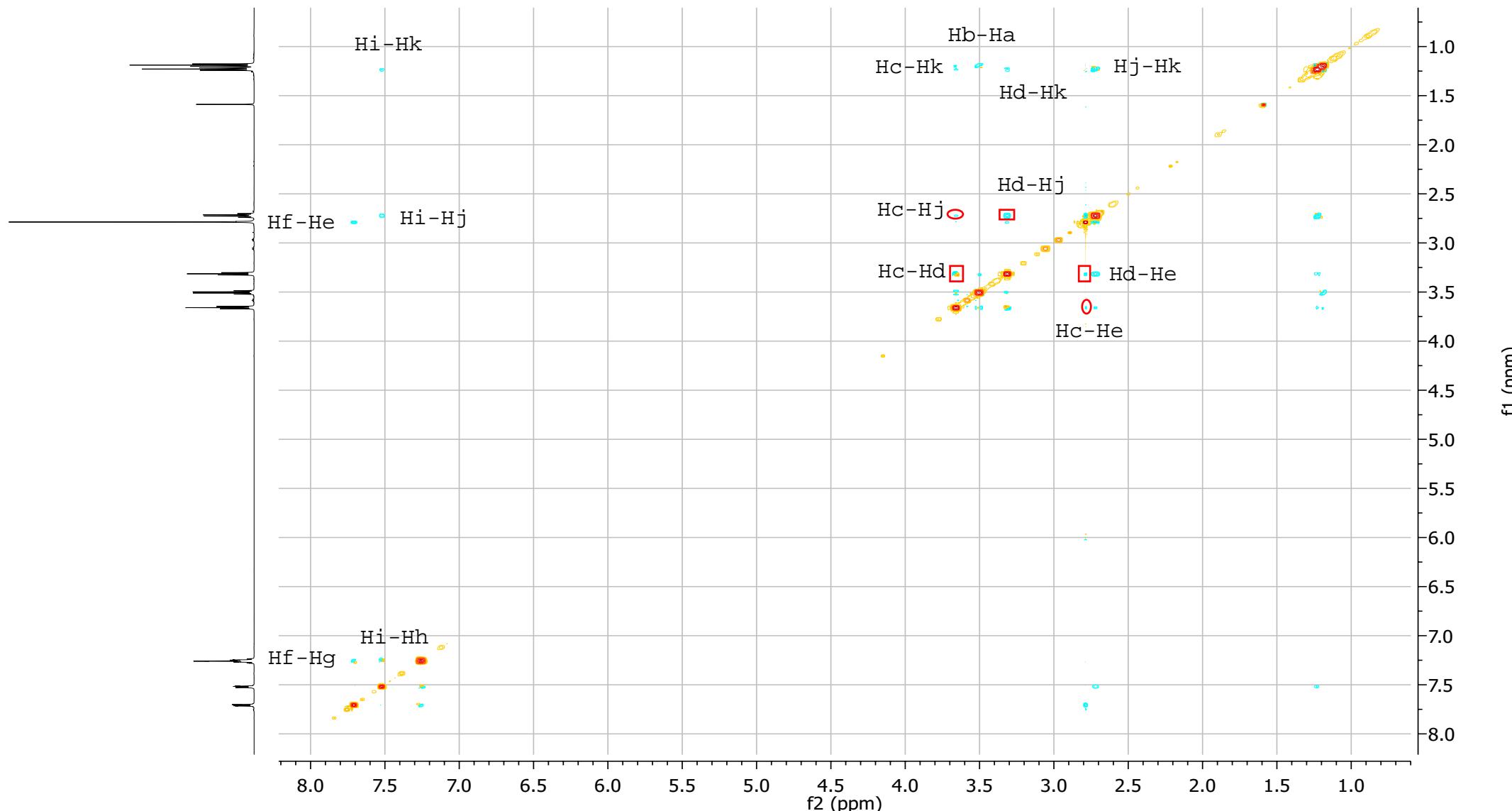
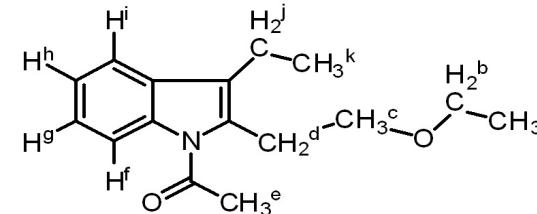
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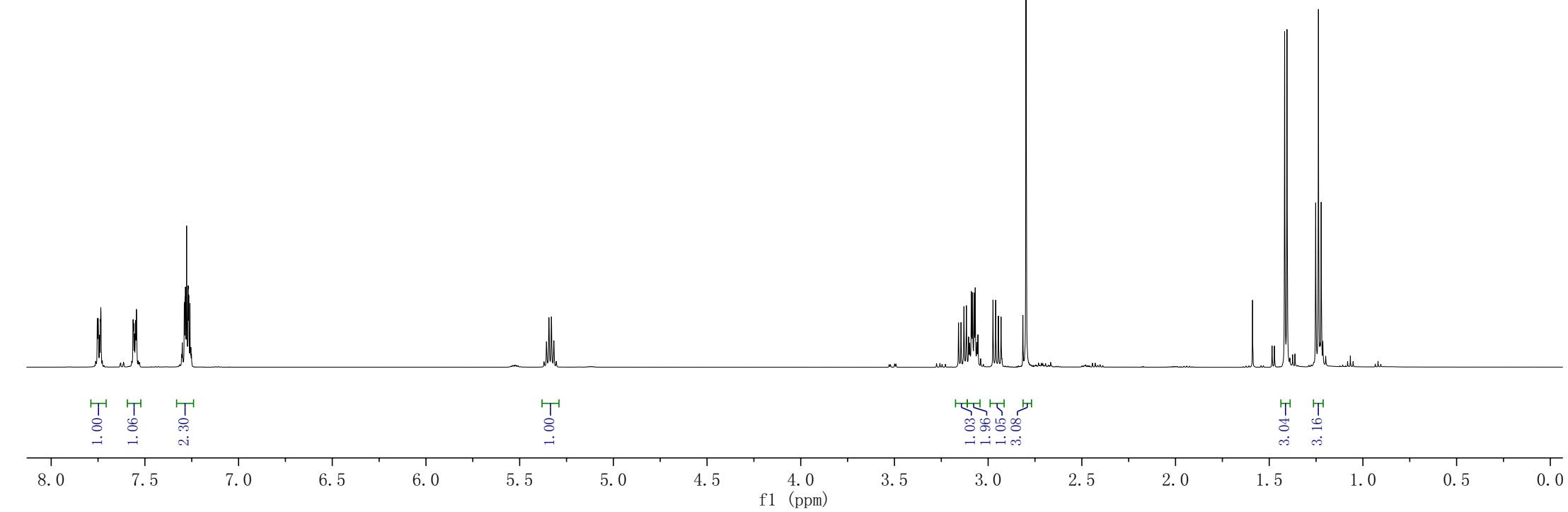
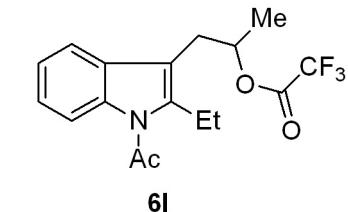
Parameter

Value (f2, f1)

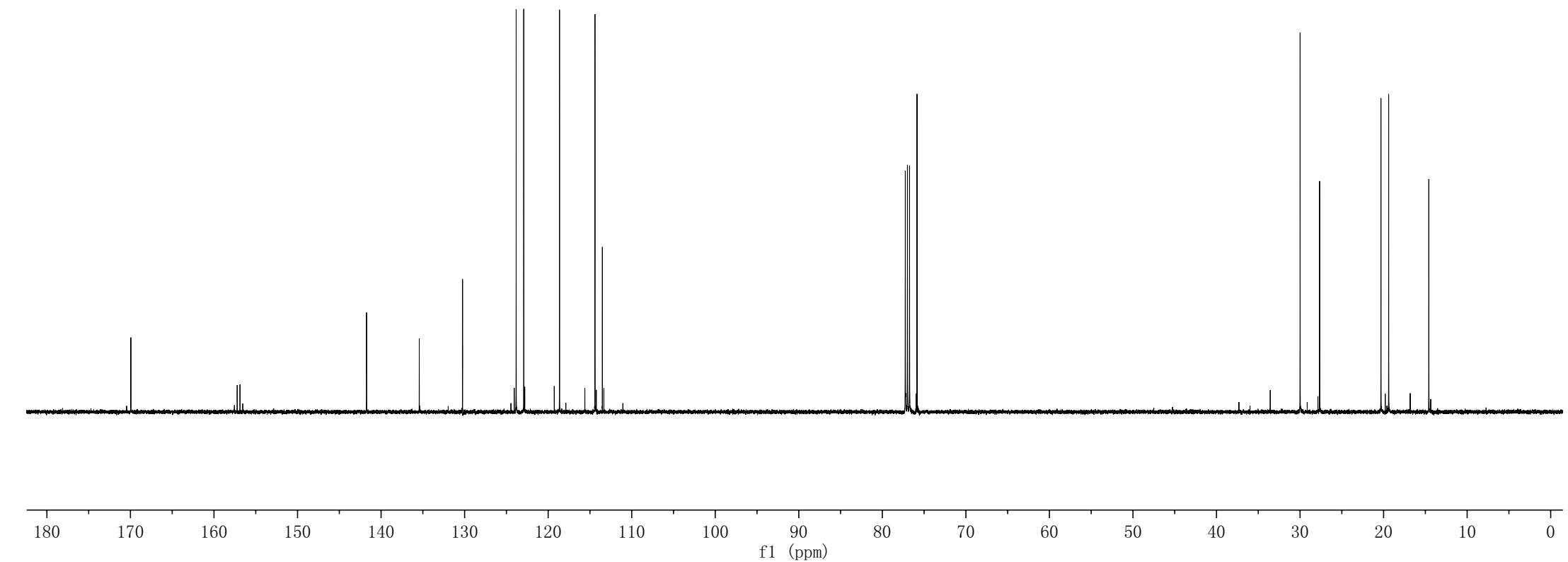
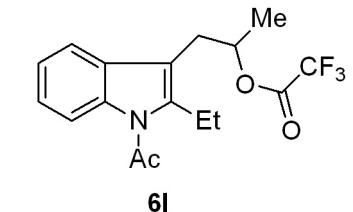
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2 Solvent cdcl3
3 Spectrometer Frequency (599.63, 599.63)



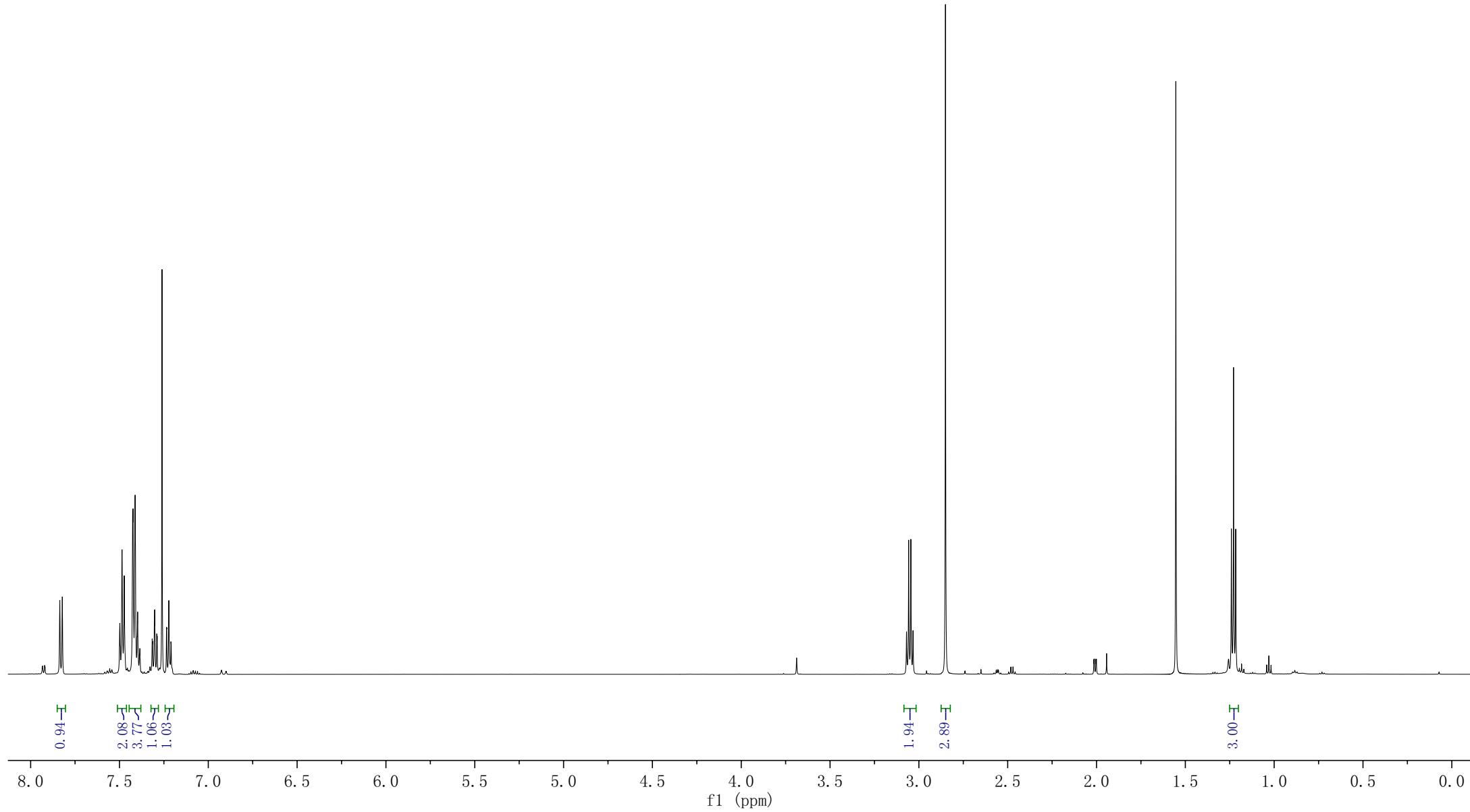
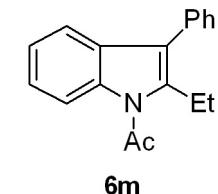
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3 Solvent	CDCl ₃
4 Spectrometer Frequency	499.86
5 Nucleus	¹ H



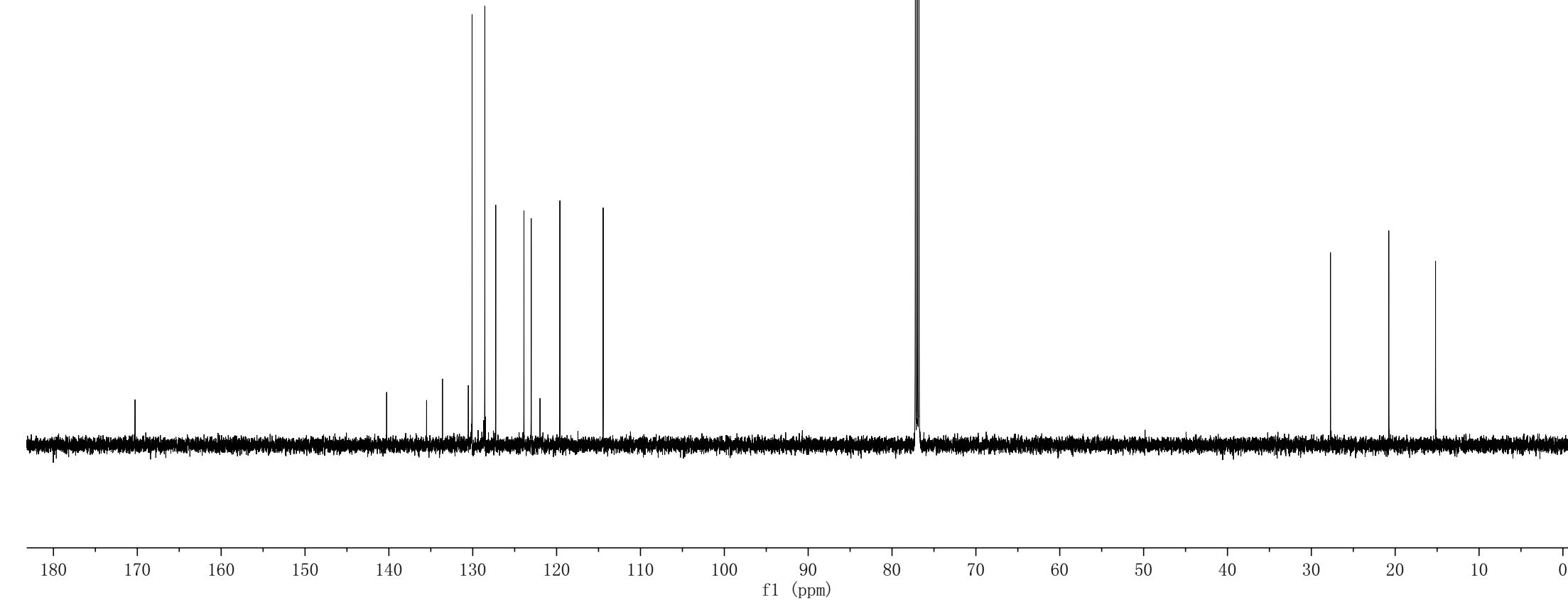
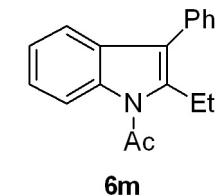
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3 Solvent	CDCl ₃
4 Spectrometer Frequency	125.70
5 Nucleus	¹³ C



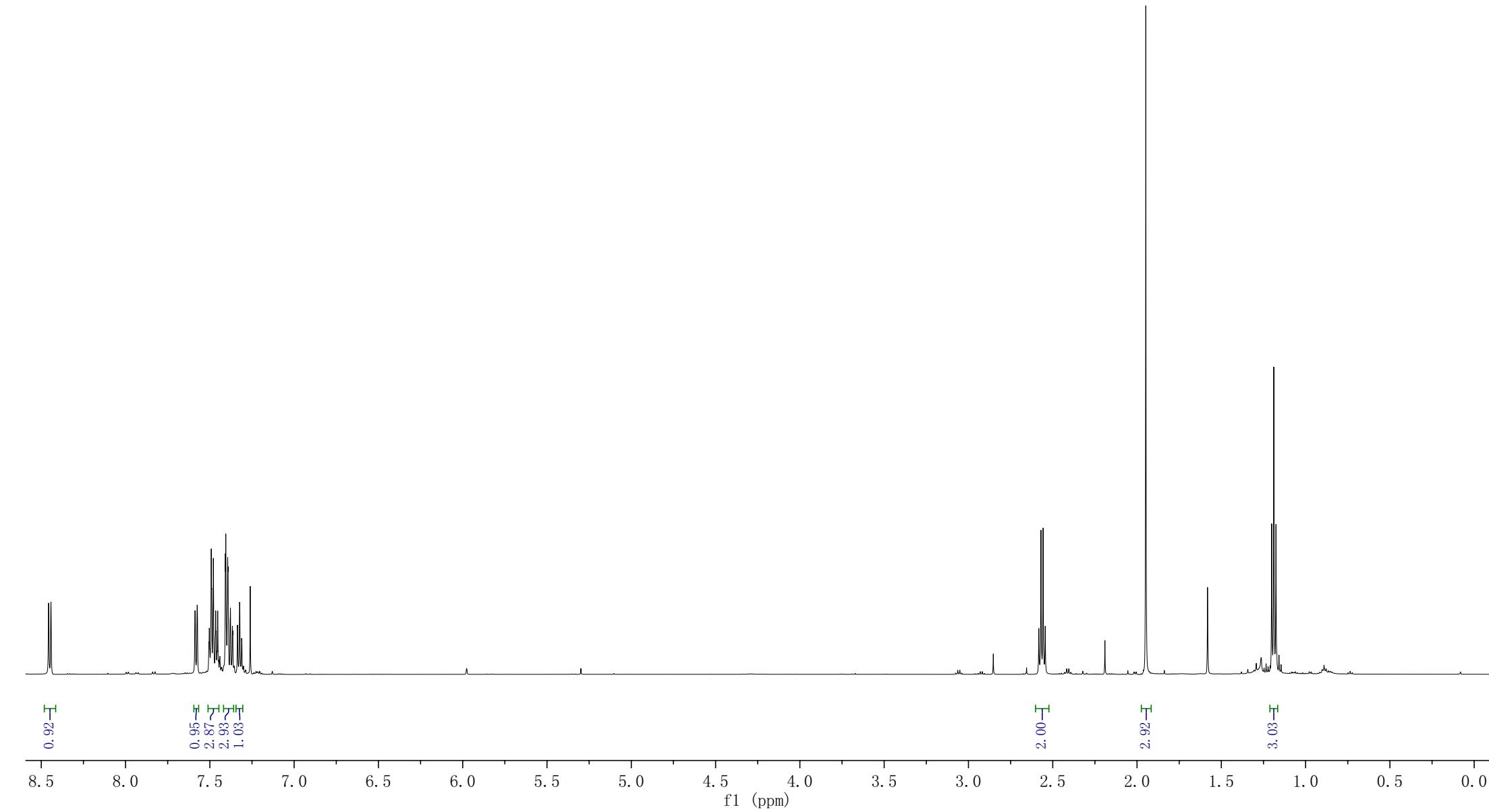
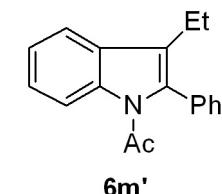
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4 Spectrometer Frequency	599. 63
5 Nucleus	1H



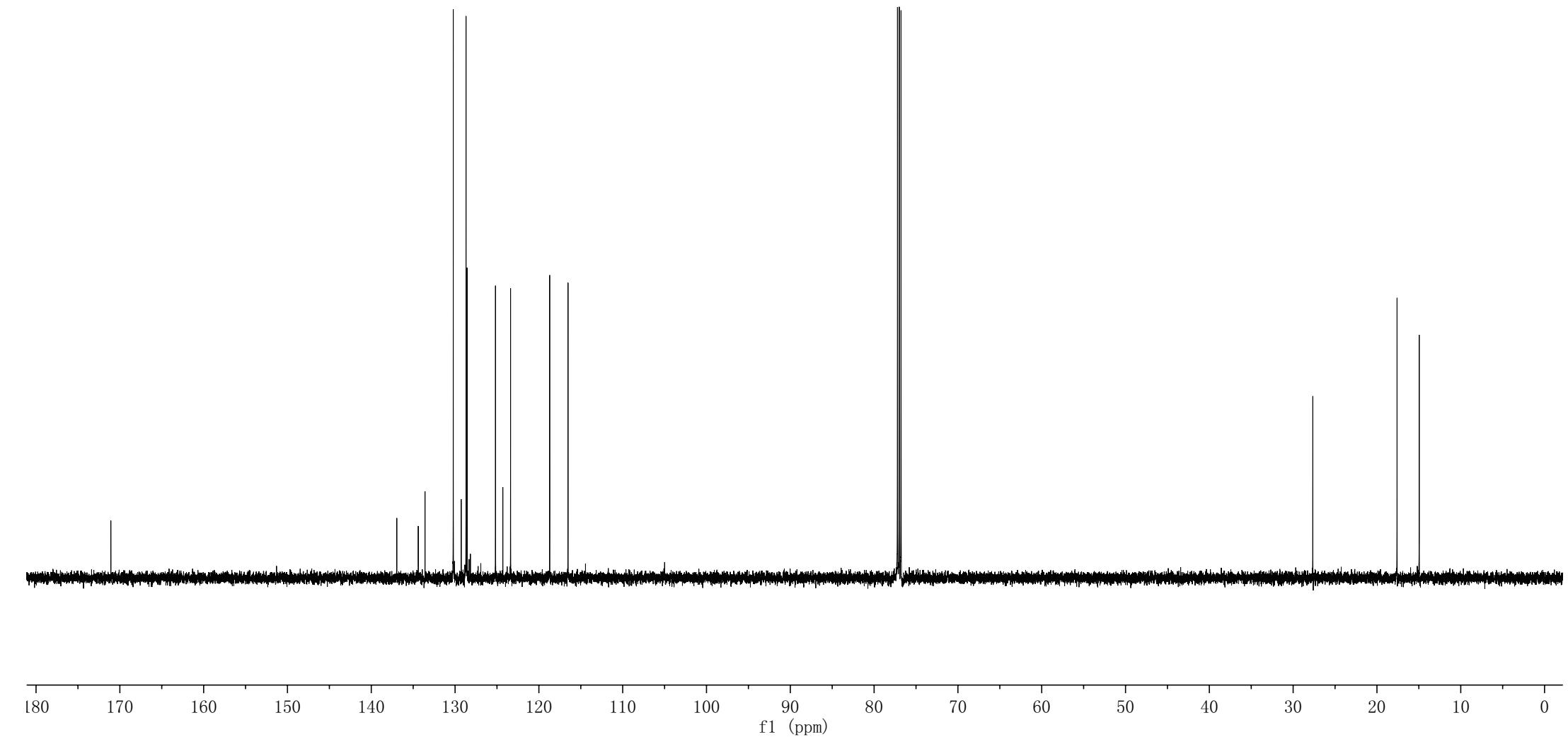
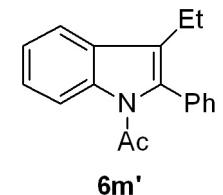
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3 Solvent	cdcl3
4 Spectrometer Frequency	150.79
5 Nucleus	13C



Parameter	Value
1 Data File Name	F:/ Yanzhao Wang Research/ Proj 2 Protected Indoles/ NMR Internal Indoles/ wyz5-193-pro1-h1.fid/ fid
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4 Spectrometer Frequency	599. 63
5 Nucleus	1H

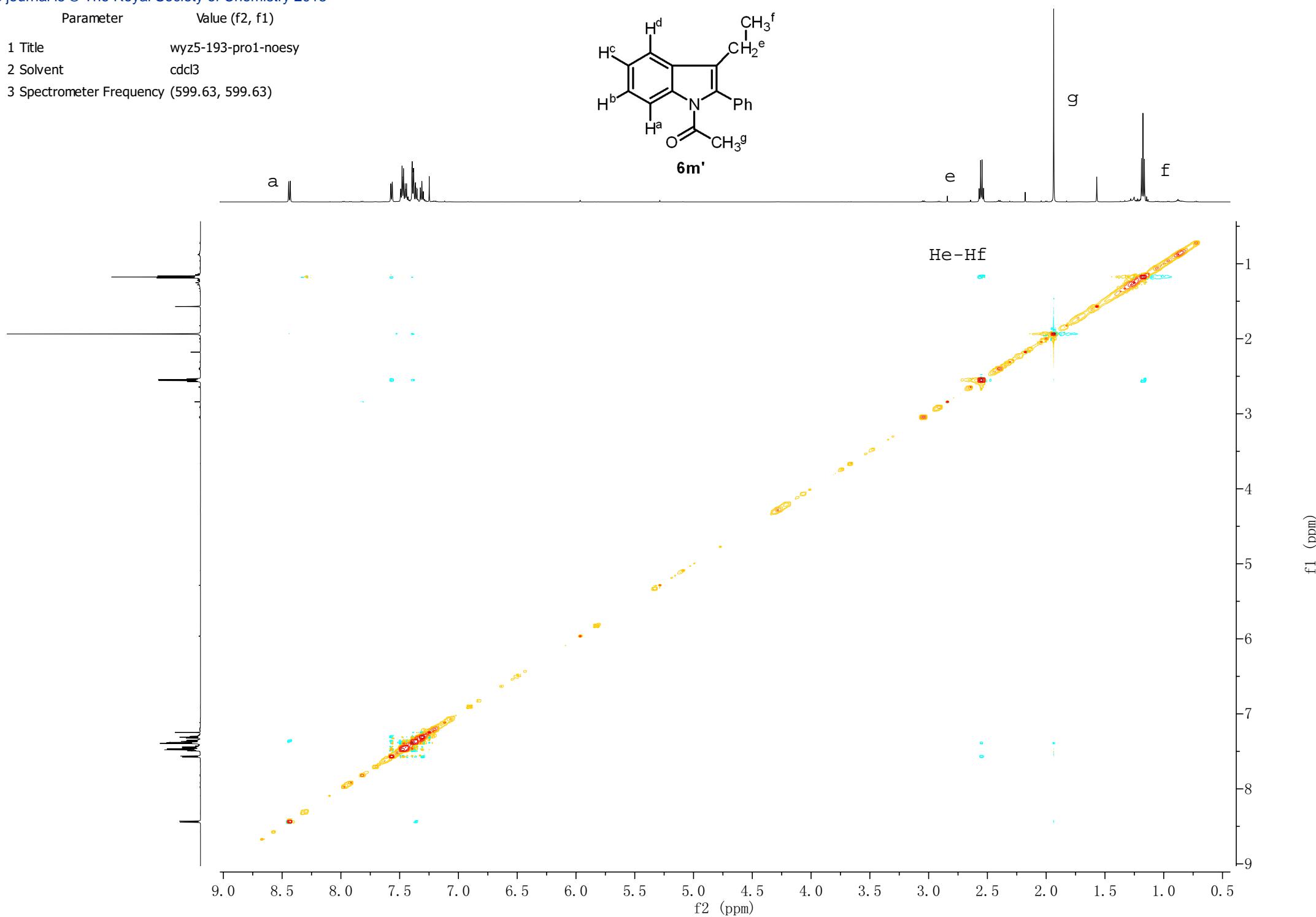
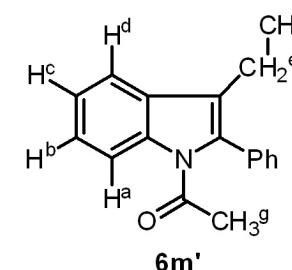


Parameter	Value
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2 Title	wyz5-193-pro1-c13
3 Solvent	cdcl3
4 Spectrometer Frequency	150.79
5 Nucleus	¹³ C



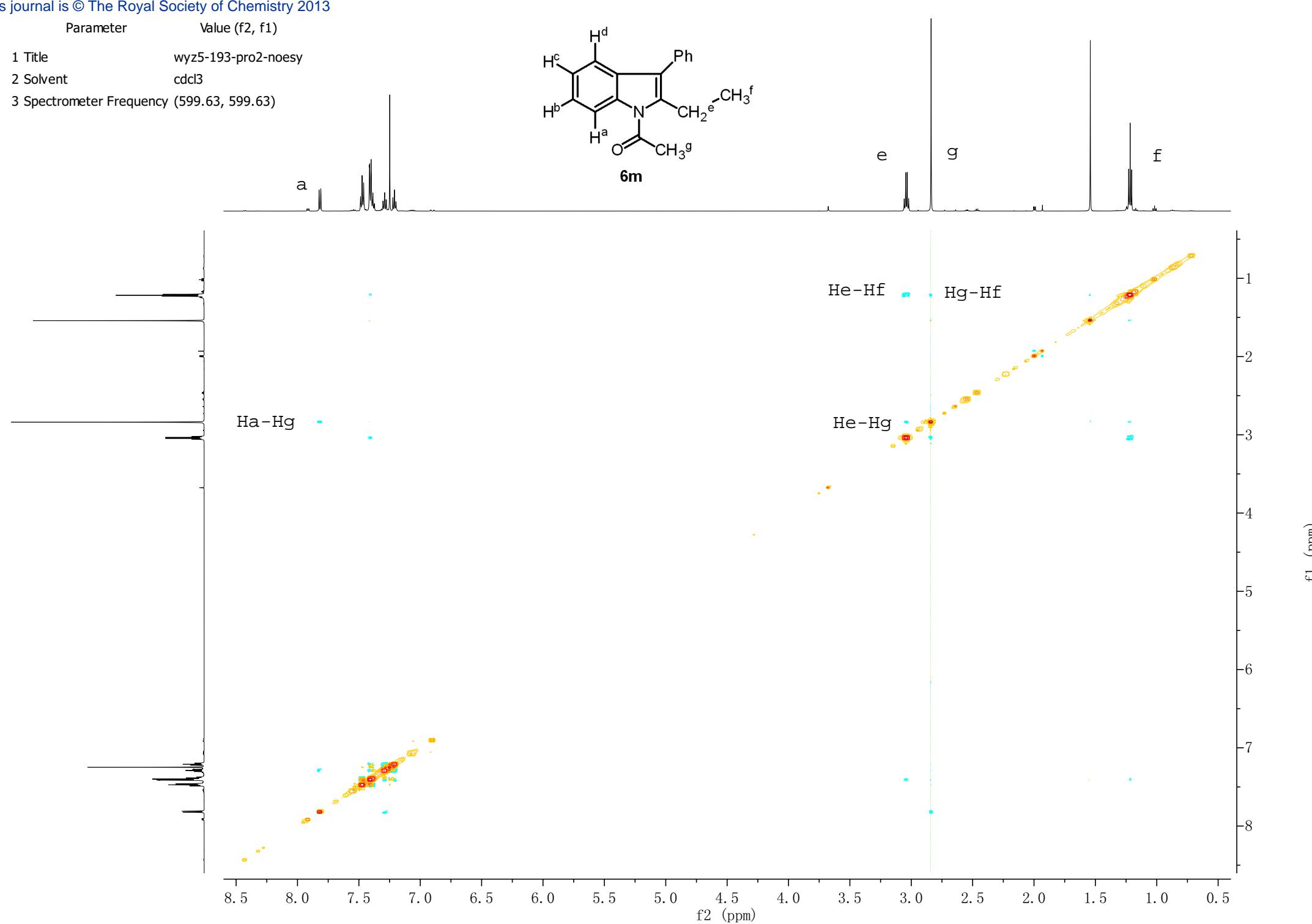
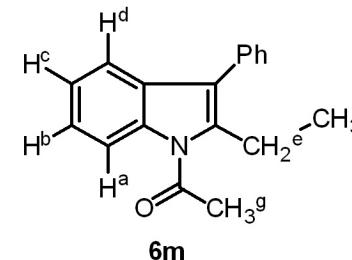
Parameter Value (f2, f1)

- 1 Title wyz5-193-pro1-noesy
2 Solvent cdcl3
3 Spectrometer Frequency (599.63, 599.63)

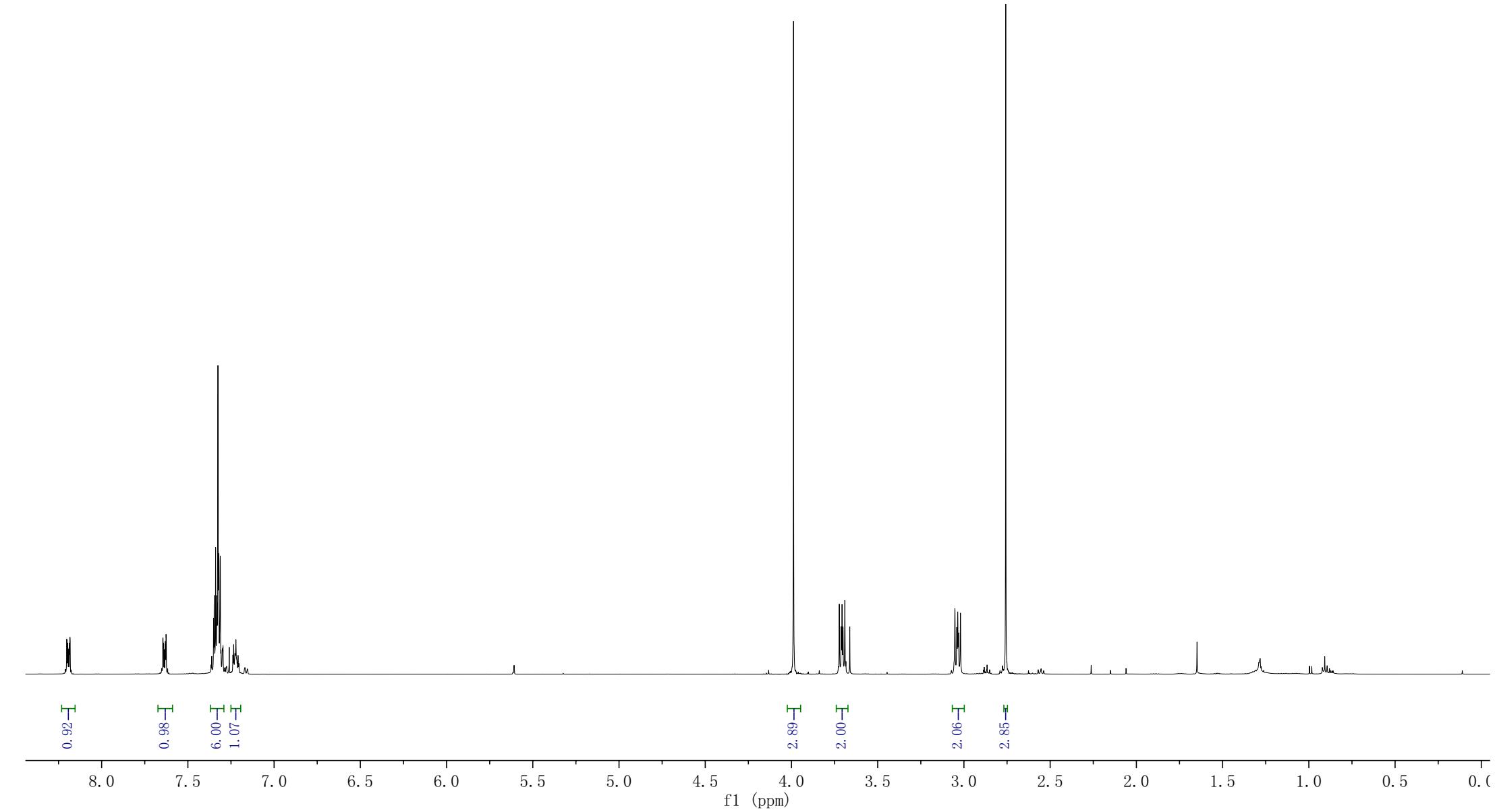
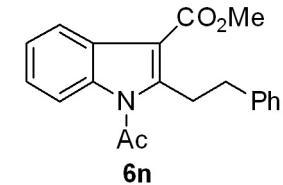


Parameter Value (f₂, f₁)

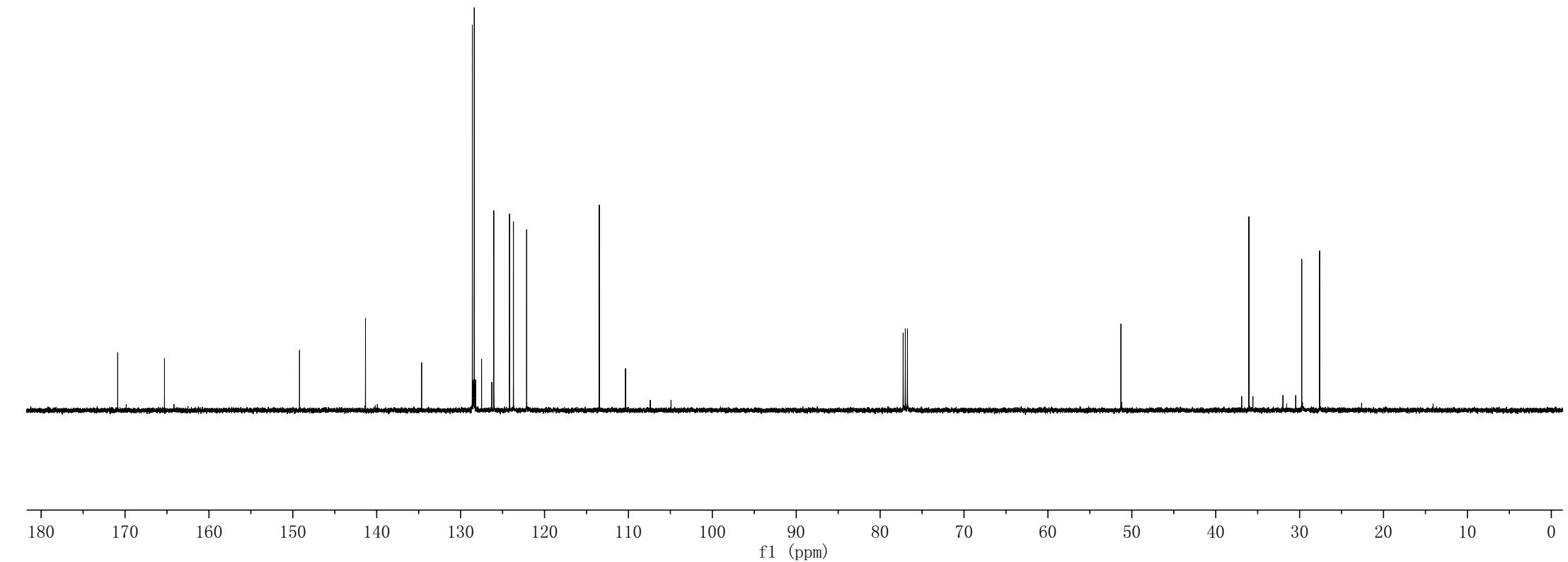
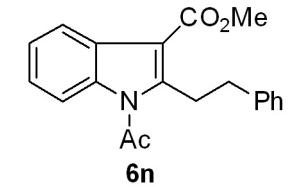
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2 Solvent cdcl₃
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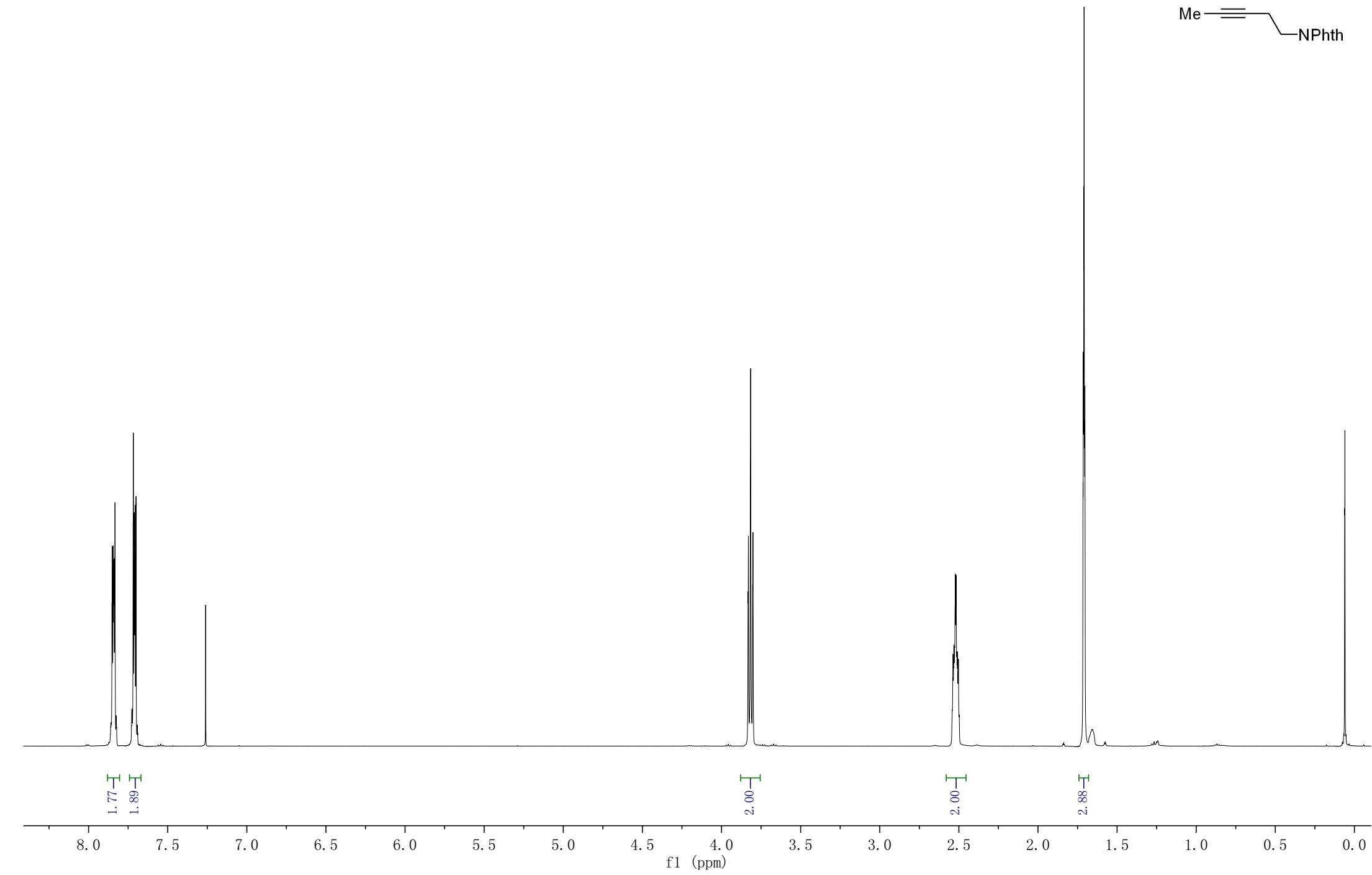
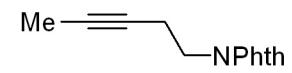


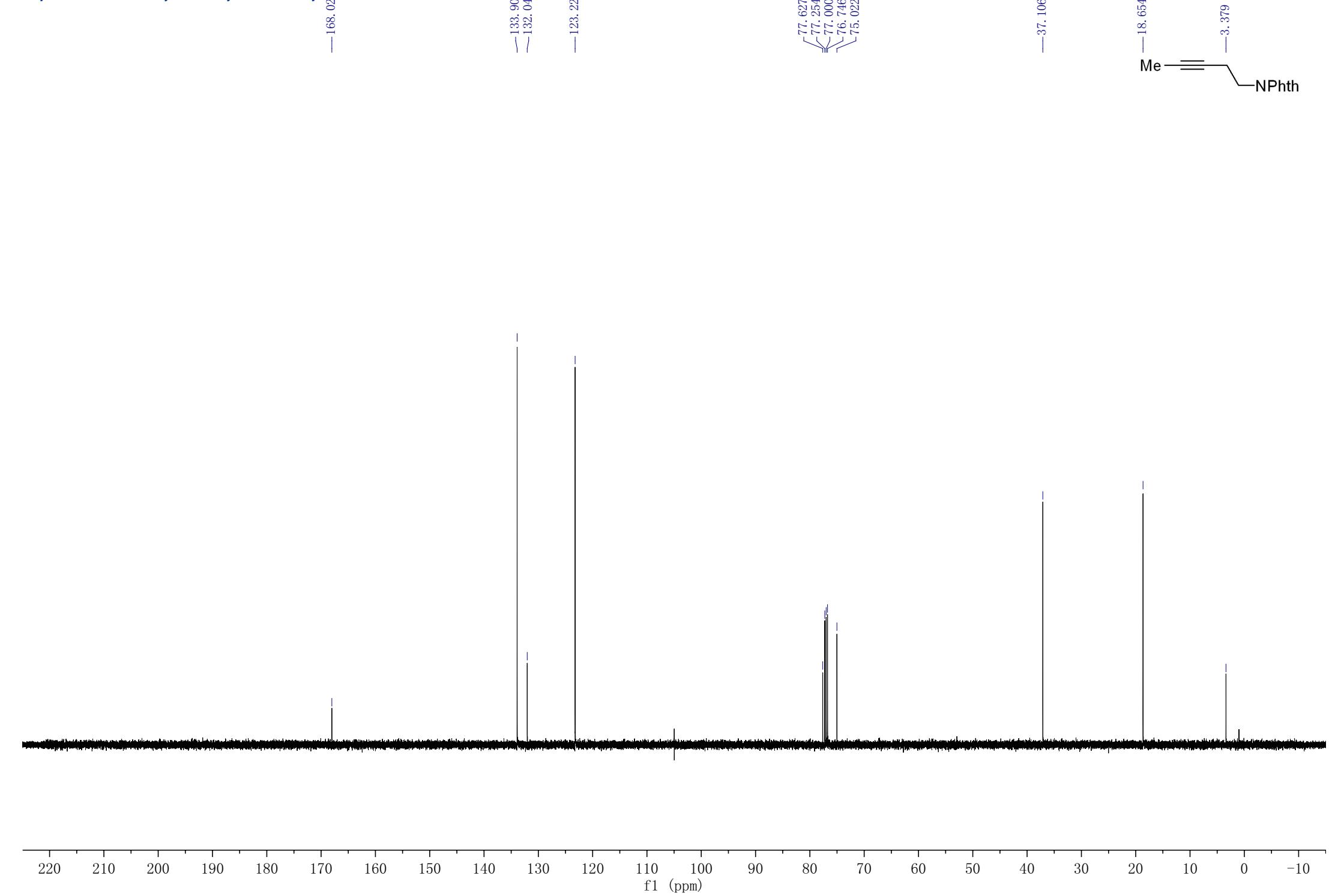
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3 Solvent	CDCl ₃
4 Spectrometer Frequency	499.86
5 Nucleus	¹ H

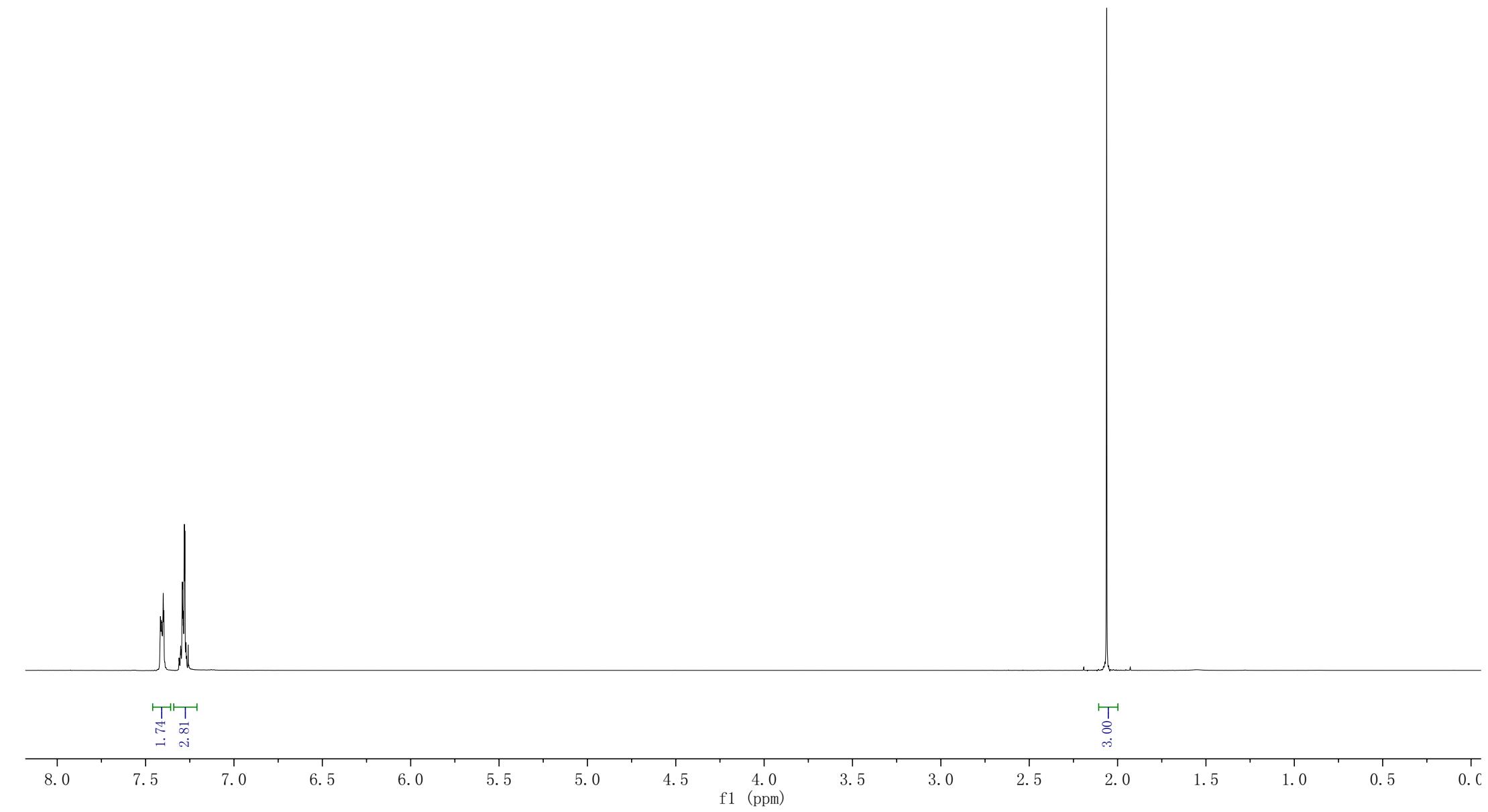
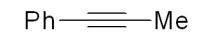


Parameter	Value
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3 Solvent	CDCl ₃
4 Spectrometer Frequency	125.70
5 Nucleus	¹³ C





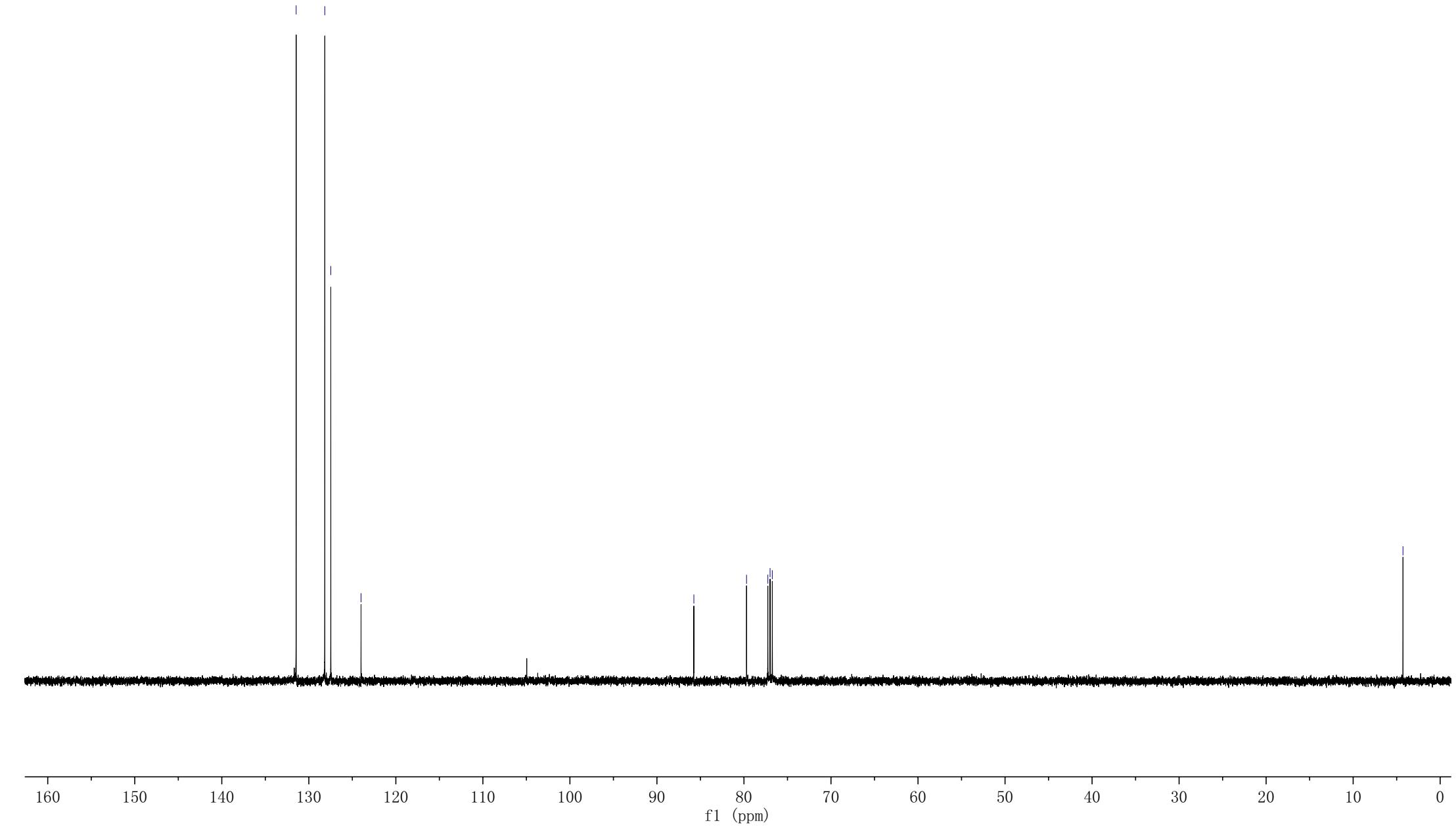




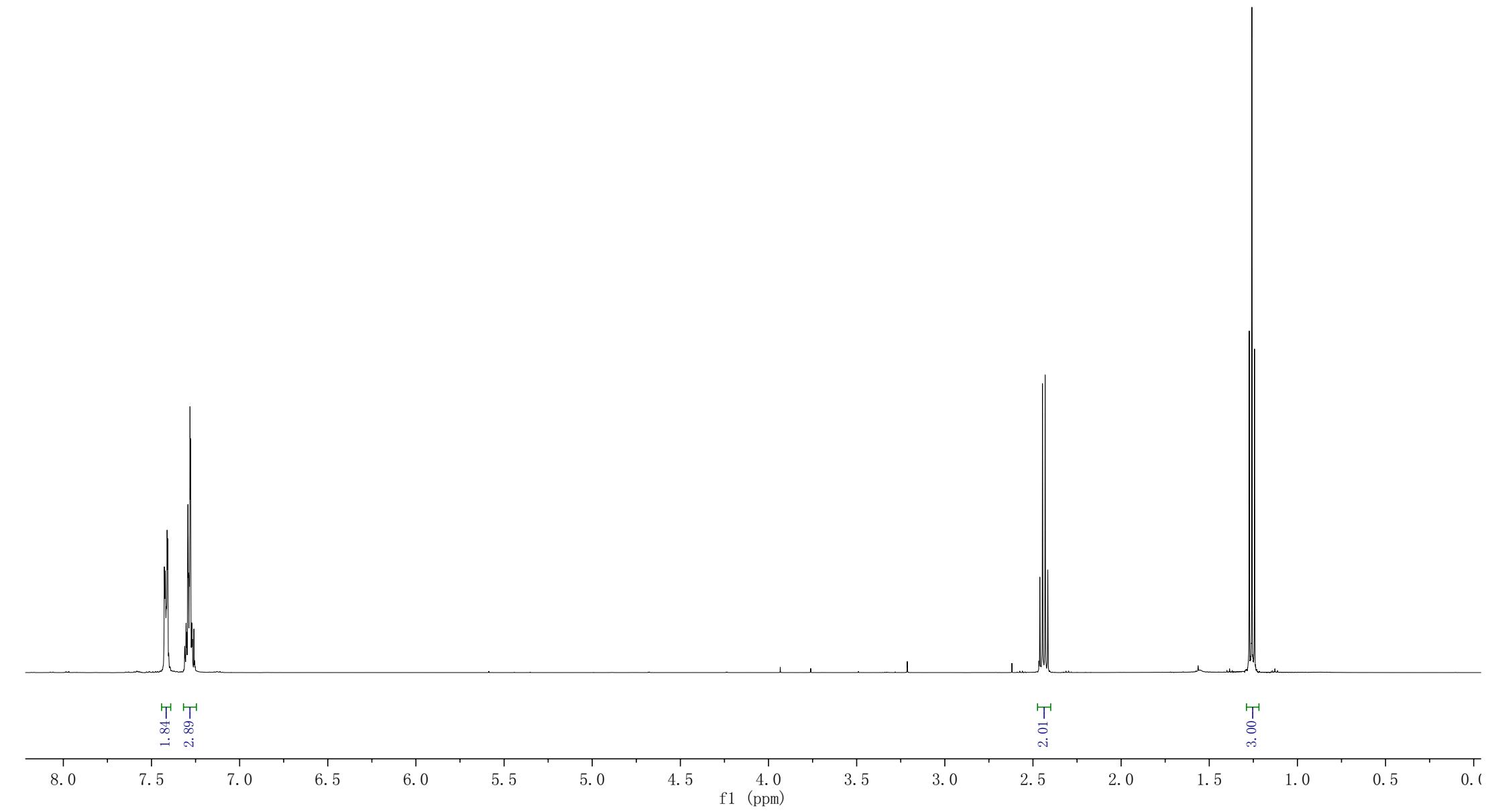
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— 127.47
— 124.00

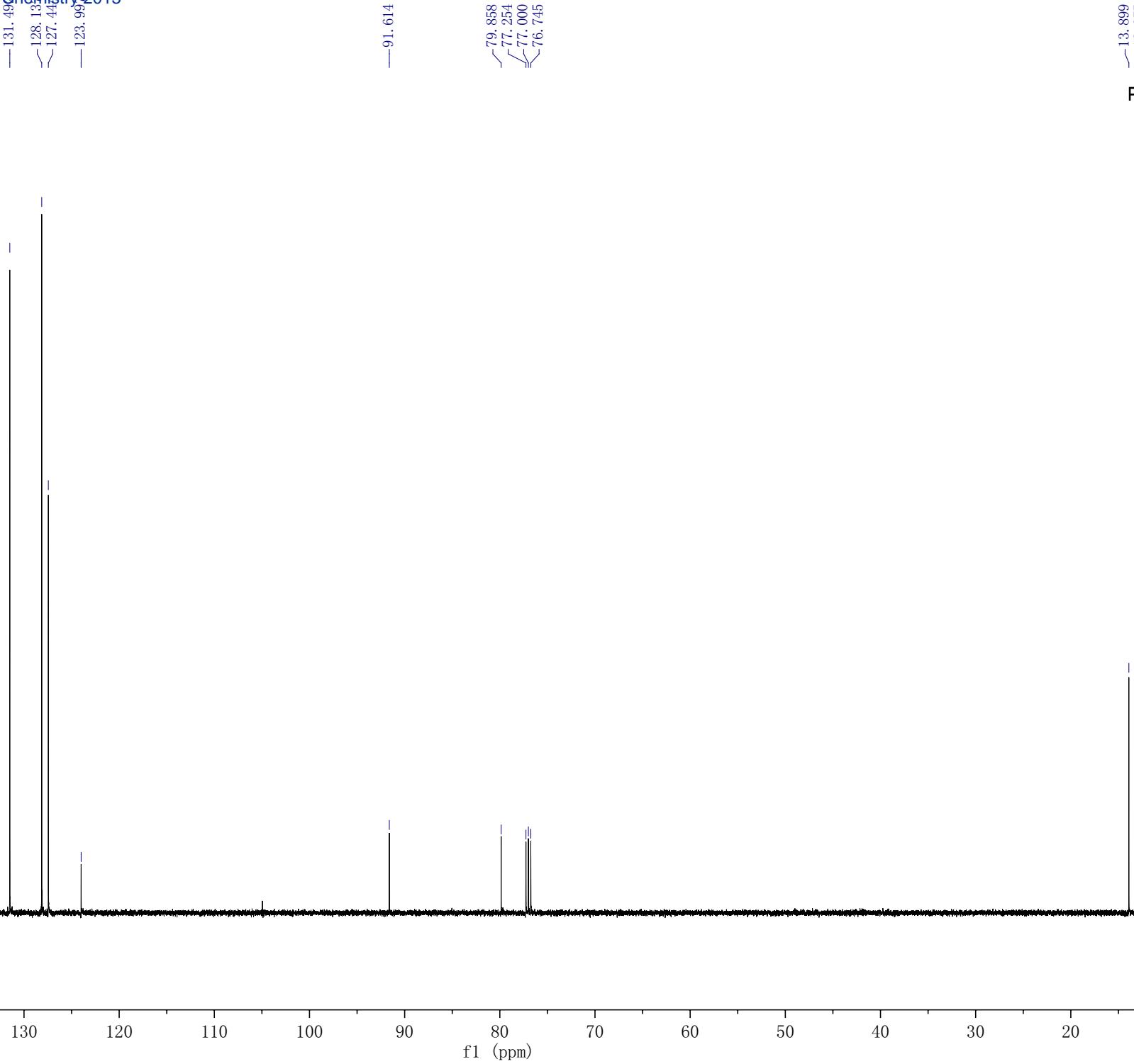
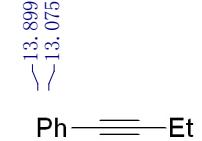
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— 77.000
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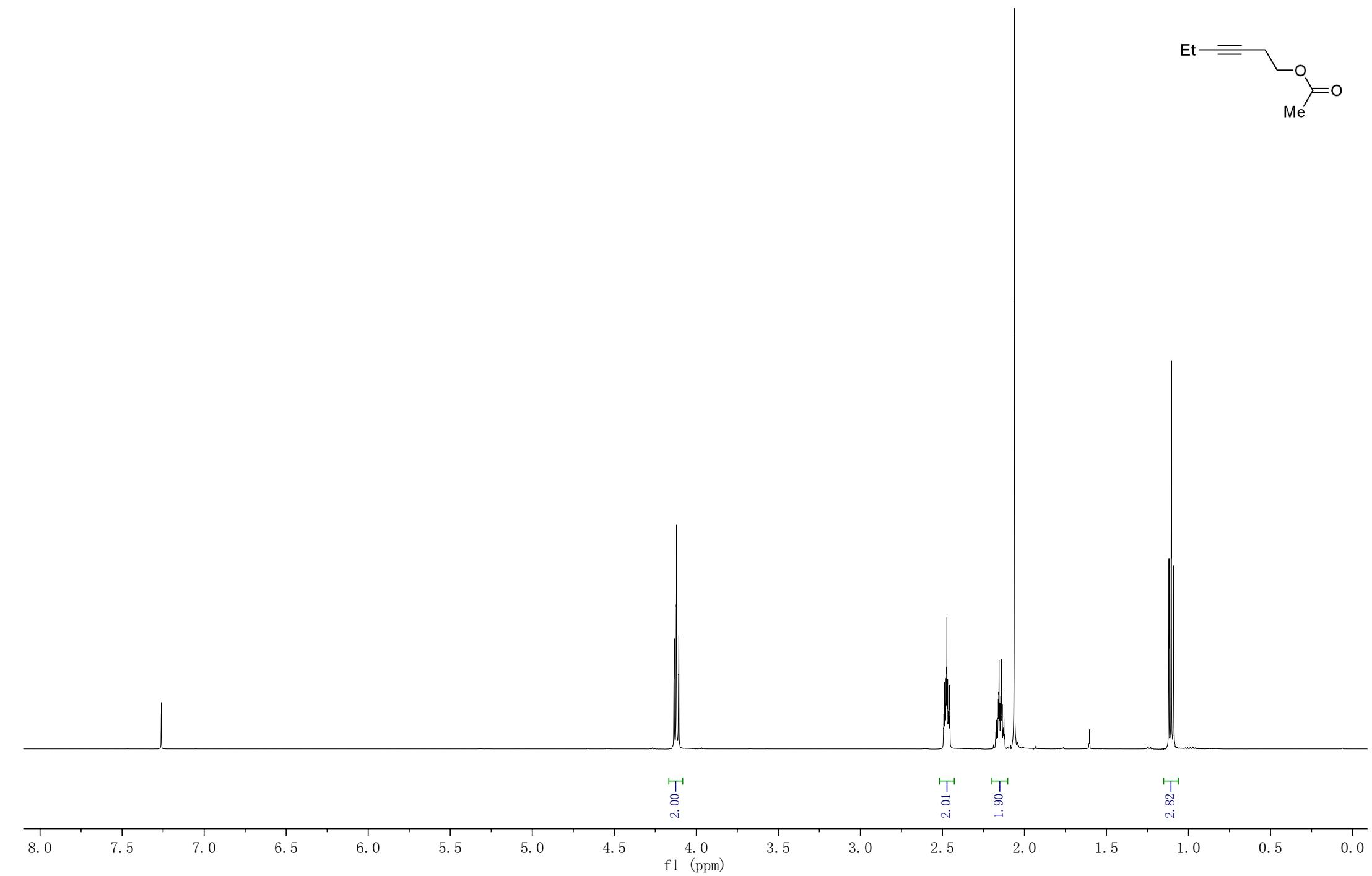
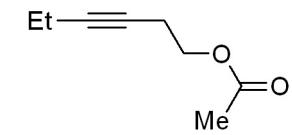
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