

Supporting Information

Symmetry-Driven Synthesis of 9-Demethyl-10,15-dideoxyryanodol

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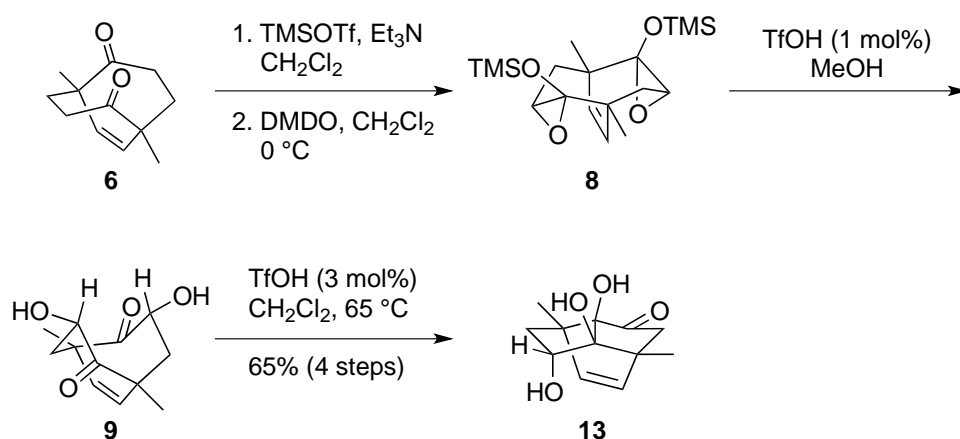
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1. Experimental

General: All reactions sensitive to air or moisture were carried out under argon atmosphere in dry solvents under anhydrous conditions, unless otherwise noted. THF, CH₂Cl₂, DMF and Et₂O were purified by Glass Contour solvent dispensing system (Nikko Hansen & Co., Ltd., Osaka, Japan). All other reagents were used as supplied. Analytical thin-layer chromatography (TLC) was performed using E. Merck Silica gel 60 F254 pre-coated plates. Flash chromatography was performed using 50-60 μm Silica Gel 60 (Kanto Chemical Co., Inc.). Melting points were measured on a Yanaco MP-S3 micro melting point apparatus or a Yanaco MP-J3 micro melting point apparatus, and are uncorrected. Infrared (IR) spectra were recorded on JASCO FT/IR-4100 spectrometer. ¹H and ¹³C NMR spectra were recorded on JEOL JNM-ECX-500, JNM-ECA-500, or JNM-ECS-400 spectrometer. Chemical shifts were reported in ppm on the δ scale relative to CHCl₃ (δ = 7.26 for ¹H NMR), CDCl₃ (δ = 77.0 for ¹³C NMR), C₆D₅H (δ = 7.16 for ¹H NMR), C₆D₆ (δ = 128.0 for ¹³C NMR), CD₂HOD (δ = 3.31 for ¹H NMR and δ = 49.0 for ¹³C NMR) as internal references. Signal patterns are indicated as s, singlet; d, doublet; t, triplet; q, quartet, m, multiplet. High resolution mass spectra were measured on Bruker microTOFII or JEOL JMS-T100LP. The numbering corresponds to those of 9-demethyl-10,15-dideoxyrynanodol (**1**).



Epoxide 8. TMSOTf (12 mL, 66 mmol) was added to a mixture of ketone **6** (4.09 g, 21.3 mmol) and Et₃N (18 mL, 130 mmol) in CH₂Cl₂ (200 mL) at room temperature. The reaction mixture was stirred for 1.5 h at room temperature, and successively diluted with hexane and saturated aqueous NaHCO₃. The resultant mixture was extracted with hexane (200 mL x3). The combined organic layers were washed with brine (200 mL), dried over Na₂SO₄, filtered and

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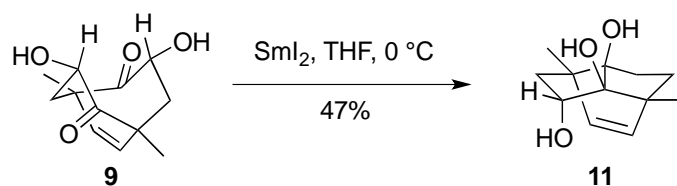
concentrated to afford the crude TMS enol ether, which was used for the next reaction without further purification.

DMDO (71 mM in acetone, 600 mL, 43 mmol) was added to a solution of the above crude TMS enol ether in CH₂Cl₂ (200 mL) at 0 °C. The reaction mixture was stirred for 10 min at 0 °C, and concentrated to afford crude epoxide **8**, which was used for the next reaction without further purification.

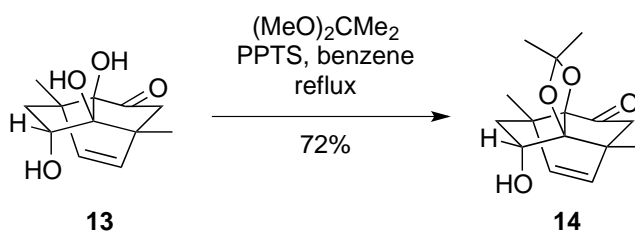
α-Hydroxy ketone 9. TfOH (19 μL, 0.21 μmol) was added to a solution of the above crude epoxide **8** in MeOH (200 mL) at room temperature. The reaction mixture was stirred for 2 h at room temperature, and concentrated to afford crude α-hydroxy ketone **9**, which was used for the next reaction without further purification. For a characterization, a small amount of **9** was purified by flash chromatography on silica gel (hexane/EtOAc 2:1): colorless crystal; m.p. 133 °C; IR (neat) ν_{\max} 3450, 1699, 1081, 923 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 1.35 (6H, s, CH₃ x 2), 1.95 (2H, dd, *J* = 14.9, 10.3 Hz, CH_AH_B x 2), 2.58 (2H, dd, *J* = 14.9, 8.6 Hz, CH_AH_B x 2), 3.59 (2 H, br s, OH x 2), 4.79 (2H, dd, *J* = 10.3, 8.6 Hz, CHOH x 2), 5.68 (2H, s, CH=C x 2); ¹³C NMR (125 MHz, CDCl₃) δ 25.8, 46.5, 48.0, 75.1, 137.1, 206.9; HRMS (ESI) calcd for C₁₂H₁₆O₄Na 247.0946 (M+Na⁺), found 247.0944.

Triol 13. TfOH (19 μL, 0.21 μmol) was added to a solution of the above crude α-hydroxy ketone **9** in CH₂Cl₂ (430 mL) at room temperature. The reaction mixture was stirred for 9 h at 65 °C in a sealed tube, and additional TfOH (19 μL, 0.21 μmol) was added. After the reaction mixture was stirred for further 24 h, TfOH (19 μL, 0.21 μmol) was added again. The reaction mixture was stirred for further 15 h and concentrated. The residue was purified by flash chromatography on silica gel (90 g, hexane/EtOAc 3:1 to 1:1 to 1:2) to afford triol **13** (3.12 g, 13.9 mmol) in 65% yield over 4 steps: colorless solid; m.p. 170 °C; IR (neat) ν_{\max} 3443, 2559, 1742, 1104 cm⁻¹; ¹H NMR (500 MHz, CD₃OD) δ 0.97 (3H, s, CH₃), 1.36 (3H, s, CH₃), 1.67 (1H, dd, *J* = 13.2, 1.2 Hz, CH_AH_BCHOH), 2.22 (1H, d, *J* = 17.2 Hz, CH_AH_BCO), 2.48 (1H, dd, *J* = 13.2, 9.8 Hz, CH_AH_BCHOH), 2.57 (1H, d, *J* = 17.2 Hz, CH_AH_BCO), 4.38 (1H, dd, *J* = 9.8, 1.2 Hz, CHOH), 5.45 (1H, d, *J* = 9.2 Hz, CH_A=CH_B), 5.65 (1H, d, *J* = 9.2 Hz, CH_A=CH_B); ¹³C NMR (125 MHz, CD₃OD) δ 18.1, 18.6, 46.8, 51.0, 52.1, 56.7, 80.7, 92.6, 92.9, 134.2, 136.7, 217.6; HRMS (ESI) calcd for C₁₂H₁₆O₄Na 247.0946 (M+Na⁺), found 247.0943.

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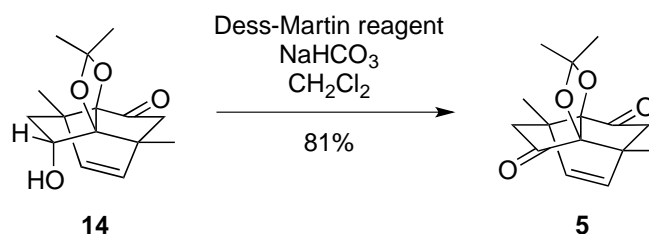
Triol 11. SmI₂ (0.1M in THF, 1.4 mL, 140 μmol) was added to a solution of α-hydroxy ketone **9** (7.6 mg, 34 μmol) in THF (1.2 mL) at 0 °C. The reaction mixture was stirred for 5 min at 0 °C, and then 0.5 M HCl solution was added. The resultant solution was extracted with EtOAc (x3), and the combined organic layers were washed with saturated aqueous Na₂S₂O₃ and brine, dried over Na₂SO₄, and concentrated. The residue was purified by flash chromatography on silica gel (hexane/EtOAc 5:1 to 3:1) to afford triol **11** (3.3 mg, 16 μmol) in 47% yield: white solid; m.p. 149 °C; IR (neat) ν_{max} 3476, 3499, 2930, 1457, 1278, 1110, 1019 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 0.99 (3H, s, CH₃), 1.21 (3H, s, CH₃), 1.41-1.52 (2H, m, CH_AH_BCH_AH_B), 1.58 (1H, d, *J* = 13.2 Hz, CH_AH_BCHOH), 1.90 (1H, br s, OH), 2.40 (1H, br s, OH), 2.82 (1H, br s, OH), 2.01-2.12 (2H, m, CH_AH_BCH_AH_B), 2.54 (1H, dd, *J* = 13.2, 9.2 Hz, CH_AH_BCHOH), 4.26 (1H, d, *J* = 9.2 Hz, CH₂CHOH), 5.49 (1H, d, *J* = 9.2 Hz, CH_A=CH_B), 5.51 (1H, d, *J* = 9.2 Hz, CH_A=CH_B); ¹³C NMR (125 MHz, CDCl₃) δ 16.2, 18.8, 29.5, 40.5, 50.2, 50.5, 50.8, 81.0, 90.9, 91.4, 133.7, 135.8; HRMS (ESI) calcd for C₁₂H₁₈O₃Na 233.1154 (M+Na⁺), found 233.1144.



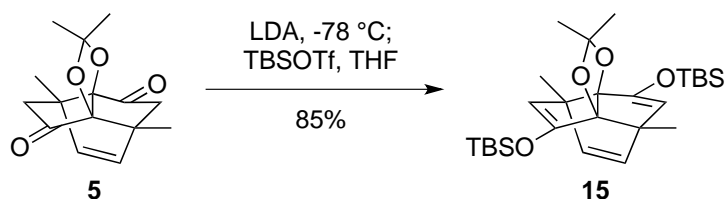
Acetonide 14. A solution of triol **13** (3.12 g, 13.9 mmol), PPTS (3.49 g, 13.9 mmol) in a mixture of 2,2-dimethoxypropane (140 mL) in benzene (140 mL) was refluxed for 3.5 days. After addition of MeOH (100 mL), the resultant solution was concentrated. The procedure was repeated three times. The resultant residue was purified by flash chromatography on silica gel (150 g, hexane/EtOAc 5:1 to 3:1 to 1:1) to afford acetonide **14** (2.65 g, 10.0 mmol) in 72% yield: colorless solid; m.p. 90 °C; IR (neat) ν_{max} 3494, 2993, 1750, 1068, 1031 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 1.12 (3H, s, CH₃), 1.34 (3H, s, CH₃), 1.46 (3H, s, CH₃), 1.50 (3H, s, CH₃), 1.74 (1H, d, *J* = 13.7 Hz, CH_AH_BCHOH), 2.03 (1H, br d, *J* = 8.0 Hz, OH), 2.38 (1H, d, *J* = 17.2 Hz, CH_AH_BCO), 2.71 (1H, dd, *J* = 13.7,

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9.2 Hz, $\text{CH}_\text{A}\text{H}_\text{B}\text{CHOH}$), 2.77 (1H, d, $J = 17.2$ Hz, $\text{CH}_\text{A}\text{H}_\text{B}\text{CO}$), 4.39 (1H, br dd, $J = 9.2, 8.0$ Hz, CHOH), 5.61 (1H, d, $J = 9.8$ Hz, $\text{CH}_\text{A}=\text{CH}_\text{B}$), 5.84 (1H, d, $J = 9.2$ Hz, $\text{CH}_\text{A}=\text{CH}_\text{B}$); ^{13}C NMR (125 MHz, CDCl_3) δ 16.7, 17.7, 28.0, 28.1, 45.3, 49.7, 52.0, 56.5, 78.7, 96.9, 100.3, 115.4, 135.0, 137.4, 214.7; HRMS (ESI) calcd for $\text{C}_{15}\text{H}_{20}\text{O}_4\text{Na}$ 287.1259 ($\text{M}+\text{Na}^+$), found 287.1253.



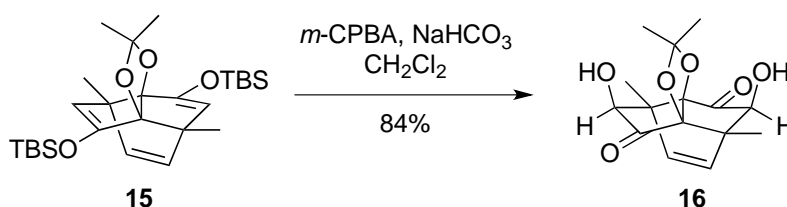
Ketone 5. Dess-Martin periodinane (6.40 g, 15.1 mmol) was added to a suspension of acetonide **14** (2.65 g, 10.0 mmol) in CH_2Cl_2 (100 mL) at room temperature. The reaction mixture was stirred for 80 min at room temperature, and saturated aqueous $\text{Na}_2\text{S}_2\text{O}_3$ (50 mL) and NaHCO_3 (50 mL) were successively added. The resultant mixture was extracted with EtOAc (100 mL x2), and the combined organic layers were dried over Na_2SO_4 , filtered and concentrated. The residue was purified by flash chromatography on silica gel (50 g, hexane/EtOAc 5:1) to afford ketone **5** (2.12 g, 8.09 mmol) in 81% yield: colorless solid; m.p. 179 °C; IR (neat) ν_{max} 2931, 1750, 1075 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 1.35 (6H, s, $\text{CH}_3 \times 2$), 1.40 (6H, s, $\text{CH}_3 \times 2$), 2.48 (2H, d, $J = 17.2$ Hz, $\text{CH}_\text{A}\text{H}_\text{B} \times 2$), 2.83 (2H, d, $J = 17.2$ Hz, $\text{CH}_\text{A}\text{H}_\text{B} \times 2$), 5.68 (2H, s, $\text{CH}=\text{C} \times 2$); ^{13}C NMR (125 MHz, CDCl_3) δ 16.4, 27.5, 44.9, 55.0, 94.6, 116.9, 135.4, 212.3; HRMS (ESI) calcd for $\text{C}_{15}\text{H}_{18}\text{O}_4\text{Na}$ 285.1103 ($\text{M}+\text{Na}^+$), found 285.1098.



TBS-enol ether 15. $n\text{-BuLi}$ (1.6 M in hexane, 11 mL, 18 mmol) was added to a solution of $i\text{-Pr}_2\text{NH}$ (2.5 μL , 18 mmol) in THF (10 mL) at -78 °C. The mixture was stirred for 30 min at 0 °C, and cooled to -78 °C. A solution of ketone **5** (465 mg, 1.77 μmol) in THF (8 mL) was added to the mixture via cannula. The reaction mixture was stirred for further 30 min at -78 °C, and then TBSOTf (2.0 mL, 8.7 mmol) was added. The reaction mixture was allowed to warm to room temperature and stirred for further 1 h, and then saturated aqueous NH_4Cl (15

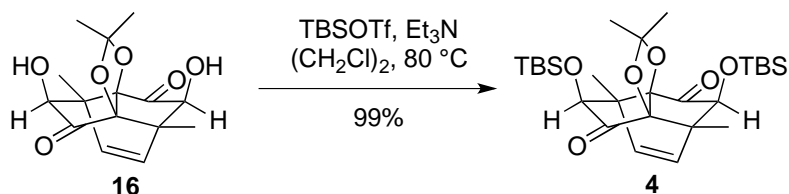
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mL) was added. The resultant mixture was extracted with Et₂O (20 mL x3), and the combined organic layers were washed with brine (20 mL), dried over Na₂SO₄, filtered and concentrated. The residue was purified by flash chromatography on silica gel (Fujisilysia BW-300, 50 g, hexane/EtOAc 1:0 to 70:1 to 30:1) to afford TBS-enol ether **15** (733 mg, 1.50 mmol) in 85% yield: pale yellow solid; m.p. 182-183 °C; IR (neat) ν_{\max} 2935, 1624, 1319, 1247, 1138 cm⁻¹; ¹H NMR (400 MHz, C₆D₆) δ 0.11 (6H, s, CH₃ of TBS x 2), 0.15 (6H, s, CH₃ of TBS x 2), 0.98 (18H, s, *t*-Bu of TBS x 2), 1.31 (6H, s, CH₃ x 2), 1.73 (6H, s, CH₃ x 2), 5.06 (2H, s, CH=COTBS x 2), 5.92 (2H, s, CH=C x 2); ¹³C NMR (100 MHz, C₆D₆) δ -4.7, -4.3, 17.2, 18.1, 25.7, 28.5, 48.3, 102.6, 115.1, 119.1, 141.5, 149.3; HRMS (ESI) calcd for C₂₇H₄₆O₄Si₂Na 513.2832 (M+Na⁺), found 513.2831.

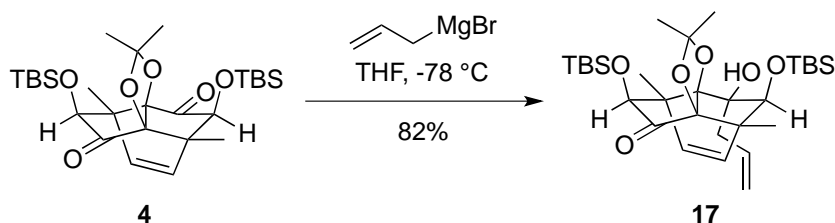


α -Hydroxy ketone 16. A solution of *m*-CPBA (purified by washing with pH 7 phosphate buffer and drying in vacuo, 1.0 g, 5.8 mmol) was added to a mixture of TBS enol ether **15** (733 mg, 1.50 mmol) and NaHCO₃ (1.3 g, 15 mmol) in CH₂Cl₂ (15 mL) at 0 °C. The reaction mixture was stirred for 1.5 h at room temperature, and saturated aqueous Na₂S₂O₃ (15 mL) was added. The resultant mixture was extracted with EtOAc (50 mL x3), and the combined organic layers were dried over Na₂SO₄, filtered and concentrated. The residue was purified by flash column chromatography on silica gel (50 g, hexane/EtOAc 3:1 to 0:1) to afford α -hydroxy ketone **16** (369 mg, 1.26 mmol) in 84% yield: colorless solid; m.p. 220 °C; IR (neat) ν_{\max} 3416, 2996, 1745, 1445, 1379, 1220, 1078 cm⁻¹; ¹H NMR (500 MHz, CD₃OD) δ 1.33 (6H, s, CH₃ x 2), 1.39 (6H, s, CH₃ x 2), 3.65 (2H, s, CHOH x 2), 5.67 (2H, s, CH=C x 2); ¹³C NMR (125 MHz, CD₃OD) δ 12.1, 27.9, 51.2, 83.1, 95.9, 118.9, 136.6, 216.0; HRMS (ESI) calcd for C₁₅H₁₈O₆Na 317.1001 (M+Na⁺), found 317.1003.

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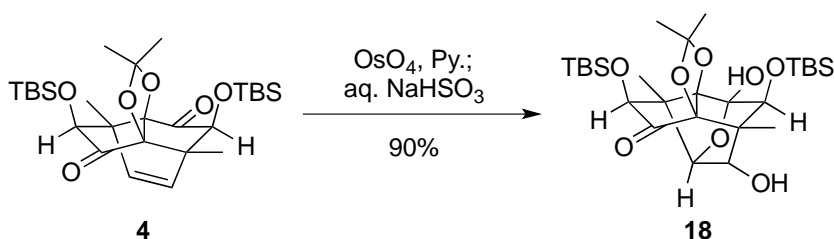
TBS-ether 4. TBSOTf (700 μ L, 3.1 mmol) was added to a mixture of α -hydroxy ketone **16** (90 mg, 310 μ mol) and Et₃N (860 μ L, 6.1 mmol) in (CH₂Cl)₂ (3.1 mL) at room temperature. The reaction mixture was stirred for 4 h at 80 °C, and then cooled to 0 °C. Saturated aqueous NH₄Cl (3 mL) was added, and the resultant mixture was concentrated to remove (CH₂Cl)₂. The aqueous solution was extracted with EtOAc (3 mL x3), and the combined organic layers were washed with brine (3 mL), dried over MgSO₄, filtered and concentrated. The residue was purified by flash column chromatography on silica gel (10 g; hexane/EtOAc 20:1) to afford TBS-ether **4** (160 mg, 306 μ mol) in 99% yield: colorless solid; m.p. 150-152 °C; IR (neat) ν_{\max} 2931, 1765, 1257, 1102 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 0.09 (6H, s, CH₃ of TBS x 2), 0.10 (6H, s, CH₃ of TBS x 2), 0.88 (18H, s, *t*-Bu of TBS x 2), 1.33 (6H, s, CH₃ x 2), 1.39 (6H, s, CH₃ x 2), 3.64 (2H, s, CHOTBS x 2), 5.57 (2H, s, CH=C x 2); ¹³C NMR (125 MHz, CDCl₃) δ -5.5, -4.6, 12.3, 18.2, 25.6, 27.6, 50.7, 82.1, 94.2, 117.9, 135.5, 213.8; HRMS (ESI) calcd for C₂₇H₄₆O₆Si₂Na 545.2731 (M+Na⁺), found 545.2714.



Allyl alcohol 17. Allyl magnesium bromide (1.0 M in Et₂O, 180 μ L, 180 μ mol) was added to a solution of **4** (4.7 mg, 9.0 μ mol) in THF (0.9 mL) at -78 °C. The reaction mixture was stirred for 1 h at -78 °C, and then saturated aqueous NH₄Cl (5 mL) was added. The resultant mixture was stirred at room temperature, and extracted with CH₂Cl₂ (3 mL x3). The combined organic layers were dried over Na₂SO₄, filtered and concentrated. The residue was purified by flash column chromatography on silica gel (2 g, hexane/EtOAc 30:1 to 10:1) to afford allyl alcohol **17** (4.2 mg, 7.4 μ mol) in 82% yield: colorless oil; IR (neat) ν_{\max} 3469, 2936, 1755, 1464, 1373, 1255, 1097 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 0.10 (9H, s, CH₃ of TBS x 3), 0.11 (3H, s, CH₃ of TBS), 0.90 (9H, s, *t*-Bu of TBS), 0.94 (9H, s, *t*-Bu of TBS), 1.17 (3H, s, CH₃), 1.42 (3H, s, CH₃), 1.46 (3H, s, CH₃), 1.67 (3H,

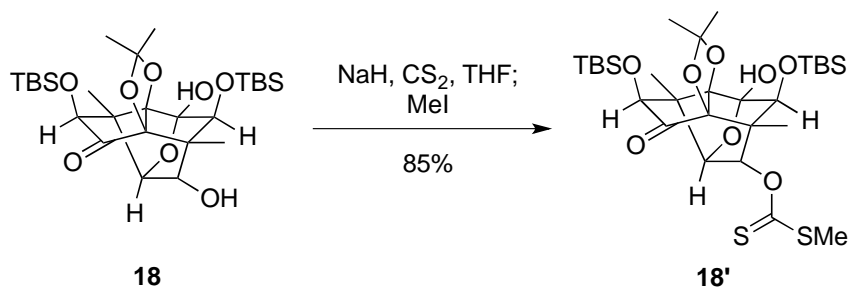
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s, CH_3), 2.34 (1H, dd, $J = 13.5, 10.0$ Hz, $CH_2=CHCH_AH_B$), 2.92 (1H, ddt, $J = 13.5, 5.0, 1.5$ Hz, $CH_2=CHCH_AH_B$), 3.51 (1H, s, $CH(OTBS)CO$), 3.54 (1H, s, OH), 3.79 (1H, s, $CH(OTBS)COH$), 5.06 (1H, br d, $J = 17.5$ Hz, $CH_AH_B=CH$), 5.09 (1H, dt, $J = 10.0, 1.5$ Hz, $CH_AH_B=CH$), 5.33 (1H, d, $J = 9.0$ Hz, $CH=CHCC(OTBS)C(OH)$), 5.65 (1H, d, $J = 9.0$ Hz, $CH=CHCC(OTBS)C(OH)$), 5.95 (1H, dtd, $J = 17.5, 10.0, 5.0$ Hz, $CH_2=CHCH_AH_B$); ^{13}C NMR (100 MHz, $CDCl_3$) δ -5.3, -4.6, -4.1, -3.7, 14.0, 14.5, 18.3, 18.4, 25.8, 25.9, 27.7, 29.4, 47.3, 51.2, 51.6, 79.0, 82.3, 83.7, 96.8, 98.3, 117.3, 117.7, 132.5, 136.1, 138.0, 217.5; HRMS (ESI) calcd for $C_{30}H_{52}O_6Si_2Na$ 587.3195 ($M+Na^+$), found 587.3205.

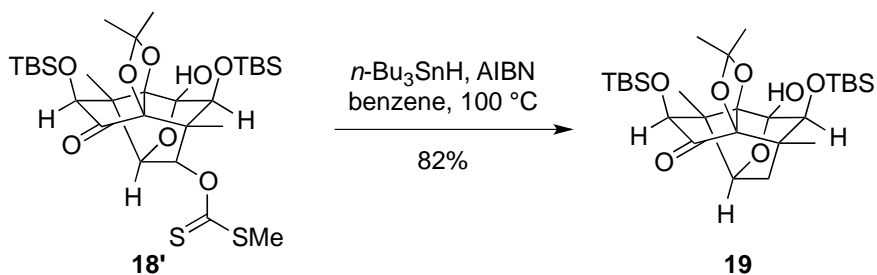


Hemiacetal 18. OsO_4 (609 mg, 2.40 mmol) was added to a solution of TBS-ether **4** (501 mg, 958 μmol) in pyridine (9.6 mL) at room temperature. The reaction mixture was stirred for 5 h at room temperature, and then saturated aqueous $NaHSO_3$ (20 mL) was added. The resultant mixture was diluted with EtOAc (20 mL) and stirred for further 17 h. The solution was extracted with EtOAc (20 mL x3). The combined organic layers were washed with brine (10 mL), dried over Na_2SO_4 , filtered and concentrated. The residue was purified by flash column chromatography on silica gel (15 g, hexane/EtOAc 3:1) to afford hemiacetal **18** (480 mg, 862 μmol) in 90% yield: colorless solid; m.p. 223-225 $^\circ\text{C}$; IR (neat) ν_{max} 3457, 2931, 1759, 1643, 1252 cm^{-1} ; 1H NMR (500 MHz, $CDCl_3$) δ 0.09 (3H, s, CH_3 of TBS), 0.11 (3H, s, CH_3 of TBS), 0.12 (3H, s, CH_3 of TBS), 0.13 (3H, s, CH_3 of TBS), 0.87 (9H, s, t -Bu of TBS), 0.90 (9H, s, t -Bu of TBS), 1.17 (3H, s, CH_3), 1.31 (3H, s, CH_3), 1.38 (3H, s, CH_3), 1.56 (3H, s, CH_3), 2.58 (1H, br s, OH), 3.34 (1H, br s, $CHOH$), 3.65 (1H, br s, OH), 3.72 (1H, s, $CHOTBSCO$), 3.86 (1H, d, $J = 1.2$ Hz, $CHOCO$), 4.15 (1H, s, $CHOTBSCO$); ^{13}C NMR (125 MHz, $CDCl_3$) δ -5.5, -5.2, -4.6, -4.4, 10.1, 12.3, 18.3, 18.6, 25.6, 25.9, 27.6, 27.7, 50.8, 55.1, 73.4, 75.7, 81.1, 83.4, 96.9, 97.1, 105.3, 121.2, 215.2; HRMS (ESI) calcd for $C_{27}H_{48}O_8Si_2Na$ 579.2785 ($M+Na^+$), found 579.2792.

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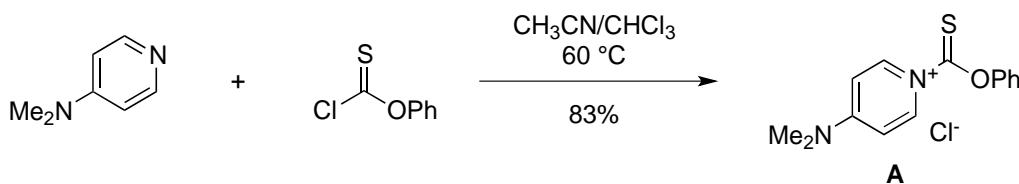
Xanthate 18'. NaH in mineral oil (>60%) was washed with hexane and dried in vacuo before use. The purified NaH (119 mg, 5.17 mmol) and CS₂ (600 μ L, 9.9 mmol) were successively added to a solution of the hemiacetal **18** (554 mg, 995 μ mol) in THF (20 mL) at 0 °C. The reaction mixture was stirred for 30 min at room temperature, and then MeI (930 μ L, 15.0 mmol) was added. The reaction mixture was stirred for further 30 min at room temperature, and cooled to 0 °C. Saturated aqueous NH₄Cl (20 mL) was added, and the resultant mixture was extracted with Et₂O (20 mL x3). The combined organic layers were washed with brine (10 mL), dried over Na₂SO₄, filtered and concentrated. The residue was purified by flash column chromatography on silica gel (20 g, hexane/EtOAc 20:1 to 10:1) to afford xanthate **18'** (550 mg, 850 μ mol) in 85% yield: pale yellow solid; m.p. 77-78 °C; IR (neat) ν_{\max} 3487, 2930, 1762, 1252, 1072 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 0.12 (3H, s, CH₃ of TBS), 0.14 (3H, s, CH₃ of TBS), 0.15 (3H, s, CH₃ of TBS), 0.16 (3H, s, CH₃ of TBS), 0.89 (9H, s, *t*-Bu of TBS), 0.92 (9H, s, *t*-Bu of TBS), 1.14 (3H, s, CH₃), 1.33 (3H, s, CH₃), 1.40 (3H, s, CH₃), 1.59 (3H, s, CH₃), 2.58 (3H, s, SCH₃), 3.57 (1H, s, OH), 3.89 (1H, s, CHOTBSCO), 4.20 (1H, d, *J* = 1.2 Hz, CHOCOH), 4.39 (1H, s, CHOTBSCO), 5.34 (1H, d, *J* = 1.2 Hz, CHOCSSCH₃); ¹³C NMR (125 MHz, CDCl₃) δ -5.5, -5.2, -4.6, -4.4, 10.1, 12.1, 18.3, 18.6, 18.9, 25.6, 25.9, 27.6, 27.7, 51.9, 54.3, 75.4, 80.5, 81.7, 81.9, 96.7, 97.3, 104.9, 121.4, 214.7, 215.3; HRMS (ESI) calcd for C₂₉H₅₀O₈S₂Si₂Na 669.2383 (M+Na⁺), found 669.2393.



Hemiacetal 19. A mixture of the xanthate **18'** (550 mg, 850 μ mol), *n*-Bu₃SnH (1.8 mL, 6.8 mmol) and AIBN (42 mg, 260 μ mol) in benzene (85 mL) was

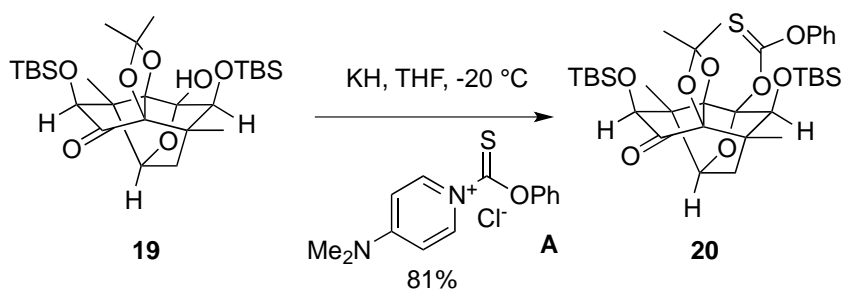
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degassed by a freeze-thaw procedure (x3). The reaction mixture was stirred for 3 h at reflux temperature, cooled to room temperature, and concentrated. The residue was purified by flash column chromatography on 10% (w/w) KF contained silica gel (15 g, hexane/EtOAc 20:1 to 10:1) to afford hemiacetal **19** (375 mg, 693 μmol) in 82% yield: colorless solid; m.p. 145-146 $^{\circ}\text{C}$; IR (neat) ν_{max} 3487, 2931, 1759, 1461, 1251 cm^{-1} ; ^1H NMR (500 MHz, CDCl_3) δ 0.09 (3H, s, CH_3 of TBS), 0.11 (3H, s, CH_3 of TBS), 0.12 (3H, s, CH_3 of TBS), 0.14 (3H, s, CH_3 of TBS), 0.89 (9H, s, *t*-Bu of TBS), 0.91 (9H, s, *t*-Bu of TBS), 1.12 (3H, s, CH_3), 1.27 (1H, dd, $J = 15.1, 1.4$ Hz, CH_AH_B), 1.32 (3H, s, CH_3), 1.40 (3H, s, CH_3), 1.57 (3H, s, CH_3), 1.77 (1H, dd, $J = 15.1, 3.2$ Hz, CH_AH_B), 3.50 (1H, s, OH), 3.67 (1H, s, CHOTBS), 3.85 (1H, s, CHOTBS), 3.94 (1H, dd, $J = 3.2, 1.4$ Hz, CHOCH_2); ^{13}C NMR (125 MHz, CDCl_3) δ -5.4, -5.2, -4.5, -4.3, 12.5, 13.4, 18.3, 18.6, 25.6, 25.9, 27.6, 27.7, 38.4, 49.4, 54.3, 76.2, 79.2, 86.2, 97.6, 97.9, 104.9, 119.7, 215.8; HRMS (ESI) calcd for $\text{C}_{27}\text{H}_{48}\text{O}_7\text{Si}_2\text{Na}$ 563.2836 ($\text{M}+\text{Na}^+$), found 563.2857.



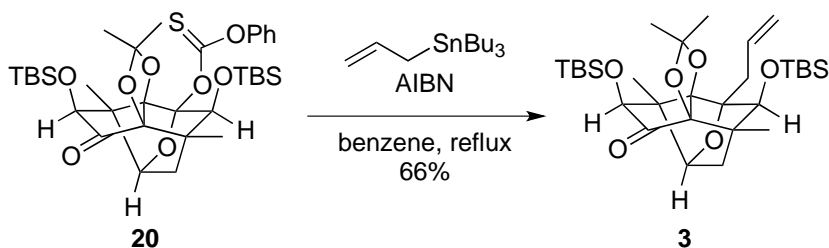
Pyridinium salt A. *O*-Phenyl chlorothionoformate (5.0 mL, 36 mmol) was added to a solution of DMAP (4.4 g, 36 mmol) in MeCN (36 mL) at room temperature. The reaction mixture was stirred at room temperature for 1 h at room temperature, and then CHCl_3 (36 mL) was added. The reaction mixture was heated $60\text{ }^{\circ}\text{C}$ until the solid was dissolved. The resultant solution was cooled to room temperature, and Et_2O (36 mL) was added to crystallize pyridinium salt **A**. The solid was filtered with Et_2O (10 mL), and the filtrate was concentrated to afford pyridinium salt **A** (8.8 g, 30 mmol) in 83% yield: yellow solid; m.p. 199-200 $^{\circ}\text{C}$; IR (neat) ν_{max} 3428, 3073, 1651, 1493, 1371, 1261, 1183 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3) δ 3.61 (6H, s, $\text{NCH}_3 \times 2$), 7.17 (2H, d, $J = 7.8$ Hz, $\text{OC}=\text{CH} \times 2$), 7.38 (1H, t, $J = 7.8$ Hz, $\text{OC}=\text{CH}-\text{CH}=\text{CH}$), 7.49 (2H, t, $J = 7.8$ Hz, $\text{OC}=\text{CH}-\text{CH} \times 2$), 7.60 (2H, d, $J = 8.7$ Hz, $\text{CH}=\text{CH}-\text{N}^+=\text{C} \times 2$), 9.12 (2H, d, $J = 8.7$ Hz, $\text{CH}=\text{CH}-\text{N}^+=\text{C} \times 2$); ^{13}C NMR (100 MHz, CDCl_3) δ 42.1, 108.9, 121.1, 127.6, 129.9, 137.1, 153.0, 158.2, 183.1; HRMS (ESI) calcd for $\text{C}_{14}\text{H}_{15}\text{N}_2\text{OS}$ 259.0900 (M^+), found 259.0900.

Supporting Information



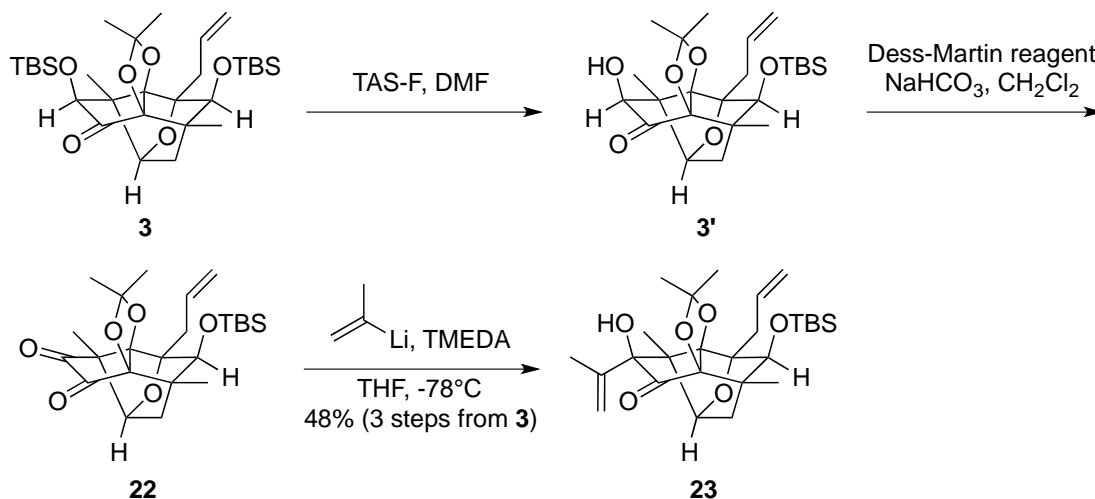
Thiocarbonate 20. KH in mineral oil (>30%) was washed with hexane and dried in vacuo. The purified KH (695 mg, 17.3 mmol) was added to a solution of hemiacetal **19** (375 mg, 693 μ mol) in THF (23 mL) at -78 °C. The mixture was stirred for 20 min at -78 °C, and then freshly prepared pyridinium salt **A** (2.0 g, 6.9 mmol) was added. The reaction mixture was allowed to warm to -20 °C and stirred for 19 h. The reaction was quenched with AcOH (1.5 mL). The resultant mixture was neutralized with saturated aqueous NaHCO₃ (15 mL), and extracted with Et₂O (20 mL x3). The combined organic layers were washed with brine (20 mL), dried over Na₂SO₄, filtered and concentrated. The residue was purified by flash column chromatography on silica gel (30 g, hexane/EtOAc 20:1 to 10:1) to afford thiocarbonate **20** (379 mg, 560 μ mol) in 81% yield: pale yellow amorphous; IR (neat) ν_{\max} 2932, 2858, 1761, 1461, 1286, 1120, 1053 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 0.13 (3H, s, CH₃ of TBS), 0.15 (6H, s, CH₃ of TBS x 2), 0.22 (3H, s, CH₃ of TBS), 0.88 (9H, s, *t*-Bu of TBS), 0.95 (9H, s, *t*-Bu of TBS), 1.22 (3H, s, CH₃), 1.30 (1H, br d, *J* = 15.6 Hz, CH_AH_B), 1.40 (3H, s, CH₃), 1.42 (3H, s, CH₃), 1.64 (3H, s, CH₃), 1.90 (1H, dd, *J* = 15.6, 3.6 Hz, CH_AH_B), 3.70 (1H, s, CHOTBSCO), 4.14 (1H, d, *J* = 3.6 Hz, CHOCH₂), 4.65 (1H, s, CHOTBSCCOSOPh), 7.15 (2H, m, aromatic), 7.29 (1H, m, aromatic), 7.42 (2H, m, aromatic); ¹³C NMR (125 MHz, CDCl₃) δ -5.5, -4.8, -4.5, -3.2, 13.1, 14.3, 18.3, 19.0, 25.6, 26.7, 27.5, 27.8, 38.2, 49.6, 53.8, 76.1, 80.8, 85.4, 97.1, 99.6, 108.8, 119.9, 122.0, 126.6, 129.5, 153.2, 190.0, 215.2; HRMS (ESI) calcd for C₃₄H₅₂O₈SSi₂Na 699.2819 (M+Na⁺), found 699.2800.

Supporting Information



Compound 3. A solution of thiocarbonate **20** (67 mg, 99 μmol), allyltributyltin (460 μL , 1.5 mmol), and AIBN (3 mg, 20 μmol) in benzene (2.0 mL) was degassed by freeze-thaw procedure (x3). The reaction mixture was stirred for 3 h at reflux temperature, cooled to room temperature, and concentrated. The residue was purified by flash chromatography on 10% (w/w) KF contained silica gel (15 g, hexane/EtOAc 30:1 to 15:1) to afford **3** with a small amount of impurity. The mixture was further purified by flash chromatography on 10% (w/w) KF contained silica gel (7 g, hexane/CH₂Cl₂ 2:1 to 1:1) to afford **3** (37 mg, 66 μmol) in 66% yield: colorless oil; IR (neat) ν_{max} 2931, 2858, 1759, 1461, 1250, 1111 cm^{-1} ; ¹H NMR (400 MHz, CDCl₃) δ 0.09 (3H, s, CH₃ of TBS), 0.10 (3H, s, CH₃ of TBS), 0.12 (3H, s, CH₃ of TBS), 0.14 (3H, s, CH₃ of TBS), 0.89 (9H, s, *t*-Bu of TBS), 0.92 (9H, s, *t*-Bu of TBS), 1.11 (3H, s, CH₃), 1.29 (1H, d, $J = 15.1$ Hz, CHOCH_AH_B), 1.29 (3H, s, CH₃), 1.37 (3H, s, CH₃), 1.56 (3H, s, CH₃), 1.78 (1H, dd, $J = 15.1, 3.2$ Hz, CHOCH_AH_B), 2.40 (1H, br dd, $J = 14.2, 8.7$ Hz, CCH_AH_BCH=CH₂), 2.61 (1H, br dd, $J = 14.2, 9.2$ Hz, CCH_AH_BCH=CH₂), 3.66 (1H, s, CHOTBS), 3.81 (1H, br d, $J = 3.2$ Hz, CHOCH₂), 3.86 (1H, s, CHOTBS), 5.14 (1H, br d, $J = 10.5$ Hz, CH₂CH=CH_AH_B), 5.15 (1H, br d, $J = 17.8$ Hz, CH₂CH=CH_AH_B), 5.95 (1H, dddd, $J = 17.8, 10.5, 9.2, 8.7$ Hz, CH₂CH=CH₂); ¹³C NMR (100 MHz, CDCl₃) δ -5.5, -4.5, -4.3, -3.9, 13.2, 14.3, 18.3, 18.4, 25.7, 26.1, 27.9 (two peaks), 34.8, 38.7, 49.9, 55.6, 76.3, 80.5, 88.7, 89.6, 99.2, 101.4, 117.7, 118.2, 134.2, 216.9; HRMS (ESI) calcd for C₃₀H₅₂O₆Si₂Na 587.3200 (M+Na⁺), found 587.3223.

Supporting Information



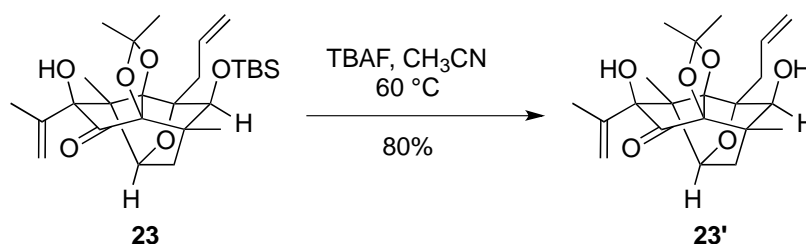
α -Hydroxy ketone 3'. A solution of **3** (84 mg, 149 μ mol) in DMF (1.5 mL) was degassed by freeze-thaw procedure (x3), and cooled to 0 °C. A solution of TAS-F (61 mg, 222 μ mol) in DMF (1.5 mL), degassed by freeze-thaw procedure (x3), was added to the solution via cannula. Then the reaction mixture was stirred for 40 min at room temperature, and cooled to 0 °C. Saturated aqueous NH₄Cl (3 mL) was added, and the resultant mixture was extracted with EtOAc (5 mL x3). The combined organic layers were washed with water (10 mL x2) and brine (10 mL), dried over Na₂SO₄, filtered and concentrated. The residue was filtered through a short pad of silica gel (3 g, hexane/EtOAc 1:1), and the filtrate was concentrated to afford crude α -hydroxy ketone **3'** (64 mg), which was used for the next reaction without further purification.

Diketone 22. Dess-Martin periodinane (188 mg, 443 μ mol) was added to a suspension of the above crude α -hydroxy ketone **3'** (64 mg) and NaHCO₃ (187 mg, 2.25 mmol) in CH₂Cl₂ (3.0 mL) at 0 °C. The reaction mixture was stirred for 50 min at 0 °C, and saturated aqueous Na₂S₂O₃ (3 mL) was added. The resultant mixture was extracted with Et₂O (5 mL x3), and the combined organic layers were washed with saturated aqueous Na₂CO₃ (10 mL x3) and brine (10 mL), dried over Na₂SO₄, filtered and concentrated to afford crude diketone **22** (69 mg) as pale red oil, which was used for the next reaction without further purification.

Allyl alcohol 23. *t*-BuLi (1.65 M in *n*-pentane, 3.6 mL, 5.9 mmol) was added to a solution of 2-bromopropene (260 μ L, 2.9 mmol) in THF (12 mL) at -78 °C. The mixture was stirred for 20 min at -78 °C, and then TMEDA (440 μ L, 2.9 mmol) was added. The mixture was stirred for 10 min, and then a solution of the above crude diketone **22** (69 mg) in THF (3 mL) was added at -78 °C via cannula. The reaction mixture was stirred for 50 min at -78 °C, and then saturated aqueous NH₄Cl (10 mL) was added. The resultant mixture was extracted with EtOAc (15

Supporting Information

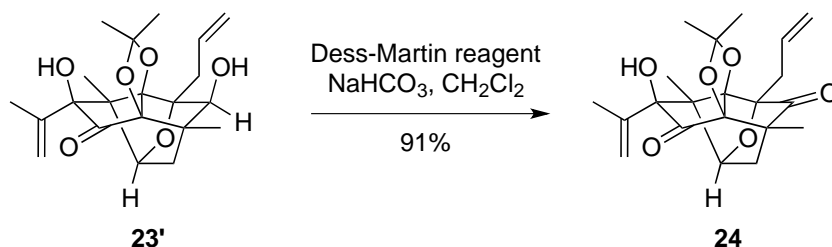
mL x3), and the combined organic layers were washed with brine (20 mL), dried over Na₂SO₄, filtered, and concentrated. The residue was purified by flash column chromatography on silica gel (3 g, hexane/EtOAc 10:1 to 5:1) to afford allyl alcohol **23** (35 mg, 71 μmol) in 48% yield over 3 steps: colorless solid; m.p. 148-149 °C; IR (neat) ν_{\max} 3376, 2934, 2858, 1750, 1249, 1110 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 0.10 (3H, s, CH₃ of TBS), 0.11 (3H, s, CH₃ of TBS), 0.93 (9H, s, *t*-Bu of TBS), 1.13 (3H, s, CH₃), 1.44 (3H, s, CH₃), 1.48 (3H, s, CH₃), 1.61 (3H, s, CH₃), 1.71 (1H, dd, *J* = 15.1, 3.2 Hz, CHOCH_AH_B), 1.78 (1H, dd, *J* = 15.1, 1.4 Hz, CHOCH_AH_B), 1.91 (3H, s, CH₃C), 2.48 (1H, br dd, *J* = 15.1, 7.4 Hz, CH_AH_BCH=CH₂), 2.51 (1H, s, OH), 2.62 (1H, br dd, *J* = 15.1, 7.3 Hz, CH_AH_BCH=CH₂), 3.87 (1H, s, CHOTBS), 3.88 (1H, dd, *J* = 3.2, 1.4 Hz, CHOCH₂), 5.09 (1H, s, CH₃C=CH_AH_B), 5.13 (1H, br s, CH₃C=CH_AH_B), 5.15 (1H, br d, *J* = 11.0 Hz, CH₂CH=CH_AH_B), 5.16 (1H, d, *J* = 16.9 Hz, CH₂CH=CH_AH_B), 5.95 (1H, dddd, *J* = 16.9, 11.0, 7.4, 7.3 Hz, CH₂CH=CH₂); ¹³C NMR (100 MHz, CDCl₃) δ -4.2, -3.8, 12.7, 14.7, 18.5, 20.4, 26.2, 27.8, 28.3, 34.7, 39.0, 50.6, 58.4, 79.8, 82.0, 89.0, 89.1, 98.8, 101.2, 117.9, 118.0, 118.5, 133.7, 140.6, 215.9; HRMS (ESI) calcd for C₂₇H₄₂O₆SiNa 513.2648 (M+Na⁺), found 513.2643.



Diene 23'. TBAF (1.0 M in THF, 1.7 mL, 1.7 mmol) was added to a solution of allyl alcohol **23** (55 mg, 110 μmol) in MeCN (5.6 mL) at room temperature. The reaction mixture was stirred for 2 h at 60 °C, and then cooled to room temperature. The mixture was filtered through a short pad of silica gel (3 g) with a mixture of hexane/EtOAc (1:1). The filtrate was concentrated, and the residue was purified by flash column chromatography on silica gel (3 g, hexane/EtOAc 3:1) to afford diene **23'** (33 mg, 88 μmol) in 80% yield: colorless solid; m.p. 203-204 °C; IR (neat) ν_{\max} 3443, 2930, 1741, 1456, 1385, 1204, 1085 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 1.21 (3H, s, CH₃), 1.47 (3H, s, CH₃), 1.53 (3H, s, CH₃), 1.64 (3H, s, CH₃), 1.76 (1H, dd, *J* = 15.1, 3.2 Hz, CHOCH_AH_B), 1.83 (1H, dd, *J* = 15.1, 1.4 Hz, CHOCH_AH_B), 1.91 (3H, s, CH₃C=CH₂), 2.39 (1H, br s, OH), 2.42 (1H, dd, *J* = 14.6, 7.8 Hz, CH_AH_BCH=CH₂), 2.78 (1H, br dd, *J* = 14.6, 6.4 Hz, CH_AH_BCH=CH₂), 3.16 (1H, d, *J* = 11.0 Hz, CHOH), 3.72 (1H, d, *J* = 11.0 Hz,

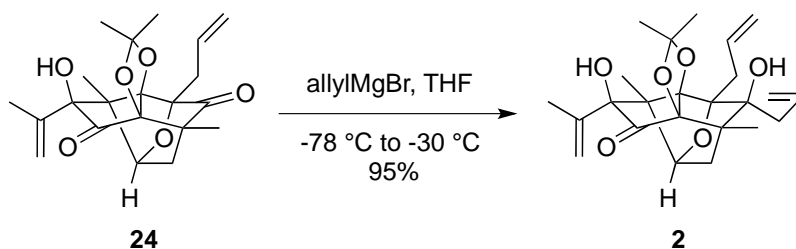
Supporting Information

CHOH), 3.91 (1H, dd, $J = 3.2, 1.4$ Hz, CHOCH₂), 5.10 (1H, br s, CH₃C=CH_AH_B), 5.15 (1H, br s, CH₃C=CH_AH_B), 5.16 (1H, br d, $J = 11.0$ Hz, CH₂CH=CH_AH_B), 5.22 (1H, dq, $J = 17.4, 1.4$ Hz, CH₂CH=CH_AH_B), 5.97 (1H, dddd, $J = 17.4, 11.0, 7.8, 6.4$ Hz, CH₂CH=CH₂); ¹³C NMR (100 MHz, CDCl₃) δ 12.6, 13.3, 20.2, 27.5, 28.1, 34.9, 38.6, 49.3, 58.0, 79.8, 81.8, 86.4, 88.8, 98.8, 100.9, 118.1, 118.2, 118.3, 133.3, 140.5, 214.0; HRMS (ESI) calcd for C₂₁H₂₈O₆Na 399.1784 (M+Na⁺), found 399.1769.

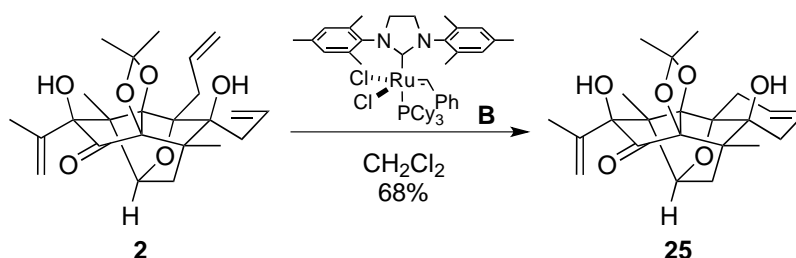


Diketone 24. Dess-Martin periodinane (190 mg, 448 μmol) was added to a suspension of **23'** (33 mg, 88 μmol) and NaHCO₃ (226 mg, 2.69 mmol) in CH₂Cl₂ (3.0 mL) at 0 °C. The reaction mixture was stirred for 12 h at 0 °C, and saturated aqueous Na₂S₂O₃ (4 mL) was added. The resultant mixture was extracted with Et₂O (7 mL x3), and the combined organic layers were washed with saturated aqueous Na₂CO₃ (10 mL x3) and brine (10 mL), dried over Na₂SO₄, filtered and concentrated. The residue was purified by flash column chromatography on silica gel (3 g, hexane/EtOAc 3:1) to afford diketone **24** (30 mg, 80 μmol) in 91% yield: colorless solid; m.p. 180-182 °C; IR (neat) ν_{\max} 3456, 2988, 1765, 1745, 1379, 1214 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 1.16 (3H, s, CH₃), 1.39 (3H, s, CH₃), 1.47 (3H, s, CH₃), 1.59 (3H, s, CH₃), 1.96 (3H, d, $J = 0.9$ Hz, CH₃C=CH₂), 2.14 (2H, d, $J = 2.3$ Hz, CHOCH₂), 2.42 (1H, dd, $J = 15.1, 7.3$ Hz, CH_AH_BCH=CH₂), 2.54 (1H, s, OH), 2.67 (1H, ddt, $J = 15.1, 7.3, 1.4$ Hz, CH_AH_BCH=CH₂), 4.05 (1H, t, $J = 2.3$ Hz, CHOCH₂), 5.11 (1H, br s, CH₃C=CH_AH_B), 5.16-5.24 (3H, m, CH₃C=CH_AH_B, CH₂CH=CH₂), 6.05 (1H, ddt, $J = 14.6, 10.1$ Hz, CH₂CH=CH₂); ¹³C NMR (100 MHz, CDCl₃) δ 11.2, 13.1, 20.2, 27.3, 28.9, 31.9, 43.1, 55.8, 56.7, 80.1, 80.5, 82.6, 91.2, 95.5, 118.3, 118.7, 118.8, 131.9, 139.9, 206.2, 214.0; HRMS (ESI) calcd for C₂₁H₂₆O₆Na 397.1627 (M+Na⁺), found 397.1608.

Supporting Information



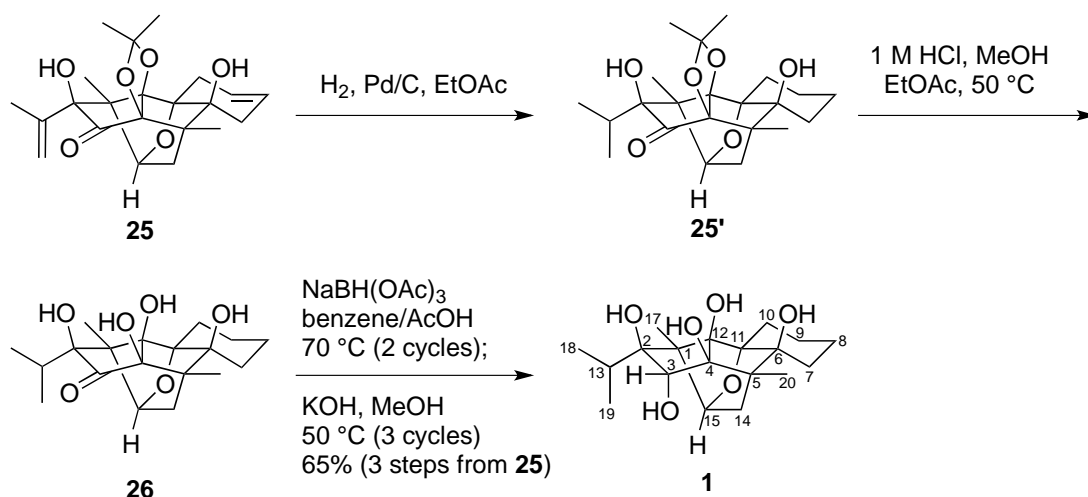
Triene 2. Allyl magnesium bromide (1.0 M in Et₂O, 1.6 mL, 1.6 mmol) was added to a solution of diketone **24** (30 mg, 80 μmol) in THF (4.0 mL) at -78 °C. The reaction mixture was allowed to warm to -30 °C, and stirred for 30 min. Saturated aqueous NH₄Cl (5 mL) was added at -30 °C. The resultant mixture was stirred at room temperature, and extracted with Et₂O (7 mL x3). The combined organic layers were washed with brine (10 mL), dried over Na₂SO₄, filtered and concentrated. The residue was purified by flash column chromatography on silica gel (3 g, hexane/EtOAc 4:1) to afford triene **2** (32 mg, 77 μmol) in 95% yield: colorless solid; m.p. 138-140 °C; IR (neat) ν_{max} 3490, 3075, 2958, 1751 1381, 1205, 1121 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 1.15 (3H, s, CH₃), 1.47 (3H, s, CH₃), 1.50 (3H, s, CH₃), 1.62 (1H, d, *J* = 15.6 Hz, CHOCH_AH_B), 1.65 (3H, s, CH₃), 1.92 (3H, s, CH₃C=CH₂), 2.00 (1H, dd, *J* = 15.6, 3.6 Hz, CHOCH_AH_B), 2.37 (1H, s, OH), 2.56-2.66 (3H, m, CH_AH_BCH=CH₂ x 2, CH_AH_B-CH=CH₂), 2.71 (1H, br dd, *J* = 15.6, 6.4 Hz, CH_AH_BCH=CH₂), 3.92 (1H, d, *J* = 3.6 Hz, CHOCH₂), 4.23 (1H, br s, OH), 5.02-5.19 (6H, m, CH₃C=CH₂, CH₂CH=CH₂ x 2), 5.93-6.09 (2H, m, CH₂CH=CH₂ x 2); ¹³C NMR (100 MHz, CDCl₃) δ 12.5, 12.8, 20.4, 27.6, 27.9, 33.1, 35.4, 36.6, 51.5, 57.3, 79.6, 81.3, 86.0, 90.1, 98.5, 100.6, 116.6, 117.9, 118.3, 118.4, 134.1, 134.6, 140.4, 214.3; HRMS (ESI) calcd for C₂₄H₃₂O₆Na 439.2097 (M+Na⁺), found 439.2074.



Compound 25. A mixture of triene **2** (32 mg, 77 μmol) and Grubbs' 2nd catalyst **B** (32 mg, 38 μmol) in CH₂Cl₂ (25 mL) was stirred for 2.5 h at room temperature. After additional Grubbs' 2nd catalyst (6.5 mg, 7.6 μmol) was added, the reaction was stirred for further 1 h at same temperature. The mixture was filtered through a short pad of silica gel (1 g, hexane/EtOAc 1:1), and the filtrate was concentrated.

Supporting Information

The residue was purified by flash column chromatography on silica gel (3 g, hexane/EtOAc 5:1 to 2:1) to afford **25** (20 mg, 52 μmol) in 68% yield: colorless solid; m.p. 243-245 $^{\circ}\text{C}$; IR (neat) ν_{max} 3462, 3370, 3027, 2995, 1750, 1459, 1383, 1217 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3) δ 1.16 (3H, s, CH_3), 1.49 (3H, s, CH_3), 1.53 (3H, s, CH_3), 1.66 (3H, s, CH_3), 1.69 (1H, d, $J = 15.6$ Hz, CHOCH_AH_B), 1.91 (3H, s, $\text{CH}_3\text{C}=\text{CH}_2$), 1.96 (1H, br d, $J = 17.8$ Hz, $\text{CCH}=\text{CHCH}_A\text{H}_B$), 1.99 (1H, dd, $J = 15.6, 3.6$ Hz, CHOCH_AH_B), 2.19 (1H, br d, $J = 16.5$ Hz, $\text{CH}_A\text{H}_B\text{CH}=\text{CHC}$), 2.44 (1H, br s, OH), 2.63 (1H, br d, $J = 16.5$ Hz, $\text{CH}_A\text{H}_B\text{CH}=\text{CHC}$), 2.79 (1H, br d, $J = 17.8$ Hz, $\text{CCH}=\text{CHCH}_A\text{H}_B$), 3.85 (1H, d, $J = 3.6$ Hz, CHOCH_2), 4.40 (1H, d, $J = 2.7$ Hz, OH), 5.13 (1H, br s, $\text{CH}_3\text{C}=\text{CH}_A\text{H}_B$), 5.15 (1H, br s, $\text{CH}_3\text{C}=\text{CH}_A\text{H}_B$), 5.68-5.78 (2H, m, $\text{CH}=\text{CH}$); ^{13}C NMR (100 MHz, CDCl_3) δ 11.4, 12.6, 20.2, 27.6, 28.1, 29.4, 31.6, 36.2, 51.3, 57.9, 78.9, 81.3, 82.5, 86.4, 98.7, 100.5, 118.2, 119.0, 123.9 (two peaks), 140.3, 214.5; HRMS (ESI) calcd for $\text{C}_{22}\text{H}_{28}\text{O}_6\text{Na}$ 411.1784 ($\text{M}+\text{Na}^+$), found 411.1777.



Compound 25'. A suspension of **25** (14 mg, 36 μmol) and 5% Pd/C (29 mg) in EtOAc (3.7 mL) was exposed to H_2 atmosphere (1 atm). Then, the reaction mixture was stirred under H_2 atmosphere for 11 h at room temperature. The resultant mixture was filtered through a pad of Celite with EtOAc. The filtrate was concentrated to afford crude **25'** (14 mg), which was used for the next reaction without further purification.

Tetraol 26. HCl solution (1.0 M in EtOAc, 2.5 mL) was added to a solution of the above crude **25'** (14 mg) in MeOH (1.2 mL) at room temperature. The reaction mixture was stirred for 2.5 h at 50 $^{\circ}\text{C}$, cooled to room temperature and then quenched with solid NaHCO_3 (ca. 500 mg). The resultant mixture was filtered through a short pad of silica gel (3 g, EtOAc/MeOH 10:1). The filtrate

Supporting Information

was concentrated to afford crude tetraol **26** (14 mg), which was used for the next reaction without further purification.

9-demethyl-10,15-dideoxyryanodol (1). NaBH(OAc)₃ (157 mg, 741 μmol) was added to a solution of the above crude tetraol **26** (14 mg) in a mixture of benzene (3.4 mL) and AcOH (0.34 mL). The reaction mixture was stirred for 3 h at 70 °C, and cooled to room temperature. MeOH (5 mL) was added, and the resultant mixture was concentrated to remove trimethyl borate azeotropically (x3). The resultant residue was filtered through a short pad silica gel (1 g, EtOAc/MeOH 10:1) and the filtrate was concentrated. The ESI-MS analysis indicated that the crude material contained a mixture of tetraol **26** and the borate of **1**. Thus, the crude mixture was again subjected to the reduction with NaBH(OAc)₃ (79 mg, 373 μmol) in a mixture of benzene (3.4 mL) and AcOH (0.34 mL) for 11 h at 70 °C. The same work up procedure of the reaction mixture as described above afforded the borate of **1**. A mixture of KOH (42 mg, 750 μmol) and the crude borate of **1** in a mixture of EtOH (1.9 mL) and H₂O (0.2 mL) was stirred for 21 h at 70 °C, and cooled to room temperature. The reaction mixture was neutralized with AcOH (0.1 mL) and concentrated. The resultant mixture was filtered through a short pad of silica gel (1 g, EtOAc/MeOH 10:1), and the filtrate was concentrated to afford a mixture of **1** and the borate. The hydrolysis procedure was repeated twice to completely convert the borate to **1**. The crude **1** was purified by flash column chromatography on silica gel (1 g, CHCl₃/MeOH 20:1 to 10:1) by using a vial made of soda-lime glassware and a flask made of quartz glassware for fraction collection to afford 9-demethyl-10,15-dideoxyryanodol (**1**) (8.5 mg, 24 μmol) in 65% yield over 3 steps: colorless solid; m.p. 220-221 °C; IR (neat) ν_{\max} 3408, 2927, 2858, 1405, 1384, 1045 cm⁻¹; ¹H NMR (400 MHz, CD₃OD) δ 0.96 (3H, d, *J* = 6.8 Hz, H18), 0.99 (3H, d, *J* = 6.4 Hz, H19), 1.10 (3H, s, H20), 1.22 (3H, s, H17), 1.27-1.37 (1H, m, H7eq), 1.37-1.43 (1H, m, H10eq), 1.43-1.55 (2H, m, H8eq and H9eq), 1.59-1.75 (2H, m, H8ax and H9ax), 1.72 (1H, dd, *J* = 14.2, 3.7 Hz, H14a), 1.98 (1H, td, *J* = 13.2, 5.5 Hz, H10ax), 2.06 (1H, td, *J* = 12.8, 5.0 Hz, H7ax), 2.12 (1H, qq, *J* = 6.8, 6.4 Hz, H13), 2.24 (1H, dd, *J* = 14.2, 1.4 Hz, H14b), 3.80 (1H, dd, *J* = 3.7, 1.4 Hz, H15), 4.16 (1H, s, H3); ¹³C NMR (100 MHz, CD₃OD) δ 13.2, 15.7, 18.9, 19.7, 20.4, 21.7, 26.5, 27.4, 30.7, 36.5, 51.2, 65.0, 80.5, 84.3, 84.7, 87.9, 92.1, 92.7, 95.6; HRMS (ESI) calcd for C₁₉H₃₀O₆Na 377.1940 (M+Na⁺), found 377.1938.

Supporting Information

Toxicity to Houseflies^{S1}

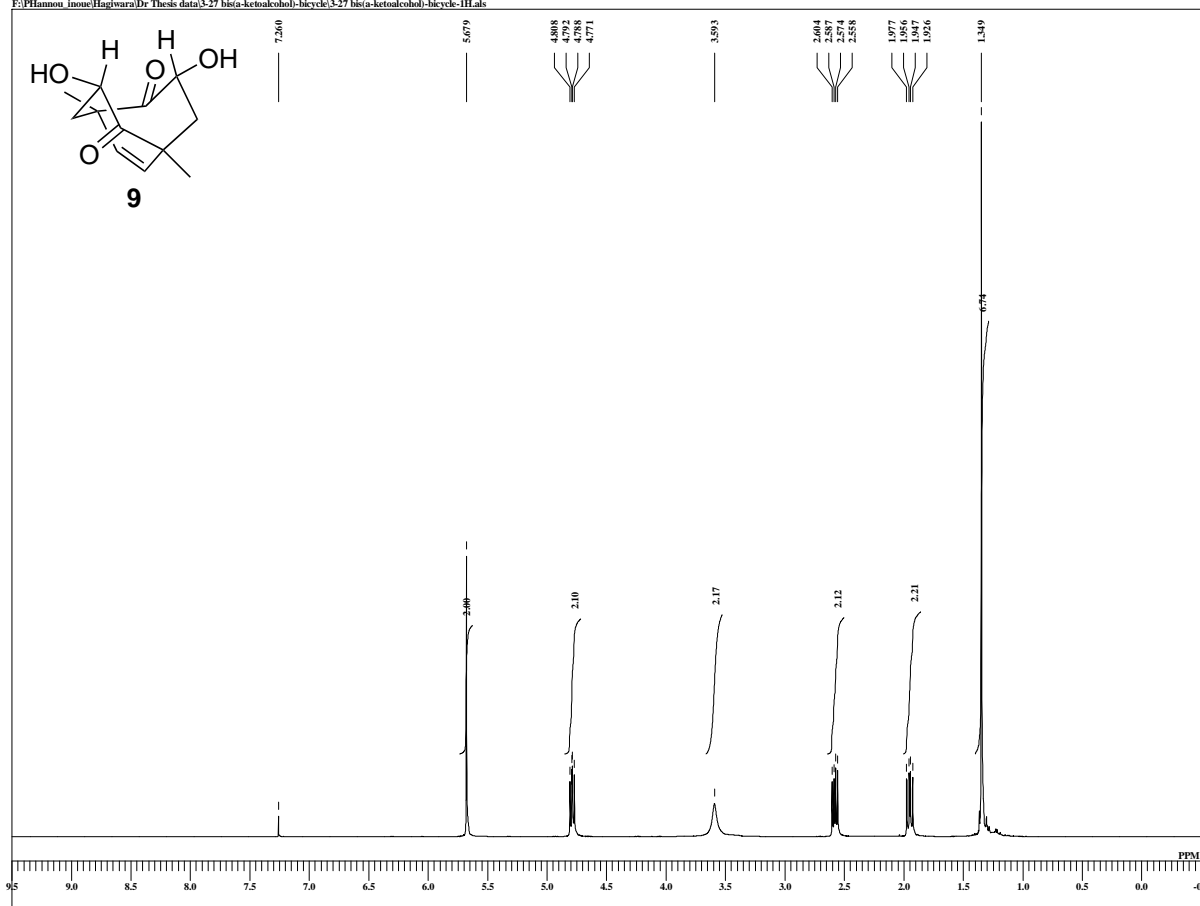
Ryanodine was purchased from Wako Pure Chemical Industries (Japan), and ryanodol was prepared according to the literature procedure.²

A filter paper of a diameter of 5.5 cm was spread on the bottom of a polyethylene cup of a diameter of 5.5 cm. A solution of varied amounts of ryanodine, ryanodol or 9-demethyl-10,15-dideoxyryanodol (**1**) in acetone (X ppm, 0.7 ml) was added dropwise onto the filter paper, and the filter paper was dried. Sucrose (30 mg) in water (0.7 mL) was applied to the filter paper as a bait. Then, five female imagoes of house flies (*Musca domestica*) were released into the polyethylene cup, and the cup was sealed with a lid. After 2 days, the number of surviving house flies was counted, and the death rate of the house flies was calculated. LD₁₀₀ of ryanodine, ryanodol and **1** were determined to be 15.6 ppm, 200 ppm and >200 ppm, respectively.

(S1) Toxicity of ryanodine and ryanodol to house flies has been known. See for examples: (a) Waterhouse, A. L.; Pessah, I. N.; Francini, A. O.; Casida, J. E. *J. Med. Chem.* **1987**, *30*, 710-716. (b) Jefferies, P. R.; Toia, R. F.; Brannigan, B.; Pessah, I.; Casida, J. E. *J. Agric. Food. Chem.* **1992**, *40*, 142-146. (c) Jefferies, P. R.; Yu, P.; Casida, J. E. *Pestic. Sci.* **1997**, *51*, 33-38.
(S2) Kelly, R. B.; Whittingham, D. J.; Wiesner, K. *Can. J. Chem.* **1951**, *29*, 905-910.

single_pulse

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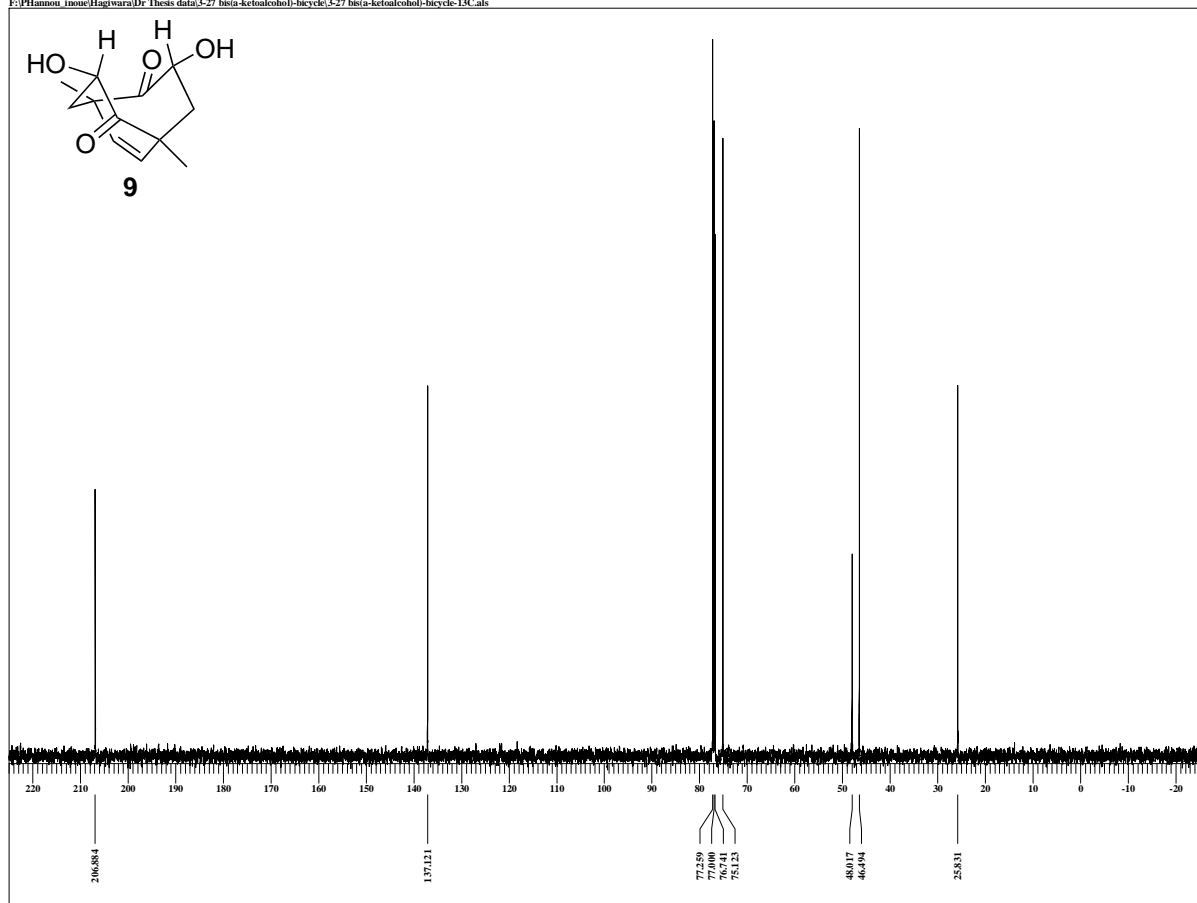


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OBFN 9.64 Hz
PW1 6.20 usec
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PREDL 0.00000 msec
IWT 1.0000 sec
POINT 13107
SPO 13107
TIMES 8
DUMMY 1
FREQU 7429.31 Hz
FLT 38000 Hz
DELAY 13.16 usec
ACQTM 1.7642 sec
PD 1.5000 sec
SCANS 8
ADBIT 16
RGAIN 38
BF 0.01 Hz
T1 0.00
T2 0.00
T3 90.00
T4 100.00
EXMOD single_pulse.ex2
EXPCM
IRNUC 1H
IFR 495.13 MHz
IRSET 4.38 KHz
IRFN 9.64 Hz
IRFPW 115 usec
IRATN 79
DFILE 3-27 bis(a-ketoalcohol)-bi
SF -601.50 KHz
LKSET -1.8 Hz
LKFN -1.8 Hz
LKLEV 0
LGAIN 0
LKPHS 0
LKSG 0
CSPED 0 Hz
FLDC
FLDF
CTEMP 19.5 c
SLVNT CDCL3
XREF 7.26 ppm
    
```

single pulse decoupled gated NOE

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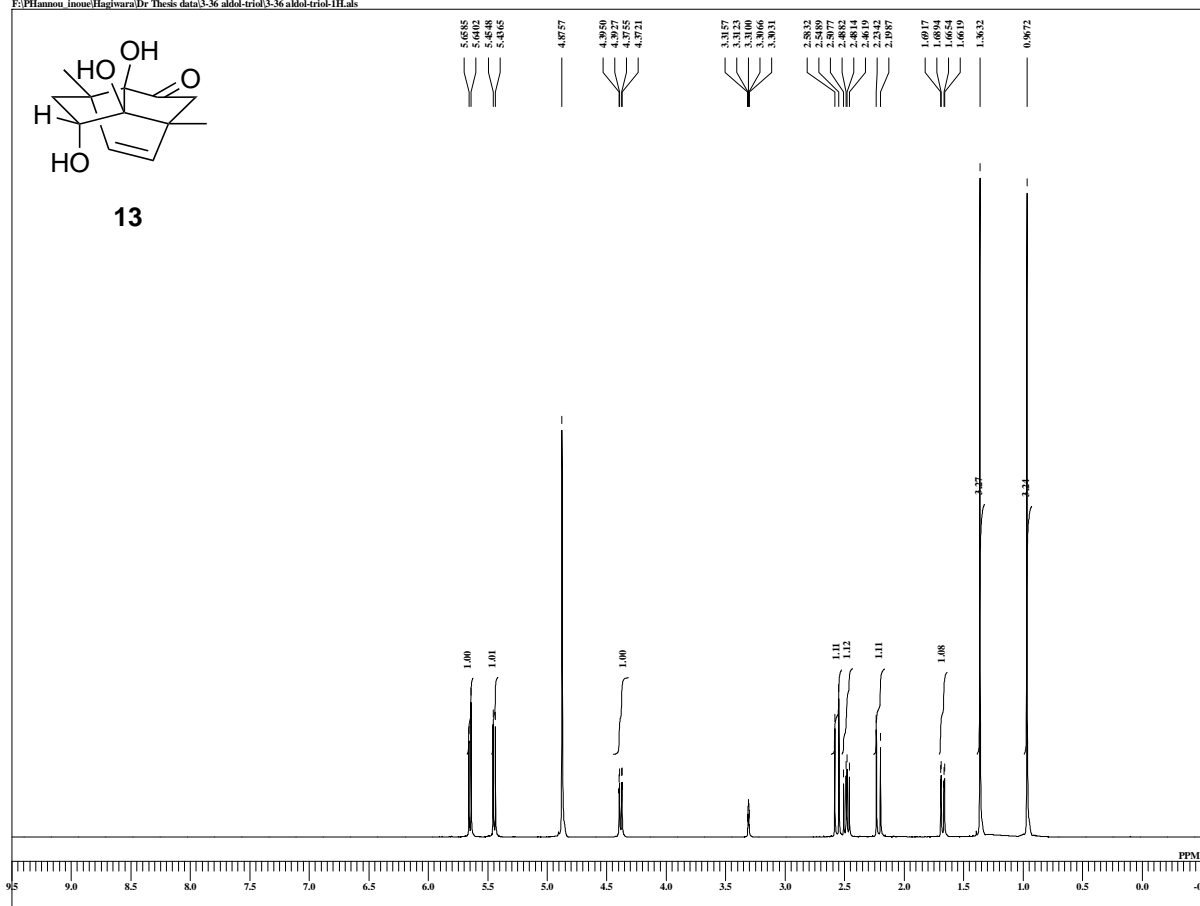


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OBFN 6.00 Hz
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PREDL 0.00000 msec
IWT 1.0000 sec
POINT 26214
SPO 26214
TIMES 8
DUMMY 4
FREQU 31249.52 Hz
FLT 157000 Hz
DELAY 20.80 usec
ACQTM 0.8389 sec
PD 5.0000 sec
SCANS 8
ADBIT 16
RGAIN 60
BF 0.01 Hz
T1 0.00
T2 0.00
T3 90.00
T4 100.00
EXMOD single_pulse_dec
EXPCM
IRNUC 1H
IFR 495.13 MHz
IRSET 4.38 KHz
IRFN 9.64 Hz
IRFPW 50 usec
IRATN 79
DFILE 3-27 bis(a-ketoalcohol)-bi
SF -601.50 KHz
LKSET -1.8 Hz
LKFN -1.8 Hz
LKLEV 0
LGAIN 0
LKPHS 0
LKSG 0
CSPED 0 Hz
FLDC
FLDF
CTEMP 20.5 c
SLVNT CDCL3
XREF 77.00 ppm
    
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single_pulse

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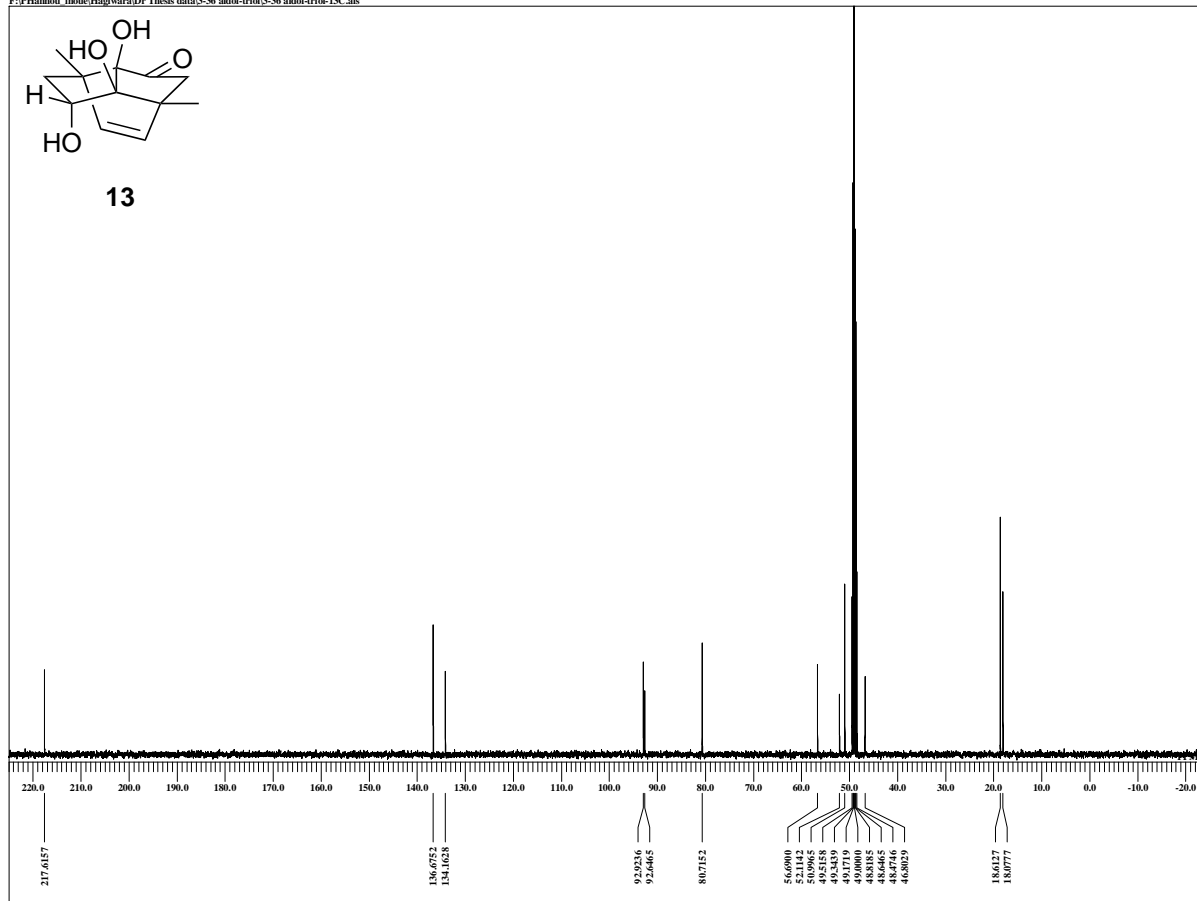


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OBFEN 7.60 Hz
PW1 8.55 usec
DEADT 0.00 usec
PREDL 0.00000 msec
IWT 1.0000 sec
POINT 13107
SPO 13107
TIMES 8
DUMMY 1
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FLT 37000 Hz
DELAY 13.52 usec
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PD 2.0000 sec
SCANS 8
ADBIT 16
RGAIN 30
BF 0.01 Hz
T1 0.00
T2 0.00
T3 90.00
T4 100.00
EXMOD single_pulse.ex2
EXPCM
IRNUC 1H
IFR 490.15 MHz
IRSET 9.16 KHz
IRFEN 7.60 Hz
IRFPW 118 usec
IRATN 79
DEFILE 3-36 aldol-triol-1H.tals
SF 70.00 KHz
LKSET 70.00 KHz
LKFN 36.6 Hz
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LGAIN 0
LKPHS 0
LKSG 0
CSPED 0 Hz
FLDC
FLDF
CTEMP 20.2 c
SLVNT CD3OD
XREF 3.31 ppm
    
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single_pulse decoupled gated NOE

F:\Phamou inoue\Hagwara\Dr Thesis data\3-36 aldol-triol\3-36 aldol-triol-13C.tals

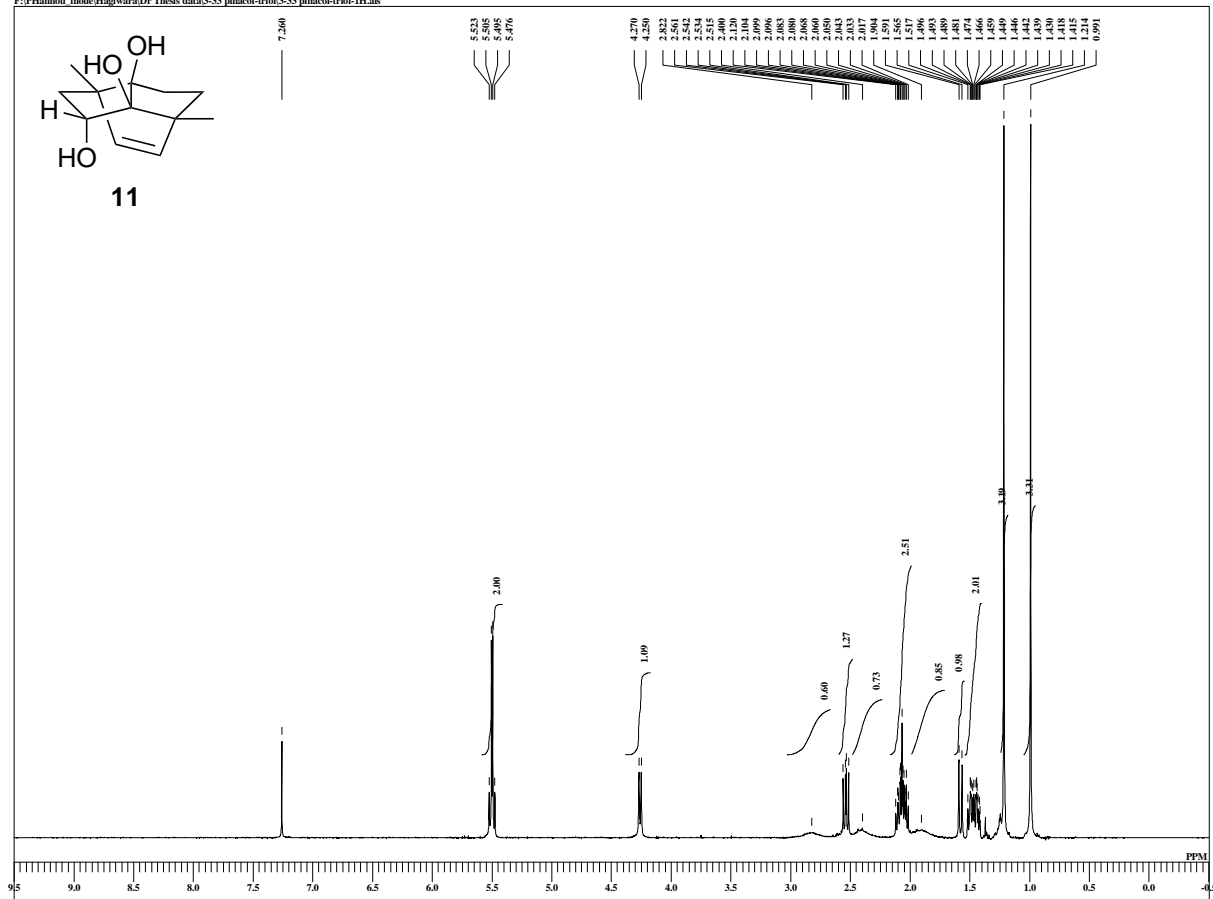


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OBSET 2.31 KHz
OBFEN 6.71 Hz
PW1 3.20 usec
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PREDL 0.00000 msec
IWT 1.0000 sec
POINT 26214
SPO 26214
TIMES 8
DUMMY 4
FREQU 30983.73 Hz
FLT 155000 Hz
DELAY 21.06 usec
ACQTM 0.8493 sec
PD 5.0000 sec
SCANS 8
ADBIT 16
RGAIN 60
BF 0.01 Hz
T1 0.00
T2 0.00
T3 90.00
T4 100.00
EXMOD single_pulse_dec
EXPCM
IRNUC 13C
IFR 125.26 MHz
IRSET 2.31 KHz
IRFEN 6.71 Hz
IRFPW 92 usec
IRATN 79
DEFILE 3-36 aldol-triol-13C.tals
SF 70.00 KHz
LKSET 70.00 KHz
LKFN 36.6 Hz
LKLEV 0
LGAIN 0
LKPHS 0
LKSG 0
CSPED 0 Hz
FLDC
FLDF
CTEMP 20.6 c
SLVNT CD3OD
XREF 49.00 ppm
    
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single_pulse

F:\Phamou inoue\Hagihara\Dr Thesis data\3-33 pinacol-triol\3-33 pinacol-triol-1H.cab

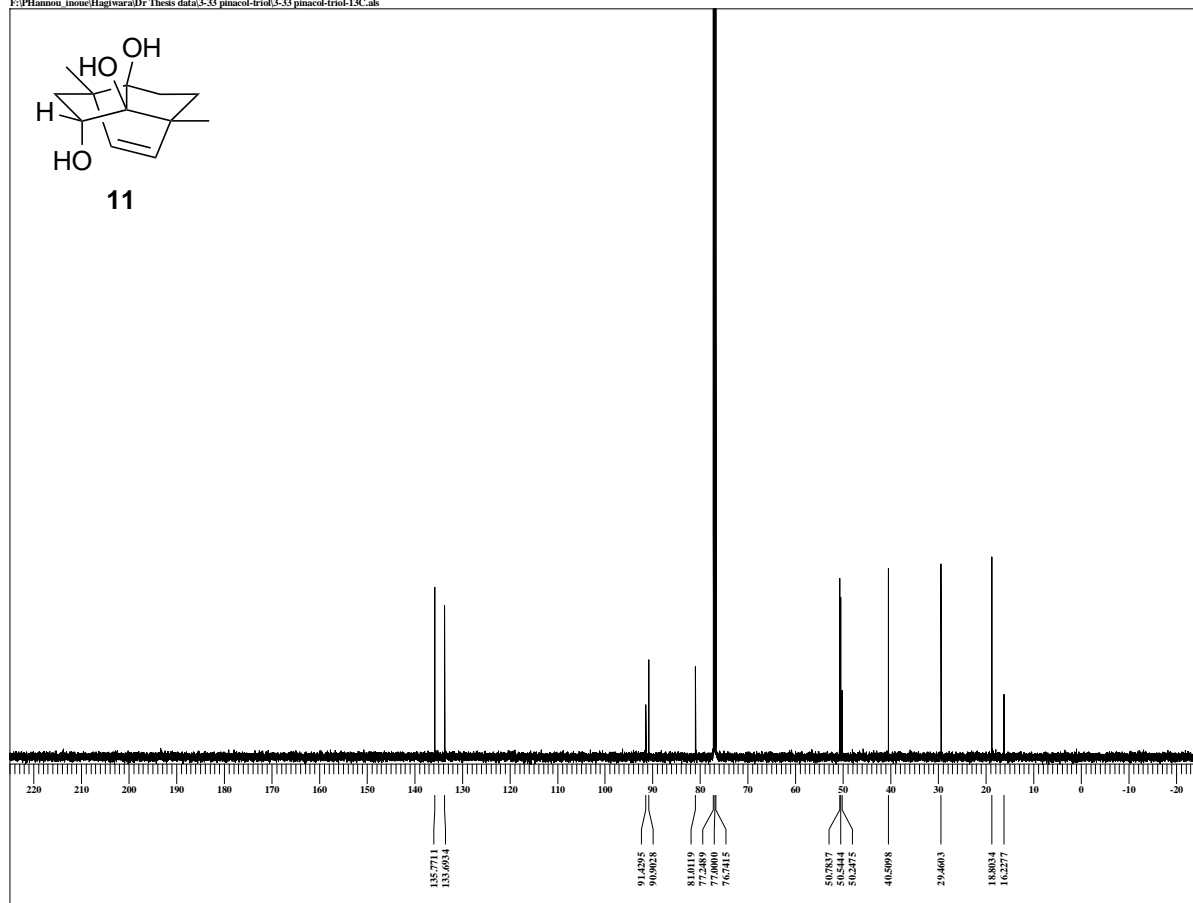


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DATIM 10-11-2008 22:10:40
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OBSET 4.38 KHz
OBFN 9.64 Hz
PWI 6.20 usec
DEADT 0.00 usec
PREDL 0.00000 msec
IWT 1.0000 sec
POINT 13107
SFO 13107
TIMES 8
DUMMY 1
FREQU 7429.31 Hz
FLT 38000 Hz
DELAY 13.16 usec
ACQTM 1.7642 sec
PD 1.5000 sec
SCANS 8
ADBIT 16
RGAIN 50
BF 0.01 Hz
T1 0.00
T2 0.00
T3 100.00
T4 100.00
EXMOD single_pulse.ex2
EXPCM
IRNUC 1H
IR 495.13 MHz
IRSET 4.38 KHz
IRFN 9.64 Hz
IRPW 50 usec
IRATN 79
DFILE 3-33 pinacol-triol-1H.cab
SF -601.50 KHz
LKFN -1.8 Hz
LKLEV 0
LGAIN 0
LKPS 0
LKSG 0
CSPED 0 Hz
FILDC
FILDF
CTEMP 19.3 c
SLVNT CDCL3
EXREF 7.26 ppm
    
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single_pulse decoupled gated NOE

F:\Phamou inoue\Hagihara\Dr Thesis data\3-33 pinacol-triol\3-33 pinacol-triol-13C.cab

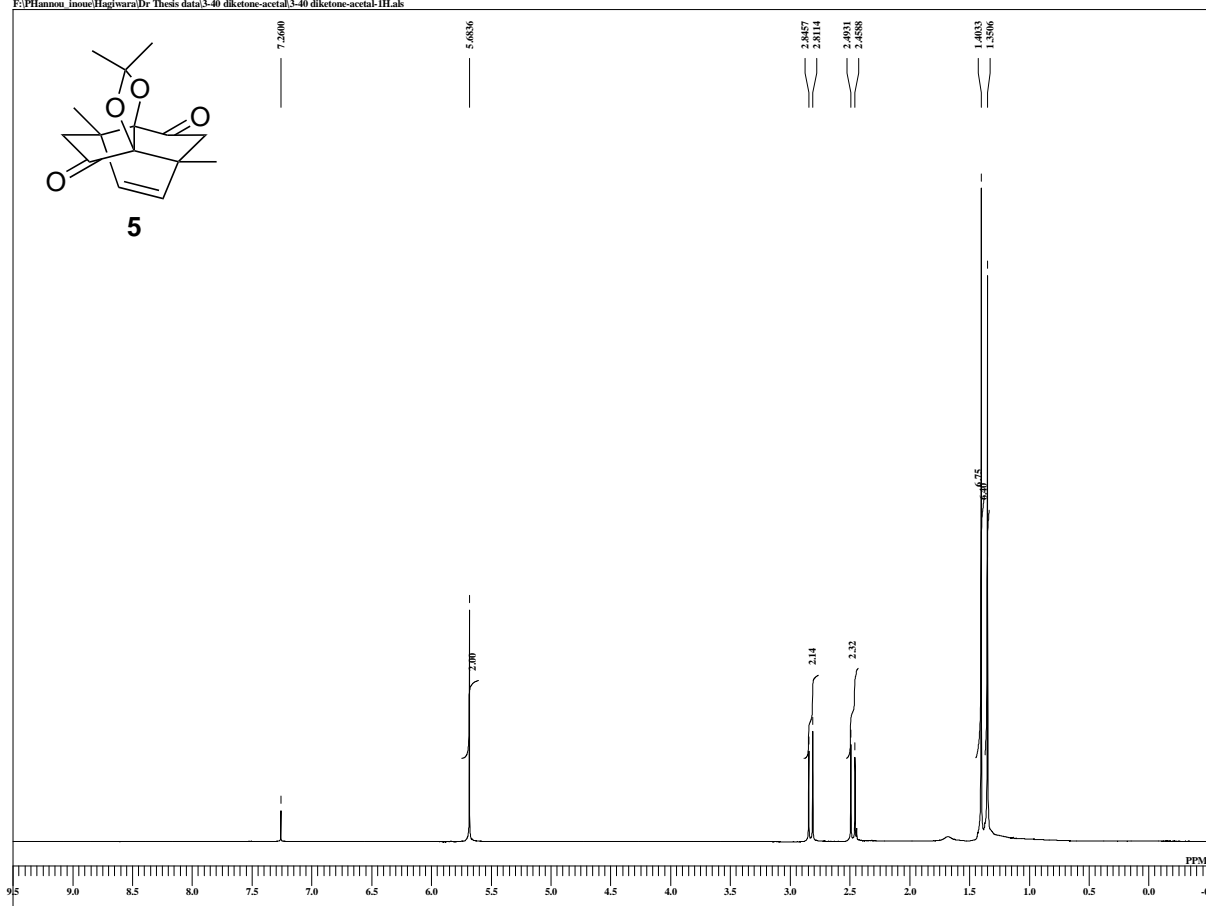


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OBSET 3.45 KHz
OBFN 6.00 Hz
PWI 3.57 usec
DEADT 0.00 usec
PREDL 0.00000 msec
IWT 1.0000 sec
POINT 26214
SFO 26214
TIMES 2056
DUMMY 4
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FLT 157000 Hz
DELAY 20.80 usec
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ADBIT 16
RGAIN 60
BF 0.01 Hz
T1 0.00
T2 0.00
T3 100.00
T4 100.00
EXMOD single_pulse_dec
EXPCM
IRNUC 1H
IR 495.13 MHz
IRSET 4.38 KHz
IRFN 9.64 Hz
IRPW 50 usec
IRATN 79
DFILE 3-33 pinacol-triol-13C.cab
SF -601.50 KHz
LKFN -1.8 Hz
LKLEV 0
LGAIN 0
LKPS 0
LKSG 0
CSPED 0 Hz
FILDC
FILDF
CTEMP 21.0 c
SLVNT CDCL3
EXREF 77.00 ppm
    
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single pulse

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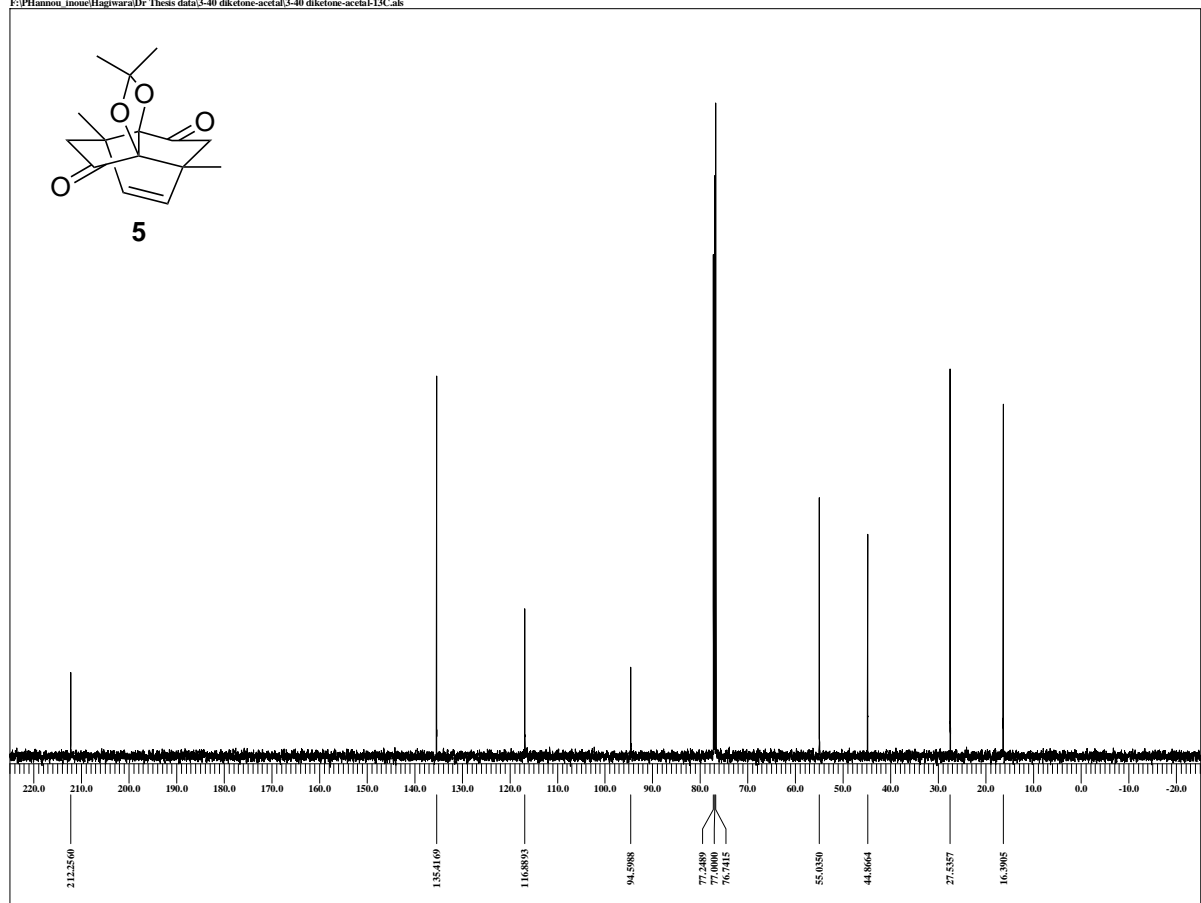


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OFRBQ 495.13 MHz
OBSET 4.38 KHz
OBFEN 9.64 Hz
PW1 6.20 usec
DEADT 0.00 msec
PREDL 0.00000 msec
IWT 1.0000 sec
POINT 13107
SFO 13107
TIMES 8
DUMMY 1
FREQU 7429.31 Hz
FLT 38000 Hz
DELAY 13.16 usec
ACQTM 1.7642 sec
PD 1.5000 sec
SCANS 8
ADBIT 16
RGAIN 46
BF 0.01 Hz
T1 0.00
T2 0.00
T3 90.00
T4 100.00
EXMOD single_pulse.ec2
EXPCM
IRNUC 1H
IFR 495.13 MHz
IRSET 4.38 KHz
IRFEN 9.64 Hz
IRRPW 50 usec
IRATN 79
DFILE 3-40 diketone-acetal-1H.
SF
LKSET -601.50 KHz
LKFN -1.8 Hz
LKLEV 0
LGAIN 0
LKPHS 0
LKSG 0
CSPED 0 Hz
FILDC
FILDF
CTEMP 19.7 c
SLVNT CDCL3
EXREF 7.26 ppm
    
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single pulse decoupled gated NOE

F:\Phannou_inoue\Hagiwara\Dr_Thesis\data\3-40 diketone-acetal\3-40 diketone-acetal-13C.ac

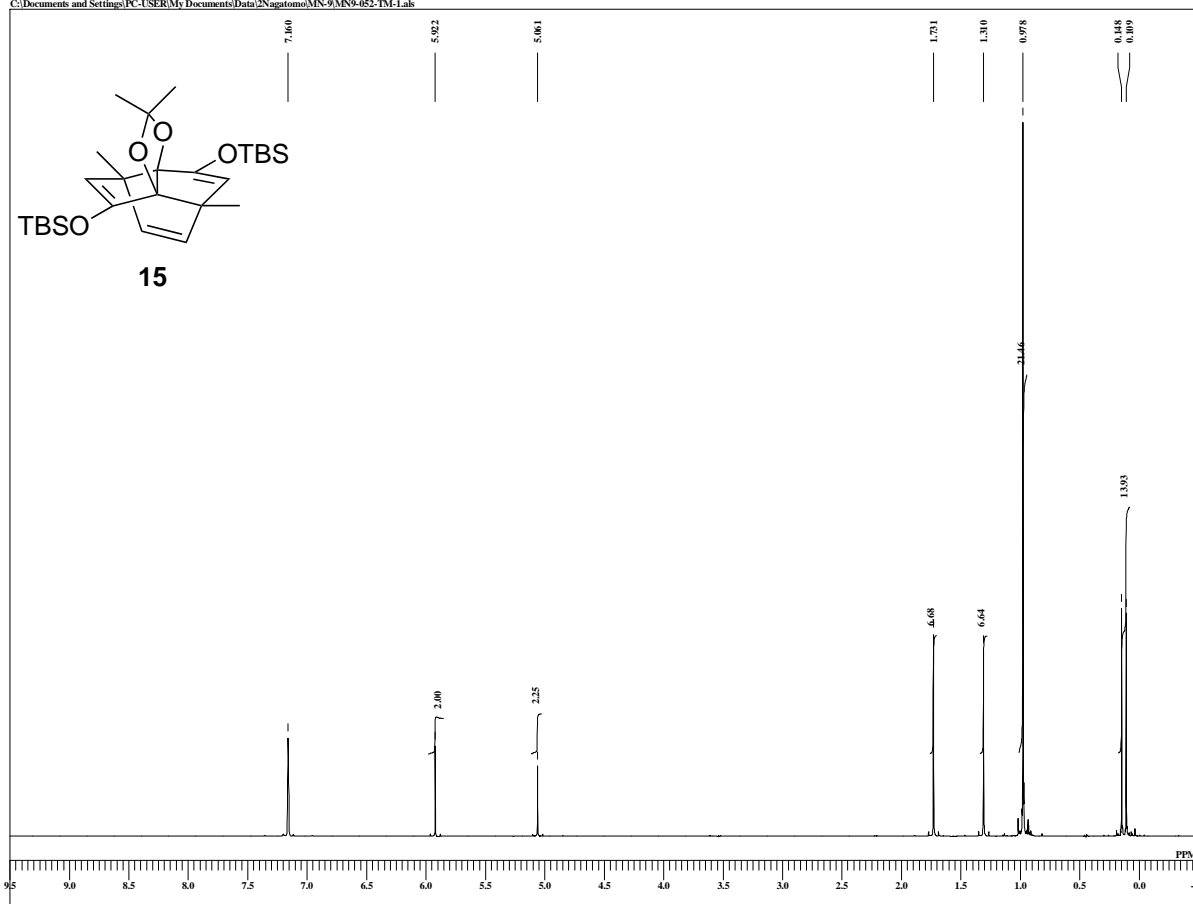


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OBFEN 6.00 Hz
PW1 3.57 usec
DEADT 0.00 msec
PREDL 0.00000 msec
IWT 1.0000 sec
POINT 26214
SFO 26214
TIMES 32
DUMMY 4
FREQU 31249.53 Hz
FLT 157000 Hz
DELAY 20.80 usec
ACQTM 0.8389 sec
PD 5.0000 sec
SCANS 32
ADBIT 16
RGAIN 60
BF 0.01 Hz
T1 0.00
T2 0.00
T3 90.00
T4 100.00
EXMOD single_pulse_dec
EXPCM
IRNUC 1H
IFR 495.13 MHz
IRSET 4.38 KHz
IRFEN 9.64 Hz
IRRPW 50 usec
IRATN 79
DFILE 3-40 diketone-acetal-13C.
SF
LKSET -601.50 KHz
LKFN -1.8 Hz
LKLEV 0
LGAIN 0
LKPHS 0
LKSG 0
CSPED 0 Hz
FILDC
FILDF
CTEMP 21.0 c
SLVNT CDCL3
EXREF 77.00 ppm
    
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MN9-052-TM

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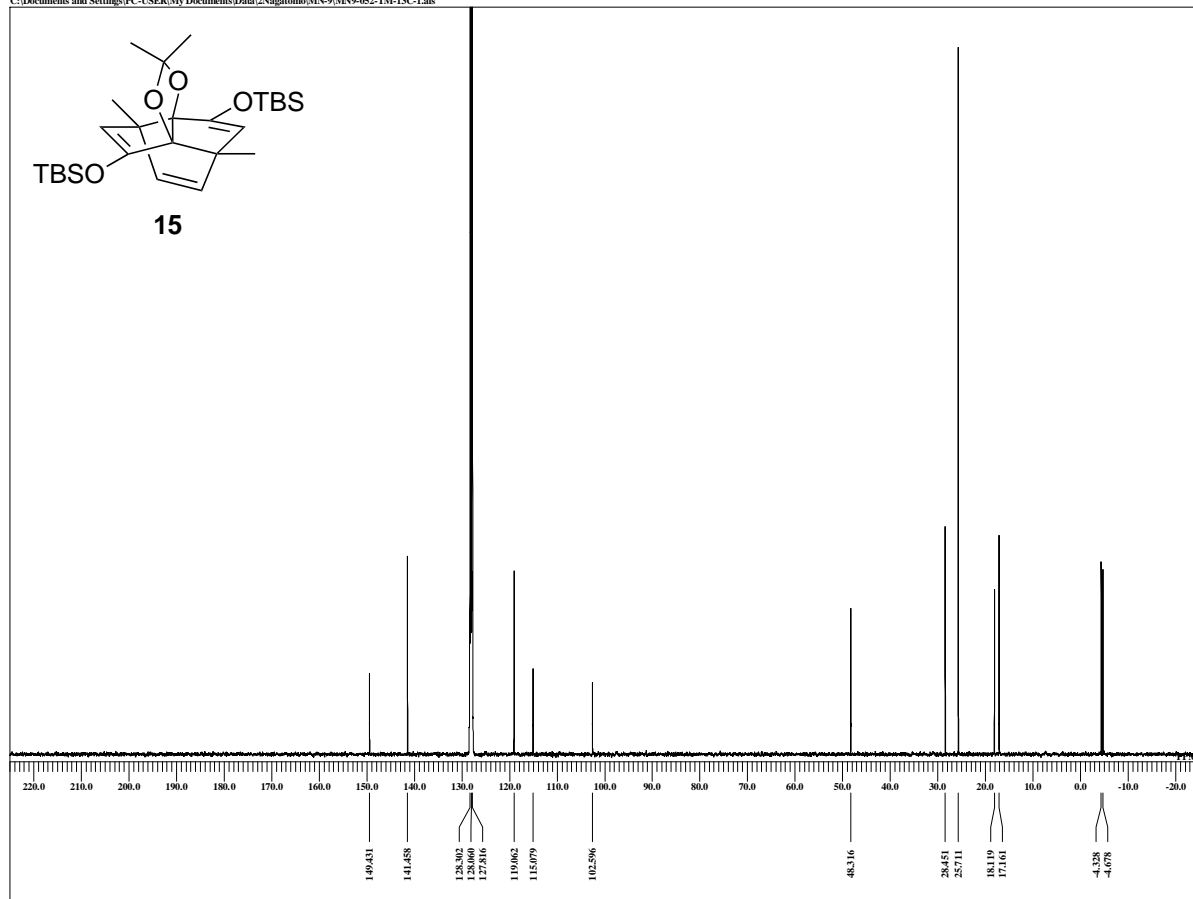


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OBFN 0.87 Hz
PW1 6.50 usec
DEADT 0.00 usec
PREDL 0.00000 msec
IWT 1.0000 sec
POINT 13107
SFO 13107
TIMES 16
DUMMY 1
FREQU 5938.15 Hz
FLT 38000 Hz
DELAY 16.68 usec
ACQTM 2.2073 sec
PD 2.0000 sec
SCANS 16
ADBIT 16
RGAIN 36
BF 0.01 Hz
T1 0.00
T2 0.00
T3 100.00
T4 100.00
EXMOD single_pulse.ex2
EXPCM
IRNUC 1H
OFR 395.88 MHz
IRSET 6.28 KHz
IRFN 0.87 Hz
IRFPW 115 usec
IRATN 79
DFILE MN9-052-TM-1.a
SF 13.20 KHz
LKSET 13.20 KHz
LKFN 69.6 Hz
LKLEV 0
LGAIN 0
LKPS 0
LKSG 0
CSPED 0 Hz
FLDC
FLDF 23.9 c
SLVNT C6D6
EXREF 7.16 ppm
    
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MN9-052-TM

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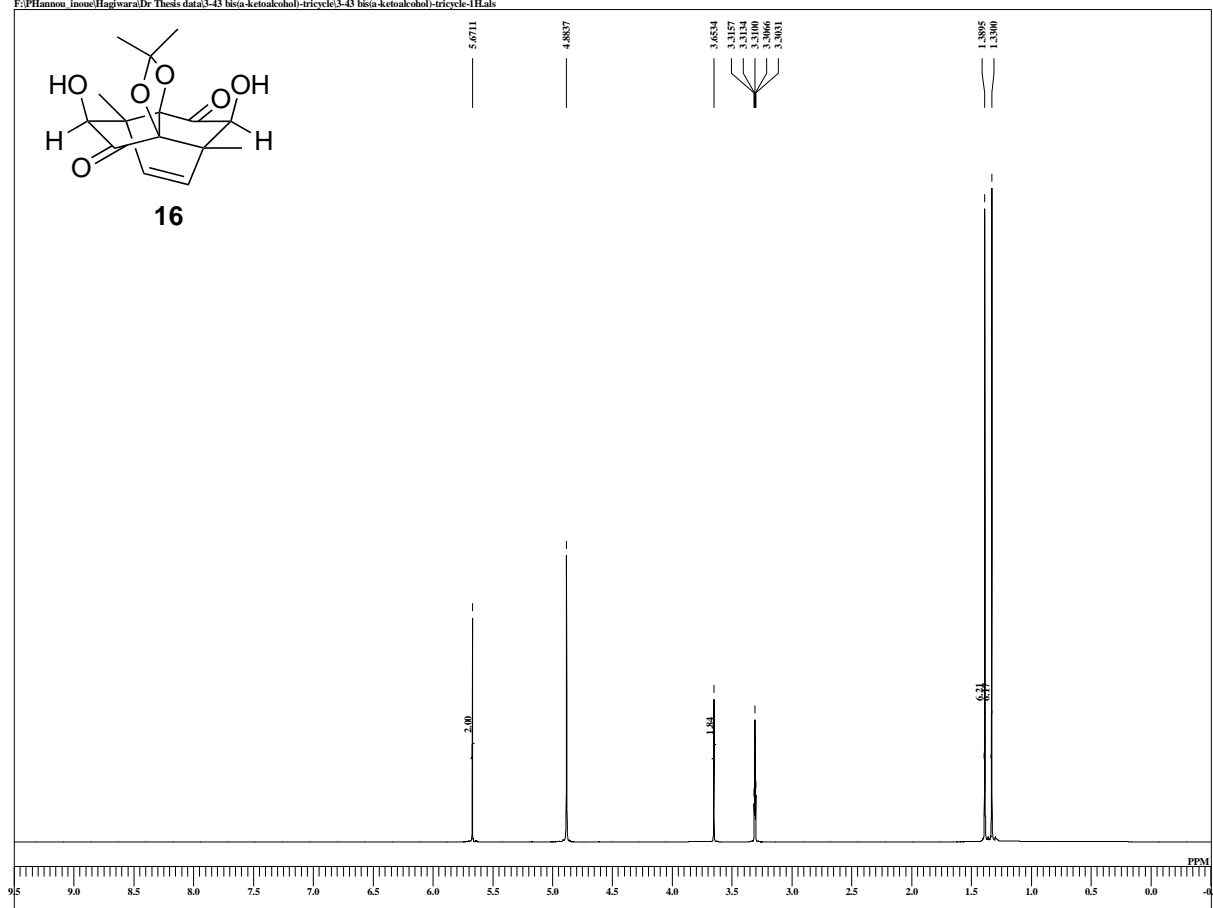


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OBSET 5.13 KHz
OBFN 0.98 Hz
PW1 3.33 usec
DEADT 0.00 usec
PREDL 0.00000 msec
IWT 1.0000 sec
POINT 104856
SFO 104856
TIMES 200
DUMMY 4
FREQU 24999.62 Hz
FLT 125000 Hz
DELAY 20.50 usec
ACQTM 1.0486 sec
PD 2.0000 sec
SCANS 200
ADBIT 16
RGAIN 36
BF 1.00 Hz
T1 0.00
T2 0.00
T3 100.00
T4 100.00
EXMOD single_pulse_dec
EXPCM
IRNUC 1H
OFR 395.88 MHz
IRSET 6.28 KHz
IRFN 0.87 Hz
IRFPW 115 usec
IRATN 79
DFILE MN9-052-TM-13C-1.a
SF 13.20 KHz
LKSET 13.20 KHz
LKFN 69.6 Hz
LKLEV 0
LGAIN 0
LKPS 0
LKSG 0
CSPED 0 Hz
FLDC
FLDF 24.3 c
SLVNT C6D6
EXREF 128.06 ppm
    
```

single_pulse

F:\Phamou Inoue\Hagiwara\Dr Thesis\data\3-43 bis(a-ketoalcohol)-tricycle\3-43 bis(a-ketoalcohol)-tricycle-1Hals

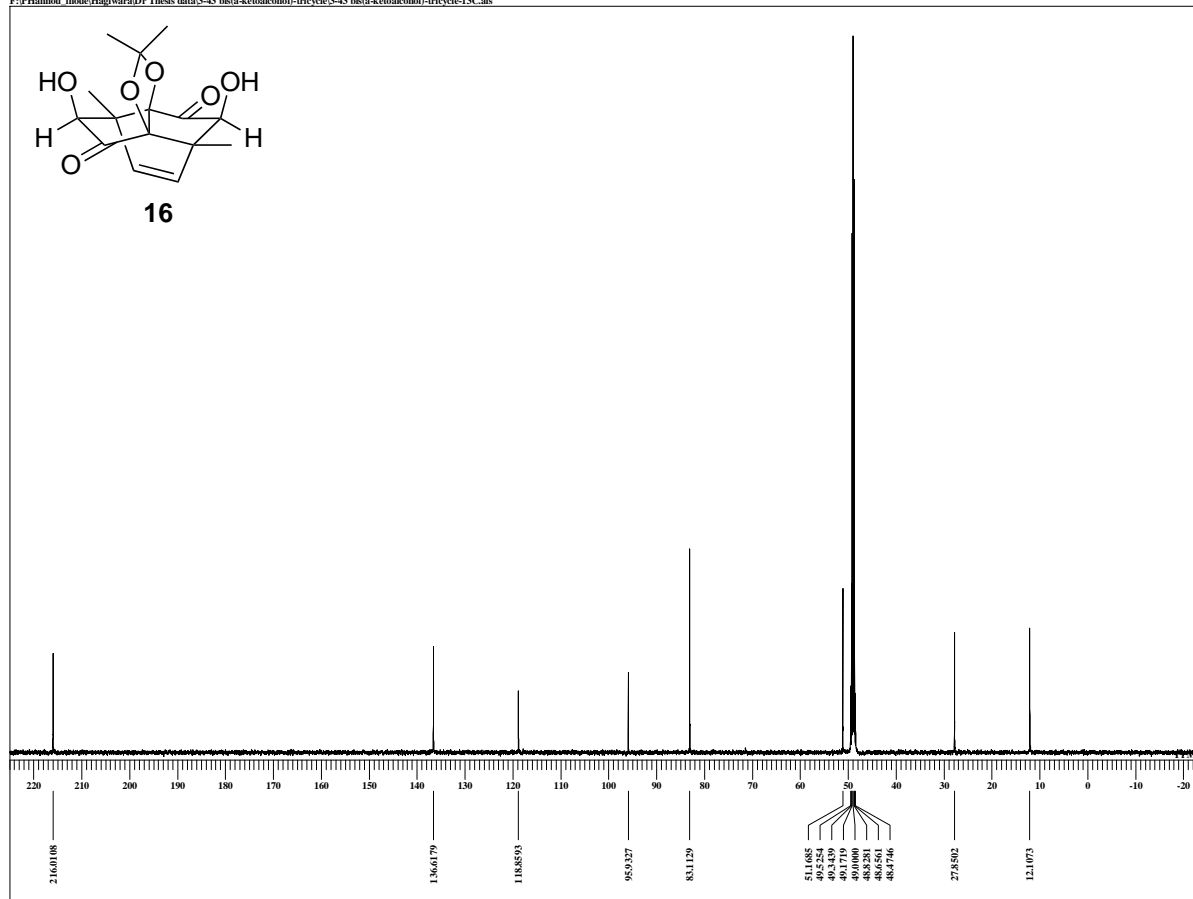


```

DFILE 3-43 bis(a-ketoalcohol)-t
COMNT single_pulse
DATIM 08-10-2008 14:06:17
MENUE
OBNUC 1H
OFR 490.15 MHz
OFRFQ 490.15 MHz
OBSET 9.16 KHz
OBFIN 7.60 Hz
PW1 8.55 usec
DEADT 0.00 usec
PREDL 0.00000 msec
IWT 1.0000 sec
POINT 13107
SFO 13107
TIMES 8
DUMMY 1
FREQU 7352.83 Hz
FLT 37000 Hz
DELAY 13.52 usec
ACQTM 1.7826 sec
PD 1.5000 sec
SCANS 8
ADBIT 16
RGAIN 44
BF 0.01 Hz
T1 0.00
T2 0.00
T3 90.00
T4 100.00
EXMOD single_pulse.ex2
EXPCM
IRNUC 1H
IFR 490.15 MHz
IRSET 9.16 KHz
IRFIN 7.60 Hz
IRRPW 118 usec
IRATN 79
DFILE 3-43 bis(a-ketoalcohol)-t
SF
LKSET 70.00 KHz
LKFIN 36.6 Hz
LKLEV 0
LGAIN 0
LKPHS 0
LKSIG 0
CSPED 0 Hz
FILDC
FILDF
CTEMP 20.8 c
SLVNT CD3OD
ENREF 3.31 ppm
    
```

single pulse decoupled gated NOE

F:\Phamou Inoue\Hagiwara\Dr Thesis\data\3-43 bis(a-ketoalcohol)-tricycle\3-43 bis(a-ketoalcohol)-tricycle-13Cals

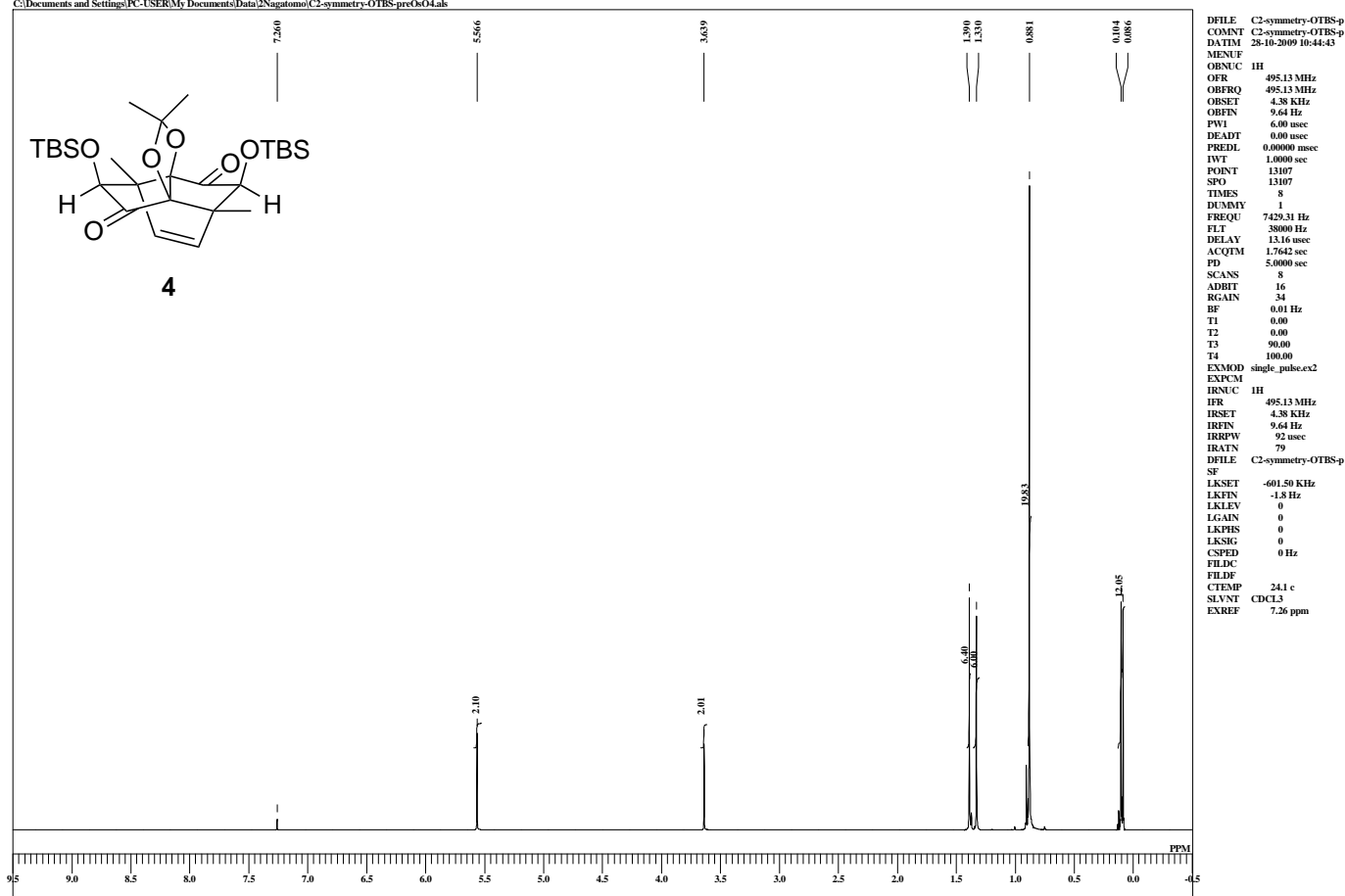


```

DFILE 3-43 bis(a-ketoalcohol)-tr
COMNT single pulse decoupled gat
DATIM 12-09-2008 10:57:44
MENUE
OBNUC 13C
OFR 123.26 MHz
OFRFQ 123.26 MHz
OBSET 2.31 KHz
OBFIN 6.71 Hz
PW1 3.20 usec
DEADT 0.00 usec
PREDL 0.00000 msec
IWT 1.0000 sec
POINT 26214
SFO 26214
TIMES 32
DUMMY 4
FREQU 30863.73 Hz
FLT 155000 Hz
DELAY 21.06 usec
ACQTM 0.8493 sec
PD 5.0000 sec
SCANS 32
ADBIT 16
RGAIN 60
BF 0.01 Hz
T1 0.00
T2 0.00
T3 90.00
T4 100.00
EXMOD single_pulse_dec
EXPCM
IRNUC 13C
IFR 123.26 MHz
IRSET 2.31 KHz
IRFIN 6.71 Hz
IRRPW 92 usec
IRATN 79
DFILE 3-43 bis(a-ketoalcohol)-tr
SF
LKSET 70.00 KHz
LKFIN 36.6 Hz
LKLEV 0
LGAIN 0
LKPHS 0
LKSIG 0
CSPED 0 Hz
FILDC
FILDF
CTEMP 20.8 c
SLVNT CD3OD
ENREF 49.00 ppm
    
```

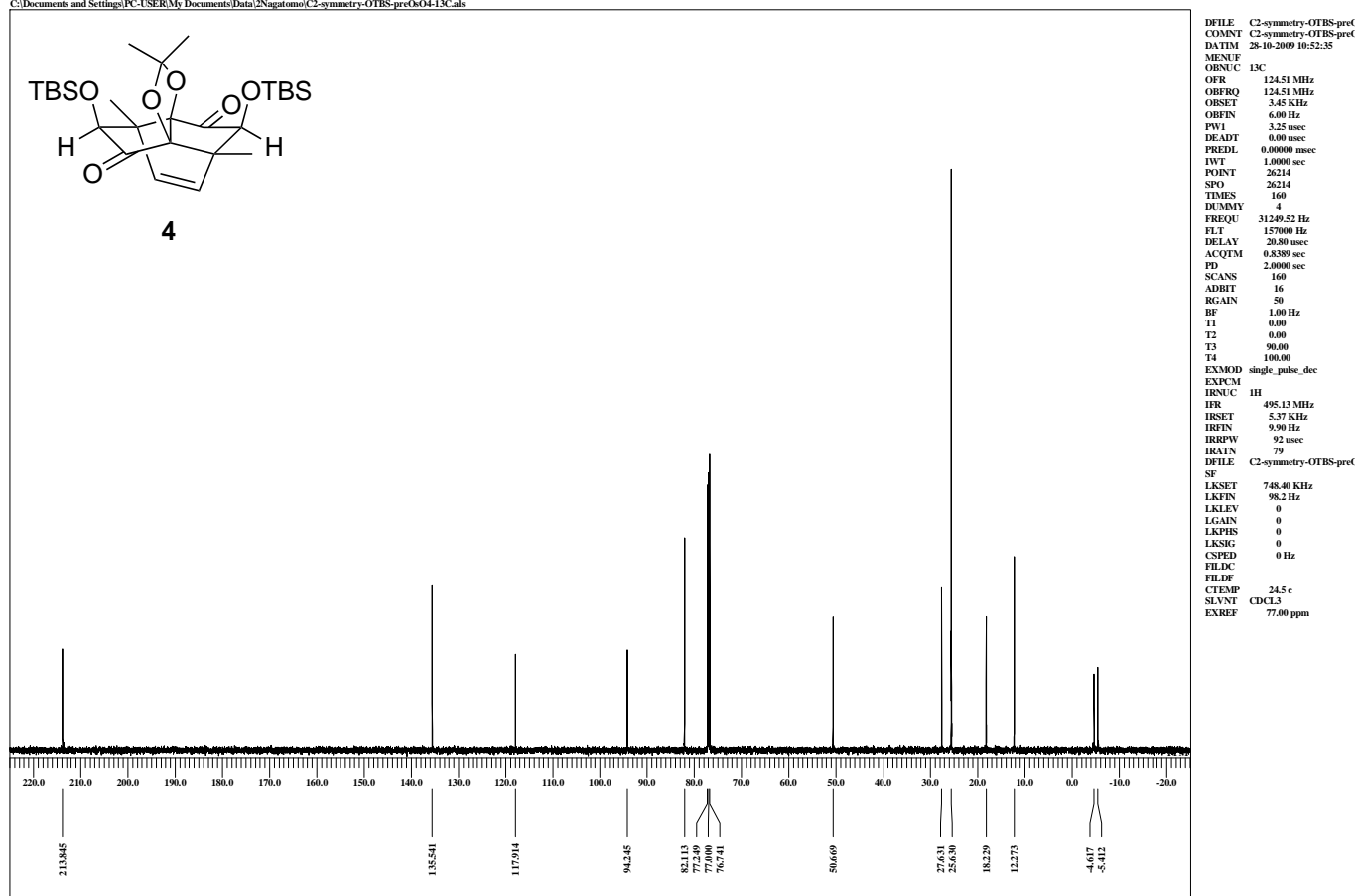
C2-symmetry-OTBS-preOsO4

C:\Documents and Settings\PC-USER\My Documents\Data\2Nagatomo\C2-symmetry-OTBS-preOsO4.aik



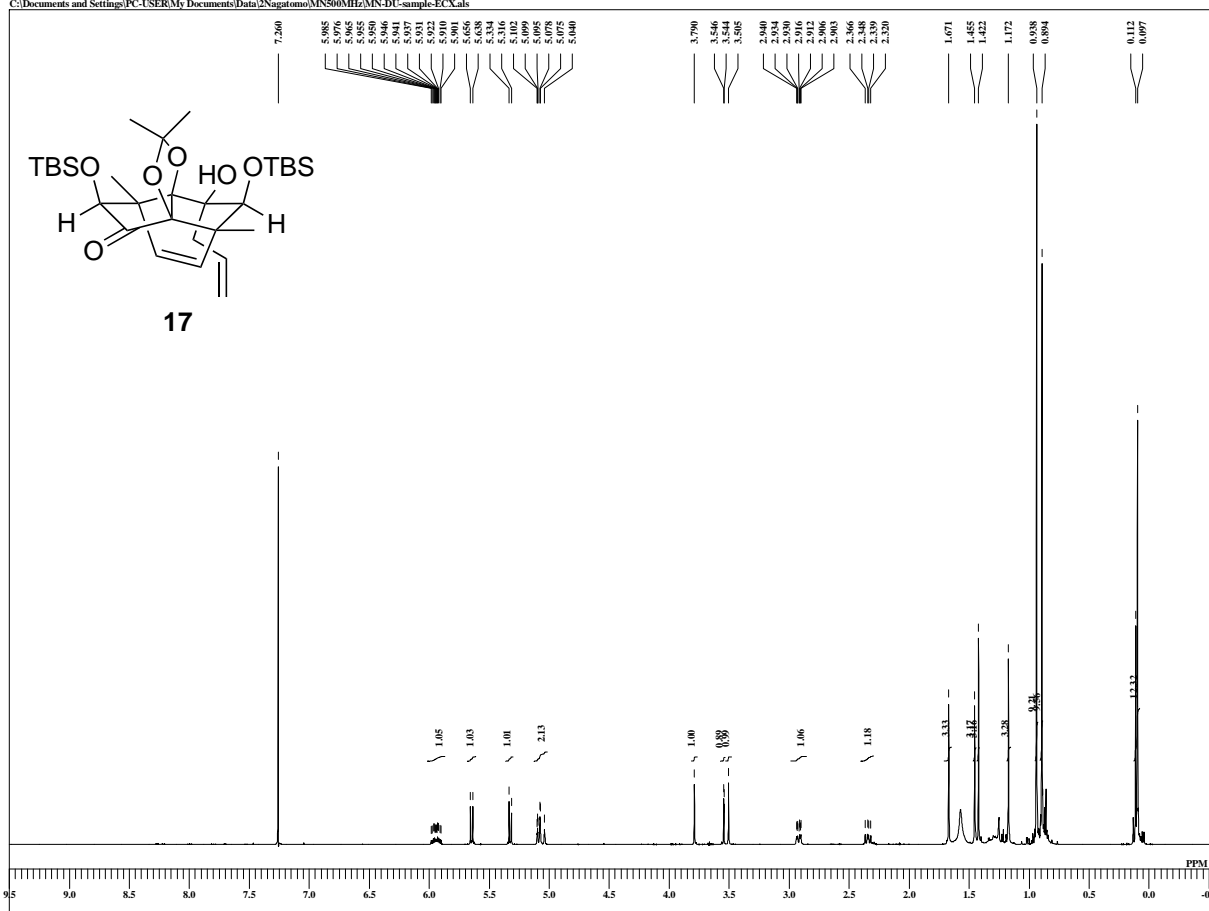
C2-symmetry-OTBS-preOsO4

C:\Documents and Settings\PC-USER\My Documents\Data\2Nagatomo\C2-symmetry-OTBS-preOsO4-13C.aik



MN-DU-sample-ECX

C:\Documents and Settings\PC-USER\My Documents\Data\2Nagatomo\MN500MHz\MN-DU-sample-ECX.xls

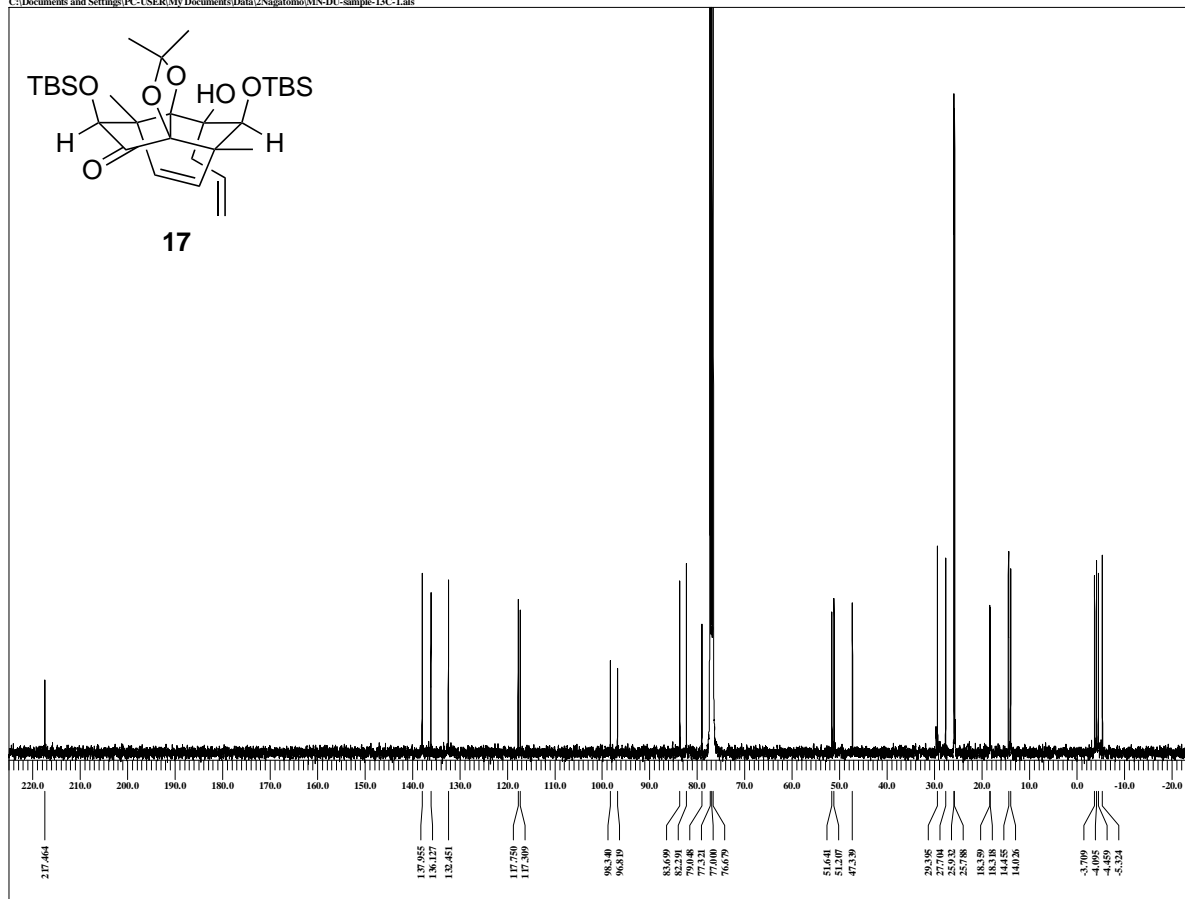


```

FILE MN-DU-sample-ECX.a1
COMNT MN-DU-sample-ECX
DATIM 03-09-2012 11:31:45
MENEF
OBNUC 1H
OFR 495.13 MHz
OFRQ 495.13 MHz
OBSET 4.38 KHz
OBFIN 9.64 Hz
PW1 6.00 usec
DEADT 0.00 usec
PREDL 0.00000 msec
IWT 1.0000 sec
POINT 13107
SFO 13107
TIMES 64
DUMMY 1
FREQU 7429.31 Hz
FLT 38000 Hz
DELAY 13.16 usec
ACQTM 1.7642 sec
PD 2.0000 sec
SCANS 64
ADBIT 16
RGAIN 50
BF 0.01 Hz
T1 0.00
T2 0.00
T3 90.00
T4 100.00
EXMOD single_pulse.ec2
EXPCM
IRNUC 1H
IFR 495.13 MHz
IRSET 4.38 KHz
IRFIN 9.64 Hz
IRRPW 92 usec
IRATN 79
DEFILE MN-DU-sample-ECX.a1
SF
LKSET -601.50 KHz
LKFN -1.5 Hz
LKLEV 0
LGAIN 0
LKPIS 0
LKSIG 0
CSPED 0 Hz
FILDC
FILDF
CTEMP 23.4 c
SLVNT CDCL3
XREF 7.26 ppm
    
```

MN-DU-sample-13C

C:\Documents and Settings\PC-USER\My Documents\Data\2Nagatomo\MN-DU-sample-13C-1.xls

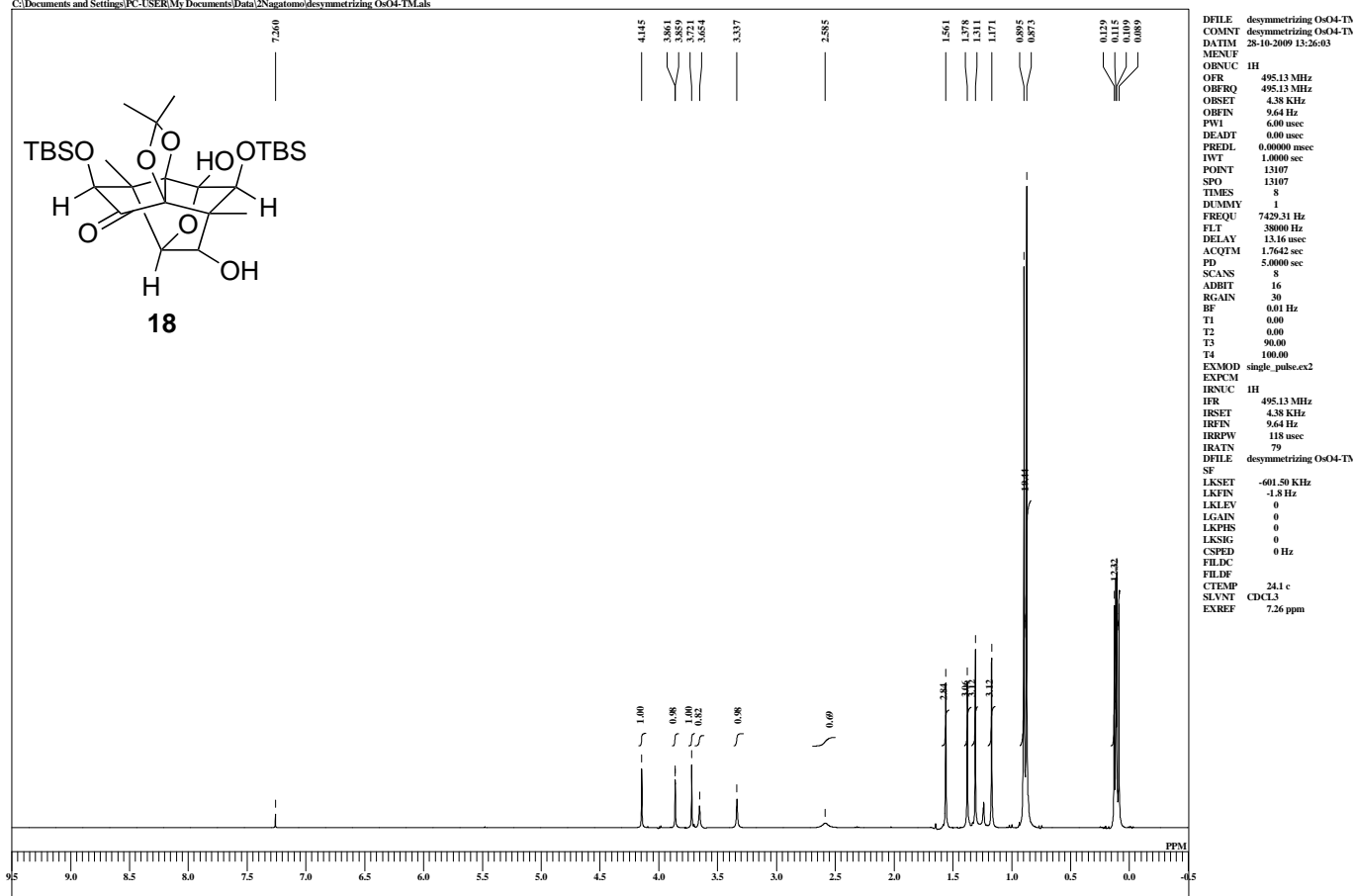


```

FILE MN-DU-sample-13C-1.xls
COMNT MN-DU-sample-13C
DATIM 04-09-2012 09:11:39
MENEF
OBNUC 13C
OFR 99.55 MHz
OFRQ 99.55 MHz
OBSET 5.13 KHz
OBFIN 0.98 Hz
PW1 3.25 usec
DEADT 0.00 usec
PREDL 0.00000 msec
IWT 1.0000 sec
POINT 104856
SFO 104856
TIMES 13926
DUMMY 4
FREQU 24999.62 Hz
FLT 125000 Hz
DELAY 20.50 usec
ACQTM 1.0486 sec
PD 2.0000 sec
SCANS 13926
ADBIT 16
RGAIN 60
BF 1.00 Hz
T1 0.00
T2 0.00
T3 100.00
T4 100.00
EXMOD single_pulse_dec
EXPCM
IRNUC 13C
IFR 395.88 MHz
IRSET 6.28 KHz
IRFIN 0.97 Hz
IRRPW 115 usec
IRATN 79
DEFILE MN-DU-sample-13C-1.xls
SF
LKSET 13.20 KHz
LKFN 75.7 Hz
LKLEV 0
LGAIN 0
LKPIS 0
LKSIG 0
CSPED 0 Hz
FILDC
FILDF
CTEMP 23.4 c
SLVNT CDCL3
XREF 77.00 ppm
    
```

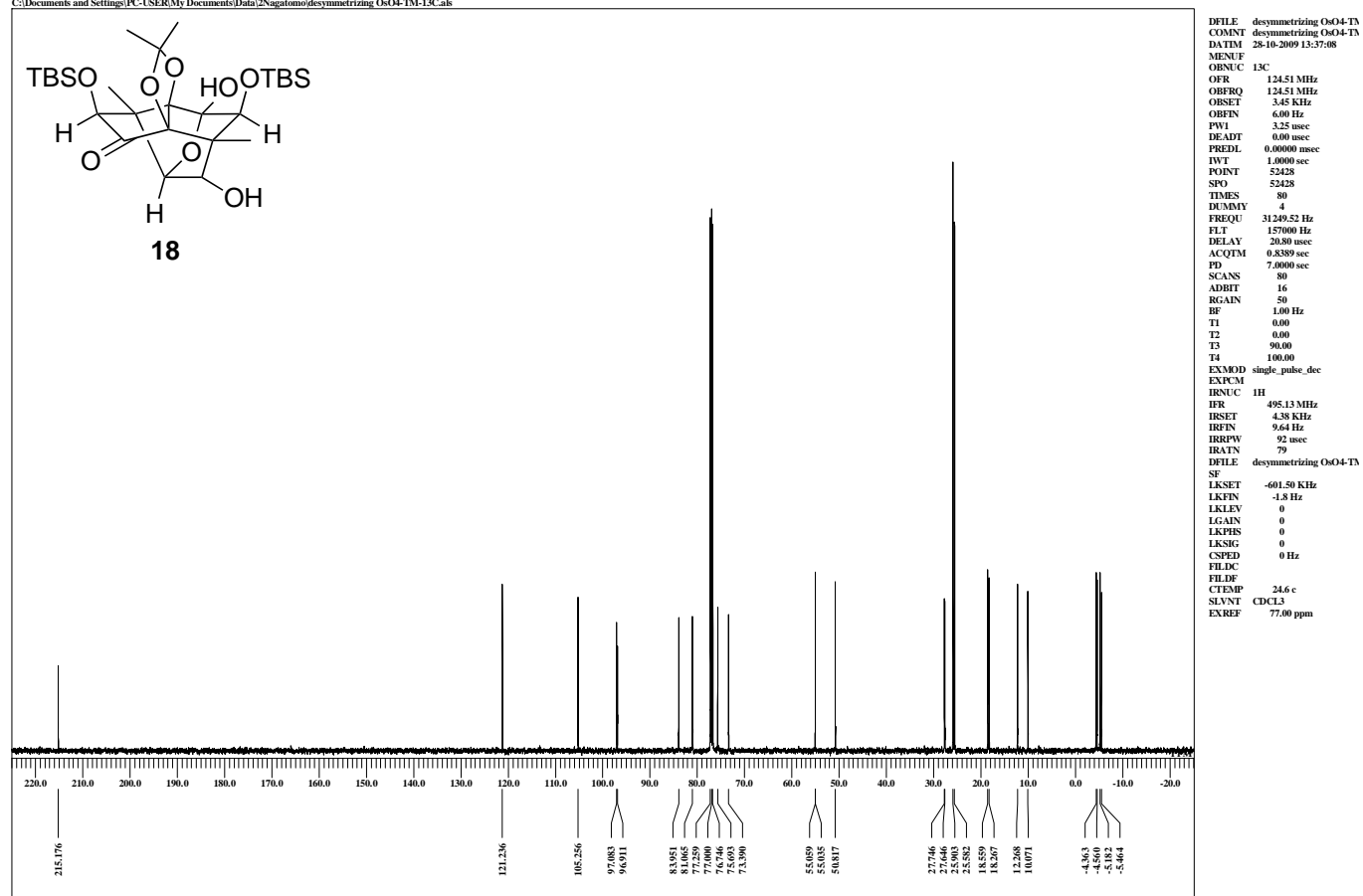
desymmetrizing OsO4-TM

C:\Documents and Settings\PC-USER\My Documents\Data\2Nagatomo\desymmetrizing OsO4-TM.cals



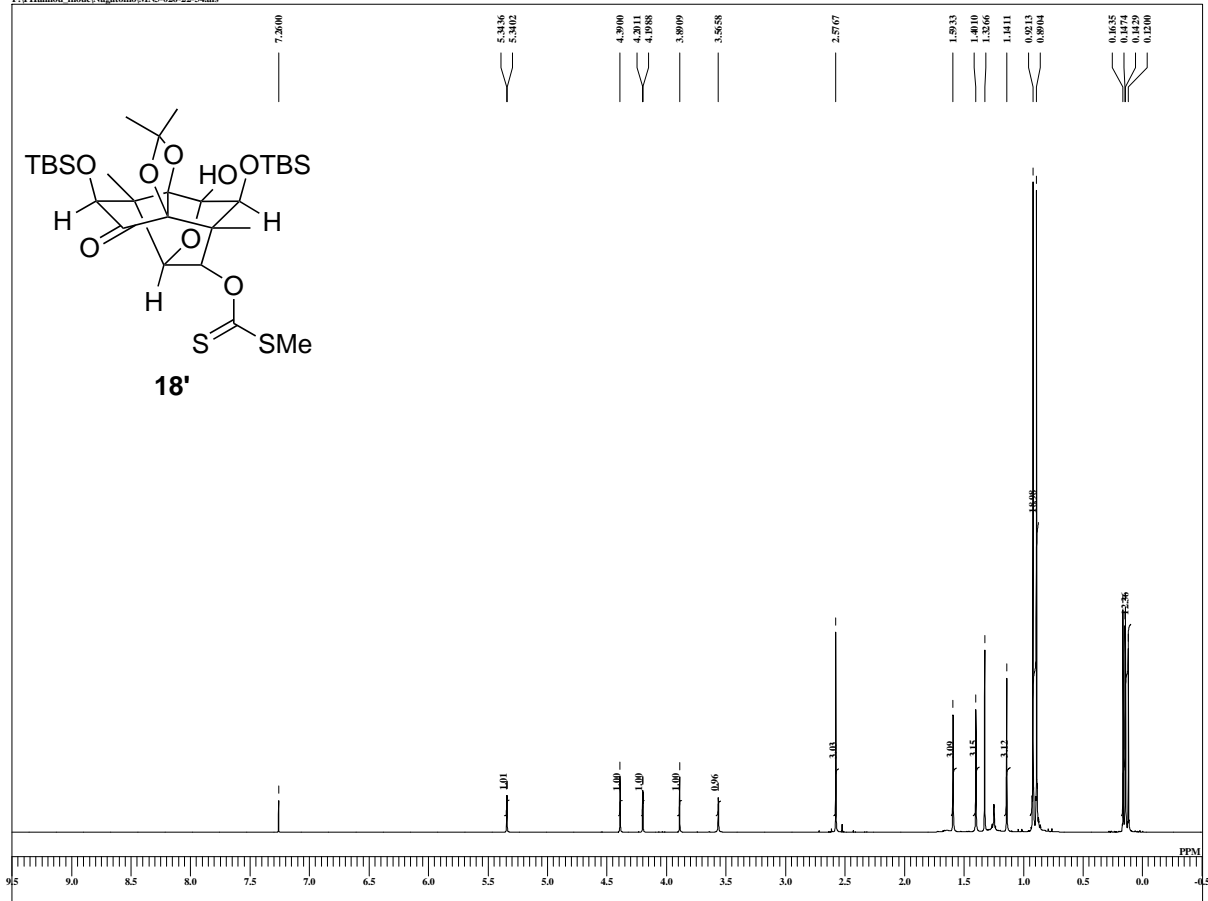
desymmetrizing OsO4-TM

C:\Documents and Settings\PC-USER\My Documents\Data\2Nagatomo\desymmetrizing OsO4-TM-13C.cals



MN3-026-22-34

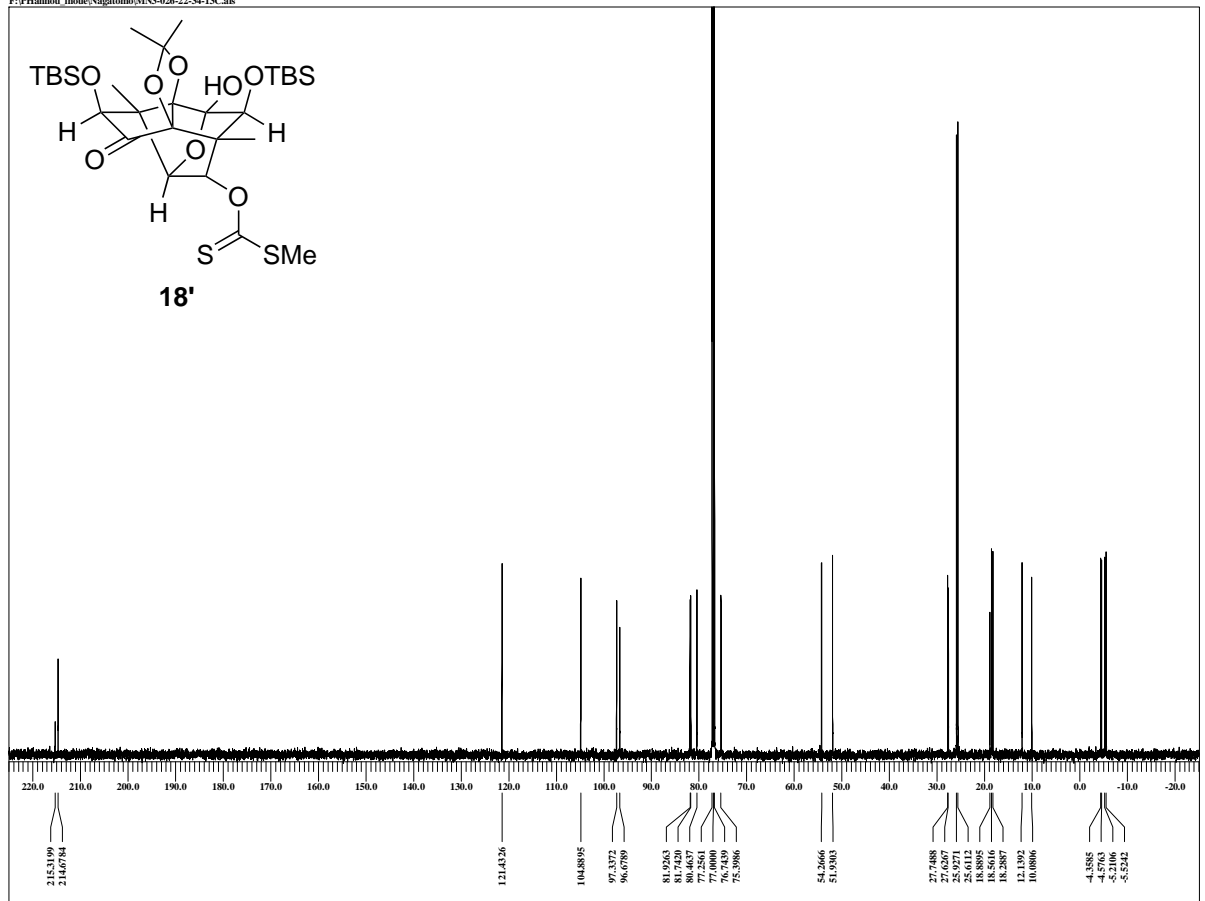
F:\PHannou inoue\Nagatomo\MN3-026-22-34.als



DFILE MN3-026-22-34.als
 COMNT MN3-026-22-34
 DATIM 29-10-2009 16:04:18
 MENTF
 OBNUC 1H
 OFR 495.13 MHz
 OFRFQ 495.13 MHz
 OBSSET 4.38 KHz
 OBFN 9.64 Hz
 PW1 6.00 usec
 DEADT 0.90 usec
 PREDL 0.00000 msec
 IWV 1.0000 sec
 POINT 13107
 SPO 13107
 TIMES 16
 DUMMY 1
 FREQU 7429.31 Hz
 FLT 38000 Hz
 DELAY 13.16 usec
 ACQTM 1.7642 sec
 PD 5.0000 sec
 SCANS 16
 ADBIT 16
 RGAIN 36
 BF 0.01 Hz
 T1 0.00
 T2 0.00
 T3 90.00
 T4 100.00
 EXMOD single_pulse.ex2
 EXPCM
 IRNUC 1H
 IFR 495.13 MHz
 IRSET 4.38 KHz
 IRFN 9.64 Hz
 IRFPW 92 usec
 IRATN 79
 DFILF
 DFILF MN3-026-22-34.als
 SF
 LKSET -601.50 KHz
 LKFN -1.8 Hz
 LKLEV 0
 LGAIN 0
 LKPS 0
 LKSG 0
 CSPED 0 Hz
 FILDC
 FILDF
 CTEMP 23.9 c
 SLVNT CDCl3
 EXREF 7.26 ppm

MN3-026-22-34

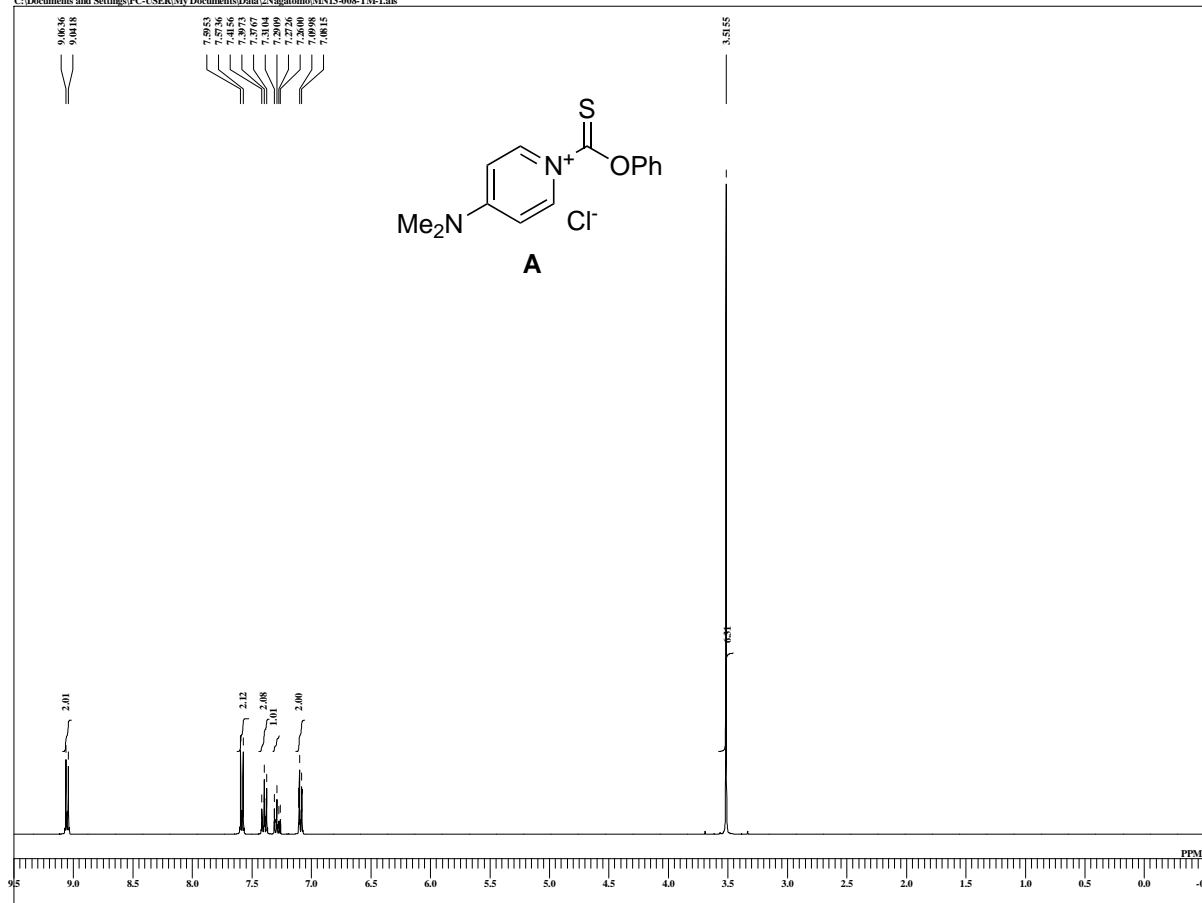
F:\PHannou inoue\Nagatomo\MN3-026-22-34-13C.als



DFILE MN3-026-22-34-13C.als
 COMNT MN3-026-22-34
 DATIM 29-10-2009 16:17:60
 MENTF
 OBNUC 13C
 OFR 124.51 MHz
 OFRFQ 124.51 MHz
 OBSSET 3.45 KHz
 OBFN 6.00 Hz
 PW1 3.25 usec
 DEADT 0.90 usec
 PREDL 0.00000 msec
 IWV 1.0000 sec
 POINT 104856
 SPO 104856
 TIMES 100
 DUMMY 4
 FREQU 31249.52 Hz
 FLT 157000 Hz
 DELAY 20.80 usec
 ACQTM 0.8389 sec
 PD 7.0000 sec
 SCANS 100
 ADBIT 16
 RGAIN 50
 BF 1.00 Hz
 T1 0.00
 T2 0.00
 T3 90.00
 T4 100.00
 EXMOD single_pulse_dec
 EXPCM
 IRNUC 1H
 IFR 495.13 MHz
 IRSET 4.38 KHz
 IRFN 9.64 Hz
 IRFPW 92 usec
 IRATN 79
 DFILF
 DFILF MN3-026-22-34-13C.als
 SF
 LKSET -601.50 KHz
 LKFN -1.8 Hz
 LKLEV 0
 LGAIN 0
 LKPS 0
 LKSG 0
 CSPED 0 Hz
 FILDC
 FILDF
 CTEMP 24.4 c
 SLVNT CDCl3
 EXREF 77.00 ppm

MN13-008-TM

C:\Documents and Settings\PC-USER\My Documents\Data\2Nagatomo\MN13-008-TM-Lals

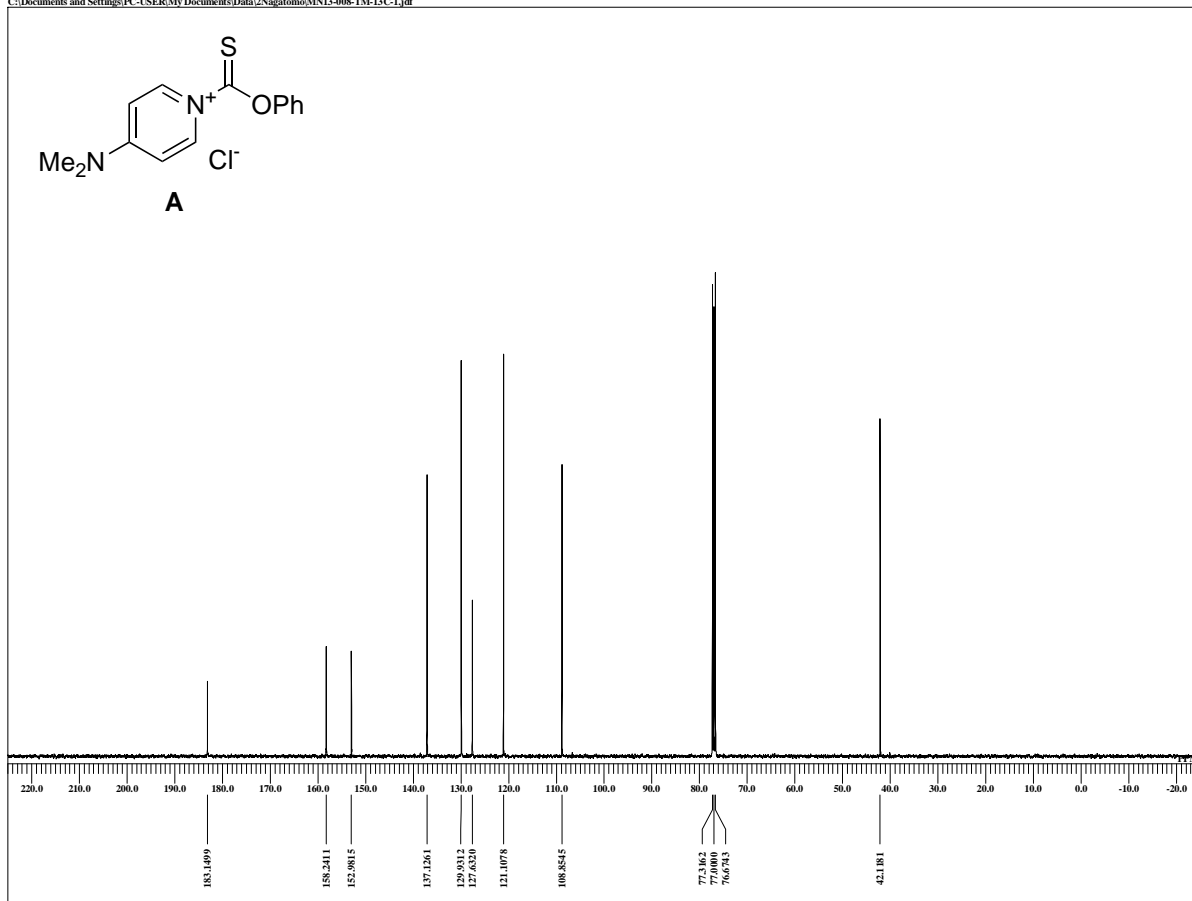


```

DFILE  MN13-008-TM-Lals
COMNT  MN13-008-TM
DATIM  30-05-2012 16:49:30
MENUF
OBNUC  1H
OFR    395.88 MHz
OBFREQ 395.88 MHz
OBSET  6.28 KHz
OBFN   0.87 Hz
PW1    6.38 usec
DEADT  0.90 usec
PREDL  0.00000 msec
IWT    1.0000 sec
POINT  13107
SPO    13107
TIMES  32
DUMMY  1
FREQU  5938.15 Hz
FLT    38000 Hz
DELAY  16.68 usec
ACQTM  2.2073 sec
PD     2.0000 sec
SCANS  32
ADBIT  16
RGAIN  30
BF     0.01 Hz
T1    0.00
T2    0.00
T3    90.00
T4    100.00
EXMOD  single_pulse.ex2
EXPCM
IRNUC  1H
IFR    395.88 MHz
IRSET  6.28 KHz
IRFN   0.87 Hz
IRFPW  115 usec
IRATN  79
DFILE  MN13-008-TM-Lals
SF     13.20 KHz
LKSET  13.20 KHz
LKFIN  75.7 Hz
LKLEV  0
LGAIN  0
LKPHS  0
LKSG   0
CSPED  0 Hz
FILDC
FILDF
CTEMP  24.8 c
SLVNT  CDCl3
XREF  7.26 ppm
    
```

MN13-008-TM

C:\Documents and Settings\PC-USER\My Documents\Data\2Nagatomo\MN13-008-TM-13C-1.jdf

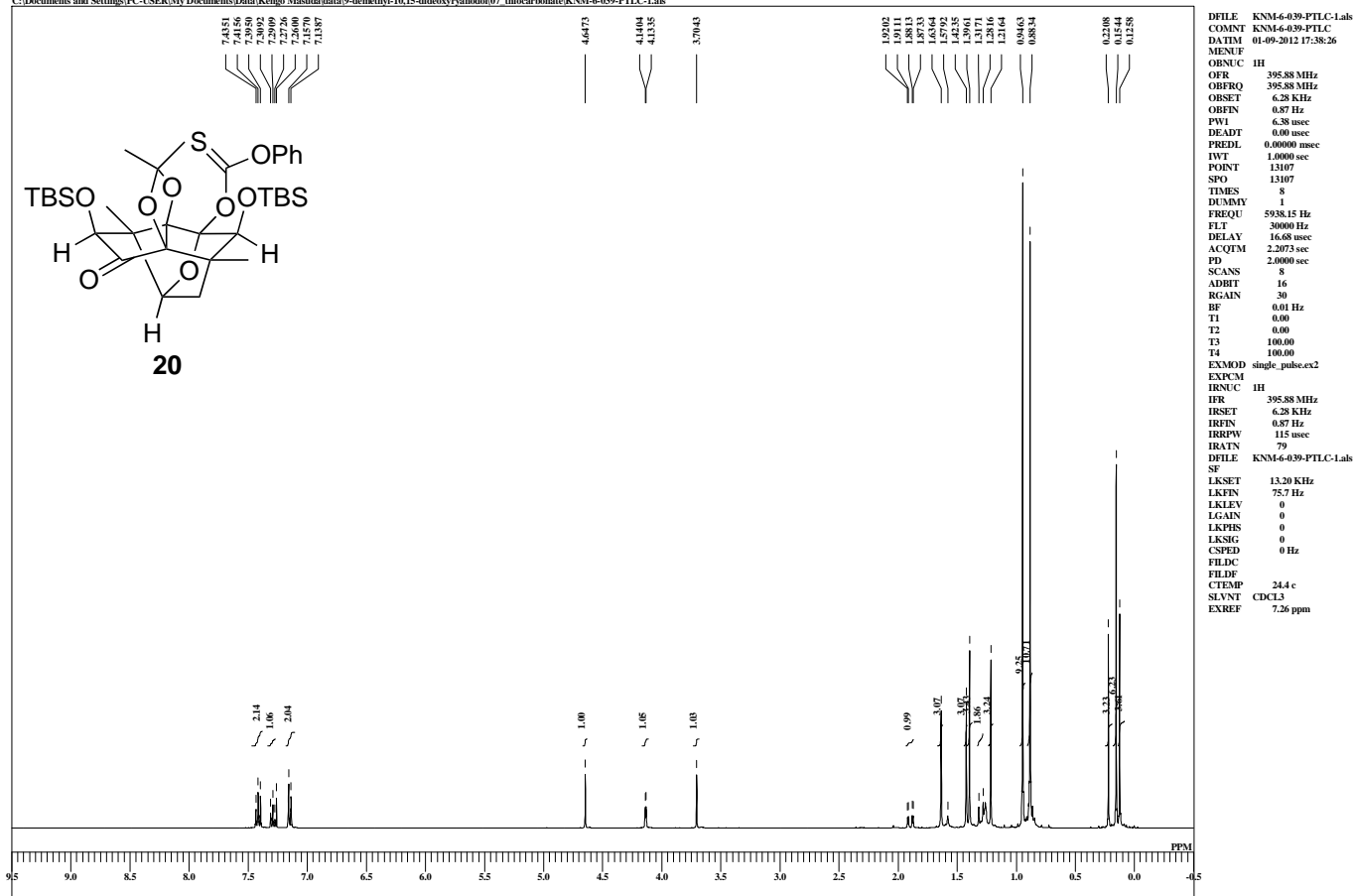


```

DFILE  MN13-008-TM-13C-1.jdf
COMNT  MN13-008-TM
DATIM  30-05-2012 17:05:02
MENUF
OBNUC  13C
OFR    99.55 MHz
OBFREQ 99.55 MHz
OBSET  5.13 KHz
OBFN   0.98 Hz
PW1    3.25 usec
DEADT  0.00 usec
PREDL  0.00000 msec
IWT    1.0000 sec
POINT  32768
SPO    32768
TIMES  301
DUMMY  4
FREQU  31250.00 Hz
FLT    125000 Hz
DELAY  20.50 usec
ACQTM  1.0486 sec
PD     2.0000 sec
SCANS  301
ADBIT  16
RGAIN  30
BF     1.00 Hz
T1    0.00
T2    0.00
T3    90.00
T4    100.00
EXMOD  single_pulse_dec
EXPCM
IRNUC  13C
IFR    99.55 MHz
IRSET  6.28 KHz
IRFN   0.87 Hz
IRFPW  115 usec
IRATN  79
DFILE  MN13-008-TM-13C-1.jdf
SF     13.20 KHz
LKSET  13.20 KHz
LKFIN  75.7 Hz
LKLEV  0
LGAIN  0
LKPHS  0
LKSG   0
CSPED  0 Hz
FILDC
FILDF
CTEMP  24.9 c
SLVNT  CDCl3
XREF  77.00 ppm
    
```


KNM-6-039-PTLC

C:\Documents and Settings\PC-USER\My Documents\Data\Kengo Masuda\data\9-demethyl-10,15-dideoxyryanodol\07_thiocarbonate\KNM-6-039-PTLC-1.ac

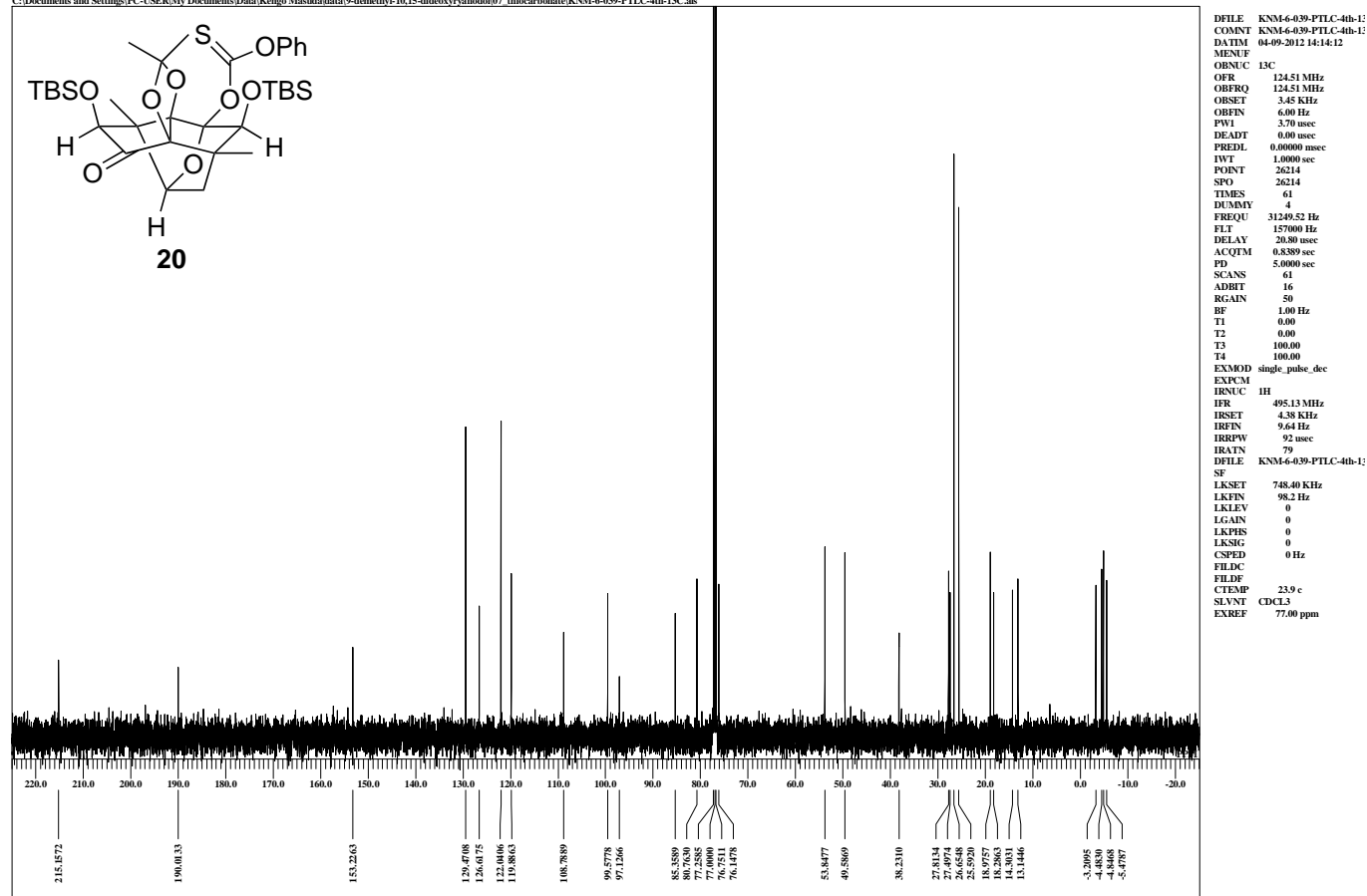


```

DFILE KNM4-039-PTLC-1.ac
COMNT KNM4-039-PTLC-1.ac
DATIM 01-09-2012 17:38:26
MENEF
OBNUC IH
OFR 395.88 MHz
OBFREQ 395.88 MHz
OBSET 6.28 kHz
OBFEN 0.87 Hz
PW1 6.38 usec
DEADT 0.00 usec
PREDL 0.00000 msec
IWT 1.0000 sec
POINT 13107
SPO 13107
TIMES 8
DUMMY 1
FREQU 5938.15 Hz
FLT 30000 Hz
DELAY 16.68 usec
ACQTM 2.2073 sec
PD 2.0000 sec
SCANS 8
ADBIT 16
RGAIN 30
BF 0.01 Hz
T1 0.00
T2 0.00
T3 100.00
T4 100.00
EXMOD single_pulse.ex2
EXPCM
IRNUC IH
IRF 395.88 MHz
IRSET 6.28 kHz
IRFEN 0.87 Hz
IRFPW 115 usec
IRATN 79
DFILE KNM4-039-PTLC-1.ac
SE
LKSET 13.20 kHz
LKFIN 75.7 Hz
LKLEV 0
LGAIN 0
LKPHS 0
LKSG 0
CSPED 0 Hz
FILDC
FILDF
CTEMP 24.4 c
SLVNT CDCL3
EXREF 7.26 ppm
    
```

KNM-6-039-PTLC-4th-13C

C:\Documents and Settings\PC-USER\My Documents\Data\Kengo Masuda\data\9-demethyl-10,15-dideoxyryanodol\07_thiocarbonate\KNM-6-039-PTLC-4th-13C.ac

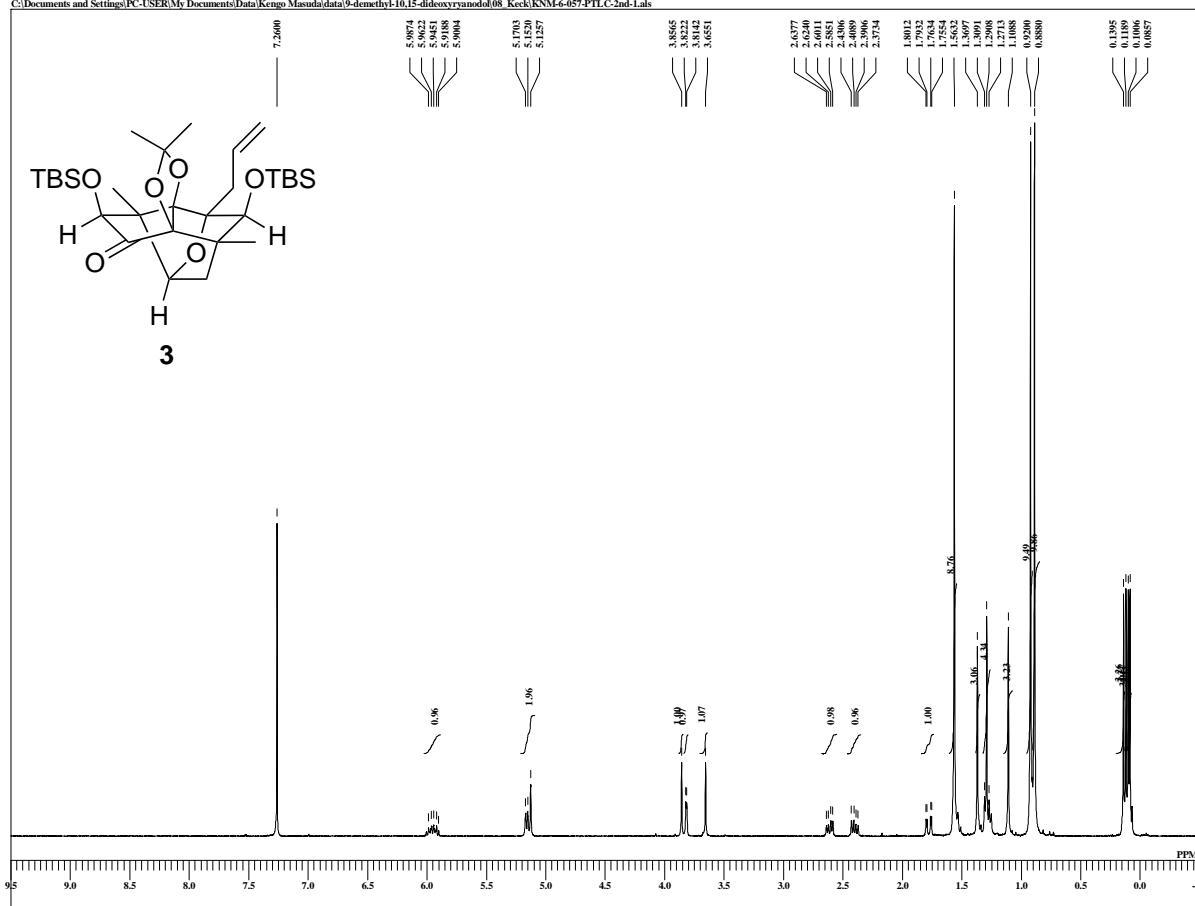


```

DFILE KNM4-039-PTLC-4th-13
COMNT KNM4-039-PTLC-4th-13
DATIM 04-09-2012 14:14:12
MENEF
OBNUC 13C
OFR 124.51 MHz
OBFREQ 124.51 MHz
OBSET 3.45 kHz
OBFEN 6.00 Hz
PW1 3.70 usec
DEADT 0.00 usec
PREDL 0.00000 msec
IWT 1.0000 sec
POINT 26214
SPO 26214
TIMES 61
DUMMY 4
FREQU 31249.52 Hz
FLT 157000 Hz
DELAY 20.80 usec
ACQTM 0.8389 sec
PD 5.0000 sec
SCANS 61
ADBIT 16
RGAIN 50
BF 1.00 Hz
T1 0.00
T2 0.00
T3 100.00
T4 100.00
EXMOD single_pulse_dec
EXPCM
IRNUC IH
IRF 495.13 MHz
IRSET 4.38 kHz
IRFEN 9.64 Hz
IRFPW 92 usec
IRATN 79
DFILE KNM4-039-PTLC-4th-13
SE
LKSET 748.40 kHz
LKFIN 98.2 Hz
LKLEV 0
LGAIN 0
LKPHS 0
LKSG 0
CSPED 0 Hz
FILDC
FILDF
CTEMP 23.9 c
SLVNT CDCL3
EXREF 77.00 ppm
    
```

KNM-6-057-PTLC-2nd

C:\Documents and Settings\PC-USER\My Documents\Data\Kengo Masuda\data\9-demethyl-10,15-dideoxyryanodol\08_Keck\KNM-6-057-PTLC-2nd-1.xls

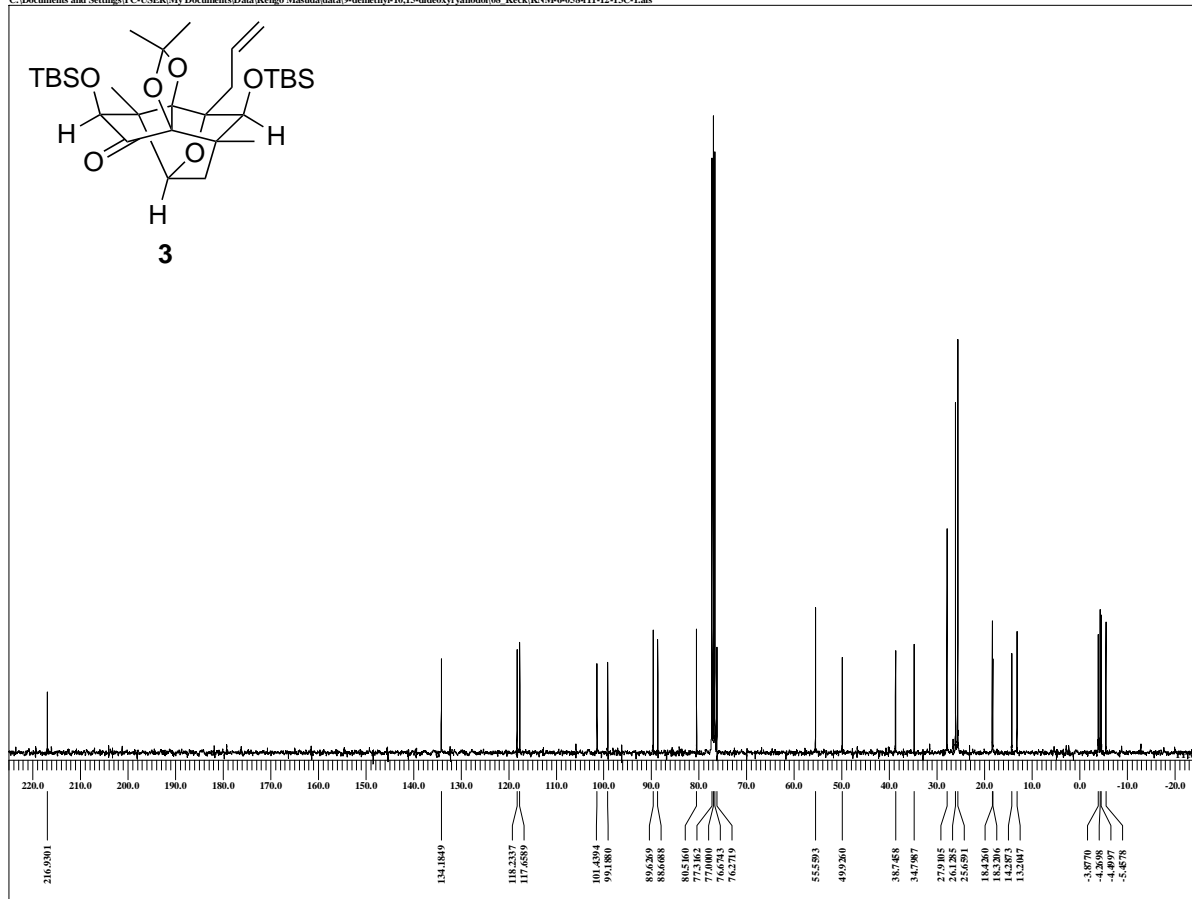


```

FILE KNM-6-057-PTLC-2nd-1
COMNT KNM-6-057-PTLC-2nd
DATIM 10-09-2012 11:39:02
MENEF
OBNUC 1H
OFR 395.88 MHz
OFRFQ 395.88 MHz
OBSET 6.28 KHz
OBFN 0.87 Hz
PWI 6.38 usec
DEADT 0.00 usec
PREDL 0.00000 msec
IWT 1.0000 sec
POINT 13107
SFO 13107
TIMES 16
DUMMY 1
FREQU 5938.15 Hz
FLT 30000 Hz
DELAY 16.68 usec
ACQTM 2.2073 sec
PD 2.0000 sec
SCANS 16
ADBIT 16
RGAIN 46
BF 0.01 Hz
T1 0.00
T2 0.00
T3 100.00
T4 100.00
EXMOD single_pulse.ex2
EXPCM
IRNUC 1H
IFR 395.88 MHz
IRSET 6.28 KHz
IRFN 0.87 Hz
IRFPV 1 usec
IRATN 10
DFILE KNM-6-057-PTLC-2nd-1
SF 13.20 KHz
LKSET 13.20 KHz
LKFN 75.7 Hz
LKLEV 0
LGAIN 0
LKPS 0
LKSG 0
CSPED 0 Hz
FILEC
FLDF
CTEMP 21.0 c
SLVNT CDCl3
EXREF 7.26 ppm
    
```

KNM-6-058-f11-12-13C

C:\Documents and Settings\PC-USER\My Documents\Data\Kengo Masuda\data\9-demethyl-10,15-dideoxyryanodol\08_Keck\KNM-6-058-f11-12-13C-1.xls

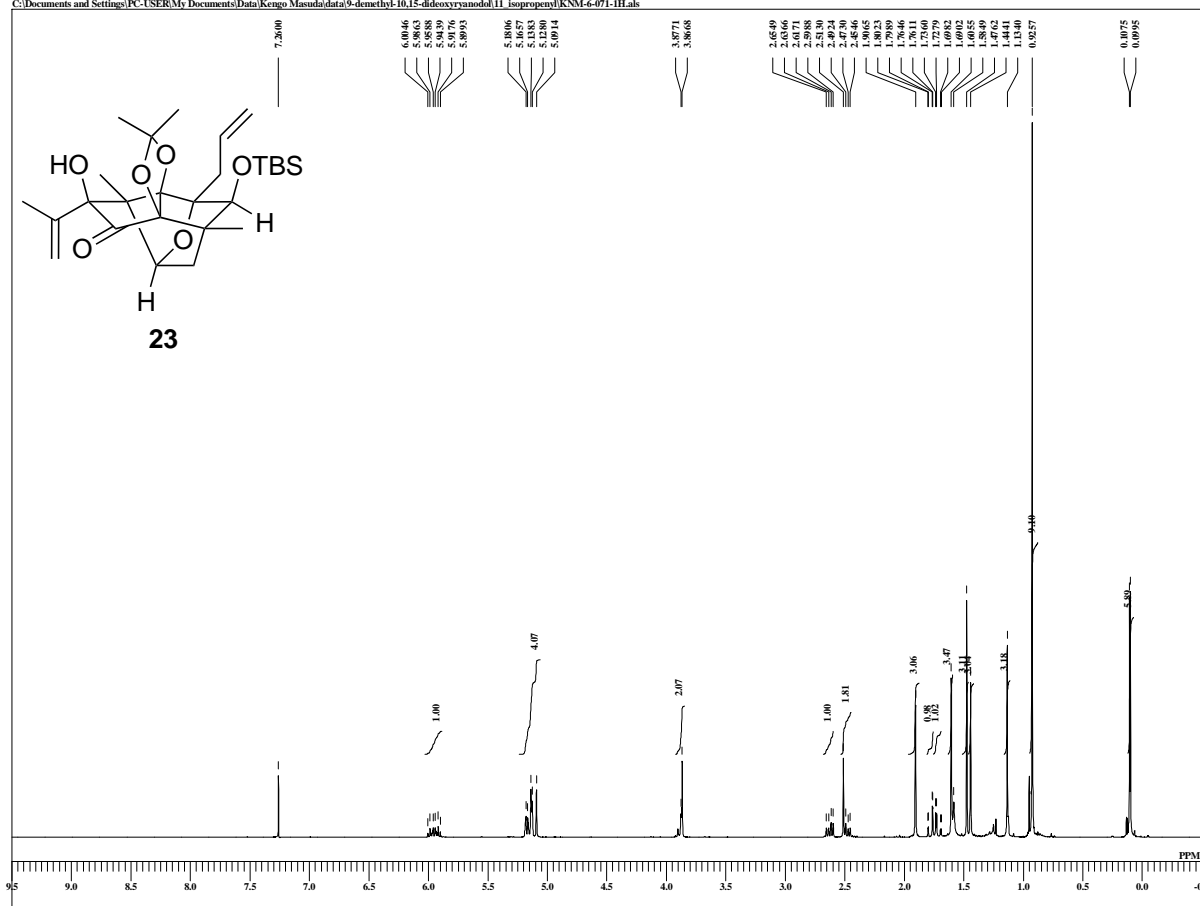


```

FILE KNM-6-058-f11-12-13C-1
COMNT KNM-6-058-f11-12-13C
DATIM 11-09-2012 10:03:12
MENEF
OBNUC 13C
OFR 99.55 MHz
OFRFQ 99.55 MHz
OBSET 5.13 KHz
OBFN 0.98 Hz
PWI 3.25 usec
DEADT 0.00 usec
PREDL 0.00000 msec
IWT 1.0000 sec
POINT 26214
SFO 26214
TIMES 60
DUMMY 4
FREQU 24999.62 Hz
FLT 125000 Hz
DELAY 20.50 usec
ACQTM 1.0486 sec
PD 5.0000 sec
SCANS 60
ADBIT 16
RGAIN 60
BF 1.00 Hz
T1 0.00
T2 0.00
T3 100.00
T4 100.00
EXMOD single_pulse_dec
EXPCM
IRNUC 13C
IFR 99.55 MHz
IRSET 5.13 KHz
IRFN 0.97 Hz
IRFPV 115 usec
IRATN 79
DFILE KNM-6-058-f11-12-13C-1
SF 13.20 KHz
LKSET 13.20 KHz
LKFN 75.7 Hz
LKLEV 0
LGAIN 0
LKPS 0
LKSG 0
CSPED 0 Hz
FILEC
FLDF
CTEMP 23.1 c
SLVNT CDCl3
EXREF 77.00 ppm
    
```

KNM-6-071-1H

C:\Documents and Settings\PC-USER\My Documents\Data\Kengo Masuda\data\9-demethyl-10,15-dideoxyryanodol\11_isopropenyl\KNM-6-071-1H.lab

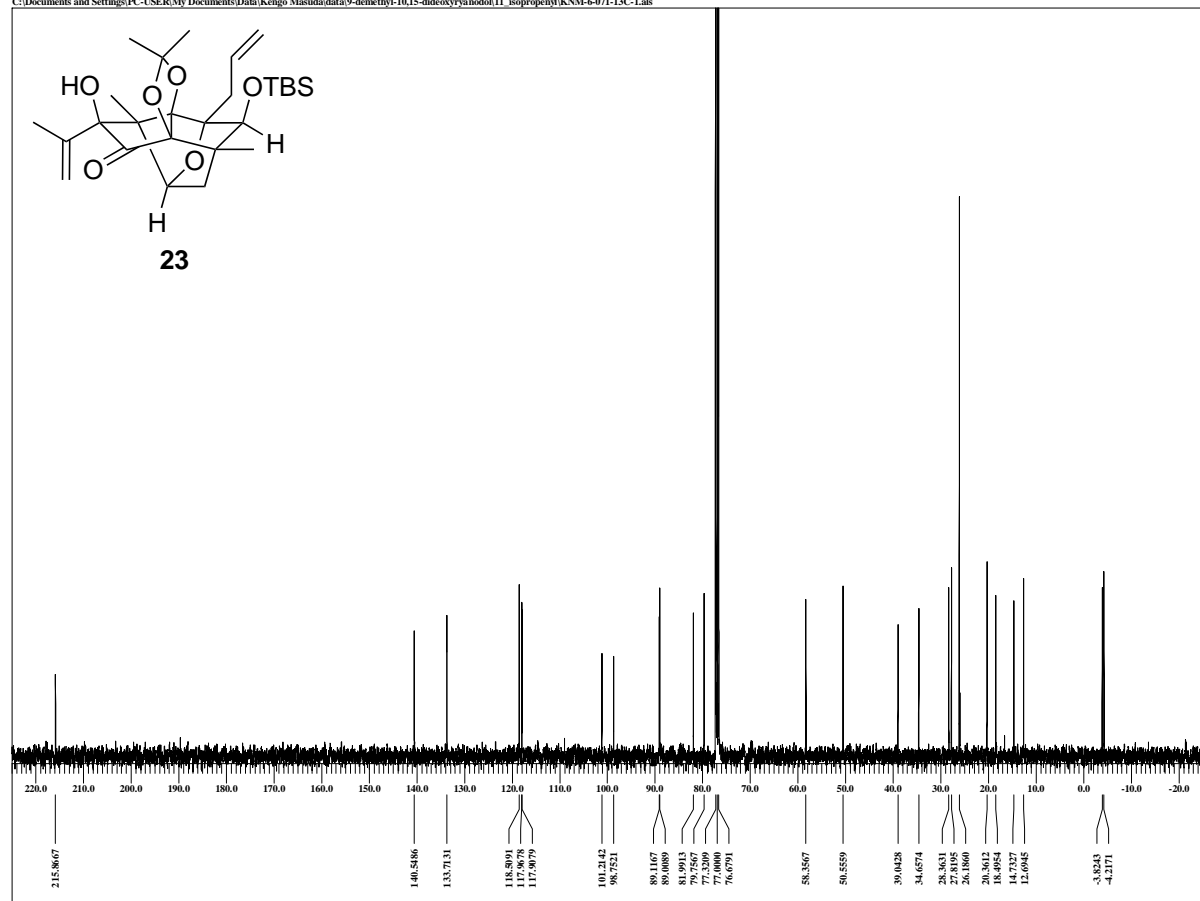


```

FILE KNM-6-071-1H.lab
COMT KNM-6-071-1H
DATIM 12-09-2012 11:50:11
MENUF
OBNUC 1H
OFR 395.88 MHz
OFRFQ 395.88 MHz
OBSET 6.28 KHz
OBFN 0.87 Hz
PW1 6.38 usec
DEADT 0.00 usec
PREDL 0.00000 msec
IWT 1.0000 sec
POINT 13107
SPO 13107
TIMES 5
DUMMY 1
FREQU 5938.15 Hz
FLT 30000 Hz
DELAY 16.68 usec
ACQTM 2.2073 sec
PD 2.0000 sec
SCANS 5
ADBIT 16
RGAIN 36
BF 0.01 Hz
T1 0.00
T2 0.00
T3 100.00
T4 100.00
EXMOD single_pulse.ex2
EXPCM
IRNUC 1H
IRF 395.88 MHz
IRSET 6.28 KHz
IRFN 0.87 Hz
IRFPW 115 usec
IRATN 79
DFILE KNM-6-071-1H.lab
SF 13.20 KHz
LKSET 13.20 KHz
LKFN 75.7 Hz
LKLEV 0
LGAIN 0
LKPIS 0
LKSG 0
CSPED 0 Hz
FILDC
FILDF
CTEMP 23.5 c
SLVNT CDCL3
EXREF 7.26 ppm
    
```

KNM-6-071-13C

C:\Documents and Settings\PC-USER\My Documents\Data\Kengo Masuda\data\9-demethyl-10,15-dideoxyryanodol\11_isopropenyl\KNM-6-071-13C-1.lab

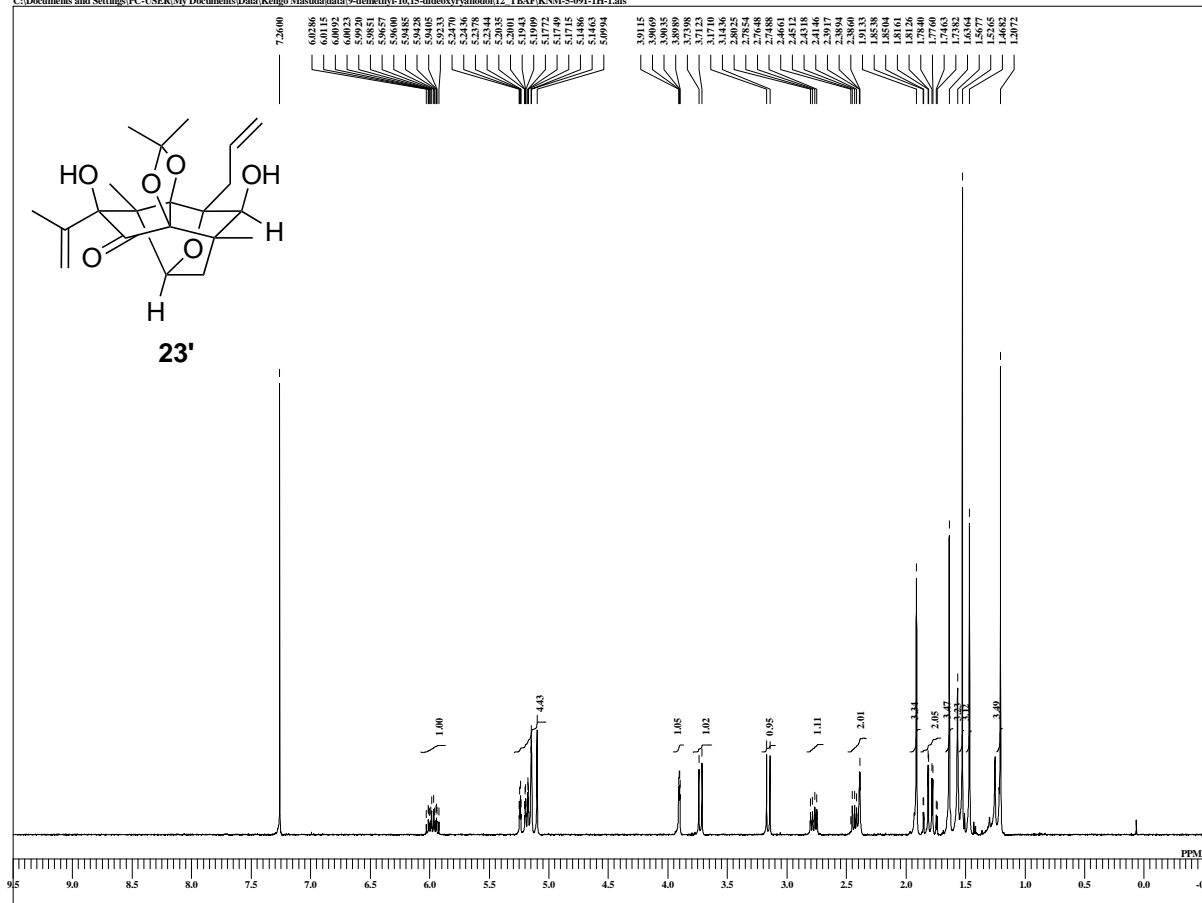


```

FILE KNM-6-071-13C-1.lab
COMT KNM-6-071-13C
DATIM 12-09-2012 12:02:09
MENUF
OBNUC 13C
OFR 99.55 MHz
OFRFQ 99.55 MHz
OBSET 5.13 KHz
OBFN 0.98 Hz
PW1 3.25 usec
DEADT 0.00 usec
PREDL 0.00000 msec
IWT 1.0000 sec
POINT 104856
SPO 104856
TIMES 99
DUMMY 4
FREQU 24999.62 Hz
FLT 125000 Hz
DELAY 20.50 usec
ACQTM 1.0486 sec
PD 5.0000 sec
SCANS 99
ADBIT 16
RGAIN 60
BF 1.00 Hz
T1 0.00
T2 0.00
T3 100.00
T4 100.00
EXMOD single_pulse_dec
EXPCM
IRNUC 13C
IRF 99.55 MHz
IRSET 5.13 KHz
IRFN 0.87 Hz
IRFPW 115 usec
IRATN 79
DFILE KNM-6-071-13C-1.lab
SF 13.20 KHz
LKSET 13.20 KHz
LKFN 75.7 Hz
LKLEV 0
LGAIN 0
LKPIS 0
LKSG 0
CSPED 0 Hz
FILDC
FILDF
CTEMP 23.8 c
SLVNT CDCL3
EXREF 77.00 ppm
    
```

KNM-5-091-1H

C:\Documents and Settings\PC-USER\My Documents\Data\Kengo Masuda\data\9-demethyl-10,15-dideoxyryanodol\12_TBAFKNM-5-091-1H-1als

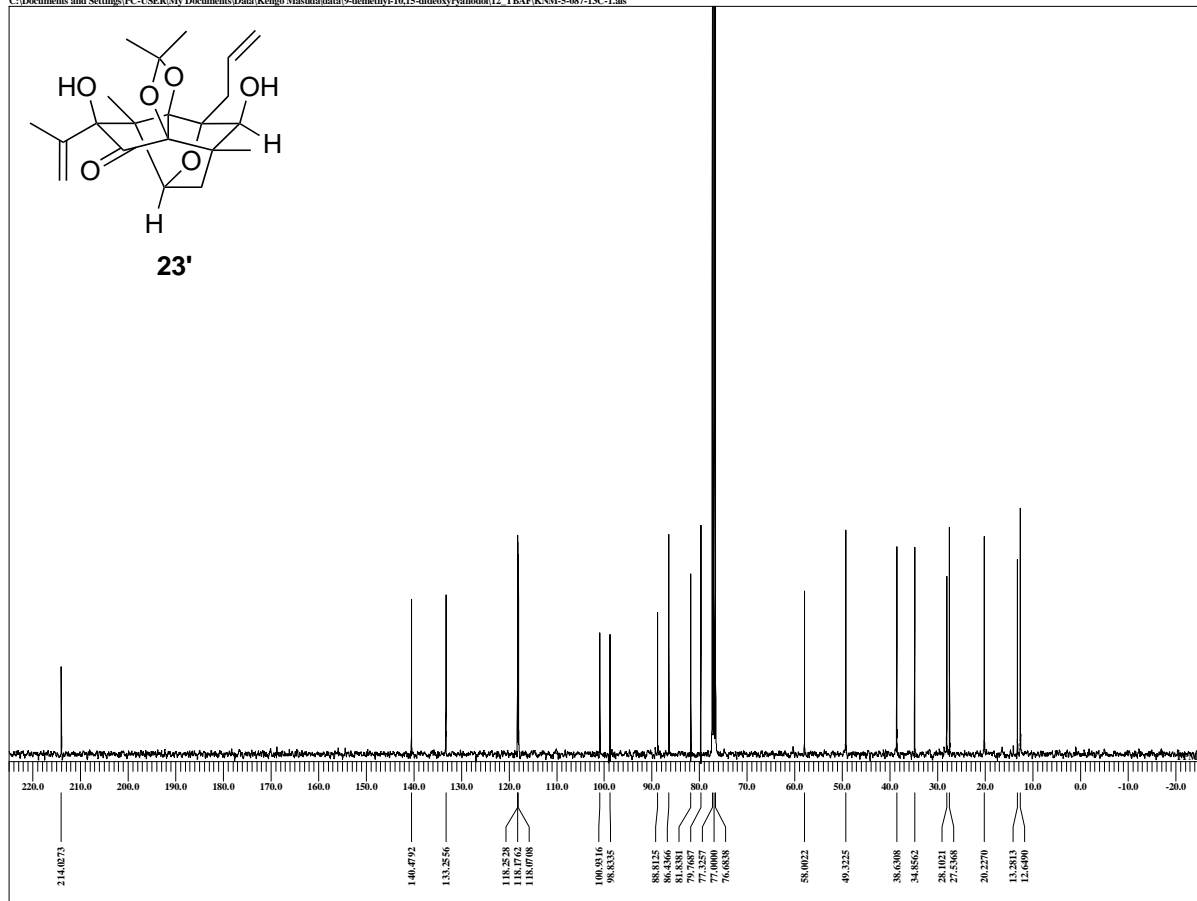


```

DFILE KNM-5-091-1H-1als
COMNT KNM-5-091-1H
DATIM 20-06-2012 20:15:33
MENEF
OBNUC 1H
OFR 395.88 MHz
OFRFQ 395.88 MHz
OBSET 6.28 KHz
OBFN 0.87 Hz
PWI 6.38 usec
DEADT 0.00 usec
PREDL 0.00000 msec
IWT 1.0000 sec
POINT 13107
SPO 13107
TIMES 8
DUMMY 1
FREQU 5938.15 Hz
FLT 38000 Hz
DELAY 16.68 usec
ACQTM 2.2073 sec
PD 2.0000 sec
SCANS 8
ADBIT 16
RGAIN 44
BF 0.01 Hz
T1 0.00
T2 0.00
T3 90.00
T4 100.00
EXMOD single_pulse.exe2
EXPCM
IRNUC 1H
IRF 395.88 MHz
IRSET 6.28 KHz
IRFN 0.87 Hz
IRPW 115 usec
IRATN 79
DFILE KNM-5-091-1H-1als
SF 13.20 KHz
LKSET 13.20 KHz
LKFN 75.7 Hz
LKLEV 0
LGAIN 0
LKPHS 0
LKSG 0 Hz
CSPED 0 Hz
FILDC
FILDF
CTEMP 24.6 c
SOLNT CDCL3
XREF 7.26 ppm
    
```

KNM-5-087-13C

C:\Documents and Settings\PC-USER\My Documents\Data\Kengo Masuda\data\9-demethyl-10,15-dideoxyryanodol\12_TBAFKNM-5-087-13C-1als

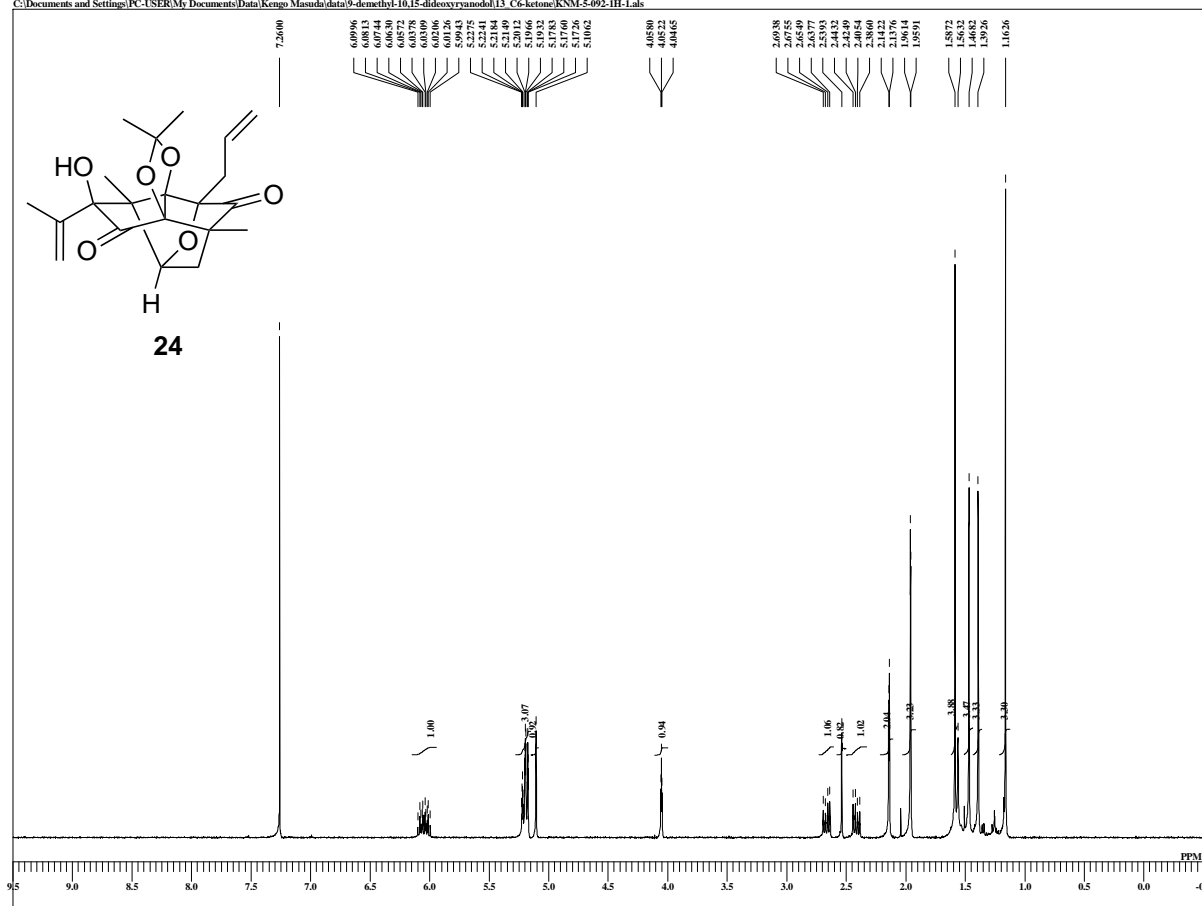


```

DFILE KNM-5-087-13C-1als
COMNT KNM-5-087-13C
DATIM 19-06-2012 21:27:15
MENEF
OBNUC 13C
OFR 99.55 MHz
OFRFQ 99.55 MHz
OBSET 5.13 KHz
OBFN 0.98 Hz
PWI 3.25 usec
DEADT 0.00 usec
PREDL 0.00000 msec
IWT 1.0000 sec
POINT 26214
SPO 26214
TIMES 69
DUMMY 4
FREQU 24999.62 Hz
FLT 125000 Hz
DELAY 20.50 usec
ACQTM 1.0486 sec
PD 5.0000 sec
SCANS 69
ADBIT 16
RGAIN 60
BF 1.00 Hz
T1 0.00
T2 0.00
T3 90.00
T4 100.00
EXMOD single_pulse_dec
EXPCM
IRNUC 13C
IRF 99.55 MHz
IRSET 5.13 KHz
IRFN 0.87 Hz
IRPW 115 usec
IRATN 79
DFILE KNM-5-087-13C-1als
SF 13.20 KHz
LKSET 13.20 KHz
LKFN 75.7 Hz
LKLEV 0
LGAIN 0
LKPHS 0
LKSG 0 Hz
CSPED 0 Hz
FILDC
FILDF
CTEMP 24.7 c
SOLNT CDCL3
XREF 77.00 ppm
    
```

KNM-5-092-1H

C:\Documents and Settings\PC-USER\My Documents\Data\Kengo Masuda\data\9-demethyl-10,15-dideoxyryanodol\13_C6-ketone\KNM-5-092-1H-1.ac

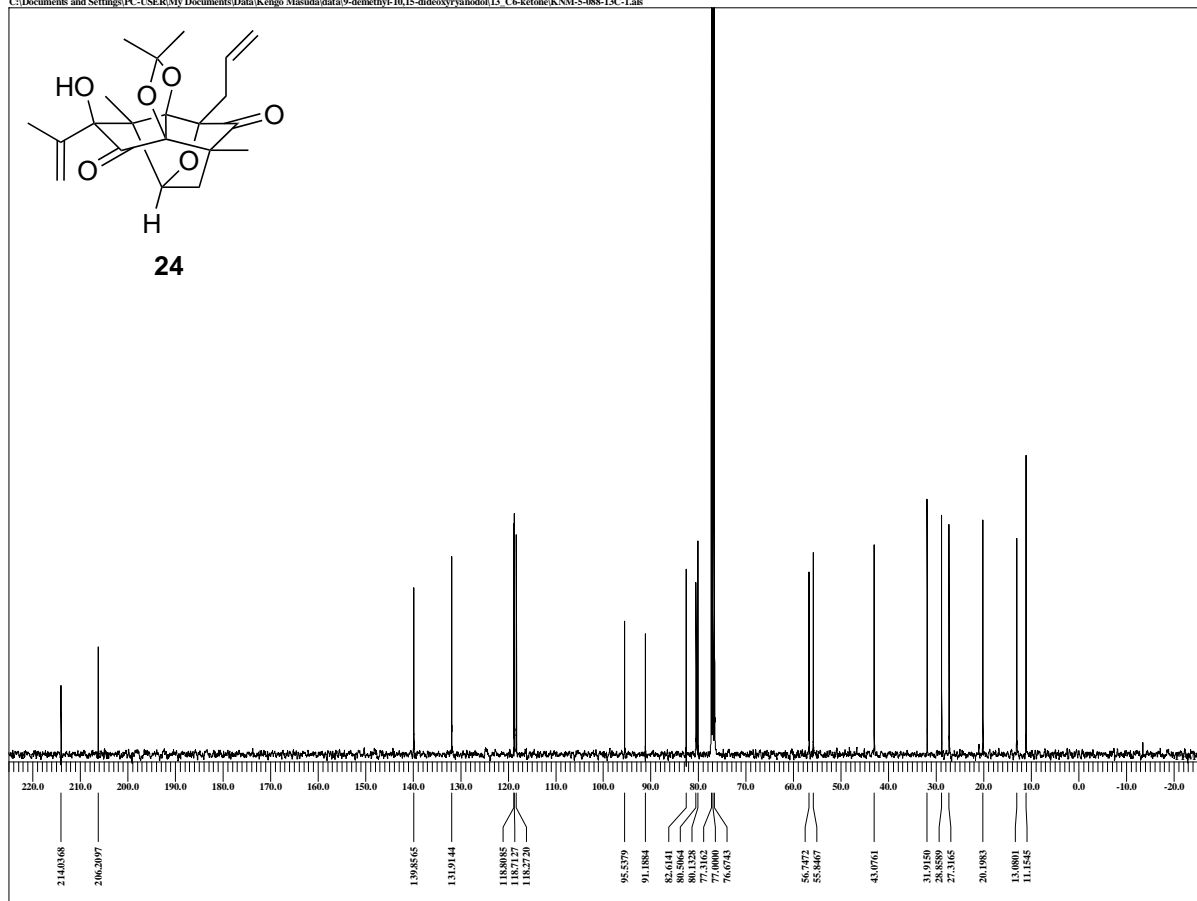


```

DEFILE KNM-5-092-1H-1.ac
COMNT KNM-5-092-1H
DATIM 21-06-2012 15:29:47
MENEF
OBNUC 1H
OFR 395.88 MHz
OBRFQ 395.88 MHz
OBSET 6.28 KHz
OBFN 0.87 Hz
PW1 6.38 usec
DEADT 0.00 usec
PREDL 0.00000 msec
IWT 1.0000 sec
POINT 13107
SFO 13107
TIMES 8
DUMMY 1
FREQU 5938.15 Hz
FLT 38000 Hz
DELAY 16.68 usec
ACQTM 2.2073 sec
PD 2.0000 sec
SCANS
ADBIT 16
RGAIN 44
BF 0.01 Hz
TI 0.00
T2 0.00
T3 90.00
T4 100.00
EXMOD single_pulse.ex2
EXPCM
IRNUC 1H
IFR 395.88 MHz
IRSET 6.28 KHz
IRFN 0.87 Hz
IRFPW 115 usec
IRATN 79
DEFILE KNM-5-092-1H-1.ac
SF
LKSET 13.20 KHz
LKFN 75.7 Hz
LKLEV 0
LGAIN 0
LKPHS 0
LKSG 0
CSPED 0 Hz
FILDC
FILDF
CTEMP 24.7 c
SLVNT CDCL3
XNREF 7.26 ppm
    
```

KNM-5-088-13C

C:\Documents and Settings\PC-USER\My Documents\Data\Kengo Masuda\data\9-demethyl-10,15-dideoxyryanodol\13_C6-ketone\KNM-5-088-13C-1.ac

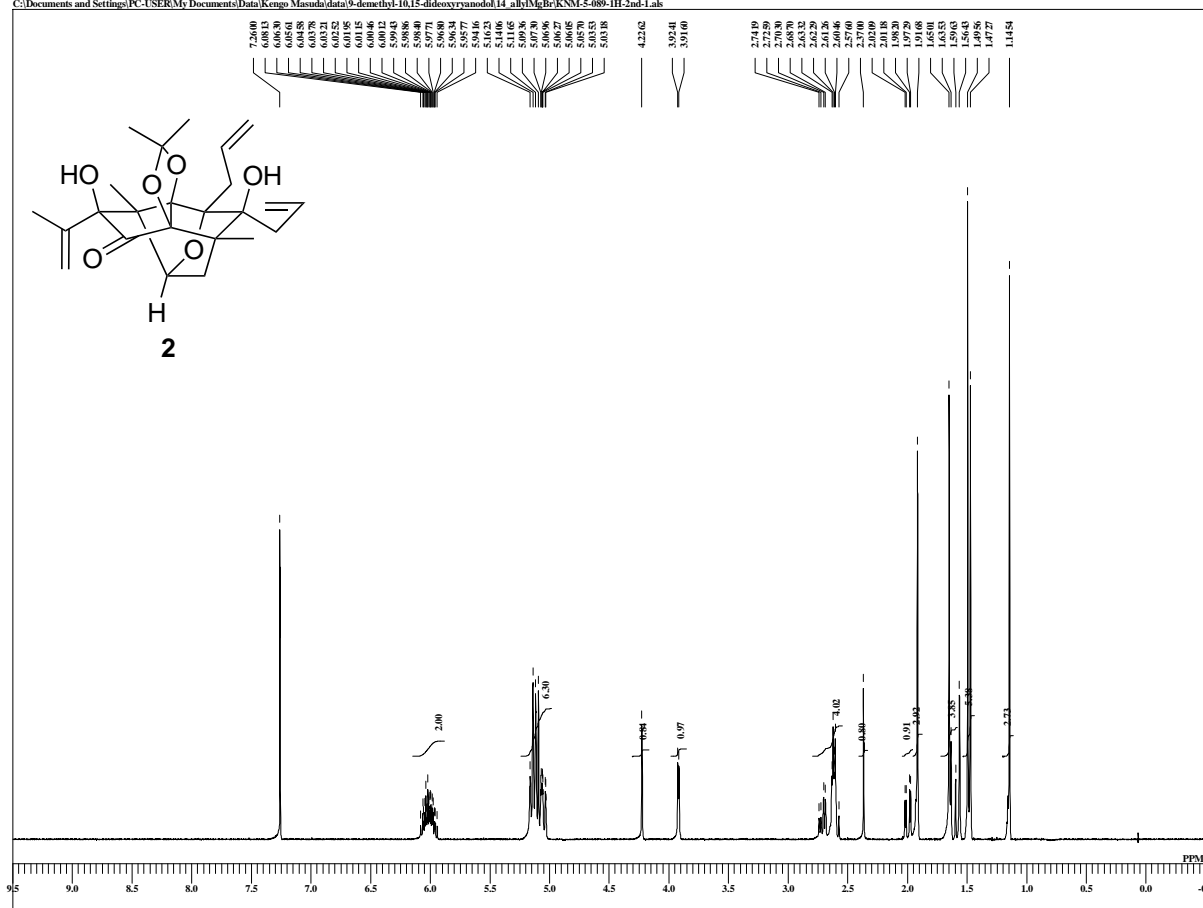


```

DEFILE KNM-5-088-13C-1.ac
COMNT KNM-5-088-13C
DATIM 20-06-2012 14:48:43
MENEF
OBNUC 13C
OFR 99.55 MHz
OBRFQ 99.55 MHz
OBSET 5.13 KHz
OBFN 0.98 Hz
PW1 3.25 usec
DEADT 0.00 usec
PREDL 0.00000 msec
IWT 1.0000 sec
POINT 26214
SFO 26214
TIMES 74
DUMMY 4
FREQU 24999.62 Hz
FLT 125000 Hz
DELAY 20.50 usec
ACQTM 1.0486 sec
PD 5.0000 sec
SCANS
ADBIT 16
RGAIN 60
BF 1.00 Hz
TI 0.00
T2 0.00
T3 90.00
T4 100.00
EXMOD single_pulse_dec
EXPCM
IRNUC 13C
IFR 99.55 MHz
IRSET 5.13 KHz
IRFN 0.87 Hz
IRFPW 115 usec
IRATN 79
DEFILE KNM-5-088-13C-1.ac
SF
LKSET 13.20 KHz
LKFN 75.7 Hz
LKLEV 0
LGAIN 0
LKPHS 0
LKSG 0
CSPED 0 Hz
FILDC
FILDF
CTEMP 25.0 c
SLVNT CDCL3
XNREF 77.00 ppm
    
```

KNM-5-092-1H

C:\Documents and Settings\PC-USER\My Documents\Data\Kengo Masuda\data\9-demethyl-10,15-dideoxyryanodol 14_allylMgBr\KNM-5-089-1H-2nd-1-Lab

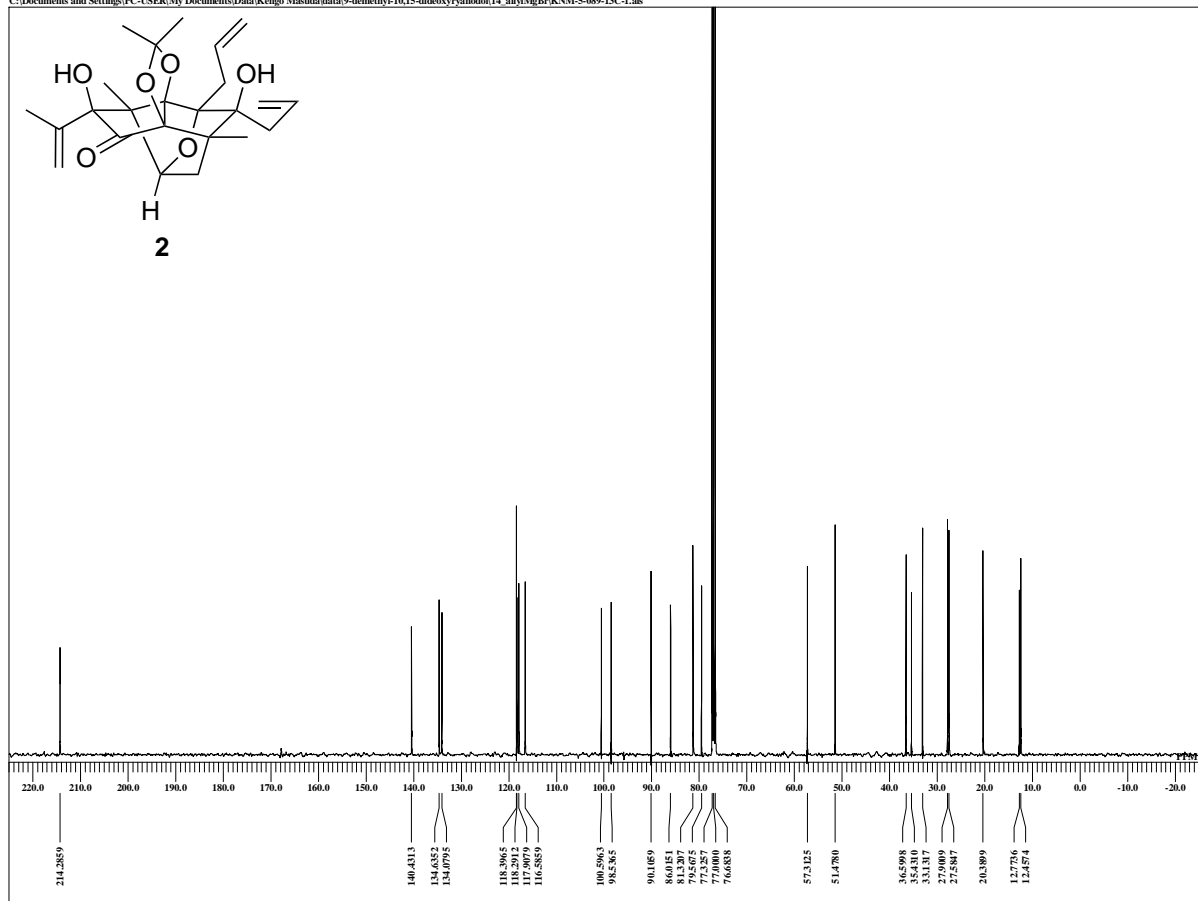


```

DFILE KNM-5-089-1H-2nd-1-Lab
COMENT KNM-5-092-1H
DATIM 21-06-2012 15:25:06
MENEF
OBNUC 1H
OFR 395.88 MHz
OBFREQ 395.88 MHz
OBSET 6.28 KHz
OBFIN 0.87 Hz
PW1 6.38 usec
DEADT 0.00 usec
PREDL 0.00000 msec
IWT 1.0000 sec
POINT 13107
SFO 13107
TIMES 8
DUMMY 1
FREQU 5938.15 Hz
FLT 30000 Hz
DELAY 16.68 usec
ACQTM 2.2073 sec
PD 2.0000 sec
SCANS 8
ADBIT 16
RGAIN 42
BF 0.01 Hz
T1 0.00
T2 0.00
T3 90.00
T4 100.00
EXMOD single_pulse.ex2
EXPCM
IRNUC 1H
IFR 395.88 MHz
IRSET 6.28 KHz
IRFIN 0.87 Hz
IRPW 115 usec
IRATN 79
DFILE KNM-5-089-1H-2nd-1-Lab
SE
LKSET 13.20 KHz
LKFIN 75.7 Hz
LKLEV 0
LGAIN 0
LKPHS 0
LKSG 0
CSPED 0 Hz
FLDC
FLDF
CTEMP 24.5 c
SLVNT CDCl3
XREF 7.26 ppm
    
```

KNM-5-091-13C

C:\Documents and Settings\PC-USER\My Documents\Data\Kengo Masuda\data\9-demethyl-10,15-dideoxyryanodol 14_allylMgBr\KNM-5-089-13C-1-Lab

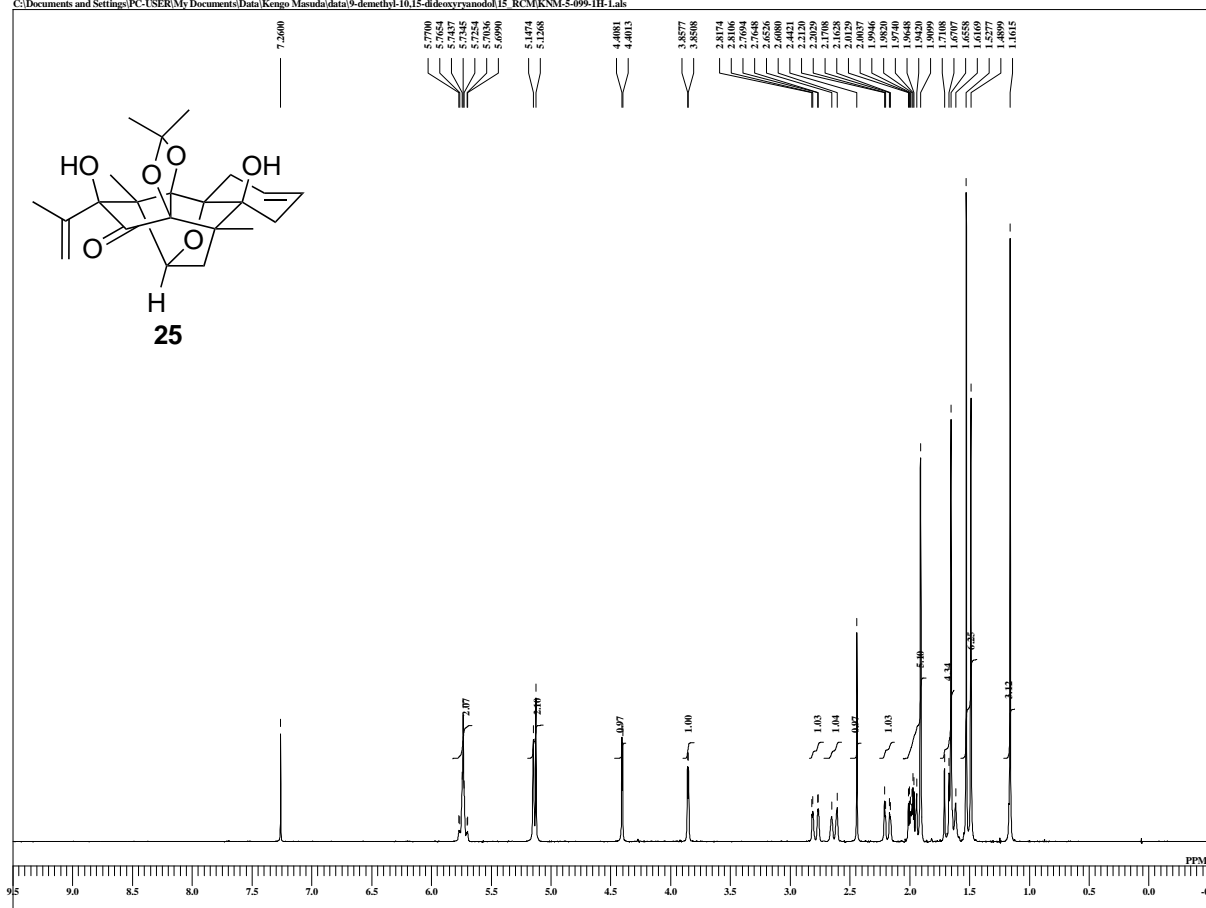


```

DFILE KNM-5-089-13C-1-Lab
COMENT KNM-5-091-13C
DATIM 21-06-2012 10:08:56
MENEF
OBNUC 13C
OFR 99.55 MHz
OBFREQ 99.55 MHz
OBSET 5.13 KHz
OBFIN 0.98 Hz
PW1 3.25 usec
DEADT 0.00 usec
PREDL 0.00000 msec
IWT 1.0000 sec
POINT 26214
SFO 26214
TIMES 29
DUMMY 4
FREQU 24999.62 Hz
FLT 125000 Hz
DELAY 20.50 usec
ACQTM 1.0486 sec
PD 5.5000 sec
SCANS 29
ADBIT 16
RGAIN 60
BF 1.00 Hz
T1 0.00
T2 0.00
T3 90.00
T4 100.00
EXMOD single_pulse_dec
EXPCM
IRNUC 13C
IFR 99.55 MHz
IRSET 6.28 KHz
IRFIN 0.87 Hz
IRPW 115 usec
IRATN 79
DFILE KNM-5-089-13C-1-Lab
SE
LKSET 13.20 KHz
LKFIN 75.7 Hz
LKLEV 0
LGAIN 0
LKPHS 0
LKSG 0
CSPED 0 Hz
FLDC
FLDF
CTEMP 24.2 c
SLVNT CDCl3
XREF 77.00 ppm
    
```

KNM-5-099-1H

C:\Documents and Settings\PC-USER\My Documents\Data\Kengo Masuda\data\9-demethyl-10,15-dideoxyryanodol.15 RCMKNM-5-099-1H-1.Lab

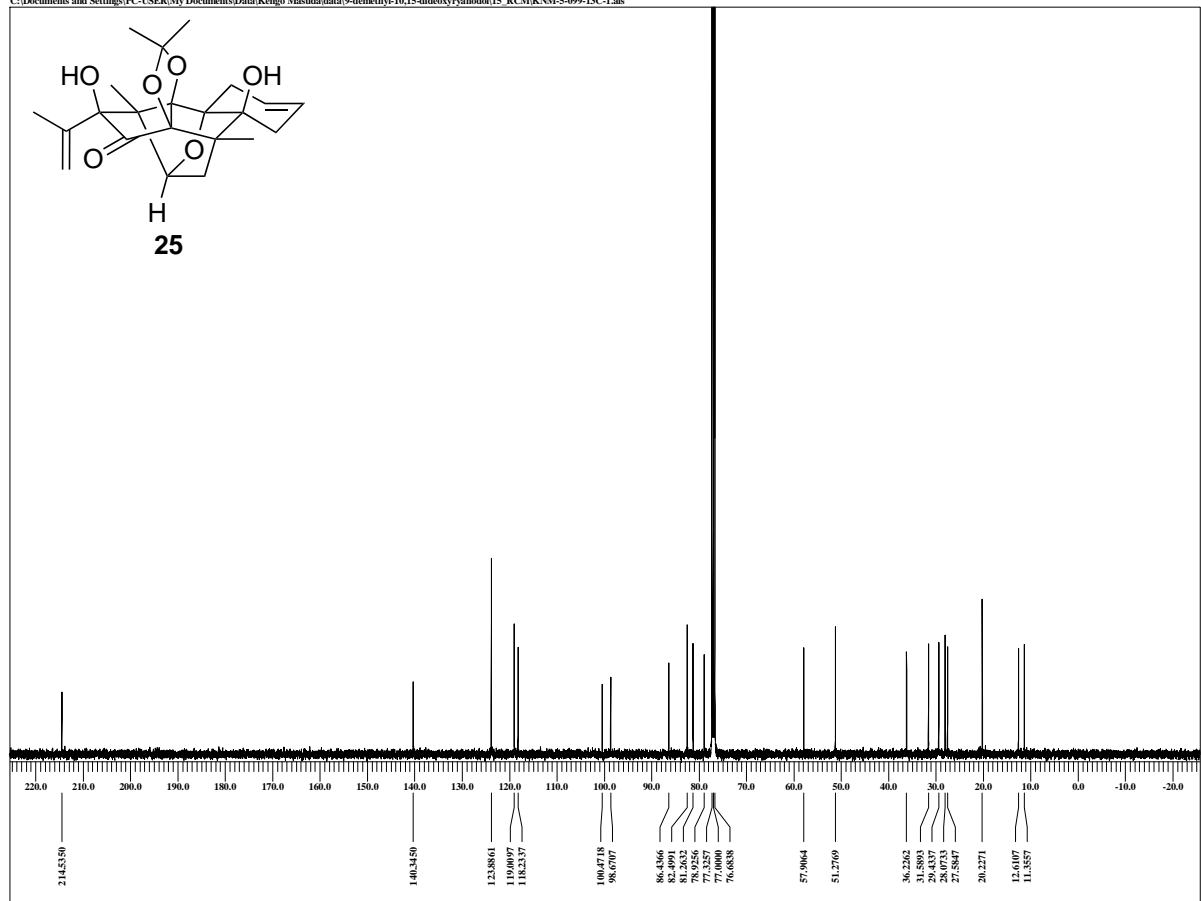


```

DFILE KNM-5-099-1H-1.Lab
COMNT KNM-5-099-1H
DATIM 24-06-2012 16:42:18
MENUE
OBNUC 1H
OFR 395.88 MHz
OBFREQ 395.88 MHz
OBSET 6.28 KHz
OBFEN 0.87 Hz
PWI 6.38 usec
DEADT 0.00 usec
PREDL 0.00000 msec
IWT 1.0000 sec
POINT 13107
SFO 13107
TIMES 8
DUMMY 1
FREQU 5938.15 Hz
FLT 30000 Hz
DELAY 16.68 usec
ACQTM 2.2073 sec
PD 2.0000 sec
SCANS 8
ADBIT 16
RGAIN 36
BF 0.01 Hz
T1 0.00
T2 0.00
T3 90.00
T4 100.00
EXMOD single_pulse.ec2
EXPCM
IRNUC 1H
IFR 395.88 MHz
IRSET 6.28 KHz
IRFEN 0.87 Hz
IRRPW 115 usec
IRATN 79
DFILE KNM-5-099-1H-1.Lab
SF 13.20 KHz
LKSET 13.20 KHz
LKFN 75.7 Hz
LKLEV 0
LGAIN 0
LKPHS 0
LKSG 0
CSPED 0 Hz
FILDC
FILDF
CTEMP 23.1 c
SLVNT CDCL3
EXREF 7.26 ppm
    
```

KNM-5-099-13C

C:\Documents and Settings\PC-USER\My Documents\Data\Kengo Masuda\data\9-demethyl-10,15-dideoxyryanodol.15 RCMKNM-5-099-13C-1.Lab

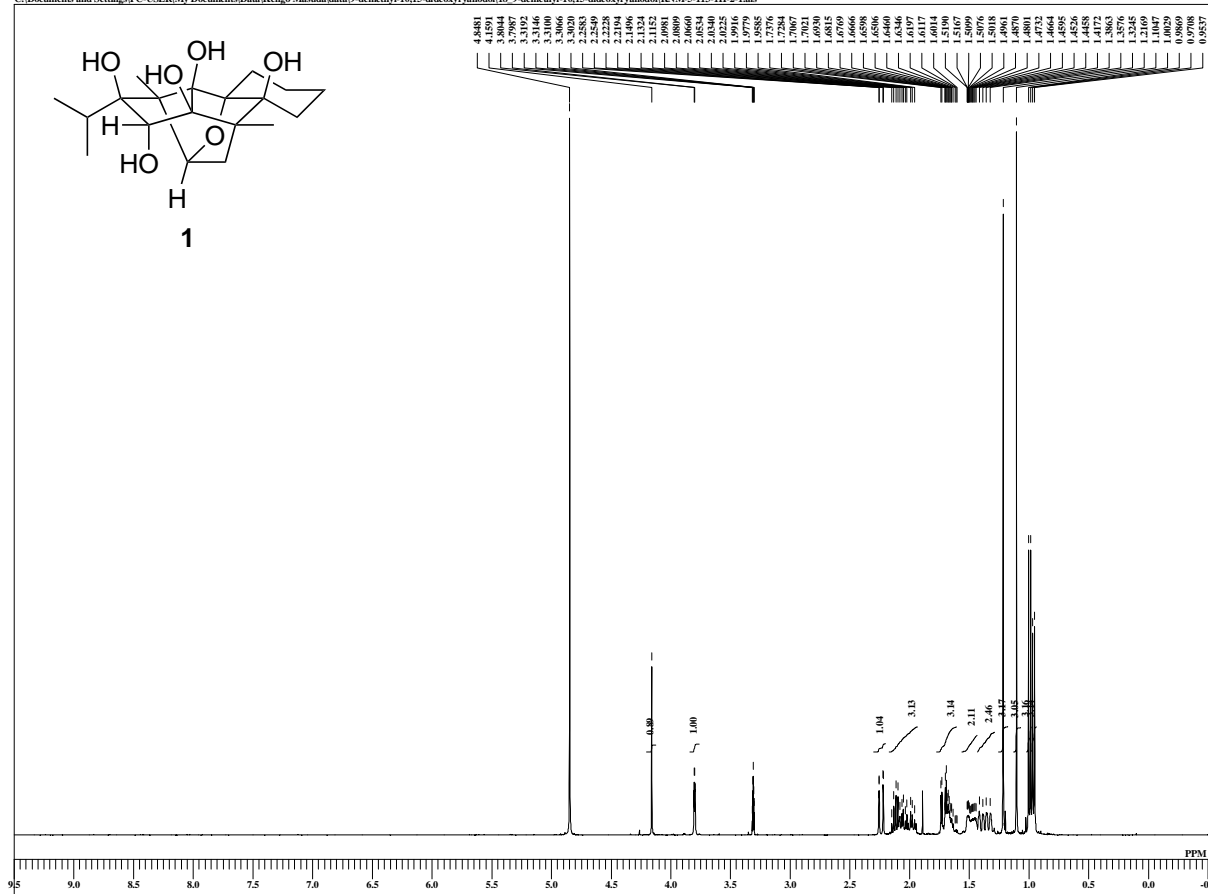


```

DFILE KNM-5-099-13C-1.Lab
COMNT KNM-5-099-13C
DATIM 24-06-2012 17:50:39
MENUE
OBNUC 13C
OFR 99.55 MHz
OBFREQ 99.55 MHz
OBSET 5.13 KHz
OBFEN 0.98 Hz
PWI 3.25 usec
DEADT 0.00 usec
PREDL 0.00000 msec
IWT 1.0000 sec
POINT 26214
SFO 26214
TIMES 500
DUMMY 4
FREQU 24999.62 Hz
FLT 125000 Hz
DELAY 20.50 usec
ACQTM 1.0486 sec
PD 7.0000 sec
SCANS 500
ADBIT 16
RGAIN 60
BF 1.00 Hz
T1 0.00
T2 0.00
T3 90.00
T4 100.00
EXMOD single_pulse_dec
EXPCM
IRNUC 13C
IFR 99.55 MHz
IRSET 5.13 KHz
IRFEN 0.87 Hz
IRRPW 115 usec
IRATN 79
DFILE KNM-5-099-13C-1.Lab
SF 13.20 KHz
LKSET 13.20 KHz
LKFN 75.7 Hz
LKLEV 0
LGAIN 0
LKPHS 0
LKSG 0
CSPED 0 Hz
FILDC
FILDF
CTEMP 23.1 c
SLVNT CDCL3
EXREF 77.00 ppm
    
```

KNM-5-113-1H-2

C:\Documents and Settings\PC-USER\My Documents\Data\Kengo Masuda\data\9-demethyl-10,15-dideoxyryanodol\9-demethyl-10,15-dideoxyryanodol\KNM-5-113-1H-2-Lab

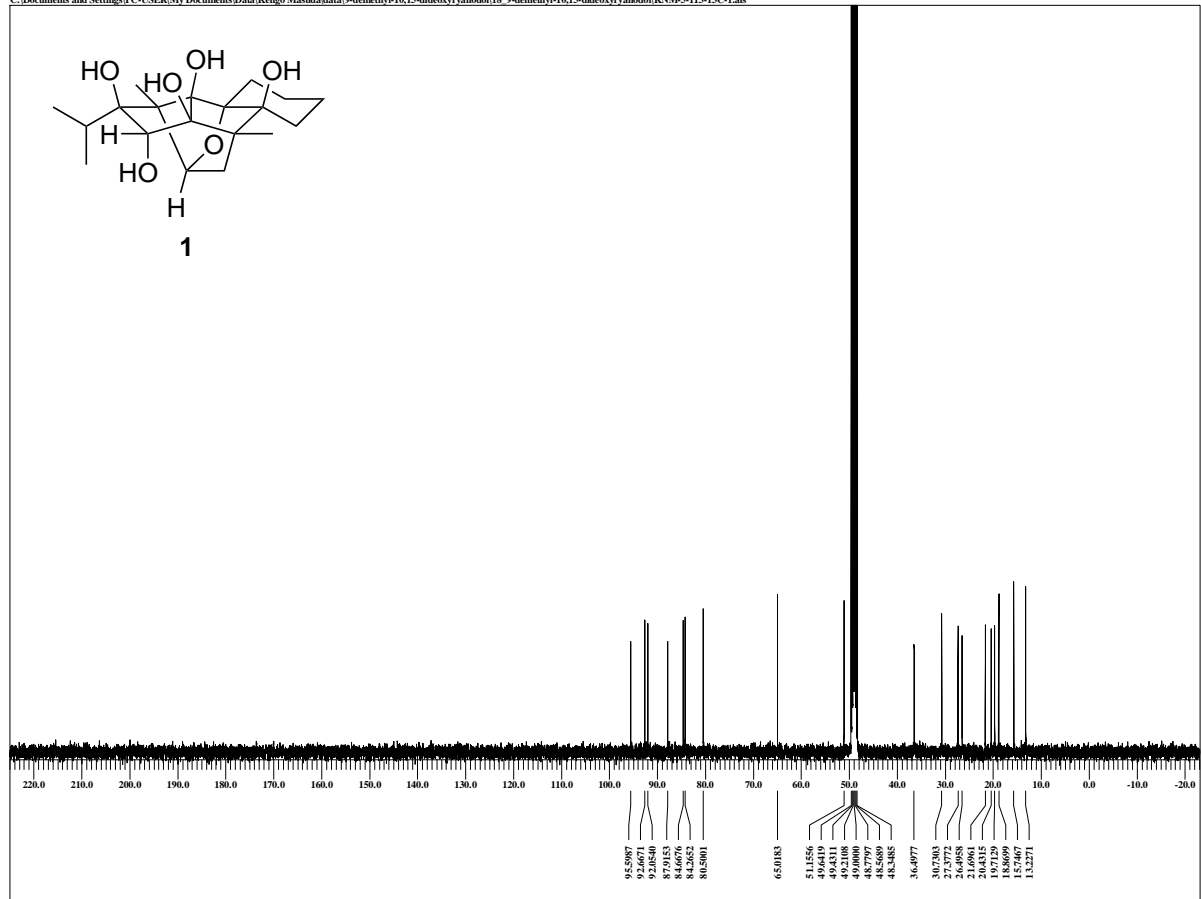


```

DFILE KNM-5-113-1H-2-Lab
COMNT KNM-5-113-1H-2
DATIM 07-07-2012 18:14:26
NAME 1
OBNUC 1H
OFR 395.88 MHz
OBFREQ 395.88 MHz
OBSET 6.28 KHz
OBFIN 0.87 Hz
PW1 6.38 usec
DEADT 0.00 usec
PREDL 0.00000 msec
IWT 1.0000 sec
POINT 13107
SFO 395.88 MHz
TIMES 8
DUMMY 1
FREQU 5938.15 Hz
FLT 30000 Hz
DELAY 16.68 usec
ACQTM 2.2073 sec
PD 2.0000 sec
SCANS 8
ADBIT 16
RGAIN 42
BF 0.01 Hz
T1 0.00
T2 0.00
T3 90.00
T4 100.00
EXMOD single_pulse.ex2
EXPCM
IRNUC 1H
IR 395.88 MHz
IRSET 6.28 KHz
IRFIN 0.87 Hz
IRFPW 147 usec
IRATN 79
DFILE KNM-5-113-1H-2-Lab
SE
LKSET 13.00 KHz
LKFIN 35.6 Hz
LKLEV 0
LGAIN 0
LKPHS 0
LKSG 0
CSPED 0 Hz
FLDC
FLDF
CTEMP 25.0 c
SLVNT CD3OD
EXREF 3.31 ppm
    
```

KNM-5-113-13C

C:\Documents and Settings\PC-USER\My Documents\Data\Kengo Masuda\data\9-demethyl-10,15-dideoxyryanodol\9-demethyl-10,15-dideoxyryanodol\KNM-5-113-13C-Lab



```

DFILE KNM-5-113-13C-Lab
COMNT KNM-5-113-13C
DATIM 07-07-2012 03:12:14
NAME 1
OBNUC 13C
OFR 99.55 MHz
OBFREQ 99.55 MHz
OBSET 5.13 KHz
OBFIN 0.98 Hz
PW1 3.25 usec
DEADT 0.00 usec
PREDL 0.00000 msec
IWT 1.0000 sec
POINT 26214
SFO 99.55 MHz
TIMES 1000
DUMMY 4
FREQU 24999.62 Hz
FLT 125000 Hz
DELAY 20.50 usec
ACQTM 1.0486 sec
PD 5.0000 sec
SCANS 1000
ADBIT 16
RGAIN 60
BF 1.00 Hz
T1 0.00
T2 0.00
T3 90.00
T4 100.00
EXMOD single_pulse_dec
EXPCM
IRNUC 1H
IR 395.88 MHz
IRSET 6.28 KHz
IRFIN 0.87 Hz
IRFPW 115 usec
IRATN 79
DFILE KNM-5-113-13C-Lab
SE
LKSET 13.00 KHz
LKFIN 35.6 Hz
LKLEV 0
LGAIN 0
LKPHS 0
LKSG 0
CSPED 0 Hz
FLDC
FLDF
CTEMP 24.6 c
SLVNT CD3OD
EXREF 49.00 ppm
    
```