

Phosphine Oxide Functional Group Based Three-Station Molecular Shuttle

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SUPPORTING INFORMATION

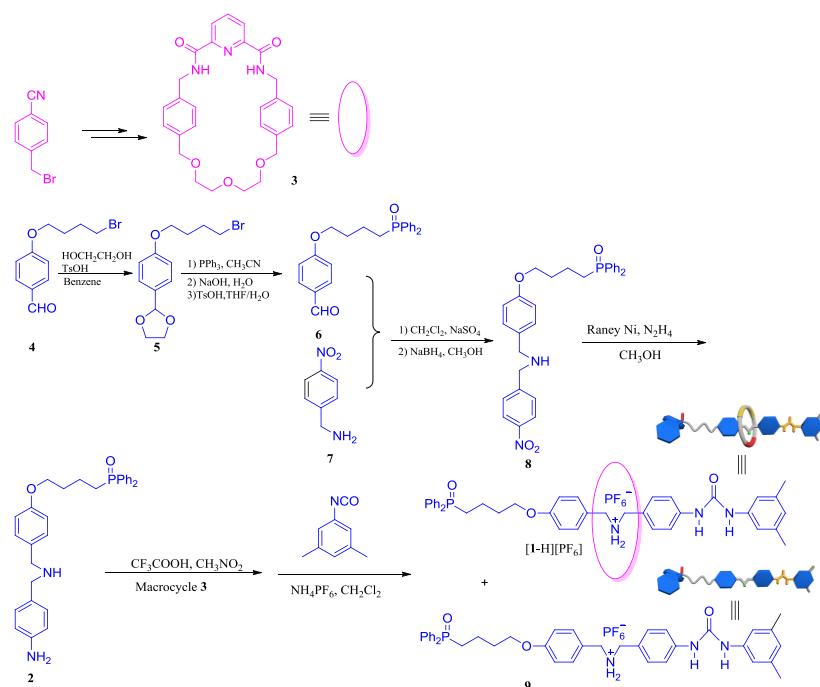
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1. Materials and methods

All reactions were performed in open atmosphere unless otherwise stated. All reagents, unless otherwise indicated, were obtained from commercial sources. Anhydrous CH_2Cl_2 and CH_3CN were obtained by distillation from CaH_2 under N_2 atmosphere. Melting points (M.p.) were determined using a Focus X-4 apparatus and were not corrected. All yields were given as isolated yields. NMR spectra were recorded on a Bruker DPX 300 MHz or 400 MHz spectrometer with internal standard tetramethylsilane (TMS) and solvent signals as internal references, and the chemical shifts (δ) were expressed in ppm and J values were given in Hz. 2D COSY and 2D ROESY experiments were performed on a Bruker DPX 400 MHz spectrometer. Low-resolution electrospray ionization mass spectra (LR-ESI-MS) were obtained on Finnigan MatTSQ 7000 instruments. High-resolution electrospray ionization mass spectra (HR-ESI-MS) were recorded on an Agilent 6540Q-TOF LCMS equipped with an electrospray ionization (ESI) probe operating in positive-ion mode with direct infusion.

2. Experimental procedures



Scheme S1. Synthesis of [2]rotaxane [1-H][PF₆]

2-(4-(4-bromobutoxy)phenyl)-1,3-dioxolane 5: *p*-Toluenesulfonic acid monohydrate (0.10 g, 0.5 mmol) as a catalyst was added to a solution of **4** (2.57 g, 10 mmol) and ethylene glycol (6.20 g, 100 mmol) in benzene (60 mL) and then the mixture was refluxed for 24 h in an apparatus fitted with a Dean-Stark condenser under N₂. The reaction mixture was subsequently poured into aqueous sodium hydrogen carbonate solution, extracted with diethyl ether (2 × 100 mL) and the combined extracts were washed with aqueous sodium hydrogen carbonate solution and water, dried over MgSO₄ and concentrated under vacuum to give dioxolane **5** as yellow oil. The crude dioxolane was used directly for the next step without further purification.

4-(4-(diphenylphosphoryl)butoxy)benzaldehyde 6^{S1}: A mixture of **5** (2.88 g, 9.6 mmol) and triphenylphosphine (2.51 g, 9.6 mmol) in anhydrous acetonitrile (60 mL) was heated at 80 °C for 24 h and then the solvent was evaporated under reduced pressure. Sodium hydroxide (1.92 g, 48 mmol) was added to the residue in water (30 mL) and the reaction mixture was refluxed for 12 h. 2 N HCl was added to acidify the solution until pH = 2, and extracted with chloroform (3 × 50 mL). The combined extracts were washed with water, dried over MgSO₄ and concentrated under vacuum. *p*-Toluenesulfonic acid monohydrate (0.10 g, 0.5 mmol) as a catalyst was added to a solution of the residue in THF (30 mL) and water (30 mL) then the mixture was refluxed for 12 h and cooled to RT. After THF was removed under reduced pressure, saturated sodium hydrogen carbonate solution (30 mL) was added. The mixture was extracted with dichloromethane (3 × 50 mL). The combined extracts were washed with water, dried over MgSO₄ and concentrated under vacuum. The residue was purified by silica-gel column chromatography (CH₂Cl₂/CH₃OH, 100:1) to afford **6** as a white solid (2.72 g, 72% yield in two steps). M.p. 86-87 °C; The ¹H & ¹³C NMR spectra of **6** are shown in Fig. S3-S4. ¹H NMR (400 MHz, CDCl₃): δ = 9.87 (s, 1H, CHO), 7.81 (d, 2H, *J* = 8.8 Hz, ArH), 7.77-7.72 (m, 4H, ArH), 7.55-7.44 (m, 6H, ArH), 6.93 (d, 2H, *J* = 8.7 Hz, ArH), 4.03 (t, 2H, *J* = 6.0 Hz, OCH₂), 2.38-2.31 (m, 2H, O=PCH₂), 1.98-1.91 (m, 2H, CH₂), 1.90-1.79 (m, 2H, CH₂); ¹³C NMR (100 MHz, CDCl₃): δ = 190.6, 163.8, 132.8 (d, ¹J (C, P) = 98 Hz, ArCP), 131.8, 131.7 (d, ⁴J (C, P) = 2 Hz, ArCH), 130.6 (d, ³J (C, P) = 9 Hz, ArCH), 129.7, 128.6 (d, ²J (C, P) = 12 Hz, ArCH), 114.6, 67.4, 29.9 (d, ²J (C, P) = 14 Hz, O=PCH₂CH₂), 29.2 (d, ¹J (C, P) = 72 Hz, O=PCH₂), 18.3 (d, ³J (C, P) = 3 Hz, O=PCH₂CH₂CH₂); ³¹P NMR (160 MHz, CDCl₃): δ = 32.2; HR-MS (ESI): m/z calcd for [6+Na]⁺ C₂₃H₂₃NaO₅P⁺, 401.1283, found

401.1311.

Nitro compound 8: A mixture of **6** (0.88 g, 2.3 mmol), Na₂SO₄ (3.27 g, 23 mmol) and (4-nitrophenyl)methanamine **7** (0.35 g, 2.3 mmol) in anhydrous dichloromethane (25 mL) was stirred at RT for 24 h. Then Na₂SO₄ was filtered off. The solvent was evaporated under reduced pressure. NaBH₄ (0.27 g, 7.0 mmol) was added to the residue in methanol (30 mL) under ice-bath. Then the ice-bath was removed and the mixture was stirred for 5 h. After removing most of the solvent, 50 mL H₂O was added to quench the reaction. The mixture was extracted with dichloromethane (3 × 50 mL). The combined extracts were washed with water, dried over MgSO₄ and concentrated under vacuum. The residue was purified by silica-gel column chromatography (CH₂Cl₂/CH₃OH, 100:1) to afford **8** as yellow oil (1.03 g, 87%). The ¹H & ¹³C NMR spectra of **8** are shown in Fig. S5-S6. ¹H NMR (400 MHz, CDCl₃): δ = 8.17 (d, 2H, *J* = 8.7 Hz, ArH), 7.76-7.70 (m, 4H, ArH), 7.53 (d, 2H, *J* = 8.8 Hz, ArH), 7.52-7.44 (m, 6H, ArH), 7.22 (d, 2H, *J* = 8.6 Hz, ArH), 6.80 (d, 2H, *J* = 8.6 Hz, ArH), 3.93 (t, 2H, *J* = 6.0 Hz, OCH₂), 3.89 (s, 2H, CH₂), 3.75 (s, 2H, CH₂), 2.37-2.31 (m, 2H, O=PCH₂), 2.05 (br, 1H, NH), 1.91-1.78 (m, 4H, CH₂); ¹³C NMR (100 MHz, CDCl₃): δ = 158.1, 148.3, 147.0, 133.0 (d, ¹J(C, P) = 98 Hz, ArCP), 131.9, 131.7 (d, ⁴J(C,P) = 2 Hz, ArCH), 130.8 (d, ³J(C,P) = 9 Hz, ArCH), 129.3, 128.7, 128.7 (d, ²J(C,P) = 11 Hz, ArCH), 123.6, 114.5, 67.1, 56.2, 52.6, 30.3 (d, ²J(C,P) = 14 Hz, O=PCH₂CH₂), 29.4 (d, ¹J(C,P) = 72 Hz, O=PCH₂), 18.5 (d, ³J(C,P) = 3 Hz, O=PCH₂CH₂CH₂); ³¹P NMR (160 MHz, CDCl₃): δ = 32.6; HR-MS (ESI): m/z calcd for [8+Na]⁺ C₃₀H₃₁N₂NaO₄P⁺, 537.1919, found 537.1923.

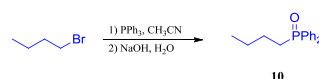
Amine 2: A mixture of **8** (1.03 g, 2 mmol) and Raney Ni (Cat.) in methanol was heated to reflux at 80 °C. Then N₂H₄ H₂O (1.8 mL, 40 mmol) was added. The reaction was refluxed for 6 h. Then the solution was cooled to RT. The catalyst was filtered off. The solvent was evaporated under reduced pressure. The residue was dissolved in CH₂Cl₂ (30 mL), and the organic phase was washed with water, dried over MgSO₄ and concentrated under vacuum. The residue was purified by silica-gel column chromatography (CH₂Cl₂/CH₃OH, 20:1) to afford **2** as a yellow solid (0.87 g, 90%). M.p. 56-58 °C; The ¹H & ¹³C NMR spectra of **2** are shown in Fig. S7-S8. ¹H NMR (400 MHz, CD₃CN): δ = 7.70-7.67 (m, 4H, ArH), 7.54-7.47 (m, 6H, ArH), 7.27 (d, 2H, *J* = 8.6 Hz, ArH), 7.10 (d, 2H, *J* = 8.4 Hz, ArH), 6.78 (d, 2H, *J* = 8.6 Hz, ArH), 6.58 (d, 2H, *J* = 8.4 Hz, ArH),

4.13 (br, 3H, NH), 3.90 (t, 2H, J = 6.3 Hz, OCH₂), 3.70 (s, 2H, CH₂), 3.65 (s, 2H, CH₂), 2.38-2.29 (m, 2H, O=PCH₂), 1.86-1.73 (m, 2H, CH₂), 1.66-1.55 (m, 2H, CH₂); ¹³C NMR (100 MHz, CDCl₃): δ = 158.5, 146.5, 132.8 (d, ¹J (C, P) = 98 Hz, ArCP), 131.8 (d, ⁴J (C, P) = 3 Hz, ArCH), 130.7 (d, ³J (C, P) = 9 Hz, ArCH), 130.5, 130.4, 128.7 (d, ²J (C, P) = 12 Hz, ArCH), 127.8, 125.0, 115.0, 114.5, 67.0, 50.8, 50.4, 30.2 (d, ²J (C, P) = 14 Hz, O=PCH₂CH₂), 29.3 (d, ¹J (C, P) = 72 Hz, O=PCH₂), 18.4 (d, ³J (C, P) = 4 Hz, O=PCH₂CH₂CH₂); ³¹P NMR (160 MHz, CDCl₃) : δ = 32.9; HR-MS (ESI): m/z calcd for [2+H]⁺ C₃₀H₃₄N₂O₂P⁺, 485.2358, found 485.2471.

[1-H][PF₆]^{S2}: Trifluoroacetic (37 μ L, 0.5 mmol) was added to a solution of amine **2** (0.24g, 0.5 mmol) and macrocycle **3**^{S3, S4} (0.24g, 0.5 mmol) in anhydrous CH₃NO₂. The mixture was stirred for 1 h. After 3,5-dimethylphenyl isocyanate (87 μ L, 0.6 mmol) was added, the resulting mixture was stirred at RT for 48 h and then the solvent was evaporated under reduced pressure. The residue was dissolved in CH₂Cl₂ (30 mL). Then saturated NH₄PF₆ solution (10 mL) was added. The mixture was stirred for 1 h and then water (20 mL) was added. The aqueous solution was extracted with CH₂Cl₂ (3 \times 50 mL). The combined extracts were washed with water, dried over MgSO₄ and concentrated under vacuum. The residue was purified by silica-gel column chromatography (CH₂Cl₂/CH₃OH, 20:1) to afford [2]Rotaxane [1-H][PF₆] as a white solid and dumbbell-shaped salt **9** as a yellow solid. [1-H][PF₆] (75 mg, 12%). M.p. 185-187 °C; The ¹H & ¹³C NMR spectra of [1-H][PF₆] are shown in Fig. S9-S10. ¹H NMR (400 MHz, CD₃CN/CDCl₃ (ν/ν = 1:1)) : δ = 9.01 (br, 2H, CONH), 8.43 (d, 2H, J = 7.8 Hz, ArH), 8.18 (t, 1H, J = 7.8 Hz, ArH), 7.76-7.71 (m, 4H, ArH), 7.56-7.49 (m, 7H, ArH and NHCONH), 7.35 (d, 2H, J = 8.5 Hz, ArH), 7.27 (br, 1H, NHCONH), 7.21 (br, 2H, NH₂), 7.08 (d, 4H, J = 7.8 Hz, ArH), 7.04 (s, 2H, ArH), 6.96 (d, 2H, J = 8.9 Hz, ArH), 6.94 (d, 2H, J = 9.3 Hz, ArH), 6.82 (d, 4H, J = 7.9 Hz, ArH), 6.67 (s, 1H, ArH), 6.65 (d, 2H, J = 8.7 Hz, ArH), 4.73-4.58 (m, 4H, ArCH₂), 4.24-4.19 (m, 4H, ArCH₂), 3.91 (t, 2H, J = 6.3 Hz, OCH₂), 3.65-3.57 (m, 4H, OCH₂), 3.31-3.21 (m, 4H, OCH₂), 2.50 (br, 4H, ArCH₂), 2.44 -2.37 (m, 2H, POCH₂), 2.38-2.29 (m, 2H, O=PCH₂), 2.25 (s, 6H, ArCH₃), 1.91-1.86 (m, 2H, CH₂), 1.81-1.72 (m, 2H, CH₂); ¹³C NMR (100 MHz, CDCl₃): δ = 164.0, 159.0, 153.6, 149.4, 141.4, 140.8, 138.9, 138.7, 134.4, 132.6 (d, ¹J (C, P) = 99 Hz, ArCP), 132.2, 130.9 (d, ³J (C, P) = 9, ArCH), 129.4, 129.2, 129.0 (d, ²J (C, P) = 12 Hz, ArCH), 128.5, 127.8, 125.5, 124.9, 124.2, 123.4, 118.3, 117.4, 114.2, 74.5, 70.5, 69.6, 67.2, 51.5, 50.5, 42.4, 30.4 (d, ²J (C, P)

= 14, POCH_2CH_2), 29.4 (d, 1J (C, P) = 71, $\text{O}=\text{PCH}_2$), 21.5, 18.6 (d, 3J (C, P) = 2, $\text{POCH}_2\text{CH}_2\text{CH}_2$); ^{31}P NMR (160 MHz, CD_3CN): δ = 30.6; LR-ESI-MS: calcd. for $[\mathbf{1}+\text{H}]^+$ $\text{C}_{66}\text{H}_{72}\text{N}_6\text{O}_8\text{P}^+$: 1107.51, found m/z = 1107.35; HR-MS (ESI): m/z calcd for $[\mathbf{1}+\text{H}]^+$ $\text{C}_{66}\text{H}_{72}\text{N}_6\text{O}_8\text{P}^+$, 1107.5144, found 1107.5152; $[\mathbf{1}+\text{Na}]^+$ $\text{C}_{66}\text{H}_{71}\text{N}_6\text{NaO}_8\text{P}^+$, 1129.4963, found 1129.4975.

9: 116 mg, 30%. M.p. 152-155 °C; The ^1H & ^{13}C NMR spectra of $[\mathbf{2}-\text{H}][\text{PF}_6]$ are shown in Fig. S11-S12. ^1H NMR (400 MHz, $\text{CD}_3\text{CN}/\text{CDCl}_3$ (ν/ν = 1:1)) : δ = 7.76 (br, 1H, NHCONH), 7.70-7.66 (m, 4H, ArH), 7.51-7.48 (m, 7H, ArH), 7.40 (d, 2H, J = 8.5 Hz, ArH), 7.29 (d, 2H, J = 8.6 Hz, ArH), 7.24 (d, 2H, J = 8.5 Hz, ArH), 7.04 (s, 2H, ArH), 6.81 (d, 2H, J = 8.5 Hz, ArH), 6.68 (s, 2H, ArH), 4.09 (br, 4H, ArCH_2), 3.92 (t, 2H, J = 5.9 Hz, OCH_2) 2.41-2.30 (m, 2H, $\text{O}=\text{PCH}_2$), 2.26 (s, 6H, ArCH_3), 1.89-1.82 (m, 2H, CH_2), 1.71-1.61 (m, 2H, CH_2); ^{13}C NMR (100 MHz, $\text{CD}_3\text{CN}/\text{CDCl}_3$ (ν/ν = 1:1)): δ = 160.7, 153.8, 141.8, 140.1, 139.5, 133.5, 133.2 (d, 4J (C, P) = 2 Hz), 133.0 (d, 1J (C, P) = 99 Hz), 132.7, 131.9, 131.6 (d, 3J (C, P) = 10 Hz), 129.9 (d, 2J (C, P) = 12 Hz), 125.4, 123.9, 119.7, 117.9, 115.6, 68.1, 51.8, 51.6, 30.7 (d, 2J (C, P) = 15 Hz), 28.9 (d, 1J (C, P) = 72 Hz), 21.5, 19.1 (d, 3J (C, P) = 4 Hz); ^{31}P NMR (160 MHz, CD_3CN): δ = 34.0; LR-ESI-MS: calcd. for $[\mathbf{9}+\text{H}]^+$ $\text{C}_{39}\text{H}_{43}\text{N}_3\text{O}_3\text{P}^+$ m/z 632.30, found 632.20; HR-MS (ESI): m/z calcd for $[\mathbf{9}+\text{H}]^+$ $\text{C}_{39}\text{H}_{43}\text{N}_3\text{O}_3\text{P}^+$, 632.3037, found 632.3064; $[\mathbf{9}+\text{Na}]^+$ $\text{C}_{39}\text{H}_{42}\text{N}_3\text{NaO}_3\text{P}^+$, 654.2856, found 654.2865.



Scheme S2. Synthesis of butyldiphenylphosphine oxide **10**

10^{S1}: A mixture of 1-bromo-butane (1.37 g, 10.0 mmol) and triphenylphosphine (2.621 g, 10.0 mmol) in anhydrous acetonitrile (60 mL) was heated at 80 °C for 24 h and then the solution was cooled to RT. After solvent was evaporated under reduced pressure, sodium hydroxide (2.0 g, 50 mmol) was added to the residue in water (30 mL) and the reaction mixture was refluxed for 12 h. 2 N HCl was added to acidify the solution until pH = 2, and extracted with chloroform (3 × 50 mL). The combined extracts were washed with water, dried over MgSO_4 and concentrated under vacuum. The residue was purified by silica-gel column chromatography ($\text{CH}_2\text{Cl}_2/\text{CH}_3\text{OH}$, 100:1) to afford **10** as a white solid (1.64 g, 64%). The ^1H NMR spectra of **10** are shown in Fig. S17. ^1H NMR (400 MHz, CDCl_3): δ = 7.79-7.73 (m, 4H, ArH), 7.52-7.49 (m, 6H, ArH), 2.38-2.31 (m, 2H,

O=PCH₂), 1.63-1.61 (m, 2H, CH₂), 1.46-1.41 (m, 2H, CH₂), 0.90 (t, 3H, CH₃); ³¹P NMR (160 MHz, CDCl₃): δ = 32.6; LR-ESI-MS: calcd. for [10+H]⁺ C₁₆H₂₀OP⁺: 259.12, found *m/z* = 259.10.

3. Dilution isotherm/Job plot for macrocycle 3 and phosphine oxide derivative 10 in CDCl₃

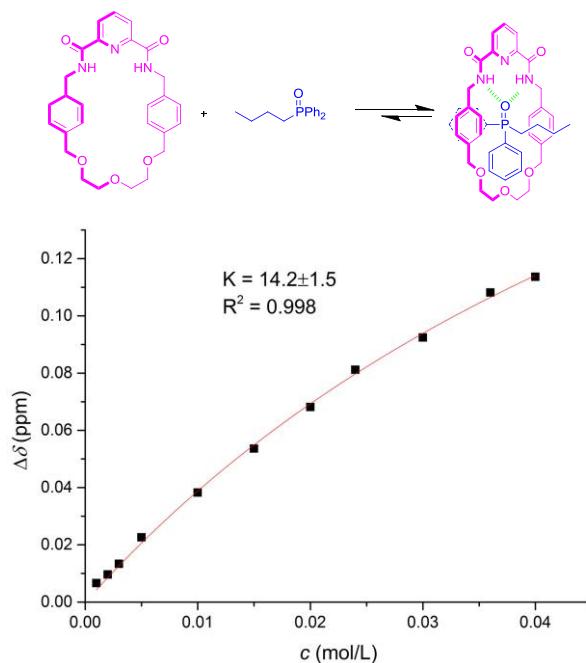


Fig. S1 The non-linear curve-fitting (NMR titrations) for the complexation of macrocycle 3 (host, 2 mM) with phosphine oxide derivative 10 (guest) in CDCl₃ at 298 K. The concentration of 10 was 1.0, 2.0, 3.0, 5.0, 10.0, 15.0, 18.0, 20.0, 24.0, 30.0, 36.0, 40.0 mM. Using the signal of free macrocycle 3 at δ 8.3489 as the reference.⁵⁵

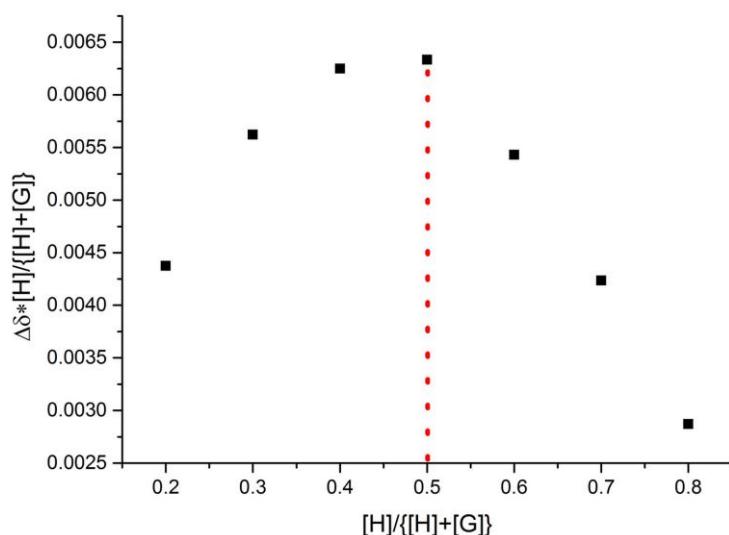


Fig. S2 The Job Plot (NMR titrations) for the complexation of macrocycle 3 (host) with phosphine oxide derivative 10 (guest) in CDCl₃ at 298 K. Using the signal of free macrocycle 3 at δ 7.8304 as the reference. $([H] + [G]) = 8$ mM.⁵⁵

4. ^1H NMR and ^{13}C NMR for new compounds

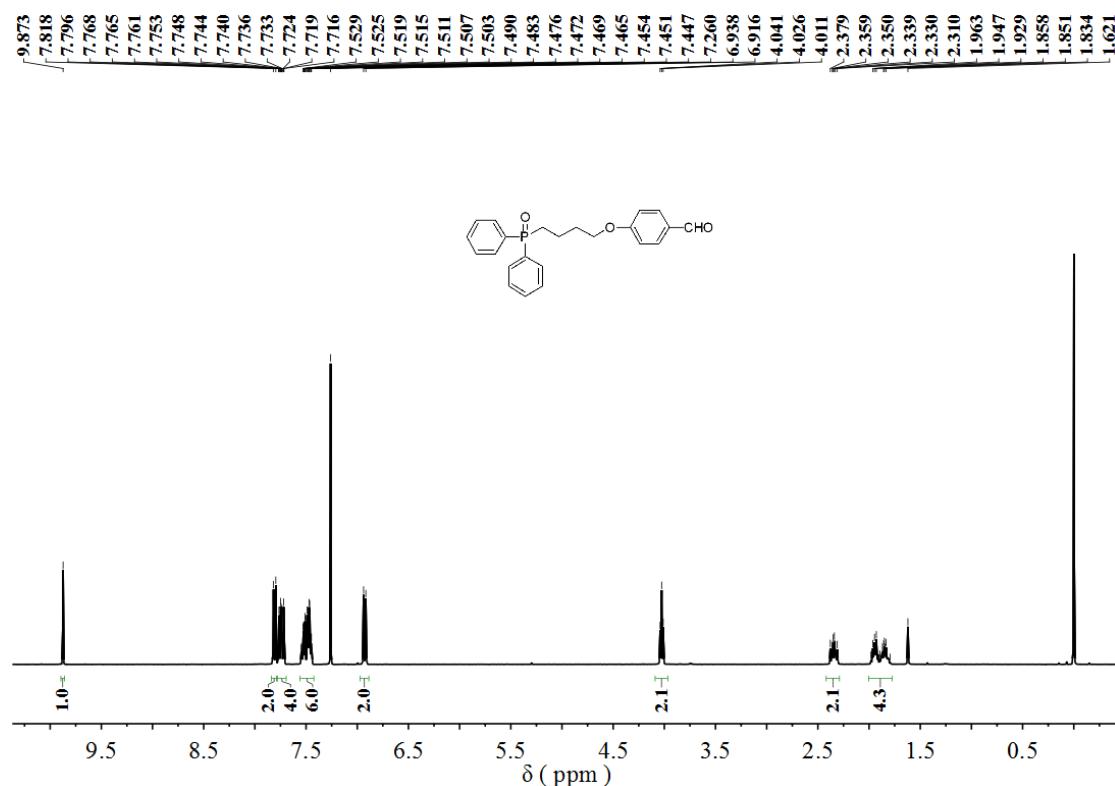


Fig. S3 ^1H NMR spectrum (CDCl_3 , 400 MHz, 298 K) of **6**.

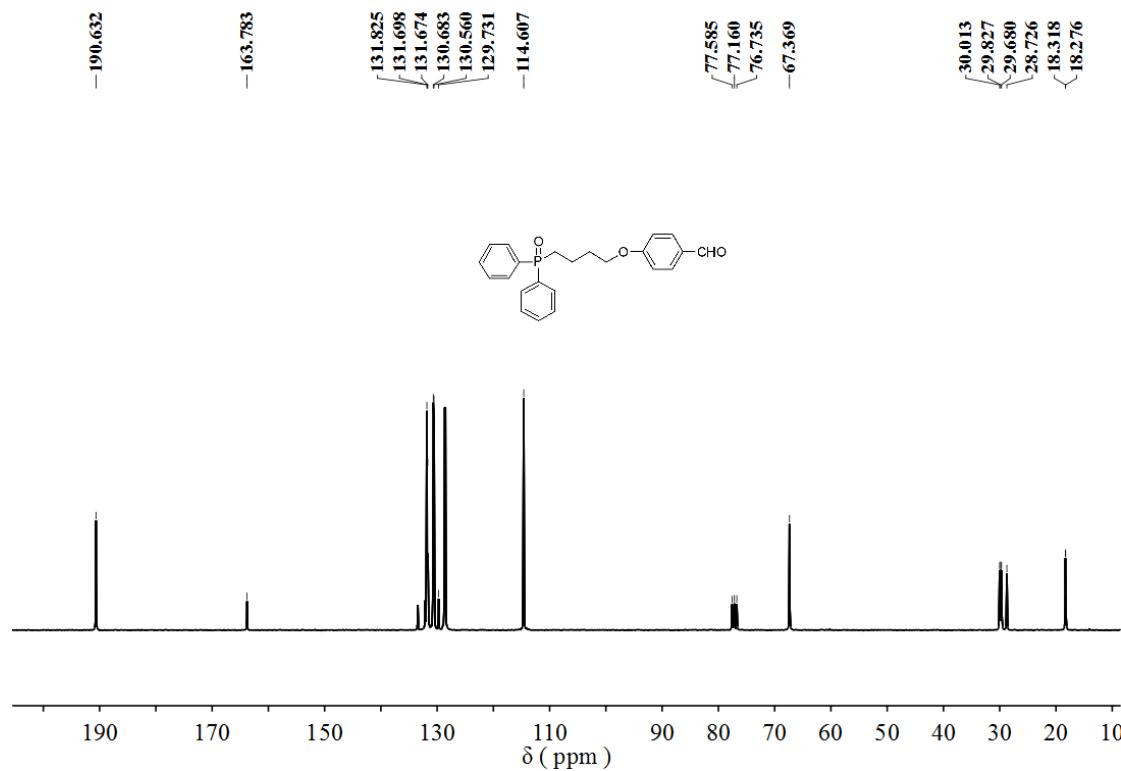


Fig. S4 ^{13}C NMR spectrum (CDCl_3 , 100 MHz, 298 K) of **6**.

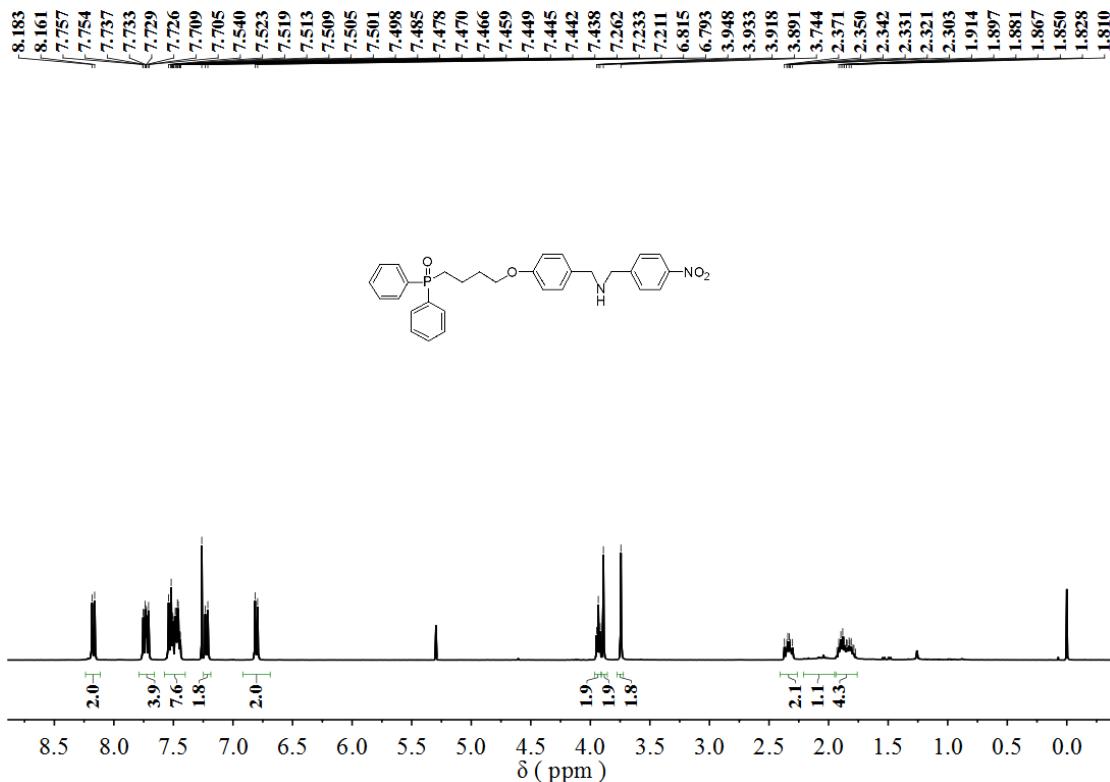


Fig. S5 ^1H NMR spectrum (CDCl_3 , 400 MHz, 298 K) of **8**.

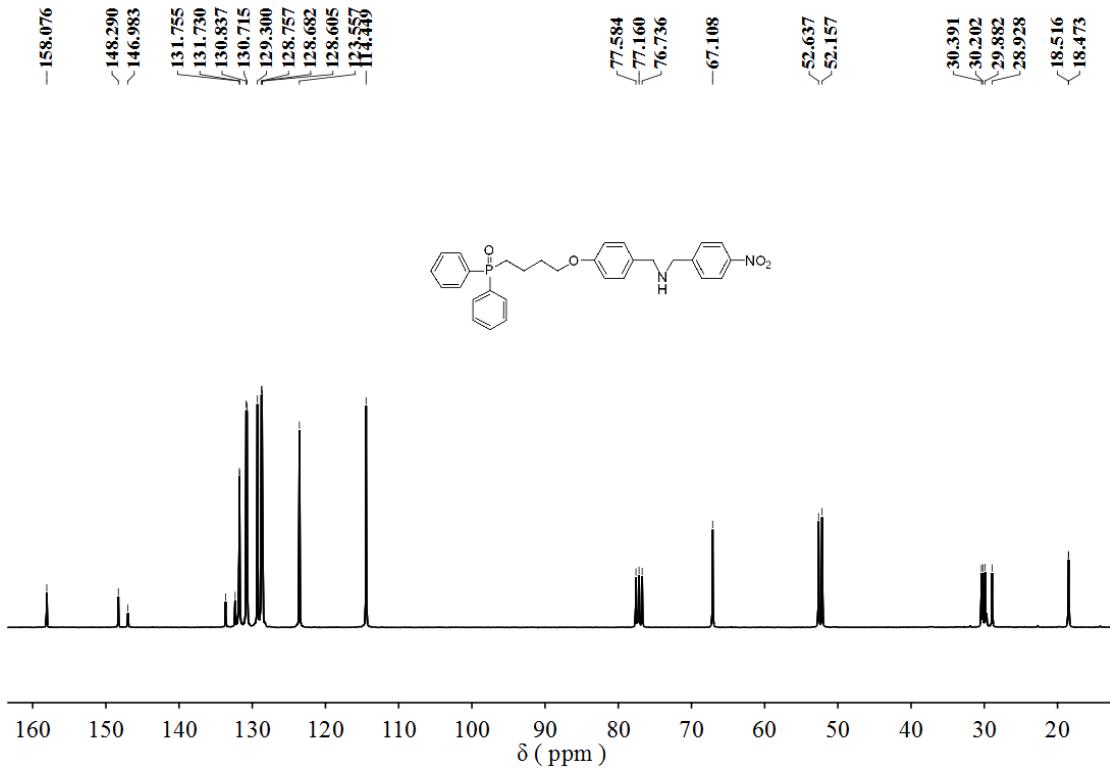


Fig. S6 ^{13}C NMR spectrum (CDCl_3 , 100 MHz, 298 K) of **8**.

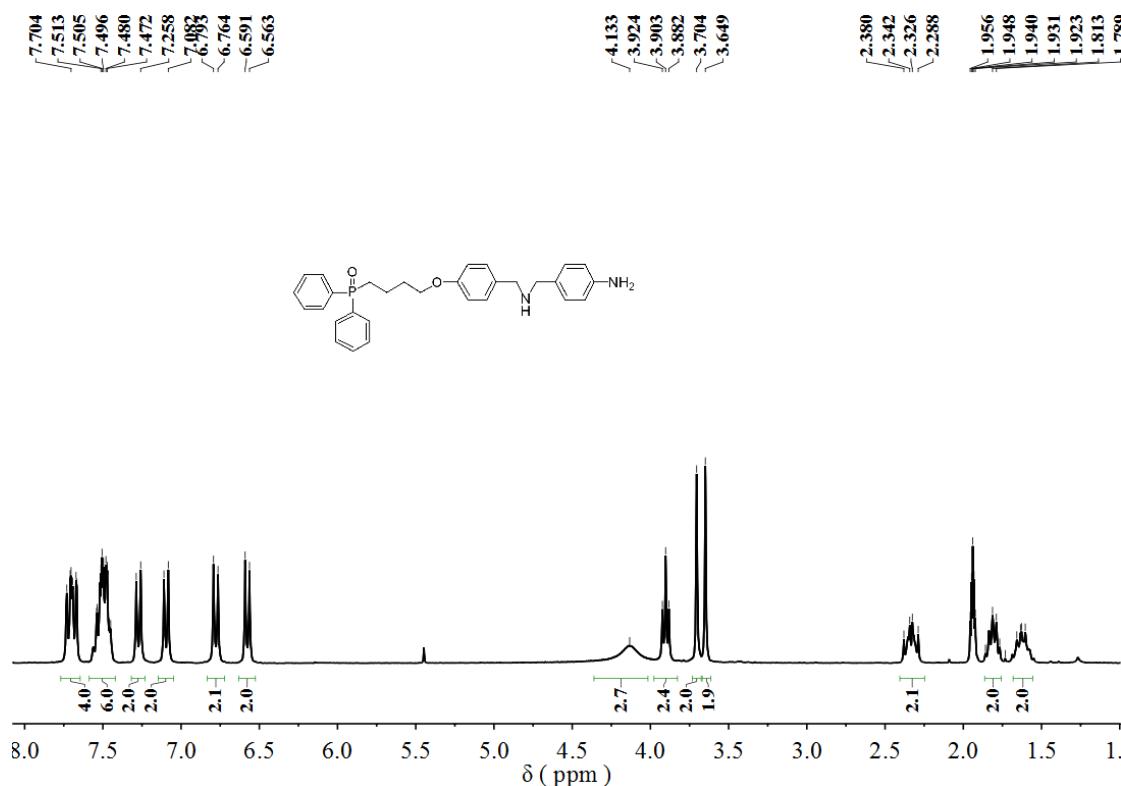


Fig. S7 ¹H NMR spectrum (CD₃CN, 400 MHz, 298 K) of 2.

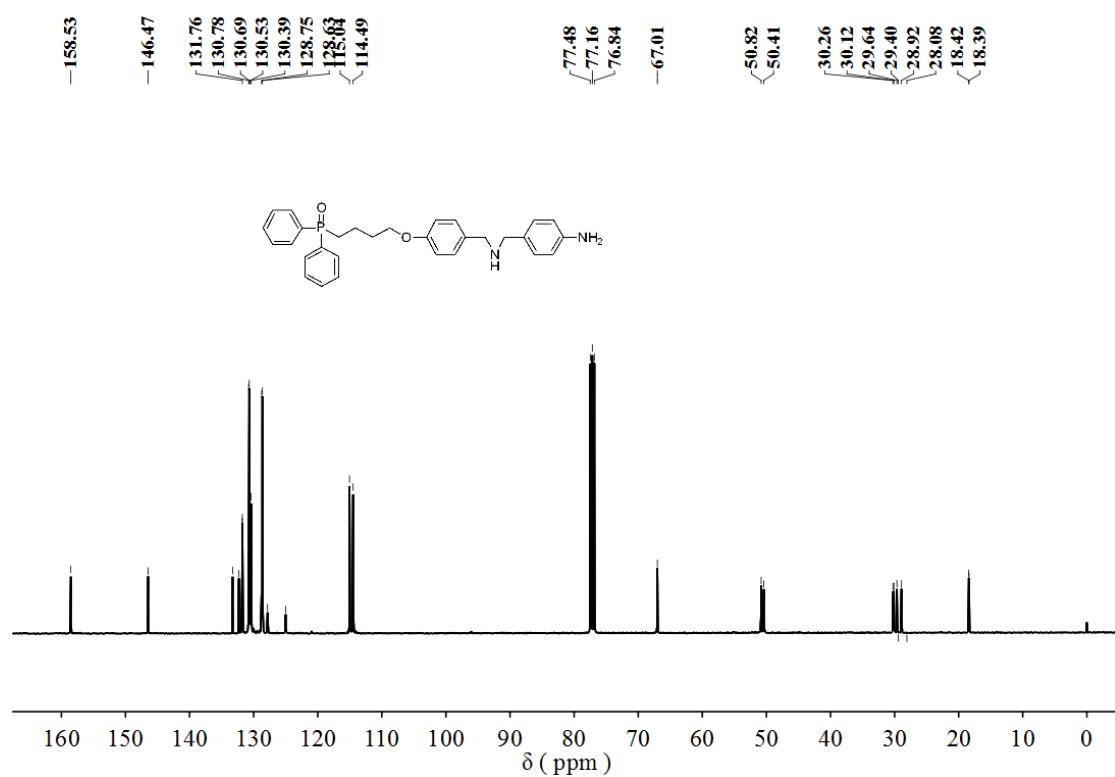


Fig. S8 ¹³C NMR spectrum (CDCl₃, 100 MHz, 298 K) of 2.

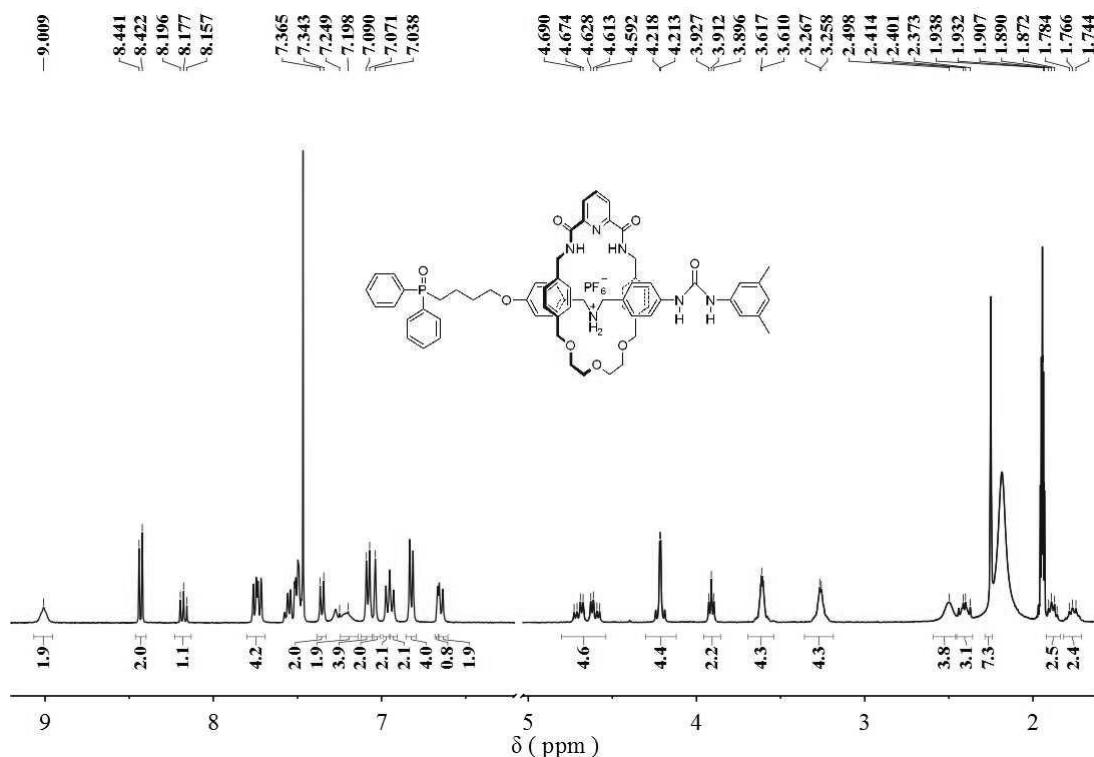


Fig. S9 ^1H NMR spectrum ($\text{CDCl}_3/\text{CD}_3\text{CN} = 1:1$, 400 MHz, 298 K) of **[1-H][PF₆]**.

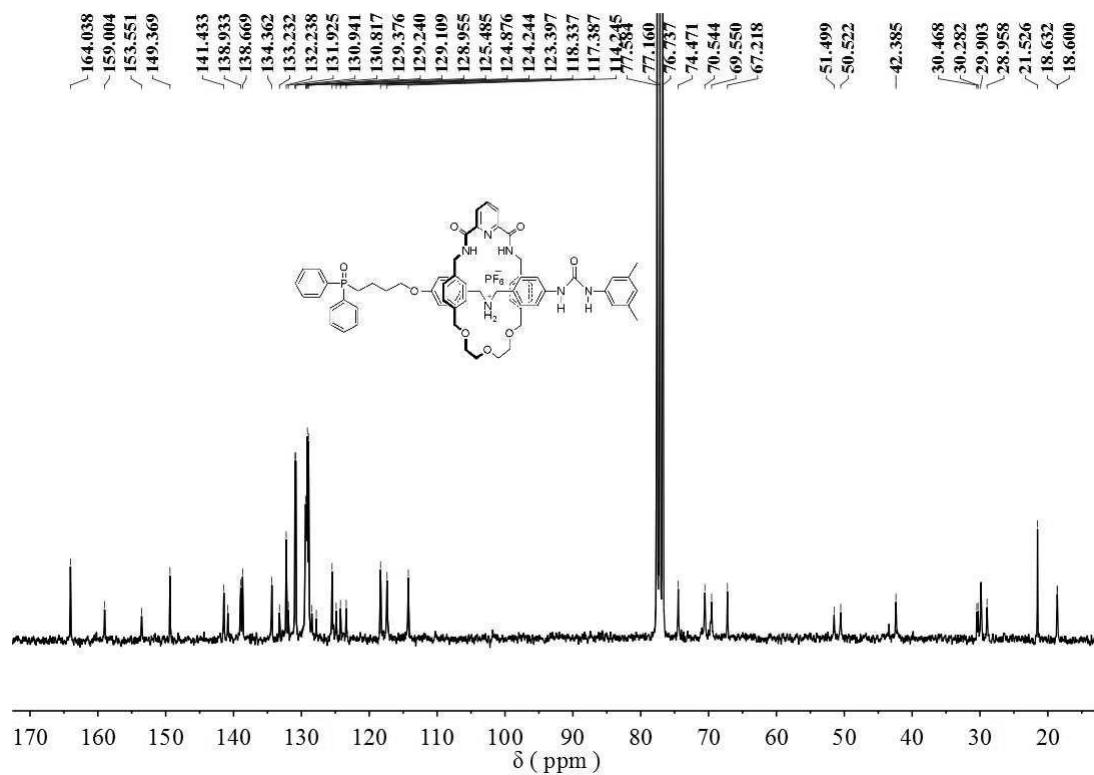


Fig. S10 ^{13}C NMR spectrum (CDCl_3 , 100 MHz, 298 K) of [1-H][PF_6^-].

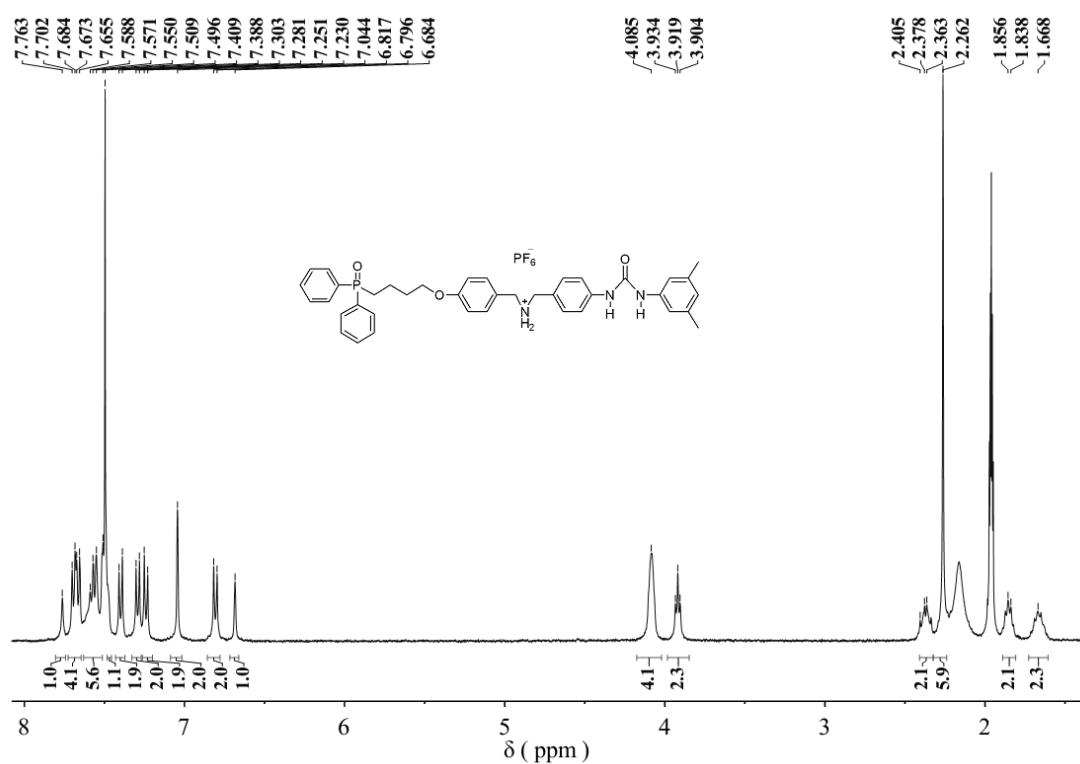


Fig. S11 ¹H NMR spectrum (CDCl₃/CD₃CN = 1:1, 400 MHz, 298 K) of **9**.

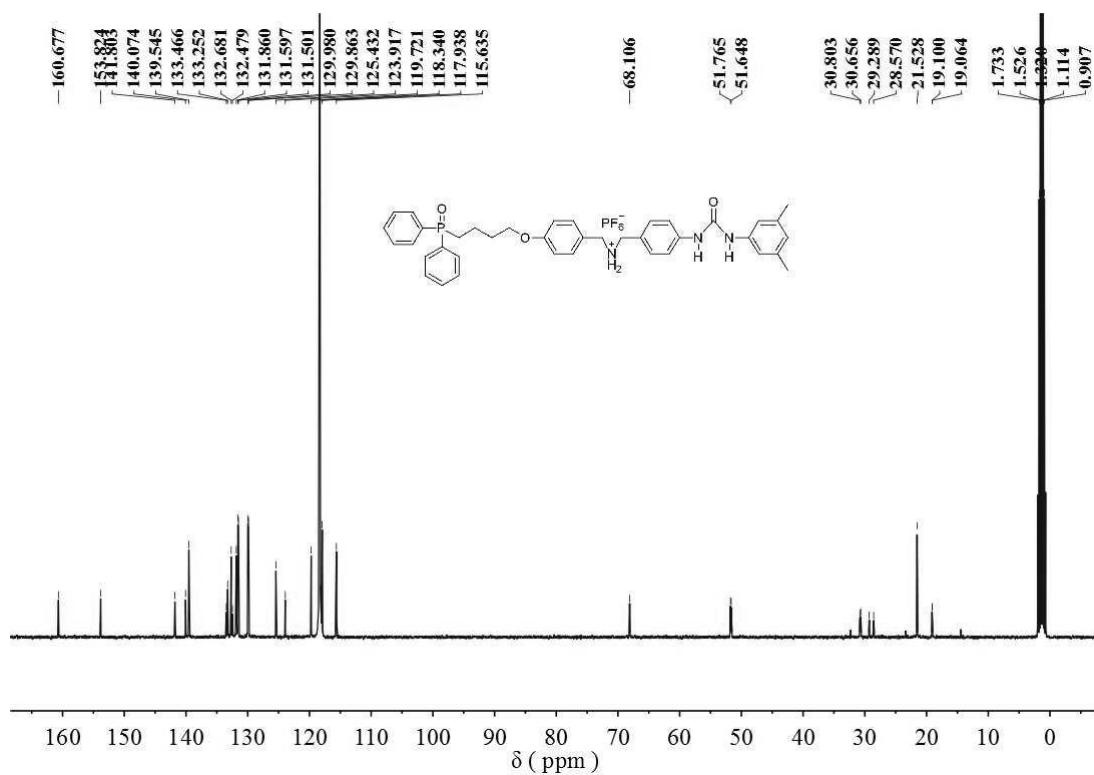


Fig. S12 ¹³C NMR spectrum (CDCl₃/CD₃CN = 1:1, 100 MHz, 298 K) of **9**.

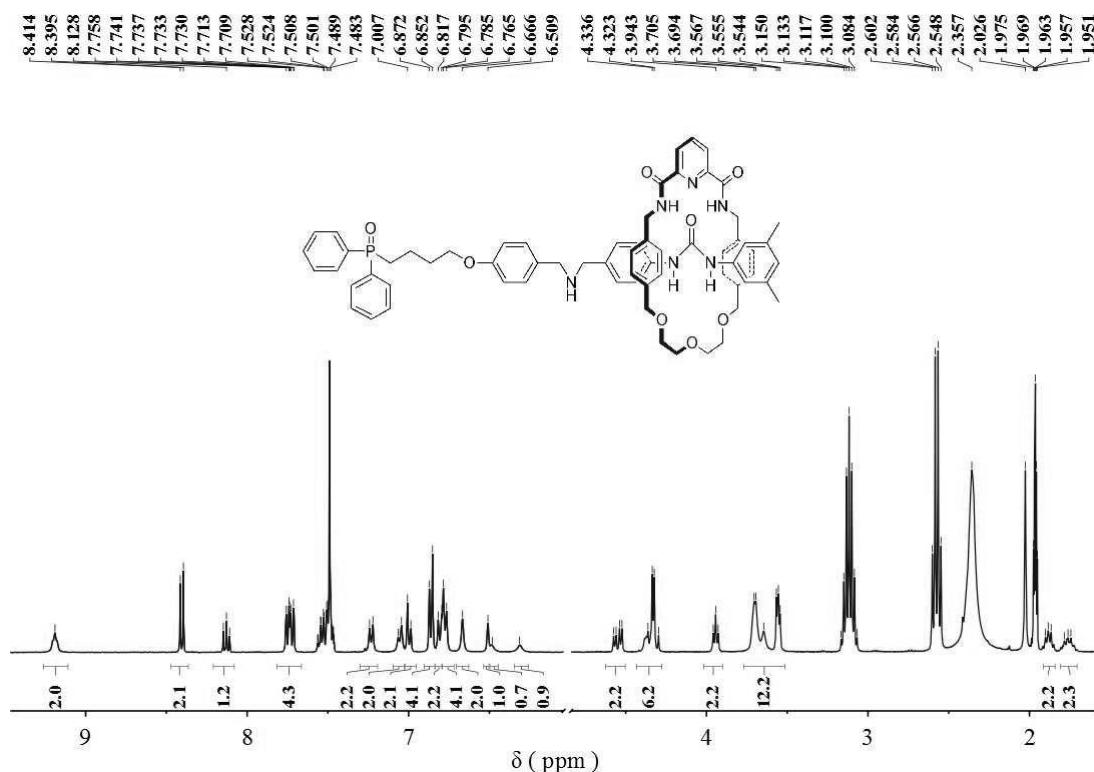


Fig. S13 ¹H NMR spectrum (CDCl₃/CD₃CN = 1:1, 400 MHz, 298 K) of [1-H][PF₆] after the addition of 1.5 equiv. of iPr₂NEt.

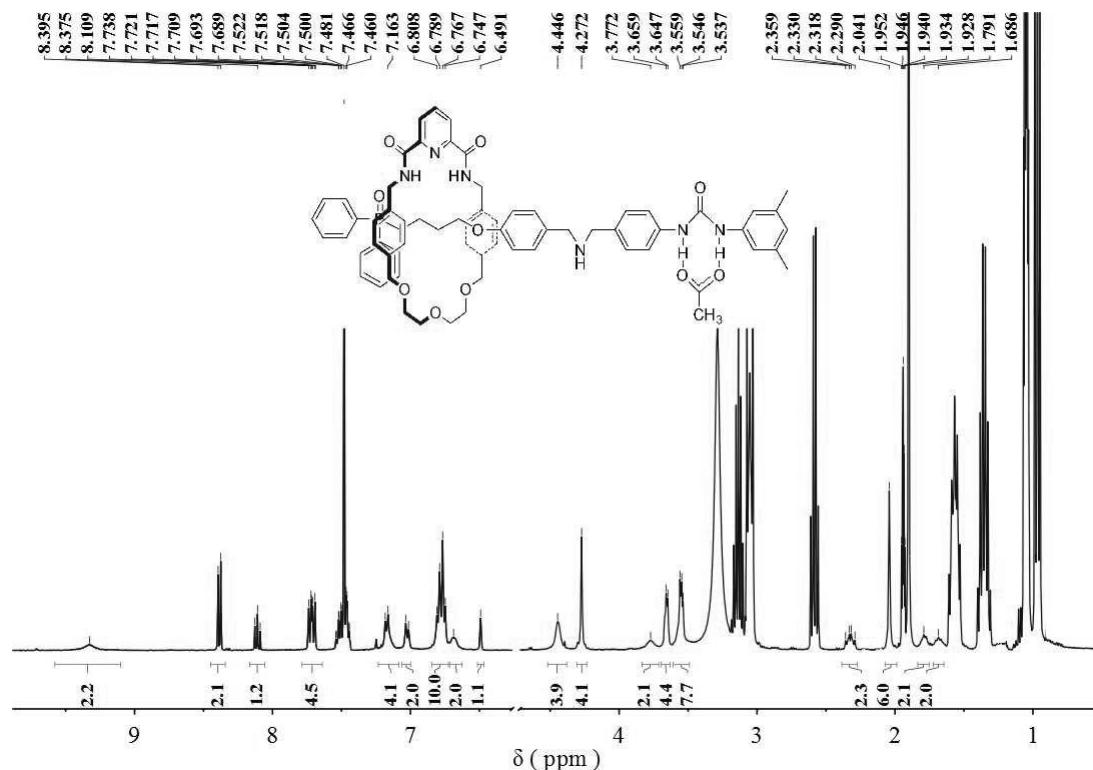


Fig. S14 ¹H NMR spectrum (CDCl₃/CD₃CN = 1:1, 400 MHz, 298 K) of [1-H][PF₆] after the addition of 1.5 equiv. of iPr₂NEt and 2.0 equiv. of TBAA.

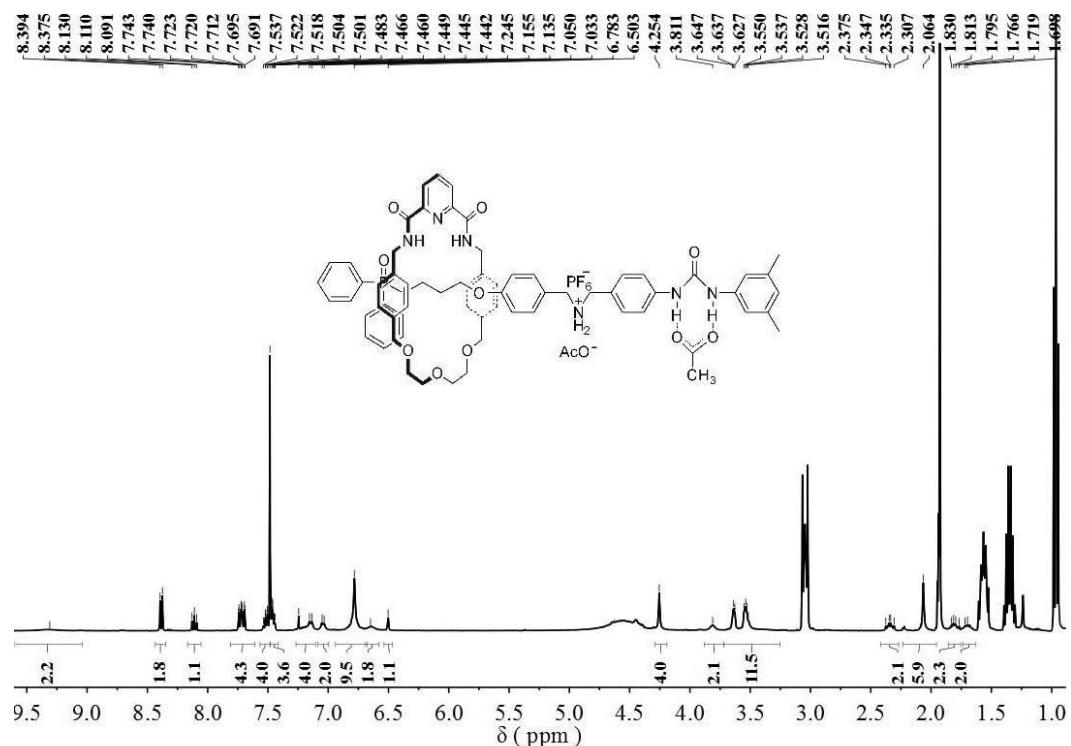


Fig. S15 ¹H NMR spectrum (CDCl₃/CD₃CN = 1:1, 400 MHz, 298 K) of [1-H][PF₆] after the addition of 3.0 equiv. of TBAA.

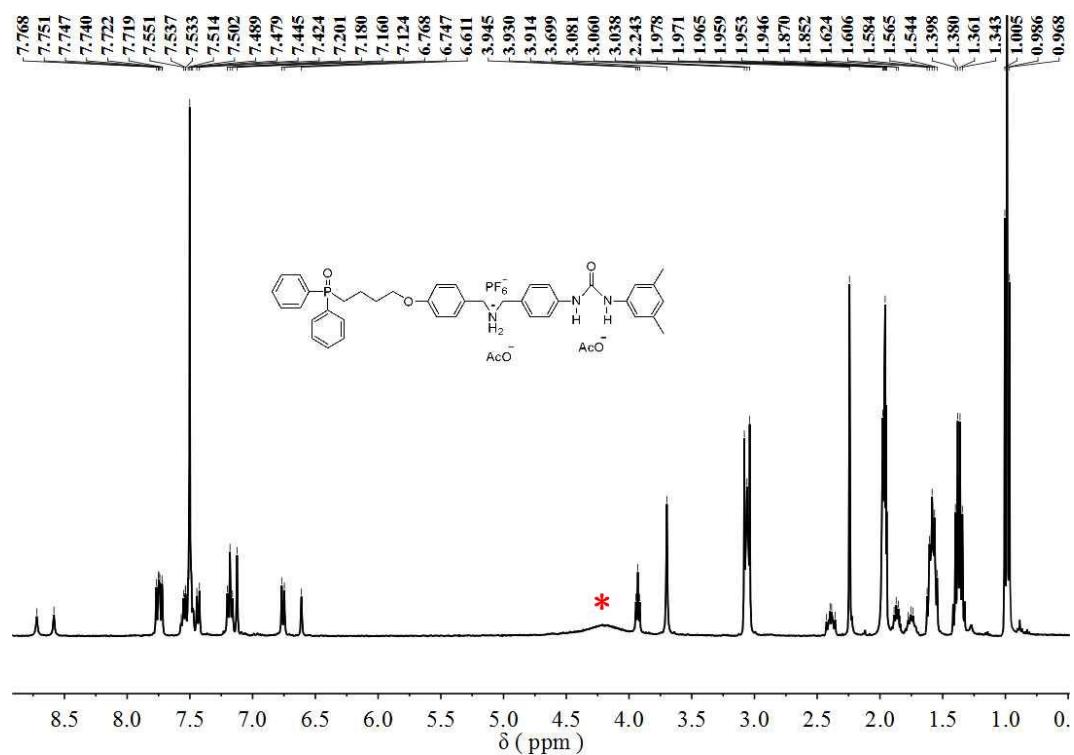


Fig. S16 ¹H NMR spectrum (CDCl₃/CD₃CN = 1:1, 400 MHz, 298 K) of 9 after the addition of 2.0 equiv. of TBAA (The peaks marked with * are ascribed to H₂O.).

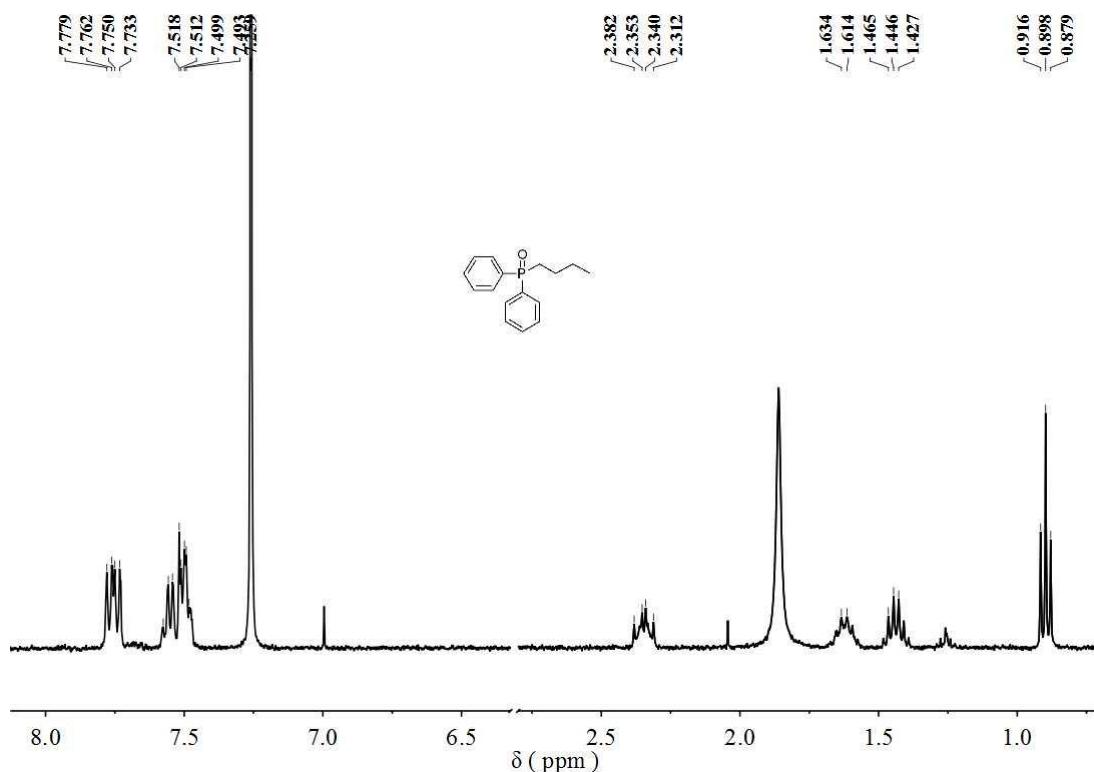


Fig. S17 ^1H NMR spectrum (CDCl_3 , 400 MHz, 298 K) of **10**.

5. ^1H NMR for the shuttling process

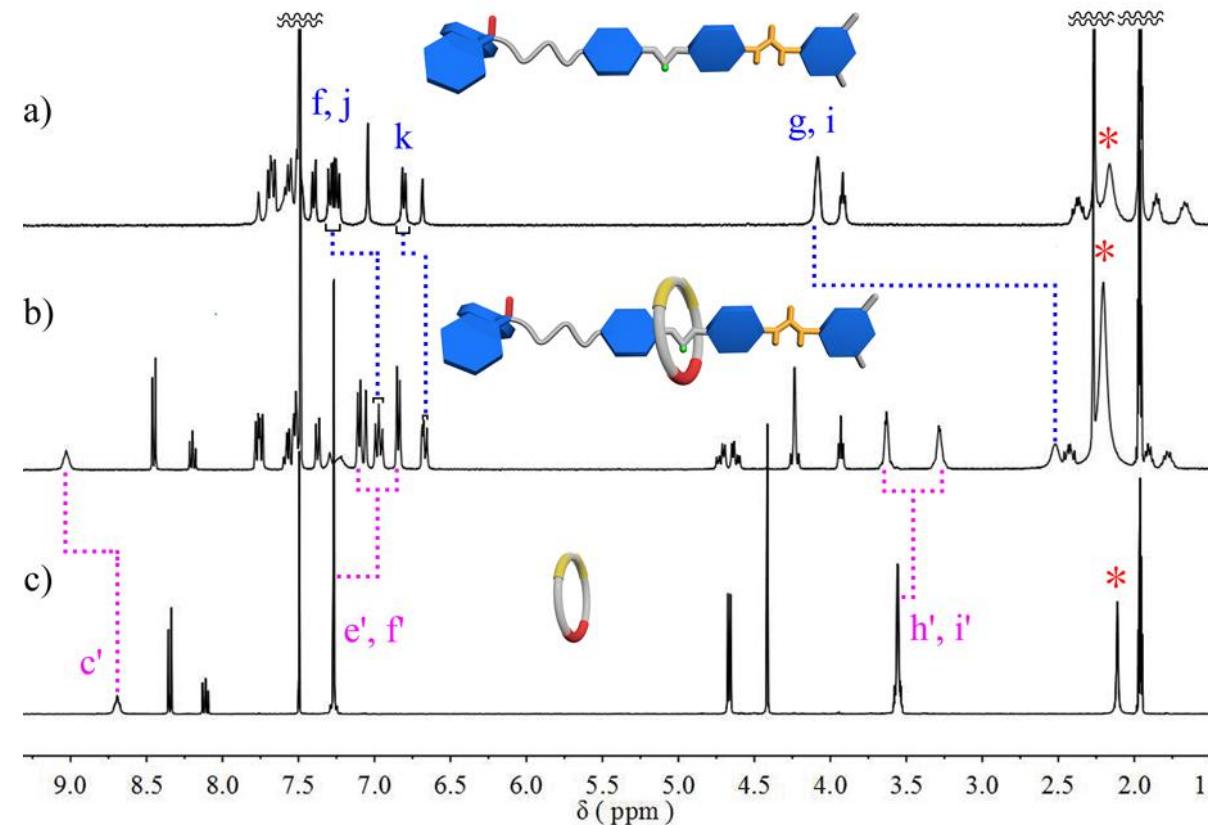


Fig. S18 Partial ^1H NMR spectra (400 MHz, $\text{CDCl}_3/\text{CD}_3\text{CN} = 1:1$, 298 K) of a) thread **9**, b) [2]rotaxane **[1-H][PF₆]**, and c) macrocycle **3**. (The peaks marked with * are ascribed to H_2O .)

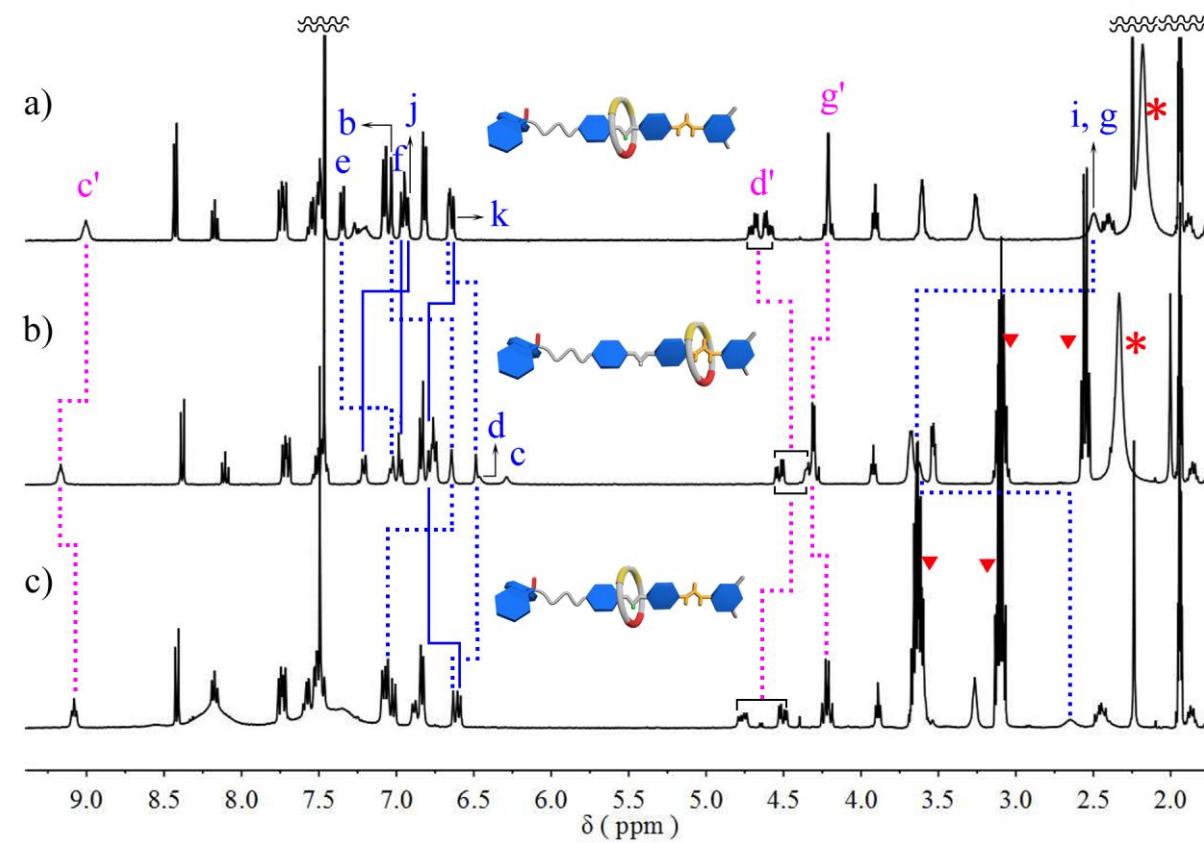


Fig. S19 Partial ¹H NMR spectra (400 MHz, $\text{CDCl}_3/\text{CD}_3\text{CN} = 1:1$, 298 K) of a) molecular shuttle **[1-H][PF₆], b) the mixture obtained after adding $i\text{Pr}_2\text{NEt}$ (1.5 equiv.) to the solution in a), c) the mixture obtained after adding TFA (2.0 equiv.) to the solution in b). (The peaks marked with * are ascribed to H_2O and the peaks marked with ▼ are ascribed to $i\text{Pr}_2\text{NEt}$).**

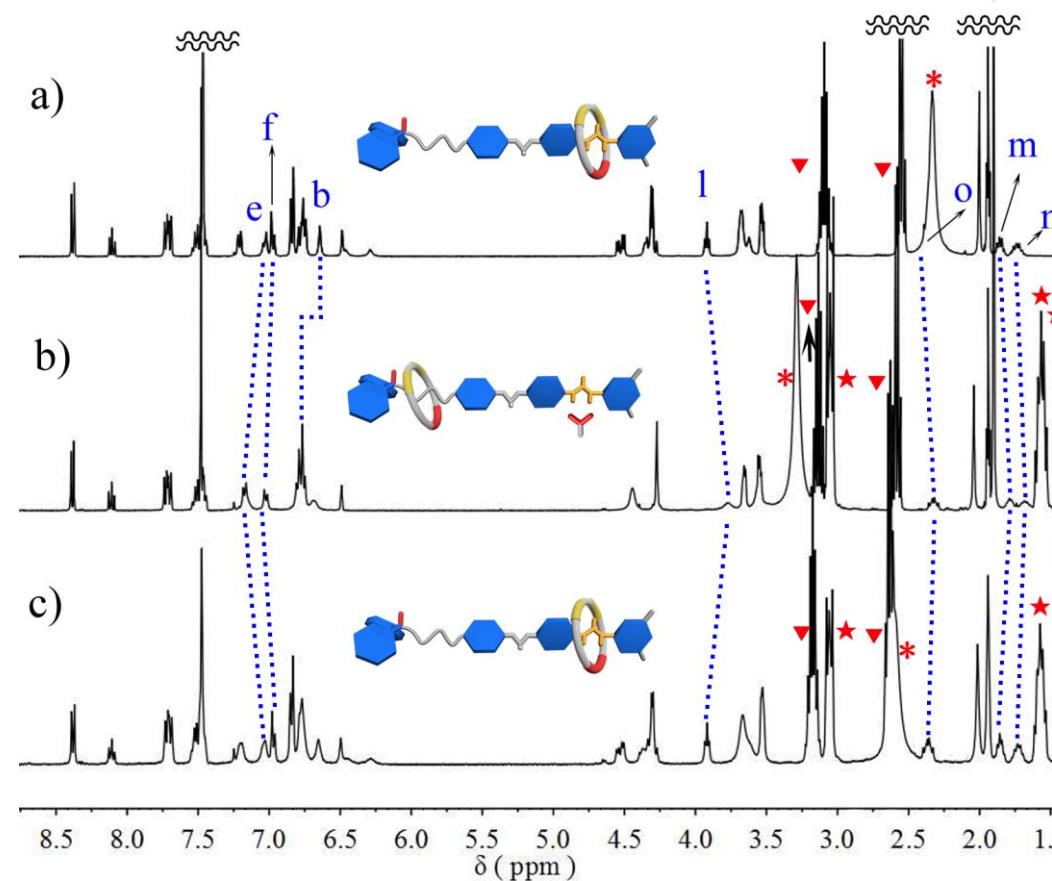


Fig. S20 Partial ^1H NMR spectra (400 MHz, $\text{CDCl}_3/\text{CD}_3\text{CN} = 1:1$, 298 K) of a) molecular shuttle **[1-H][PF₆]** after adding $i\text{Pr}_2\text{NEt}$ (1.5 equiv.), b) the mixture obtained after adding TBAA (2.0 equiv.) to the solution in a), c) the mixture obtained after adding NaClO₄ (3.0 equiv.) to the solution in b). (The peaks marked with * are ascribed to H_2O , the peaks marked with ▼ are ascribed to $i\text{Pr}_2\text{NEt}$ and the peaks marked with ★ are ascribed to TBAA)

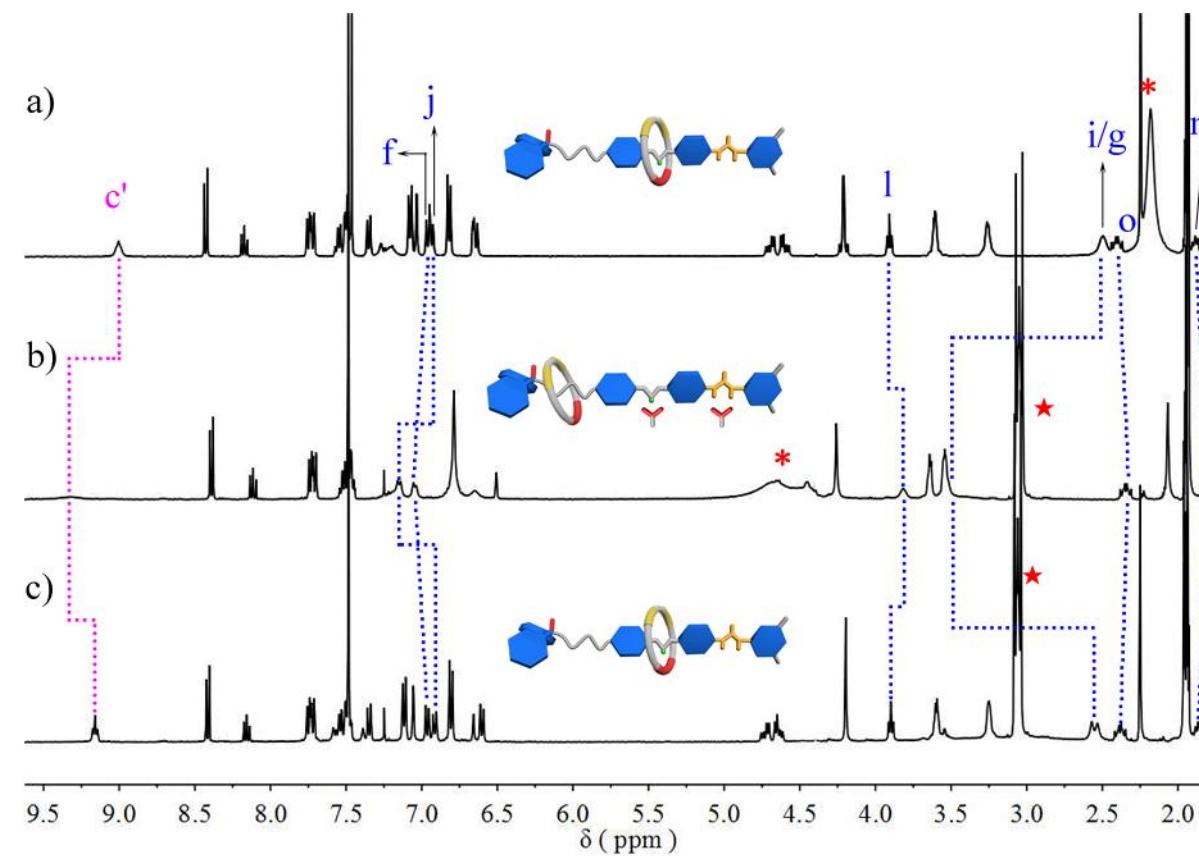


Fig. S21 Partial ¹H NMR spectra (400 MHz, $\text{CDCl}_3/\text{CD}_3\text{CN} = 1:1$, 298 K) of a) molecular shuttle **[1-H][PF₆]**, b) the mixture obtained after adding TBAA (3.0 equiv.) to the solution in a), c) the mixture obtained after adding NaClO₄ (6.0 equiv.) to the solution in b). (The peaks marked with * are ascribed to H₂O and the peaks marked with ★ are ascribed to TBAA)

6. LR-ESI-MS of [1-H][PF₆] and 9

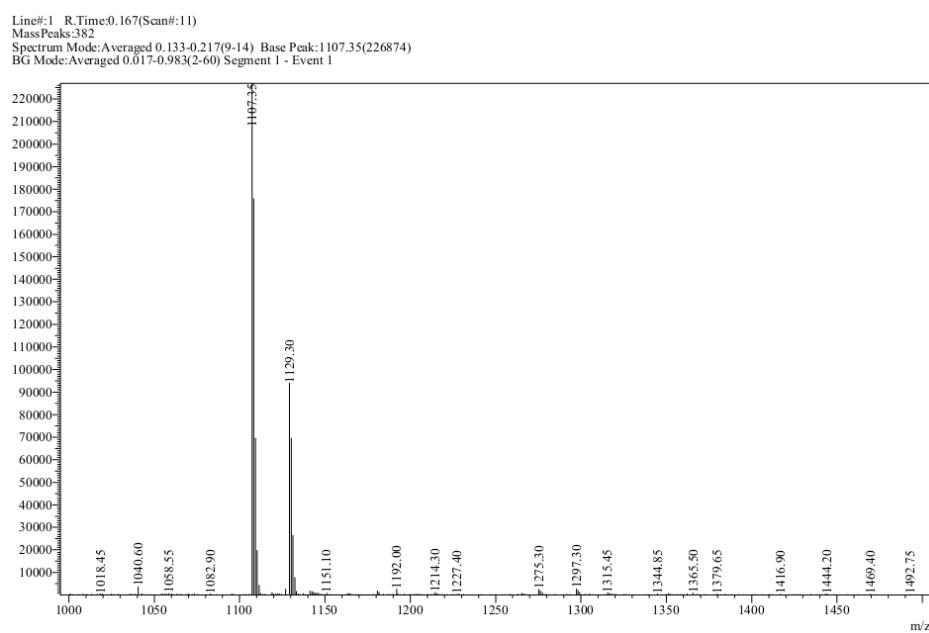


Fig. S22 LR-ESI-MS of [1-H][PF₆].

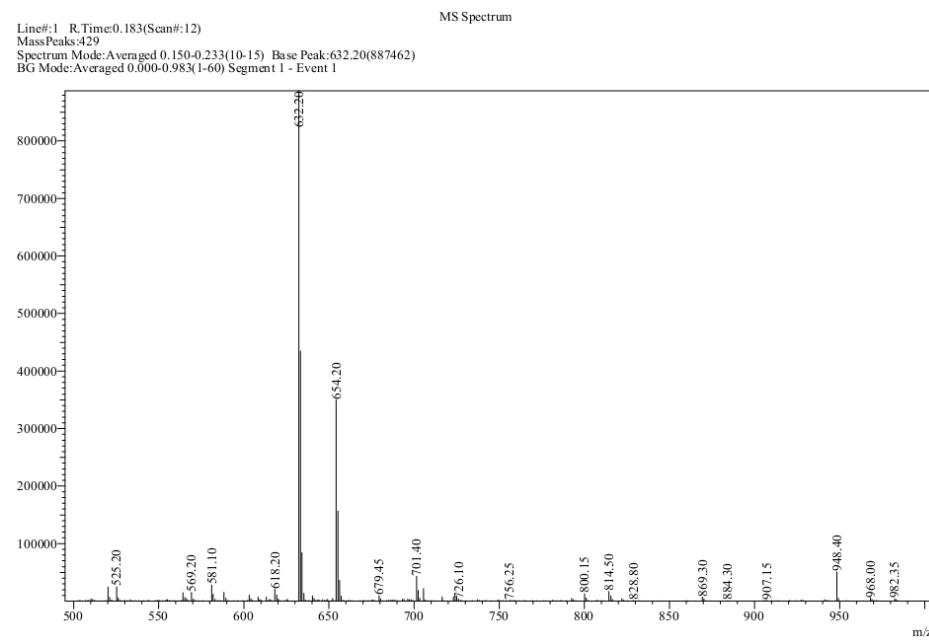


Fig. S23 LR-ESI-MS of 9.

7. Partial 2D COSY and ROESY in different States

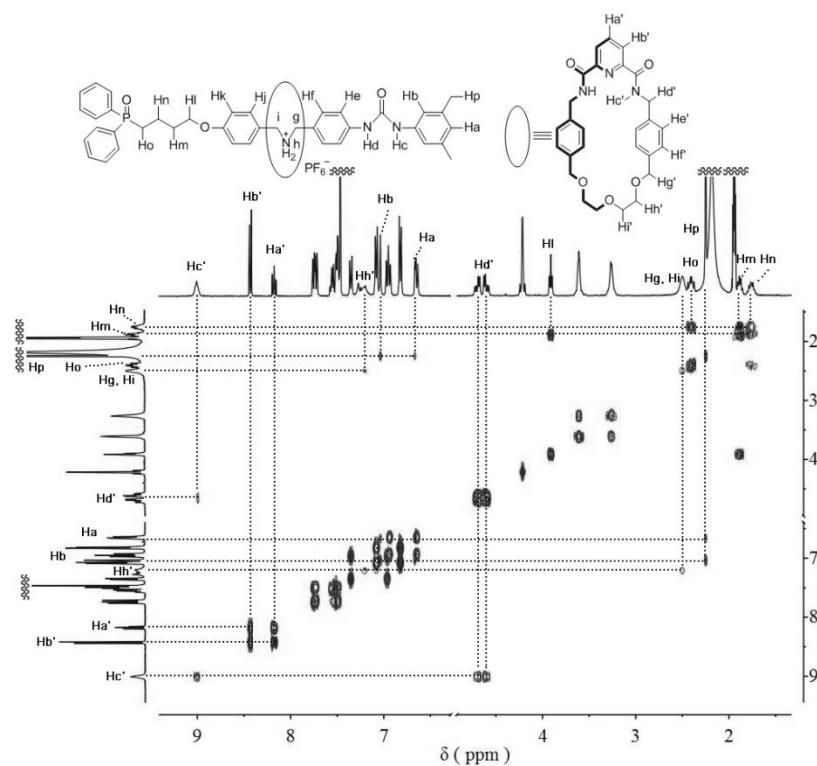


Fig. S24 Partial 2D COSY spectrum ($\text{CDCl}_3/\text{CD}_3\text{CN} = 1:1$, 400 MHz, 298 K) of **[1-H][PF₆]**.

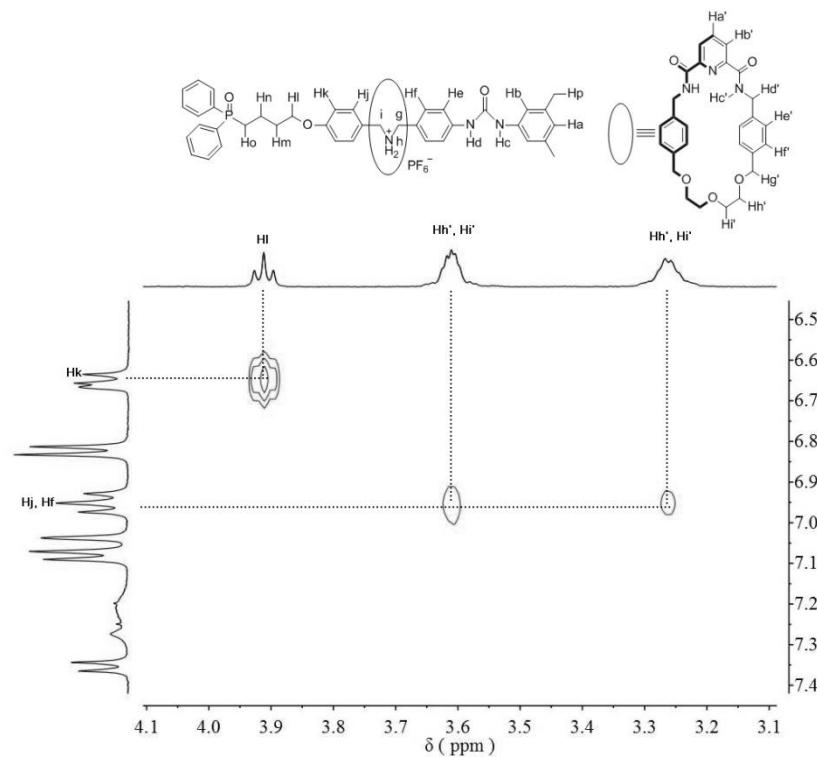


Fig. S25 Partial 2D ROESY spectrum ($\text{CDCl}_3/\text{CD}_3\text{CN} = 1:1$, 400 MHz, 298 K) of **[1-H][PF₆]**.

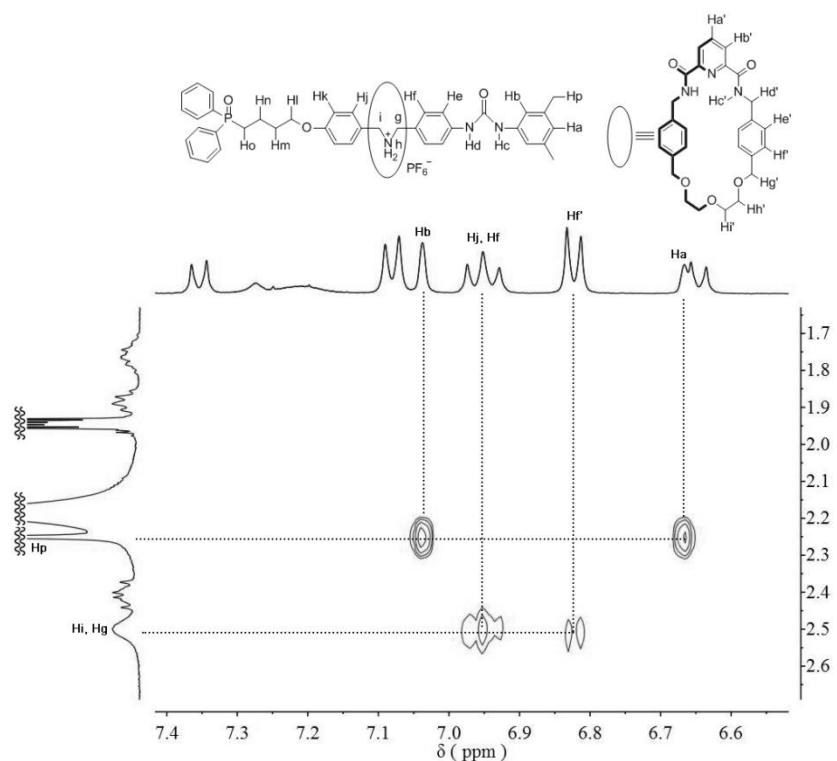


Fig. S26 Partial 2D ROESY spectrum (CDCl₃/CD₃CN = 1:1, 400 MHz, 298 K) of [1-H][PF₆].

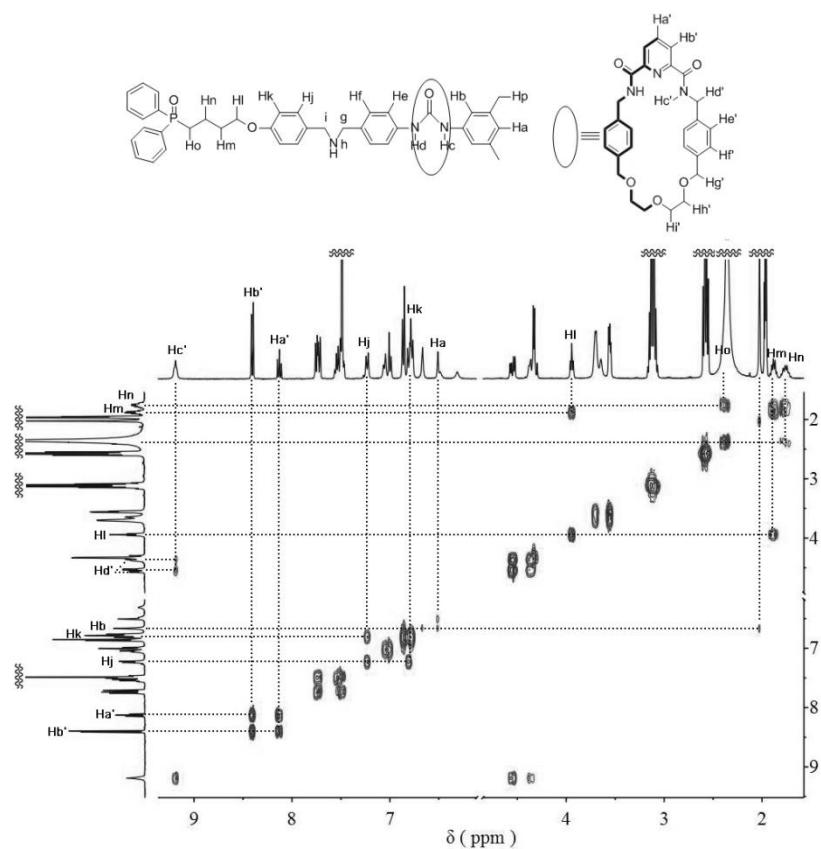


Fig. S27 Partial 2D COSY spectrum (CDCl₃/CD₃CN = 1:1, 400 MHz, 298 K) of [1-H][PF₆] after the addition of 1.5 equiv. of iPr₂NEt.

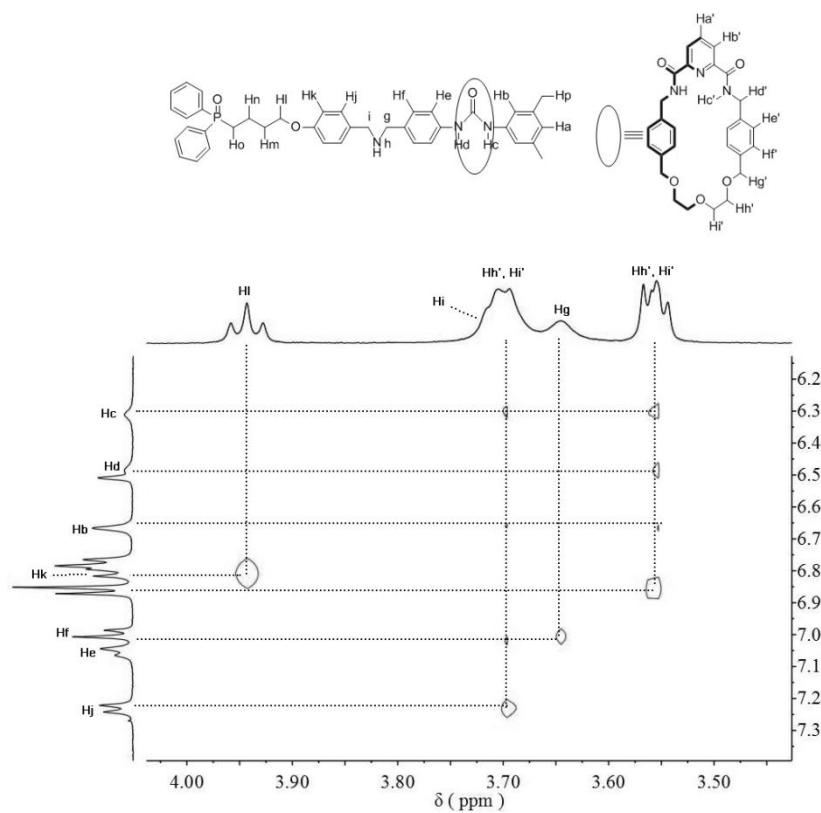


Fig. S28 Partial 2D ROESY spectrum ($\text{CDCl}_3/\text{CD}_3\text{CN} = 1:1$, 400 MHz, 298 K) of [1-H][PF₆] after the addition of 1.5 equiv. of *i*Pr₂NEt.

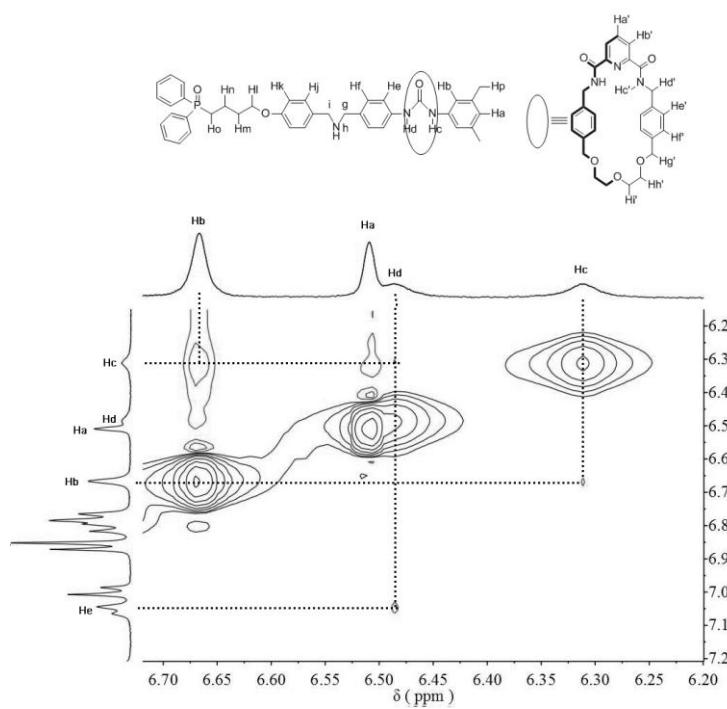


Fig. S29 Partial 2D ROESY spectrum ($\text{CDCl}_3/\text{CD}_3\text{CN} = 1:1$, 400 MHz, 298 K) of **[1-H][PF₆]** after the addition of 1.5 equiv. of *i*Pr₂NEt.

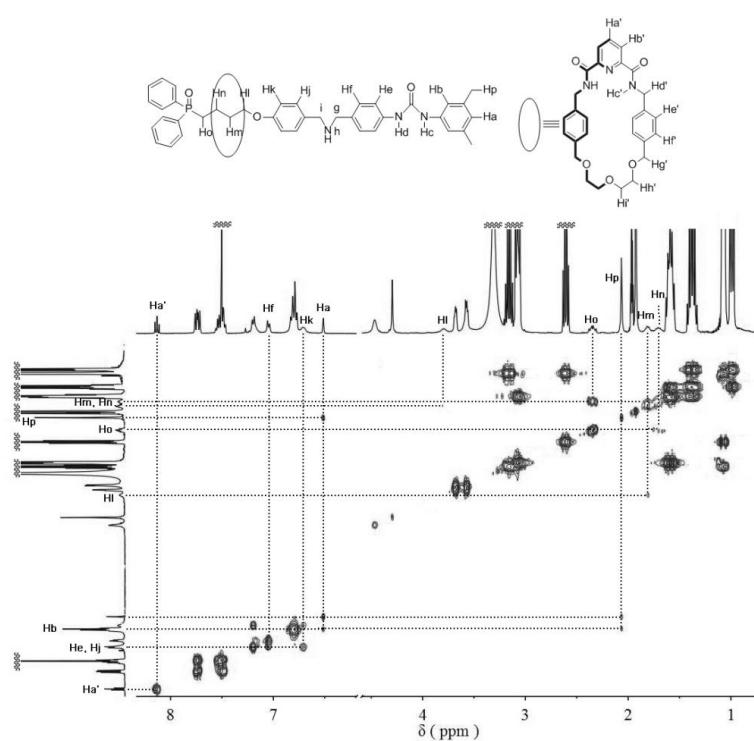


Fig. S30 Partial 2D COSY spectrum ($\text{CDCl}_3/\text{CD}_3\text{CN} = 1:1$, 400 MHz, 298 K) of **[1-H][PF₆]** after the addition of 1.5 equiv. of *i*Pr₂NEt and 2.0 equiv. of TBAA.

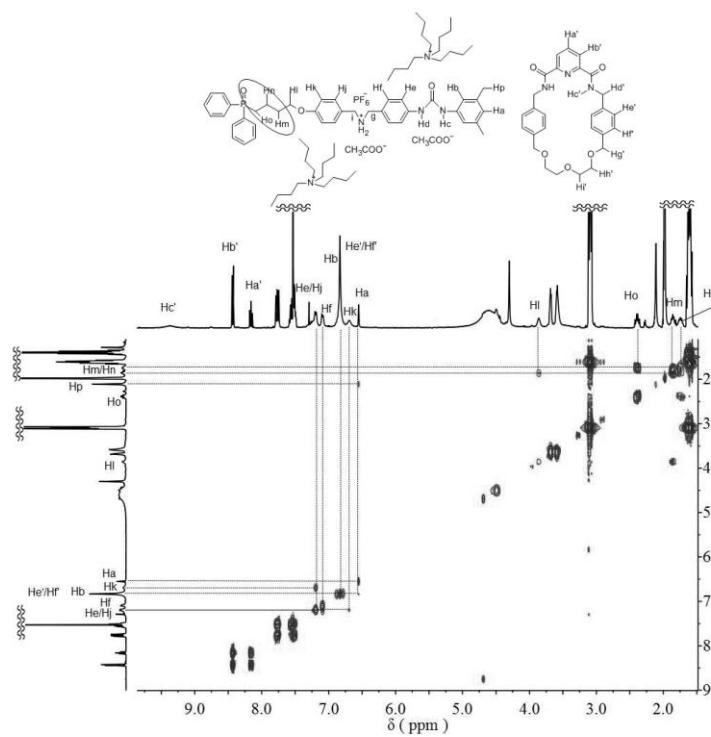


Fig. S31 Partial 2D COSY spectrum ($\text{CDCl}_3/\text{CD}_3\text{CN} = 1:1$, 400 MHz, 298 K) of **[1-H][PF₆]** after the addition of 3.0 equiv. of TBAA.

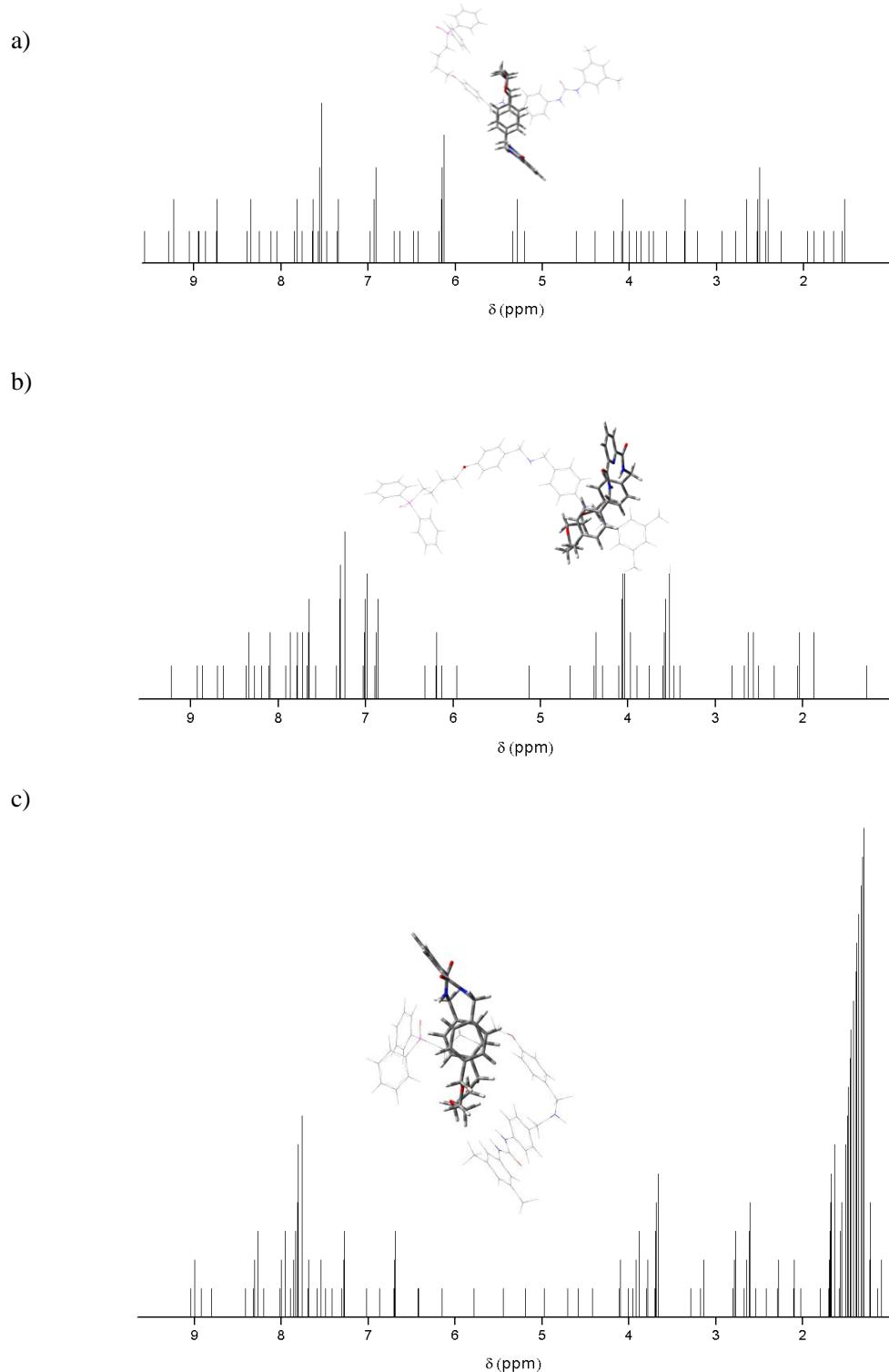
8. X-Ray Data for [1-H][PF₆]

Table 1. Crystal data and structure refinement for [1-H][PF₆]

CCDC number	900778
Empirical formula	C ₆₆ H ₇₄ N ₆ O ₁₀ F ₆ P ₂
Formula weight	1289.27
Temperature	296(2)
Wavelength	0.71073 Å
Crystal system	Triclinic
Space group	P ⁻ 1
<i>a</i>	14.9639(11)
<i>b</i>	16.2074(10)
<i>c</i>	16.6904(12)
α	68.0740(10)
β	65.602(2)
γ	86.2270(10)
Volume	3400.9(4) Å ³
<i>Z</i>	2
Density (calculated)	1.259
Absorption coefficient	0.140
F(000)	1356.0
Crystal size	0.28 × 0.24 × 0.22 mm ³
Theta range for data collection	2.34 to 22.15 °
Index ranges	-16<=h<=18, -18<=k<=19, -0<=l<=20
Reflections collected	37275
Independent reflections	13369 [R(int) = 0.0425]
Completeness to theta = 26.00 °	100.0 %
Absorption correction	multi-scan
Refinement method	Full-matrix least-squares on <i>F</i> 2
Goodness-of-fit on <i>F</i> 2	1.117
Final R indices [<i>I</i> > 2sigma(<i>I</i>)]	R1 = 0.0577, <i>wR</i> 2 = 0.1132
<i>R</i> indices (all data)	R1 = 0.0837, <i>wR</i> 2 = 0.1186
Largest diff. peak and hole	0.185 and -0.599 e Å ⁻³

9. Theoretical calculations

All calculations were performed with the Gaussian 09 program suite^[S6].



d)

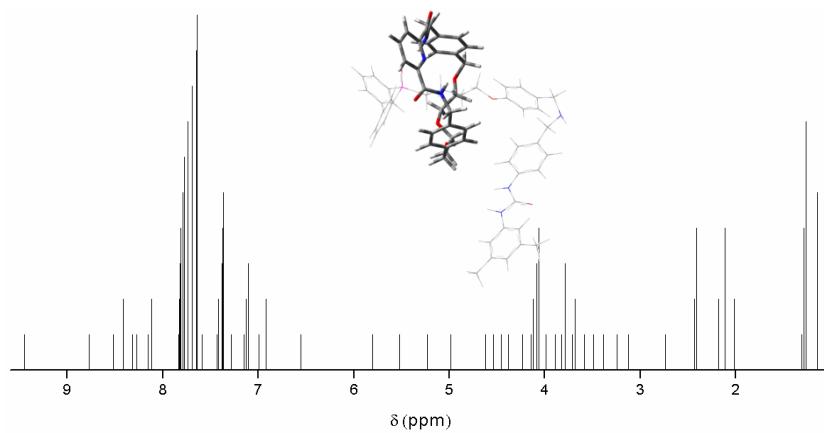


Fig. S32 The optimized structures and DFT-calculated chemical shifts of ^1H of three different stations a) State **I**, b) State **II**, c) State **III**, and d) State **III'**.

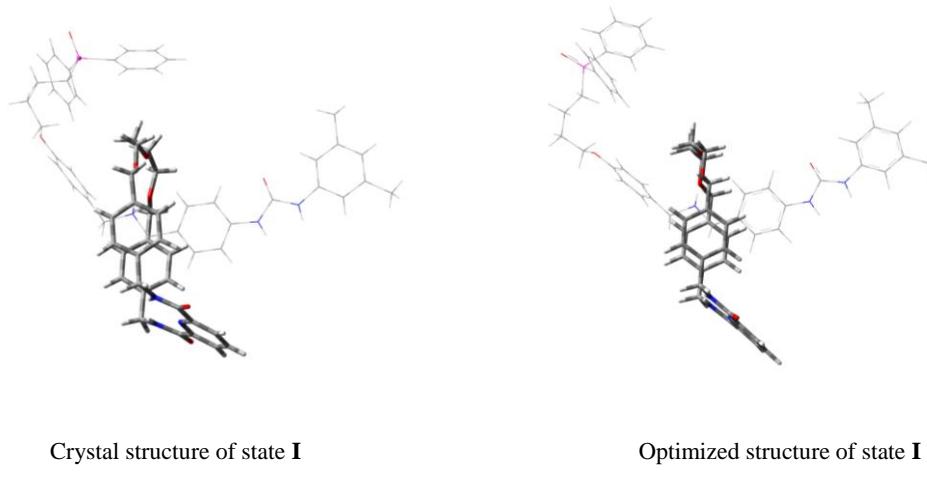


Fig. S33 The crystal structure (left) and the geometry optimized structure by theoretical calculations (right) for molecular shuttle at State **I**.

10. References

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