

Synthesis and Anticancer Activity of Epipolythiodiketopiperazine Alkaloids

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Supporting Material

General procedures	S3
Materials	S3
Instrumentation	S3
Positional numbering system	S4
Figure S1: List of dimeric epipolythiodiketopiperazines and diketopiperazines	S5
Figure S2: List of C3-substituted epipolythiodiketopiperazines and diketopiperazines	S5
Scheme S1: Synthesis of 3 , 14 , 18 , and 21–23	S6
Scheme S2: Synthesis of 4–6 and 15–17	S6
General reagents and methods for biological assays	S7
Cell culture information	S7
IC ₅₀ value determination for adherent cells using Sulforhodamine B	S7
IC ₅₀ value determination for non-adherent cells using MTS	S7
Hemolysis assay using human erythrocytes	S8
Figure S3: Percent hemolysis following treatment with ETPs from Table 2	S8
Apoptosis in U-937 cells with annexin V-FITC and propidium iodide	S8
Apoptosis in U-937 cells by western blot analysis	S8
C3-(5-Bromo-1-TIPS-indol-3'-yl)-pyrrolidinoindoline S12	S10
C3-(indol-3'-yl)-pyrrolidinoindoline (+)- 59	S12
C3-(Indol-3'-yl)-hexacyclic diol (–)- 56	S14
C3-(Indol-3'-yl)-epidithiodiketopiperazine 26	S16
General procedure for the Friedel–Crafts nucleophile substitution	S18
General procedure for the regio- and stereoselective hydroxylation	S18
C3-Bromo epidithiodiketopiperazines 30 and 34	S19
C3-Fluoro epidithiodiketopiperazines 31 and 35	S21
C3-(Pyrrol-3'-yl)-epidithiodiketopiperazine 32	S23–
C3-(<i>p</i> -Methoxyphenyl)-epidithiodiketopiperazine 33	S25
Hexacyclic triphenylmethanedisulfide (+)- 71	S27
(+)-12-Deoxybionectin A 10	S29
C3-(Indol-3'-yl)-epitrithiodiketopiperazine 29	S30
C3-(Indol-3'-yl)-C11-thiohemiaminal 48	S32
C3-(Indol-3'-yl)-epitrithiodiketopiperazine 27	S33
C3-(Indol-3'-yl)-epitetrathiodiketopiperazine 28	S35
C3-(<i>N</i> -Boc-indol-3'-yl)-bis(benzylthioether) 43	S36

C3-(<i>N</i> -Boc-indol-3'-yl)-epidithiodiketopiperazine 24	S38
C3-(<i>N</i> -Boc-indol-3'-yl)-bis(<i>S</i> -MOM)ether 40	S39
C3-(Indol-3'-yl)-bis(<i>S</i> -MOM)ether 41	S41
C3-(<i>N</i> -Boc-indol-3'-yl)-bis(<i>S</i> -MEM)ether 42 and C3-(<i>N</i> -Boc-indol-3'-yl)- <i>S</i> 15-MEM ether 47	S43
C3-(Indol-3'-yl)-dithiepanethione 36	S45
C3-(Indol-3'-yl)-dithiocarbonate 37	S47
C3-(Indol-3'-yl)-dithioacetal 38	S49
C3-(Indol-3'-yl)-epimonothiodiketopiperazine 25	S51
C3-(Indol-3'-yl)-bisthioacetate 44	S52
C3-(Indol-3'-yl)- <i>N</i> -(thiomethyl) bis(methylsulfane) 45	S54
C3-(Indol-3'-yl)-bis(methylsulfane) 46	S56
C3-(Indol-3'-yl)-pyrrolidinoindoline 74	S58
C3-(Indol-3'-yl)-dithiepanethiones 64 and 66	S59
C3-(Indol-3'-yl)-epidithiodiketopiperazines 60 and 62	S61
Dimeric bisdithiepanethione 18	S63
Dimeric epidithiodiketopiperazine 14	S64
C3-Propyl-dithiepanethiones 65 and 67	S65
β -C3-Propyl-epidithiodiketopiperazine 61	S67
α -C3-Propyl-epidithiodiketopiperazine 63	S68
Dimeric bis(triphenylmethanetrifluoride) 19	S69
Copies of ^1H , ^{13}C , and ^{19}F NMR spectra	S71

General Procedures. All reactions were performed in oven-dried or flame-dried round-bottom flasks. The flasks were fitted with rubber septa, and reactions were conducted under a positive pressure of argon. Cannulae or gas-tight syringes with stainless steel needles were used to transfer air- or moisture-sensitive liquids. Where necessary (so noted), solutions were deoxygenated by sparging with argon for a minimum of 10 min. Flash column chromatography was performed as described by Still *et al.* using granular silica gel (60-Å pore size, 40–63 μm, 4–6% H₂O content, Zeochem).¹ Analytical thin layer chromatography (TLC) was performed using glass plates pre-coated with 0.25 mm 230–400 mesh silica gel impregnated with a fluorescent indicator (254 nm). TLC plates were visualized by exposure to short wave ultraviolet light (254 nm), reversibly stained with iodine (I₂ absorbed on silica) vapor, and irreversibly stained by treatment with an aqueous solution of ceric ammonium molybdate (CAM) followed by heating (~ 1 min) on a hot plate (~ 250 °C). Organic solutions were concentrated at 29–30 °C on rotary evaporators capable of achieving a minimum pressure of ~2 torr. The benzenesulfonyl photodeprotection was accomplished by irradiation in a Rayonet RMR-200 photochemical reactor (Southern New England Ultraviolet Company, Branford, CT, USA) equipped with 16 lamps (RPR-3500, 24 W, λ_{max} = 350 nm, bandwidth ~ 20 nm).

Materials. Commercial reagents and solvents were used as received with the following exceptions: dichloromethane, acetonitrile, tetrahydrofuran, methanol, pyridine, toluene, and triethylamine were purchased from J.T. Baker (Cycletainer™) and were purified by the method of Grubbs *et al.* under positive argon pressure.² Nitromethane and nitroethane (from Sigma–Aldrich) were purified by fractional distillation over calcium hydride and were stored over Linde 3 Å molecular sieves in Schlenk flasks sealed with septa and Teflon tape under argon atmosphere.³ Hünig’s base and benzene were dried by distillation from calcium hydride under an inert argon atmosphere and used directly. 1,4-Dimethoxynaphthalene, hafnium (IV) trifluoromethanesulfonate hydrate, and iodomethane were purchased from Alfa Aesar; 1-(triisopropylsilyl)-*1H*-pyrrole was purchased from Combi-Block; triphenylmethanesulfonyl chloride was purchased from TCI America, Inc; 2,6-di-*tert*-butyl-4-methylpyridine (DTBMP) was purchased from OChem Incorporation. All other solvents and chemicals were purchased from Sigma–Aldrich. Silver tetrafluoroborate (≥99.99% trace metals basis) and hydrogen sulfide (≥99.5%) were purchased from Sigma–Aldrich. 1,4-Dimethoxynaphthalene was purified by crystallization from absolute ethanol.

Instrumentation. Proton nuclear magnetic resonance (¹H NMR) spectra were recorded with a Bruker AVANCE-600 NMR spectrometer (with a Magnex Scientific superconducting actively-shielded magnet) or a Varian inverse probe 500 INOVA spectrometer, are reported in parts per million on the δ scale, and are referenced from the residual protium in the NMR solvent (CDCl₃: δ 7.26 (CHCl₃), acetone-*d*₆: δ 2.05 (acetone-*d*₅), acetonitrile-*d*₃: δ 2.13 (acetonitrile-*d*₂), DMSO-*d*₆: δ 2.50 (DMSO-*d*₅), methanol-*d*₄: δ 3.31 (methanol-*d*₃)).⁴ Data are reported as follows: chemical shift [multiplicity (br = broad, s = singlet, d = doublet, t = triplet, q = quadruplet, sp = septet, m = multiplet), coupling constant(s) in Hertz, integration, assignment]. Carbon-13 nuclear magnetic resonance (¹³C NMR) spectra were recorded with a Bruker AVANCE-600 NMR spectrometer (with a Magnex Scientific superconducting actively-shielded magnet), a Bruker AVANCE-400 NMR spectrometer (with a Magnex Scientific superconducting magnet), or a Varian 500 INOVA spectrometer, are reported in parts per million on the δ scale, and are referenced from the carbon resonances of the solvent (CDCl₃:

¹ Still, W. C.; Kahn, M.; Mitra, A. *J. Org. Chem.* **1978**, *43*, 2923.

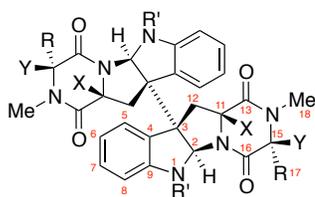
² Pangborn, A. B.; Giardello, M. A.; Grubbs, R. H.; Rosen, R. K.; Timmers, F. J. *Organometallics* **1996**, *15*, 1518.

³ Armarego, W. L. F.; Chai, C. L. L. *Purification of Laboratory Chemicals*, 5th ed.; Butterworth–Heinemann: London, 2003.

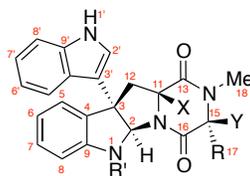
⁴ Fulmer, G. R.; Miller, A. J. M.; Sherden, N. H.; Gottlieb, H. E.; Nudelman, A.; Stoltz, B. M.; Bercaw, J. E.; Goldberg, K. I. *Organometallics* **2010**, *29*, 2176.

δ 77.23, acetone- d_6 : δ 29.84, acetonitrile- d_3 : δ 118.26, DMSO- d_6 : δ 39.52). Data are reported as follows: chemical shift (multiplicity,⁵ coupling constant in Hertz,⁵ assignment). Fluorine-19 nuclear magnetic resonance (¹⁹F NMR) spectra were recorded with a Varian Mercury 300 spectrometer, are reported in parts per million on the δ scale, and are referenced from the fluorine resonance of neat trichlorofluoromethane (CFCl₃; δ 0). Data are reported as follows: chemical shift. Infrared data (IR) were obtained with a Perkin-Elmer 2000 FTIR and are reported as follows: frequency of absorption (cm⁻¹), intensity of absorption (s = strong, m = medium, w = weak, br = broad). Optical Rotations were recorded on a Jasco P-1010 Polarimeter (chloroform, Aldrich, Chromasolv Plus 99.9%; acetone, Aldrich, Chromasolv Plus 99.9%) and specific rotations are reported as follows: [wavelength of light, temperature (°C), specific rotation, concentration in grams/100 mL of solution, solvent]. Preparative HPLC was performed on a Waters system with the 1525 Binary HPLC Pump, 2489 UV/Vis Detector, 3100 Mass Detector, System Fluidics Organizer, and 2767 Sample Manager components. We are grateful to Dr. Li Li and Deborah Bass for obtaining the mass spectrometric data at the Department of Chemistry's Instrumentation Facility, Massachusetts Institute of Technology. High-resolution mass spectra (HRMS) were recorded on a Bruker Daltonics APEXIV 4.7 Tesla FT-ICR-MS using an electrospray (ESI) ionization source.

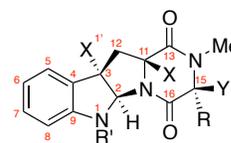
Positional Numbering System. At least three numbering systems for dimeric diketopiperazine alkaloids exist in the literature.⁶ In assigning the ¹H and ¹³C NMR data of all intermediates en route to our different naturally occurring ETPs and their synthetic analogues, we wished to employ a uniform numbering scheme. For ease of direct comparison, particularly between early intermediates, non-thiolated diketopiperazines, and advanced compounds, the numbering system used by Barrow for (+)-WIN-64821 (using positional numbers 1–21) is optimal and used throughout this report. In key instances, the products are accompanied by the numbering system as shown below.



The numbering system used for all dimeric diketopiperazines in this report



The numbering system used for all C3-(indol-3'-yl) diketopiperazines in this report



The numbering system used for all C3-substituted diketopiperazines in this report

⁵ Given if applicable.

⁶ (a) Von Hauser, D.; Weber, H. P.; Sigg, H. P. *Helv. Chim. Acta* **1970**, *53*, 1061. (b) Barrow, C. J.; Cai, P.; Snyder, J. K.; Sedlock, D. M.; Sun, H. H.; Cooper, R. *J. Org. Chem.* **1993**, *58*, 6016. (c) Springer, J. P.; Büchi, G.; Kobbe, B.; Demain, A. L.; Clardy, J. *Tetrahedron Lett.* **1977**, *28*, 2403.

A. Dimeric ETP derivatives from *N*-methyl-L-alanine/serine and L-tryptophan *cyclo*-dipeptide

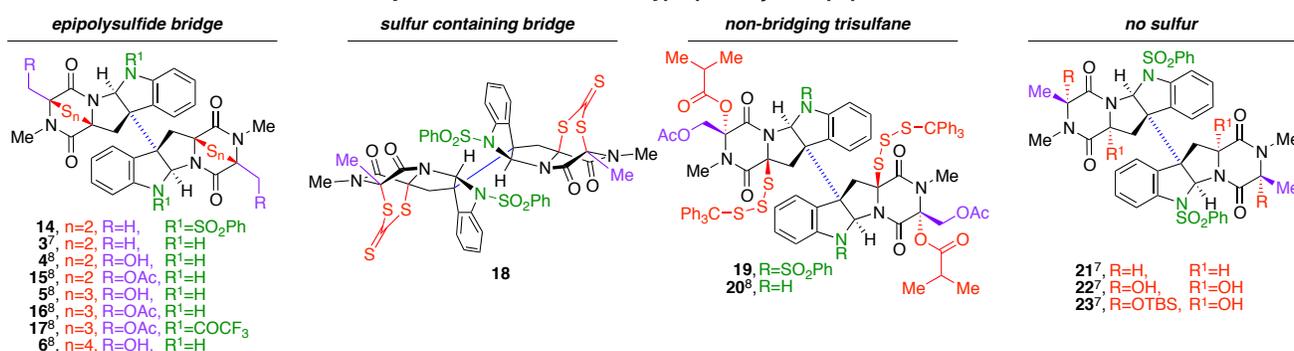
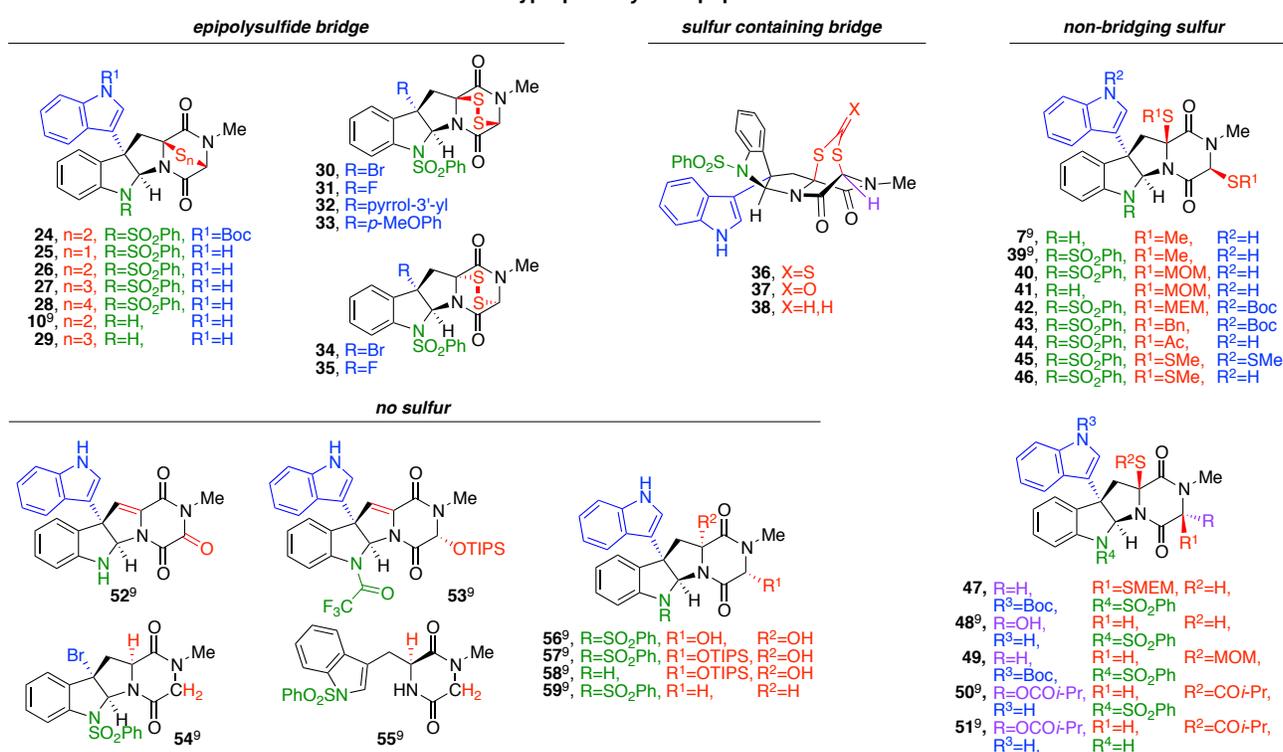


Figure S1. List of dimeric epipolythiodiketopiperazines and diketopiperazines.^{7,8}

B. Monomeric ETP derivatives from sarcosine and L-tryptophan *cyclo*-dipeptide



C. Monomeric ETP derivatives from *N*-methyl-L-alanine and L-tryptophan *cyclo*-dipeptide

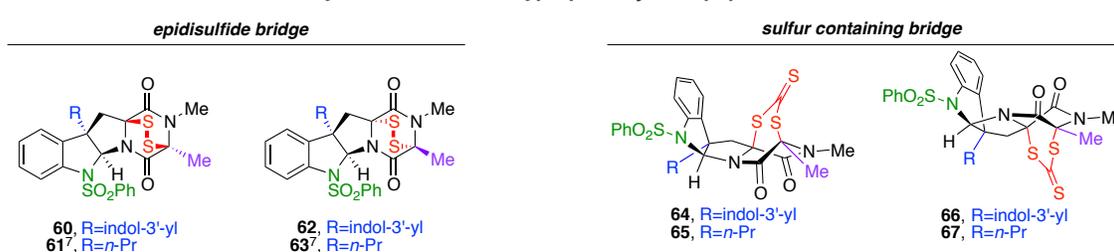


Figure S2. List of C3-substituted epipolythiodiketopiperazines and diketopiperazines.⁹

⁷ For the experimental procedure and characterization data, see: Kim, J.; Ashenhurst, J. A.; Movassaghi, M. *Science* **2009**, 324, 238.

⁸ For the experimental procedure and characterization data, see: Kim, J.; Movassaghi, M. *J. Am. Chem. Soc.* **2010**, 132, 14376.

⁹ For the experimental procedure and characterization data, see: Boyer, N.; Movassaghi, M. *Chem. Sci.* **2012**, 3, 1798.

N₂H₄, THF, 0 °C; NaH, Ph₃CSCl, 90%; (e) BF₃•OEt₂, DTBMP, Et₃SiH, CH₂Cl₂, 82%; (f) Otera's cat., MeOH, PhMe, 85 °C, 92%; (g) N₂H₄, THF, 0 °C; TrSSCl, NEt₃, 86%; (h) N₂H₄, THF, 0 °C, 93%; (i) TrSSCl, NEt₃, 80%; (j) TFAA, DTBMP, MeCN; BF₃•OEt₂, 91%; (k) HCO₂Ac; MeCN, BF₃•OEt₂, 60%; (l) Otera's cat., MeOH, PhMe, 90 °C; N₂H₄, 95%; (m) Ac₂O, CH₂Cl₂, 70%; (n) HCl, MeOH, 52%.

General Reagents and Methods for Biological Assays. For biological assays, propidium iodide and phenazine methosulfate were purchased from Sigma–Aldrich. The 3-(4,5-dimethylthiazol-2-yl)-5-(3-carboxymethoxyphenyl)-2-(4-sulfophenyl)-2H-tetrazolium salt was obtained from Promega. Human erythrocytes were purchased from Bioreclamation and used within three days of receipt. Optical densities were recorded on a Spectramax Plus 384 (Molecular Devices, Sunnyvale, CA). Flow cytometry was performed on a BD Biosciences LSR II (San Jose, CA) and the data was analyzed as described using FACSDiva software (San Jose, CA).

Cell Culture Information. Cells were grown in media supplemented with fetal bovine serum (FBS) and antibiotics (100 µg/mL penicillin and 100 U/mL streptomycin). Specifically, experiments were performed using the following cell lines and media compositions: U-937, HeLa, H460, and 786-O (RPMI-1640 + 10% FBS), and MCF7 (EMEM + 10% FBS). Cells were incubated at 37 °C in a 5% CO₂, 95% humidity atmosphere.

IC₅₀ Value Determination for Adherent Cells using Sulforhodamine B (SRB). Adherent cells (HeLa, H460, 786-O, and MCF7) were added into 96-well plates (5,000 cells/well for HeLa cell line; 2,000 cells/well for H460, 786-O, and MCF7 cell lines) in 100 µL media and were allowed to adhere for 2-3 hours. Compounds were solubilized in DMSO as 100x stocks, added directly to the cells (100 µL final volume), and tested over a range of concentrations in triplicate (1% DMSO final) on a half-log scale. Concentrations tested ranged from 1 pM to 10 µM, depending on the potency of the compound. DMSO and cell-free wells served as the live and dead control, respectively. After 72 hours of continuous exposure, the plates were evaluated using the SRB colorimetric assay as described previously.¹⁰ Briefly, media was removed from the plate, and cells were fixed by the addition of 100 µL cold 10% trichloroacetic acid in water. After incubating at 4 °C for an hour, the plates were washed in water and allowed to dry. Sulforhodamine B was added as a 0.057% solution in 1% acetic acid (100 µL), and the plates were incubated at room temperature for 30 minutes, washed in 1% acetic acid, and allowed to dry. The dye was solubilized by adding 10 mM Tris base solution (pH 10.5, 200 µL) and incubating at room temperature for 30 minutes. Plates were read at λ = 510 nm. IC₅₀ values were determined from three or more independent experiments using TableCurve (San Jose, CA).

IC₅₀ Value Determination for Non-Adherent Cells using MTS. In a 96-well plate, compounds were pre-added as DMSO stocks in triplicate to achieve a final concentration of 1%. DMSO and cell-free wells served as the live and dead control, respectively. U-937 (5,000 cells/well) cells were distributed in 100 µL media to the compound-containing plate. After 72 hours, cell viability was assessed by adding 20 µL of a PMS/MTS solution¹¹ to each well, allowing the dye to develop at 37 °C until the live

¹⁰ Vichai, V.; Kirtikara, K. *Nature Prot.* **2006**, *1*, 1112.

¹¹ Cory, A. H.; Owen, T. C.; Barltrop, J. A.; Cory, J. G. *Cancer Commun.* **1991**, *3*, 207.

control had processed MTS, and reading the absorbance at $\lambda = 490$ nm. IC_{50} values were determined from three or more independent experiments using TableCurve (San Jose, CA).

Hemolysis Assay using Human Erythrocytes. To prepare the erythrocytes, 0.1 mL of human blood was centrifuged (10,000 g, 2 min). The pellet was washed three times with saline (0.9% NaCl) via gentle resuspension and centrifugation (10,000 g, 2 min). Following the final wash, the erythrocytes were resuspended in 0.8 mL red blood cell (RBC) buffer (10 mM Na_2HPO_4 , 150 mM NaCl, 1 mM $MgCl_2$, pH 7.4).

DMSO stocks of compounds were added to 0.5 mL tubes in singlicate (1 μ L, 3.3% DMSO final). The stocks were diluted with 19 μ L RBC buffer. Positive control tubes contained DMSO in water, and negative control tubes contained DMSO in RBC buffer. A suspension of washed erythrocytes (10 μ L) was added to each tube, and samples were incubated at 37 °C for 2 hours. Samples were centrifuged (10,000 g, 2 min), and the supernatant was transferred to a clear, sterile 384-well plate. The absorbance of the supernatants was measured at $\lambda = 540$ nm, and percent hemolysis was calculated relative to the average absorbance values measured for the controls.

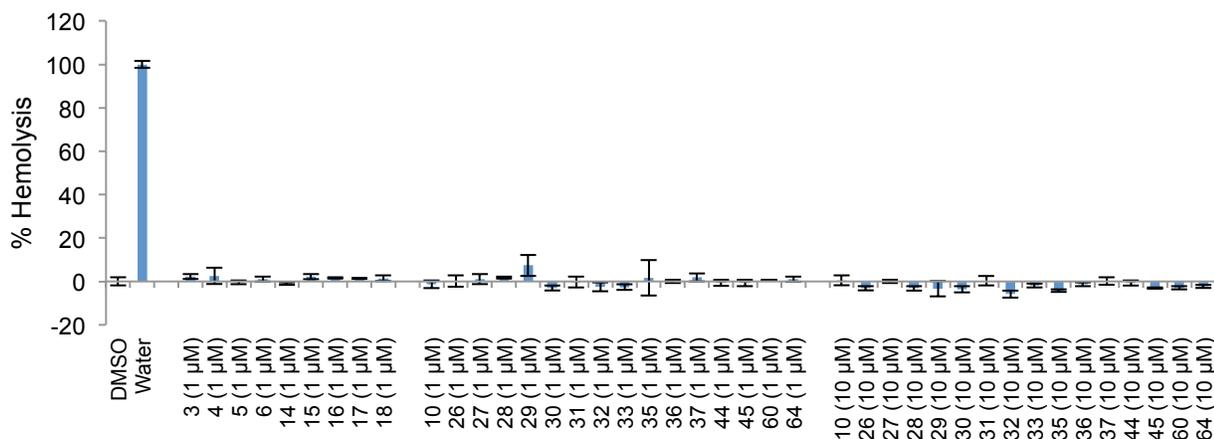


Figure S3. Percent hemolysis following treatment with ETPs from Table 2. Error bars represent standard error of the mean, $n \geq 3$.

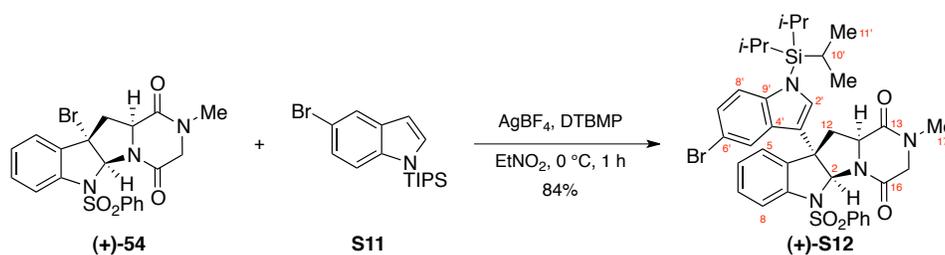
Apoptosis in U-937 Cells with Annexin V-FITC and Propidium Iodide (AnnV/PI). DMSO stocks of compounds were added to a 24-well plate in singlicate (0.2% DMSO final). After compound addition, 0.5 mL of a U-937 cell suspension (250,000 cells/mL) was added and allowed to incubate for 24 hours. Following treatment, the cell suspensions were transferred to flow cytometry tubes and pelleted (500 g, 3 min). The media was removed by aspiration, and cells were resuspended in 200 μ L AnnV binding buffer (10 mM HEPES, pH 7.4, 140 mM NaCl, 2.5 mM $CaCl_2$) with 5 μ g/mL PI and 1:90 dilution of AnnV. Samples were analyzed using flow cytometry.

Apoptosis in U-937 Cells by Western Blot Analysis. In a 24-well plate, compounds were added as DMSO stocks (0.2% DMSO final) in singlicate. After compound addition, 1.5 mL of a U-937 cell suspension (250,000 cells/mL) was added and allowed to incubate for 24 hours. The cell suspensions were transferred to 1.5 mL tubes and pelleted (600 g, 3 min). The media was removed via aspiration, and the cells were lysed by adding 40 μ L of RIPA buffer (50 mM Tris, pH 8.0, 150 mM NaCl, 1% TX-100, 0.5% sodium deoxycholate, 0.1% SDS) with 1% Protease Inhibitor Cocktail Set III. Each sample

was then vigorously vortexed twice for 15 seconds, with a 15-minute incubation on ice following each agitation. The cellular debris was pelleted (16,100 g, 5 min), and then 33 μ L of the protein suspension was transferred to fresh 0.5 mL tubes. The protein levels were quantified using a standard BCA (Thermo Scientific), after which the samples were diluted with deionized water to achieve equal protein concentrations for all samples.

Prior to analyzing the samples, 6x Laemmli sample buffer (350 mM Tris, pH 6.8, 12% SDS, 0.012% bromophenol blue, 47% glycerol) with 5% β -mercaptoethanol was added to each sample to achieve a final 1x concentration, after which the samples were incubated at 95 °C for 5 minutes to denature the protein samples. 20–30 μ g of protein was added to a 15-well 4–20% Tris-HCl gel and run for 1 hour at 120 V. The gel was equilibrated PBS (pH 7.4) for 5 minutes, and then transferred to a PVDF membrane for 2 hours at 45 V.

Generally, blots were probed as follows. The blot was blocked overnight at 4 °C with a blocking agent in 0.05% Tris-Buffered Saline Tween-20 (TBST) and then probed for the primary antibody at a 1:1000 dilution with a blocking agent in TBST overnight at 4 °C. The blot was washed with TBST, and then probed with a secondary rabbit HRP antibody (1:10,000, Cell Signaling) in TBST for 1 hour at room temperature. The blot was washed with TBST and PBS, and then visualized with Pico luminescent substrate kit (Thermo Scientific). Caspase 3 and PARP were blocked in 5% milk, and actin was blocked in 5% BSA.



C3-(5-Bromo-1-TIPS-indol-3'-yl)-pyrrolidinoindoline (+)-S12:

A round-bottom flask was charged with *endo*-tetracyclic bromide (+)-**54** (5.00 g, 10.5 mmol, 1 equiv), 2,6-di-*tert*-butyl-4-methylpyridine (DTBMP, 2.59 g, 12.6 mmol, 1.20 equiv), and 5-bromo-1-triisopropylsilyl-1*H*-indole¹² (**S11**, 14.8 g, 42.0 mmol, 4.00 equiv), and the mixture was dried azeotropically (concentration of a benzene solution, 2 × 30 mL) under reduced pressure and placed under an argon atmosphere. Anhydrous nitroethane (120 mL) was introduced via syringe, and the mixture was cooled to 0 °C in an ice–water bath. A solution of silver(I) tetrafluoroborate (6.30 g, 32.4 mmol, 3.09 equiv) in anhydrous nitroethane (40 mL) at 0 °C was introduced via cannula to the solution containing the tetracyclic bromide (+)-**54** over 20 min. After 5 min, a white precipitate was observed in the clear yellow reaction solution. The reaction flask was covered in aluminum foil, and the suspension was maintained at 0 °C. After 1 h, saturated aqueous sodium chloride solution (25 mL) was introduced, and the resulting biphasic mixture was vigorously stirred for 30 min at 0 °C. The reaction mixture was diluted with ethyl acetate (150 mL), was filtered through a Celite pad, and the solid was washed with ethyl acetate (3 × 50 mL). The combined filtrates were washed with 5% aqueous citric acid solution (2 × 100 mL), water (3 × 100 mL), and saturated aqueous sodium chloride solution (75 mL). The organic layer was dried over anhydrous sodium sulfate, was filtered, and was concentrated under reduced pressure. The resulting orange residue was purified by flash column chromatography (eluent: gradient, 2 → 10% acetone in dichloromethane) to afford the indole adduct (+)-**S12** (6.56 g, 83.6%) as a white foam. Structural assignments were made with additional information from gCOSY, HSQC, gHMBC, and NOESY data.

¹H NMR (600 MHz, CDCl₃, 20 °C):

δ 8.04 (app-d, *J* = 7.4, 2H, SO₂Ph-*o*-H), 7.77 (d, *J* = 8.3, 1H, C₈H), 7.56 (app-t, *J* = 7.5, 1H, SO₂Ph-*p*-H), 7.42 (app-dd, *J* = 7.8, 8.0, 2H, SO₂Ph-*m*-H), 7.30 (d, *J* = 8.9, 1H, C₈H), 7.29 (app-dt, *J* = 1.1, 7.9, 1H, C₇H), 7.15 (app-dd, *J* = 1.8, 8.8, 1H, C₇H), 6.98 (app-t, *J* = 7.5, 1H, C₆H), 6.94 (s, 1H, C₂H), 6.84 (d, *J* = 7.4, 1H, C₅H), 6.55 (d, *J* = 1.3, 1H, C₅H), 6.28 (s, 1H, C₂H), 4.47 (dd, *J* = 8.0, 9.5, 1H, C₁₁H), 4.07 (d, *J* = 17.8, 1H, C₁₅H_a), 3.94 (d, *J* = 17.8, 1H, C₁₅H_b), 3.03 (dd, *J* = 7.6, 13.8, 1H, C₁₂H_a), 3.00 (s, 3H, C₁₇H₃), 2.86 (dd, *J* = 10.0, 13.9, 1H, C₁₂H_b), 1.59 (app-sp, *J* = 7.5, 3H, C₁₀H), 1.08 (app-d, *J* = 8.5, 18H, C₁₁H).

¹² 5-Bromo-1-triisopropylsilyl-1*H*-indole **S11** was prepared in quantitative yield by silylation of commercially available 5-bromoindole using triisopropylsilyl chloride and sodium hydride in tetrahydrofuran. For preparation and characterization, see: Brown, D. A.; Mishra, M.; Zhang, S.; Biswas, S.; Parrington, I.; Antonio, T.; Reith, M. E. A.; Dutta, A. K. *Bioorg. Med. Chem.* **2009**, *17*, 3923.

^{13}C NMR (100 MHz, CDCl_3 , 20 °C):	δ 167.7 (C_{13}), 166.8 (C_{16}), 141.3 (C_9), 139.7 (C_9), 137.1 ($\text{SO}_2\text{Ph-}i\text{phso-C}$), 134.2 ($\text{SO}_2\text{Ph-}p\text{-C}$), 134.0 (C_4), 130.9 (C_2), 130.3 (C_4), 129.6 (C_7), 129.3 ($\text{SO}_2\text{Ph-}m\text{-C}$), 127.9 ($\text{SO}_2\text{Ph-}o\text{-C}$), 125.4 (C_7), 124.6 (C_6), 124.0 (C_5), 121.9 (C_5), 116.0 (C_8), 115.7 (C_8), 115.1 (C_3), 113.5 (C_6), 82.7 (C_2), 59.5 (C_{11}), 55.4 (C_3), 54.6 (C_{15}), 37.6 (C_{12}), 33.8 (C_{17}), 18.2 (C_{11}), 12.9 (C_{10}).
FTIR (thin film) cm^{-1} :	2949 (m), 2869 (m), 1681 (s), 1447 (m), 1396 (m), 1366 (m), 1178 (s), 1092 (w), 987 (w), 732 (m), 690 (w).
HRMS (ESI) (m/z):	calc'd for $\text{C}_{37}\text{H}_{44}\text{BrN}_4\text{O}_4\text{SSi}$ $[\text{M}+\text{H}]^+$: 747.2030, found: 747.2025.
$[\alpha]_{\text{D}}^{24}$:	+93.6 ($c = 0.26$, CHCl_3).
TLC (10% acetone in dichloromethane), R_f :	0.67 (UV, CAM).



C3-(indol-3'-yl)-pyrrolidinoindoline (+)-59:

A mixture of anhydrous methanol and ethyl acetate (3:2 v/v, 160 mL) was introduced into a round-bottom flask charged with the indole adduct (+)-**S12** (6.56 g, 8.77 mmol, 1 equiv) and palladium on activated charcoal (10% w/w, 0.50 g, 0.47 mmol, 0.05 equiv). The flask was purged by three cycles of vacuum and dihydrogen and sealed under an atmosphere of hydrogen gas (15 psi). Triethylamine (1.50 mL, 10.7 mmol, 1.22 equiv) was introduced to the flask via syringe, and the resulting suspension was vigorously stirred at 23 °C. Upon completion of the reaction (*ca* 8 h) as monitored by TLC, the flask was purged by three cycles of vacuum and argon and sealed under argon atmosphere. Neat triethylamine trihydrofluoride¹³ (3.00 mL, 18.4 mmol, 2.15 equiv) was introduced to the flask via syringe and the resulting suspension was stirred at 23 °C. After 13 h, the reaction mixture was filtered through a pad of Celite. The solids were washed with ethyl acetate (3 × 50 mL). The combined filtrates were concentrated under reduced pressure. The resulting pale yellow solid was diluted in ethyl acetate (400 mL) and washed sequentially with an aqueous hydrochloric acid solution (1 N, 2 × 100 mL), water (2 × 100 mL), and saturated aqueous sodium chloride solution (50 mL). The organic layer was dried over anhydrous sodium sulfate, was filtered, and was concentrated under reduced pressure. The resulting residue was purified by flash column chromatography (eluent: 15% acetone in dichloromethane) to afford the indole adduct (+)-**59** (4.59 g, 99.9%) as a white solid. Structural assignments were made with additional information from gCOSY, HSQC, gHMBC, and NOESY data.

¹H NMR (600 MHz, CDCl₃, 20 °C):

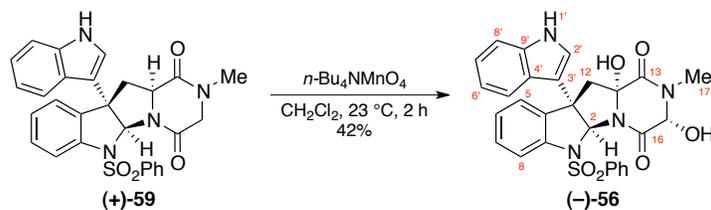
δ 8.03 (br-s, 1H, N₁H), 7.75 (d, *J* = 8.2, 1H, C₈H), 7.50 (d, *J* = 7.6, 2H, SO₂Ph-*o*-H), 7.38 (t, *J* = 7.5, 1H, SO₂Ph-*p*-H), 7.35 (d, *J* = 8.2, 1H, C₈H), 7.30 (app-dt, *J* = 1.1, 7.8, 1H, C₇H), 7.19 (app-t, *J* = 7.6, 1H, C₇H), 7.10 (app-t, *J* = 7.9, 2H, SO₂Ph-*m*-H), 7.09–7.06 (m, 1H, C₅H), 7.06 (app-t, *J* = 7.4, 1H, C₆H), 6.93 (app-t, *J* = 7.4, 1H, C₆H), 6.89 (d, *J* = 7.9, 1H, C₅H), 6.37 (s, 1H, C₂H), 6.16 (d, *J* = 2.3, 1H, C₂H), 4.56 (app-t, *J* = 8.1, 1H, C₁₁H), 4.13 (d, *J* = 17.5, 1H, C₁₅H_a), 3.85 (d, *J* = 17.5, 1H, C₁₅H_b), 3.09 (dd, *J* = 8.9, 14.1, 1H, C₁₂H_a), 3.03 (dd, *J* = 7.2, 14.1, 1H, C₁₂H_b), 2.90 (s, 3H, C₁₇H₃).

¹³C NMR (150 MHz, CDCl₃, 20 °C):

δ 167.5 (C₁₃), 165.9 (C₁₆), 139.6 (C₉), 137.6 (SO₂Ph-*ipso*-C), 137.4 (C₉), 135.9 (C₄), 133.1 (SO₂Ph-*p*-C), 129.3 (C₇), 128.6 (SO₂Ph-*m*-C), 127.6 (SO₂Ph-*o*-C), 125.2 (C₆), 124.8 (C₅), 124.6 (C₄), 123.6 (C₂), 122.9 (C₇), 120.3 (C₆), 119.0 (C₅), 117.1 (C₈), 115.0 (C₃), 112.0 (C₈), 83.8 (C₂), 58.8 (C₁₁), 55.4 (C₃), 54.6 (C₁₅), 36.1 (C₁₂), 33.8 (C₁₇).

¹³ McClinton, M. A. *Aldrichimica Acta* **1995**, 28, 31.

FTIR (thin film) cm^{-1} :	3384 (br-m), 3013 (w), 2925 (w), 1681 (s), 1457 (m), 1399 (m), 1355 (m), 1169 (m), 1091 (w), 751 (m).
HRMS (ESI) (m/z):	calc'd for $\text{C}_{28}\text{H}_{24}\text{N}_4\text{NaO}_4\text{S}$ $[\text{M}+\text{Na}]^+$: 535.1410, found: 535.1413.
$[\alpha]_{\text{D}}^{23}$:	+70.0 ($c = 0.15$, CHCl_3).
TLC (25% acetone in dichloromethane), R_f :	0.41 (UV, CAM).



C3-(Indol-3'-yl) hexacyclic diol (–)-56:

Freshly prepared tetra-*n*-butylammonium permanganate^{14,15,16} (767 mg, 2.12 mmol, 3.79 equiv) was added as a solid to a solution of the indole adduct (+)-59 (287 mg, 0.56 mmol, 1 equiv) in dichloromethane (20 mL) at 23 °C. After 30 min, the dark purple solution was diluted with saturated aqueous sodium sulfite solution (20 mL) and then with ethyl acetate (160 mL). The resulting mixture was washed sequentially with saturated aqueous sodium hydrogenocarbonate solution (50 mL), water (2 × 50 mL), and saturated aqueous sodium chloride solution (30 mL). The aqueous layer was extracted with ethyl acetate (2 × 100 mL), and the combined organic layers were dried over anhydrous sodium sulfate, were filtered, and were concentrated under reduced pressure. The resulting yellow residue was purified by flash column chromatography (eluent: gradient, 10 → 25% acetone in dichloromethane) to afford the diol (–)-56 (127 mg, 41.6%) as a white solid.¹⁷ Structural assignments were made with additional information from gCOSY, HSQC, gHMBC, and NOESY data.

¹H NMR (600 MHz, acetone-*d*₆, 20 °C):

δ 9.85 (br-s, 1H, N₁H), 8.01 (d, *J* = 8.2, 1H, C₅H), 7.56 (d, *J* = 8.1, 1H, C₈H), 7.49 (d, *J* = 8.1, 1H, C₈H), 7.41 (d, *J* = 7.5, 1H, C₅H), 7.35 (app-t, *J* = 7.5, 1H, SO₂Ph-*p*-H), 7.35 (app-t, *J* = 7.5, 1H, C₇H), 7.24 (app-t, *J* = 7.6, 1H, C₇H), 7.20 (app-t, *J* = 7.5, 1H, C₆H), 7.17 (app-t, *J* = 7.5, 1H, C₆H), 7.04 (d, *J* = 7.5, 2H, SO₂Ph-*o*-H), 6.98 (app-t, *J* = 7.8, 2H, SO₂Ph-*m*-H), 6.80 (d, *J* = 6.2, 1H, C₁₅OH), 6.66 (s, 1H, C₂H), 6.22 (s, 1H, C₁₁OH), 5.65 (d, *J* = 2.5, 1H, C₂H), 5.15 (d, *J* = 6.0, 1H, C₁₅H), 3.64 (d, *J* = 15.1, 1H, C₁₂H_a), 3.01 (d, *J* = 15.1, 1H, C₁₂H_b), 2.95 (s, 3H, C₁₇H₃).

¹³C NMR (150 MHz, acetone-*d*₆, 20 °C):

δ 168.1 (C₁₃), 165.7 (C₁₆), 140.4 (C₉), 139.3 (SO₂Ph-*ipso*-C), 138.8 (C₄), 138.6 (C₉), 133.7 (SO₂Ph-*p*-C), 129.8 (C₇), 128.9 (SO₂Ph-*m*-C), 127.5 (SO₂Ph-*o*-C), 126.3 (C₅), 126.2 (C₆), 125.7 (C₂), 125.2 (C₄), 122.9 (C₇), 120.4 (C₆), 119.6 (C₅), 118.2 (C₈), 115.7 (C₃), 113.0 (C₈), 88.6 (C₁₁), 85.3 (C₂), 83.9 (C₁₅), 55.3 (C₃), 45.1 (C₁₂), 31.8 (C₁₇).

FTIR (thin film) cm⁻¹:

3392 (br-m), 1700 (s), 1460 (w), 1400 (w), 1360 (m), 1169 (m), 1091 (w), 750 (w).

¹⁴ Sala, T.; Sargent, M. V. *J. Chem. Soc., Chem. Commun.* **1978**, 253.

¹⁵ Tetra-*n*-butylammonium permanganate was prepared according to a literature procedure (Karaman, H.; Barton, R. J.; Robertson, B. E.; Lee, D. G. *J. Org. Chem.* **1984**, *49*, 4509) and dried under reduced pressure at room temperature.

¹⁶ (a) Gardner, K. A.; Mayer, J. M. *Science* **1995**, *269*, 1849. (b) Strassner, T.; Houk, K. N. *J. Am. Chem. Soc.* **2000**, *122*, 7821. (c) Shi, S.; Wang, Y.; Xu, A.; Wang, H.; Zhu, D.; Roy, S. B.; Jackson, T. A.; Busch, D. H.; Yin, G. *Angew. Chem. Int. Ed.* **2011**, *50*, 7321.

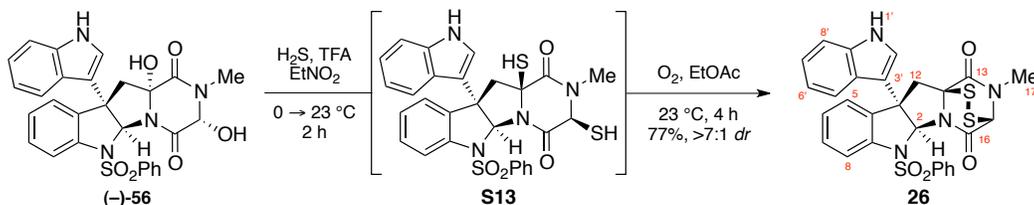
¹⁷ Analytically pure samples of polar diol (–)-56 could be obtained by trituration with minimal amount of chloroform.

HRMS (ESI) (m/z): calc'd for $C_{28}H_{24}N_4NaO_6S$ $[M+Na]^+$: 567.1309,
found: 567.1315.

$[\alpha]_D^{24}$: -71.4 ($c = 0.114$, acetone).

m.p.: 212 °C.

TLC (20% acetone in dichloromethane), R_f : 0.24 (UV, CAM).



C3-(Indol-3'-yl) epidithiodiketopiperazine 26:

A slow stream of hydrogen sulfide gas was introduced into a solution of diol (–)-**56** (254 mg, 466 μmol , 1 equiv) in anhydrous nitroethane (20 mL) at 0 °C, providing a saturated hydrogen sulfide solution. After 20 min, trifluoroacetic acid (TFA, 15 mL) was added slowly via syringe, and the slow introduction of hydrogen sulfide into the mixture was maintained for another 20 min. The reaction mixture was left under an atmosphere of hydrogen sulfide. The ice–water bath was removed, and the yellow solution was allowed to warm to 23 °C. After 2 h, a slow stream of argon gas was introduced into the solution. After 15 min, the reaction mixture was diluted with ethyl acetate (150 mL) and slowly poured into saturated aqueous sodium hydrogenocarbonate solution (70 mL) at 23 °C. The organic layer was sequentially washed with water (3 \times 40 mL) and saturated aqueous sodium chloride solution (25 mL), was dried over anhydrous sodium sulfate, was filtered, and was concentrated under reduced pressure to afford the corresponding bithiol **S13** that was used in the next step without further purification.

The orange residue was dissolved in ethyl acetate (120 mL). A slow stream of dioxygen gas was introduced into the solution. After 4 h, the yellow solution was concentrated under reduced pressure. The orange residue was purified by flash column chromatography on silica gel (eluent: gradient, 5 \rightarrow 15% ethyl acetate in dichloromethane) to afford the epidithiodiketopiperazine **26** (205 mg, 76.7%) as a white solid. Structural assignments were made with additional information from gCOSY, HSQC, and gHMBC data.

^1H NMR (600 MHz, acetone- d_6 , 20 °C):

δ 10.05 (br-s, 1H, N_1H), 7.65 (d, $J = 8.1$, 1H, C_8H), 7.55 (d, $J = 7.5$, 1H, C_5H), 7.50 (d, $J = 8.0$, 1H, C_5H), 7.48 (d, $J = 8.8$, 1H, C_8H), 7.46 (app-dt, $J = 1.0, 7.5$, 1H, C_7H), 7.39 (t, $J = 7.4$, 1H, $\text{SO}_2\text{Ph-}p\text{-H}$), 7.30 (app-t, $J = 0.8, 7.5$, 1H, C_6H), 7.22 (dd, $J = 7.2, 8.0$, 1H, C_7H), 7.12 (app-dd, $J = 1.0, 8.4$, 2H, $\text{SO}_2\text{Ph-}o\text{-H}$), 7.10 (dd, $J = 7.3, 7.9$, 1H, C_6H), 7.00 (dd, $J = 7.5, 8.2$, 2H, $\text{SO}_2\text{Ph-}m\text{-H}$), 6.63 (s, 1H, C_2H), 5.98 (d, $J = 2.6$, 1H, C_2H), 5.80 (s, 1H, C_{15}H), 3.95 (d, $J = 15.6$, 1H, C_{12}H_a), 3.17 (s, 3H, C_{17}H_3), 2.92 (d, $J = 15.7$, 1H, C_{12}H_b).

^{13}C NMR (150 MHz, acetone- d_6 , 20 °C):

δ 165.9 (C_{13}), 161.0 (C_{16}), 141.5 (C_9), 138.7 ($\text{SO}_2\text{Ph-}ipso\text{-C}$), 138.5 (C_9), 138.1 (C_4), 134.0 ($\text{SO}_2\text{Ph-}p\text{-C}$), 130.1 (C_7), 129.0 ($\text{SO}_2\text{Ph-}m\text{-C}$), 127.7 ($\text{SO}_2\text{Ph-}o\text{-C}$), 126.6 (C_6), 125.9 (C_5), 125.8 (C_2), 125.0 (C_4), 123.0 (C_7), 120.6 (C_6), 119.2 (C_8), 119.1 (C_5), 114.1 (C_3), 113.1 (C_8), 85.7 (C_2), 75.5 (C_{11}), 69.1 (C_{15}), 56.4 (C_3), 42.6 (C_{12}), 31.8 (C_{17}).

FTIR (thin film) cm^{-1} : 3392 (w), 3060 (w), 2990 (w), 1693 (s), 1447 (w), 1358 (m), 1234 (w), 1169 (m), 1089 (w), 1052 (w), 964 (w), 736 (m), 587 (m).

HRMS (ESI) (m/z): calc'd for $\text{C}_{28}\text{H}_{23}\text{N}_4\text{O}_4\text{S}_3$ $[\text{M}+\text{H}]^+$: 575.0876, found 575.0885; calc'd for $\text{C}_{28}\text{H}_{22}\text{N}_4\text{NaO}_4\text{S}_3$ $[\text{M}+\text{Na}]^+$: 597.0695, found 597.0704.

TLC (20% ethyl acetate in dichloromethane), R_f : 0.62 (UV, CAM).

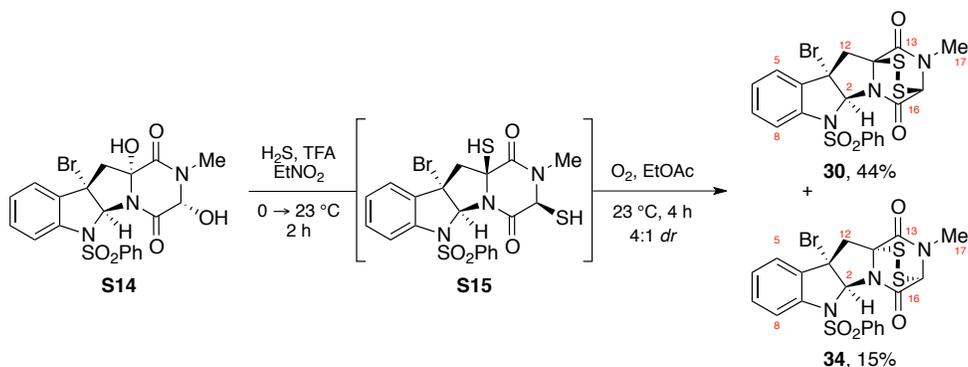
General Procedure for the Friedel–Crafts Nucleophilic Substitution. A round-bottom flask was charged with *endo*-tetracyclic bromide (+)-**54**⁹ (1 equiv), 2,6-di-*tert*-butyl-4-methylpyridine (DTBMP, 2.10 equiv), and the nucleophile (for **68**¹⁸: tetrafluoroborate as nucleophilic fluorine source, for **69**¹⁹: 1-(triisopropylsilyl)-*1H*-pyrrole, for **70**²⁰: anisole), and the mixture was dried azeotropically (concentration of an anhydrous benzene solution, 2 × 10 mL) under reduced pressure and placed under an argon atmosphere. Anhydrous nitroethane (4 mL) was introduced via syringe, and the mixture was cooled to 0 °C in an ice–water bath. A solution of silver(I) tetrafluoroborate (2.30 equiv) in anhydrous nitroethane (1 mL) at 0 °C was introduced via syringe to the solution containing the tetracyclic bromide (+)-**54** over 1 min. The reaction flask was covered in aluminum foil. The ice–water bath was removed, and the reaction mixture was allowed to warm to 23 °C. After 1 h, saturated aqueous sodium chloride solution (10 mL) was introduced, and the resulting biphasic mixture was vigorously stirred for 30 min at 23 °C. The reaction mixture was diluted with ethyl acetate (50 mL), was filtered through a Celite pad, and the solids were washed with ethyl acetate (3 × 15 mL). The combined filtrates were washed with 5% aqueous citric acid solution (2 × 20 mL), water (3 × 20 mL), and saturated aqueous sodium chloride solution (15 mL). The organic layer was dried over anhydrous sodium sulfate, was filtered, and was concentrated under reduced pressure.

General Procedure for the Regio- and Stereoselective Hydroxylation. Freshly prepared tetra-*n*-butylammonium permanganate (4.0 equiv) was added as a solid to a solution of the corresponding diketopiperazine (**54**, **68–70**) (1 equiv) in dichloromethane (0.05 M) at 23 °C. After 2 h, the dark purple solution was diluted with saturated aqueous sodium sulfite solution (20 mL) and then with ethyl acetate (120 mL). The resulting mixture was washed sequentially with saturated aqueous sodium hydrogenocarbonate solution (20 mL), water (4 × 20 mL), and saturated aqueous sodium chloride solution (20 mL). The organic layer was dried over anhydrous sodium sulfate, was filtered, and was concentrated under reduced pressure.

¹⁸ C3-Fluoro Friedel–Crafts adduct **68**: ¹H NMR (600 MHz, CDCl₃, 20 °C): δ 7.79 (app-dd, *J* = 0.9, 8.2, 2H, SO₂Ph-*o*-H), 7.56 (d, *J* = 8.2, 1H, C₈H), 7.50 (t, *J* = 7.5, 1H, SO₂Ph-*p*-H), 7.39–7.35 (m, 1H, C₇H), 7.38 (dd, *J* = 7.9, 8.3, 2H, SO₂Ph-*m*-H), 7.34 (d, *J* = 7.7, 1H, C₅H), 7.15 (dd, *J* = 7.5, 7.6, 1H, C₆H), 6.07 (d, *J* = 14.5, 1H, C₂H), 4.53 (dd, *J* = 8.2, 8.4, 1H, C₁₁H), 4.17 (d, *J* = 17.6, 1H, C₁₅H_a), 3.86 (d, *J* = 17.6, 1H, C₁₅H_b), 3.06–2.97 (m, 1H, C₁₂H_a), 2.93–2.83 (m, 1H, C₁₂H_b), 2.90 (s, 3H, C₁₇H₃). ¹⁹F NMR (282.4 MHz, CDCl₃, 20 °C): δ -133.3. MS (ESI) (*m/z*): [M+H]⁺: 416.22, [M+Na]⁺: 438.25, [2M+H]⁺: 833.73, [2M+Na]⁺: 853.59. TLC (20% acetone in dichloromethane), R_f: 0.46 (UV, CAM).

¹⁹ C3-(*N*-TIPS-Pyrrol-3'-yl) Friedel–Crafts adduct **69**: ¹H NMR (600 MHz, CDCl₃, 20 °C): δ 8.03 (app-dd, *J* = 1.0, 7.3, 2H, SO₂Ph-*o*-H), 7.63 (d, *J* = 7.7, 1H, C₈H), 7.54 (app-dt, *J* = 1.5, 7.5, 1H, SO₂Ph-*p*-H), 7.43 (app-t, *J* = 7.6, 2H, SO₂Ph-*m*-H), 7.16–7.11 (m, 1H, C₇H), 7.05–6.99 (m, 2H, C₅H + C₆H), 6.69–6.65 (m, 1H, C₅H), 6.53–5.49 (m, 1H, C₄H), 6.09 (s, 1H, C₂H), 5.83–5.79 (m, 1H, C₂H), 4.33 (dd, *J* = 8.2, 8.9, 1H, C₁₁H), 4.10 (d, *J* = 17.8, 1H, C₁₅H_a), 3.95 (app-dd, *J* = 2.0, 17.6, 1H, C₁₅H_b), 2.99 (s, 3H, C₁₇H₃), 2.84 (dd, *J* = 7.4, 13.3, 1H, C₁₂H_a), 2.73 (dd, *J* = 10.0, 13.3, 1H, C₁₂H_b), 1.40 (app-dsp, *J* = 1.6, 7.5, 3H, SiCH(CH₃)₂), 1.08 (d, *J* = 7.6, 9H, SiCH(CH₃)₂), 1.07 (d, *J* = 6.3, 9H, SiCH(CH₃)₂). ¹³C NMR (150 MHz, CDCl₃, 20 °C): δ 167.7 (C₁₃), 166.8 (C₁₆), 139.5 (C₉), 137.6 (SO₂Ph-*ipso*-C), 135.9 (C₄), 133.4 (SO₂Ph-*p*-C), 129.0 (SO₂Ph-*m*-C), 128.9 (C₇), 128.2 (SO₂Ph-*o*-C), 125.7 (C₅), 125.0 (C₃), 124.6 (C₆), 124.0 (C₅), 121.2 (C₂), 115.6 (C₈), 109.4 (C₄), 84.8 (C₂), 59.5 (C₁₁), 55.3 (C₃), 54.5 (C₁₅), 39.6 (C₁₂), 33.6 (C₁₇), 17.9 (SiCH(CH₃)₂), 11.7 (SiCH(CH₃)₂). MS (ESI) (*m/z*): [M+H]⁺: 619.49, [M+Na]⁺: 641.49, [2M+Na]⁺: 1261.37. TLC (20% acetone in dichloromethane), R_f: 0.48 (UV, CAM).

²⁰ C3-(*p*-Methoxyphenyl) Friedel–Crafts adduct **70**: ¹H NMR (600 MHz, CDCl₃, 20 °C): δ 7.60 (app-dd, *J* = 0.7, 8.1, 1H, C₈H), 7.46 (app-dd, *J* = 1.1, 8.4, 2H, SO₂Ph-*o*-H), 7.34 (app-dt, *J* = 1.1, 7.5, 1H, SO₂Ph-*p*-H), 7.30–7.26 (m, 1H, C₇H), 7.14–7.11 (m, 2H, C₅H + C₆H), 7.11 (app-t, *J* = 7.5, 2H, SO₂Ph-*m*-H), 6.67 (d, *J* = 8.9, 2H, C₂H), 6.63 (d, *J* = 8.9, 1H, C₃H), 6.15 (s, 1H, C₂H), 4.42 (dd, *J* = 7.6, 8.2, 1H, C₁₁H), 4.12 (d, *J* = 17.5, 1H, C₁₅H_a), 3.84 (d, *J* = 17.5, 1H, C₁₅H_b), 3.78 (s, 3H, C₅H₃), 3.10 (dd, *J* = 6.8, 14.2, 1H, C₁₂H_a), 2.91–2.85 (m, 1H, C₁₂H_b), 2.90 (s, 3H, C₁₇H₃). MS (ESI) (*m/z*): [M+Na]⁺: 526.31, [2M+Na]⁺: 1029.94. TLC (20% acetone in dichloromethane), R_f: 0.37 (UV, CAM).



C3-Bromo epidithiodiketopiperazines **30** and **34**:

This compound was prepared in two steps starting from bishemiaminal **S14**²¹ (13.5 mg, 26.6 μmol)²² using the methodology developed to access the corresponding C3-(indol-3'-yl) epidithiodiketopiperazine **26**²³. The orange residue was purified by flash column chromatography on silica gel (eluent: gradient, 15 \rightarrow 40% ethyl acetate in dichloromethane) to afford the β -epimer of epidithiodiketopiperazine **30** (6.3 mg, 44%) as a colorless oil and its α -epimer **34** (2.1 mg, 15%) as a colorless oil. Structural assignments were made with additional information from gCOSY, HSQC, and gHMBC data.

β -epimer **30**:²⁴

¹H NMR (600 MHz, CDCl₃, 20 °C):

δ 7.82 (d, $J = 8.0$, 2H, SO₂Ph-*o*-H), 7.60 (d, $J = 8.2$, 1H, C₈H), 7.52 (app-dd, $J = 7.4$, 7.6, 1H, SO₂Ph-*p*-H), 7.42–7.38 (m, 1H, C₇H), 7.40 (app-t, $J = 7.7$, 2H, SO₂Ph-*m*-H), 7.35 (d, $J = 7.7$, 1H, C₅H), 7.25 (app-t, $J = 7.6$, 1H, C₆H), 6.47 (s, 1H, C₂H), 5.22 (s, 1H, C₁₅H), 3.82 (d, $J = 15.4$, 1H, C₁₂H_a), 3.19 (d, $J = 15.4$, 1H, C₁₂H_b), 3.11 (s, 3H, C₁₇H₃).

¹³C NMR (150 MHz, CDCl₃, 20 °C):

δ 164.3 (C₁₃), 159.6 (C₁₆), 140.2 (C₉), 138.1 (SO₂Ph-*ipso*-C), 135.1 (C₄), 134.1 (SO₂Ph-*p*-C), 131.5 (C₇), 129.2 (SO₂Ph-*m*-C), 128.3 (SO₂Ph-*o*-C), 127.1 (C₆), 124.3 (C₅), 118.9 (C₈), 87.4 (C₂), 74.0 (C₁₁), 68.3 (C₁₅), 58.2 (C₃), 46.7 (C₁₂), 32.3 (C₁₇).

²¹ **S14**: ¹H NMR (600 MHz, MeOD-*d*₄, 20 °C): δ 7.89 (app-dd, $J = 0.8$, 8.2, 2H, SO₂Ph-*o*-H), 7.56 (t, $J = 7.5$, 1H, SO₂Ph-*p*-H), 7.47 (d, $J = 8.3$, 1H, C₈H), 7.44 (dd, $J = 7.5$, 8.2, 2H, SO₂Ph-*m*-H), 7.38 (d, $J = 7.7$, 1H, C₅H), 7.33 (app-dt, $J = 1.0$, 7.7, 1H, C₇H), 7.16 (app-dt, $J = 0.6$, 7.5, 1H, C₆H), 6.55 (s, 1H, C₂H), 4.99 (s, 1H, C₁₅H), 3.71 (d, $J = 15.4$, 1H, C₁₂H_a), 3.09 (d, $J = 15.4$, 1H, C₁₂H_b), 2.86 (s, 3H, C₁₇H₃). MS (ESI) (m/z): [2M+Na]⁺: 1039.24. TLC (20% acetone in dichloromethane), R_f: 0.40 (UV, CAM).

²² Please see pages S14 and S18 for experimental details.

²³ Please see page S16 for experimental details.

²⁴ The relative stereochemistry of the epidisulfide bridge of the β -epimer **30** has been confirmed by key NOESY cross-peaks on the corresponding bis(methylthioether). Our assignment is supported by key NOESY signals (¹H,¹H) in ppm: (1.86,3.40), (3.40,7.36), (3.11,6.68). This derivatized compound was prepared in one step using the methodology developed to access (+)-gliocladin B (see reference 9). ¹H NMR (600 MHz, CDCl₃, 20 °C): δ 7.82 (d, $J = 8.1$, 2H, SO₂Ph-*o*-H), 7.59 (d, $J = 8.2$, 1H, C₈H), 7.55 (app-dd, $J = 7.3$, 7.6, 1H, SO₂Ph-*p*-H), 7.45 (dd, $J = 7.7$, 7.8, 2H, SO₂Ph-*m*-H), 7.39 (app-t, $J = 7.9$, 1H, C₇H), 7.36 (d, $J = 7.8$, 1H, C₅H), 7.19 (app-t, $J = 7.6$, 1H, C₆H), 6.68 (s, 1H, C₂H), 4.52 (s, 1H, C₁₅H), 3.40 (d, $J = 14.5$, 1H, C₁₂H_a), 3.11 (d, $J = 14.5$, 1H, C₁₂H_b), 3.06 (s, 3H, C₁₇H₃), 2.27 (s, 3H, C₁₅SCH₃), 1.86 (s, 3H, C₁₁SCH₃). ¹³C NMR (150 MHz, CDCl₃, 20 °C): δ 164.0 (C₁₃), 163.5 (C₁₆), 140.5 (C₉), 138.8 (SO₂Ph-*ipso*-C), 137.2 (C₄), 133.6 (SO₂Ph-*p*-C), 131.1 (C₇), 129.3 (SO₂Ph-*m*-C), 127.6 (SO₂Ph-*o*-C), 125.9 (C₆), 123.8 (C₅), 117.9 (C₈), 86.8 (C₂), 69.7 (C₁₁), 67.3 (C₁₅), 58.0 (C₃), 49.9 (C₁₂), 32.7 (C₁₇), 17.2 (C₁₅SCH₃), 15.4 (C₁₁SCH₃).

FTIR (thin film) cm^{-1} : 2926 (m), 2857 (w), 1771 (m), 1697 (s), 1551 (w), 1449 (m), 1368 (s), 1170 (s), 1090 (w), 1055 (w), 756 (s).

HRMS (ESI) (m/z): calc'd for $\text{C}_{20}\text{H}_{16}\text{BrN}_3\text{NaO}_4\text{S}_3$ $[\text{M}+\text{Na}]^+$: 559.9379, found 559.9392.

TLC (20% ethyl acetate in dichloromethane), R_f : 0.47 (UV, I_2 , CAM).

α -epimer 34:

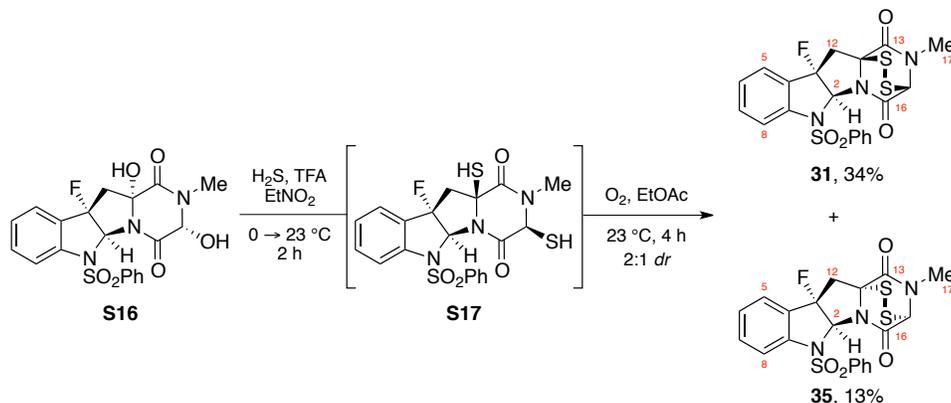
^1H NMR (600 MHz, CDCl_3 , 20 °C): δ 7.91 (d, $J = 8.1$, 2H, $\text{SO}_2\text{Ph-}o\text{-H}$), 7.53–7.51 (m, 1H, $\text{SO}_2\text{Ph-}p\text{-H}$), 7.52 (d, $J = 7.9$, 1H, C_8H), 7.41 (app-t, $J = 7.7$, 2H, $\text{SO}_2\text{Ph-}m\text{-H}$), 7.38 (d, $J = 7.9$, 1H, C_5H), 7.32 (dd, $J = 7.6$, 8.0, 1H, C_7H), 7.17 (app-t, $J = 7.6$, 1H, C_6H), 6.61 (s, 1H, C_2H), 5.16 (s, 1H, C_{15}H), 4.25 (d, $J = 15.0$, 1H, C_{12}H_a), 3.09 (d, $J = 15.0$, 1H, C_{12}H_b), 2.95 (s, 3H, C_{17}H_3).

^{13}C NMR (150 MHz, CDCl_3 , 20 °C): δ 164.1 (C_{13}), 160.9 (C_{16}), 138.8 (C_9), 138.2 ($\text{SO}_2\text{Ph-}ipso\text{-C}$), 134.1 ($\text{SO}_2\text{Ph-}p\text{-C}$), 133.6 (C_4), 131.5 (C_7), 129.2 ($\text{SO}_2\text{Ph-}m\text{-C}$), 128.4 ($\text{SO}_2\text{Ph-}o\text{-C}$), 126.8 (C_6), 125.2 (C_5), 117.7 (C_8), 87.7 (C_2), 73.8 (C_{11}), 68.9 (C_{15}), 58.2 (C_3), 45.0 (C_{12}), 31.9 (C_{17}).

FTIR (thin film) cm^{-1} : 3296 (w), 3008 (m), 2925 (s), 2855 (s), 1771 (m), 1699 (s), 1552 (m), 1463 (s), 1447 (s), 1368 (s), 1171 (s), 1091 (s), 1057 (m), 757 (s).

HRMS (ESI) (m/z): calc'd for $\text{C}_{20}\text{H}_{16}\text{BrN}_3\text{NaO}_4\text{S}_3$ $[\text{M}+\text{Na}]^+$: 559.9379, found 559.9396.

TLC (20% ethyl acetate in dichloromethane), R_f : 0.56 (UV, I_2 , CAM).



C3-Fluoro epidithiodiketopiperazines **31** and **35**:

This compound was prepared in two steps starting from bishemiaminal **S16**²⁵ (15.1 mg, 33.7 μmol)²² using the methodology developed to access the corresponding C3-(indol-3'-yl) epidithiodiketopiperazine **26**²³. The orange residue was purified by flash column chromatography on silica gel (eluent: gradient, 15 \rightarrow 40% ethyl acetate in dichloromethane) to afford the β -epimer of epidithiodiketopiperazine **31** (5.4 mg, 34%) as a colorless oil and its α -epimer **35** (2.1 mg, 13%) as a colorless oil. Structural assignments were made with additional information from gCOSY, HSQC, and gHMBC data.

β -epimer **31**:²⁶

¹H NMR (600 MHz, CDCl₃, 20 °C):

δ 7.68 (app-dd, $J = 1.1, 7.4$, 2H, SO₂Ph-*o*-H), 7.64 (d, $J = 8.2$, 1H, C₈H), 7.51 (t, $J = 7.5$, 1H, SO₂Ph-*p*-H), 7.50 (app-dt, $J = 1.1, 6.7$, 1H, C₇H), 7.38 (dd, $J = 7.6, 8.1$, 2H, SO₂Ph-*m*-H), 7.40–7.36 (m, 1H, C₆H), 7.28 (d, $J = 7.6$, 1H, C₅H), 6.31 (d, $J = 11.8$, 1H, C₂H), 5.23 (s, 1H, C₁₅H), 3.65 (app-t, $J = 15.2$,

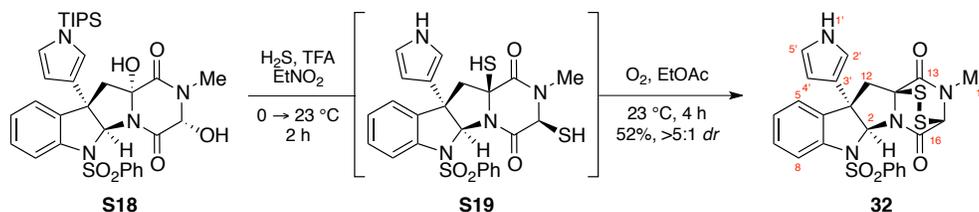
²⁵ **S16**: ¹H NMR (600 MHz, CDCl₃, 20 °C): δ 7.68 (d, $J = 7.6$, 2H, SO₂Ph-*o*-H), 7.56 (d, $J = 8.2$, 1H, C₈H), 7.50 (t, $J = 8.2$, 1H, SO₂Ph-*p*-H), 7.43 (dd, $J = 7.6, 8.2$, 1H, C₇H), 7.36 (app-t, $J = 7.9$, 2H, SO₂Ph-*m*-H), 7.36–7.33 (m, 1H, C₅H), 7.18 (app-t, $J = 7.5$, 1H, C₆H), 6.44 (d, $J = 13.2$, 1H, C₂H), 5.75 (br-s, 2H, C₁₁OH + C₁₅OH), 5.13 (s, 1H, C₁₅H), 3.49 (dd, $J = 8.6, 15.6$, 1H, C₁₂H_a), 3.02 (s, 3H, C₁₇H₃), 2.97 (dd, $J = 15.6, 20.8$, 1H, C₁₂H_b). ¹³C NMR (100 MHz, CDCl₃, 20 °C): δ 166.7 (C₁₃), 166.4 (C₁₆), 141.8 (d, $J = 4.6$, C₉), 136.8 (SO₂Ph-*ipso*-C), 134.0 (SO₂Ph-*p*-C), 132.2 (d, $J = 3.2$, C₇), 130.6 (d, $J = 23.5$, C₄), 129.2 (SO₂Ph-*m*-C), 128.0 (SO₂Ph-*o*-C), 126.8 (C₆), 125.5 (C₅), 118.5 (C₈), 101.7 (d, $J = 202.3$, C₃), 88.5 (d, $J = 4.1$, C₁₁), 83.1 (d, $J = 33.0$, C₂), 83.0 (C₁₅), 42.9 (d, $J = 29.7$, C₁₂), 32.6 (C₁₇). ¹⁹F NMR (282.4 MHz, CDCl₃, 20 °C): δ -133.2. FTIR (thin film) cm⁻¹: 3365 (br-m), 1695 (br-s), 1447 (m), 1402 (m), 1365 (m), 1342 (m), 1173 (m), 1087 (w), 1023 (w), 912 (w), 729 (m), 600 (m). HRMS (ESI) (m/z): calc'd for C₂₀H₁₉FN₃O₆S [M+H]⁺: 448.0973, found 448.0963; calc'd for C₂₀H₁₈FN₃NaO₆S [M+Na]⁺: 470.0793, found 470.0780. TLC (20% acetone in dichloromethane), R_f: 0.29 (UV, CAM).

²⁶ The relative stereochemistry of the epidisulfide bridge of the β -epimer **31** has been confirmed by key NOESY cross-peaks on the corresponding bis(methylthioether). Our assignment is supported by key NOESY signals (¹H,¹H) in ppm: (1.93,3.11), (3.11,7.41), (2.94,6.45). This derivatized compound was prepared in one step using the methodology developed to access (+)-gliocladin B (see reference 9). ¹H NMR (600 MHz, CDCl₃, 20 °C): δ 7.94 (d, $J = 8.0$, 2H, SO₂Ph-*o*-H), 7.72 (d, $J = 8.3$, 1H, C₈H), 7.56 (app-dd, $J = 7.4, 7.5$, 1H, SO₂Ph-*p*-H), 7.49–7.45 (m, 1H, C₇H), 7.47 (app-t, $J = 7.7$, 2H, SO₂Ph-*m*-H), 7.41 (d, $J = 7.7$, 1H, C₅H), 7.20 (app-t, $J = 7.5$, 1H, C₆H), 6.45 (d, $J = 17.5$, 1H, C₂H), 4.58 (s, 1H, C₁₅H), 3.11 (app-t, $J = 14.3$, 1H, C₁₂H_a), 3.09 (s, 3H, C₁₇H₃), 2.94 (dd, $J = 14.3, 20.1$, 1H, C₁₂H_b), 2.30 (s, 3H, C₁₉H₃), 1.93 (s, 3H, C₂₀H₃). ¹³C NMR (150 MHz, CDCl₃, 20 °C): δ 164.3 (C₁₃), 160.2 (C₁₆), 144.1 (d, $J = 5.1$, C₉), 138.3 (SO₂Ph-*ipso*-C), 133.6 (SO₂Ph-*p*-C), 132.4 (d, $J = 3.2$, C₇), 129.3 (SO₂Ph-*m*-C), 128.8 (d, $J = 23.9$, C₄), 127.5 (SO₂Ph-*o*-C), 125.3 (d, $J = 2.7$, C₆), 124.2 (C₅), 117.1 (d, $J = 1.8$, C₈), 102.8 (d, $J = 200.8$, C₃), 82.0 (d, $J = 32.5$, C₂), 70.6 (d, $J = 6.5$, C₁₁), 67.1 (C₁₅), 45.5 (d, $J = 31.8$, C₁₂), 32.8 (C₁₇), 16.9 (C₁₅SCH₃), 15.2 (C₁₁SCH₃). ¹⁹F NMR (282.4 MHz, CDCl₃, 20 °C): δ -135.0. MS (ESI) (m/z): [M+Na]⁺: 530.52, [2M+Na]⁺: 1038.00.

	1H, C ₁₂ H _a), 3.13 (s, 3H, C ₁₇ H ₃), 2.89 (app-d, <i>J</i> = 15.1, 1H, C ₁₂ H _b).
¹³ C NMR (150 MHz, CDCl ₃ , 20 °C):	δ 164.5 (C ₁₃), 160.0 (C ₁₆), 143.2 (d, <i>J</i> = 4.8, C ₉), 137.3 (SO ₂ Ph- <i>ipso</i> -C), 133.8 (SO ₂ Ph- <i>p</i> -C), 132.8 (d, <i>J</i> = 3.4, C ₇), 129.9 (d, <i>J</i> = 23.3, C ₄), 129.1 (SO ₂ Ph- <i>m</i> -C), 127.9 (SO ₂ Ph- <i>o</i> -C), 126.8 (d, <i>J</i> = 2.8, C ₆), 124.8 (C ₅), 119.4 (d, <i>J</i> = 2.2, C ₈), 102.3 (d, <i>J</i> = 205.5, C ₃), 82.7 (d, <i>J</i> = 31.8, C ₂), 74.4 (d, <i>J</i> = 6.2, C ₁₁), 68.4 (C ₁₅), 39.2 (d, <i>J</i> = 32.3, C ₁₂), 32.3 (C ₁₇).
¹⁹ F NMR (282 MHz, CDCl ₃ , 20 °C):	δ -137.7.
FTIR (thin film) cm ⁻¹ :	2999 (w), 2920 (w), 1693 (s), 1447 (w), 1368 (m), 1173 (m), 1088 (w), 1040 (w), 914 (w), 719 (w).
HRMS (ESI) (<i>m/z</i>):	calc'd for C ₂₀ H ₁₇ FN ₃ O ₄ S ₃ [M+H] ⁺ : 478.0360, found 478.0375; calc'd for C ₂₀ H ₁₆ FN ₃ NaO ₄ S ₃ [M+Na] ⁺ : 500.0179, found 500.0198.
TLC (20% ethyl acetate in dichloromethane), R _f :	0.27 (UV, I ₂ , CAM).

α-epimer 35:

¹ H NMR (600 MHz, CDCl ₃ , 20 °C):	δ 7.74 (d, <i>J</i> = 8.5, 2H, SO ₂ Ph- <i>o</i> -H), 7.59 (d, <i>J</i> = 8.2, 1H, C ₈ H), 7.51 (app-dt, <i>J</i> = 1.1, 7.6, 1H, SO ₂ Ph- <i>p</i> -H), 7.43 (dd, <i>J</i> = 7.5, 7.6, 1H, C ₇ H), 7.40 (d, <i>J</i> = 7.5, 1H, C ₅ H), 7.39 (app-t, <i>J</i> = 7.6, 2H, SO ₂ Ph- <i>m</i> -H), 7.20 (dd, <i>J</i> = 7.5, 7.6, 1H, C ₆ H), 6.43 (d, <i>J</i> = 11.5, 1H, C ₂ H), 5.21 (s, 1H, C ₁₅ H), 3.89 (dd, <i>J</i> = 5.2, 15.2, 1H, C ₁₂ H _a), 3.01 (s, 3H, C ₁₇ H ₃), 2.85 (app-ddd, <i>J</i> = 0.5, 15.9, 16.6, 1H, C ₁₂ H _b).
¹³ C NMR (150 MHz, CDCl ₃ , 20 °C):	δ 164.2 (C ₁₃), 161.6 (C ₁₆), 141.9 (d, <i>J</i> = 3.7, C ₉), 137.0 (SO ₂ Ph- <i>ipso</i> -C), 134.0 (SO ₂ Ph- <i>p</i> -C), 132.8 (d, <i>J</i> = 2.8, C ₇), 129.2 (SO ₂ Ph- <i>m</i> -C), 128.6 (d, <i>J</i> = 19.5, C ₄), 128.0 (SO ₂ Ph- <i>o</i> -C), 126.8 (C ₆), 125.4 (C ₅), 118.5 (C ₈), 101.8 (d, <i>J</i> = 170.3, C ₃), 83.1 (d, <i>J</i> = 26.9, C ₂), 74.0 (d, <i>J</i> = 4.0, C ₁₁), 68.6 (C ₁₅), 39.2 (d, <i>J</i> = 26.4, C ₁₂), 31.9 (C ₁₇).
¹⁹ F NMR (282 MHz, CDCl ₃ , 20 °C):	δ -134.1.
FTIR (thin film) cm ⁻¹ :	3069 (w), 2991 (w), 1699 (s), 1448 (w), 1367 (m), 1335 (m), 1173 (m), 1089 (w), 908 (w), 730 (m), 720 (m).
HRMS (ESI) (<i>m/z</i>):	calc'd for C ₂₀ H ₁₇ FN ₃ O ₄ S ₃ [M+H] ⁺ : 478.0360, found 478.0372; calc'd for C ₂₀ H ₁₆ FN ₃ NaO ₄ S ₃ [M+Na] ⁺ : 500.0179, found 500.0199.
TLC (20% ethyl acetate in dichloromethane), R _f :	0.16 (UV, I ₂ , CAM).



C3-(Pyrrol-3'-yl) epidithiodiketopiperazine **32**:

This compound was prepared in two steps starting from bishemiaminal **S18**²⁷ (308 mg, 473 μmol)²² using the methodology developed to access the corresponding C3-(indol-3'-yl) epidithiodiketopiperazine **26**.²³ The orange residue was purified by flash column chromatography on silica gel (eluent: gradient, 10 \rightarrow 40% ethyl acetate in dichloromethane) to afford the epidithiodiketopiperazine **32** (128 mg, 51.5%) as a pale yellow solid. Structural assignments were made with additional information from gCOSY, HSQC, and gHMBC data.²⁸

¹H NMR (600 MHz, CDCl₃, 20 °C):

δ 7.86 (br-s, 1H, N₁H), 7.57 (d, $J = 8.1$, 1H, C₈H), 7.49 (d, $J = 8.4$, 2H, SO₂Ph-*o*-H), 7.36 (app-dt, $J = 1.1$, 7.6, 1H, SO₂Ph-*p*-H), 7.35 (app-t, $J = 8.2$, 1H, C₇H), 7.23 (d, $J = 7.5$, 1H, C₅H), 7.19 (dd, $J = 7.4$, 7.5, 1H, C₆H), 7.15 (app-dt, $J = 0.9$, 7.4, 2H, SO₂Ph-*m*-H), 6.72–6.69 (m, 1H, C₅H), 6.28 (s, 1H, C₂H), 6.03–5.99 (m, 1H, C₄H), 5.58–5.54 (m, 1H, C₂H), 5.22 (s, 1H, C₁₅H), 3.60 (d, $J = 15.5$, 1H, C₁₂H_a), 3.13 (s, 3H, C₁₇H₃), 2.82 (d, $J = 15.5$, 1H, C₁₂H_b).

¹³C NMR (150 MHz, CDCl₃, 20 °C):

δ 165.4 (C₁₃), 160.3 (C₁₆), 140.9 (C₉), 138.6 (SO₂Ph-*ipso*-C), 137.2 (C₄), 132.9 (SO₂Ph-*p*-C), 129.5 (C₇), 128.5 (SO₂Ph-*m*-C), 127.6 (SO₂Ph-*o*-C), 126.0 (C₆), 124.5 (C₅), 123.5 (C₃), 119.6 (C₅), 118.5 (C₈), 117.1 (C₂), 106.4 (C₄), 87.1 (C₂), 74.4 (C₁₁), 68.4 (C₁₅), 55.4 (C₃), 44.2 (C₁₂), 32.2 (C₁₇).

FTIR (thin film) cm⁻¹:

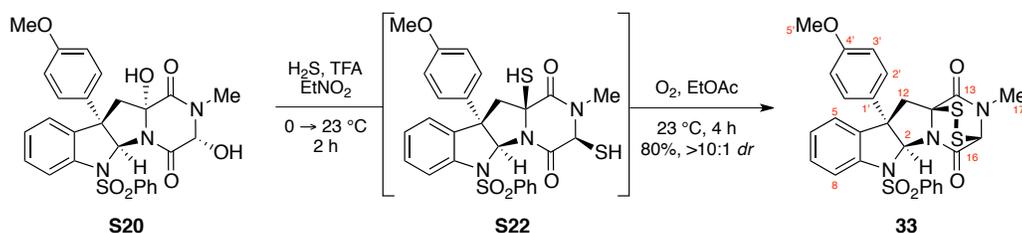
3391 (w), 2925 (w), 1699 (s), 1458 (m), 1360 (m), 1169 (m), 1090 (w), 749 (m).

²⁷ **S18**: ¹H NMR (600 MHz, acetone-*d*₆, 20 °C): δ 7.75 (app-dd, $J = 0.8$, 7.5, 2H, SO₂Ph-*o*-H), 7.54 (app-dt, $J = 0.9$, 7.5, 1H, SO₂Ph-*p*-H), 7.37 (app-dt, $J = 0.7$, 7.6, 2H, SO₂Ph-*m*-H), 7.30 (d, $J = 7.6$, 1H, C₈H), 7.26 (app-dd, $J = 0.4$, 7.7, 1H, C₃H), 7.22 (app-dt, $J = 1.1$, 7.6, 1H, C₆H), 7.12 (app-dt, $J = 1.1$, 7.4, 1H, C₇H), 6.72–6.69 (m, 1H, C₅H), 6.66–6.63 (m, 1H, C₄H), 6.39 (s, 1H, C₂H), 6.25 (br-s, 1H, C₁₁OH), 6.09 (br-s, 1H, C₁₅OH), 5.71–5.68 (m, 1H, C₂H), 5.05 (s, 1H, C₁₅H), 3.35 (d, $J = 14.7$, 1H, C₁₂H_a), 2.92 (s, 3H, C₁₇H₃), 2.85 (d, $J = 14.7$, 1H, C₁₂H_b), 1.46 (sp, $J = 7.5$, 3H, SiCH(CH₃)₂), 1.08 (d, $J = 7.5$, 18H, SiCH(CH₃)₂). MS (ESI) (m/z): [M+Na]⁺: 547.0539. TLC (20% acetone in dichloromethane), R_f: 0.44 (UV, I₂, CAM).

²⁸ The relative stereochemistry of the epidisulfide bridge **32** has been confirmed by key NOESY cross-peaks on the corresponding bis(methylthioether). Our assignment is supported by key NOESY signals (¹H,¹H) in ppm: (1.89,3.06), (2.91,5.86–5.82), (2.91,6.07–6.04), (2.91,6.47). This derivatized compound was prepared in one step using the methodology developed to access (+)-gliocladin B (see reference 9). ¹H NMR (600 MHz, CDCl₃, 20 °C): δ 8.07 (br-s, 1H, N₁H), 7.84 (d, $J = 7.5$, 2H, SO₂Ph-*o*-H), 7.50 (d, $J = 8.2$, 1H, C₈H), 7.47 (t, $J = 7.5$, 1H, SO₂Ph-*p*-H), 7.35 (app-t, $J = 7.9$, 2H, SO₂Ph-*m*-H), 7.28 (app-dt, $J = 1.1$, 7.8, 1H, C₇H), 7.19 (d, $J = 7.4$, 1H, C₅H), 7.09 (dd, $J = 7.4$, 7.5, 1H, C₆H), 6.65–6.62 (m, 1H, C₅H), 6.47 (s, 1H, C₂H), 6.07–6.04 (m, 1H, C₂H), 5.86–5.82 (m, 1H, C₄H), 4.50 (s, 1H, C₁₅H), 3.06 (d, $J = 14.4$, 1H, C₁₂H_a), 3.06 (s, 3H, C₁₈H₃), 2.91 (d, $J = 14.4$, 1H, C₁₂H_b), 2.23 (s, 3H, C₁₅SCH₃), 1.89 (s, 3H, C₁₁SCH₃). ¹³C NMR (150 MHz, CDCl₃, 20 °C): δ 165.3 (C₁₃), 162.6 (C₁₆), 142.1 (C₉), 140.0 (SO₂Ph-*ipso*-C), 137.5 (C₄), 132.8 (SO₂Ph-*p*-C), 128.9 (SO₂Ph-*m*-C), 128.8 (C₇), 127.1 (SO₂Ph-*o*-C), 125.7 (C₃), 124.9 (C₆), 123.5 (C₅), 119.3 (C₅), 117.0 (C₈), 115.6 (C₂), 106.3 (C₄), 86.0 (C₂), 69.7 (C₁₁), 67.7 (C₁₅), 53.1 (C₃), 45.7 (C₁₂), 32.5 (C₁₇), 17.3 (C₁₅SCH₃), 15.5 (C₁₁SCH₃).

HRMS (ESI) (m/z): calc'd for $C_{24}H_{20}N_4NaO_4S_3$ $[M+Na]^+$: 547.0539,
found 547.0560.

TLC (20% ethyl acetate in dichloromethane), R_f : 0.33 (UV, I_2 , CAM).



C3-(*p*-Methoxyphenyl) epidithiodiketopiperazine 33:

This compound was prepared in two steps starting from bishemiaminal **S20**²⁹ (380 mg, 709 μmol)²² using the methodology developed to access the corresponding C3-(indol-3'-yl) epidithiodiketopiperazine **26**.²³ The orange residue was purified by flash column chromatography on silica gel (eluent: gradient, 5 \rightarrow 25% ethyl acetate in dichloromethane) to afford the epidithiodiketopiperazine **33** (321 mg, 80.0%) as a pale yellow solid. Structural assignments were made with additional information from gCOSY, HSQC, and gHMBC data.³⁰

¹H NMR (600 MHz, CDCl₃, 20 °C):

δ 7.64 (d, J = 8.0, 1H, C₈H), 7.40 (app-dt, J = 1.4, 7.0, 1H, C₇H), 7.33 (d, J = 8.0, 2H, SO₂Ph-*o*-H), 7.29 (t, J = 7.5, 1H, SO₂Ph-*p*-H), 7.28–7.23 (m, 2H, C₅H + C₆H), 7.02 (dd, J = 7.6, 7.8, 2H, SO₂Ph-*m*-H), 6.76 (d, J = 8.7, 2H, C₂H), 6.62 (d, J = 8.7, 1H, C₃H), 6.39 (s, 1H, C₂H), 5.28 (s, 1H, C₁₅H), 3.78 (s, 3H, C₅H₃), 3.63 (d, J = 15.6, 1H, C₁₂H_a), 3.13 (s, 3H, C₁₇H₃), 2.87 (d, J = 15.6, 1H, C₁₂H_b).

¹³C NMR (150 MHz, CDCl₃, 20 °C):

δ 165.2 (C₁₃), 160.2 (C₁₆), 158.9 (C₄), 141.3 (C₉), 138.3 (SO₂Ph-*ipso*-C), 135.8 (C₄), 133.1 (SO₂Ph-*p*-C), 131.2 (C₁), 129.9 (C₇), 128.6 (SO₂Ph-*m*-C), 128.0 (C₂), 127.3 (SO₂Ph-*o*-C), 126.2 (C₆), 125.8 (C₅), 119.1 (C₈), 114.5 (C₃), 87.8 (C₂), 74.6 (C₁₁), 68.4 (C₁₅), 59.5 (C₃), 55.5 (C₅), 45.6 (C₁₂), 32.2 (C₁₇).

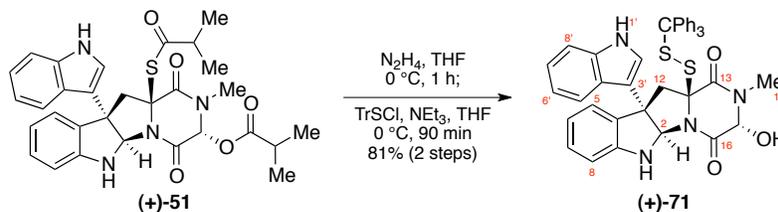
²⁹ **S20**: ¹H NMR (600 MHz, DMSO-*d*₆, 20 °C): δ 7.44 (t, J = 7.5, 1H, SO₂Ph-*p*-H), 7.38 (d, J = 7.8, 1H, C₈H), 7.34 (app-t, J = 8.8, 1H, C₇H), 7.26 (d, J = 7.4, 2H, SO₂Ph-*o*-H), 7.21 (app-dt, J = 0.6, 7.3, 1H, C₆H), 7.14 (dd, J = 7.6, 8.1, 2H, SO₂Ph-*m*-H), 7.01 (d, J = 7.5, 1H, C₅H), 6.76 (d, J = 8.8, 2H, C₂H), 6.66 (d, J = 8.8, 1H, C₃H), 6.22 (s, 1H, C₂H), 5.00 (d, J = 7.4, 1H, C₁₅H), 3.74 (s, 3H, C₅H₃), 3.19 (d, J = 15.0, 1H, C₁₂H_a), 2.77 (s, 3H, C₁₇H₃), 2.67 (d, J = 15.0, 1H, C₁₂H_b). ¹³C NMR (100 MHz, DMSO-*d*₆, 20 °C): δ 166.6 (C₁₃), 165.8 (C₁₆), 158.0 (C₄), 139.4 (C₉), 138.0 (SO₂Ph-*ipso*-C), 137.8 (C₄), 133.4 (C₁), 133.2 (SO₂Ph-*p*-C), 128.9 (C₇), 128.7 (SO₂Ph-*m*-C), 128.0 (C₂), 126.7 (SO₂Ph-*o*-C), 126.7 (C₅), 125.7 (C₆), 117.0 (C₈), 114.0 (C₃), 87.3 (C₂), 86.0 (C₁₁), 80.9 (C₁₅), 57.4 (C₃), 55.1 (C₅), 49.7 (C₁₂), 30.5 (C₁₇). MS (ESI) (m/z): [M+H]⁺: 537.39, [M+Na]⁺: 558.43, [2M+Na]⁺: 1094.13. TLC (20% acetone in dichloromethane), R_f: 0.50 (UV, CAM).

³⁰ The relative stereochemistry of the epidisulfide bridge **33** has been confirmed by key NOESY cross-peaks on the corresponding bis(methylthioether). Our assignment is supported by key NOESY signals (¹H,¹H) in ppm: (1.89,3.13), (3.13,7.13–7.07), (2.98,6.89), (2.98,6.47). This derivatized compound was prepared in one step using the methodology developed to access (+)-gliocladin B (see reference 9). ¹H NMR (600 MHz, CDCl₃, 20 °C): δ 7.85 (app-dd, J = 0.7, 7.7, 2H, SO₂Ph-*o*-H), 7.54 (d, J = 8.1, 1H, C₈H), 7.48 (t, J = 7.3, 1H, SO₂Ph-*p*-H), 7.35 (app-t, J = 7.8, 2H, SO₂Ph-*m*-H), 7.30 (app-dt, J = 1.4, 7.5, 1H, C₇H), 7.13–7.07 (m, 2H, C₅H + C₆H), 6.89 (d, J = 8.8, 2H, C₂H), 6.70 (d, J = 8.8, 2H, C₃H), 6.64 (s, 1H, C₂H), 4.48 (s, 1H, C₁₅H), 3.75 (s, 3H, C₅H₃), 3.13 (d, J = 14.3, 1H, C₁₂H_a), 3.06 (s, 3H, C₁₇H₃), 2.98 (d, J = 14.3, 1H, C₁₂H_b), 2.20 (s, 3H, C₁₅SCH₃), 1.89 (s, 3H, C₁₁SCH₃). ¹³C NMR (150 MHz, CDCl₃, 20 °C): δ 165.1 (C₁₃), 162.3 (C₁₆), 158.8 (C₄), 142.3 (C₉), 140.1 (SO₂Ph-*ipso*-C), 136.7 (C₄), 134.2 (C₁), 132.9 (SO₂Ph-*p*-C), 129.1 (SO₂Ph-*m*-C), 127.1 (C₂), 127.1 (C₇), 127.0 (SO₂Ph-*o*-C), 124.9 (C₆), 123.8 (C₅), 117.1 (C₈), 114.4 (C₃), 85.8 (C₂), 69.8 (C₁₁), 67.6 (C₁₅), 57.0 (C₃), 55.5 (C₅), 45.7 (C₁₂), 32.5 (C₁₇), 17.2 (C₁₅SCH₃), 15.5 (C₁₁SCH₃).

FTIR (thin film) cm^{-1} : 3065 (w), 3006 (w), 2931 (w), 2839 (w), 1698 (s),
1512 (m), 1459 (m), 1363 (m), 1255 (m), 1170 (m),
1035 (w), 755 (m).

HRMS (ESI) (m/z): calc'd for $\text{C}_{27}\text{H}_{23}\text{N}_3\text{NaO}_5\text{S}_3$ $[\text{M}+\text{Na}]^+$: 588.0692,
found 588.0694.

TLC (20% ethyl acetate in dichloromethane), R_f : 0.42 (UV, I_2 , CAM).



Hexacyclic triphenylmethanedisulfide (+)-71:⁹

Anhydrous hydrazine (150 μ L, 4.77 mmol, 11.1 equiv) was added via syringe to a solution of aminothioisobutyrate (+)-**51**⁹ (240 mg, 428 μ mol, 1 equiv) in anhydrous tetrahydrofuran (50 mL) at 0 $^{\circ}$ C. After 1 h, the reaction mixture was diluted sequentially with saturated aqueous ammonium chloride solution (20 mL) and ethyl acetate (120 mL). The organic layer was sequentially washed with saturated aqueous ammonium chloride solution (50 mL), water (2 \times 50 mL), and saturated aqueous sodium chloride solution (30 mL). The aqueous layer was extracted with ethyl acetate (2 \times 50 mL). The combined organic layers were dried over anhydrous sodium sulfate, were filtered, and were concentrated under reduced pressure to afford the hexacyclic aminothiols that were used in the next step without further purification.³¹

Triethylamine (600 μ L, 4.27 mmol, 10.0 equiv) and solid triphenylmethanesulfonyl chloride (665 mg, 2.14 mmol, 5.00 equiv) were sequentially added to a solution of hexacyclic aminothiols in anhydrous tetrahydrofuran (60 mL) at 0 $^{\circ}$ C under an argon atmosphere. After 90 min, the solution was partitioned between saturated aqueous ammonium chloride (50 mL) and ethyl acetate (130 mL). The aqueous layer was extracted with diethyl ether (2 \times 50 mL), and the combined organic layers were washed sequentially with water (2 \times 50 mL) and saturated aqueous sodium chloride solution (30 mL), were dried over anhydrous sodium sulfate, were filtered, and were concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (eluent: gradient, 10 \rightarrow 30% ethyl acetate in dichloromethane) to afford triphenylmethanedisulfide (+)-**71** (242 mg, 81.4%)³² as a white solid. Structural assignments were made with additional information from gCOSY, HSQC, and gHMBC data.³³

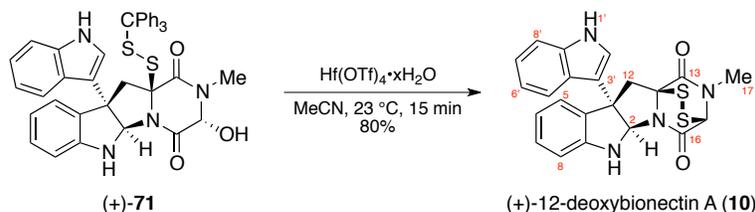
¹H NMR (600 MHz, acetonitrile-*d*₃, 20 $^{\circ}$ C): δ 9.16 (br-s, 1H, N₁H), 7.37 (d, *J* = 7.4, 1H, C₈H), 7.36 (d, *J* = 7.6, 1H, C₅H), 7.34–7.30 (m, 6H, C(Ph-*o*-H)₃), 7.34–7.30 (m, 3H, C(Ph-*p*-H)₃), 7.18–7.15 (m, 6H, C(Ph-*m*-H)₃), 7.15–7.11 (m, 1H, C₇H), 7.10

³¹ This hexacyclic aminothiols can be purified by flash column chromatography on silica gel (eluent: gradient, 1 \rightarrow 3% methanol in dichloromethane). ¹H NMR (600 MHz, CDCl₃, 20 $^{\circ}$ C): δ 8.16 (br-s, 1H, N₁H), 7.40 (d, *J* = 8.1, 1H, C₅H), 7.31 (d, *J* = 8.2, 1H, C₈H), 7.23 (d, *J* = 7.4, 1H, C₅H), 7.19 (app-dt, *J* = 1.0, 7.7, 1H, C₇H), 7.17 (app-t, *J* = 7.4, 1H, C₇H), 7.02 (app-t, *J* = 7.6, 1H, C₆H), 6.89 (d, *J* = 2.6, 1H, C₂H), 6.85 (app-t, *J* = 7.0, 1H, C₆H), 6.77 (d, *J* = 7.8, 1H, C₈H), 5.92 (s, 1H, C₂H), 5.36 (s, 1H, C₁₅H), 5.20 (br-s, 1H, N₁H), 3.76 (d, *J* = 14.3, 1H, C₁₂H_a), 3.75 (br-s, 1H, C₁₅OH), 3.30 (d, *J* = 14.3, 1H, C₁₂H_b), 3.09 (s, 3H, C₁₄H₃), 2.57 (br-s, 1H, C₁₁SH). ¹³C NMR (150 MHz, CDCl₃, 20 $^{\circ}$ C): δ 166.6 (C₁₃), 166.6 (C₁₆), 148.2 (C₉), 137.4 (C₉), 131.8 (C₄), 129.4 (C₇), 125.0 (C₄), 125.0 (C₅), 122.7 (C₇), 122.2 (C₂), 120.4 (C₆), 120.2 (C₆), 119.7 (C₅), 117.3 (C₃), 111.8 (C₈), 110.4 (C₈), 82.5 (C₂), 77.2 (C₁₅), 69.0 (C₁₁), 54.2 (C₃), 50.9 (C₁₂), 29.3 (C₁₈). TLC (5% methanol in dichloromethane), R_f: 0.27 (UV, CAM).

³² This sequence can also be combined as a sequential single-flask two-step process to afford (+)-**71** in 74% yield.

³³ Triphenylmethanedisulfide (+)-**71** has also been characterized by NMR in CDCl₃: ¹H NMR (600 MHz, CDCl₃, 20 $^{\circ}$ C): δ 8.00 (br-s, 1H, N₁H), 7.31 (d, *J* = 7.8, 1H, C₅H), 7.30 (d, *J* = 7.8, 1H, C₈H), 7.29–7.26 (m, 6H, C(Ph-*o*-H)₃), 7.29–7.26 (m, 3H, C(Ph-*p*-H)₃), 7.20–7.17 (m, 6H, C(Ph-*m*-H)₃), 7.16 (app-t, *J* = 7.7, 1H, C₇H), 7.15 (app-t, *J* = 8.1, 1H, C₇H), 7.02 (app-t, *J* = 7.5, 1H, C₆H), 6.83 (d, *J* = 2.5, 1H, C₂H), 6.74–6.68 (m, 1H, C₃H), 6.74–6.68 (m, 1H, C₆H), 6.74–6.68 (m, 1H, C₈H), 5.82 (s, 1H, C₂H), 5.24 (d, *J* = 3.6, 1H, C₁₅H), 4.99 (br-s, 1H, N₁H), 4.07 (d, *J* = 3.6, 1H, C₁₅OH), 3.43 (d, *J* = 14.7, 1H, C₁₂H_a), 3.00 (s, 3H, C₁₇H₃), 2.57 (d, *J* = 14.7, 1H, C₁₂H_b). ¹³C NMR (150 MHz, CDCl₃, 20 $^{\circ}$ C): δ 167.3 (C₁₆), 163.7 (C₁₃), 147.6 (C₉), 143.9 (C(Ph-*ipso*-C)₃), 137.3 (C₉), 131.6 (C₄), 130.8 (C(Ph-*m*-C)₃), 129.5 (C₇), 127.9 (C(Ph-*o*-C)₃), 127.5 (C(Ph-*p*-C)₃), 125.2 (C₅), 125.1 (C₄), 122.6 (C₇), 122.0 (C₂), 120.2 (C₆), 120.0 (C₅), 119.9 (C₆), 117.5 (C₃), 111.6 (C₈), 110.1 (C₃), 82.6 (C₂), 77.6 (CPh₃), 72.9 (C₁₅), 69.7 (C₁₁), 54.0 (C₃), 48.0 (C₁₂), 29.4 (C₁₈).

	(app-dt, $J = 0.8$, 7.6, 1H, C ₇ H), 6.97 (d, $J = 2.7$, 1H, C ₂ H), 6.96 (app-t, $J = 8.0$, 1H, C ₆ H), 6.68 (d, $J = 7.9$, 1H, C ₈ H), 6.64–6.60 (m, 1H, C ₅ H), 6.64–6.60 (m, 1H, C ₆ H), 5.75 (d, $J = 1.0$, 1H, C ₂ H), 5.60 (br-s, 1H, N ₁ H), 5.11 (s, 1H, C ₁₅ H), 4.59 (br-s, 1H, C ₁₅ OH), 3.32 (d, $J = 14.5$, 1H, C ₁₂ H _a), 2.89 (s, 3H, C ₁₇ H ₃), 2.70 (d, $J = 14.5$, 1H, C ₁₂ H _b).
¹³ C NMR (150 MHz, acetonitrile- <i>d</i> ₃ , 20 °C):	δ 166.9 (C ₁₃), 164.1 (C ₁₆), 149.2 (C ₉), 145.0 (C(Ph- <i>ipso</i> -C) ₃), 138.1 (C ₉), 133.3 (C ₄), 131.2 (C(Ph- <i>m</i> -C) ₃), 129.4 (C ₇), 128.8 (C(Ph- <i>o</i> -C) ₃), 128.4 (C(Ph- <i>p</i> -C) ₃), 125.8 (C ₄), 125.4 (C ₅), 122.8 (C ₇), 122.7 (C ₂), 120.3 (C ₆), 120.3 (C ₅), 120.1 (C ₆), 118.6 (C ₃), 112.7 (C ₈), 110.6 (C ₈), 83.1 (C ₂), 78.4 (CPh ₃), 78.4 (C ₁₅), 73.1 (C ₁₁), 54.3 (C ₃), 49.4 (C ₁₂), 29.1 (C ₁₈).
FTIR (thin film) cm ⁻¹ :	3345 (br-m), 3056 (w), 2926 (w), 1674 (s), 1483 (m), 1459 (m), 1442 (m), 1388 (m), 745 (s), 700 (s).
HRMS (ESI) (<i>m/z</i>):	calc'd for C ₄₁ H ₃₅ N ₄ O ₃ S ₂ [M+H] ⁺ : 695.2145, found: 695.2147.
[α] _D ²⁴ :	+165.2 (<i>c</i> = 0.12, CHCl ₃).
TLC (5% methanol in dichloromethane), R _f :	0.44 (UV, CAM).



(+)-12-Deoxybionectin A (10):^{9,34}

Hafnium(IV) trifluoromethanesulfonate hydrate (800 mg) was added as a solid to a colorless solution of hexacyclic triphenylmethanedisulfide (+)-71 (100 mg, 144 μmol , 1 equiv) in anhydrous acetonitrile (40 mL) at 23 °C. A bright yellow coloration was observed immediately after the addition. The suspension was stirred at 23 °C under an argon atmosphere. After 15 min, the reaction mixture was partitioned between saturated aqueous sodium bicarbonate (60 mL) and ethyl acetate (100 mL). The aqueous layer was extracted with ethyl acetate (2 \times 50 mL). The combined organic layers were washed sequentially with water (3 \times 50 mL) and saturated aqueous sodium chloride solution (30 mL), were dried over anhydrous sodium sulfate, were filtered, and were concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (eluent: gradient, 1 \rightarrow 6% acetone in dichloromethane) to afford (+)-12-deoxybionectin A (10) (50.2 mg, 80.3 %) as a colorless oil. Structural assignments were made with additional information from gCOSY, HSQC, and gHMBC data.³⁵

¹H NMR (600 MHz, CDCl₃, 20 °C):

δ 8.07 (br-s, 1H, N₁H), 7.48 (d, J = 8.0, 1H, C₅H), 7.37 (d, J = 8.2, 1H, C₈H), 7.25 (d, J = 8.3, 1H, C₅H), 7.20 (app-dt, J = 0.7, 7.7, 1H, C₇H), 7.20 (app-dt, J = 0.7, 7.7, 1H, C₇H), 7.09 (app-t, J = 7.6, 1H, C₆H), 6.95 (d, J = 2.5, 1H, C₂H), 6.88 (app-t, J = 7.4, 1H, C₆H), 6.76 (d, J = 7.9, 1H, C₈H), 5.95 (s, 1H, C₂H), 5.30 (br-s, 1H, N₁H), 5.21 (s, 1H, C₁₅H), 4.10 (d, J = 15.4, 1H, C₁₂H_a), 3.15 (s, 3H, C₁₇H₃), 2.95 (d, J = 15.4, 1H, C₁₂H_b).

¹³C NMR (150 MHz, CDCl₃, 20 °C):

δ 165.8 (C₁₃), 162.2 (C₁₆), 148.2 (C₉), 137.5 (C₉), 132.0 (C₄), 129.4 (C₇), 125.1 (C₄), 124.3 (C₅), 122.9 (C₇), 122.9 (C₂), 120.4 (C₆), 120.1 (C₆), 119.6 (C₅), 116.7 (C₃), 111.9 (C₈), 110.4 (C₈), 83.0 (C₂), 74.8 (C₁₁), 68.4 (C₁₅), 56.1 (C₃), 43.6 (C₁₂), 32.2 (C₁₇).

FTIR (thin film) cm⁻¹:

3358 (br-w), 3006 (w), 2926 (w), 1684 (s), 1609 (w), 1460 (w), 1383 (w), 1232 (m), 748 (m).

HRMS (ESI) (m/z):

calc'd for C₂₂H₁₉N₄O₂S₂ [M+H]⁺: 435.0944, found: 435.0943.

[α]_D²⁴:

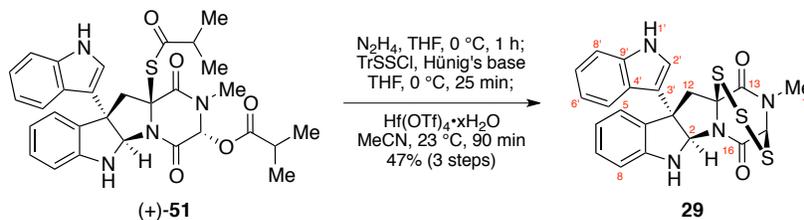
+387.3 (c = 0.10, CHCl₃).

TLC (10% acetone in dichloromethane), R_f:

0.54 (UV, CAM).

³⁴ Zheng, C.-J.; Kim, C.-J.; Bae, K. S.; Kim, Y.-H.; Kim, W.-G. *J. Nat. Prod.* **2006**, *69*, 1816.

³⁵ The relative stereochemistry of the episulfide bridge **10** has been confirmed by key NOESY signals (¹H,¹H) in ppm: (1.99,3.31), (3.31,7.16), (3.20,6.06) on the corresponding bis(methylthioether) – *i.e.*, (+)-gliocladin B (**7**, see reference 9).



C3-(Indol-3'-yl) epitritiodiketopiperazine 29:

This compound was prepared in two steps starting from aminothioisobutyrate (+)-**51**⁹ (26.5 mg, 47.3 μmol) using the methodology developed to access the corresponding C3-(indol-3'-yl) epidithiodiketopiperazine (+)-12-deoxybionectin A (**10**) (Please see pages S27 and S29 for details). The residue was purified by flash column chromatography on silica gel (eluent: gradient, 2 \rightarrow 10% acetone in dichloromethane) to afford epitritiodiketopiperazine **29** (10.3 mg, 46.7%) as a colorless oil. Structural assignments were made with additional information from gCOSY, HSQC, and gHMBC data.³⁶

¹H NMR (600 MHz, CDCl₃, 20 °C):

Major conformer: δ 8.10 (br-s, 1H, N₁H), 7.46 (d, J = 8.1, 1H, C₅H), 7.35 (d, J = 8.0, 1H, C₈H), 7.21 (app-dt, J = 0.7, 6.9, 1H, C₇H), 7.18 (app-t, J = 7.6, 1H, C₇H), 7.14 (d, J = 7.3, 1H, C₅H), 7.06 (app-t, J = 7.5, 1H, C₆H), 6.92 (d, J = 2.4, 1H, C₂H), 6.81 (d, J = 8.3, 1H, C₈H), 6.80 (app-t, J = 7.5, 1H, C₆H), 5.85 (s, 1H, C₂H), 4.87 (s, 1H, C₁₅H), 3.80 (d, J = 14.6, 1H, C₁₂H_a), 3.20 (s, 3H, C₁₇H₃), 3.16 (d, J = 14.6, 1H, C₁₂H_b).³⁷

Minor conformer: δ 8.11 (br-s, 1H, N₁H), 7.54 (d, J = 8.1, 1H, C₅H), 7.36 (d, J = 7.9, 1H, C₈H), 7.22–7.18 (m, 1H, C₇H), 7.12 (d, J = 7.4, 1H, C₅H), 7.11 (dd, J = 7.6, 7.7, 1H, C₇H), 7.09 (app-t, J = 7.6, 1H, C₆H), 6.94 (d, J = 2.4, 1H, C₂H), 6.78 (app-t, J = 7.5, 1H, C₆H), 6.71 (d, J = 7.7, 1H, C₈H), 6.19 (s, 1H, C₂H), 5.21 (s, 1H, C₁₅H), 3.70 (d, J = 14.7, 1H, C₁₂H_a), 3.09 (d, J = 14.7, 1H, C₁₂H_b), 3.02 (s, 3H, C₁₇H₃).³⁷

¹³C NMR (150 MHz, CDCl₃, 20 °C):

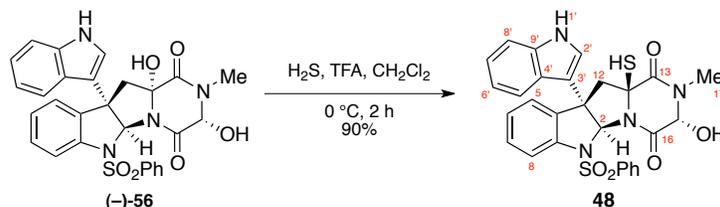
Major conformer: δ 168.9 (C₁₃), 164.5 (C₁₆), 149.6 (C₉), 137.3 (C₉), 130.8 (C₄), 129.9 (C₇), 125.0 (C₄), 124.8 (C₅), 122.8 (C₇), 122.5 (C₂), 120.3 (C₆), 120.0 (C₆), 119.7 (C₅), 116.5 (C₃), 111.8 (C₈), 110.6 (C₈), 82.1 (C₂), 79.3 (C₁₁), 67.2 (C₁₅), 54.2 (C₃), 49.2 (C₁₂), 31.2 (C₁₇).

Minor conformer: δ 167.4 (C₁₃), 163.2 (C₁₆), 148.2 (C₉), 137.4 (C₉), 131.4 (C₄), 129.2 (C₇), 125.1 (C₄), 124.3 (C₅), 122.8 (C₇), 122.5 (C₂), 120.3 (C₆),

³⁶ Upon concentration or in concentrated solution, the epitritiodiketopiperazine **29** tends to degrade, thus rendering its isolation and characterization particularly arduous.

³⁷ The resonance for N₁H was not observed.

	120.2 (C ₆), 119.7 (C ₅), 116.7 (C ₃), 111.9 (C ₈), 109.8 (C ₈), 83.7 (C ₂), 74.8 (C ₁₁), 71.2 (C ₁₅), 54.3 (C ₃), 46.8 (C ₁₂), 32.5 (C ₁₇).
FTIR (thin film) cm ⁻¹ :	3397 (br-m), 3061 (w), 2922 (w), 2852 (w), 1693 (s), 1458 (m), 1382 (m), 1265 (w), 1170 (m), 1092 (w), 1026 (w), 737 (m).
HRMS (ESI) (<i>m/z</i>):	calc'd for C ₂₂ H ₁₉ N ₄ O ₂ S ₃ [M+H] ⁺ : 467.0665, found: 467.0669.
TLC (10% acetone in dichloromethane), R _f :	0.61 (UV, I ₂ , CAM).



C3-(Indol-3'-yl) C11-thiohemiaminal 48:

A slow stream of hydrogen sulfide gas was introduced into a solution of diol (-)-**56** (185 mg, 340 μmol , 1 equiv) in anhydrous dichloromethane (30 mL) at 0 °C, providing a saturated hydrogen sulfide solution. After 20 min, trifluoroacetic acid (6 mL) was added via syringe over 10 min, and the slow introduction of hydrogen sulfide into the mixture was maintained for another 10 min. The reaction mixture was left under an atmosphere of hydrogen sulfide for an additional 2 h at 0 °C. A slow stream of argon gas was introduced into the solution. After 15 min, the reaction mixture was diluted with ethyl acetate (150 mL) and slowly poured into saturated aqueous sodium hydrogenocarbonate solution (50 mL). The organic layer was sequentially washed with water (3 \times 40 mL) and saturated aqueous sodium chloride solution (40 mL). The organic layer was dried over anhydrous sodium sulfate, was filtered, and was concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (eluent: gradient, 5 \rightarrow 25% acetone in dichloromethane) to afford thiohemiaminal **48** (171 mg, 89.8 %) as an orange solid. Structural assignments were made with additional information from gCOSY, HSQC, and gHMBC data.

^1H NMR (600 MHz, CDCl_3 , 20 °C):

δ 7.89 (br-s, 1H, N_1H), 7.87 (d, $J = 8.2$, 1H, C_8H), 7.45 (d, $J = 7.7$, 2H, $\text{SO}_2\text{Ph-}o\text{-H}$), 7.44–7.39 (m, 1H, C_7H), 7.34 (t, $J = 7.4$, 1H, $\text{SO}_2\text{Ph-}p\text{-H}$), 7.31 (d, $J = 8.2$, 1H, C_8H), 7.21–7.18 (m, 2H, $\text{C}_5\text{H} + \text{C}_6\text{H}$), 7.17 (dd, $J = 7.4$, 7.8, 1H, C_7H), 7.03 (app-t, $J = 7.8$, 2H, $\text{SO}_2\text{Ph-}m\text{-H}$), 6.92 (dd, $J = 7.5$, 7.6, 1H, C_6H), 6.73 (d, $J = 8.0$, 1H, C_5H), 6.61 (s, 1H, C_2H), 6.22 (d, $J = 2.3$, 1H, C_2H), 5.42 (s, 1H, C_{15}H), 4.53 (br-s, 1H, C_{15}OH), 3.82 (d, $J = 14.9$, 1H, C_{12}H_a), 3.11 (s, 3H, C_{17}H_3), 2.99 (d, $J = 14.9$, 1H, C_{12}H_b), 2.61 (s, 1H, C_{11}SH).

^{13}C NMR (150 MHz, CDCl_3 , 20 °C):

δ 166.2 (C_{13}), 165.8 (C_{16}), 140.9 (C_9), 137.3 ($\text{SO}_2\text{Ph-}ipso\text{-C}$), 136.8 (C_9), 135.9 (C_4), 133.3 ($\text{SO}_2\text{Ph-}p\text{-C}$), 129.4 (C_7), 128.6 ($\text{SO}_2\text{Ph-}m\text{-C}$), 127.5 ($\text{SO}_2\text{Ph-}o\text{-C}$), 126.0 (C_6), 125.0 (C_5), 124.1 (C_4), 123.9 (C_2), 122.9 (C_7), 120.5 (C_6), 118.7 (C_5), 118.4 (C_8), 114.2 (C_3), 111.9 (C_8), 84.5 (C_2), 77.3 (C_{15}), 69.5 (C_{11}), 53.8 (C_3), 51.8 (C_{12}), 29.3 (C_{17}).

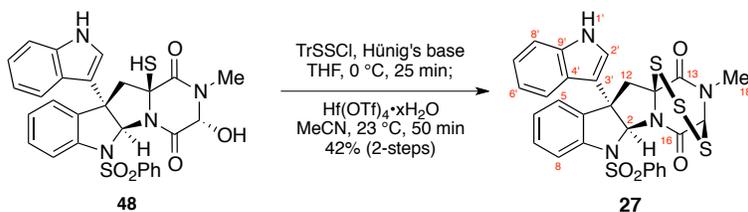
FTIR (thin film) cm^{-1} :

3394 (br-w), 2926 (w), 2547 (w), 1700 (s), 1662 (s), 1457 (m), 1359 (m), 1168 (s), 1090 (m), 1024 (w), 734 (m).

HRMS (ESI) (m/z):

calc'd for $\text{C}_{28}\text{H}_{24}\text{N}_4\text{NaO}_5\text{S}_2$ [$\text{M}+\text{Na}$] $^+$: 583.1080, found: 583.1095.

TLC (20% ethyl acetate in dichloromethane), R_f : 0.09 (UV, CAM).



C3-(Indol-3'-yl) epitritiodiketopiperazine **27**:

This compound was prepared in two steps starting from thiohemiaminal **48** (25.0 mg, 44.6 μmol) using the methodology developed to access the corresponding C3-(indol-3'-yl) epidithiodiketopiperazine (+)-12-deoxybionectin A (**10**) (Please see pages S27 and S29 for details). The residue was purified by flash column chromatography on silica gel (eluent: gradient, 5 \rightarrow 30% ethyl acetate in dichloromethane) to afford epitritiodiketopiperazine **27** (11.3 mg, 41.8 %) as a white solid. Structural assignments were made with additional information from gCOSY, HSQC, and gHMBC data. Based on ^1H NMR analysis at 20 $^\circ\text{C}$ in CDCl_3 , the product exists as a 3:7 mixture of minor:major conformers.³⁸

^1H NMR (600 MHz, CDCl_3 , 20 $^\circ\text{C}$):

Major conformer: δ 7.89 (br-s, 1H, N_1H), 7.81 (d, J = 8.1, 1H, C_8H), 7.53 (d, J = 7.5, 2H, $\text{SO}_2\text{Ph-}o\text{-H}$), 7.41 (app-ddd, J = 2.3, 6.5, 8.1, 1H, C_7H), 7.37 (t, J = 7.7, 1H, $\text{SO}_2\text{Ph-}p\text{-H}$), 7.33 (d, J = 8.1, 1H, C_8H), 7.19 (dd, J = 6.9, 7.9, 1H, C_7H), 7.16–7.12 (m, 2H, $\text{C}_5\text{H} + \text{C}_6\text{H}$), 7.09 (dd, J = 7.8, 8.0, 2H, $\text{SO}_2\text{Ph-}m\text{-H}$), 6.96 (dd, J = 7.4, 7.7, 1H, C_6H), 6.89 (d, J = 8.0, 1H, C_5H), 6.56 (s, 1H, C_2H), 6.25 (d, J = 2.5, 1H, C_2H), 4.91 (s, 1H, C_{15}H), 3.83 (d, J = 15.2, 1H, C_{12}H_a), 3.21 (s, 3H, C_{17}H_3), 2.84 (d, J = 15.2, 1H, C_{12}H_b).

Minor conformer: δ 7.77 (br-s, 1H, N_1H), 7.68 (d, J = 8.0, 1H, C_8H), 7.38–7.34 (m, 2H, $\text{C}_5\text{H} + \text{C}_8\text{H}$), 7.34–7.32 (m, 1H, C_7H), 7.27–7.22 (m, 2H, $\text{C}_5\text{H} + \text{SO}_2\text{Ph-}p\text{-H}$), 7.22–7.19 (m, 2H, $\text{SO}_2\text{Ph-}o\text{-H}$), 7.19–7.16 (m, 1H, C_6H), 7.15–7.12 (m, 1H, C_6H), 6.98–6.94 (m, 1H, C_7H), 6.95 (s, 1H, C_2H), 6.92–6.86 (m, 2H, $\text{SO}_2\text{Ph-}m\text{-H}$), 5.88 (d, J = 2.6, 1H, C_2H), 5.26 (s, 1H, C_{15}H), 3.62 (d, J = 15.1, 1H, C_{12}H_a), 3.03 (s, 3H, C_{17}H_3), 2.85 (d, J = 15.1, 1H, C_{12}H_b).

^{13}C NMR (150 MHz, CDCl_3 , 20 $^\circ\text{C}$):

Major conformer: δ 168.2 (C_{13}), 162.6 (C_{16}), 142.0 (C_9), 137.7 ($\text{SO}_2\text{Ph-}ipso\text{-C}$), 137.3 (C_9), 135.3 (C_4), 133.1 ($\text{SO}_2\text{Ph-}p\text{-C}$), 130.2 (C_7), 128.6 ($\text{SO}_2\text{Ph-}m\text{-C}$), 127.5 ($\text{SO}_2\text{Ph-}o\text{-C}$), 125.8 (C_6), 124.9 (C_5), 124.2 (C_4), 124.0 (C_2), 123.0 (C_7), 120.5 (C_6), 118.9 (C_5), 118.8 (C_8), 113.8 (C_3), 112.0 (C_8), 84.4 (C_2), 79.5 (C_{11}), 67.2 (C_{15}), 53.7 (C_3), 48.8 (C_{12}), 32.8 (C_{17}).

³⁸ Upon concentration or in concentrated solution, the epitritiodiketopiperazine **27** tends to degrade, thus rendering its isolation and characterization particularly arduous. One of the by-products has been identified as the corresponding epidithiodiketopiperazine **26**.

Minor conformer: δ 169.9 (C₁₃), 161.5 (C₁₆), 141.2 (C₉), 138.1 (SO₂Ph-*ipso*-C), 137.1 (C₉), 136.6 (C₄), 132.9 (SO₂Ph-*p*-C), 129.7 (C₇), 128.2 (SO₂Ph-*m*-C), 127.4 (SO₂Ph-*o*-C), 126.5 (C₆), 124.7 (C₅), 124.2 (C₂), 123.9 (C₄), 123.2 (C₇), 120.8 (C₆), 119.4 (C₈), 118.7 (C₅), 114.2 (C₃), 112.0 (C₈), 85.4 (C₂), 75.0 (C₁₁), 71.4 (C₁₅), 54.1 (C₃), 46.3 (C₁₂), 33.2 (C₁₇).

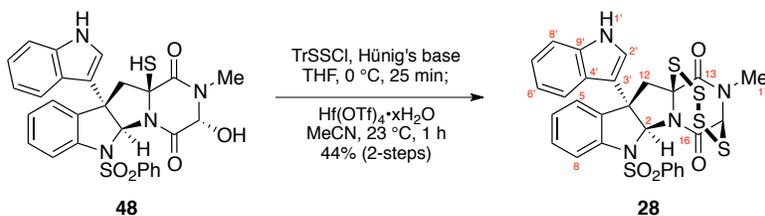
FTIR (thin film) cm⁻¹:

3394 (br-m), 3017 (w), 2922 (w), 2852 (w), 1699 (s), 1460 (m), 1364 (m), 1236 (w), 1169 (m), 1082 (m), 1049 (w), 750 (m).

HRMS (ESI) (*m/z*):

calc'd for C₂₈H₂₃N₄O₄S₄ [M+H]⁺: 607.0597, found 607.0611; calc'd for C₂₈H₂₂N₄NaO₄S₄ [M+Na]⁺: 629.0416, found 629.0435.

TLC (10% ethyl acetate in dichloromethane), R_f: 0.46 (UV, CAM).



C3-(Indol-3'-yl) epitetrathiodiketopiperazine **28**:

This compound was prepared in two steps starting from thiohemiaminal **48** (49.3 mg, 88.0 μmol) using the methodology developed to access the corresponding C3-(indol-3'-yl) epidithiodiketopiperazine (+)-12-deoxybionectin A (**10**) (Please see pages S27 and S29 for details). The residue was purified by flash column chromatography on silica gel (eluent: gradient, 5 \rightarrow 30% acetate in dichloromethane) to afford epitetrathiodiketopiperazine **28** (25.0 mg, 44.4 %) as a white solid. Structural assignments were made with additional information from gCOSY, HSQC, and gHMBC data.³⁹

¹H NMR (600 MHz, CDCl₃, 20 °C):

δ 7.92 (br-s, 1H, N₁H), 7.69 (d, $J = 7.8$, 2H, SO₂Ph-*o*-H), 7.58 (d, $J = 8.1$, 1H, C₈H), 7.40 (t, $J = 7.4$, 1H, SO₂Ph-*p*-H), 7.34 (d, $J = 8.2$, 1H, C₈H), 7.31 (app-t, $J = 7.8$, 1H, C₇H), 7.22–7.16 (m, 4H, C₅H + C₇H + SO₂Ph-*m*-H), 7.11 (app-t, $J = 7.4$, 1H, C₆H), 7.04 (d, $J = 7.8$, 1H, C₅H), 7.01 (dd, $J = 7.1$, 7.7, 1H, C₆H), 6.95 (s, 1H, C₂H), 6.45 (d, $J = 2.2$, 1H, C₂H), 5.23 (s, 1H, C₁₅H), 3.47 (d, $J = 14.8$, 1H, C₁₂H_a), 3.06 (s, 3H, C₁₇H₃), 3.03 (d, $J = 14.8$, 1H, C₁₂H_b).

¹³C NMR (150 MHz, CDCl₃, 20 °C):

δ 168.2 (C₁₃), 162.8 (C₁₆), 141.8 (C₉), 138.5 (SO₂Ph-*ipso*-C), 137.3 (C₉), 136.4 (C₄), 133.2 (SO₂Ph-*p*-C), 129.7 (C₇), 128.8 (SO₂Ph-*m*-C), 127.7 (SO₂Ph-*o*-C), 125.7 (C₆), 124.6 (C₅), 124.3 (C₄), 123.0 (C₂), 123.0 (C₇), 120.7 (C₆), 118.8 (C₅), 117.3 (C₈), 115.8 (C₃), 112.0 (C₈), 85.2 (C₂), 76.0 (C₁₁), 68.3 (C₁₅), 53.6 (C₃), 49.1 (C₁₂), 32.5 (C₁₇).

FTIR (thin film) cm⁻¹:

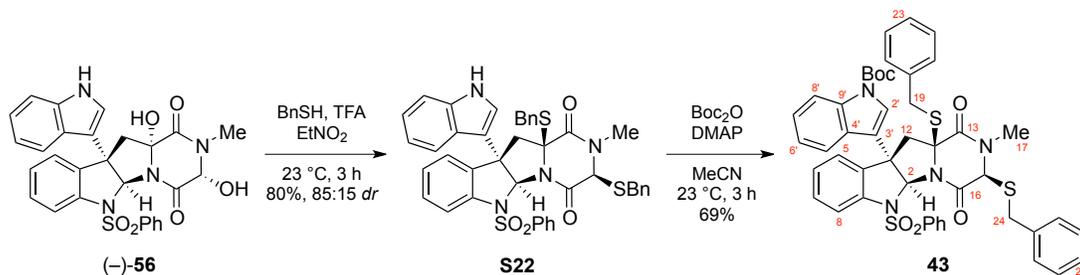
3395 (br-w), 3061 (w), 2924 (w), 2853 (w), 1690 (s), 1458 (w), 1382 (m), 1240 (w), 1170 (m), 1023 (w), 734 (m), 591 (m).

HRMS (ESI) (m/z):

calc'd for C₂₈H₂₂N₄NaO₄S₅ [M+Na]⁺: 661.0137, found 661.0120.

TLC (10% ethyl acetate in dichloromethane), R_f: 0.30 (UV, I₂, CAM).

³⁹ The isolation and purification of epitetrathiodiketopiperazine **28** were complicated by its instability in solution.



C3-(*N*-Boc-indol-3'-yl) bis(benzylthioether) **43**:

Trifluoroacetic acid (4 mL) was slowly added via syringe to a stirred solution of diol (**(-)-56**) (70.0 mg, 128.6 μmol , 1 equiv) and benzyl mercaptan (BnSH, 600 μL , 5.12 mmol, 39.7 equiv) in anhydrous nitroethane (5 mL) at 23 °C. After 3 h, the reaction mixture was diluted with ethyl acetate (100 mL) and slowly poured into saturated aqueous sodium hydrogenocarbonate solution (40 mL) at 23 °C. The organic layer was sequentially washed with water (3 \times 40 mL) and saturated aqueous sodium chloride solution (25 mL). The combined aqueous layers were extracted with ethyl acetate (2 \times 30 mL). The combined organic layers were dried over anhydrous sodium sulfate, were filtered, and were concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (eluent: gradient, 10 \rightarrow 40% ethyl acetate in hexanes) to afford the bis(benzylthioether) **S22**⁴⁰ (77.8 mg, 79.9%) as a pale yellow oil. {A minor diastereomer was also isolated from this reaction (13.0 mg, 13.3%)}

4-Dimethylaminopyridine (DMAP, 8.0 mg, 65.5 μmol , 0.83 equiv) was added as a solid to a solution of bis(benzylthioether) **S22** (60.0 mg, 79.3 μmol , 1 equiv) and di-*tert*-butyl dicarbonate (Boc₂O, 60.0 mg, 275 μmol , 3.47 equiv) in anhydrous acetonitrile (4 mL) at 23 °C. After 2 h, another portion of DMAP (2.5 mg, 20.5 μmol , 0.26 equiv) was added. After 1 h, the reaction mixture was diluted with ethyl acetate (60 mL). The resulting mixture was sequentially washed with aqueous 5% citric acid solution (30 mL), water (2 \times 20 mL), and saturated aqueous sodium chloride solution (20 mL). The organic layer was dried over anhydrous sodium sulfate, was filtered, and was concentrated under reduced pressure. The resulting residue was purified by flash column chromatography (eluent: gradient, 10 \rightarrow 50% ethyl acetate in hexanes) to afford the *N*-Boc-indole adduct **43** (47.0 mg, 69.2%) as a colorless oil. Structural assignments were made with additional information from gCOSY, HSQC, gHMBC, and NOESY data.

¹H NMR (600 MHz, CDCl₃, 20 °C):

δ 8.03 (br-s, 1H, C₈H), 7.79 (d, J = 8.2, 1H, C₈H), 7.47 (d, J = 7.4, 2H, C₂₁H), 7.45–7.39 (m, 3H, C₇H + SO₂Ph-*o*-H), 7.37 (dd, J = 7.5, 7.6, 2H, C₂₂H), 7.32–7.28 (m, 2H, C₇H + C₂₁H), 7.24 (dd, J = 7.4, 7.5, 1H, C₂₈H), 7.22–7.17 (m, 3H, C₂₆H + SO₂Ph-*p*-H), 7.17 (app-t, J = 7.5, 1H, C₆H), 7.11 (app-t, J = 7.6, 1H, C₆H), 7.01 (d, J = 7.5, 1H, C₅H), 7.00–6.92 (m, 5H, C₅H + C₂₇H + SO₂Ph-*m*-H), 6.68 (s, 1H, C₂H), 6.51 (br-s, 1H, C₂H), 4.47 (s, 1H, C₁₅H), 3.96 (d, J = 14.0, 1H, C₁₉H_a), 3.85 (d, J = 14.0, 1H,

⁴⁰ **S22**: ¹H NMR (600 MHz, CDCl₃, 20 °C): δ 7.85 (d, J = 8.0, 1H, C₈H), 7.75 (d, J = 7.4, 1H, C₈H), 7.41 (t, J = 7.5, 1H, SO₂Ph-*p*-H), 7.38–7.27 (m, 7H), 7.27–7.23 (m, 1H), 7.22–7.15 (m, 4H), 7.18 (app-t, J = 7.5, 2H, SO₂Ph-*m*-H), 7.14–7.10 (m, 2H, C₆H + C₆H), 7.10–7.05 (m, 2H), 6.80–6.76 (m, 2H), 6.71 (s, 1H, C₂H), 6.41 (d, J = 2.5, 1H, C₂H), 4.48 (s, 1H, C₁₅H), 4.06 (d, J = 12.9, 1H, C₁₉H_a), 3.81 (d, J = 13.6, 1H, C₂₄H_a), 3.79 (d, J = 12.8, 1H, C₁₉H_b), 3.76 (d, J = 13.7, 1H, C₂₄H_b), 3.39 (d, J = 14.4, 1H, C₁₂H_a), 2.83 (d, J = 14.4, 1H, C₁₂H_b), 2.53 (s, 3H, C₁₇H₃). MS (ESI) (m/z): [M+H]⁺: 757.56, [M+Na]⁺: 779.60, [M+K]⁺: 795.55. TLC (50% ethyl acetate in hexanes), R_f: 0.40 (UV, CAM).

$C_{19}\mathbf{H}_b$), 3.70 (d, $J = 12.1$, 1H, $C_{24}\mathbf{H}_a$), 3.51 (d, $J = 12.1$, 1H, $C_{24}\mathbf{H}_b$), 3.17 (d, $J = 14.7$, 1H, $C_{12}\mathbf{H}_a$), 2.86 (d, $J = 14.7$, 1H, $C_{12}\mathbf{H}_b$), 2.57 (s, 3H, $C_{17}\mathbf{H}_3$), 1.66 (s, 9H, $\text{OC}(\text{CH}_3)_3$).

^{13}C NMR (150 MHz, CDCl_3 , 20 °C):

δ 165.2 (C_{13}), 163.4 (C_{16}), 140.9 ($\text{C}_{\text{carbamate}}$), 142.1 (C_9), 138.3 (C_9), 137.3 ($\text{SO}_2\text{Ph-}i\text{pso-C}$), 136.0 (C_4), 136.0 (C_{25}), 135.7 (C_{20}), 132.7 ($\text{SO}_2\text{Ph-}p\text{-C}$), 129.9 (C_{21}), 129.7 (C_{26}), 129.7 ($\text{SO}_2\text{Ph-}m\text{-C}$), 129.5 (C_7), 128.9 (C_{22}), 128.5 ($\text{SO}_2\text{Ph-}o\text{-C}$), 128.4 (C_{27}), 127.8 (C_{23}), 127.4 (C_4), 127.2 (C_{28}), 126.0 (C_6), 125.1 (C_7), 124.7 (C_2), 124.1 (C_5), 123.3 (C_6), 120.0 (C_3), 119.2 (C_5), 119.1 (C_8), 115.9 (C_8), 84.4 ($\text{OC}(\text{CH}_3)_3$), 83.6 (C_2), 70.6 (C_{11}), 63.4 (C_{15}), 53.2 (C_3), 45.5 (C_{12}), 37.5 (C_{19}), 37.0 (C_{24}), 31.5 (C_{17}), 28.4 ($\text{OC}(\text{CH}_3)_3$).

FTIR (thin film) cm^{-1} :

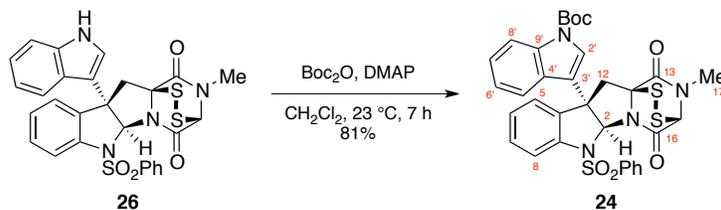
3214 (br-w), 3062 (w), 3027 (w), 2979 (w), 2930 (w), 2856 (w), 1734 (s), 1696 (s), 1668 (s), 1476 (m), 1454 (s), 1373 (s), 1270 (s), 1235 (s), 1171 (s), 1158 (s), 1097 (m), 1026 (m), 754 (s), 703 (m).

HRMS (ESI) (m/z):

calc'd for $\text{C}_{47}\text{H}_{44}\text{N}_4\text{NaO}_6\text{S}_3$ $[\text{M}+\text{Na}]^+$: 879.2315, found 879.2303.

TLC (30% ethyl acetate in hexanes), R_f :

0.33 (UV, CAM).



C3-(*N*-Boc-indol-3'-yl) epidithiodiketopiperazine 24:

A solution of DMAP in anhydrous dichloromethane (0.17 M, 25 μ L, 2.5 mol%) was added via syringe to a solution of epidithiodiketopiperazine **26** (98.3 mg, 171 μ mol, 1 equiv) and di-*tert*-butyl dicarbonate (77.6 mg, 355 μ mol, 2.08 equiv) in anhydrous dichloromethane (20 mL) at 23 $^{\circ}$ C. After 2 h, another portion of DMAP solution (25 μ L, 2.5 mol%) was added. After 5 h, the reaction mixture was diluted with ethyl acetate (100 mL). The resulting mixture was sequentially washed with aqueous 5% citric acid solution (50 mL), water (2 \times 50 mL), and saturated aqueous sodium chloride solution (30 mL). The organic layer was dried over anhydrous sodium sulfate, was filtered, and was concentrated under reduced pressure. The resulting residue was purified by flash column chromatography (eluent: gradient, 30 \rightarrow 60% ethyl acetate in hexanes) to afford the *N*-Boc-epidithiodiketopiperazine **24** (93.3 mg, 80.9%) as a pale yellow oil. Structural assignments were made with additional information from gCOSY, HSQC, and gHMBC data.

^1H NMR (600 MHz, CDCl_3 , 20 $^{\circ}$ C):

δ 8.05 (br-s, 1H, C_8H), 7.85 (d, $J = 8.1$, 1H, C_8H), 7.48 (app-dt, $J = 4.5, 8.1$, 1H, C_7H), 7.38 (app-dt, $J = 1.3, 7.7$, 1H, $\text{SO}_2\text{Ph-}p\text{-H}$), 7.55 (d, $J = 7.1$, 1H, C_5H), 7.34–7.30 (m, 2H, $\text{C}_5\text{H} + \text{C}_6\text{H}$), 7.28 (dd, $J = 7.1, 7.3$, 1H, C_7H), 7.17 (app-t, $J = 7.4$, 1H, C_6H), 7.13 (d, $J = 7.6$, 2H, $\text{SO}_2\text{Ph-}o\text{-H}$), 6.82 (dd, $J = 7.6, 8.1$, 2H, $\text{SO}_2\text{Ph-}m\text{-H}$), 6.55 (s, 1H, C_2H), 6.18 (br-s, 1H, C_2H), 5.29 (s, 1H, C_{15}H), 3.88 (d, $J = 15.6$, 1H, C_{12}H_a), 3.17 (s, 3H, C_{17}H_3), 2.67 (d, $J = 15.6$, 1H, C_{12}H_b), 1.66 (s, 9H, $\text{OC}(\text{CH}_3)_3$).

^{13}C NMR (150 MHz, CDCl_3 , 20 $^{\circ}$ C):

δ 165.1 (C_{13}), 160.3 (C_{16}), 149.2 ($\text{C}_{\text{carbamate}}$), 141.0 (C_9), 137.5 ($\text{SO}_2\text{Ph-}ipso\text{-C}$), 137.5 (C_9), 135.9 (C_4), 132.8 ($\text{SO}_2\text{Ph-}p\text{-C}$), 130.3 (C_7), 128.1 ($\text{SO}_2\text{Ph-}m\text{-C}$), 127.1 ($\text{SO}_2\text{Ph-}o\text{-C}$), 126.7 (C_6), 125.6 (C_4), 125.4 (C_5), 124.5 (C_2), 123.6 (C_7), 123.6 (C_6), 120.1 (C_5), 119.0 (C_8), 118.5 (C_3), 116.0 (C_8), 84.6 ($\text{OC}(\text{CH}_3)_3$), 84.1 (C_2), 74.4 (C_{11}), 68.5 (C_{15}), 55.2 (C_3), 42.2 (C_{12}), 32.3 (C_{17}), 28.4 ($\text{OC}(\text{CH}_3)_3$).

FTIR (thin film) cm^{-1} :

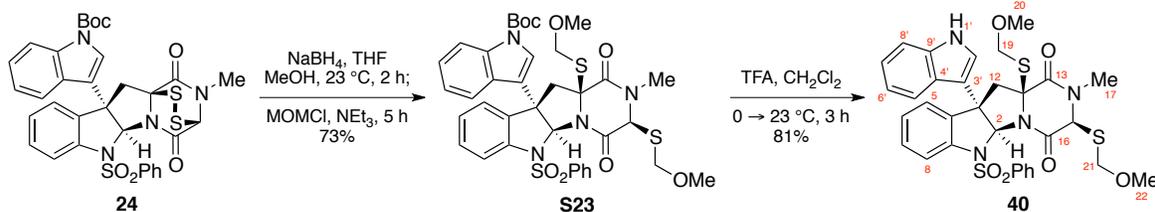
2978 (w), 2929 (w), 1733 (s), 1677 (m), 1454 (m), 1371 (s), 1256 (m), 1157 (s), 1092 (m), 751 (s).

HRMS (ESI) (m/z):

calc'd for $\text{C}_{33}\text{H}_{30}\text{N}_4\text{NaO}_6\text{S}_3$ $[\text{M}+\text{Na}]^+$: 697.1220, found 697.1231.

TLC (50% ethyl acetate in hexanes), R_f :

0.39 (UV, I_2 , CAM).



C3-(*N*-Boc-Indol-3'-yl) bis(*S*-MOM)ether **40**:

Sodium borohydride (50.0 mg, 1.32 mmol, 6.06 equiv) was added as a solid to a solution of epidithiodiketopiperazine **24** (147 mg, 218 μ mol, 1 equiv) in anhydrous tetrahydrofuran (15 mL) and anhydrous methanol (60.0 μ L) at 23 °C. After 2 h, chloromethyl methyl ether (MOMCl, 500 μ L, 1.42 mmol, 30.4 equiv) was added to the reaction mixture. After 1 h, triethylamine (200 μ L, 1.42 mmol, 6.53 equiv) was added to the reaction mixture. After 4 h, the white reaction mixture was diluted with ethyl acetate (100 mL) and washed with saturated aqueous ammonium chloride solution (30 mL). The aqueous layer was extracted with ethyl acetate (2 \times 20 mL). The combined organic layers were washed sequentially with water (2 \times 30 mL) and saturated aqueous sodium chloride solution (20 mL), were dried over anhydrous sodium sulfate, were filtered, and were concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (eluent: gradient, 30 \rightarrow 70% ethyl acetate in hexanes) to afford the bis(*S*-MOM) derivative **S23** (123 mg, 73.4%) as a colorless oil.⁴¹

Trifluoroacetic acid (2 mL) was added to a solution of the *N*-Boc-indole **S23** (6.1 mg, 7.8 μ mol, 1 equiv) in anhydrous dichloromethane (5 mL) at 0 °C. After 30 min, the ice–water bath was removed, and the solution was allowed to warm to 23 °C. After 3 h, the reaction mixture was diluted with ethyl acetate (50 mL) and slowly poured into saturated aqueous sodium hydrogenocarbonate solution (25 mL). The organic layer was sequentially washed with water (3 \times 15 mL) and saturated aqueous sodium chloride solution (15 mL). The combined aqueous layers were extracted with ethyl acetate (2 \times 20 mL). The combined organic layers were dried over anhydrous sodium sulfate, were filtered, and were concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (eluent: gradient, 10 \rightarrow 40% ethyl acetate in hexanes) to afford the bis(*S*-MOM)ether **40** (4.3 mg, 81%) as a pale yellow oil. Structural assignments were made with additional information from gCOSY, HSQC, gHMBC, and NOESY data.

¹H NMR (600 MHz, CDCl₃, 20 °C):

δ 7.87 (br-s, 1H, C₁H), 7.79 (d, J = 8.2, 1H, C₈H), 7.72 (d, J = 7.7, 2H, SO₂Ph-*o*-H), 7.41 (app-dd, J = 7.4, 7.5, 1H, SO₂Ph-*p*-H), 7.32 (app-dt, J = 0.9, 7.8, 1H, C₇H), 7.29 (d, J = 8.1, 1H, C₈H), 7.19 (dd, J = 7.7, 8.0, 2H, SO₂Ph-*m*-H), 7.14 (app-t, J = 7.5, 1H, C₇H), 7.14 (d, J = 7.4, 1H, C₅H), 7.08 (dd, J = 7.4, 7.7, 1H, C₆H), 6.83 (dd, J = 7.4, 7.8, 1H, C₆H), 6.70 (d, J = 8.0, 1H, C₅H), 6.68 (s, 1H, C₂H), 6.51 (d, J = 2.5, 1H, C₂H), 5.17 (d, J = 11.8, 1H, C₂₁H_a), 5.11

⁴¹ **S23**: ¹H NMR (600 MHz, CDCl₃, 20 °C): δ 8.03 (br-s, 1H, C₈H), 7.84 (app-dd, J = 0.5, 8.2, 1H, C₈H), 7.53 (br-d, J = 7.4, 2H, SO₂Ph-*o*-H), 7.38 (app-ddd, J = 1.5, 7.4, 8.2, 1H, C₇H), 7.30 (app-t, J = 7.6, 1H, C₇H), 7.27 (app-dt, J = 0.9, 8.3, 1H, C₆H), 7.19 (app-dd, J = 0.8, 7.5, 1H, C₅H), 7.15 (app-dt, J = 0.9, 7.4, 1H, C₆H), 7.05 (dd, J = 7.7, 7.8, 2H, SO₂Ph-*m*-H), 7.00 (dd, J = 7.4, 7.6, 2H, SO₂Ph-*p*-H), 6.73 (d, J = 7.8, 1H, C₅H), 6.72 (s, 1H, C₂H), 6.65 (s, 1H, C₂H), 5.21 (d, J = 11.8, 1H, C₂₁H_a), 5.08 (app-d, J = 12.7, 1H, C₁₉H_a), 4.95 (s, 1H, C₁₅H), 4.46 (d, J = 11.8, 1H, C₂₁H_b), 4.30 (d, J = 12.7, 1H, C₁₉H_b), 3.46 (s, 3H, C₂₂H₃), 3.39 (d, J = 14.7, 1H, C₁₂H_a), 3.23 (d, J = 14.7, 1H, C₁₂H_b), 3.08 (s, 3H, C₁₇H₃), 2.92 (s, 3H, C₂₀H₃), 1.66 (s, 9H, OC(CH₃)₃). TLC (50% ethyl acetate in hexanes), R_f: 0.49 (UV, CAM).

(d, $J = 12.7$, 1H, C₁₉H_a), 4.91 (s, 1H, C₁₅H), 4.44 (d, $J = 11.8$, 1H, C₂₁H_b), 4.35 (d, $J = 12.7$, 1H, C₁₉H_b), 3.51 (d, $J = 14.7$, 1H, C₁₂H_a), 3.45 (s, 3H, C₂₂H₃), 3.29 (d, $J = 14.7$, 1H, C₁₂H_b), 3.07 (s, 3H, C₁₇H₃), 2.93 (s, 3H, C₂₀H₃).

¹³C NMR (150 MHz, CDCl₃, 20 °C):

δ 165.7 (C₁₃), 163.2 (C₁₆), 141.6 (C₉), 138.4 (SO₂Ph-*ipso*-C), 137.3 (C₉), 136.4 (C₄), 133.0 (SO₂Ph-*p*-C), 129.0 (C₇), 128.8 (SO₂Ph-*m*-C), 127.5 (SO₂Ph-*o*-C), 125.2 (C₆), 125.0 (C₅), 124.5 (C₄), 122.9 (C₂), 122.7 (C₇), 120.4 (C₆), 119.1 (C₅), 117.0 (C₈), 116.5 (C₃), 111.7 (C₈), 84.6 (C₂), 76.5 (C₂₁), 75.5 (C₁₉), 70.5 (C₁₁), 64.9 (C₁₅), 56.8 (C₂₀), 56.6 (C₂₂), 53.7 (C₃), 49.2 (C₁₂), 32.3 (C₁₇).

FTIR (thin film) cm⁻¹:

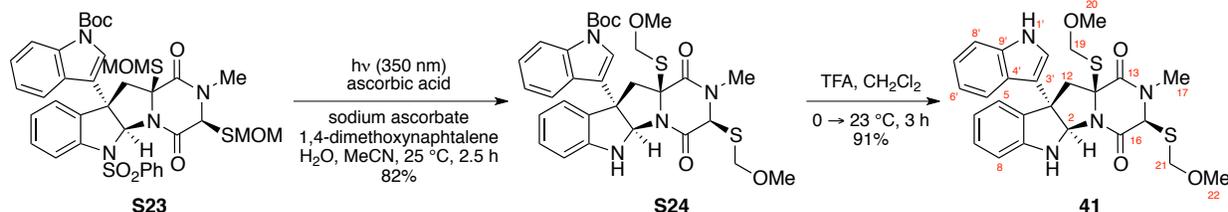
3390 (w), 3004 (w), 2927 (w), 2823 (w), 1693 (s), 1666 (s), 1461 (m), 1392 (s), 1364 (s), 1312 (m), 1265 (w), 1235 (w), 1181 (s), 1084 (s), 751 (s).

HRMS (ESI) (m/z):

calc'd for C₃₂H₃₂N₄NaO₆S₃ [M+Na]⁺: 687.1376, found: 687.1378.

TLC (50% ethyl acetate in hexanes), R_f:

0.38 (UV, CAM).



C3-(Indol-3'-yl) bis(*S*-MOM)ether **41**:

A 20 × 150 mm Pyrex tube was sequentially charged with bis(*S*-MOM)ether **S23** (92.2 mg, 121 μmol, 1 equiv), L-ascorbic acid (310 mg, 1.76 mmol, 14.6 equiv), sodium L-ascorbate (380 mg, 1.92 mmol, 15.9 equiv), and 1,4-dimethoxynaphthalene (1.25 g, 6.64 mmol, 55.1 equiv), and the mixture was placed under an argon atmosphere. A solution of water in acetonitrile (20% v/v, 24 mL) that was purged with argon for 15 min at 23 °C was transferred to the flask via cannula. The system was vigorously stirred under an argon atmosphere and irradiated with a Rayonet photoreactor equipped with 16 lamps emitting at 350 nm at 25 °C. After 2.5 h, the lamps were turned off, and the reaction mixture was diluted with ethyl acetate (100 mL) and diethyl ether (50 mL). The resulting solution was sequentially washed with saturated aqueous sodium hydrogenocarbonate solution (50 mL), water (2 × 40 mL), and saturated aqueous sodium chloride solution (40 mL). The aqueous layer was extracted with ethyl acetate (2 × 50 mL). The combined organic layers were dried over anhydrous sodium sulfate, were filtered, and were concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (eluent: gradient, 20 → 60% ethyl acetate in hexanes) to afford aniline **S24** (61.7 mg, 81.9%) as a pale yellow oil.⁴²

Trifluoroacetic acid (2 mL) was added to a solution of the *N*-Boc-indole **S24** (6.0 mg, 9.6 μmol, 1 equiv) in anhydrous dichloromethane (5 mL) at 0 °C. After 30 min, the ice–water bath was removed, and the solution was allowed to warm to 23 °C. After 3 h, the reaction mixture was diluted with ethyl acetate (50 mL) and slowly poured into saturated aqueous sodium hydrogenocarbonate solution (25 mL) at 23 °C. The organic layer was sequentially washed with water (3 × 15 mL) and saturated aqueous sodium chloride solution (15 mL). The combined aqueous layers were extracted with ethyl acetate (2 × 20 mL). The combined organic layers were dried over anhydrous sodium sulfate, were filtered, and were concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (eluent: gradient, 20 → 60% ethyl acetate in hexanes) to afford the bis(*S*-MOM)ether **41** (4.6 mg, 91%) as a colorless oil. Structural assignments were made with additional information from gCOSY, HSQC, gHMBC, and NOESY data.

¹H NMR (600 MHz, CDCl₃, 20 °C):

δ 7.98 (br-s, 1H, N₁H), 7.38 (d, *J* = 8.0, 1H, C₅H), 7.32 (d, *J* = 8.2, 1H, C₈H), 7.16 (app-t, *J* = 7.2, 1H, C₇H), 7.14 (d, *J* = 6.9, 1H, C₅H), 7.10 (app-dt, *J* = 1.0, 7.8, 1H, C₇H), 7.02 (app-t, *J* = 7.2, 1H, C₆H), 7.01 (d, *J* = 2.7, 1H, C₂H), 6.73 (dd, *J* = 7.3, 7.5, 1H, C₆H), 6.71 (d, *J* = 7.8, 1H, C₈H), 6.05 (s, 1H, C₂H), 5.22 (d, *J* = 11.7, 1H, C₂₁H_a), 5.13 (d, *J* =

⁴² **S24**: ¹H NMR (600 MHz, CDCl₃, 20 °C): δ 8.11 (br-s, 1H, N₁H), 7.46 (br-s, 1H, C₈H), 7.37 (d, *J* = 7.9, 1H, C₅H), 7.26 (app-t, *J* = 7.6, 1H, C₇H), 7.10 (d, *J* = 7.2, 1H, C₅H), 7.13–7.08 (m, 3H, C₆H + C₇H + C₈H), 6.74–6.67 (m, 2H, C₂H + C₆H), 6.04 (s, 1H, C₂H), 5.19 (d, *J* = 11.7, 1H, C₂₁H_a), 5.16 (d, *J* = 12.6, 1H, C₁₉H_a), 4.90 (s, 1H, C₁₅H), 4.52 (d, *J* = 11.7, 1H, C₂₁H_b), 4.31 (d, *J* = 12.6, 1H, C₁₉H_b), 3.55 (d, *J* = 14.1, 1H, C₁₂H_a), 3.45 (d, *J* = 14.1, 1H, C₁₂H_b), 3.47 (s, 3H, C₂₂H₃), 3.06 (s, 3H, C₁₇H₃), 2.91 (s, 3H, C₂₀H₃), 1.65 (s, 9H, OC(CH₃)₃). HRMS (ESI) (*m/z*): calc'd for C₃₁H₃₆N₄NaO₆S₂ [M+Na]⁺: 647.1968, found: 647.1976. TLC (50% ethyl acetate in hexanes), R_f: 0.74 (UV, CAM).

12.6, 1H, C₁₉H_a), 4.93 (s, 1H, C₁₅H), 4.53 (d, *J* = 11.7, 1H, C₂₁H_b), 4.34 (d, *J* = 12.6, 1H, C₁₉H_b), 3.48 (s, 2H, C₁₂H), 3.48 (s, 3H, C₂₂H₃), 3.07 (s, 3H, C₁₇H₃), 2.95 (s, 3H, C₂₀H₃).³⁷

¹³C NMR (150 MHz, CDCl₃, 20 °C):

δ 166.1 (C₁₃), 165.4 (C₁₆), 148.5 (C₉), 137.4 (C₉), 132.8 (C₄), 128.6 (C₇), 125.3 (C₄), 124.9 (C₅), 122.6 (C₇), 121.6 (C₂), 120.2 (C₆), 119.9 (C₅), 119.6 (C₆), 119.4 (C₃), 111.6 (C₈), 109.3 (C₈), 82.6 (C₂), 77.0 (C₂₁), 75.7 (C₁₉), 69.3 (C₁₁), 65.1 (C₁₅), 57.0 (C₂₀), 56.5 (C₂₂), 54.3 (C₃), 48.0 (C₁₂), 32.0 (C₁₇).

FTIR (thin film) cm⁻¹:

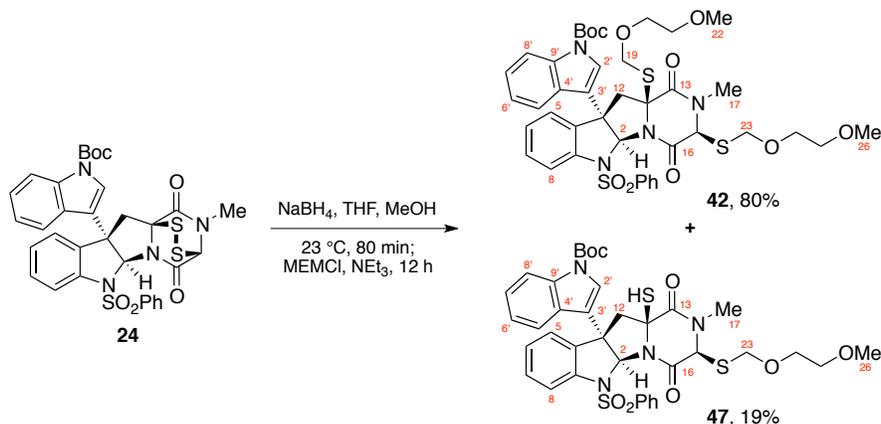
3394 (br-w), 3013 (w), 2928 (w), 2823 (w), 1693 (s), 1669 (s), 1461 (m), 1393 (m), 1363 (m), 1265 (w), 1180 (s), 752 (s).

HRMS (ESI) (*m/z*):

calc'd for C₂₆H₂₈N₄NaO₄S₂ [M+Na]⁺: 547.1444, found: 547.1434.

TLC (50% ethyl acetate in hexanes), R_f:

0.43 (UV, CAM).



C3-(*N*-Boc-Indol-3'-yl) bis(*S*-MEM)ether **42 and C3-(*N*-Boc-indol-3'-yl) *S*15-MEM ether **47**:**

Sodium borohydride (9.8 mg, 250 μmol, 3.6 equiv) was added as a solid to a solution of epidithiodiketopiperazine **24** (47.0 mg, 69.6 μmol, 1 equiv) in anhydrous tetrahydrofuran (8 mL) and anhydrous methanol (50 μL) at 23 °C. After 80 min, 2-methoxyethoxymethyl chloride (MEMCl, 300 μL, 2.63 mmol, 37.7 equiv) followed by triethylamine (400 μL, 2.85 mmol, 40.9 equiv) were added to the reaction mixture. After 12 h, the yellow reaction mixture was partitioned between aqueous 5% citric acid solution (30 mL) and ethyl acetate (80 mL). The isolated organic layer was washed sequentially with water (2 × 30 mL) and saturated aqueous sodium chloride solution (20 mL), was dried over anhydrous sodium sulfate, was filtered, and was concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (eluent: gradient, 10 → 25% ethyl acetate in dichloromethane) to afford the bis(*S*-MEM)ether adduct **42** (57.6 mg, 80.2%) and the *S*15-MEM-adduct **47** (10.0 mg, 18.8%) as colorless oils. Structural assignments were made with additional information from gCOSY, HSQC, gHMBC, and NOESY data.

C3-(*N*-Boc-indol-3'-yl) bis(*S*-MEM)ether **42:**

¹H NMR (600 MHz, CDCl₃, 20 °C):

δ 8.02 (br-s, 1H, C₈H), 7.83 (d, *J* = 8.2, 1H, C₈H), 7.56 (br-d, *J* = 6.2, 2H, SO₂Ph-*o*-H), 7.36 (dd, *J* = 7.7, 8.0, 1H, C₇H), 7.32 (t, *J* = 7.3, 1H, SO₂Ph-*p*-H), 7.24 (d, *J* = 7.8, 1H, C₅H), 7.16 (d, *J* = 7.4, 1H, C₅H), 7.10 (app-t, *J* = 7.6, 1H, C₆H), 7.08 (app-t, *J* = 7.5, 2H, SO₂Ph-*m*-H), 6.96 (app-t, *J* = 7.5, 1H, C₆H), 6.76 (s, 1H, C₂H), 6.67–6.61 (m, 1H, C₇H), 6.62 (s, 1H, C₂H), 5.21 (d, *J* = 12.0, 1H, C₂₃H_a), 5.13 (d, *J* = 12.8, 1H, C₁₉H_a), 5.00 (s, 1H, C₁₅H), 4.63 (d, *J* = 12.0, 1H, C₂₃H_b), 4.47 (d, *J* = 12.8, 1H, C₁₉H_b), 4.00–3.94 (m, 1H, C₂₄H_a), 3.67–3.64 (m, 2H, C₂₄H_b + C₂₅H_a), 3.61–3.56 (m, 1H, C₂₅H_b), 3.41 (d, *J* = 14.9, 1H, C₁₂H_a), 3.39 (s, 3H, C₂₆H₃), 3.38–3.33 (m, 2H, C₂₁H), 3.31 (s, 3H, C₂₂H₃), 3.28–3.23 (m, 1H, C₂₀H_a), 3.23 (d, *J* = 14.9, 1H, C₁₂H_b), 3.20–3.14 (m, 1H, C₂₀H_b), 3.09 (s, 3H, C₁₇H₃), 1.69 (s, 9H, (OC(CH₃)₃)).

¹³C NMR (150 MHz, CDCl₃, 20 °C):

δ 165.3 (C₁₃), 163.1 (C₁₆), 149.3 (C_{carbamate}), 141.7 (C₉), 137.9 (SO₂Ph-*ipso*-C), 136.3 (C₉), 135.7 (C₄), 133.0 (SO₂Ph-*p*-C), 129.3 (C₇), 128.6 (SO₂Ph-*m*-C),

	127.4 (SO ₂ Ph- <i>o</i> -C), 127.2 (C _{4'}), 125.5 (C ₆), 125.0 (C ₅), 124.9 (C ₅), 124.5 (C ₇), 123.2 (C ₆), 120.3 (C ₃), 119.1 (C ₂), 117.9 (C ₈), 115.7 (C ₈), 84.4 (OC(CH ₃) ₃), 83.8 (C ₂), 75.2 (C ₂₃), 74.0 (C ₁₉), 71.6 (C ₂₅), 71.6 (C ₂₁), 70.3 (C ₁₁), 68.2 (C ₂₀), 68.1 (C ₂₄), 64.9 (C ₁₅), 59.3 (C ₂₂), 59.2 (C ₂₆), 53.3 (C ₃), 48.6 (C ₁₂), 32.2 (C ₁₇), 28.4 (OC(CH ₃) ₃).
FTIR (thin film) cm ⁻¹ :	2920 (m), 2851 (m), 1734 (s), 1699 (s), 1668 (s), 1454 (s), 1373 (s), 1310 (m), 1272 (m), 1158 (s), 1088 (s), 1025 (m), 752 (s).
HRMS (ESI) (<i>m/z</i>):	calc'd for C ₄₁ H ₄₈ N ₄ NaO ₁₀ S ₃ [M+Na] ⁺ : 875.2425, found: 875.2411.
TLC (20% ethyl acetate in dichloromethane), R _f :	0.44 (UV, I ₂ , CAM).

C3-(N-Boc-indol-3'-yl) S15-MEM ether 47:

¹H NMR (600 MHz, CDCl₃, 20 °C):

δ 8.04 (br-s, 1H, C₈H), 7.85 (d, *J* = 8.1, 1H, C₈H), 7.45 (app-dt, *J* = 1.0, 8.0, 1H, C₇H), 7.38 (br-d, *J* = 6.2, 2H, SO₂Ph-*o*-H), 7.31 (app-t, *J* = 7.8, 1H, C₇H), 7.29–7.21 (m, 3H, C₅H + C₆H + SO₂Ph-*p*-H), 7.12 (dd, *J* = 7.4, 7.6, 1H, C₆H), 6.95 (app-t, *J* = 7.7, 2H, SO₂Ph-*m*-H), 6.91 (d, *J* = 7.8, 1H, C₅H), 6.67 (s, 1H, C₂H), 6.53 (s, 1H, C₂H), 5.25 (d, *J* = 11.9, 1H, C₂₃H_a), 5.09 (s, 1H, C₁₅H), 4.71 (d, *J* = 11.9, 1H, C₂₃H_b), 4.00–3.96 (m, 1H, C₂₄H_a), 3.70–3.62 (m, 2H, C₂₄H_b + C₂₅H_a), 3.62–3.58 (m, 1H, C₂₅H_b), 3.43 (d, *J* = 14.6, 1H, C₁₂H_a), 3.40 (s, 3H, C₂₆H₃), 3.13 (s, 3H, C₁₇H₃), 2.87 (d, *J* = 14.6, 1H, C₁₂H_b), 1.66 (s, 9H, (OC(CH₃)₃)).

¹³C NMR (150 MHz, CDCl₃, 20 °C):

δ 167.5 (C₁₃), 162.2 (C₁₆), 149.2 (C_{carbamate}), 142.2 (C₉), 137.8 (SO₂Ph-*ipso*-C), 135.6 (C₉), 135.6 (C₄), 132.8 (SO₂Ph-*p*-C), 129.9 (C₇), 128.3 (SO₂Ph-*m*-C), 127.3 (SO₂Ph-*o*-C), 126.9 (C_{4'}), 126.3 (C₆), 125.2 (C₇), 124.9 (C₅), 124.8 (C₂), 123.4 (C₆), 119.6 (C₃), 119.3 (C₈), 119.0 (C₅), 115.9 (C₈), 84.5 (OC(CH₃)₃), 83.8 (C₂), 74.0 (C₂₃), 71.6 (C₂₅), 68.4 (C₂₄), 68.1 (C₁₁), 64.3 (C₁₅), 59.3 (C₂₆), 53.4 (C₃), 51.2 (C₁₂), 32.7 (C₁₇), 28.4 (OC(CH₃)₃).

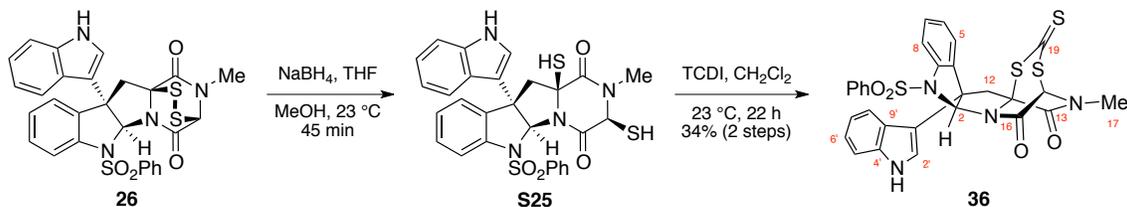
FTIR (thin film) cm⁻¹:

2978 (w), 2922 (w), 1734 (m), 1697 (m), 1454 (m), 1372 (s), 1272 (w), 1235 (w), 1157 (m), 1091 (m), 752 (s).

HRMS (ESI) (*m/z*):

calc'd for C₃₇H₄₀N₄NaO₈S₃ [M+Na]⁺: 787.1900, found: 787.1897.

TLC (20% ethyl acetate in dichloromethane), R_f: 0.24 (UV, I₂, CAM).



C3-(Indol-3'-yl) dithiepanethione 36:

Sodium borohydride (4.9 mg, 0.13 mmol, 3.4 equiv) was added as a solid to a solution of epidithiodiketopiperazine **26** (22.0 mg, 38.3 μ mol, 1 equiv) in anhydrous tetrahydrofuran (5 mL) and anhydrous methanol (50 μ L) at 23 °C. After 45 min, the reaction mixture was diluted with ethyl acetate (40 mL) and washed with saturated aqueous ammonium chloride solution (20 mL). The aqueous layer was extracted with ethyl acetate (2 \times 20 mL) and the combined organic layers were washed sequentially with water (2 \times 20 mL) and saturated aqueous sodium chloride solution (20 mL), were dried over anhydrous sodium sulfate, were filtered, and were concentrated under reduced pressure to afford the hexacyclic bisthiol **S25** that was used in the next step without further purification.

1,1'-Thiocarbonyldiimidazole (TCDI, 108 mg, 606 μ mol, 15.8 equiv) was added as a solid to the solution of bisthiol **S25** in anhydrous dichloromethane (6 mL) at 23 °C. After 22 h, the volatiles were removed under reduced pressure. The residue was purified by flash column chromatography on silica gel (eluent: gradient, 5 \rightarrow 25% ethyl acetate in dichloromethane) to afford the dithiepanethione **36** (8.4 mg, 34%) as a pale yellow oil. Structural assignments were made with additional information from gCOSY, HSQC, and gHMBC data.⁴³

¹H NMR (600 MHz, CDCl₃, 20 °C):

δ 7.86 (br-s, 1H, N₁H), 7.75 (d, J = 8.2, 1H, C₈H), 7.49 (app-dd, J = 1.0, 8.3, 2H, SO₂Ph-*o*-H), 7.43–7.36 (m, 2H, C₇H + SO₂Ph-*p*-H), 7.35 (d, J = 8.1, 1H, C₈H), 7.23–7.17 (m, 3H, C₃H + C₆H + C₇H), 7.10 (dd, J = 7.6, 8.1, 2H, SO₂Ph-*m*-H), 6.98 (app-dt, J = 0.5, 7.5, 1H, C₆H), 6.89 (d, J = 8.0, 1H, C₅H), 6.64 (s, 1H, C₂H), 6.36 (d, J = 2.5, 1H, C₂H), 5.05 (s, 1H, C₁₅H), 3.96 (d, J = 15.6, 1H, C₁₂H_a), 3.18 (s, 3H, C₁₇H₃), 2.80 (d, J = 15.6, 1H, C₁₂H_b).

¹³C NMR (150 MHz, CDCl₃, 20 °C):

δ 214.3 (C₁₉), 164.3 (C₁₃), 159.4 (C₁₆), 140.9 (C₉), 137.5 (SO₂Ph-*ipso*-C), 137.3 (C₉), 135.1 (C₄), 133.2 (SO₂Ph-*p*-C), 130.2 (C₇), 128.7 (SO₂Ph-*m*-C), 127.4 (SO₂Ph-*o*-C), 126.2 (C₆), 124.6 (C₅), 124.2 (C₄), 124.1 (C₂), 123.2 (C₇), 120.7 (C₆), 118.8 (C₅), 118.5 (C₈), 113.8 (C₃), 112.0 (C₈), 85.3 (C₂), 75.3 (C₁₁), 69.5 (C₁₅), 54.6 (C₃), 45.8 (C₁₂), 32.7 (C₁₇).

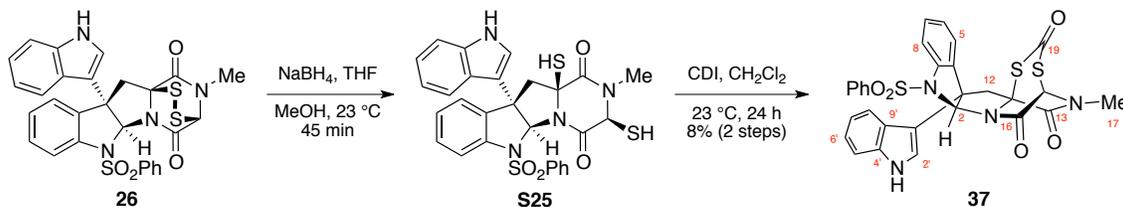
FTIR (thin film) cm⁻¹:

3393 (br-w), 2921 (m), 2851 (w), 1703 (s), 1459 (m), 1361 (m), 1168 (m), 1089 (w), 1016 (w), 907 (w), 733 (m).

⁴³ Upon concentration or in concentrated solution, the dithiepanethione **36** tends to degrade, thus rendering its isolation and characterization particularly arduous.

HRMS (ESI) (m/z): calc'd for $C_{29}H_{23}N_4O_4S_4$ $[M+H]^+$: 619.0597, found 619.0609; calc'd for $C_{29}H_{22}N_4NaO_4S_4$ $[M+Na]^+$: 641.0416, found 641.0424.

TLC (20% ethyl acetate in dichloromethane), R_f : 0.68 (UV, I_2 , CAM).



C3-(Indol-3'-yl) dithiocarbonate 37:

Sodium borohydride (4.9 mg, 0.13 mmol, 3.3 equiv) was added as a solid to a solution of epidithiodiketopiperazine **26** (22.6 mg, 39.3 μ mol, 1 equiv) in anhydrous tetrahydrofuran (5 mL) and anhydrous methanol (50 μ L) at 23 °C. After 45 min, the reaction mixture was diluted with ethyl acetate (40 mL) and washed with saturated aqueous ammonium chloride solution (20 mL). The aqueous layer was extracted with ethyl acetate (2 \times 20 mL) and the combined organic layers were washed sequentially with water (2 \times 20 mL) and saturated aqueous sodium chloride solution (20 mL), were dried over anhydrous sodium sulfate, were filtered, and were concentrated under reduced pressure to afford the hexacyclic bithiol **S25** that was used in the next step without further purification.

1,1'-Carbonyldiimidazole (CDI, 80.0 mg, 493 μ mol, 12.0 equiv) was added as a solid to the solution of bithiol **S25** in anhydrous dichloromethane (10 mL) at 23 °C. After 24 h, the volatiles were removed under reduced pressure. The residue was purified by flash column chromatography on silica gel (eluent: gradient, 5 \rightarrow 20% ethyl acetate in dichloromethane) to afford the dithiocarbonate **37** along with epidithiodiketopiperazine **26**. Both compounds were separated by preparative HPLC [Waters X-Bridge preparative HPLC column, C18, 5 μ m, 19 \times 250 mm; 20.0 mL/min; gradient, 20 \rightarrow 90% acetonitrile in water, 20 min; t_R (**37**) = 15.35 min, t_R (**26**) = 14.50 min] to afford **37** (2.0 mg, 8%) as a pale yellow oil.⁴⁴ Structural assignments were made with additional information from gCOSY, HSQC, and gHMBC data.

¹H NMR (600 MHz, CDCl₃, 20 °C):

δ 7.84 (br-s, 1H, N₁H), 7.78 (d, J = 8.2, 1H, C₈H), 7.45 (d, J = 8.4, 2H, SO₂Ph-*o*-H), 7.41 (app-ddd, J = 1.6, 7.3, 7.7, 1H, C₇H), 7.37 (app-dt, J = 1.0, 6.4, 1H, SO₂Ph-*p*-H), 7.35 (d, J = 8.2, 1H, C₈H), 7.21 (app-ddd, J = 0.8, 7.2, 7.6, 1H, C₇H), 7.19 (app-dt, J = 0.9, 7.3, 1H, C₆H), 7.16 (app-dd, J = 1.1, 7.6, 1H, C₅H), 7.06 (dd, J = 7.6, 8.3, 2H, SO₂Ph-*m*-H), 6.99 (app-dt, J = 0.7, 7.5, 1H, C₆H), 6.90 (d, J = 7.9, 1H, C₅H), 6.64 (s, 1H, C₂H), 6.26 (d, J = 2.6, 1H, C₂H), 5.17 (s, 1H, C₁₅H), 3.92 (d, J = 15.5, 1H, C₁₂H_a), 3.20 (s, 3H, C₁₇H₃), 2.78 (d, J = 15.5, 1H, C₁₂H_b).

¹³C NMR (150 MHz, CDCl₃, 20 °C):

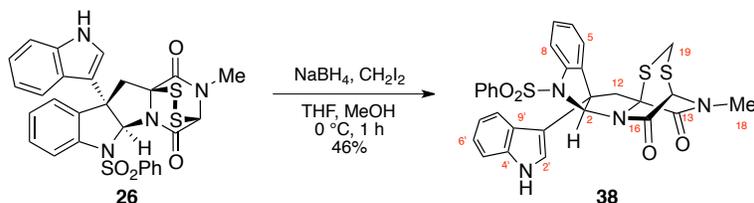
δ 185.4 (C₁₉), 165.0 (C₁₃), 160.0 (C₁₆), 141.0 (C₉), 137.3 (SO₂Ph-*ipso*-C), 137.3 (C₉), 135.2 (C₄), 133.2 (SO₂Ph-*p*-C), 130.2 (C₇), 128.7 (SO₂Ph-*m*-C), 127.5 (SO₂Ph-*o*-C), 126.2 (C₆), 124.6 (C₅), 124.1 (C₄), 124.1 (C₂), 123.2 (C₇), 120.7 (C₆), 118.7 (C₅), 118.7 (C₈), 113.8 (C₃), 112.0 (C₈), 85.3 (C₂), 72.6 (C₁₁), 66.6 (C₁₅), 54.5 (C₃), 46.5 (C₁₂), 32.6 (C₁₇).

⁴⁴ Epidithiodiketopiperazine **26** was also recovered (2.9 mg, 12%).

FTIR (thin film) cm^{-1} : 3396 (br-m), 2924 (m), 2853 (w), 1696 (m), 1460 (m), 1383 (m), 1169 (m), 1091 (w), 1051 (w), 735 (m).

HRMS (ESI) (m/z): calc'd for $\text{C}_{29}\text{H}_{22}\text{N}_4\text{NaO}_5\text{S}_3$ $[\text{M}+\text{Na}]^+$: 625.0645, found 625.0652.

TLC (20% ethyl acetate in dichloromethane), R_f : 0.57 (UV, I_2 , CAM).



C3-(Indol-3'-yl) dithioacetal **38:**

Sodium borohydride (15.0 mg, 0.400 mmol, 9.88 equiv) was added as a solid to a solution of epidithiodiketopiperazine **26** (23.1 mg, 40.2 μ mol, 1 equiv) in anhydrous THF (5 mL) and diiodomethane (0.2 mL) at 0 °C under an argon atmosphere.⁴⁵ After 5 min, anhydrous methanol (50 μ L) was added. After 50 min, the reaction mixture was partitioned between aqueous hydrochloric acid solution (1 N, 25 mL) and ethyl acetate (80 mL). The aqueous layer was extracted with ethyl acetate (2 \times 20 mL), and the combined organic layers were washed sequentially with water (2 \times 30 mL) and saturated aqueous sodium chloride solution (30 mL), were dried over anhydrous sodium sulfate, were filtered, and were concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (eluent: gradient, 5 \rightarrow 20% ethyl acetate in dichloromethane) to afford dithioacetal **38** (10.8 mg, 45.6%) as a colorless oil. Structural assignments were made with additional information from gCOSY, HSQC, and gHMBC data. Based on ¹H NMR analysis at 20 °C in CDCl₃, the product exists as a 1:4 mixture of minor:major conformers.

¹H NMR (600 MHz, CDCl₃, 20 °C):

Major conformer: δ 7.89 (br-s, 1H, N₁H), 7.87 (d, J = 8.2, 1H, C₈H), 7.65 (d, J = 7.7, 2H, SO₂Ph-*o*-H), 7.43 (app-dd, J = 7.4, 7.5, 1H, SO₂Ph-*p*-H), 7.37 (app-ddd, J = 1.3, 7.4, 8.2, 1H, C₇H), 7.31 (d, J = 8.2, 1H, C₈H), 7.15 (dd, J = 7.7, 7.9, 2H, SO₂Ph-*m*-H), 7.14 (app-t, J = 7.3, 1H, C₇H), 7.10 (app-t, J = 7.5, 1H, C₆H), 7.02 (app-dd, J = 0.4, 7.4, 1H, C₅H), 6.81 (app-dd, J = 7.4, 7.7, 1H, C₆H), 6.57 (s, 1H, C₂H), 6.51 (d, J = 8.0, 1H, C₅H), 6.43 (d, J = 2.4, 1H, C₂H), 4.86 (s, 1H, C₁₅H), 4.55 (d, J = 14.8, 1H, C₁₉H_a), 3.86 (d, J = 14.9, 1H, C₁₂H_a), 3.71 (d, J = 14.8, 1H, C₁₉H_b), 3.08 (s, 3H, C₁₇H₃), 2.70 (d, J = 14.9, 1H, C₁₂H_b).

Minor conformer: δ 7.78 (d, J = 8.3, 1H, C₈H), 7.78 (br-s, 1H, N₁H), 7.44–7.40 (m, 1H, C₇H), 7.35 (d, J = 7.7, 2H, SO₂Ph-*o*-H), 7.32–7.29 (m, 1H, C₇H), 7.28 (d, J = 8.3, 1H, C₅H), 7.27–7.21 (m, C₈H SO₂Ph-*p*-H), 7.20 (d, J = 8.0, 1H, C₅H), 7.12–7.07 (m, 2H, C₆H + C₆H), 6.93 (dd, J = 7.7, 8.0, 2H, SO₂Ph-*m*-H), 6.57 (s, 1H, C₂H), 5.97 (d, J = 2.5, 1H, C₂H), 5.26 (s, 1H, C₁₅H), 4.01 (d, J = 15.6, 1H, C₁₉H_a), 3.56 (d, J = 14.9, 1H, C₁₉H_b), 3.16 (s, 3H, C₁₇H₃), 3.11 (d, J = 15.8, 1H, C₁₂H_a), 2.72 (d, J = 15.8, 1H, C₁₂H_b).

⁴⁵ (a) Cook, K. M.; Hilton, S. T.; Mecinović, J.; Motherwell, W. B.; Figg, W. D.; Schofield, C. J. *J. Biol. Chem.* **2009**, *284*, 26831. (b) Poisel, H.; Schmidt, U. *Chem. Ber.* **1971**, *104*, 1714.

^{13}C NMR (150 MHz, CDCl_3 , 20 °C):

Major conformer: δ 167.5 (C_{13}), 161.7 (C_{16}), 140.5 (C_9), 137.3 (C_9), 136.6 ($\text{SO}_2\text{Ph-}i\text{pso-C}$), 136.1 (C_4), 133.4 ($\text{SO}_2\text{Ph-}p\text{-C}$), 129.5 (C_7), 128.8 ($\text{SO}_2\text{Ph-}m\text{-C}$), 128.0 ($\text{SO}_2\text{Ph-}o\text{-C}$), 125.7 (C_6), 124.6 (C_5), 124.4 (C_4), 124.1 (C_2), 122.9 (C_7), 120.4 (C_6), 119.0 (C_5), 117.8 (C_8), 113.9 (C_3), 111.8 (C_8), 85.4 (C_2), 70.6 (C_{11}), 65.2 (C_{15}), 54.4 (C_3), 48.1 (C_{12}), 32.7 (C_{17}), 31.7 (C_{19}).

Minor conformer: δ 165.3 (C_{13}), 160.5 (C_{16}), 140.8 (C_9), 137.6 (C_9), 137.2 ($\text{SO}_2\text{Ph-}i\text{pso-C}$), 136.6 (C_4), 133.0 ($\text{SO}_2\text{Ph-}p\text{-C}$), 129.9 (C_7), 128.4 ($\text{SO}_2\text{Ph-}m\text{-C}$), 127.3 ($\text{SO}_2\text{Ph-}o\text{-C}$), 126.1 (C_6), 124.8 (C_5), 124.4 (C_4), 124.2 (C_2), 123.2 (C_7), 120.8 (C_6), 119.1 (C_5), 118.8 (C_8), 114.2 (C_3), 111.9 (C_8), 84.8 (C_2), 74.5 (C_{11}), 68.5 (C_{15}), 55.7 (C_3), 42.6 (C_{12}), 32.7 (C_{17}), 32.3 (C_{19}).

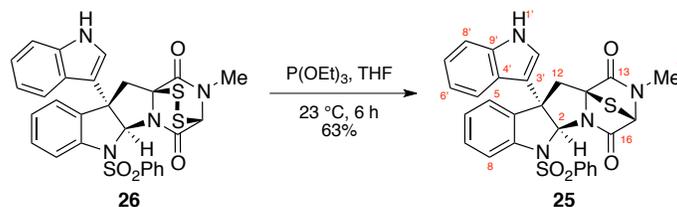
FTIR (thin film) cm^{-1} :

3392 (br-m), 3059 (w), 2977 (w), 1690 (s), 1451 (w), 1361 (m), 1266 (w), 1170 (m), 1090 (w), 1022 (m), 736 (m).

HRMS (ESI) (m/z):

calc'd for $\text{C}_{29}\text{H}_{24}\text{N}_4\text{NaO}_4\text{S}_3$ $[\text{M}+\text{Na}]^+$: 611.0852, found 611.0850.

TLC (10% ethyl acetate in dichloromethane), R_f : 0.40 (UV, I_2 , CAM).



C3-(Indol-3'-yl) epimonothiodiketopiperazine 25:⁴⁶

Triethylphosphite (10.0 μL , 58.4 μmol , 21.4 equiv) was added to the solution of epidithiodiketopiperazine **26** (8.6 mg, 15 μmol , 1 equiv) in anhydrous tetrahydrofuran (4 mL) at 23 °C. After 6 h, the reaction mixture was diluted sequentially with saturated aqueous ammonium chloride solution (20 mL) and ethyl acetate (60 mL). The organic layer was washed with water (2 \times 15 mL) and saturated aqueous sodium chloride solution (15 mL), was dried over anhydrous sodium sulfate, was filtered, and was concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (eluent: gradient, 2 \rightarrow 8% ethyl acetate in dichloromethane) to afford the epimonothiodiketopiperazine **25** (5.1 mg, 63%) as a white solid. Structural assignments were made with additional information from gCOSY, HSQC, and gHMBC data.⁴⁷

¹H NMR (600 MHz, acetone-*d*₆, 20 °C):

δ 9.95 (br-s, 1H, N₁H), 7.64 (d, J = 8.1, 1H, C₈H), 7.59 (d, J = 7.8, 1H, C₅H), 7.52 (d, J = 7.3, 1H, C₅H), 7.48 (d, J = 8.0, 1H, C₈H), 7.44 (app-dt, J = 1.1, 7.8, 1H, C₇H), 7.32 (app-tt, J = 0.9, 7.4, 1H, SO₂Ph-*p*-H), 7.28 (app-dt, J = 0.9, 7.6, 1H, C₆H), 7.26 (app-dt, J = 0.9, 7.6, 1H, C₇H), 7.20 (app-dt, J = 0.8, 7.5, 1H, C₆H), 6.98 (app-dd, J = 1.0, 8.3, 2H, SO₂Ph-*o*-H), 6.90 (app-t, J = 7.5, 2H, SO₂Ph-*m*-H), 6.19 (s, 1H, C₂H), 5.67 (d, J = 2.6, 1H, C₂H), 5.17 (s, 1H, C₁₅H), 3.70 (d, J = 15.4, 1H, C₁₂H_a), 3.11 (s, 3H, C₁₇H₃), 2.84 (d, J = 15.4, 1H, C₁₂H_b).

¹³C NMR (150 MHz, acetone-*d*₆, 20 °C):

δ 173.9 (C₁₃), 171.6 (C₁₆), 141.4 (C₉), 138.7 (SO₂Ph-*ipso*-C), 138.7 (C₉), 137.9 (C₄), 133.9 (SO₂Ph-*p*-C), 130.2 (C₇), 129.0 (SO₂Ph-*m*-C), 127.2 (SO₂Ph-*o*-C), 126.4 (C₆), 125.7 (C₅), 125.3 (C₂), 124.9 (C₄), 123.0 (C₇), 120.6 (C₆), 119.0 (C₅), 118.6 (C₈), 115.3 (C₃), 113.1 (C₈), 83.3 (C₂), 81.9 (C₁₁), 73.0 (C₁₅), 59.4 (C₃), 35.3 (C₁₂), 31.4 (C₁₇).

FTIR (thin film) cm⁻¹:

3357 (br-w), 3059 (w), 2919 (w), 2851 (w), 1740 (s), 1713 (s), 1457 (m), 1358 (m), 1261 (w), 1169 (m), 1086 (w), 971 (w), 737 (s), 685 (m).

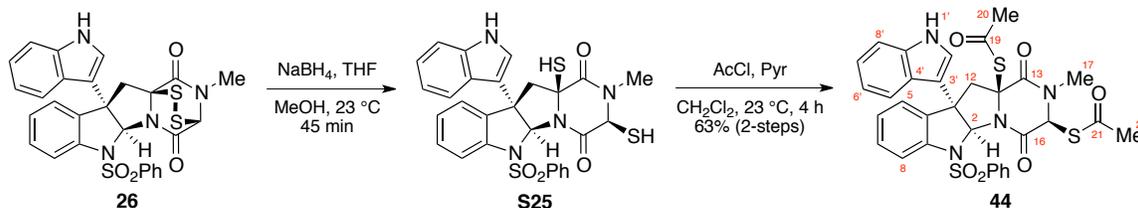
HRMS (ESI) (m/z):

calc'd for C₂₈H₂₂N₄NaO₄S₂ [M+Na]⁺: 565.0975, found 565.0971.

TLC (10% ethyl acetate in dichloromethane), R_f: 0.76 (UV, I₂, CAM).

⁴⁶ Cherblanc, F.; Lo, Y.-P.; De Gussem, E.; Alcazar-Fuoli, L.; Bignell, E.; He, Y.; Chapman-Rothe, N.; Bultinck, P.; Herrebout, W. A.; Brown, R.; Rzepa, H. S.; Fuchter, M. J. *Chem. – Eur. J.* **2011**, *17*, 11868.

⁴⁷ Limited solubility of epimonothiodiketopiperazine **25** was observed in CH₂Cl₂, CHCl₃, EtOAc, MeOH, DMSO; this low solubility resulted in difficulty to acquire high quality spectroscopic data.



C3-(Indol-3'-yl) bithioacetate 44:

Sodium borohydride (4.9 mg, 0.13 mmol, 3.3 equiv) was added as a solid to a solution of epidithiodiketopiperazine **26** (22.6 mg, 39.3 μ mol, 1 equiv) in anhydrous tetrahydrofuran (5 mL) and anhydrous methanol (50 μ L) at 23 °C. After 45 min, the reaction mixture was diluted with ethyl acetate (40 mL) and washed with saturated aqueous ammonium chloride solution (20 mL). The aqueous layer was extracted with ethyl acetate (2 \times 20 mL), and the combined organic layers were washed sequentially with water (2 \times 20 mL) and saturated aqueous sodium chloride solution (20 mL), were dried over anhydrous sodium sulfate, were filtered, and were concentrated under reduced pressure to afford the hexacyclic bithiol **S25** that was used in the next step without further purification.

Acetyl chloride (200 μ L, 2.80 mmol, 71.3 equiv) was added to the solution of bithiol **S25** in anhydrous dichloromethane (6 mL) and anhydrous pyridine (300 μ L, 3.72 mmol, 94.7 equiv) at 23 °C. After 4 h, the reaction mixture was diluted with ethyl acetate (60 mL) and washed with aqueous 5% citric acid solution (2 \times 20 mL). The aqueous layer was extracted with ethyl acetate (2 \times 20 mL), and the combined organic layers were washed sequentially with water (2 \times 20 mL), saturated aqueous sodium hydrogenocarbonate solution (20 mL), water (20 mL), and saturated aqueous sodium chloride solution (20 mL), were dried over anhydrous sodium sulfate, were filtered, and were concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (eluent: gradient, 5 \rightarrow 20% ethyl acetate in dichloromethane) to afford bithioacetate **44** (17.0 mg, 62.7%) as a pale yellow oil. Structural assignments were made with additional information from gCOSY, HSQC, and gHMBC data.

^1H NMR (600 MHz, CDCl_3 , 20 °C):

δ 8.01 (br-s, 1H, N_1H), 7.78 (d, $J = 8.1$, 1H, C_8H), 7.75 (d, $J = 8.2$, 2H, $\text{SO}_2\text{Ph-}o\text{-H}$), 7.42 (app-dt, $J = 1.0, 7.5$, 1H, $\text{SO}_2\text{Ph-}p\text{-H}$), 7.37–7.33 (m, 1H, C_7H), 7.29 (d, $J = 8.2$, 1H, C_8H), 7.21 (dd, $J = 7.6, 8.3$, 2H, $\text{SO}_2\text{Ph-}m\text{-H}$), 7.12 (dd, $J = 7.4, 8.0$, 1H, C_7H), 7.08–7.03 (m, 2H, $\text{C}_5\text{H} + \text{C}_6\text{H}$), 6.81 (dd, $J = 7.2, 7.9$, 1H, C_6H), 6.72 (s, 1H, C_2H), 6.58 (d, $J = 8.0$, 1H, C_5H), 6.55 (d, $J = 2.5$, 1H, C_2H), 6.09 (s, 1H, C_{15}H), 3.44 (d, $J = 14.7$, 1H, C_{12}H_a), 3.26 (d, $J = 14.7$, 1H, C_{12}H_b), 2.98 (s, 3H, C_{17}H_3), 2.48 (s, 3H, C_{22}H_3), 2.06 (s, 3H, C_{20}H_3).

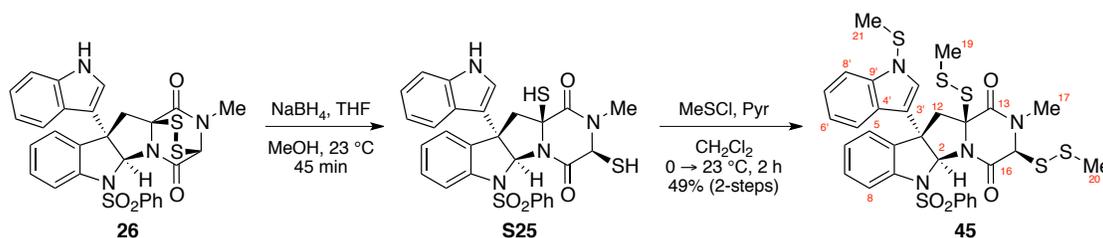
^{13}C NMR (150 MHz, CDCl_3 , 20 °C):

δ 194.0 (C_{21}), 193.9 (C_{19}), 165.1 (C_{13}), 161.9 (C_{16}), 142.0 (C_9), 137.7 ($\text{SO}_2\text{Ph-}ipso\text{-C}$), 137.3 (C_9), 135.2 (C_4), 133.3 ($\text{SO}_2\text{Ph-}p\text{-C}$), 129.6 (C_7), 129.0 ($\text{SO}_2\text{Ph-}m\text{-C}$), 127.5 ($\text{SO}_2\text{Ph-}o\text{-C}$), 125.1 (C_6), 124.9 (C_5), 124.4 (C_4), 122.9 (C_2), 122.8 (C_7), 120.4 (C_6), 118.7 (C_5), 116.6 (C_8), 115.7 (C_3), 111.9 (C_8), 84.8 (C_2), 73.3 (C_{11}), 63.5 (C_{15}), 53.6 (C_3), 49.3 (C_{12}), 32.3 (C_{17}), 30.6 (C_{20}), 30.5 (C_{22}).

FTIR (thin film) cm^{-1} : 3395 (br-m), 3063 (w), 2923 (m), 2852 (w), 1699 (br-s), 1459 (m), 1368 (m), 1311 (w), 1172 (m), 1121 (m), 1093 (m), 1025 (w), 954 (w), 734 (m).

HRMS (ESI) (m/z): calc'd for $\text{C}_{32}\text{H}_{28}\text{N}_4\text{NaO}_6\text{S}_3$ $[\text{M}+\text{Na}]^+$: 683.1063, found 683.1047.

TLC (20% ethyl acetate in dichloromethane), R_f : 0.52 (UV, I_2 , CAM).



C3-(Indol-3'-yl) *N*-(thiomethyl) bis(methylthio) 45:⁴⁸

Sodium borohydride (3.7 mg, 0.10 mmol, 5.2 equiv) was added as a solid to a solution of epidithiodiketopiperazine **26** (10.8 mg, 18.8 μ mol, 1 equiv) in anhydrous tetrahydrofuran (5 mL) and anhydrous methanol (50 μ L) at 23 °C. After 45 min, the reaction mixture was diluted with ethyl acetate (40 mL) and washed with saturated aqueous ammonium chloride solution (20 mL). The aqueous layer was extracted with ethyl acetate (2 \times 20 mL), and the combined organic layers were washed sequentially with water (2 \times 20 mL) and saturated aqueous sodium chloride solution (20 mL), were dried over anhydrous sodium sulfate, were filtered, and were concentrated under reduced pressure to afford the hexacyclic bithiol **S25** that was used in the next step without further purification.

A solution of methanesulfinyl chloride⁴⁹ in dichloromethane (1.6 M, 250 μ L, 402 μ mol, 21.4 equiv) was added to the solution of bithiol **S25** in anhydrous dichloromethane (5 mL) and anhydrous pyridine (100 μ L, 1.24 mmol, 66.0 equiv) at 0 °C. After 10 min, the ice–water bath was removed, and the yellow solution was allowed to warm to 23 °C. After 2 h, the reaction mixture was diluted sequentially with saturated aqueous ammonium chloride solution (20 mL) and ethyl acetate (60 mL). The organic layer was sequentially washed with saturated aqueous ammonium chloride solution (20 mL), water (2 \times 15 mL), and saturated aqueous sodium chloride solution (15 mL). The aqueous layer was extracted with ethyl acetate (2 \times 20 mL). The combined organic layers were dried over anhydrous sodium sulfate, were filtered, and were concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (eluent: gradient, 5 \rightarrow 30% ethyl acetate in hexanes) to afford the *N*-thiomethyl bis(methylthio) **45** (6.5 mg, 49%) as a colorless oil. Structural assignments were made with additional information from gCOSY, HSQC, and gHMBC data.

¹H NMR (600 MHz, CDCl₃, 20 °C):

δ 7.74 (d, J = 8.1, 1H, C₈H), 7.57 (d, J = 8.2, 1H, C₈H), 7.42–7.37 (m, 1H, C₇H), 7.39 (d, J = 8.2, 2H, SO₂Ph-*o*-H), 7.35 (app-dt, J = 1.0, 7.4, 1H, SO₂Ph-*p*-H), 7.31 (app-dt, J = 0.8, 7.7, 1H, C₇H), 7.27 (app-dt, J = 1.0, 7.6, 1H, C₃H), 7.21 (app-dt, J = 0.8, 7.5, 1H, C₆H), 7.09 (app-dt, J = 0.7, 7.5, 1H, C₆H), 7.01 (d, J = 7.9, 1H, C₅H), 6.97 (dd, J = 7.6, 8.2, 2H, SO₂Ph-*m*-H), 6.71 (s, 1H, C₂H), 6.08 (s, 1H, C₂H), 5.02 (s, 1H, C₁₅H), 3.29 (d, J = 15.0, 1H, C₁₂H_a), 3.25 (d, J = 15.0, 1H, C₁₂H_b), 3.17 (s, 3H, C₁₇H₃), 2.67 (s, 3H, C₂₀H₃), 2.50 (s, 3H, C₂₁H₃), 2.29 (s, 3H, C₁₉H₃).

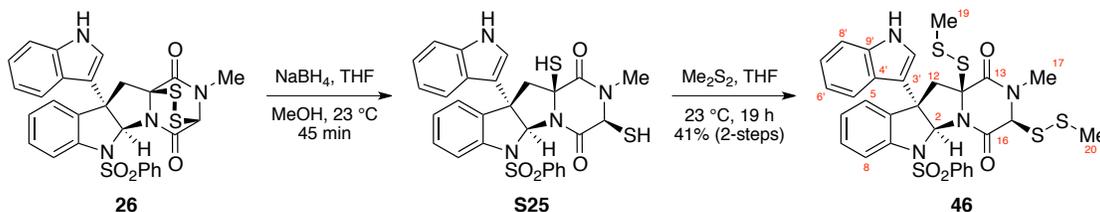
¹³C NMR (150 MHz, CDCl₃, 20 °C):

δ 165.3 (C₁₃), 162.5 (C₁₆), 142.0 (C₉), 141.2 (C₉), 137.8 (SO₂Ph-*ipso*-C), 136.0 (C₄), 133.0 (C₂), 132.9

⁴⁸ (a) Gilow, H. M.; Brown, C. S.; Copeland, J. N.; Kelly, K. E. *J. Heterocyclic Chem.* **1991**, *28*, 1025. (b) Kim, J. K.; Caserio, M. C. *J. Org. Chem.* **1979**, *44*, 1897. (c) Kharasch, N.; Parker, A. J. *J. Org. Chem.* **1959**, *24*, 1029.

⁴⁹ Douglass, I. B.; Norton, R. V.; Farah, B. S. *Org. Synth.* **1960**, *40*, 62.

	(SO ₂ Ph- <i>p</i> -C), 129.7 (C ₇), 128.4 (SO ₂ Ph- <i>m</i> -C), 127.3 (SO ₂ Ph- <i>o</i> -C), 125.9 (C ₆), 125.8 (C ₄), 124.6 (C ₅), 123.7 (C ₇), 121.6 (C ₆), 119.1 (C ₅), 118.8 (C ₈), 117.6 (C ₃), 111.7 (C ₈), 84.8 (C ₂), 79.2 (C ₁₅), 73.9 (C ₁₁), 53.5 (C ₃), 46.0 (C ₁₂), 32.7 (C ₁₇), 24.4 (C ₂₀), 24.0 (C ₂₁), 23.3 (C ₁₉).
FTIR (thin film) cm ⁻¹ :	2925 (w), 1699 (s), 1458 (m), 1359 (m), 1231 (w), 1168 (m), 1091 (w), 749 (m).
HRMS (ESI) (<i>m/z</i>):	calc'd for C ₃₁ H ₃₀ N ₄ NaO ₄ S ₆ [M+Na] ⁺ : 737.0484, found 737.0469.
TLC (50% ethyl acetate in hexanes), R _f :	0.60 (UV, I ₂ , CAM).



C3-(Indol-3'-yl) bis(methyl disulfane) 46:

Sodium borohydride (4.8 mg, 0.13 mmol, 3.7 equiv) was added as a solid to a solution of epidthiodiketopiperazine **26** (19.5 mg, 33.9 μmol , 1 equiv) in anhydrous tetrahydrofuran (5 mL) and anhydrous methanol (50 μL) at 23 °C. After 45 min, the reaction mixture was diluted with ethyl acetate (40 mL) and washed with saturated aqueous ammonium chloride solution (20 mL). The aqueous layer was extracted with ethyl acetate (2 \times 20 mL), and the combined organic layers were washed sequentially with water (2 \times 20 mL) and saturated aqueous sodium chloride solution (20 mL), were dried over anhydrous sodium sulfate, were filtered, and were concentrated under reduced pressure to afford the hexacyclic bithiol **S25** that was used in the next step without further purification.

Dimethyl disulfide⁵⁰ (200 μL , 2.23 mmol, 65.7 equiv) was added to the solution of bithiol **S25** in anhydrous tetrahydrofuran (6 mL) at 23 °C. After 19 h, the volatiles were removed under reduced pressure. The residue was purified by flash column chromatography on silica gel (eluent: gradient, 5 \rightarrow 10% ethyl acetate in dichloromethane) to afford the bis(methyl disulfane) **46** (9.3 mg, 41%) as a colorless oil. Structural assignments were made with additional information from gCOSY, HSQC, and gHMBC data.

¹H NMR (600 MHz, CDCl₃, 20 °C):

δ 7.85 (br-s, 1H, N₁H), 7.70 (d, $J = 8.1$, 1H, C₈H), 7.52 (d, $J = 7.4$, 2H, SO₂Ph-*o*-H), 7.36 (app-dt, $J = 1.3, 7.8$, 1H, C₇H), 7.33 (t, $J = 7.5$, 1H, SO₂Ph-*p*-H), 7.32 (d, $J = 8.2$, 1H, C₈H), 7.24 (app-dd, $J = 0.8, 7.5$, 1H, C₃H), 7.19 (app-ddd, $J = 2.4, 5.8, 8.2$, 1H, C₇H), 7.16 (app-dt, $J = 0.8, 7.5$, 1H, C₆H), 7.07 (app-dt, $J = 0.5, 7.9$, 2H, SO₂Ph-*m*-H), 7.01–6.96 (m, 2H, C₅H + C₆H), 6.76 (s, 1H, C₂H), 6.28 (d, $J = 2.6$, 1H, C₂H), 5.00 (s, 1H, C₁₅H), 3.38 (d, $J = 15.0$, 1H, C₁₂H_a), 3.26 (d, $J = 15.0$, 1H, C₁₂H_b), 3.17 (s, 3H, C₁₇H₃), 2.64 (s, 3H, C₂₀H₃), 2.29 (s, 3H, C₁₉H₃).

¹³C NMR (150 MHz, CDCl₃, 20 °C):

δ 165.4 (C₁₃), 162.6 (C₁₆), 141.8 (C₉), 138.2 (SO₂Ph-*ipso*-C), 137.3 (C₉), 136.4 (C₄), 133.0 (SO₂Ph-*p*-C), 129.5 (C₇), 128.6 (SO₂Ph-*m*-C), 127.4 (SO₂Ph-*o*-C), 125.7 (C₆), 124.6 (C₅), 124.2 (C₄), 123.3 (C₂), 122.9 (C₇), 120.5 (C₆), 118.9 (C₅), 118.3 (C₈), 115.7 (C₃), 111.9 (C₈), 85.2 (C₂), 79.2 (C₁₅), 74.0 (C₁₁), 53.6 (C₃), 46.4 (C₁₂), 32.7 (C₁₇), 24.4 (C₂₀), 23.4 (C₁₉).

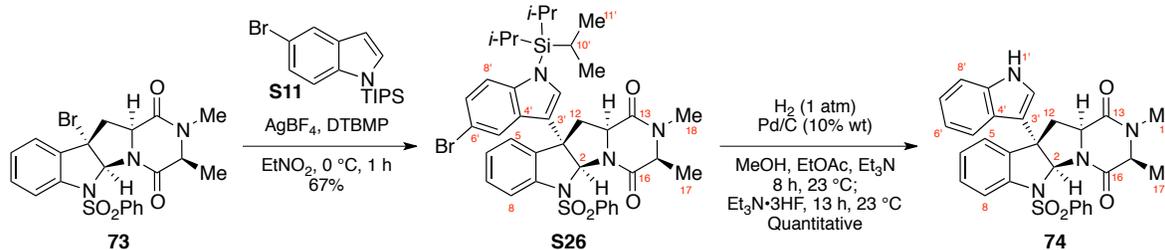
FTIR (thin film) cm⁻¹:

3392 (br-m), 3060 (w), 2921 (w), 1685 (s), 1459 (m), 1391 (m), 1266 (w), 1169 (m), 1092 (w), 1022 (w), 736 (m).

⁵⁰ Dubs, P.; Stuessi, R. *Helv. Chim. Acta* **1976**, *59*, 1307.

HRMS (ESI) (m/z): calc'd for $C_{30}H_{28}N_4NaO_4S_5$ $[M+Na]^+$: 691.0606,
found 691.0613.

TLC (20% ethyl acetate in dichloromethane), R_f : 0.67 (UV, I_2 , CAM).



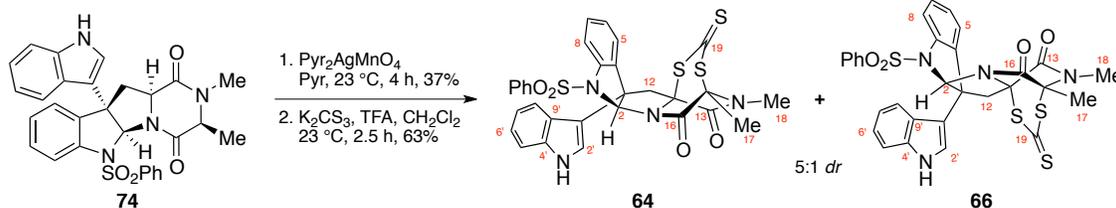
C3-(Indol-3'-yl)-pyrrolidinoindoline 74:

This compound was prepared in two steps starting from *endo*-tetracyclic bromide⁷ (+)-**73** (512.5 mg, 10.5 mmol, 1 equiv) using the methodology developed to access the corresponding C3-(5-bromo-*N*-TIPS-indol-3'-yl)-pyrrolidinoindoline (+)-**S12** (Please see page S10 for details) with DTBMP (339 mg, 1.65 mmol, 1.58 equiv), 5-bromo-1-triisopropylsilyl-1*H*-indole¹² **S11** (1.92 g, 5.45 mmol, 5.20 equiv), and silver(I) tetrafluoroborate (600 mg, 3.08 mmol, 2.95 equiv) in anhydrous nitroethane (12 mL). After 1 h, saturated aqueous sodium chloride solution (20 mL) was introduced, and the resulting biphasic mixture was vigorously stirred for 30 min at 0 °C. The reaction mixture was diluted with ethyl acetate (50 mL), was filtered through a Celite pad, and the solid was washed with ethyl acetate (3 × 15 mL). The combined filtrates were washed with 5% aqueous citric acid solution (2 × 25 mL), water (3 × 25 mL), and saturated aqueous sodium chloride solution (25 mL). The organic layer was dried over anhydrous sodium sulfate, was filtered, and was concentrated under reduced pressure. The resulting residue was purified by flash column chromatography (eluent: gradient, 1 → 10% acetone in dichloromethane) to afford the C3-(5-bromo-*N*-TIPS-indol-3'-yl)-pyrrolidinoindoline **S26** (537 mg, 67.4%) as a white foam.⁵¹

The free indole was accessed in a one-pot two-step procedure using the methodology developed to access the corresponding C3-(indol-3'-yl)-pyrrolidinoindoline (+)-**59** (Please see page S12 for details). The reaction mixture was filtered through a pad of Celite. The solids were washed with ethyl acetate (3 × 50 mL). The combined filtrates were concentrated under reduced pressure. The resulting pale yellow solid was diluted in ethyl acetate (150 mL) and washed sequentially with an aqueous hydrochloric acid solution (1 N, 2 × 50 mL), water (2 × 50 mL), and saturated aqueous sodium chloride solution (40 mL). The organic layer was dried over anhydrous sodium sulfate, was filtered, and was concentrated under reduced pressure to afford the C3-(indol-3'-yl)-pyrrolidinoindoline **74**⁵² (370 mg, 99.7%) as a white solid that was used in the next step without further purification.

⁵¹ **S26**: ¹H NMR (600 MHz, CDCl₃, 20 °C): δ 8.03 (d, *J* = 7.8, 2H, SO₂Ph-*o*-H), 7.78 (d, *J* = 8.4, 1H, C₈H), 7.54 (app-dd, *J* = 7.2, 7.8, 1H, SO₂Ph-*p*-H), 7.40 (app-t, *J* = 7.8, 2H, SO₂Ph-*m*-H), 7.30 (d, *J* = 8.9, 1H, C₈H), 7.28 (app-dt, *J* = 1.0, 7.9, 1H, C₇H), 7.15 (app-dd, *J* = 1.7, 8.8, 1H, C₇H), 6.97 (dd, *J* = 7.5, 7.6, 1H, C₆H), 6.95 (s, 1H, C₂H), 6.82 (d, *J* = 7.3, 1H, C₅H), 6.50 (br-s, 1H, C₃H), 6.30 (s, 1H, C₂H), 4.44 (dd, *J* = 7.8, 9.4, 1H, C₁₁H), 3.97 (q, *J* = 7.1, 1H, C₁₅H), 3.03 (dd, *J* = 7.5, 13.8, 1H, C₁₂H_a), 2.99 (s, 3H, C₁₈H₃), 2.88 (dd, *J* = 9.8, 13.8, 1H, C₁₂H_b), 1.66 (d, *J* = 7.1, 3H, C₁₇H), 1.59 (app-sp, *J* = 7.5, 3H, C₁₀H), 1.07 (app-d, *J* = 5.5, 18H, C₁₁H). ¹³C NMR (150 MHz, CDCl₃, 20 °C): δ 169.5 (C₁₃), 169.2 (C₁₆), 141.3 (C₉), 139.6 (C₉), 137.1 (SO₂Ph-*ipso*-C), 134.2 (SO₂Ph-*p*-C), 133.9 (C₄), 130.9 (C₂), 130.3 (C₄), 129.5 (C₇), 129.2 (SO₂Ph-*m*-C), 127.9 (SO₂Ph-*o*-C), 125.3 (C₇), 124.5 (C₆), 123.9 (C₅), 121.9 (C₅), 116.0 (C₈), 115.6 (C₈), 115.1 (C₃), 113.5 (C₆), 83.0 (C₂), 59.5 (C₁₁), 57.5 (C₁₅), 55.3 (C₃), 37.8 (C₁₂), 29.6 (C₁₈), 18.2 (C₁₁), 14.8 (C₁₁), 12.9 (C₁₀). TLC (20% acetone in dichloromethane), R_f: 0.76 (UV, CAM).

⁵² **74**: ¹H NMR (600 MHz, CDCl₃, 20 °C): δ 8.94 (br-s, 1H, N₁H), 7.74 (d, *J* = 8.2, 1H, C₈H), 7.46 (d, *J* = 8.2, 2H, SO₂Ph-*o*-H), 7.35 (app-dt, *J* = 0.9, 7.5, 1H, SO₂Ph-*p*-H), 7.34 (d, *J* = 8.3, 1H, C₈H), 7.29 (dd, *J* = 7.5, 8.1, 1H, C₇H), 7.19 (app-dt, *J* = 4.1, 8.2, 1H, C₇H), 7.12 (d, *J* = 7.5, 1H, C₃H), 7.09–7.04 (m, 3H, SO₂Ph-*m*-H + C₆H), 6.95 (app-d, *J* = 4.0, 2H, C₃H + C₆H), 6.40 (s, 1H, C₂H), 6.09 (d, *J* = 2.0, 1H, C₂H), 4.52 (app-t, *J* = 7.8, 1H, C₁₁H), 4.07 (q, *J* = 7.0, 1H, C₁₅H), 3.10 (app-d, *J* = 7.8, 2H, C₁₂H), 2.90 (s, 3H, C₁₈H₃), 1.61 (d, *J* = 7.1, 3H, C₁₇H₃). MS (ESI) (*m/z*): [M+H]⁺: 527.25; [M+Na]⁺: 549.21. TLC (20% acetone in dichloromethane), R_f: 0.27 (UV, CAM).



C3-(Indol-3'-yl) dithiepanethiones 64 and 66:

Freshly prepared bis(pyridine)silver(I) permanganate⁵³ (800 mg, 2.08 mmol, 5.45 equiv) was added as a solid to a solution of indole adduct **74** (201 mg, 382 μ mol, 1 equiv) in anhydrous pyridine (5 mL) at 23 °C. After 2 h, a second portion of bis(pyridine)silver(I) permanganate (600 mg, 1.56 mmol, 4.08 equiv) was added. After 2 h, the resulting thick brown suspension was diluted with saturated aqueous sodium sulfite solution (50 mL) and then with ethyl acetate (160 mL). The resulting mixture was washed sequentially with water (2 \times 50 mL), aqueous 5% copper sulfate solution (3 \times 50 mL), and saturated aqueous sodium chloride solution (30 mL). The combined aqueous layers were extracted with ethyl acetate (2 \times 100 mL). The combined organic layers were dried over anhydrous sodium sulfate, were filtered, and were concentrated under reduced pressure. The resulting yellow residue was purified by flash column chromatography (eluent: gradient, 2 \rightarrow 20% isopropanol in dichloromethane and hexanes (50%)) to afford the corresponding diols⁵⁴ (78.0 mg, 36.5%) as a yellow oil.

To a yellow solution of potassium trithiocarbonate⁵⁵ (250 mg, 1.34 mmol, 9.63 equiv) in anhydrous dichloromethane (6 mL) and trifluoroacetic acid (4 mL) at 23 °C was added a solution of the diol (78.0 mg, 139 μ mol, 1 equiv) in dichloromethane (1 mL). After 2.5 h, the reaction mixture was diluted with ethyl acetate (60 mL) and washed with saturated aqueous sodium bicarbonate (30 mL). The aqueous layer was extracted with ethyl acetate (2 \times 20 mL) and the combined organic layers were dried over anhydrous sodium sulfate, were filtered, and were concentrated under reduced pressure. The resulting orange residue was purified by flash column chromatography on silica gel (eluent: gradient, 2 \rightarrow 8% ethyl acetate in dichloromethane) to afford an inseparable mixture of isomeric monomeric dithiepanethiones **64** and **66** (55.7 mg, 63.3%, **64**:**66**, 5:1) as a pale yellow solid.

Isomers **64** and **66** were separated for the purpose of full and independent characterization by preparative HPLC [Waters X-Bridge preparative HPLC column, C18, 5 μ m, 19 \times 250 mm; 20.0 mL/min; gradient, 30 \rightarrow 100% acetonitrile in water, 35 min; t_R (**64**) = 21.3 min, t_R (**66**) = 23.4 min]. Structural assignments were made using additional information from gCOSY, HSQC, and HMBC experiments.

β -epimer 64:

¹H NMR (600 MHz, CDCl₃, 20 °C):

δ 7.81 (br-s, 1H, N₁H), 7.74 (d, J = 8.2, 1H, C₈H), 7.44–7.39 (m, 1H, C₇H), 7.37 (app-dd, J = 0.7, 7.5, 2H, SO₂Ph-*o*-H), 7.35 (d, J = 8.1, 1H, C₈H), 7.34 (t, J = 7.5, 1H, SO₂Ph-*p*-H), 7.26–7.21 (m, 3H, C₅H + C₆H + C₇H), 7.08–7.04 (m, 2H, C₅H + C₆H), 7.02 (app-t, J = 7.7, 2H, SO₂Ph-*m*-H), 6.74 (s, 1H, C₂H), 6.20 (d, J = 2.5, 1H, C₂H), 3.91 (d, J = 15.6, 1H,

⁵³ Firouzabadi, H.; Vessal, B.; Naderi, M. *Tetrahedron Lett.* **1982**, *23*, 1847.

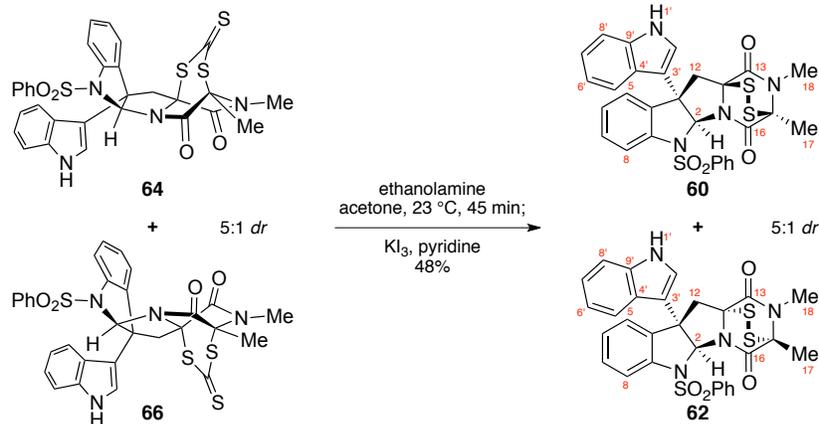
⁵⁴ The product was isolated as a mixture of isomers.

⁵⁵ Stueber, D.; Patterson, D.; Mayne, C. L.; Orendt, A. M.; Grant, D. M.; Parry, R. W. *Inorg. Chem.* **2001**, *40*, 1902.

	$C_{12}\mathbf{H}_a$), 3.13 (s, 3H, $C_{18}\mathbf{H}_3$), 2.86 (d, $J = 15.6$, 1H, $C_{12}\mathbf{H}_b$), 2.01 (s, 3H, $C_{17}\mathbf{H}_3$).
^{13}C NMR (100 MHz, CDCl_3 , 20 °C):	δ 215.9 (C_{19}), 165.0 (C_{13}), 161.0 (C_{16}), 141.1 (C_9), 137.8 ($\text{SO}_2\text{Ph-}ipso\text{-C}$), 137.3 (C_9), 135.4 (C_4), 133.1 ($\text{SO}_2\text{Ph-}p\text{-C}$), 130.2 (C_7), 128.5 ($\text{SO}_2\text{Ph-}m\text{-C}$), 127.3 ($\text{SO}_2\text{Ph-}o\text{-C}$), 126.3 (C_6), 124.7 (C_5), 124.1 (C_4), 124.0 (C_2), 123.2 (C_7), 120.8 (C_6), 119.0 (C_8), 118.8 (C_5), 114.1 (C_3), 112.0 (C_8), 85.6 (C_2), 75.1 (C_{11}), 73.5 (C_{15}), 54.1 (C_3), 46.4 (C_{12}), 28.7 (C_{18}), 20.2 (C_{17}).
FTIR (thin film) cm^{-1} :	3397 (br-m), 3061 (w), 1688 (s), 1459 (w), 1361 (s), 1241 (w), 1170 (s), 1108 (w), 1001 (m), 908 (w), 734 (m).
HRMS (ESI) (m/z):	calc'd for $\text{C}_{30}\text{H}_{25}\text{N}_4\text{O}_4\text{S}_4$ $[\text{M}+\text{H}]^+$: 633.0753, found 633.0744.
TLC (50% ethyl acetate in hexanes), R_f :	0.33 (UV, CAM).

α -epimer 66:

^1H NMR (600 MHz, CDCl_3 , 20 °C):	δ 7.80 (app-dd, $J = 1.6, 6.8$, 1H, $\text{C}_5\mathbf{H}$), 7.72 (d, $J = 8.0$, 1H, $\text{C}_8\mathbf{H}$), 7.54 (br-s, 1H, $\text{N}_1\mathbf{H}$), 7.40–7.34 (m, 3H, $\text{C}_5\mathbf{H} + \text{C}_7\mathbf{H} + \text{C}_8\mathbf{H}$), 7.34–7.29 (m, 2H, $\text{C}_6\mathbf{H} + \text{C}_7\mathbf{H}$), 7.22 (app-t, $J = 7.5$, 1H, $\text{C}_6\mathbf{H}$), 7.20 (t, $J = 7.4$, 1H, $\text{SO}_2\text{Ph-}p\text{-H}$), 7.06 (app-dd, $J = 0.9, 8.3$, 2H, $\text{SO}_2\text{Ph-}o\text{-H}$), 6.81 (dd, $J = 7.6, 8.1$, 2H, $\text{SO}_2\text{Ph-}m\text{-H}$), 6.79 (s, 1H, $\text{C}_2\mathbf{H}$), 5.55 (d, $J = 2.5$, 1H, $\text{C}_2\mathbf{H}$), 4.04 (d, $J = 15.6$, 1H, $\text{C}_{12}\mathbf{H}_a$), 3.11 (d, $J = 15.6$, 1H, $\text{C}_{12}\mathbf{H}_b$), 2.98 (s, 3H, $\text{C}_{18}\mathbf{H}_3$), 2.00 (s, 3H, $\text{C}_{17}\mathbf{H}_3$).
^{13}C NMR (100 MHz, CDCl_3 , 20 °C):	δ 209.5 (C_{19}), 164.5 (C_{13}), 161.1 (C_{16}), 139.3 (C_9), 138.3 ($\text{SO}_2\text{Ph-}ipso\text{-C}$), 137.3 (C_9), 135.8 (C_4), 132.7 ($\text{SO}_2\text{Ph-}p\text{-C}$), 130.0 (C_7), 128.1 ($\text{SO}_2\text{Ph-}m\text{-C}$), 127.0 ($\text{SO}_2\text{Ph-}o\text{-C}$), 125.9 (C_6), 125.4 (C_5), 124.9 (C_2), 123.7 (C_4), 123.5 (C_7), 121.3 (C_6), 119.1 (C_5), 118.2 (C_8), 114.4 (C_3), 112.0 (C_8), 85.4 (C_2), 74.9 (C_{11}), 73.6 (C_{15}), 54.9 (C_3), 42.0 (C_{12}), 28.8 (C_{18}), 21.3 (C_{17}).
FTIR (thin film) cm^{-1} :	3396 (br-m), 2924 (w), 1698 (s), 1458 (m), 1364 (m), 1334 (m), 1251 (w), 1169 (m), 1091 (m), 1013 (m), 912 (w), 734 (m).
HRMS (ESI) (m/z):	calc'd for $\text{C}_{30}\text{H}_{25}\text{N}_4\text{O}_4\text{S}_4$ $[\text{M}+\text{H}]^+$: 633.0753, found 633.0767.
TLC (50% ethyl acetate in hexanes), R_f :	0.33 (UV, CAM).



C3-(Indol-3'-yl) epidithiodiketopiperazines 60 and 62:

Ethanolamine (4 mL) was added via syringe to a solution of the bisdithiepanethiones **64** and **66** (33.0 mg, 52.1 μmol , 1equiv, **64:66**, 5:1) in acetone (6 mL) at 23 °C. After 45 min, the reaction mixture was partitioned between ethyl acetate (100 mL) and aqueous hydrochloric acid solution (1 N, 30 mL). The organic layer was collected, and the aqueous layer was extracted with ethyl acetate (2 \times 15 mL). A solution of potassium triiodide in pyridine (2.5% w/v) was added dropwise to the combined organic layers until a persistent yellow color was observed. The resulting mixture was washed with aqueous hydrochloric acid (1 N, 20 mL), and saturated aqueous sodium chloride solution (20 mL). The organic layer was dried over anhydrous sodium sulfate, was filtered, and was concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (eluent: gradient, 20 \rightarrow 50% ethyl acetate in hexanes) to afford an inseparable mixture of isomeric monomeric epidithiodiketopiperazines **60** and **62** (14.8 mg, 48.2%, **60:62**, 5:1) as a pale yellow solid.

Isomers **60** and **62** were separated for the purpose of full and independent characterization by preparative HPLC [Waters X-Bridge preparative HPLC column, C18, 5 μm , 19 \times 250 mm; 20.0 mL/min; gradient, 30 \rightarrow 100% acetonitrile in water, 35 min; $t_{\text{R}}(\mathbf{60}) = 18.0$ min, $t_{\text{R}}(\mathbf{62}) = 19.7$ min]. Structural assignments were made using additional information from gCOSY, HSQC, and HMBC experiments.

β -epimer 60:

^1H NMR (600 MHz, CDCl_3 , 20 °C):

δ 8.01 (d, $J = 7.3$, 1H, C_5H), 7.73 (d, $J = 8.0$, 1H, C_8H), 7.60 (br-s, 1H, N_1H), 7.41 (d, $J = 6.6$, 1H, C_5H), 7.39 (d, $J = 7.8$, 1H, C_8H), 7.43–7.38 (m, 1H, C_7H), 7.38–7.31 (m, 2H, $\text{C}_6\text{H} + \text{C}_7\text{H}$), 7.26–7.21 (m, 2H, $\text{C}_6\text{H} + \text{SO}_2\text{Ph-}p\text{-H}$), 7.11 (app-dd, $J = 0.9$, 8.3, 2H, $\text{SO}_2\text{Ph-}o\text{-H}$), 6.85 (dd, $J = 7.6$, 8.1, 2H, $\text{SO}_2\text{Ph-}m\text{-H}$), 6.84 (s, 1H, C_2H), 5.58 (d, $J = 2.5$, 1H, C_2H), 4.00 (d, $J = 15.1$, 1H, C_{12}H_a), 3.19 (d, $J = 15.1$, 1H, C_{12}H_b), 2.97 (s, 3H, C_{18}H_3), 2.04 (s, 3H, C_{17}H_3).

^{13}C NMR (100 MHz, CDCl_3 , 20 °C):

δ 166.0 (C_{13}), 161.9 (C_{16}), 140.9 (C_9), 137.6 ($\text{SO}_2\text{Ph-}ipso\text{-C}$), 137.2 (C_9), 136.9 (C_4), 133.0 ($\text{SO}_2\text{Ph-}p\text{-C}$), 129.8 (C_7), 128.3 ($\text{SO}_2\text{Ph-}m\text{-C}$), 127.2 ($\text{SO}_2\text{Ph-}o\text{-C}$), 126.1 (C_6), 124.5 (C_5), 124.3 (C_4), 124.0 (C_2), 123.1 (C_7), 120.7 (C_6), 119.3 (C_8),

118.7 (C₅), 114.1 (C₃), 112.1 (C₈), 85.1 (C₂), 73.9 (C₁₅), 73.5 (C₁₁), 55.3 (C₃), 43.0 (C₁₂), 27.8 (C₁₈), 18.4 (C₁₇).

FTIR (thin film) cm⁻¹: 3396 (br-m), 3061 (w), 2924 (w), 2851 (w), 1704 (s), 1447 (w), 1360 (m), 1332 (s), 1244 (w), 1169 (s), 1109 (m), 1090 (m), 910 (w), 735 (s).

HRMS (ESI) (*m/z*): calc'd for C₂₉H₂₅N₄O₄S₃ [M+H]⁺: 589.1032, found 589.1043.

TLC (50% ethyl acetate in hexanes), R_f: 0.27 (UV, CAM).

α-epimer 62:

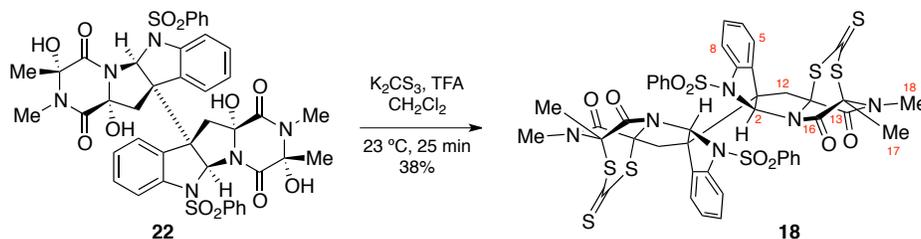
¹H NMR (600 MHz, CDCl₃, 20 °C): δ 7.99 (d, *J* = 7.4, 1H, C₅H), 7.70 (d, *J* = 8.0, 1H, C₈H), 7.57 (br-s, 1H, N₁H), 7.40–7.34 (m, 3H, C₅H + C₇H + C₈H), 7.34–7.28 (m, 2H, C₆H + C₇H), 7.21 (app-dt, *J* = 1.7, 7.6, 2H, C₆H + SO₂Ph-*p*-H), 7.08 (app-dd, *J* = 0.9, 8.3, 2H, SO₂Ph-*o*-H), 6.82 (dd, *J* = 7.6, 8.1, 2H, SO₂Ph-*m*-H), 6.82 (s, 1H, C₂H), 5.55 (d, *J* = 2.5, 1H, C₂H), 3.97 (d, *J* = 15.1, 1H, C₁₂H_a), 3.16 (d, *J* = 15.1, 1H, C₁₂H_b), 2.94 (s, 3H, C₁₈H₃), 2.01 (s, 3H, C₁₇H₃).

¹³C NMR (100 MHz, CDCl₃, 20 °C): δ 165.9 (C₁₃), 162.6 (C₁₆), 139.5 (C₉), 138.3 (SO₂Ph-*ipso*-C), 137.3 (C₉), 135.6 (C₄), 132.6 (SO₂Ph-*p*-C), 129.8 (C₇), 128.1 (SO₂Ph-*m*-C), 127.1 (SO₂Ph-*o*-C), 125.9 (C₆), 125.4 (C₅), 124.6 (C₂), 123.9 (C₄), 123.4 (C₇), 121.0 (C₆), 119.2 (C₅), 118.4 (C₈), 115.2 (C₃), 111.9 (C₈), 85.0 (C₂), 74.4 (C₁₁), 73.8 (C₁₅), 55.9 (C₃), 41.2 (C₁₂), 27.6 (C₁₈), 18.7 (C₁₇).

FTIR (thin film) cm⁻¹: 3395 (br-m), 2923 (w), 1701 (s), 1460 (w), 1359 (m), 1332 (m), 1247 (w), 1168 (m), 1090 (w), 912 (w), 734 (m).

HRMS (ESI) (*m/z*): calc'd for C₂₉H₂₅N₄O₄S₃ [M+H]⁺: 589.1032, found 589.1037.

TLC (50% ethyl acetate in hexanes), R_f: 0.27 (UV, CAM).



Dimeric bisdithiepanethione 18:

Dimeric tetraol **22** (200 mg, 226 μmol , 1 equiv) was added as a solid to a yellow solution of potassium trithiocarbonate (632 mg, 3.39 mmol, 15.0 equiv) in anhydrous dichloromethane (5.1 mL) and trifluoroacetic acid (1.7 mL) at 23 °C. After 25 min, the reaction mixture was diluted with dichloromethane (60 mL) and washed with saturated aqueous sodium bicarbonate (125 mL). The aqueous layer was extracted with dichloromethane (2 \times 30 mL) and the combined organic layers were dried over anhydrous sodium sulfate, were filtered, and were concentrated under reduced pressure to afford a yellow powder. This powder was purified by flash column chromatography on silica gel (eluent: 5% acetone in dichloromethane) to afford dimeric bisdithiepanethione **18** (88.8 mg, 38.0%) as an orange-yellow solid.

^1H NMR (500 MHz, CDCl_3 , 20 °C):

δ 7.75–7.65 (m, 2H, C_8H), 7.75–7.65 (m, 4H, $\text{SO}_2\text{Ph-}o\text{-H}$), 7.53 (app-t, $J = 7.4$, 2H, $\text{SO}_2\text{Ph-}p\text{-H}$), 7.41 (app-t, $J = 8.0$, 4H, $\text{SO}_2\text{Ph-}m\text{-H}$), 7.30–7.14 (m, 6H, C_6H , C_7H , C_5H), 6.86 (s, 2H, C_2H), 3.26 (d, $J = 14.9$, 2H, C_{12}H_a), 3.09 (d, $J = 14.9$, 2H, C_{12}H_b), 3.01 (s, 6H, C_{18}H), 1.68 (s, 6H, C_{17}H).

^{13}C NMR (125.8 MHz, CDCl_3 , 20 °C):

δ 215.1 ($\text{C}=\text{S}$), 164.1 (C_{13}), 159.7 (C_{16}), 142.7 (C_9), 141.9 ($\text{SO}_2\text{Ph-}ipso\text{-C}$), 133.1 ($\text{SO}_2\text{Ph-}p\text{-C}$), 131.3 (C_4), 129.2 ($\text{SO}_2\text{Ph-}m\text{-C}$), 129.2 (C_6), 125.5 ($\text{SO}_2\text{Ph-}o\text{-C}$), 125.2 (C_7), 124.5 (C_8), 116.1 (C_5), 81.6 (C_2), 73.9 (C_{11}), 73.6 (C_{15}), 59.1 (C_3), 44.7 (C_{12}), 28.6 (C_{18}), 19.3 (C_{17}).

FTIR (thin film) cm^{-1} :

1715 (s), 1691 (s), 1479 (m), 1462 (m), 1447 (m), 1359 (s), 1169 (s), 729 (m).

HRMS (ESI) (m/z):

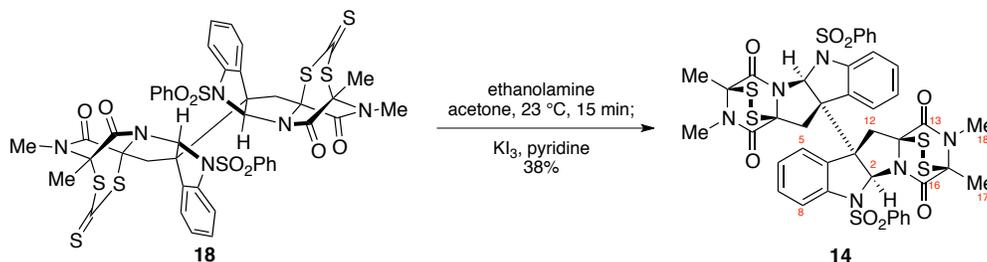
calc'd for $\text{C}_{44}\text{H}_{36}\text{N}_6\text{NaO}_8\text{S}_8$ [$\text{M}+\text{Na}$] $^+$: 1055.0252, found 1055.0255.

$[\alpha]_D^{24}$:

+ 230 (c 0.19, CHCl_3).

TLC (5% acetone in dichloromethane), R_f :

0.27 (UV, CAM).



Dimeric epidithiodiketopiperazine **14**:

Ethanolamine (500 μL) was added via syringe to a solution of dimeric bisdithiepanethione **18** (11.2 mg, 10.8 μmol , 1 equiv) in acetone (500 μL) at 23 °C. After 15 min, the reaction mixture was diluted with dichloromethane (30 mL) and aqueous hydrochloric acid solution (1 N, 30 mL). The organic layer was collected, and the aqueous layer was extracted with dichloromethane (2 \times 5 mL). A solution of potassium triiodide in pyridine (2.5% w/v) was added dropwise to the combined organic layers until a persistent yellow color was observed. The resulting mixture was washed with aqueous hydrochloric acid (1 N, 30 mL) and the layers were separated. The aqueous layer was extracted with dichloromethane (2 \times 5 mL), and the combined organic layers were dried over anhydrous sodium sulfate, were filtered, and were concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (eluent: 5% acetone in dichloromethane) to afford dimeric epidithiodiketopiperazine **14** (3.9 mg, 38%) as a white solid.

¹H NMR (500 MHz, CDCl₃, 20 °C):

δ 7.85 (dd, $J = 1.4, 7.3$, 4H, SO₂Ph-*o*-H), 7.68 (d, $J = 7.5$, 2H, C₈H), 7.54 (tt, $J = 1.2, 7.5$, 2H, SO₂Ph-*p*-H), 7.46 (app-t, $J = 8.0$, 4H, SO₂Ph-*m*-H), 7.20 (app-dt, $J = 1.3, 7.5$, 2H, C₆H), 7.16 (app-dt, $J = 1.2, 7.5$, 2H, C₇H), 7.04 (dd, $J = 1.0, 7.6$, 2H, C₅H), 6.83 (s, 2H, C₂H), 3.55 (d, $J = 15.2$, 2H, C₁₂H_a), 2.97 (s, 6H, C₁₈H), 2.95 (d, $J = 15.2$, 2H, C₁₂H_b), 1.62 (s, 6H, C₁₇H).

¹³C NMR (125.8 MHz, CDCl₃, 20 °C):

δ 164.9 (C₁₃), 160.8 (C₁₆), 142.5 (C₉), 142.4 (SO₂Ph-*ipso*-C), 132.6 (SO₂Ph-*p*-C), 130.9 (C₄), 130.6 (C₆), 129.0 (SO₂Ph-*m*-C), 125.7 (SO₂Ph-*o*-C), 125.2 (C₇), 124.7 (C₈), 116.3 (C₅), 81.9 (C₂), 73.8 (C₁₅), 73.4 (C₁₁), 60.5 (C₃), 41.9 (C₁₂), 27.8 (C₁₈), 17.9 (C₁₇).

FTIR (thin film) cm⁻¹:

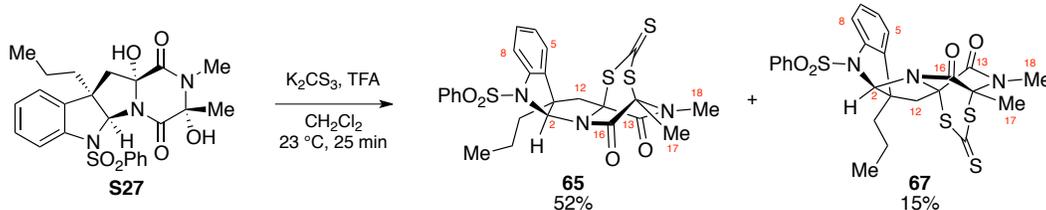
1716 (s), 1688 (s), 1480 (m), 1462 (m), 1447 (w), 1348 (s), 1168 (m).

HRMS (ESI) (m/z):

calc'd for C₄₂H₃₇N₆O₈S₆ [M+H]⁺: 945.0992, found 945.0968.

TLC (5% acetone in dichloromethane), R_f:

0.21 (UV, CAM).



C3-Propyl dithiepanethiones **65 and **67**:**

A solution of the tetracyclic diol **S27** (228 mg, 470 μmol , 1 equiv) in dichloromethane (3.5 mL) was added to a yellow solution of potassium trithiocarbonate (438 mg, 2.35 mmol, 5.00 equiv) in anhydrous dichloromethane (7 mL) and trifluoroacetic acid (3 mL) at 23 °C. An additional portion of trifluoroacetic acid (1.5 mL) was added to the reaction mixture via syringe. After 25 min, the reaction mixture was diluted with dichloromethane (60 mL) and washed with saturated aqueous sodium bicarbonate (125 mL). The aqueous layer was extracted with dichloromethane (2 \times 30 mL) and the combined organic layers were dried over anhydrous sodium sulfate, were filtered, and were concentrated under reduced pressure to yield a yellow powder. This powder was purified by flash column chromatography on silica gel (eluent: 20% acetone in dichloromethane) to afford diastereomeric dithiepanethiones **65** (137 mg, 52.0%) and **67** (38.7 mg, 14.7%) as yellow films.

β -epimer **65:**

^1H NMR (500 MHz, CDCl_3 , 20 °C):

δ 7.72 (d, $J = 7.5$, 2H, $\text{SO}_2\text{Ph-}o\text{-H}$), 7.52 (t, $J = 7.5$, 1H, $\text{SO}_2\text{Ph-}p\text{-H}$), 7.40 (app-t, $J = 7.9$, 2H, $\text{SO}_2\text{Ph-}m\text{-H}$), 7.35 (d, $J = 7.1$, 1H, C_8H), 7.29 (app-dt, $J = 1.7$, 8.2, 1H, C_7H), 7.19 (app-dt, $J = 0.9$, 7.7, 1H, C_6H), 7.16 (dd, $J = 1.4$, 7.6, 1H, C_5H), 6.29 (s, 1H, C_2H), 3.00 (s, 3H, C_{18}H), 2.98 (d, $J = 15.1$, 1H, C_{12}H_a), 2.75 (d, $J = 15.1$, 1H, C_{12}H_b), 1.79 (s, 3H, C_{17}H), 1.47–1.31 (m, 2H, $\text{CH}_2\text{CH}_2\text{CH}_3$), 1.47–1.31 (m, 1H, $\text{CH}_2\text{CH}_a\text{H}_b\text{CH}_3$), 1.19–1.06 (m, 1H, $\text{CH}_2\text{CH}_a\text{H}_b\text{CH}_3$), 0.78 (app-t, $J = 7.0$, 3H $\text{CH}_2\text{CH}_2\text{CH}_3$).

^{13}C NMR (125.8 MHz, CDCl_3 , 20 °C):

δ 215.9 ($\text{C}=\text{S}$), 164.7 (C_{13}), 160.5 (C_{16}), 141.6 (C_9), 140.4 ($\text{SO}_2\text{Ph-}ipso\text{-C}$), 135.4 (C_4), 133.3 ($\text{SO}_2\text{Ph-}p\text{-C}$), 129.7 (C_7), 129.2 ($\text{SO}_2\text{Ph-}m\text{-C}$), 126.5 ($\text{SO}_2\text{Ph-}o\text{-C}$), 125.9 (C_6), 123.6 (C_5), 117.6 (C_8), 83.7 (C_2), 74.6 (C_{11}), 73.5 (C_{15}), 54.5 (C_3), 46.1 (C_{12}), 40.5 ($\text{CH}_2\text{CH}_2\text{CH}_3$), 28.5 (C_{18}), 19.7 (C_{17}), 18.0 ($\text{CH}_2\text{CH}_2\text{CH}_3$), 14.3 ($\text{CH}_2\text{CH}_2\text{CH}_3$).

FTIR (thin film) cm^{-1} :

1711 (s), 1686 (s), 1477 (m), 1461 (m), 1447 (m), 1365 (s), 1167 (s), 732 (m).

HRMS (ESI) (m/z):

calc'd for $\text{C}_{25}\text{H}_{25}\text{N}_3\text{NaO}_4\text{S}_4$ $[\text{M}+\text{Na}]^+$: 582.0620, found 582.0646.

TLC (40% ethyl acetate in hexanes), R_f :

0.18 (UV, CAM).

α -epimer 67:

^1H NMR (500 MHz, CDCl_3 , 20 °C):

δ 7.79 (dd, $J = 1.0, 7.3$, 2H, $\text{SO}_2\text{Ph-}o\text{-H}$), 7.57 (d, $J = 8.0$, 1H, C_8H), 7.53 (t, $J = 7.5$, 1H, $\text{SO}_2\text{Ph-}p\text{-H}$), 7.40 (app-t, $J = 7.8$, 2H, $\text{SO}_2\text{Ph-}m\text{-H}$), 7.27 (app-dt, $J = 1.4, 7.8$, 1H, C_7H), 7.12 (app-dt, $J = 0.9, 7.6$, 1H, C_6H), 7.06 (dd, $J = 0.8, 7.5$, 1H, C_5H), 6.06 (s, 1H, C_2H), 3.42 (d, $J = 15.7$, 1H, C_{12}H_a), 2.97 (s, 3H, C_{18}H), 2.44 (d, $J = 15.7$, 1H, C_{12}H_b), 1.95 (s, 3H, C_{17}H), 1.37–1.26 (m, 1H, $\text{CH}_2\text{CH}_a\text{H}_b\text{CH}_3$), 1.26–1.14 (m, 1H, $\text{CH}_2\text{CH}_a\text{H}_b\text{CH}_3$), 0.97–0.83 (m, 2H, $\text{CH}_2\text{CH}_2\text{CH}_3$), 0.69 (app-t, $J = 6.8$, 3H, $\text{CH}_2\text{CH}_2\text{CH}_3$).

^{13}C NMR (125.8 MHz, CDCl_3 , 20 °C):

δ 216.8 ($\text{C}=\text{S}$), 164.3 (C_{13}), 161.4 (C_{15}), 139.3 (C_9), 138.9 ($\text{SO}_2\text{Ph-}ipso\text{-C}$), 138.3 (C_4), 133.8 ($\text{SO}_2\text{Ph-}p\text{-C}$), 129.3 ($\text{SO}_2\text{Ph-}m\text{-C}$), 129.3 (C_7), 127.7 ($\text{SO}_2\text{Ph-}o\text{-C}$), 126.3 (C_6), 124.4 (C_5), 118.3 (C_8), 84.5 (C_2), 74.8 (C_{11}), 74.1 (C_{15}), 55.0 (C_3), 42.4 (C_{12}), 40.0 ($\text{CH}_2\text{CH}_2\text{CH}_3$), 28.6 (C_{18}), 21.0 (C_{17}), 18.3 ($\text{CH}_2\text{CH}_2\text{CH}_3$), 14.2 ($\text{CH}_2\text{CH}_2\text{CH}_3$).

FTIR (thin film) cm^{-1} :

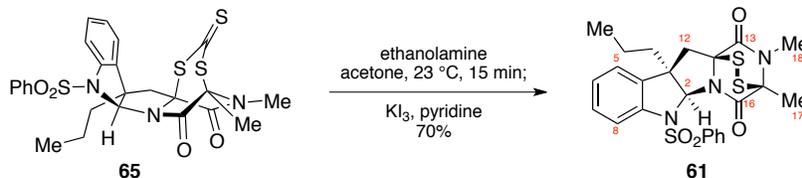
1712 (s), 1691 (s), 1476 (m), 1461 (m), 1447 (m), 1368 (s), 1333 (s), 1172 (s), 727 (w).

HRMS (ESI) (m/z):

calc'd for $\text{C}_{25}\text{H}_{25}\text{N}_3\text{NaO}_4\text{S}_4$ [$\text{M}+\text{Na}$] $^+$: 582.0620, found 582.0636.

TLC (40% ethyl acetate in hexanes), R_f :

0.50 (UV, CAM).



β-C3-Propyl epidithiodiketopiperazine 61:

Ethanolamine (500 μL) was added via syringe to a solution of dithiepanethione **65** (13.3 mg, 23.8 μmol , 1 equiv) in acetone (500 μL) at 23 °C. After 15 min, the reaction mixture was diluted with dichloromethane (30 mL) and aqueous hydrochloric acid solution (2 N, 30 mL). The organic layer was collected, and the aqueous layer was extracted with dichloromethane (2 \times 2 mL). A solution of potassium triiodide in pyridine (2.5% w/v) was added dropwise to the combined organic layers until a persistent yellow color was observed. The resulting mixture was washed with aqueous hydrochloric acid (2 N, 30 mL) and the layers were separated. The aqueous layer was extracted with dichloromethane (2 \times 5 mL), and the combined organic layers were dried over anhydrous sodium sulfate, were filtered, and were concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (eluent: 1% acetone in dichloromethane) to afford epidithiodiketopiperazine **61** (8.6 mg, 70%) as a clear film.

¹H NMR (500 MHz, CDCl₃, 20 °C):

δ 7.80 (d, $J = 7.0$, 2H, SO₂Ph-*o*-H), 7.53 (t, $J = 7.0$, 1H, SO₂Ph-*p*-H), 7.46–7.37 (m, 1H, C₈H), 7.46–7.37 (m, 2H, SO₂Ph-*m*-H), 7.29 (app-dt, $J = 1.1$, 7.7, 1H, C₇H), 7.16 (app-t, $J = 7.6$, 1H, C₆H), 7.12 (d, $J = 7.6$, 1H, C₅H), 6.09 (s, 1H, C₂H), 3.19 (d, $J = 15.2$, 1H, C₁₂H_a), 2.98 (s, 3H, C₁₈H), 2.57 (d, $J = 15.2$, 1H, C₁₂H_b), 1.87 (s, 3H, C₁₇H), 1.43–1.30 (m, 1H, CH₂CH_aH_bCH₃), 1.22–1.04 (m, 1H, CH₂CH_aH_bCH₃), 1.22–1.04 (m, 2H, CH₂CH₂CH₃), 0.77–0.68 (m, 3H, CH₂CH₂CH₃).

¹³C NMR (125.8 MHz, CDCl₃, 20 °C):

δ 165.9 (C₁₃), 161.6 (C₁₆), 141.1 (C₉), 139.8 (SO₂Ph-*ipso*-C), 137.6 (C₄), 133.4 (SO₂Ph-*p*-C), 129.3 (C₇), 129.2 (SO₂Ph-*m*-C), 127.4 (SO₂Ph-*o*-C), 125.9 (C₆), 123.6 (C₅), 118.4 (C₈), 83.7 (C₂), 73.7 (C₁₁), 73.5 (C₁₅), 55.9 (C₃), 41.8 (C₁₂), 40.0 (CH₂CH₂CH₃), 27.7 (C₁₈), 18.3 (CH₂CH₂CH₃), 18.0 (C₁₇), 14.3 (CH₂CH₂CH₃).

FTIR (thin film) cm⁻¹:

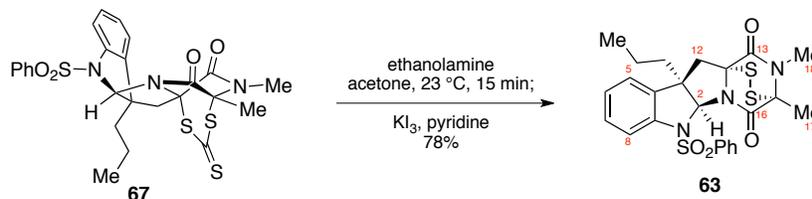
1713 (s), 1688 (s), 1478 (m), 1460 (m), 1447 (m), 1341 (s), 1172 (s), 719 (w).

HRMS (ESI) (m/z):

calc'd for C₂₄H₂₅N₃NaO₄S₃ [M+Na]⁺: 538.0899, found 538.0923.

TLC (1% acetone in dichloromethane), R_f:

0.21 (UV, CAM).



α -C3-Propyl epidithiodiketopiperazine 63:

Ethanolamine (500 μ L) was added via syringe to a solution of dithiepanethione **67** (13.3 mg, 23.8 μ mol, 1 equiv) in acetone (500 μ L) at 23 °C. After 15 min, the reaction mixture was diluted with dichloromethane (30 mL) and aqueous hydrochloric acid solution (2 N, 30 mL). The organic layer was collected, and the aqueous layer was extracted with dichloromethane (2 \times 5 mL). A solution of potassium triiodide in pyridine (2.5% w/v) was added dropwise to the combined organic layers until a persistent yellow color was observed. The resulting mixture was washed with aqueous hydrochloric acid (2 N, 30 mL) and the layers were separated. The aqueous layer was extracted with dichloromethane (2 \times 5 mL), and the combined organic layers were dried over anhydrous sodium sulfate, were filtered, and were concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (eluent: 30% ethyl acetate in dichloromethane) to afford epidithiodiketopiperazine **63** (9.6 mg, 78%) as a clear film.

^1H NMR (500 MHz, CDCl_3 , 20 °C):

δ 7.83 (dd, $J = 0.8, 8.2$, 2H, $\text{SO}_2\text{Ph-}o\text{-H}$), 7.53 (t, $J = 7.4$, 1H, $\text{SO}_2\text{Ph-}p\text{-H}$), 7.51 (d, $J = 7.9$, 1H, C_8H), 7.40 (app-t, $J = 8.1$, 2H, $\text{SO}_2\text{Ph-}m\text{-H}$), 7.28–7.19 (m, 1H, C_7H), 7.13–7.05 (m, 1H, C_6H), 7.13–7.05 (m, 1H, C_5H), 6.14 (s, 1H, C_2H), 3.57 (d, $J = 14.9$, 1H, C_{12}H_a), 2.89 (s, 3H, C_{18}H), 2.37 (d, $J = 14.9$, 1H, C_{12}H_b), 1.93 (s, 3H, C_{17}H), 1.38–1.14 (m, 2H, $\text{CH}_2\text{CH}_2\text{CH}_3$), 1.00–0.85 (m, 2H, $\text{CH}_2\text{CH}_2\text{CH}_3$), 0.70 (app-t, $J = 7.2$, 3H, $\text{CH}_2\text{CH}_2\text{CH}_3$).

^{13}C NMR (125.8 MHz, CDCl_3 , 20 °C):

δ 165.7 (C_{13}), 162.9 (C_{16}), 139.3 (C_9), 139.1 ($\text{SO}_2\text{Ph-}ipso\text{-C}$), 137.4 (C_4), 133.7 ($\text{SO}_2\text{Ph-}p\text{-C}$), 129.3 ($\text{SO}_2\text{Ph-}m\text{-C}$), 129.3 (C_7), 127.7 ($\text{SO}_2\text{Ph-}o\text{-C}$), 126.1 (C_6), 124.5 (C_5), 118.1 (C_8), 84.3 (C_2), 74.6 (C_{11}), 73.9 (C_{15}), 56.4 (C_3), 40.5 (C_{12}), 39.7 ($\text{CH}_2\text{CH}_2\text{CH}_3$), 27.5 (C_{18}), 18.7 (C_{17}), 18.1 ($\text{CH}_2\text{CH}_2\text{CH}_3$), 14.2 ($\text{CH}_2\text{CH}_2\text{CH}_3$).

FTIR (thin film) cm^{-1} :

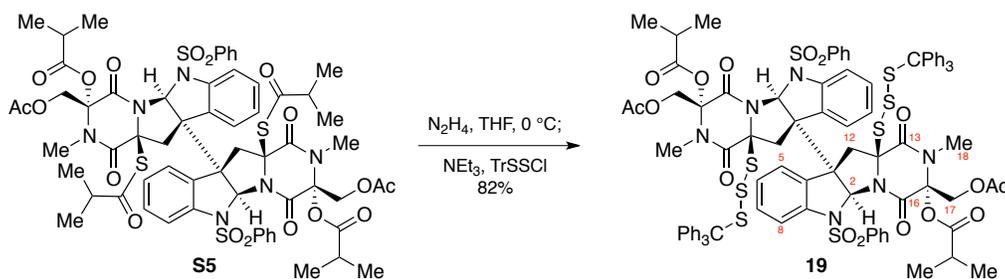
1694 (s), 1447 (m), 1366 (s), 1331 (m), 1172 (s), 722 (w).

HRMS (ESI) (m/z):

calc'd for $\text{C}_{24}\text{H}_{25}\text{N}_3\text{NaO}_4\text{S}_3$ $[\text{M}+\text{Na}]^+$: 538.0899, found 538.0920.

TLC (30% ethyl acetate in hexanes), R_f :

0.21 (UV, CAM).



Dimeric bis(triphenylmethanetrисульфид) **19**:

Anhydrous hydrazine (0.8 μL , 25 μmol , 5.00 equiv) was added via syringe to a solution of diaminodithioisobutyrate (+)-**S5** (6.6 mg, 5.0 μmol , 1 equiv) in tetrahydrofuran (2 mL) at 0 $^{\circ}\text{C}$. After 18 min, triethylamine (17.5 μL , 126 μmol , 25.0 equiv) and solid chloro(triphenylmethyl)disulfane (17.2 mg, 50.3 μmol , 10.0 equiv) were sequentially added to the reaction mixture under an inert atmosphere. After 13 min, saturated aqueous ammonium chloride (3 mL) was added to the reaction mixture. The solution was then poured into a separatory funnel containing saturated aqueous ammonium chloride (10 mL) and dichloromethane (15 mL). The aqueous layer was extracted with dichloromethane (2 \times 5 mL), and the combined organic layers were dried over anhydrous sodium sulfate, were filtered, and were concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (eluent: 35% ethyl acetate in hexanes) to afford dimeric bis(triphenylmethanetrисульфид) (+)-**19** (7.4 mg, 82%) as a slightly off-white solid.

^1H NMR (500 MHz, CDCl_3 , 20 $^{\circ}\text{C}$):

δ 8.04 (d, $J = 7.5$, 4H, $\text{SO}_2\text{Ph-}o\text{-H}$), 7.64 (t, $J = 7.5$, 2H, $\text{SO}_2\text{Ph-}p\text{-H}$), 7.52 (app-t, $J = 7.9$, 4H, $\text{SO}_2\text{Ph-}m\text{-H}$), 7.22–7.12 (m, 2H, C_8H), 7.22–7.12 (m, 18H, $\text{C}(\text{C}_6\text{H}_5)_3$), 6.99–6.90 (m, 12H, $\text{C}(\text{C}_6\text{H}_5)_3$), 6.80 (s, 2H, C_2H), 6.65 (br-s, 2H, C_5H), 6.57 (app-t, $J = 8.1$, 2H, C_7H), 6.08 (app-t, $J = 7.0$, 2H, C_6H), 4.43 (d, $J = 11.9$, 2H, C_{17}H_a), 4.23 (d, $J = 11.7$, 2H, C_{17}H_b), 3.31 (d, $J = 14.5$, 2H, C_{12}H_a), 2.92 (d, $J = 14.4$, 2H, C_{12}H_b), 2.71 (s, 6H, C_{18}H), 2.54 (app-sp, $J = 7.1$, 2H, $\text{CH}_{\text{isobutyrate}}$), 1.79 (s, 6H, $\text{CH}_{\text{acetate}}$), 1.11 (d, $J = 7.0$, 6H, $\text{CH}_{\text{isobutyrate}}$), 1.08 (d, $J = 7.1$, 6H, $\text{CH}_{\text{isobutyrate}}$).

^{13}C NMR (125.8 MHz, CDCl_3 , 20 $^{\circ}\text{C}$):

δ 174.1 ($\text{C}=\text{O}_{\text{isobutyrate}}$), 170.1 ($\text{C}=\text{O}_{\text{acetate}}$), 164.1 (C_{13}), 161.2 (C_{16}), 143.3 ($\text{C}(\text{C}_6\text{H}_5)_3$), 142.8 (C_9), 140.5 ($\text{SO}_2\text{Ph-}ipso\text{-C}$), 133.4 ($\text{SO}_2\text{Ph-}p\text{-C}$), 130.7 (C_4), 130.5 ($\text{C}(\text{C}_6\text{H}_5)_3$), 129.6 ($\text{SO}_2\text{Ph-}m\text{-C}$), 129.4 (C_7), 128.0 ($\text{C}(\text{C}_6\text{H}_5)_3$), 127.3 ($\text{C}(\text{C}_6\text{H}_5)_3$), 127.3 ($\text{SO}_2\text{Ph-}o\text{-C}$), 124.0 (C_5), 123.8 (C_6), 112.8 (C_8), 86.2 (C_{15}), 80.9 (C_2), 75.2 (C_{11}), 73.3 ($\text{C}(\text{C}_6\text{H}_5)_3$), 64.7 (C_{17}), 60.7 (C_3), 42.8 (C_{12}), 33.6 ($\text{CH}_{\text{isobutyrate}}$), 28.7 (C_{18}), 21.4 ($\text{CH}_{\text{acetate}}$), 18.8 ($\text{CH}_{\text{isobutyrate}}$).

FTIR (thin film) cm^{-1} :

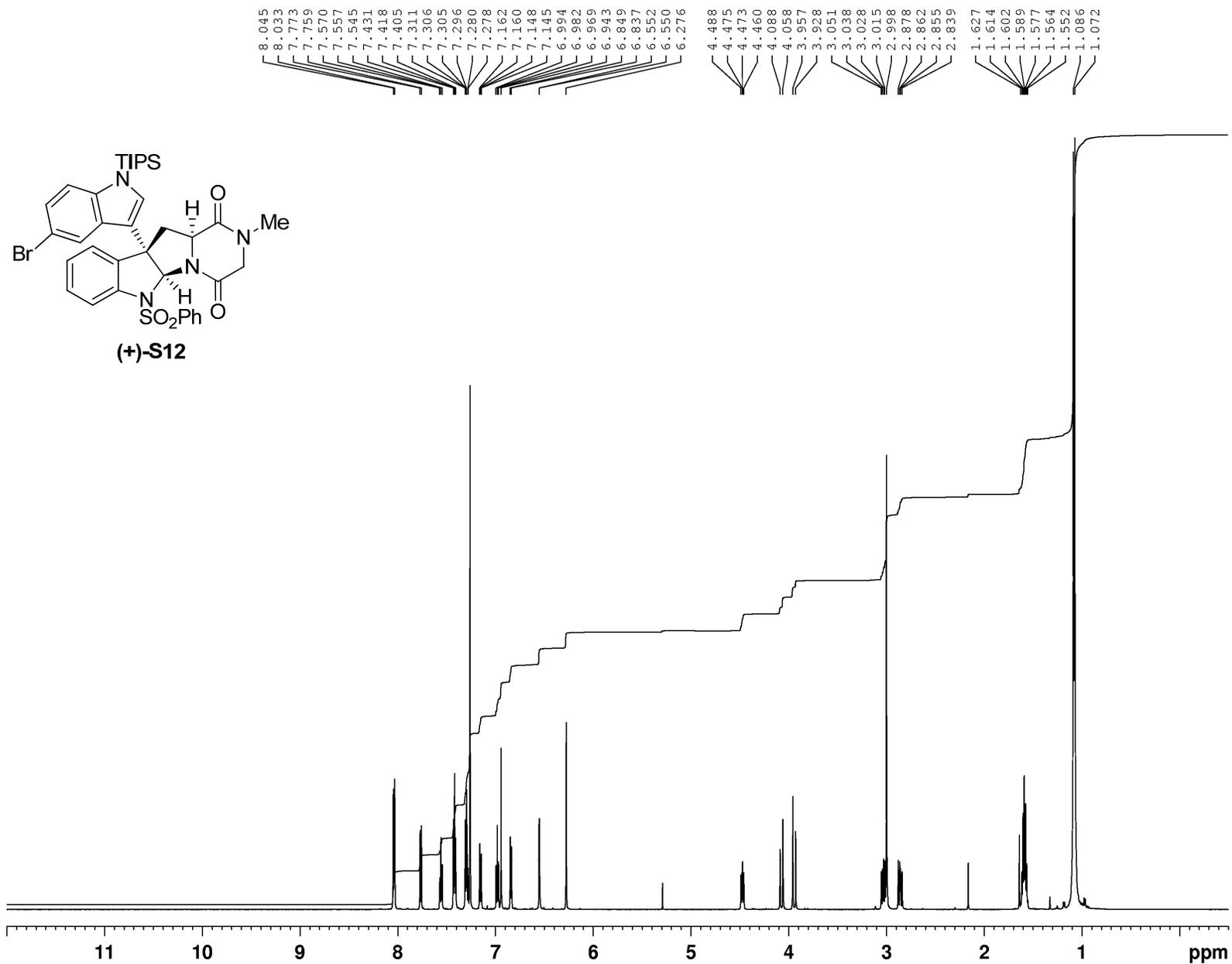
1749 (s), 1708 (s), 1480 (m), 1462 (m), 1447 (m), 1380 (s), 1220 (m), 1173 (s), 729 (m), 699 (m).

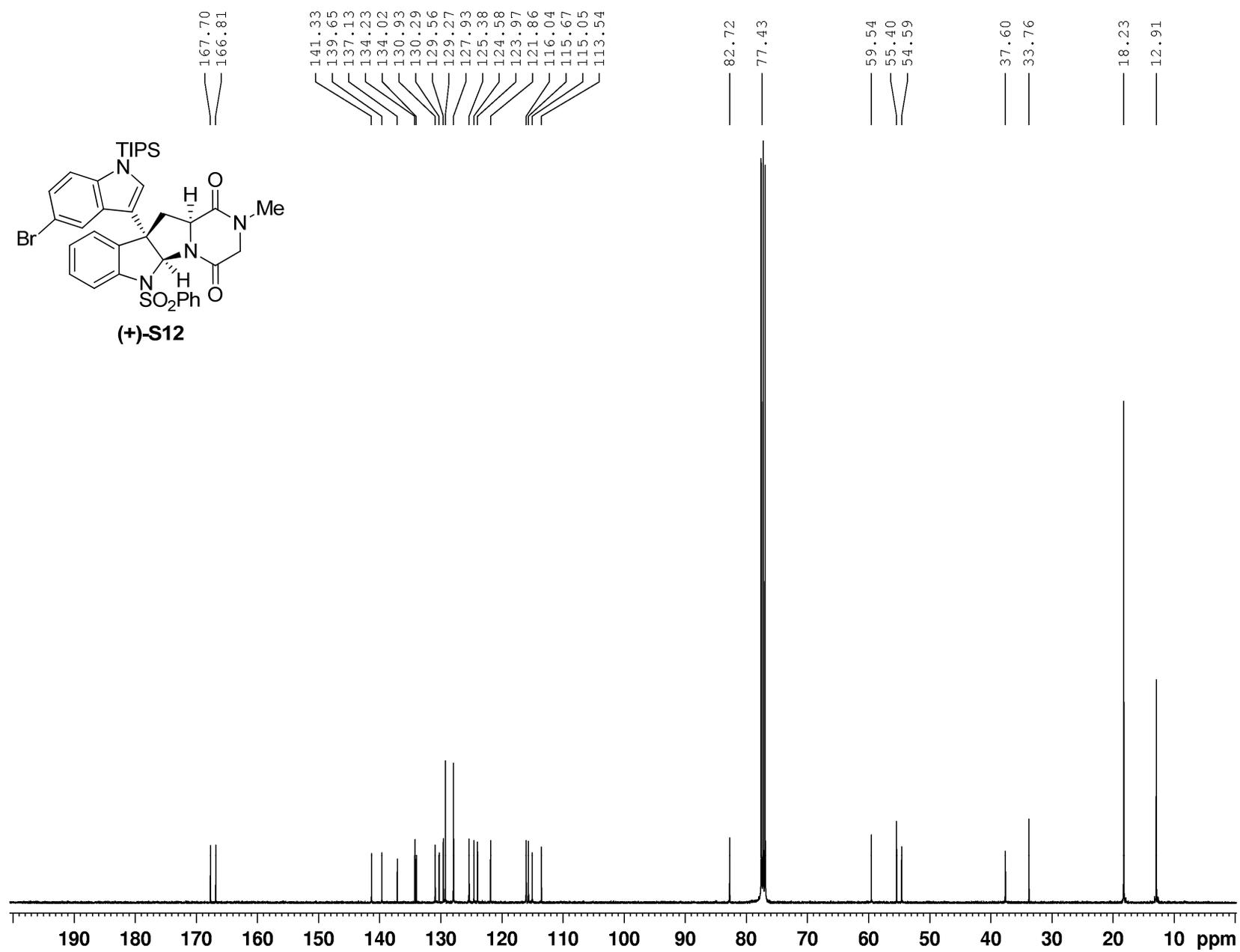
HRMS (ESI) (m/z): calc'd for $C_{92}H_{88}N_7O_{16}S_8$ $[M+NH_4]^+$: 1802.4048,
found 1802.4073.

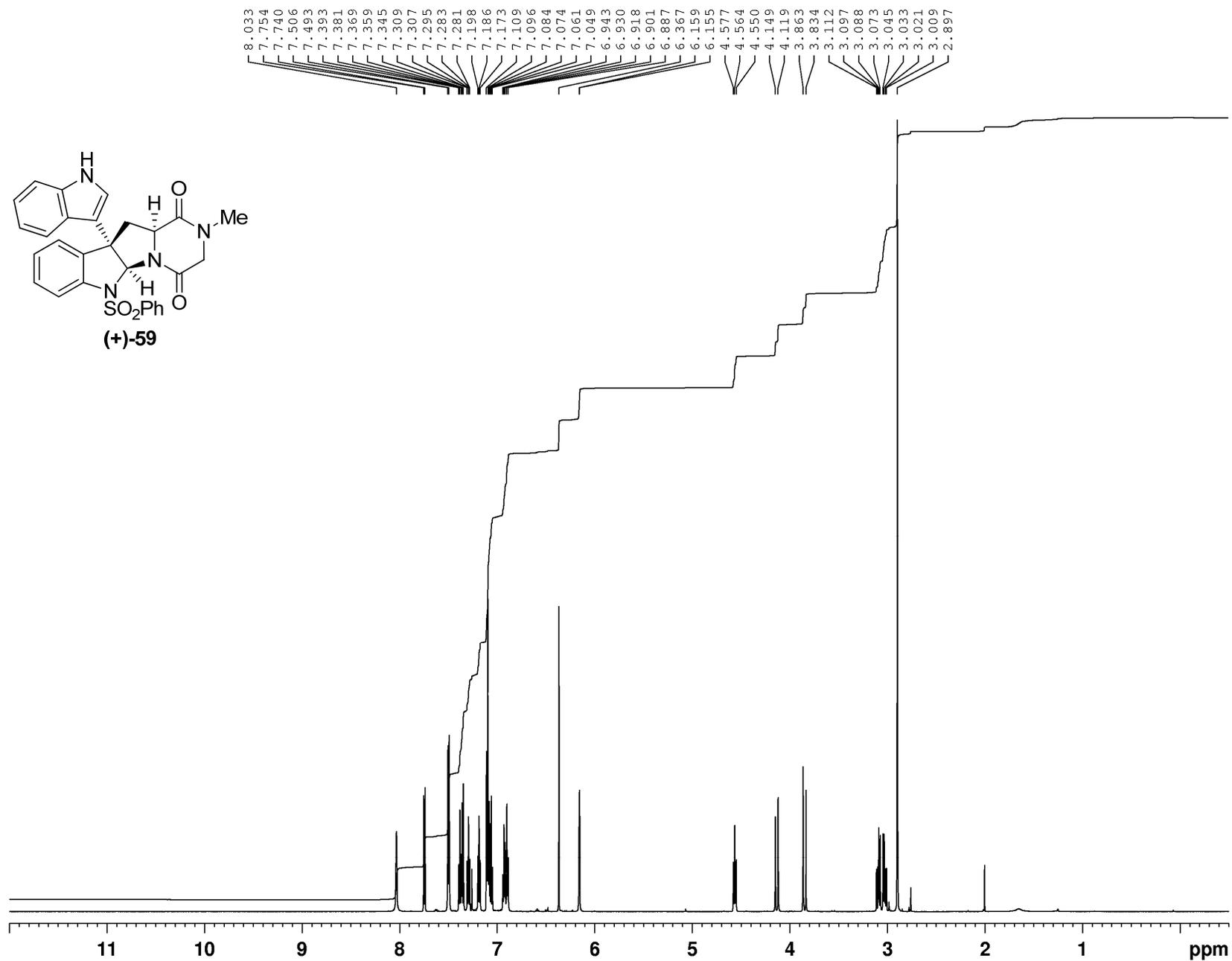
$[\alpha]_D^{24}$: + 287 (c 0.35, $CHCl_3$).

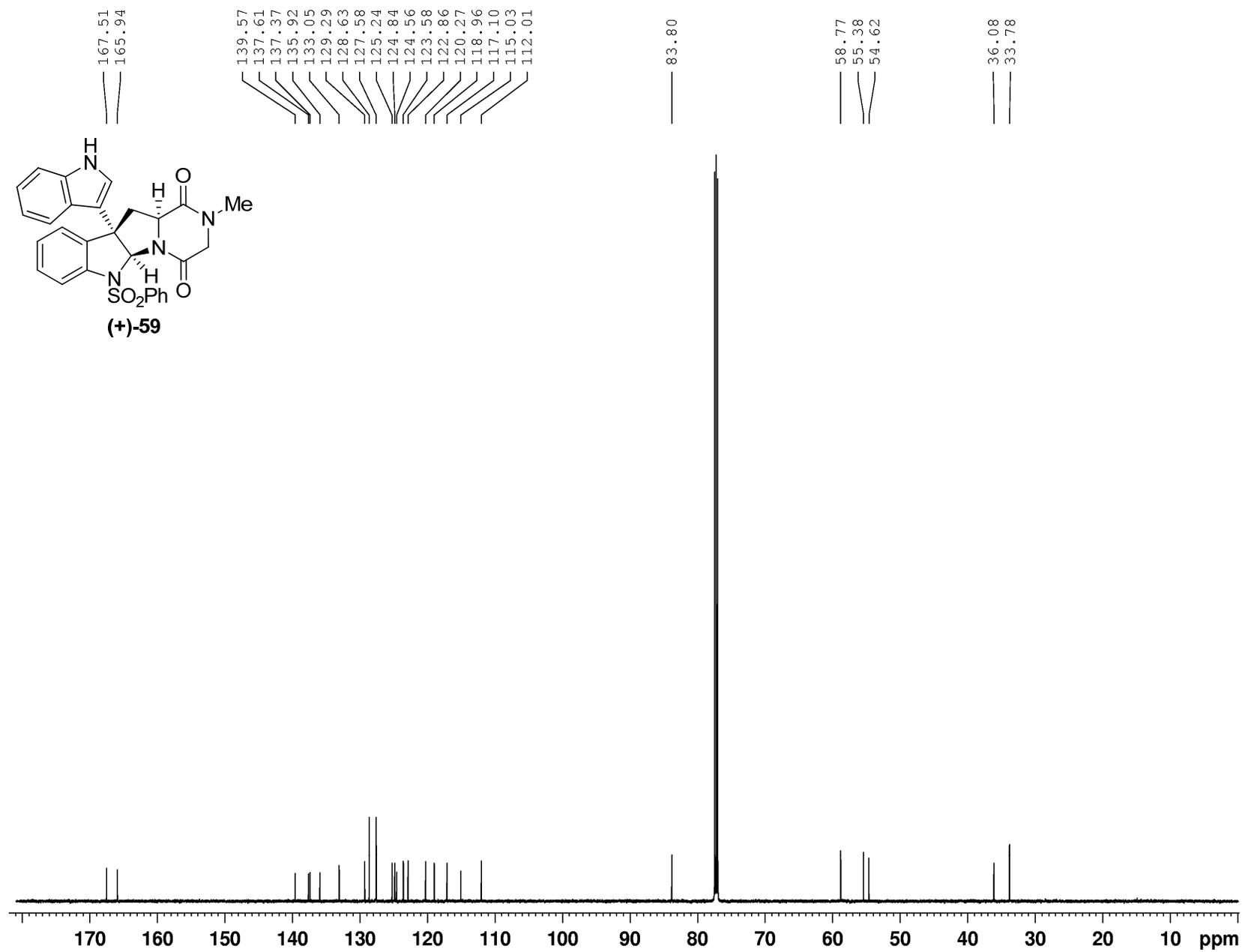
TLC (35% ethyl acetate in hexanes), R_f : 0.23 (UV, CAM).

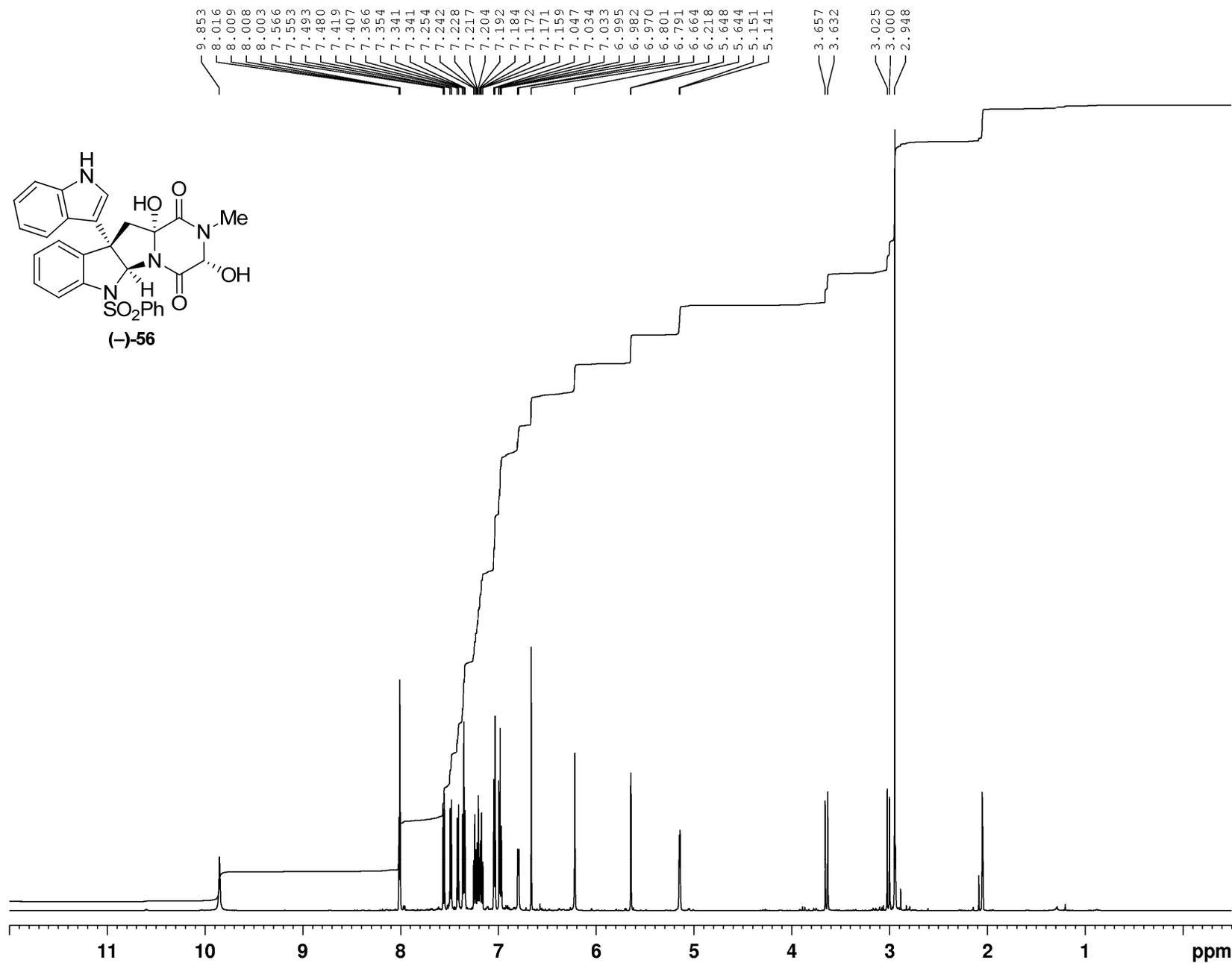
Copies of ^1H , ^{13}C and ^{19}F NMR Spectra.

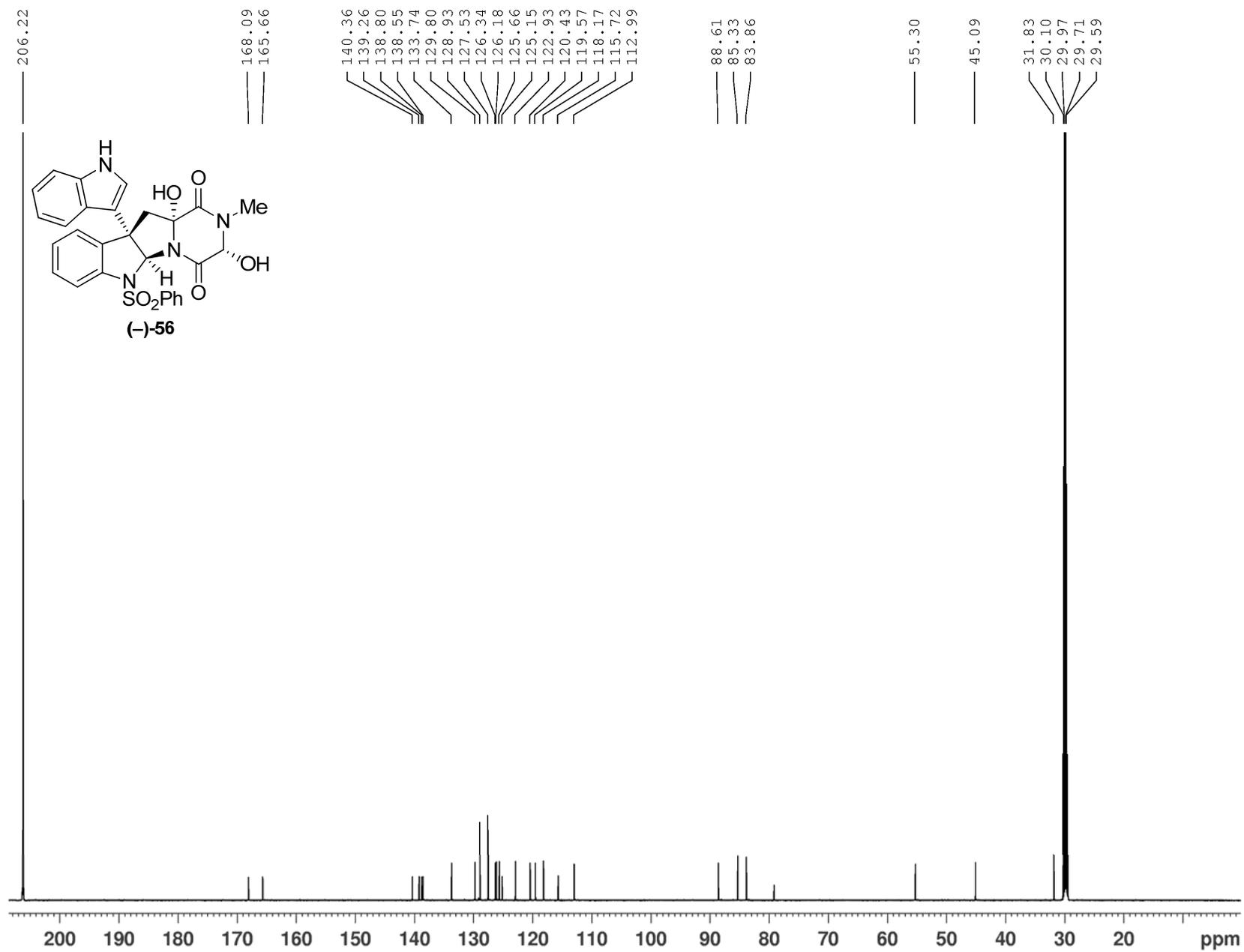


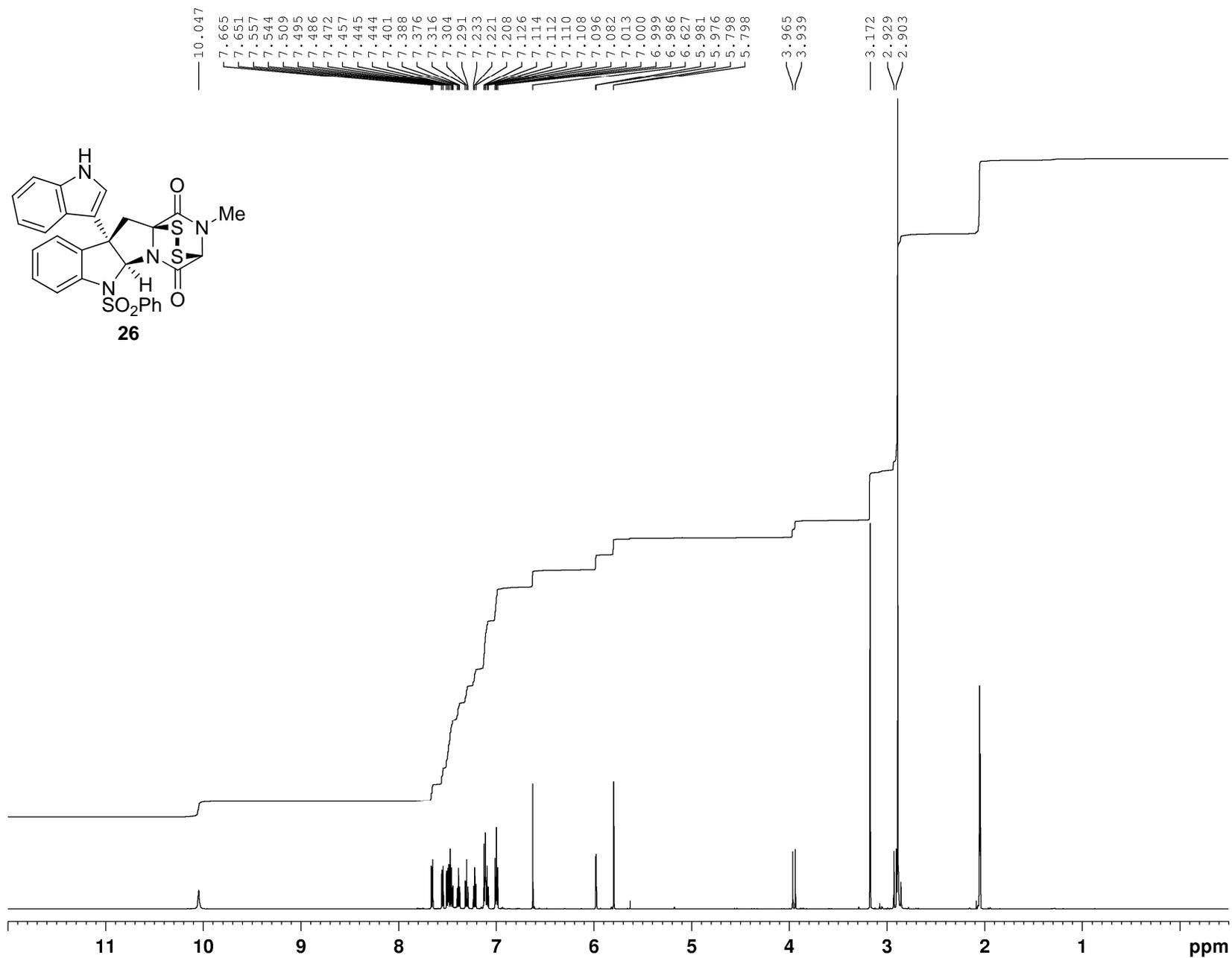


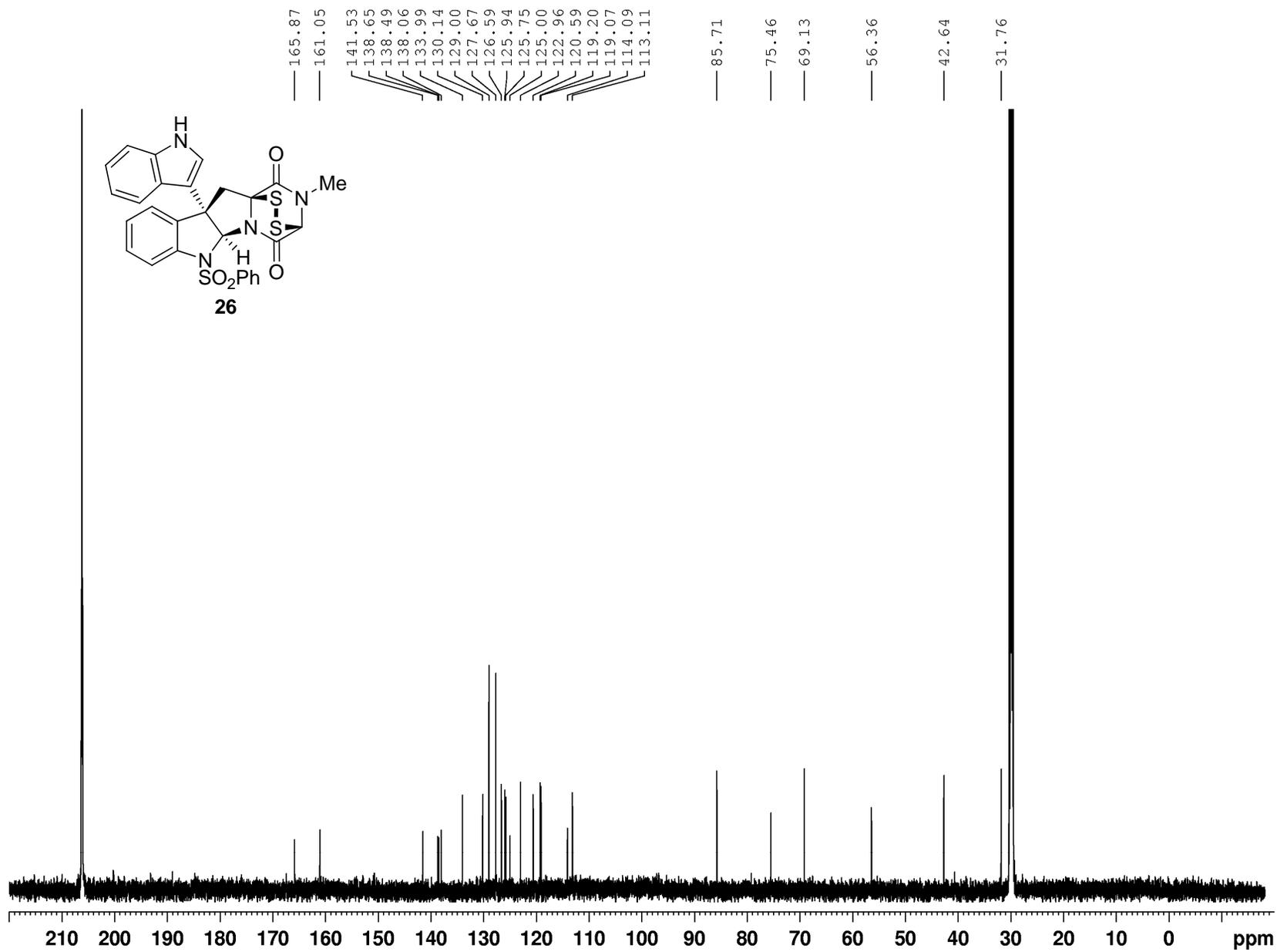


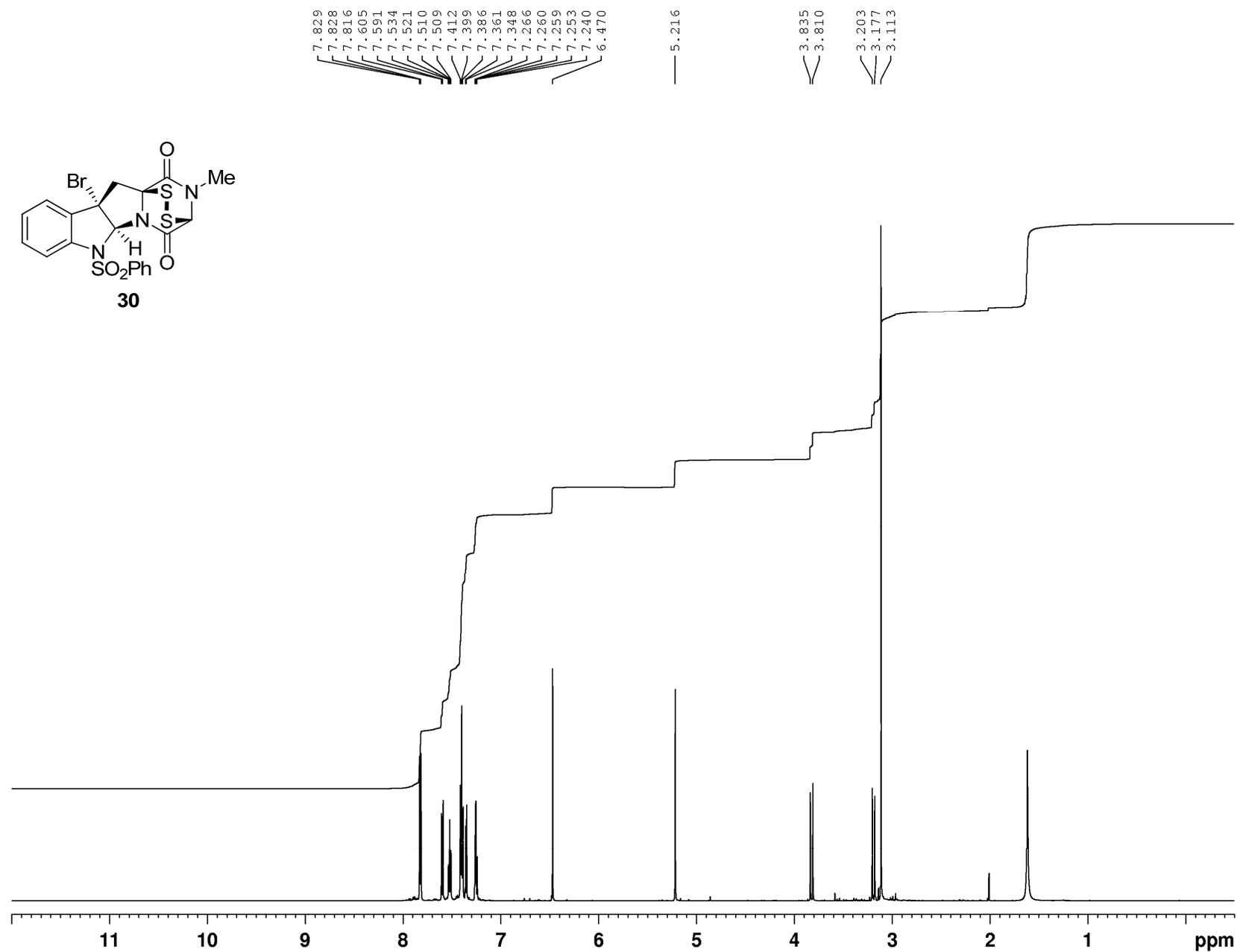


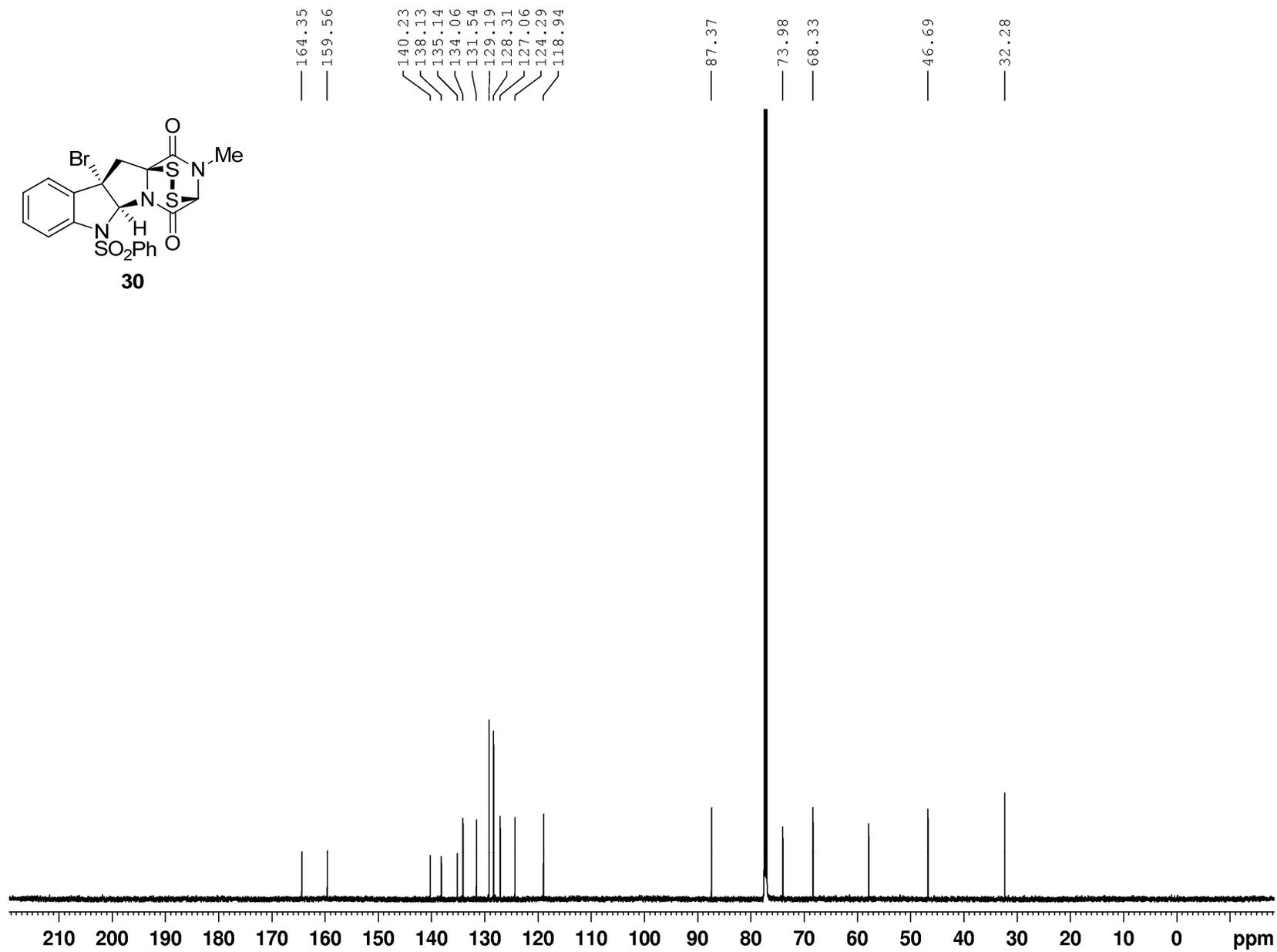


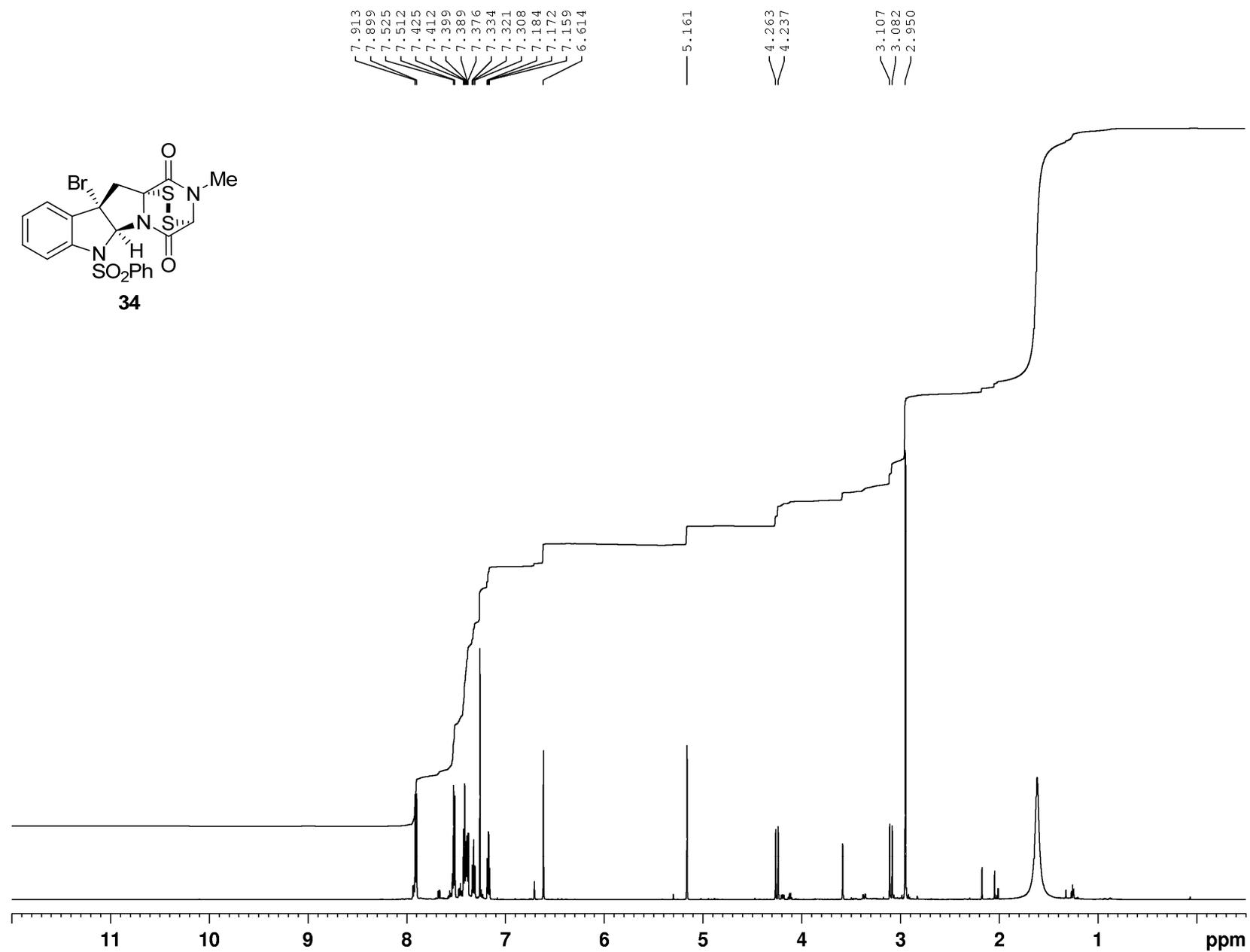


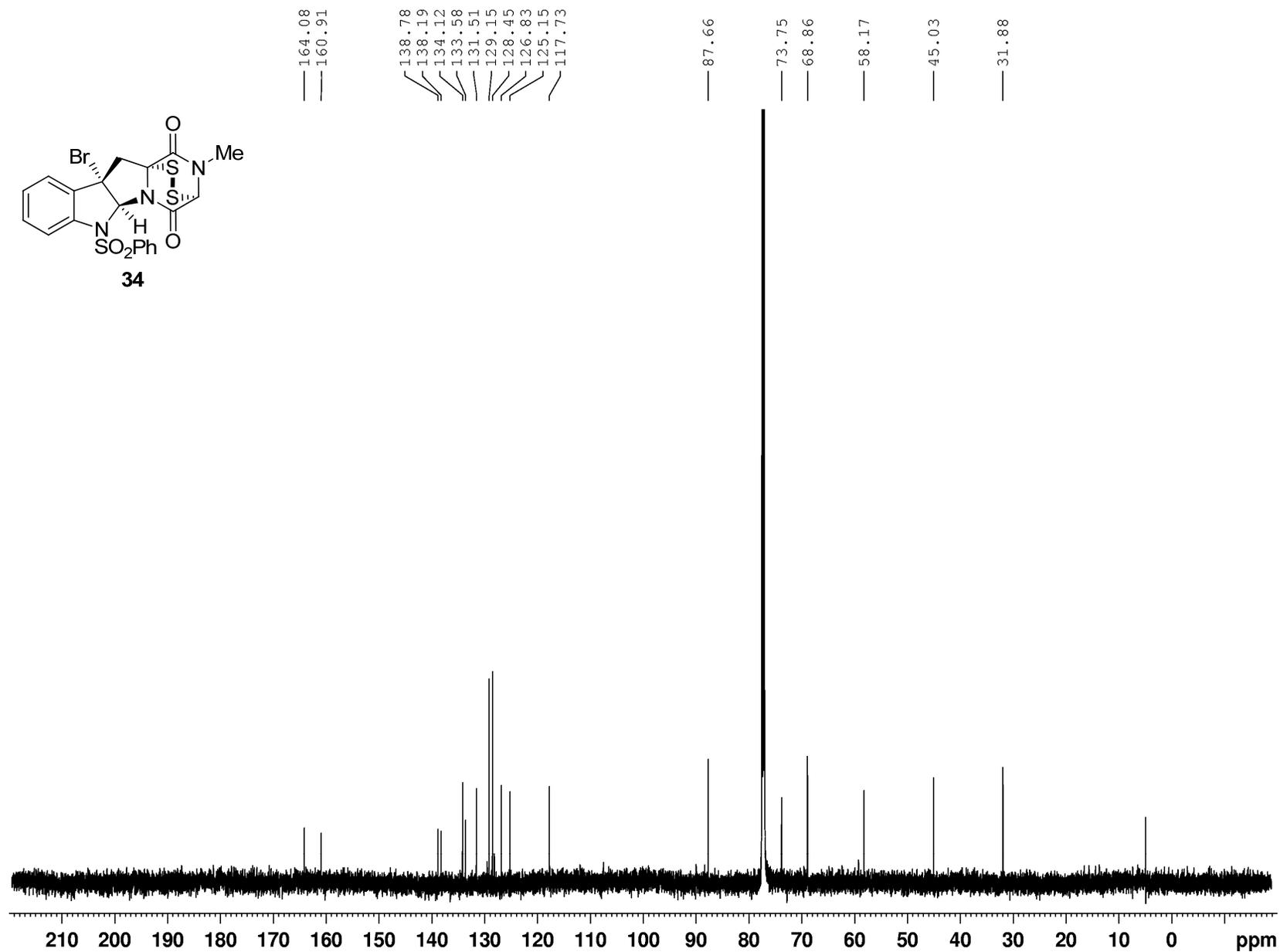


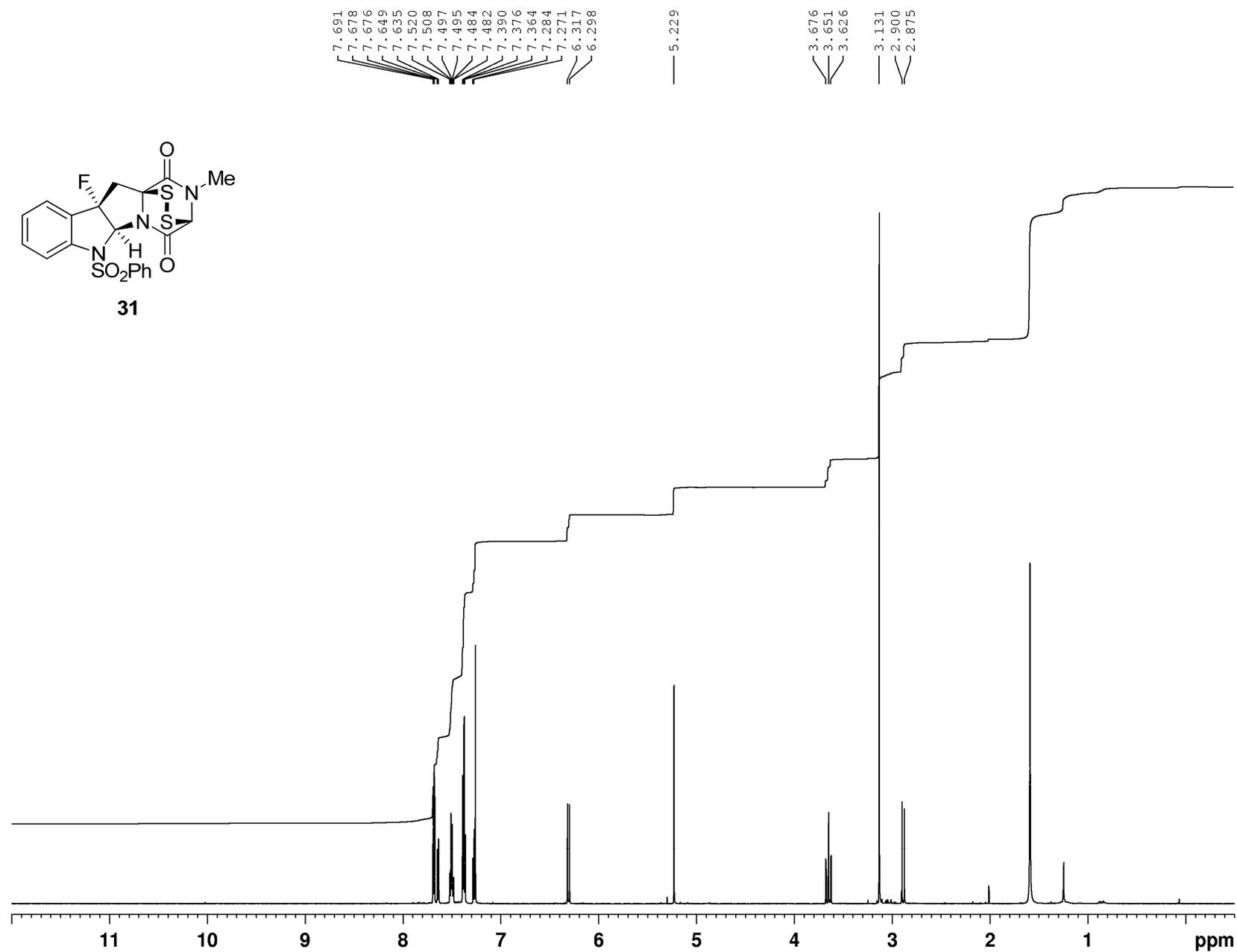


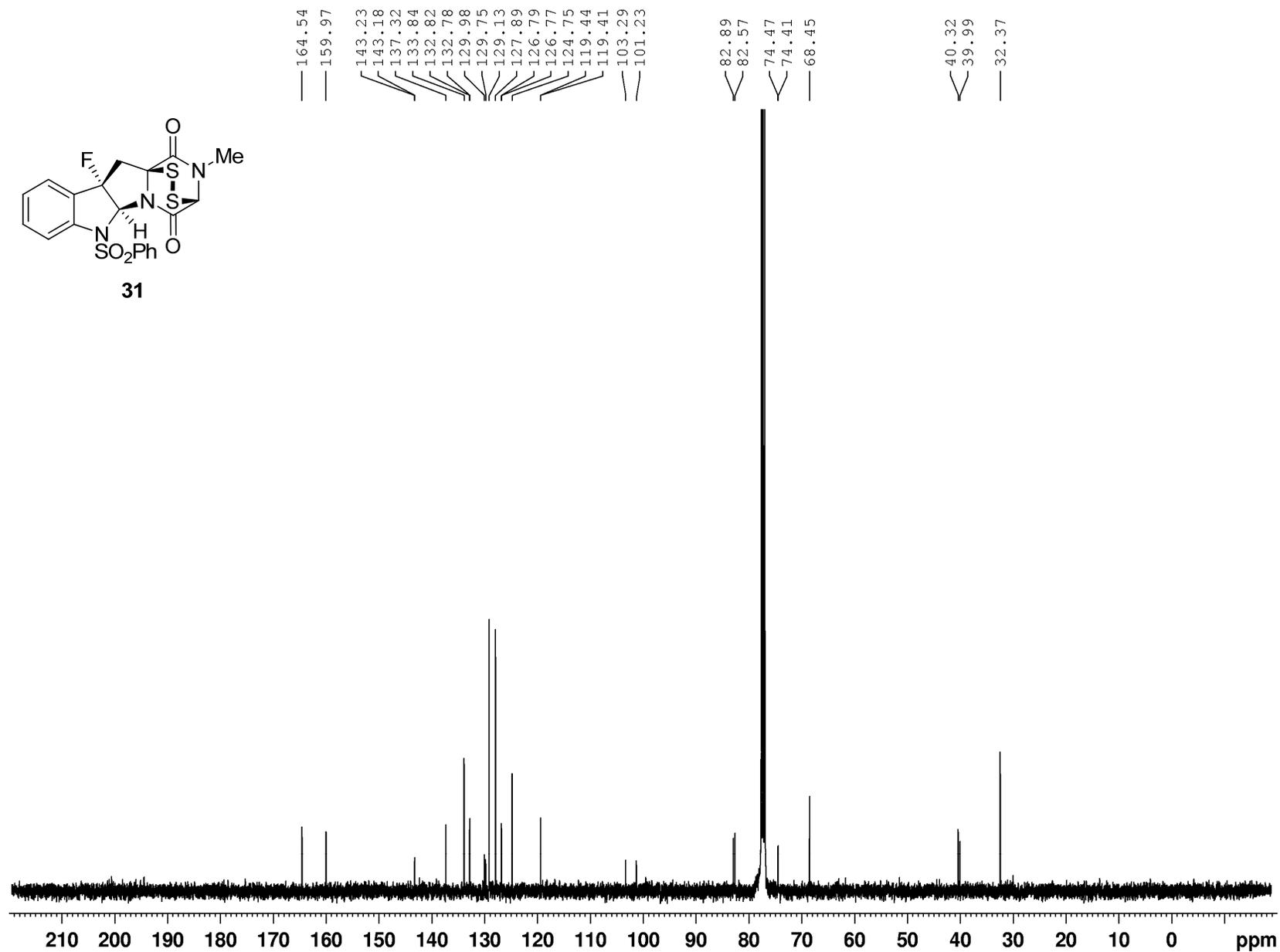




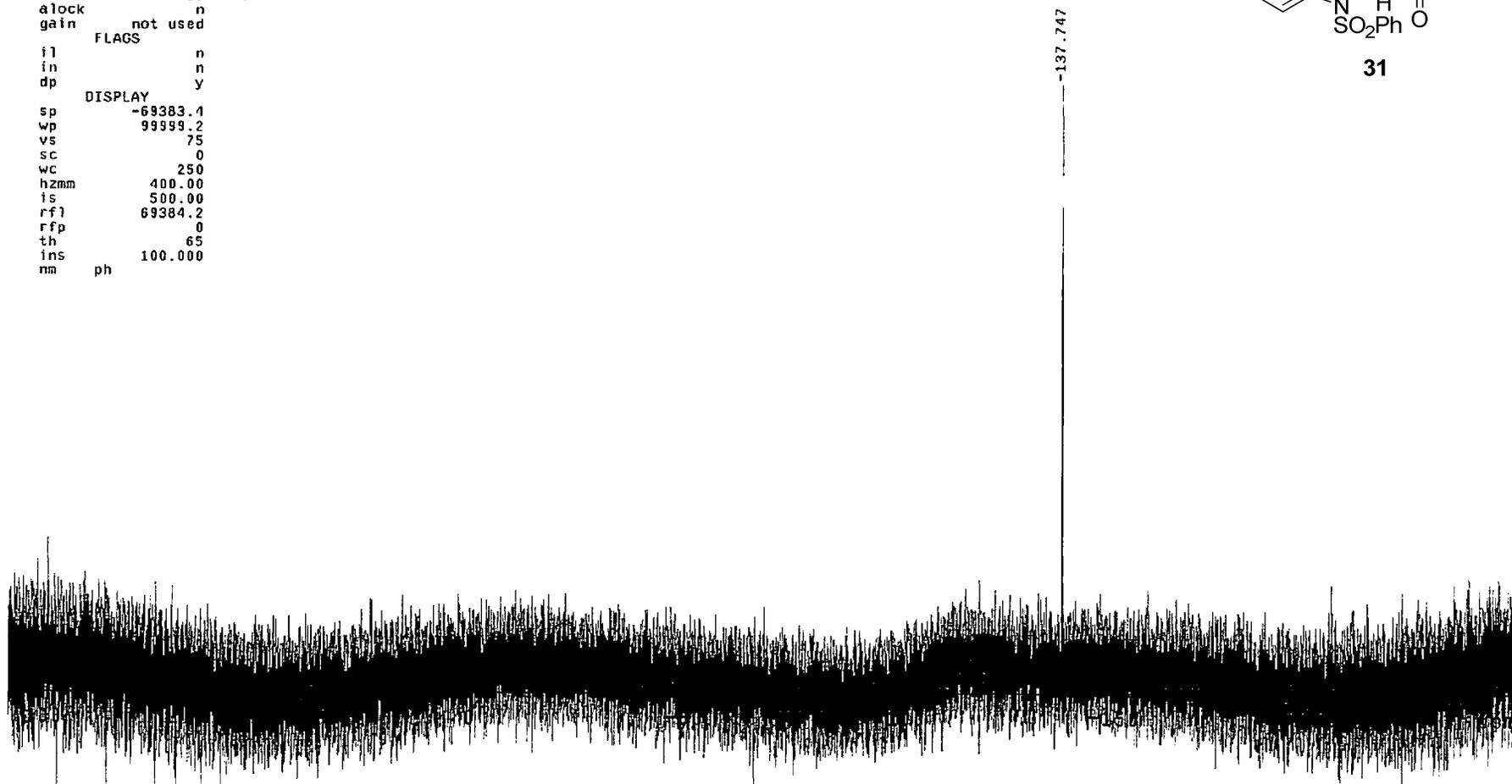
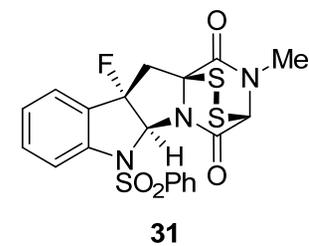


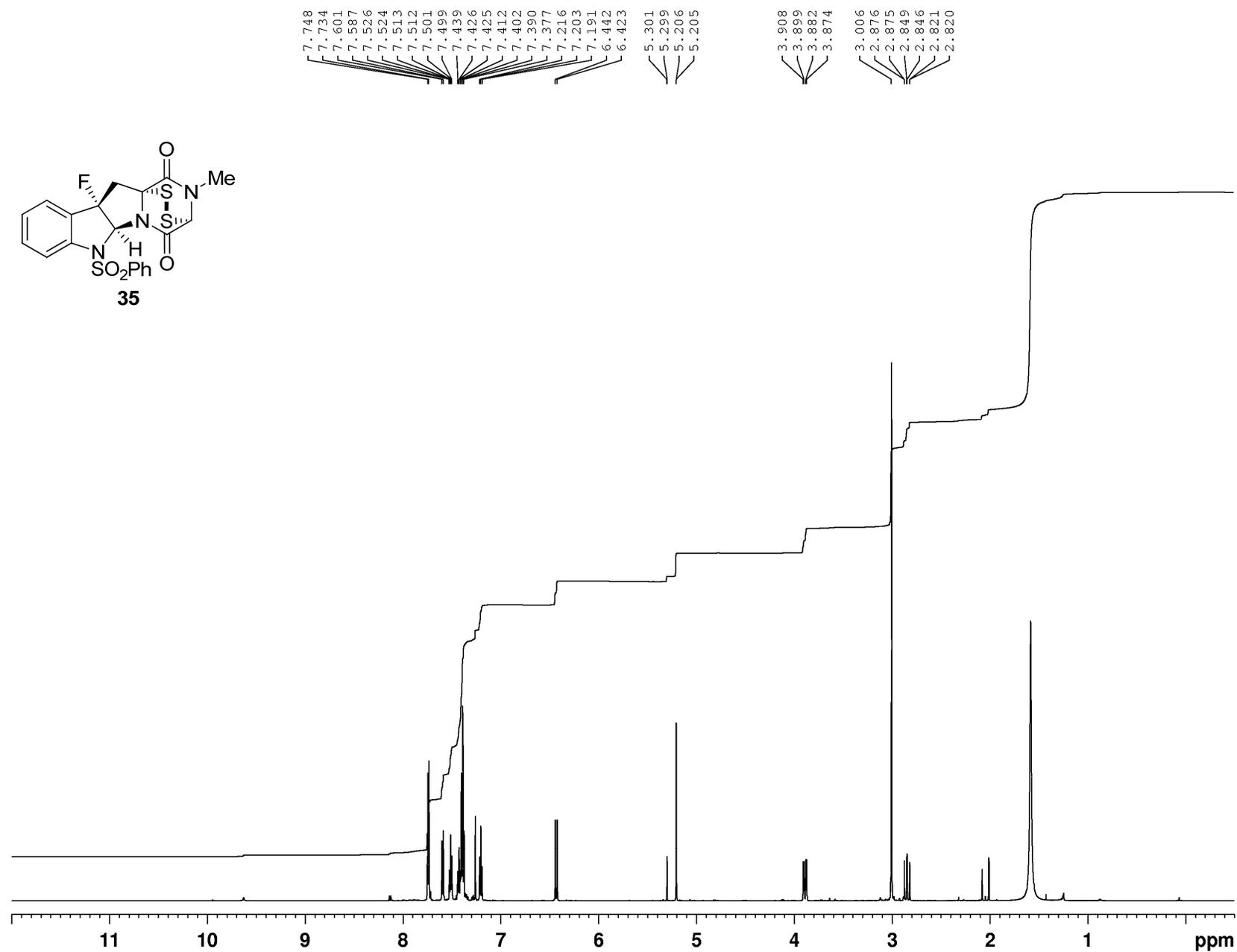


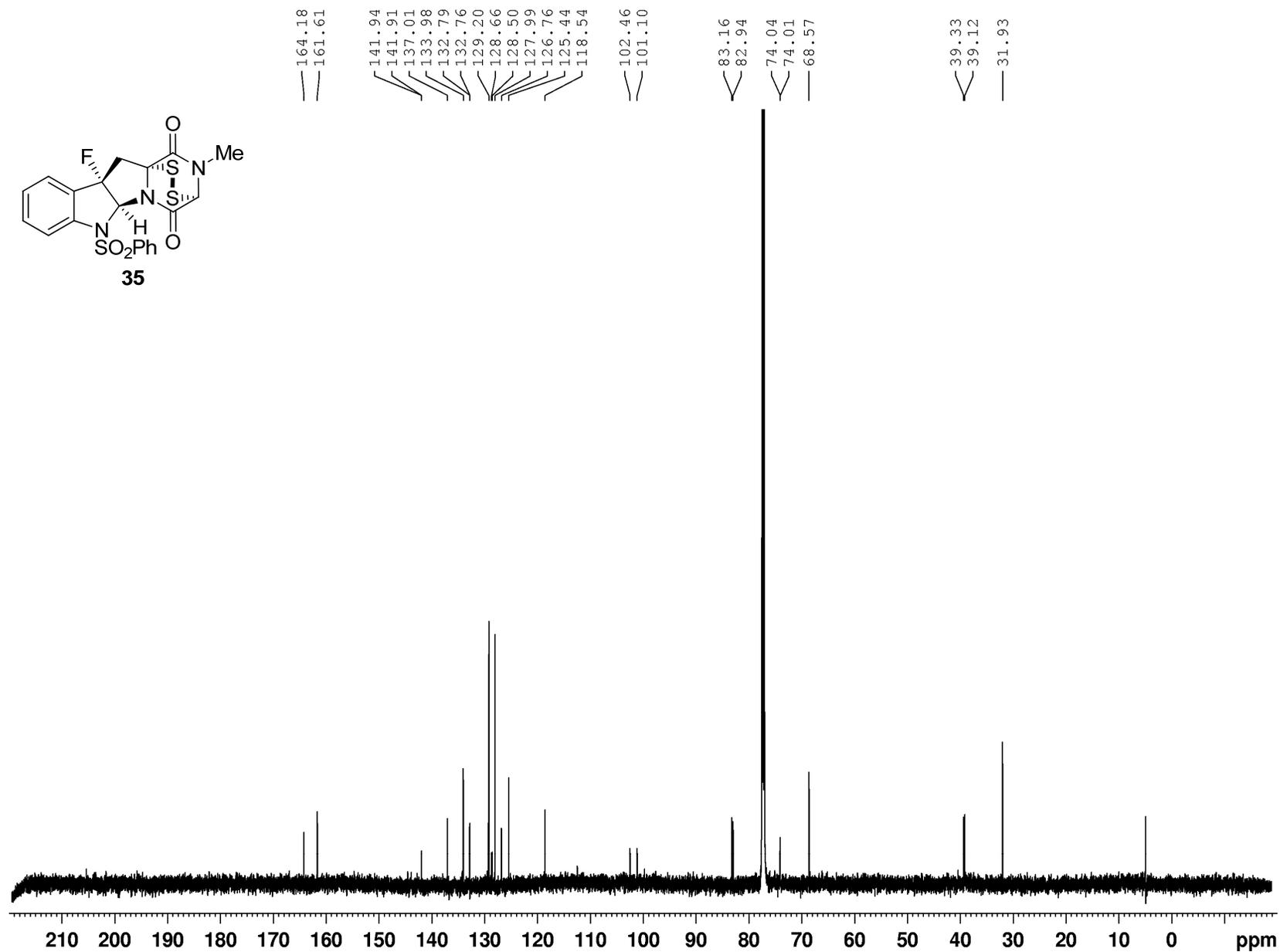




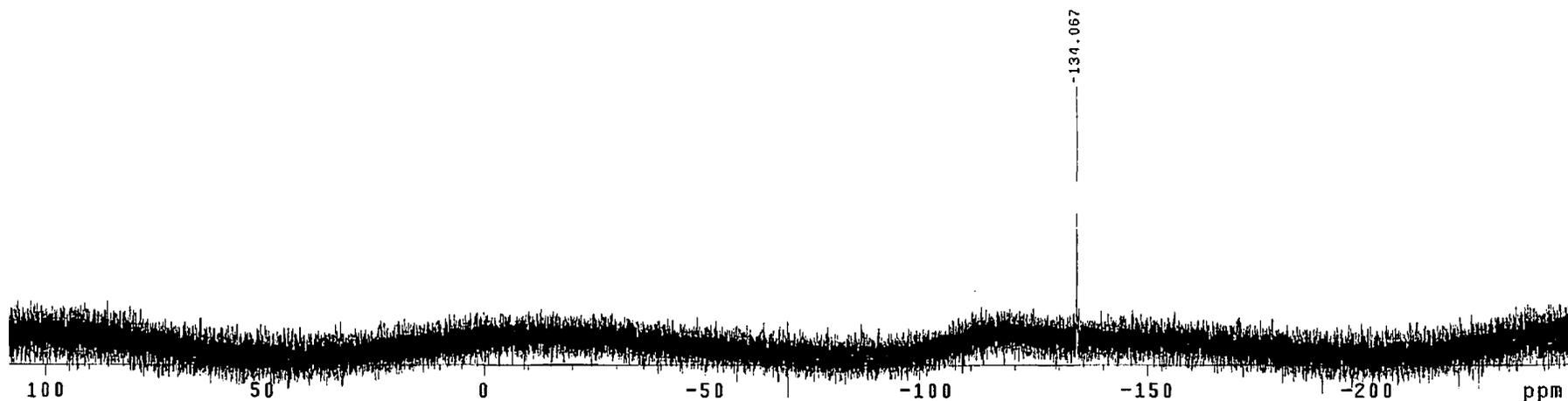
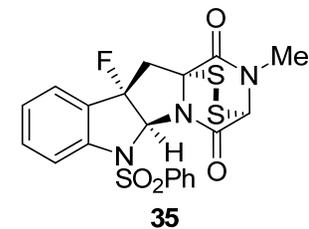
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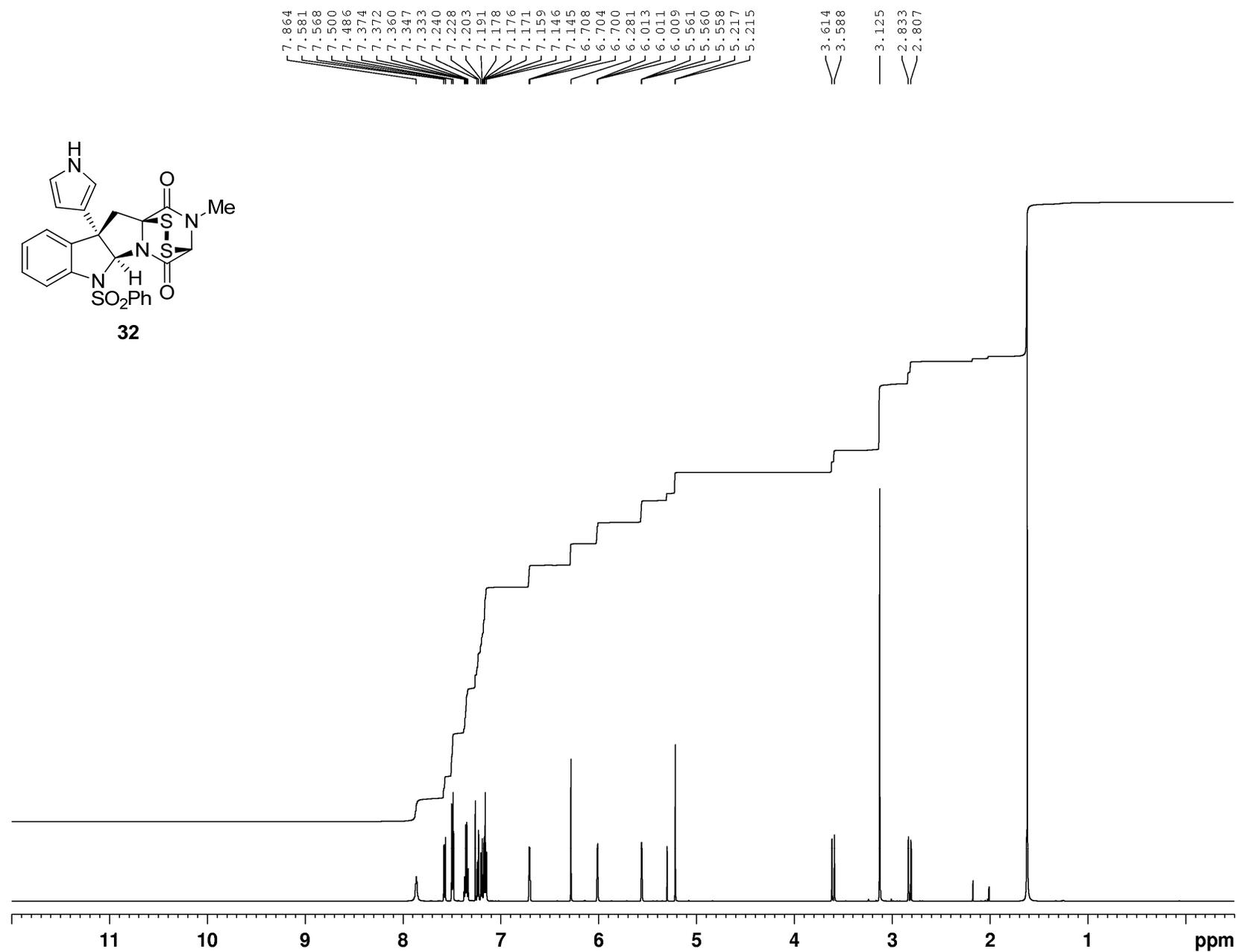


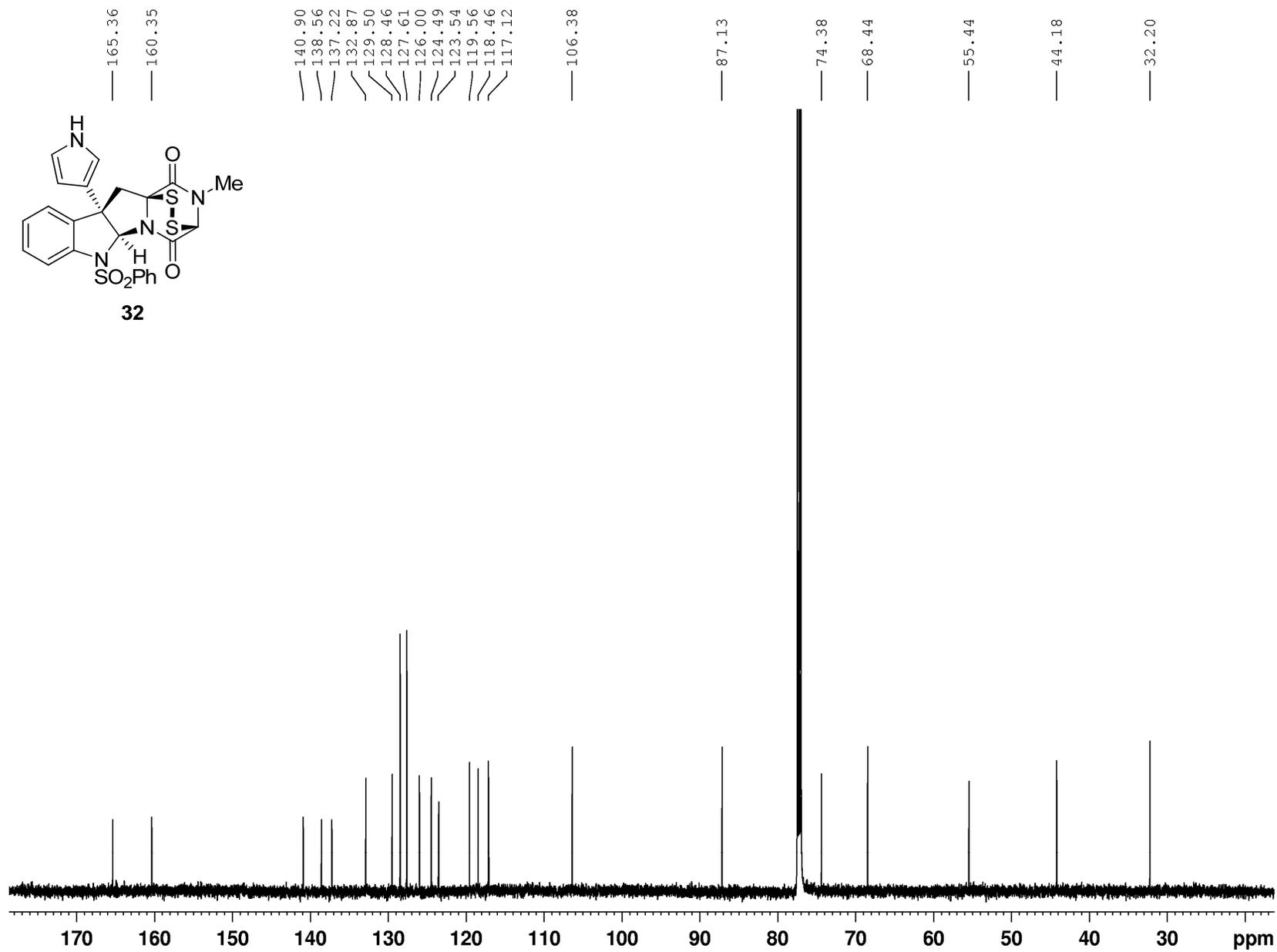


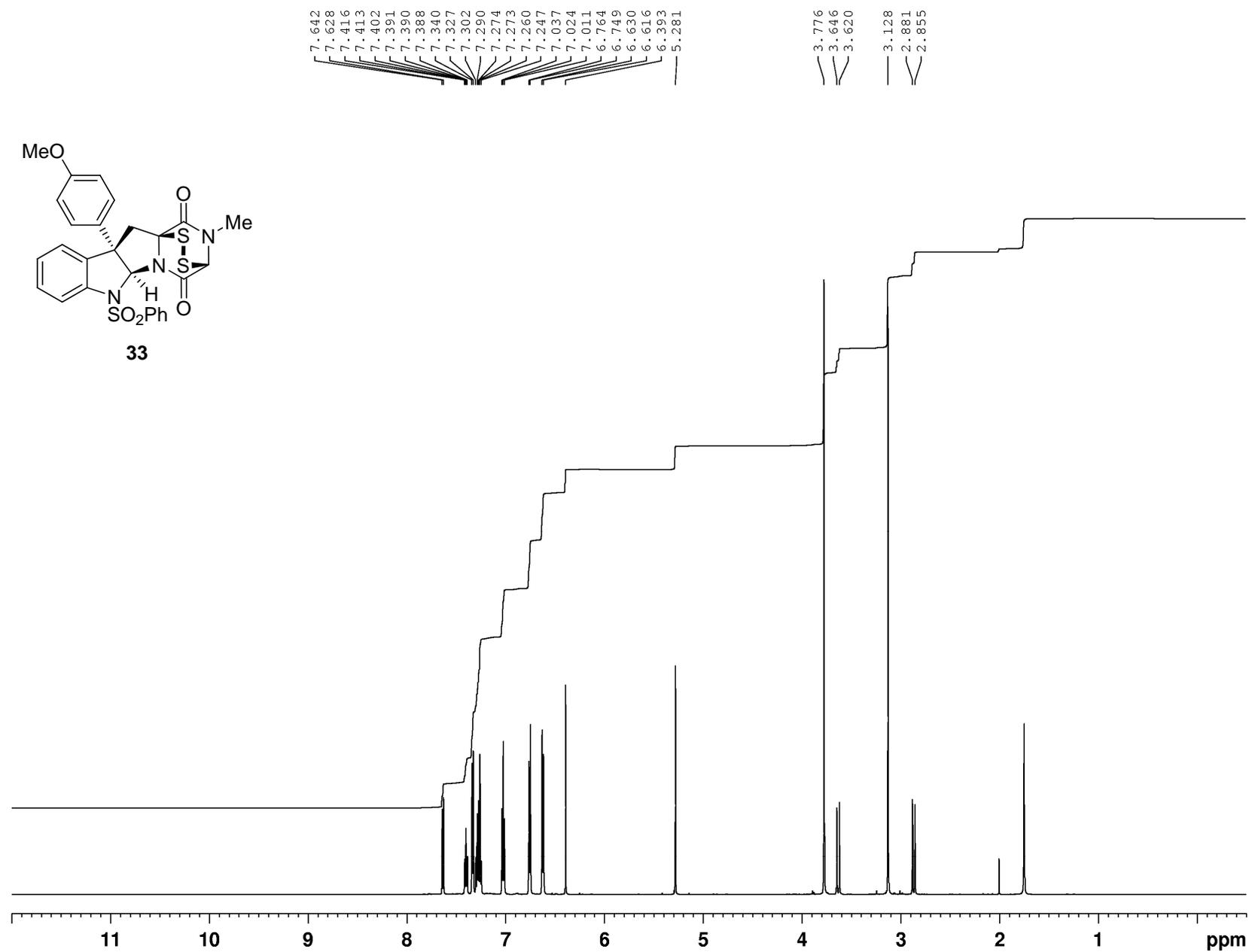


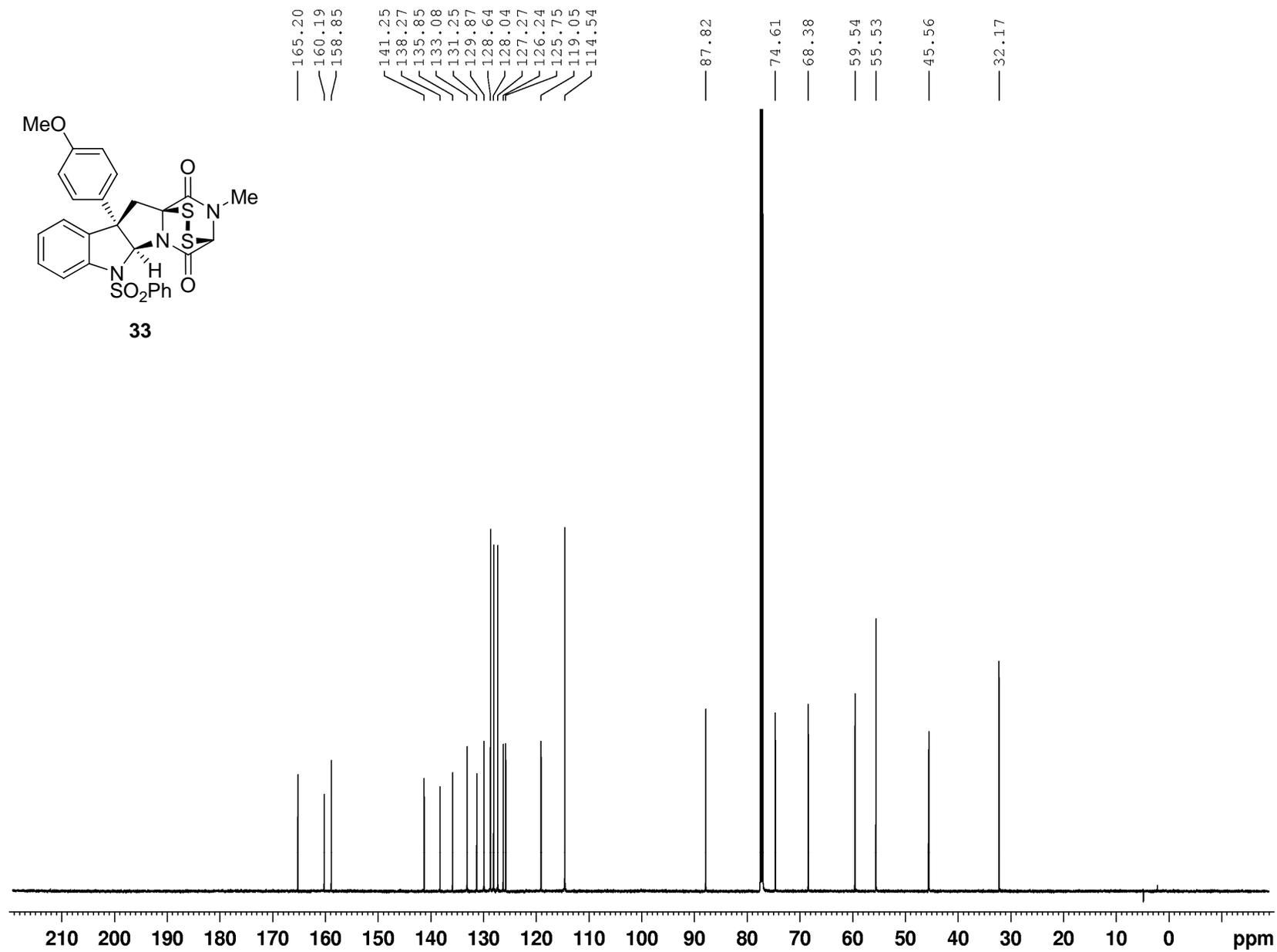
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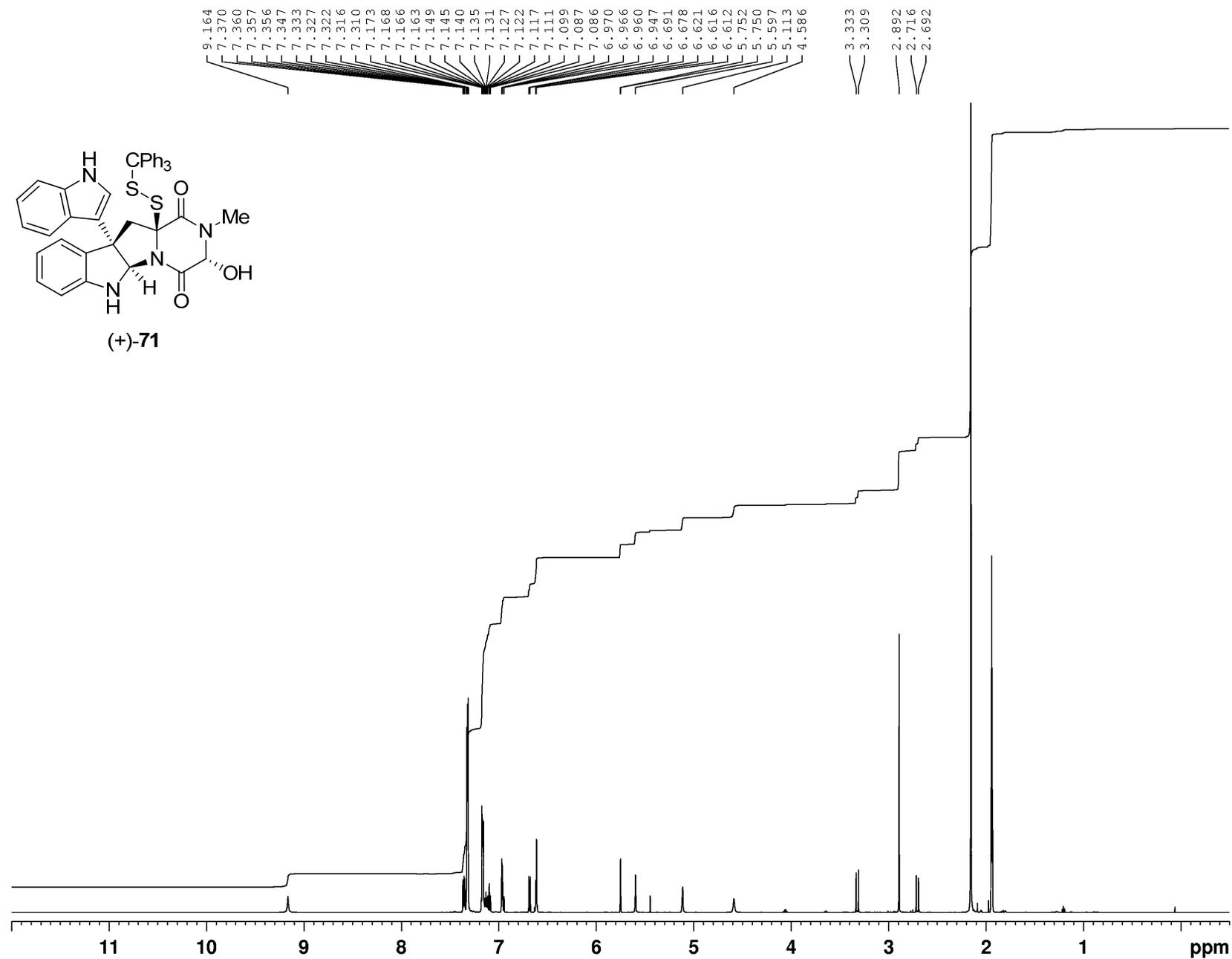


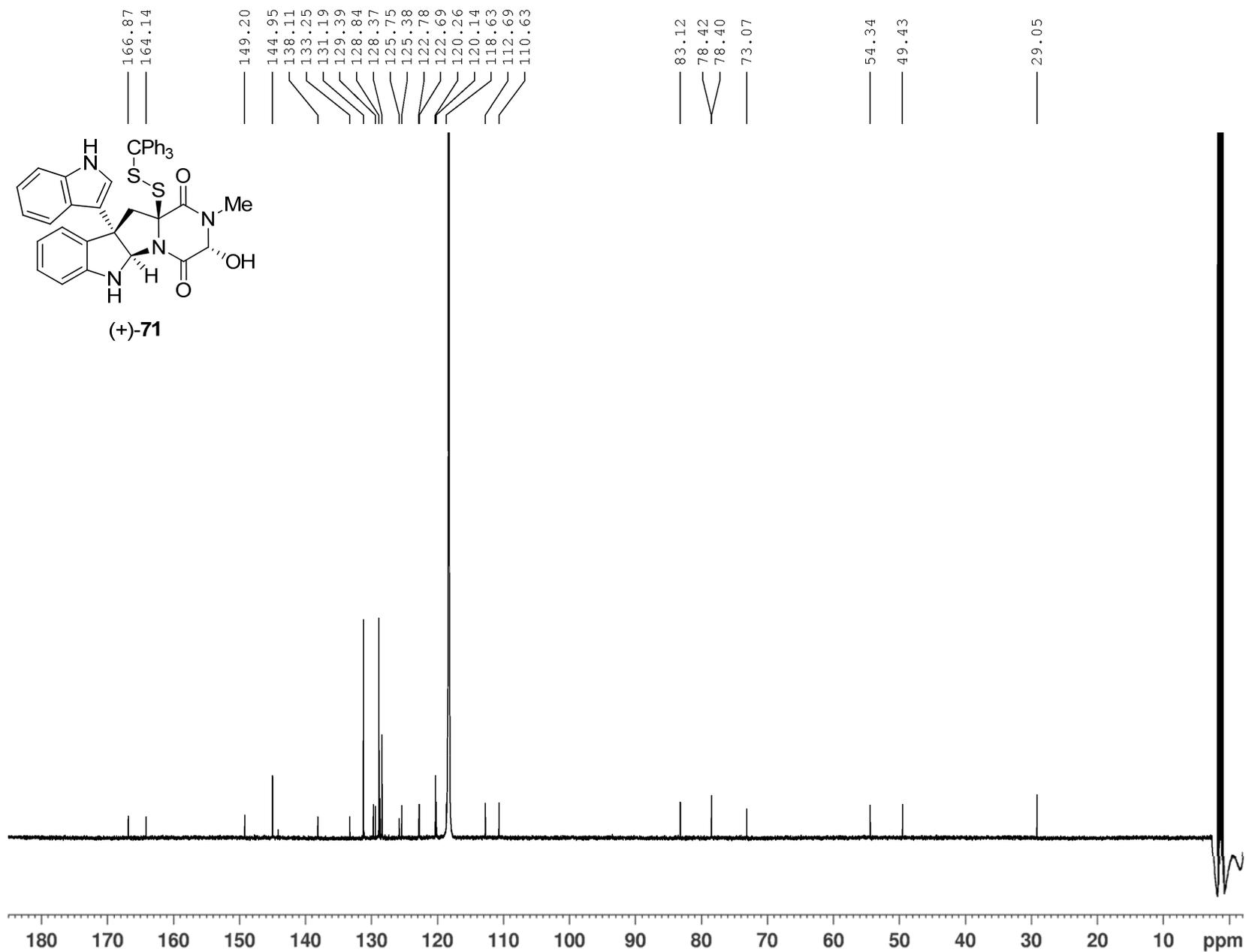


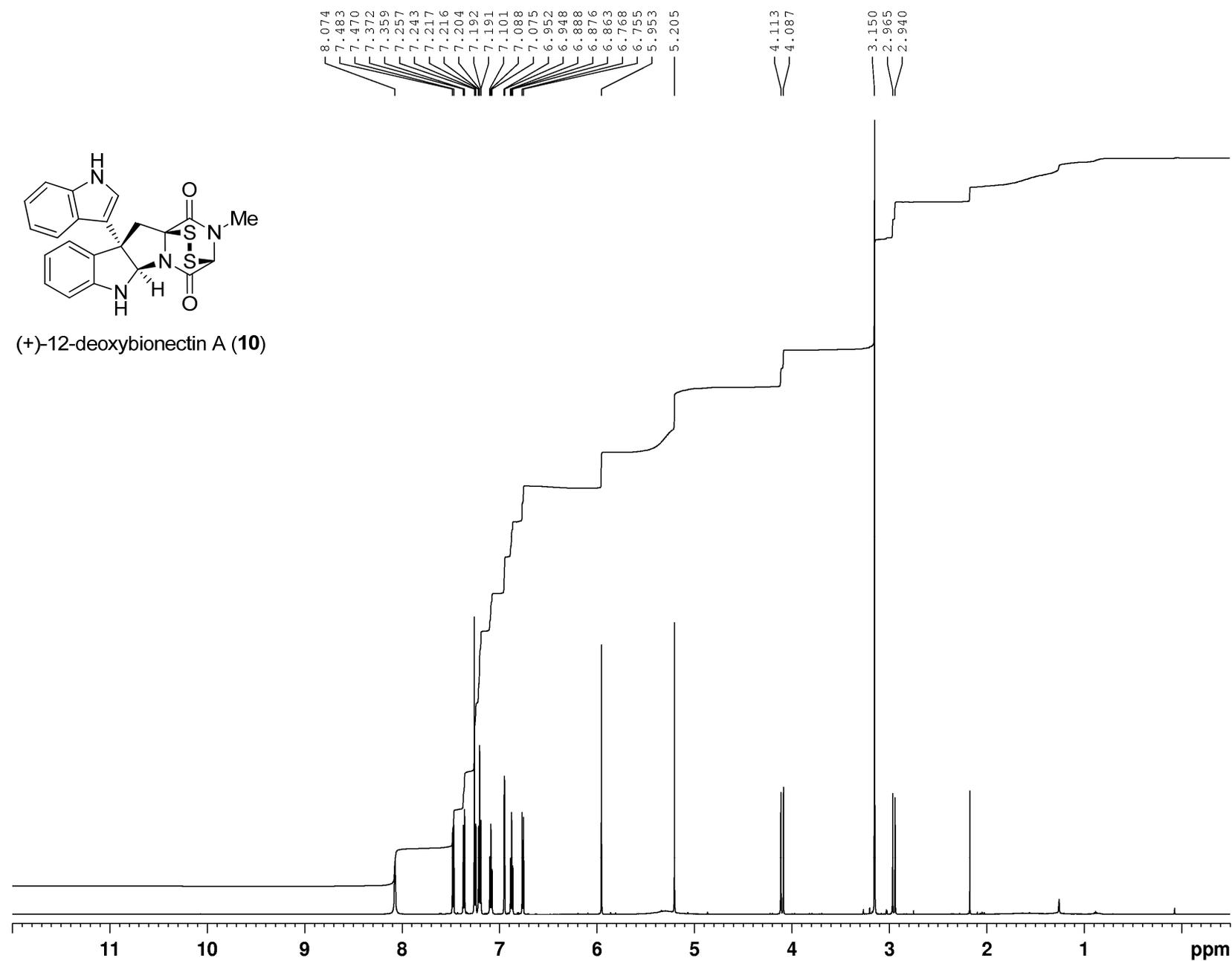


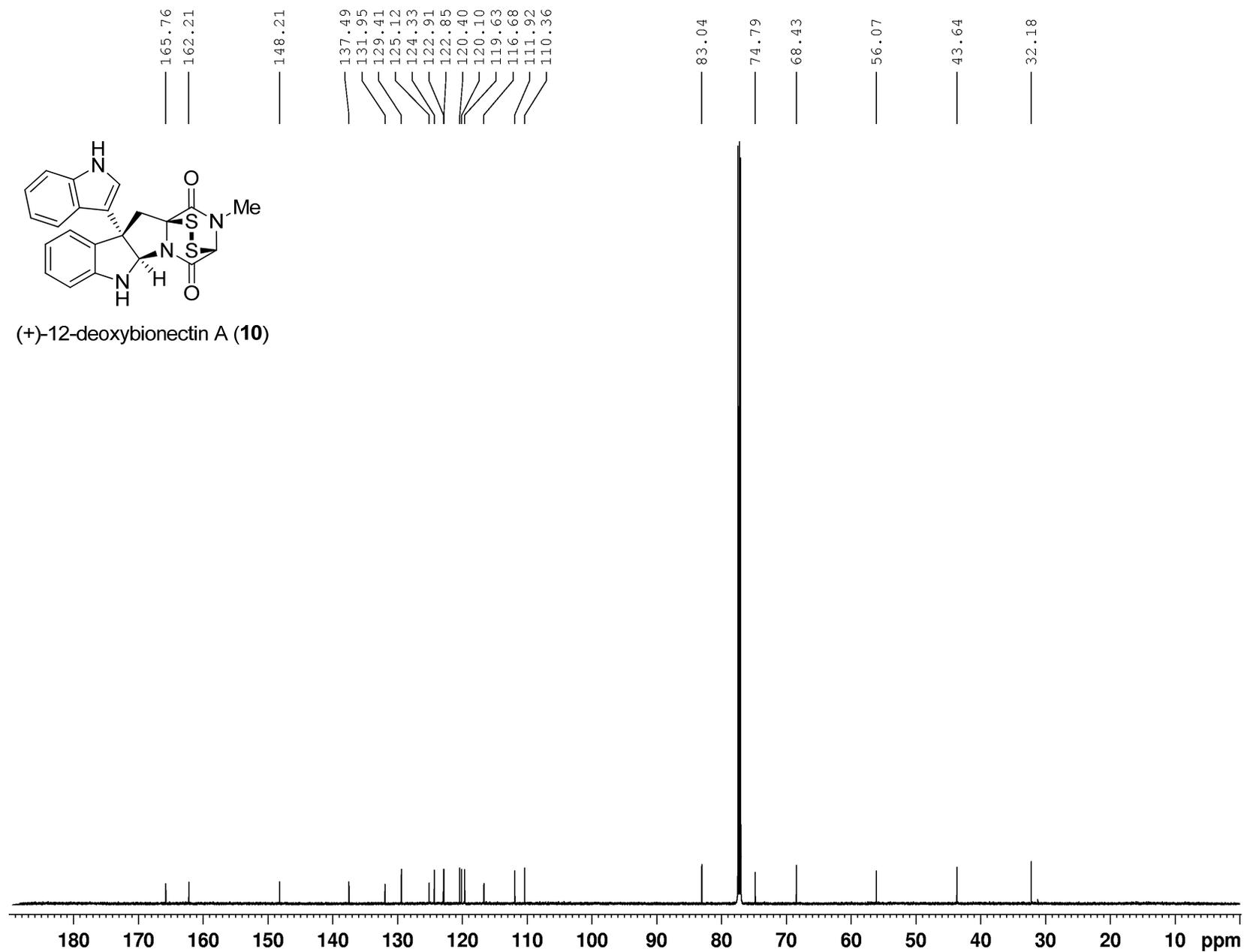


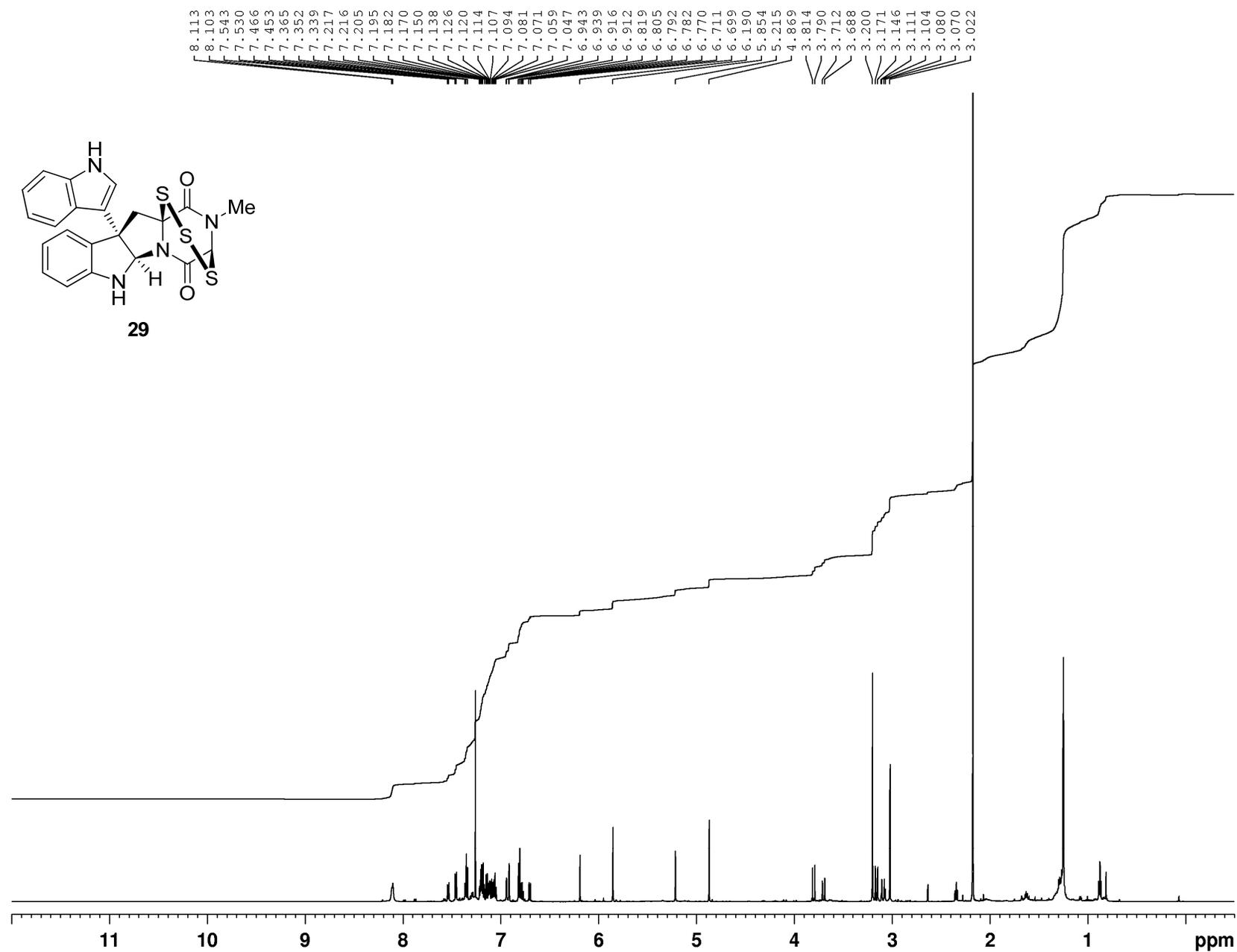


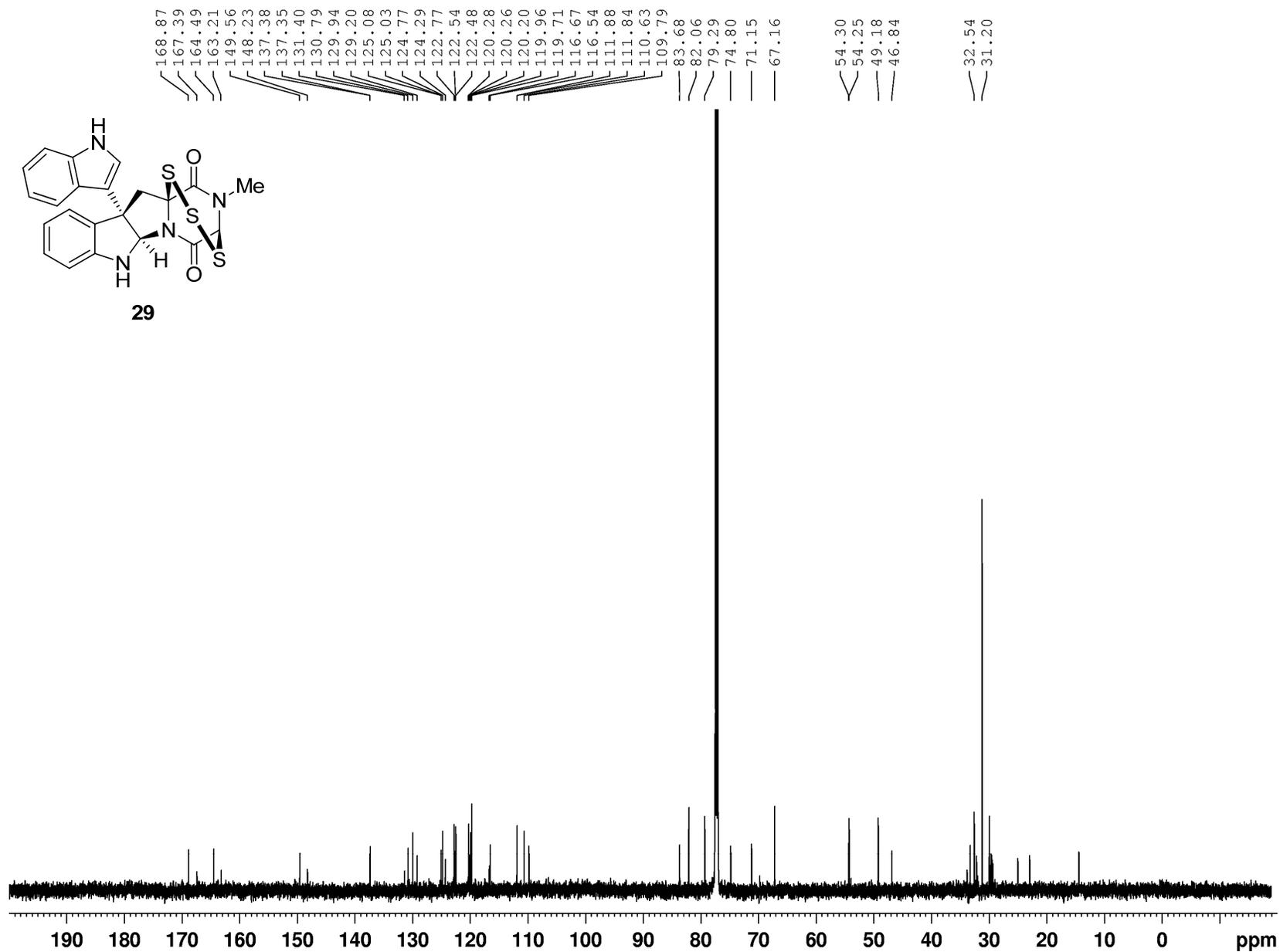


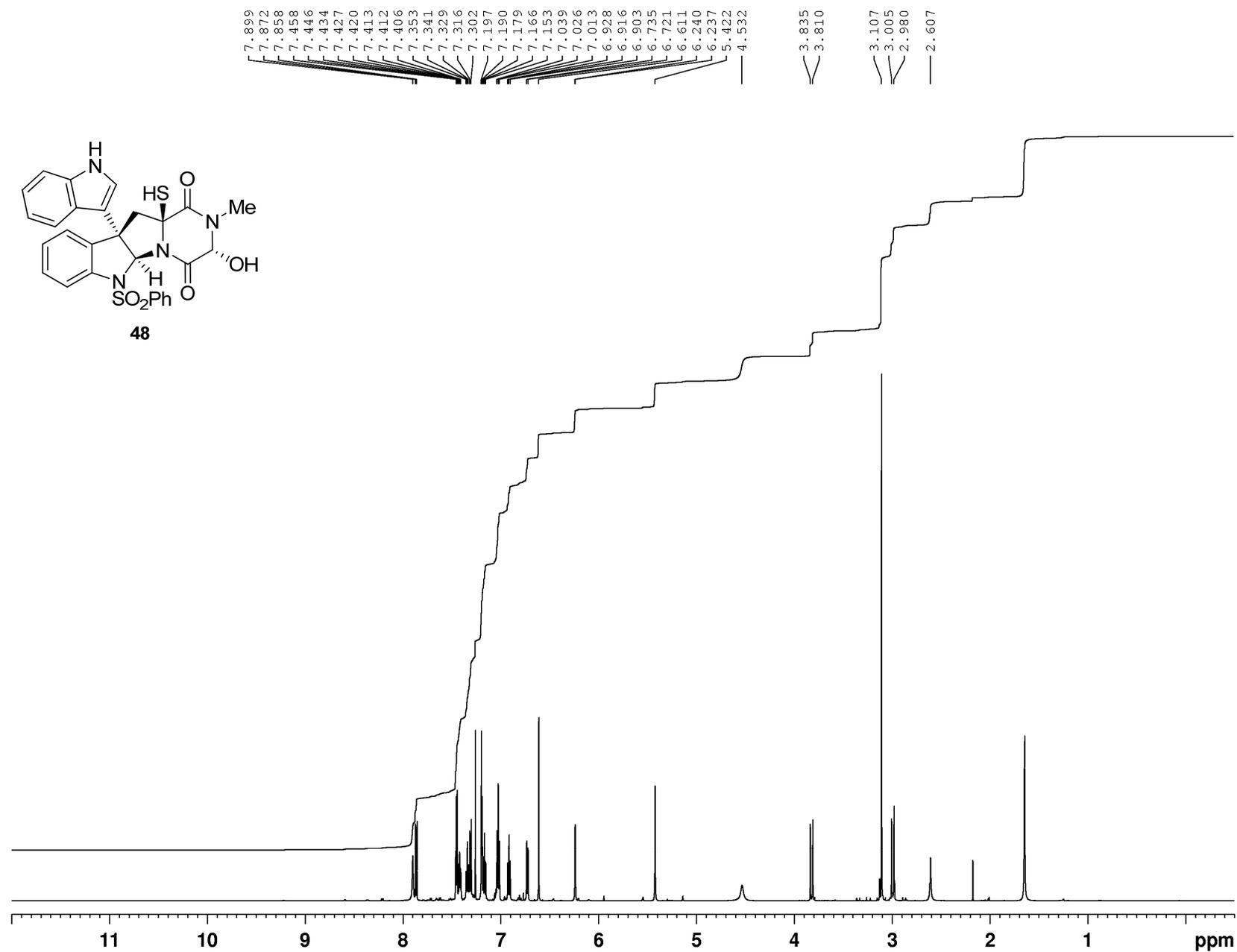


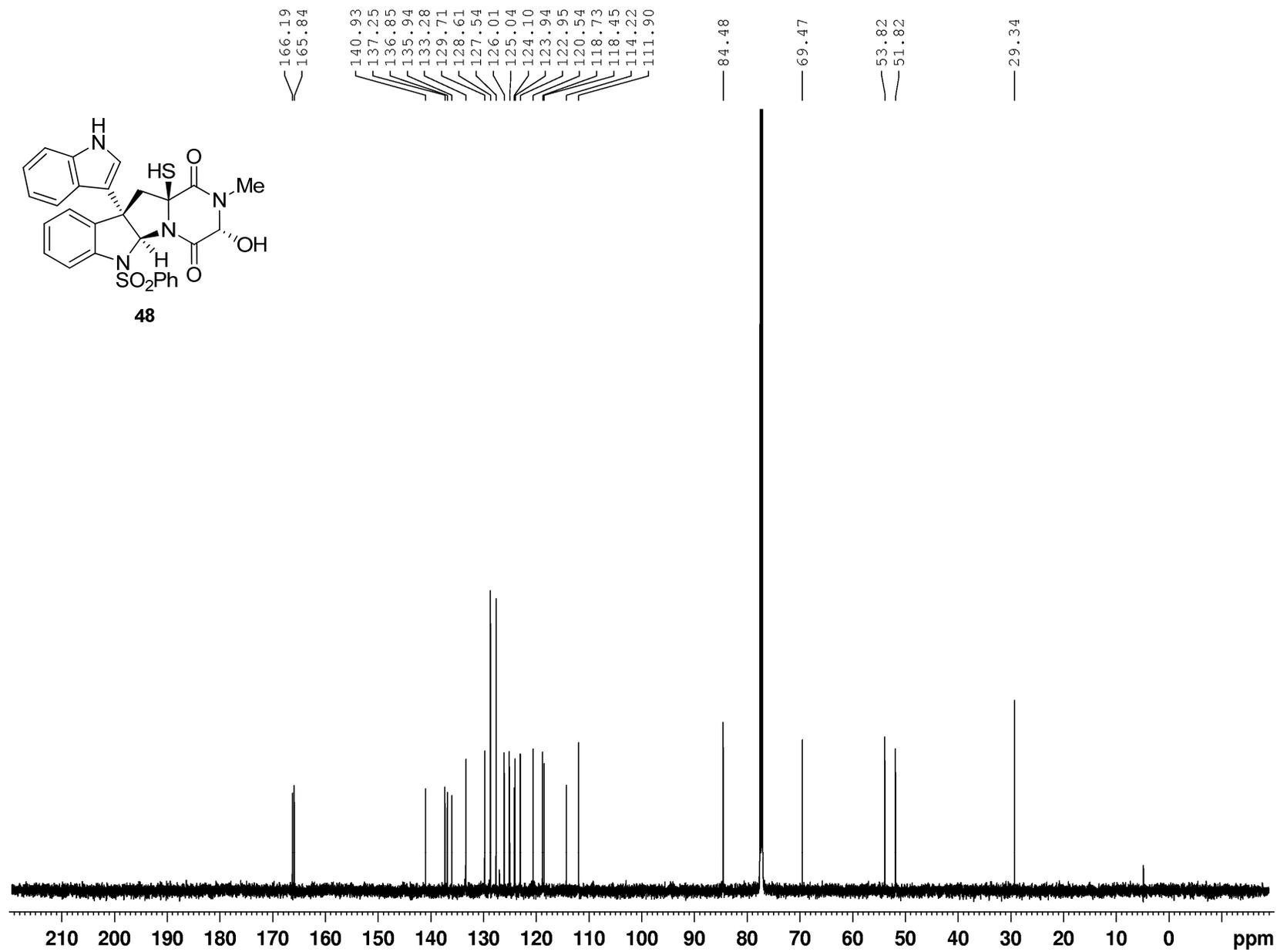


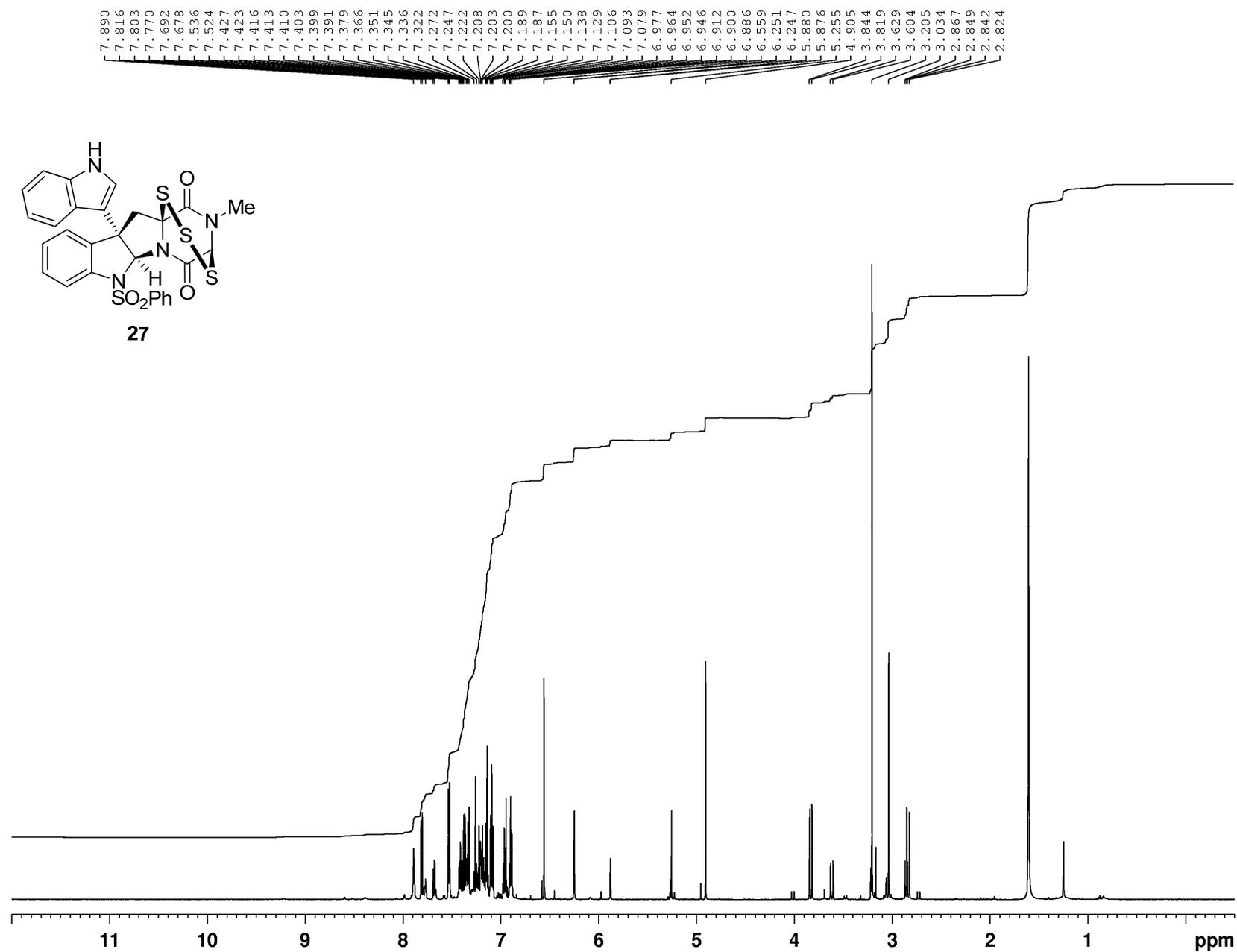


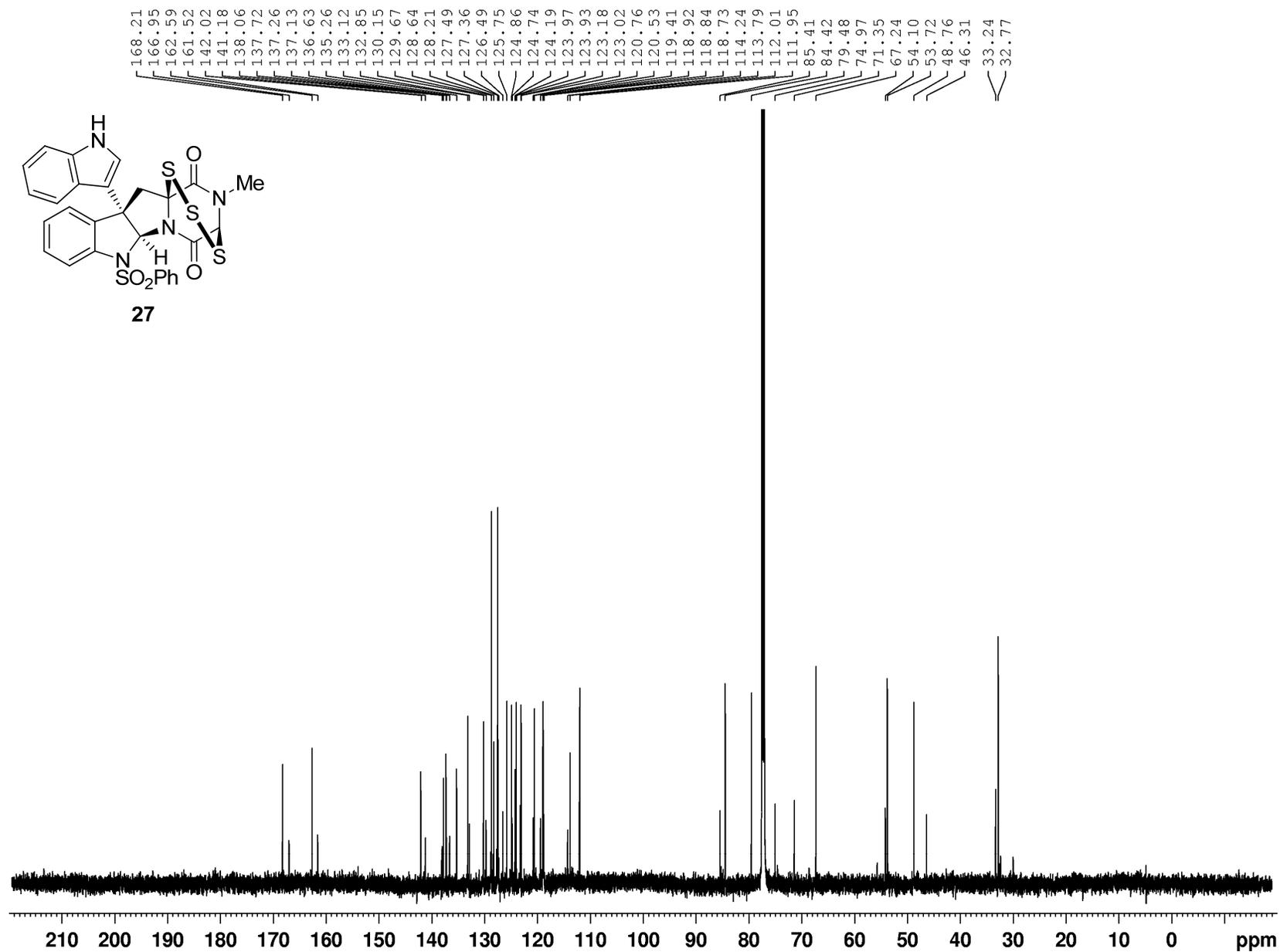


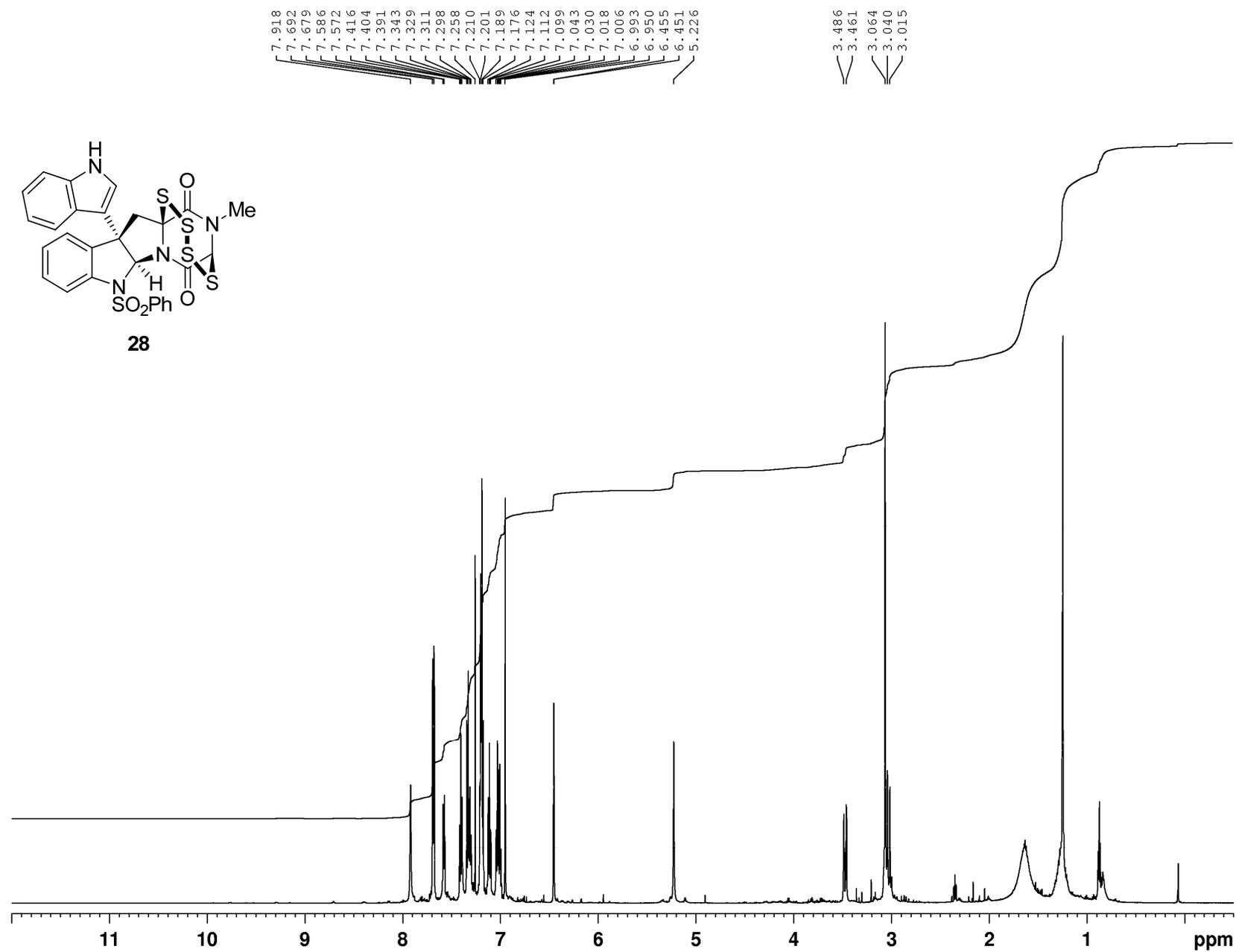


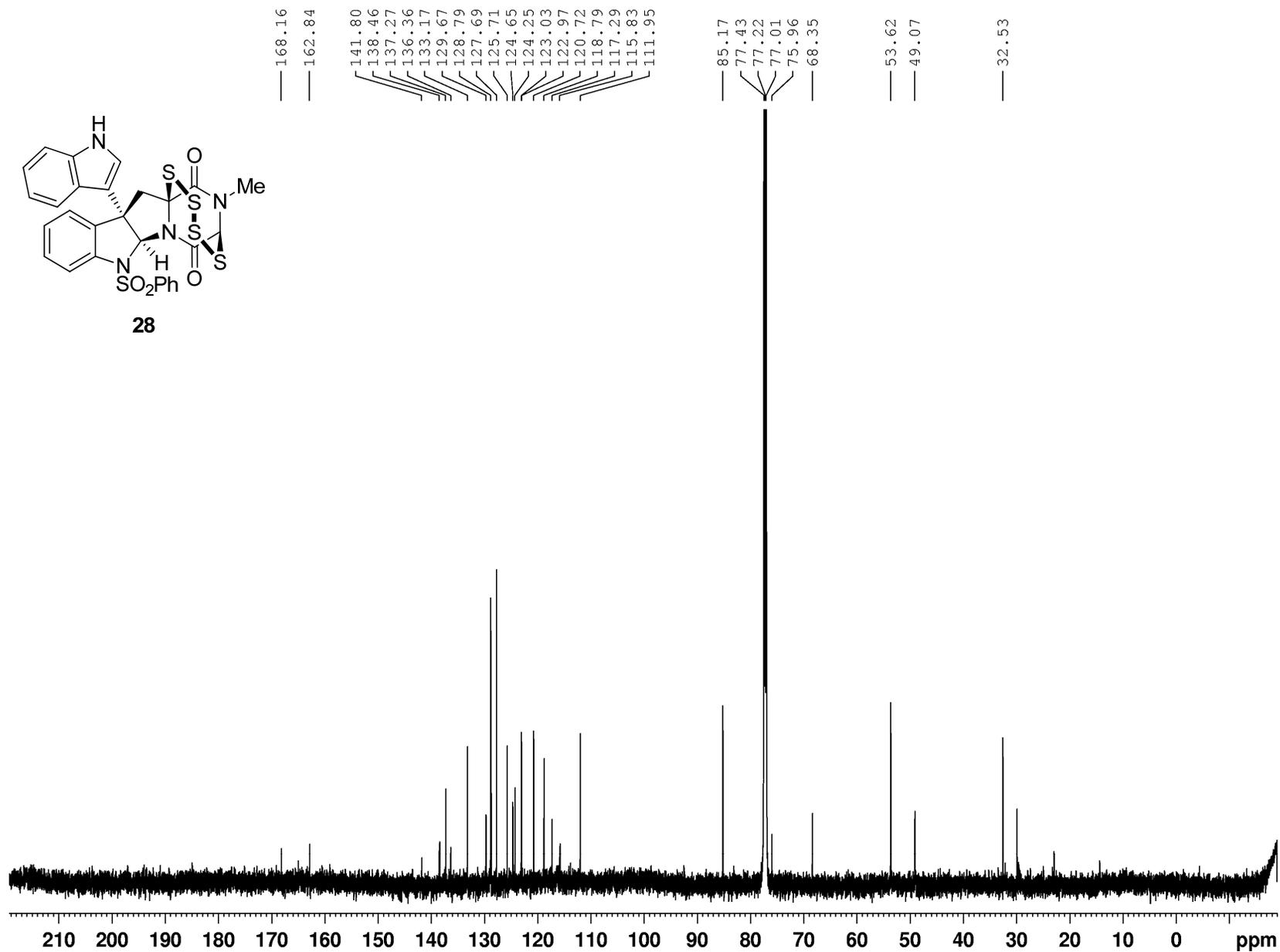


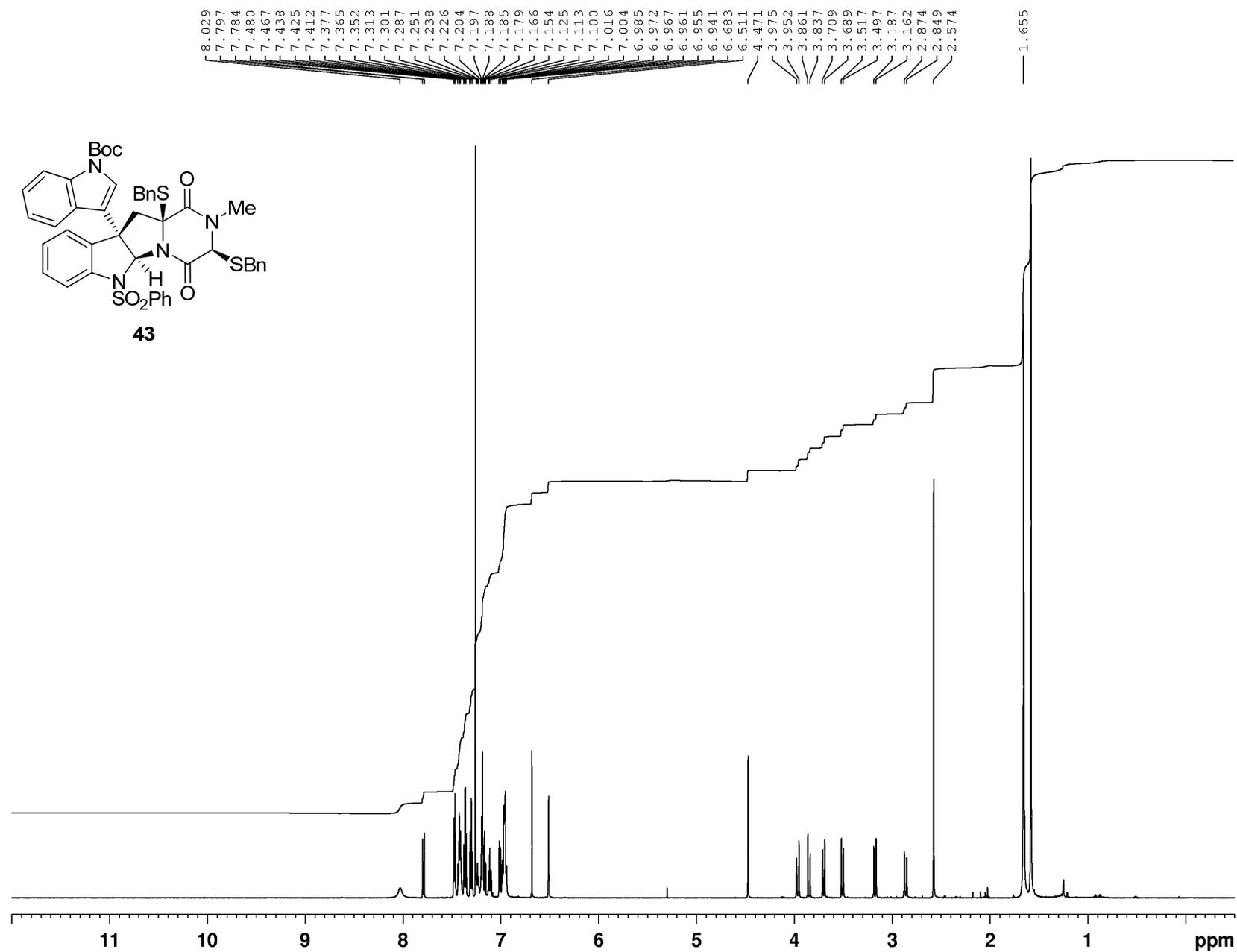


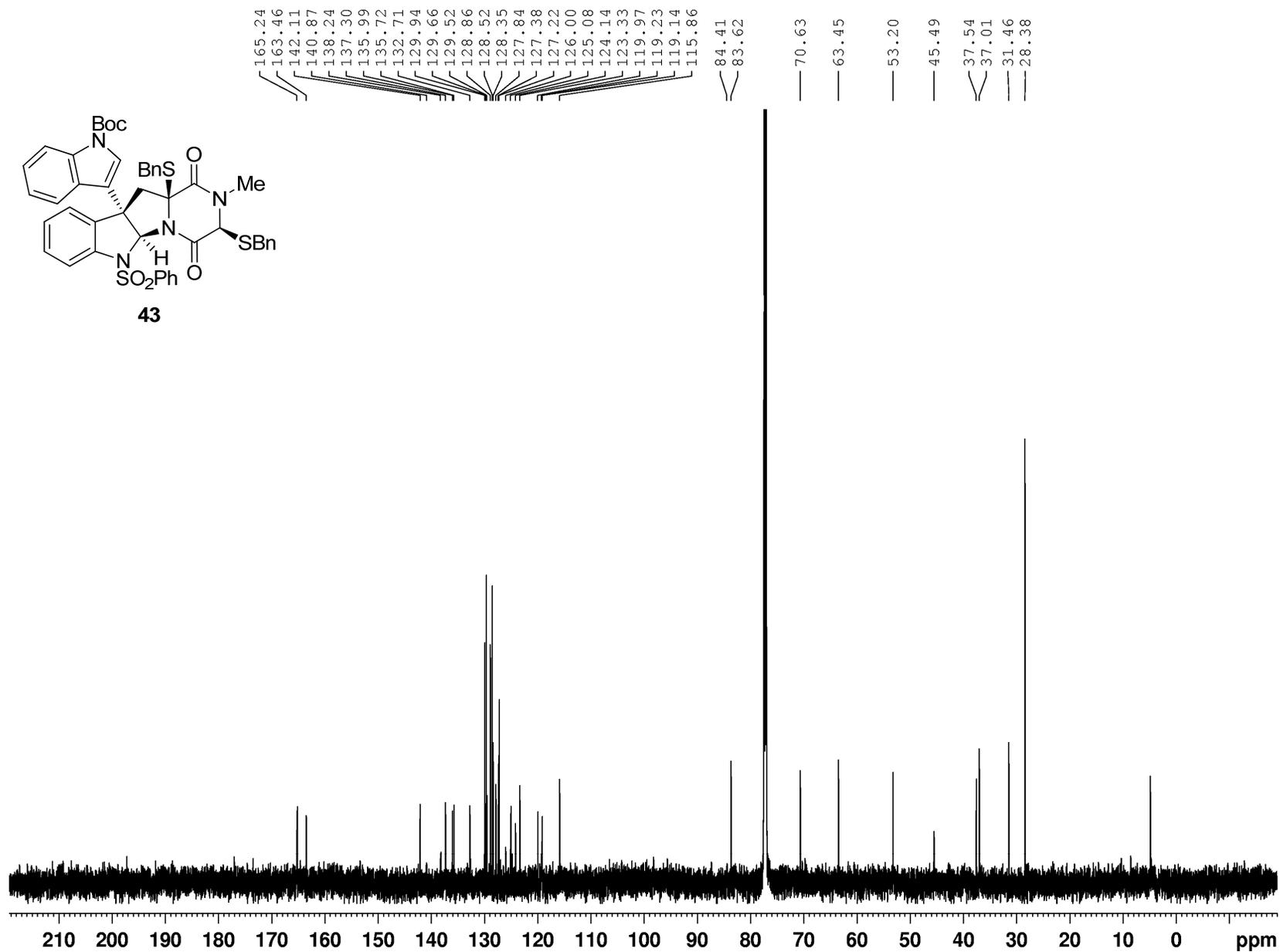


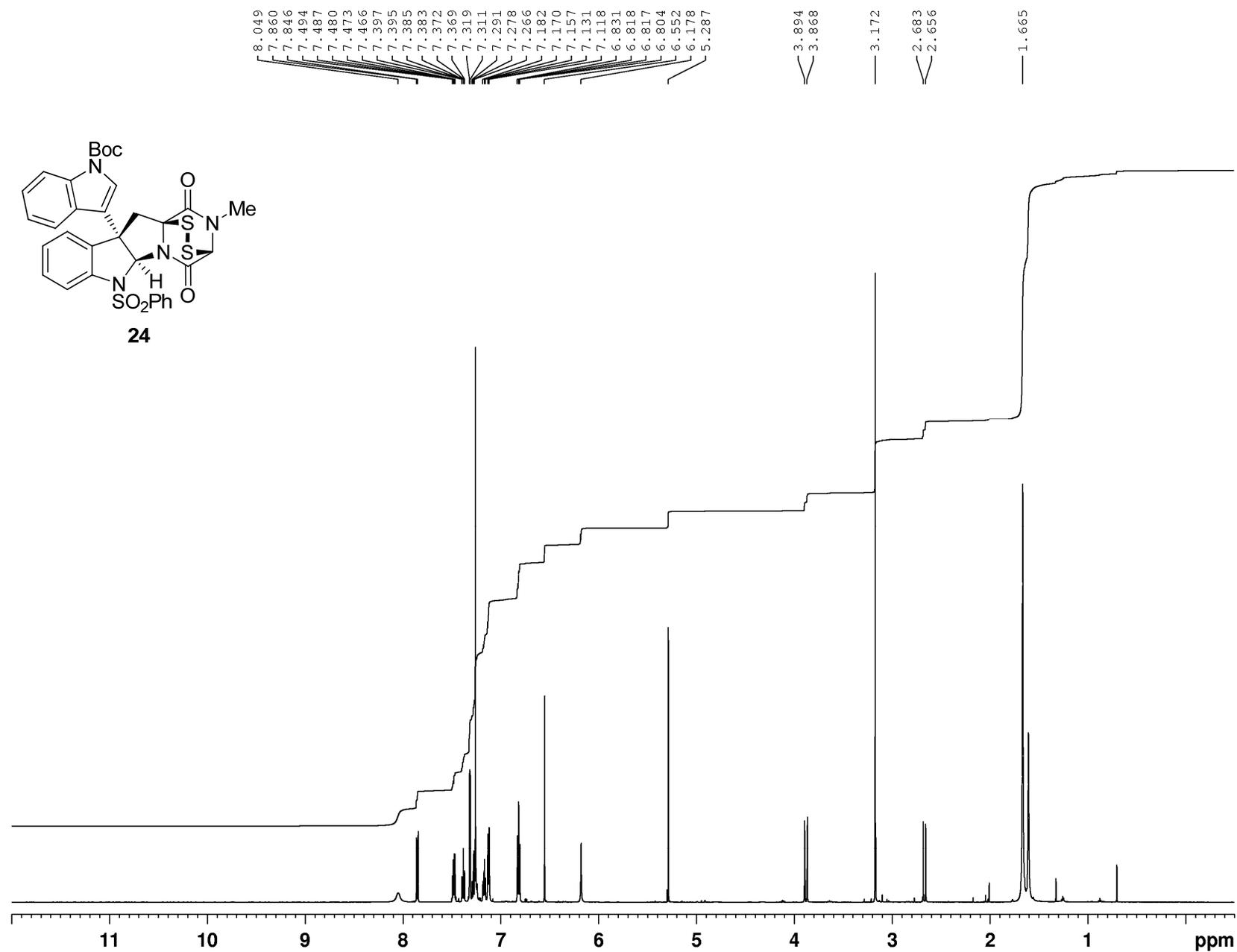


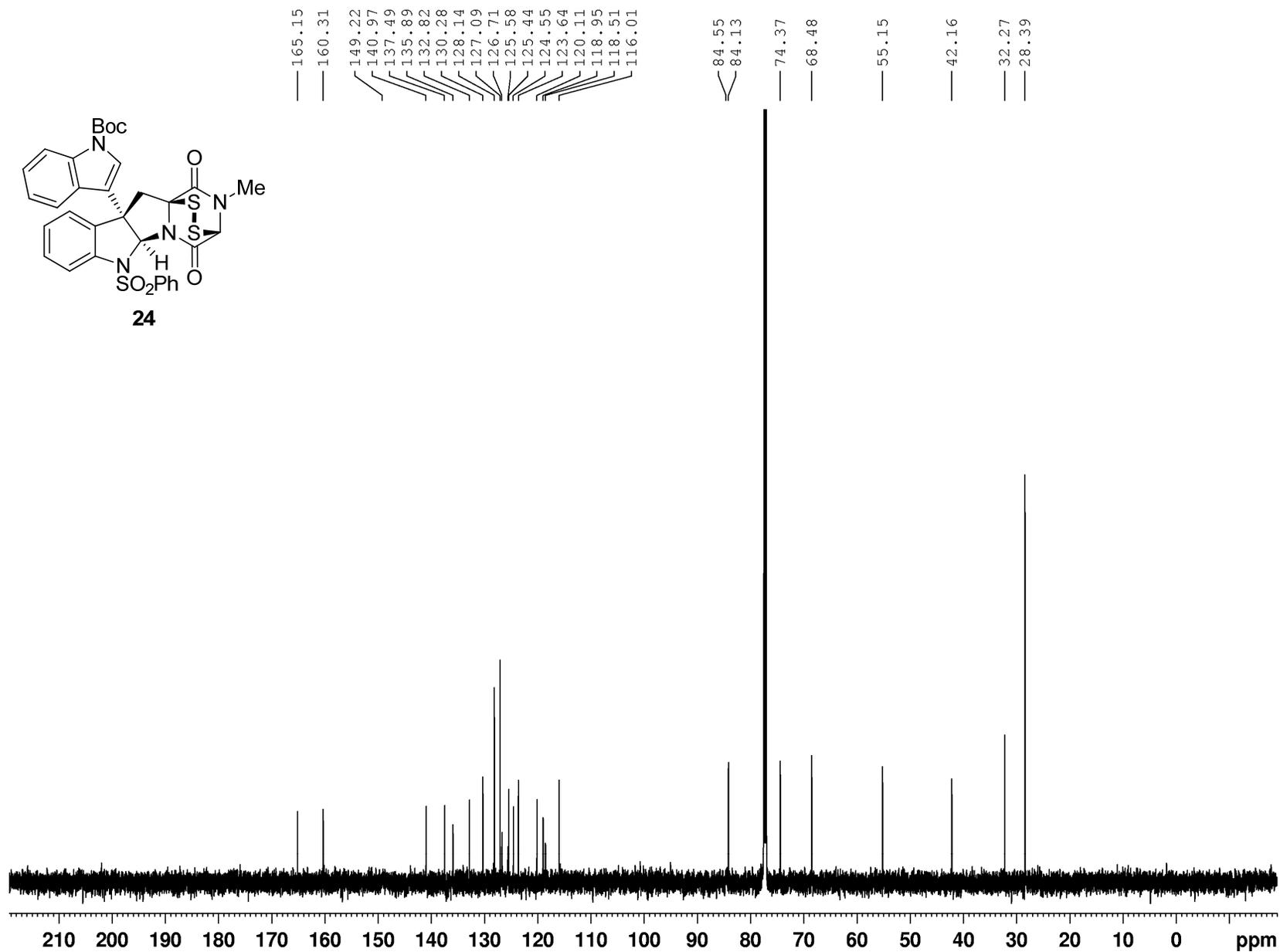


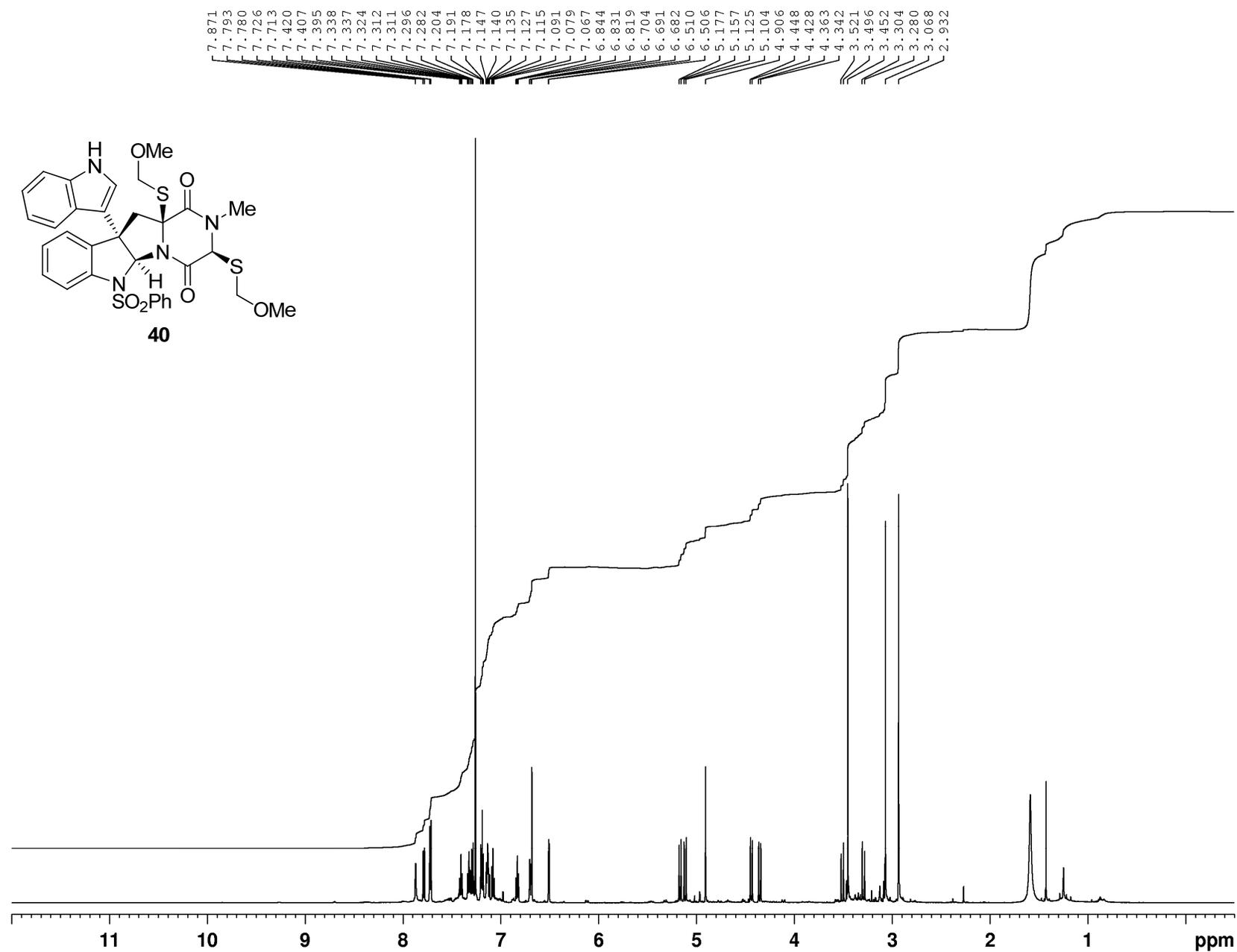


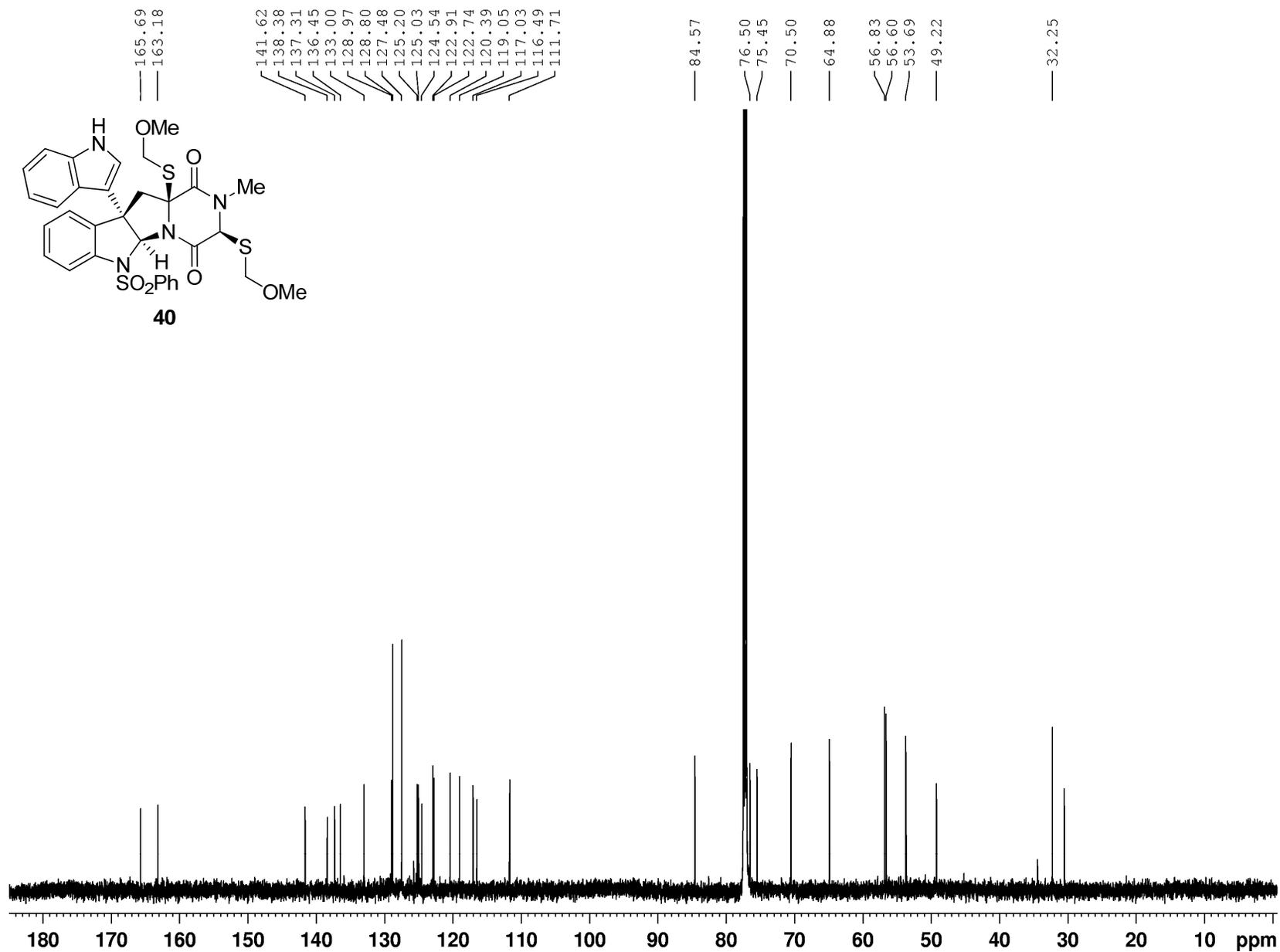


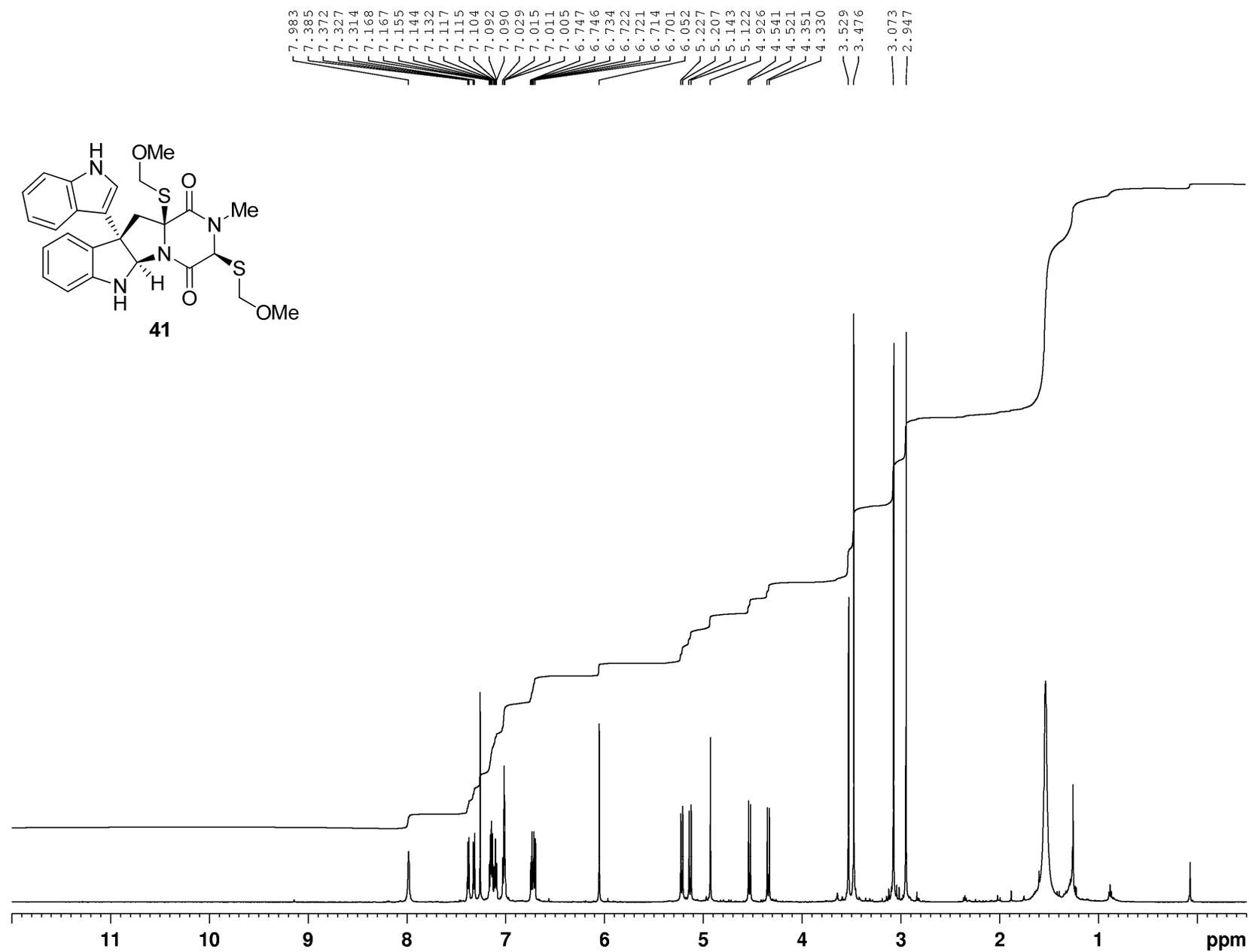


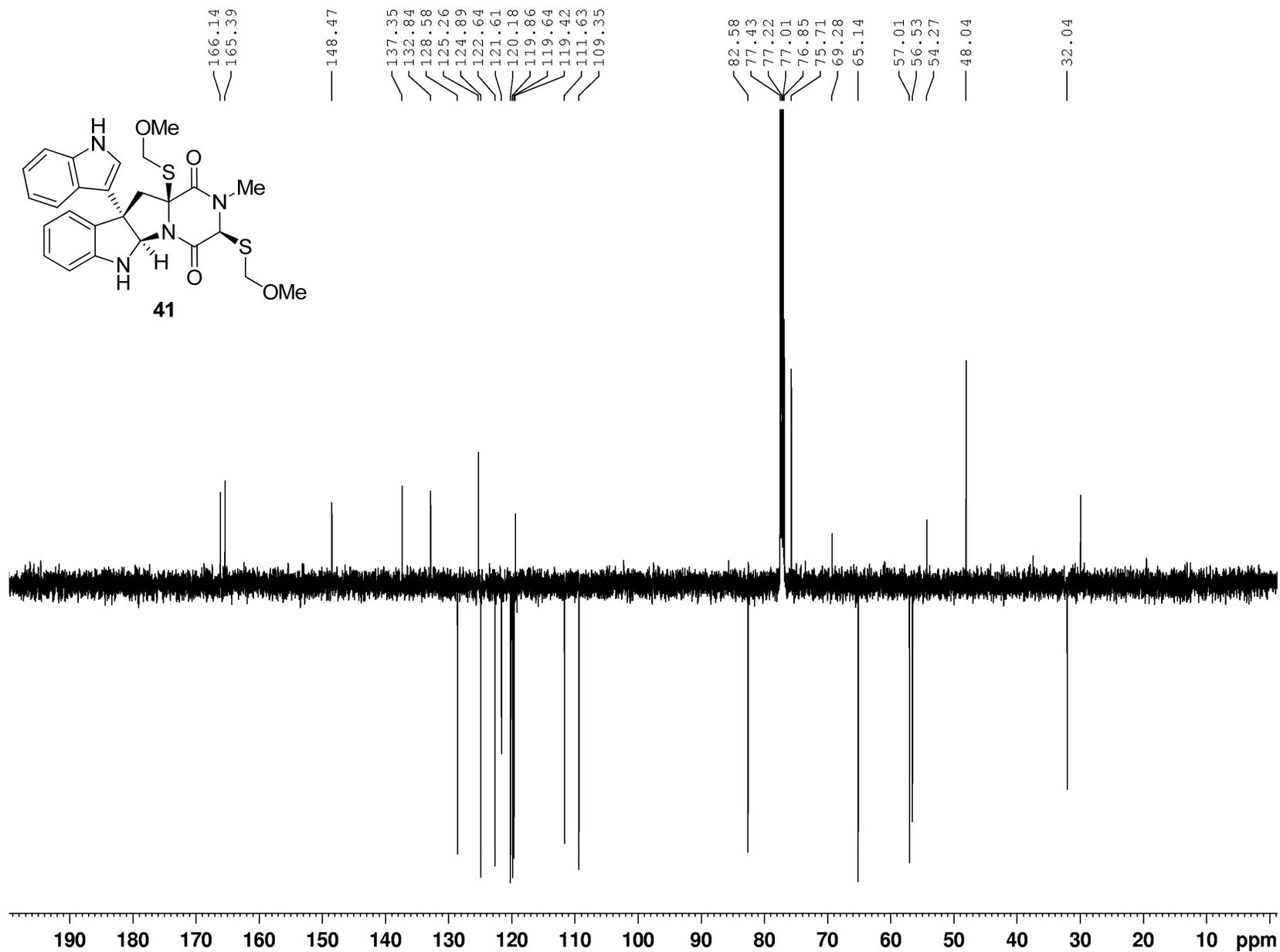


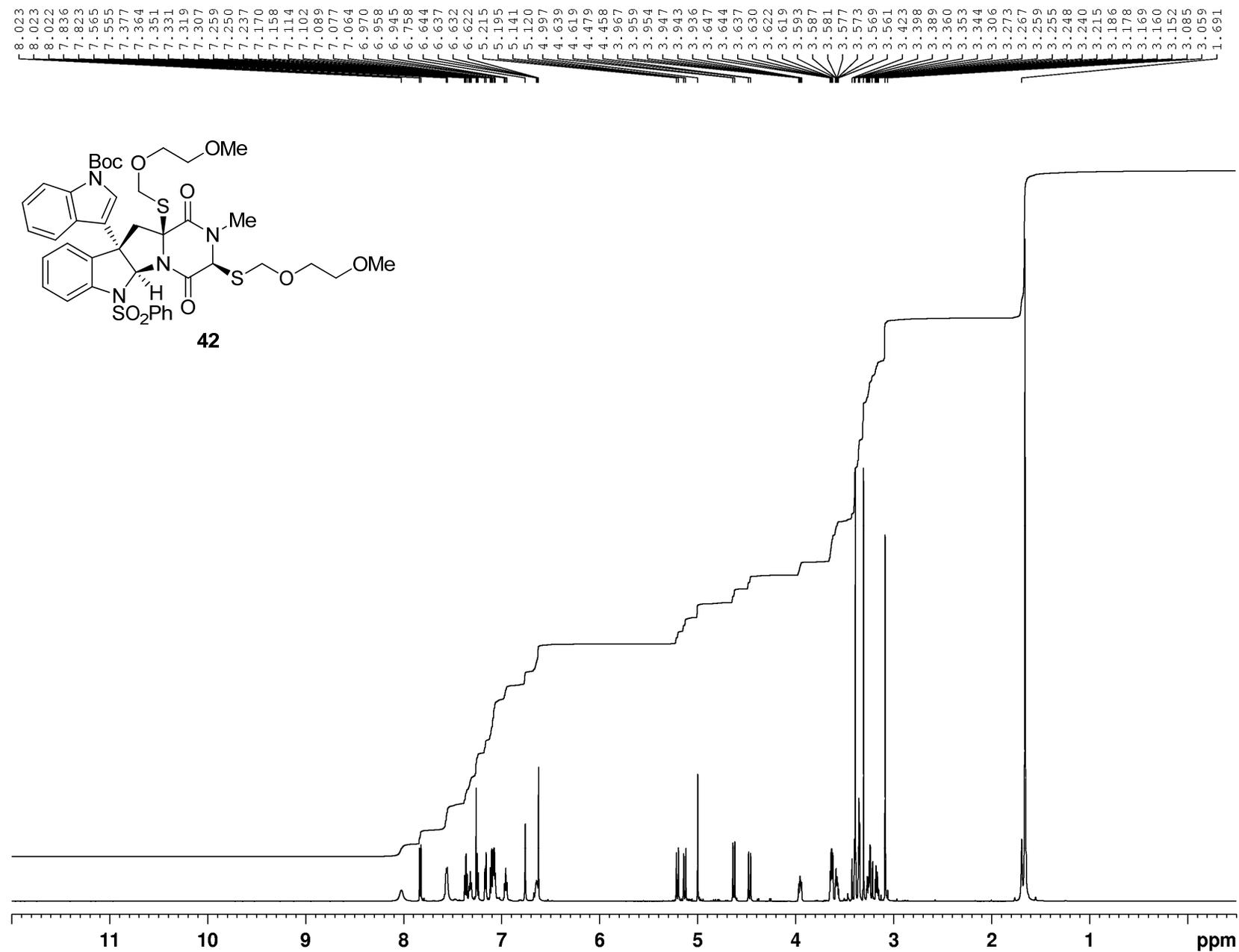


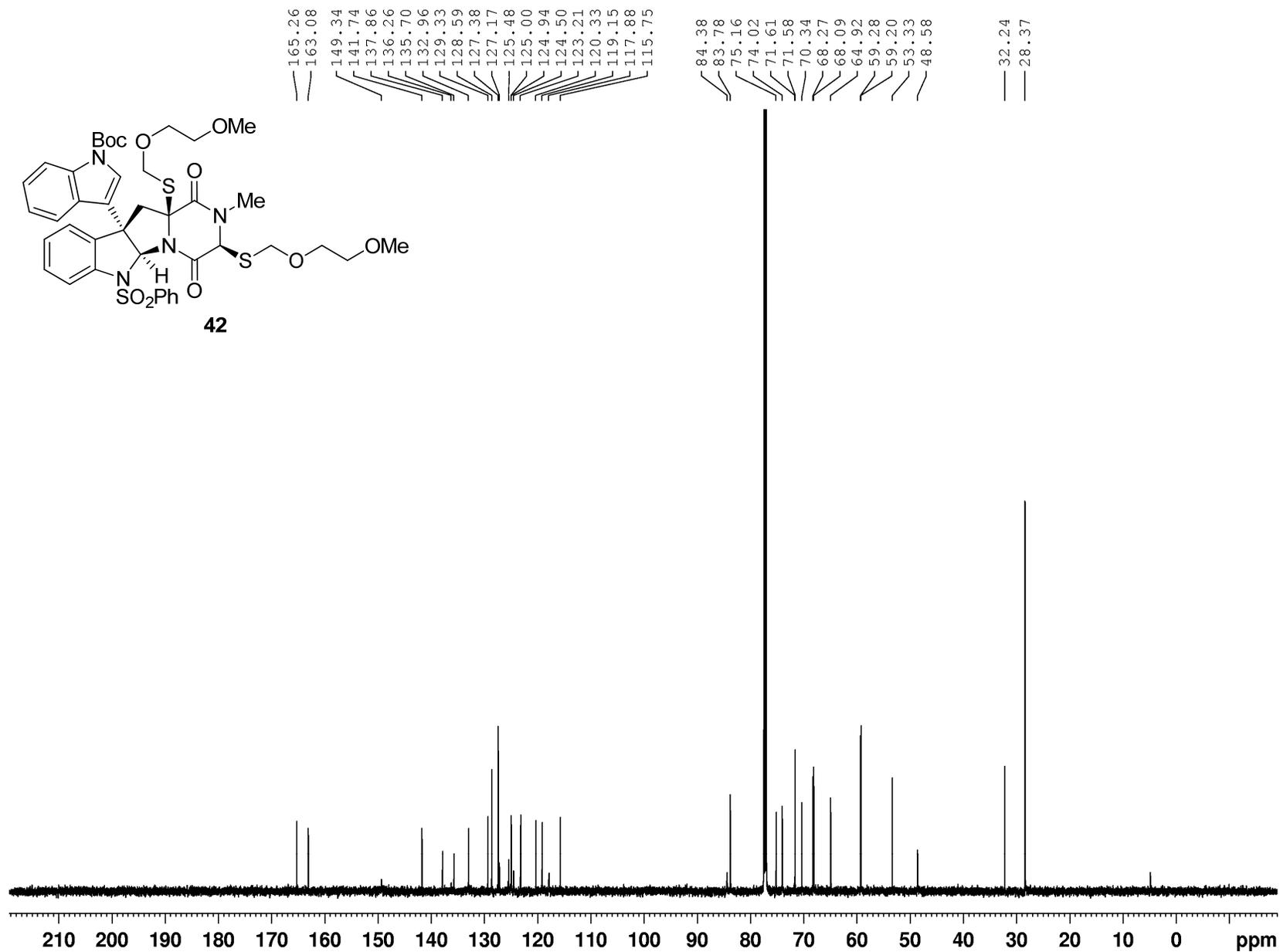


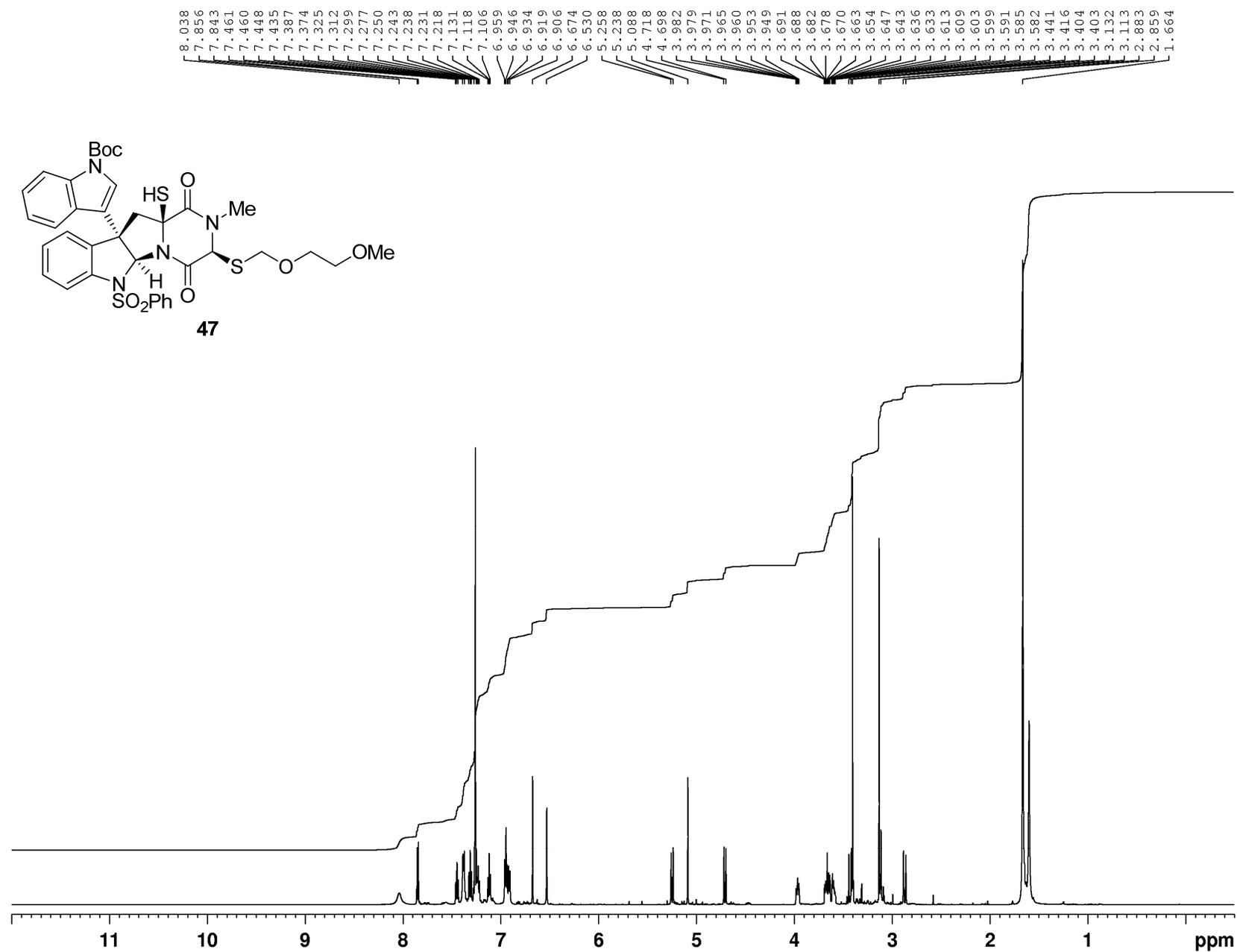


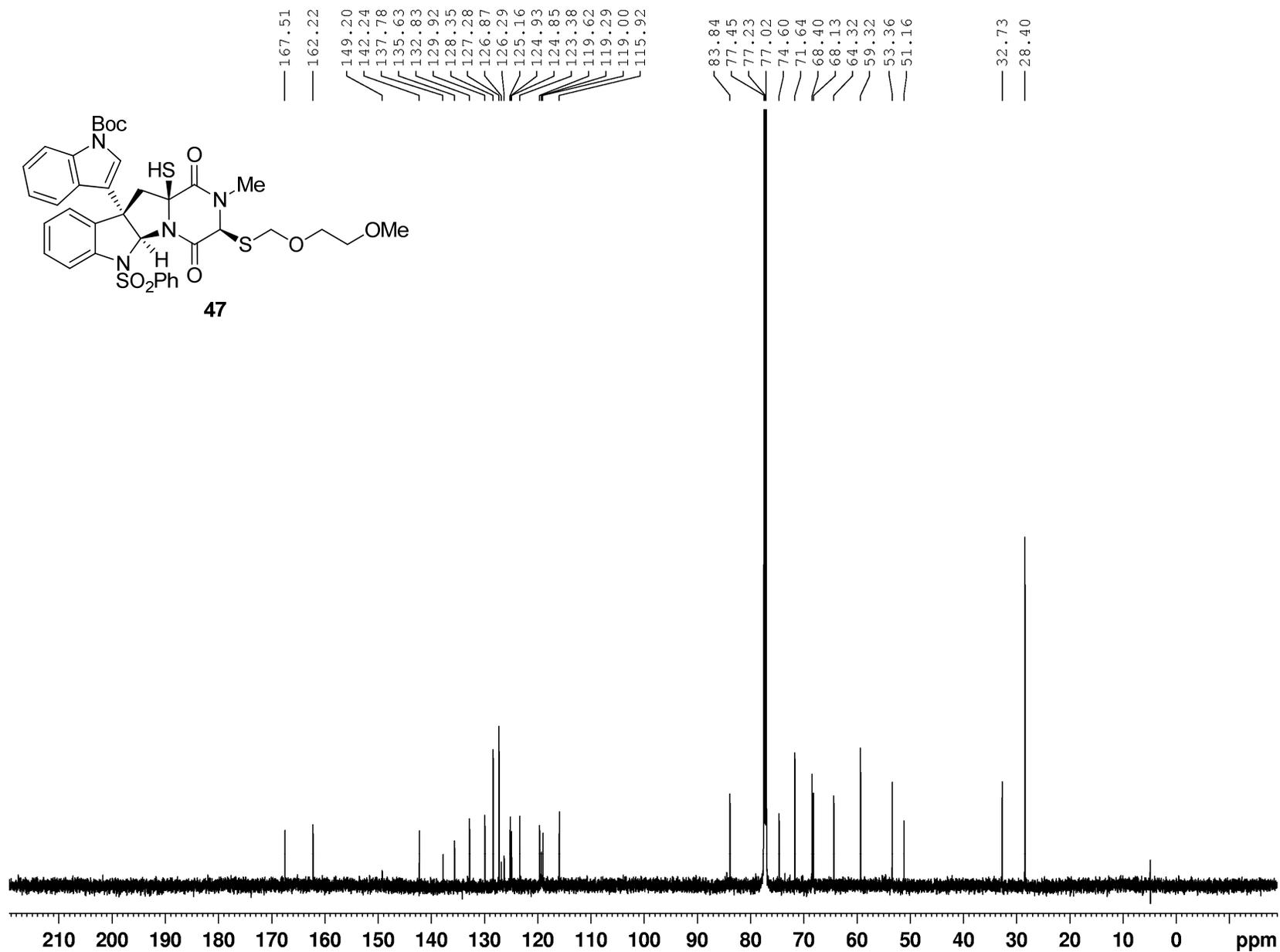


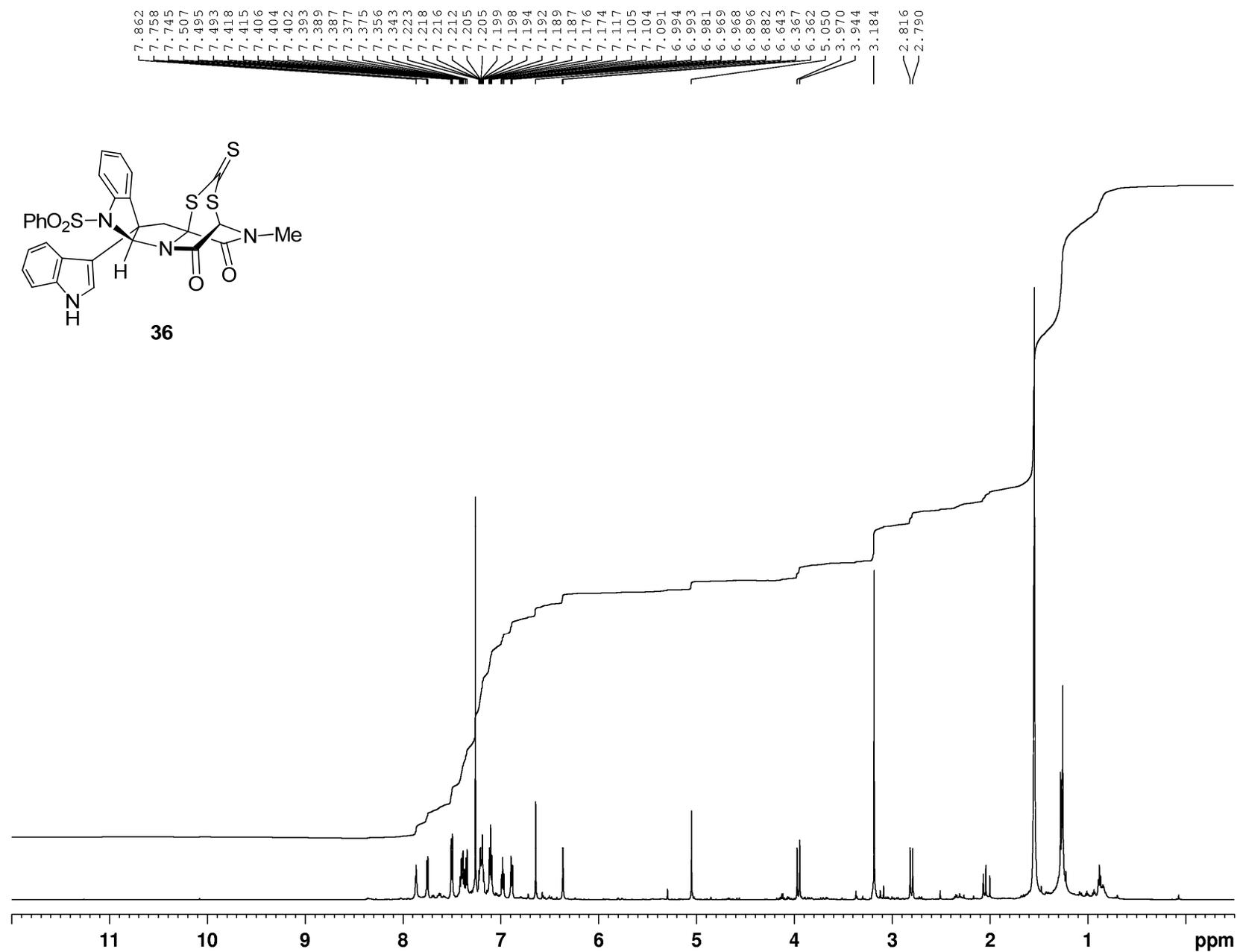


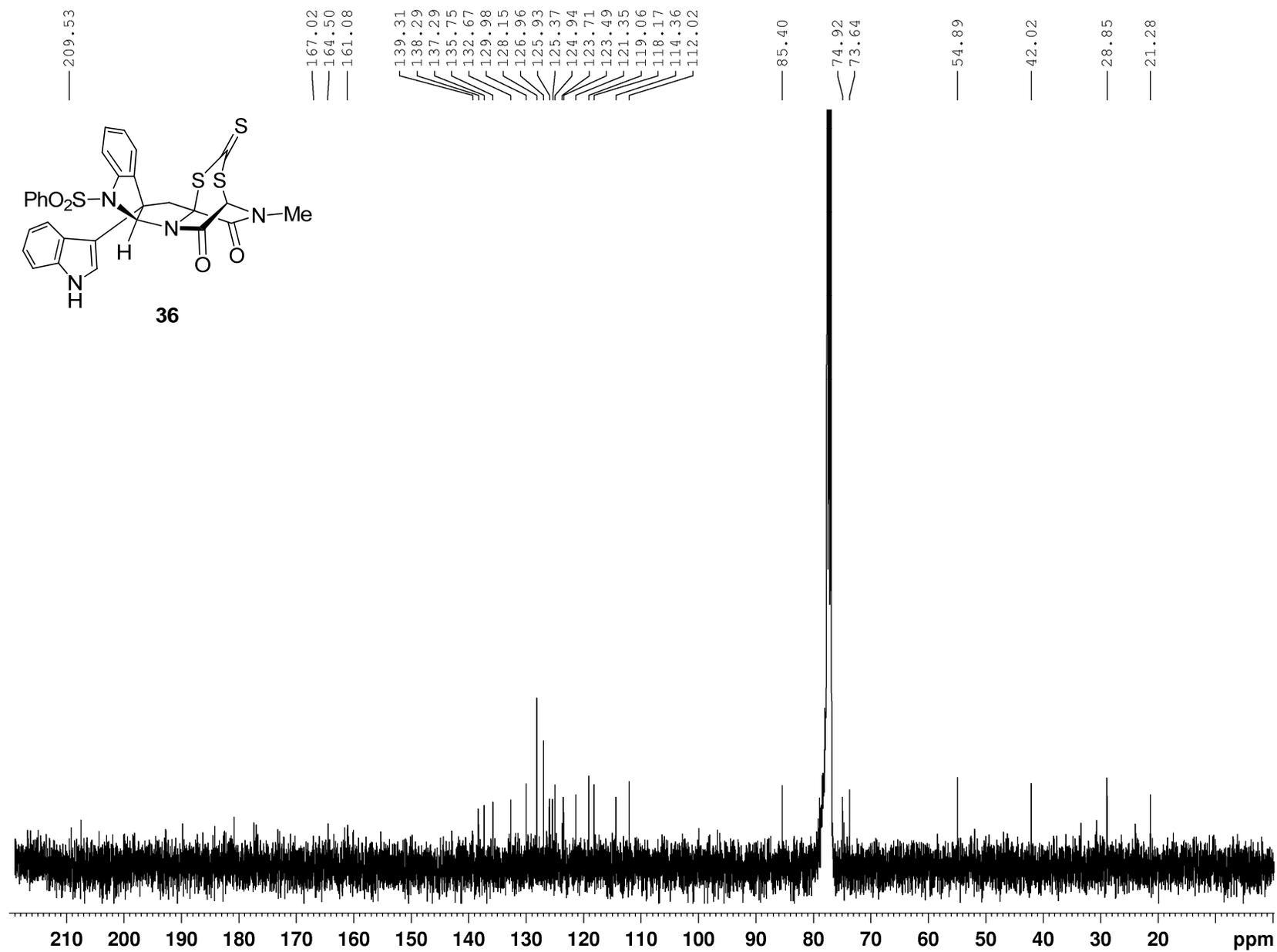


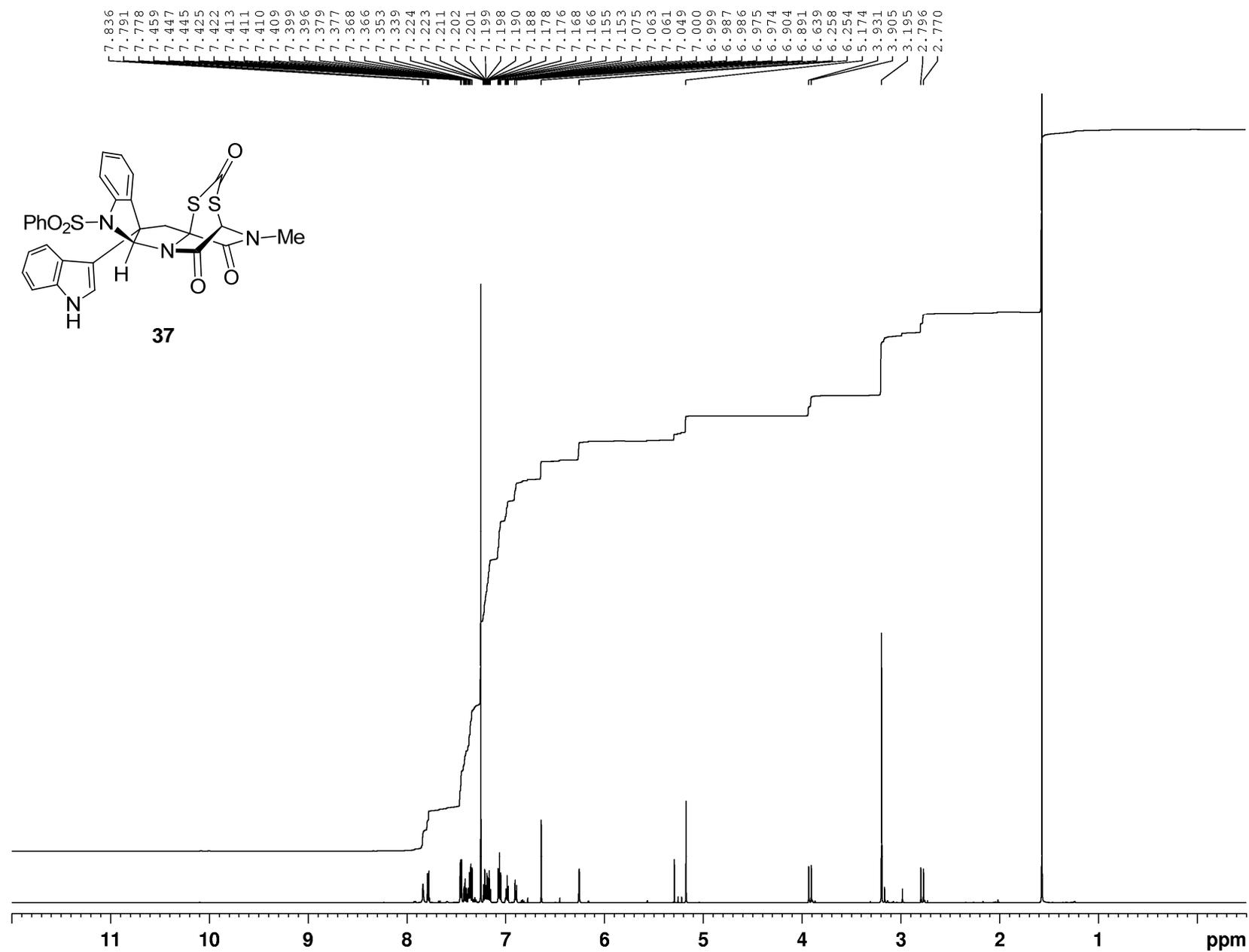


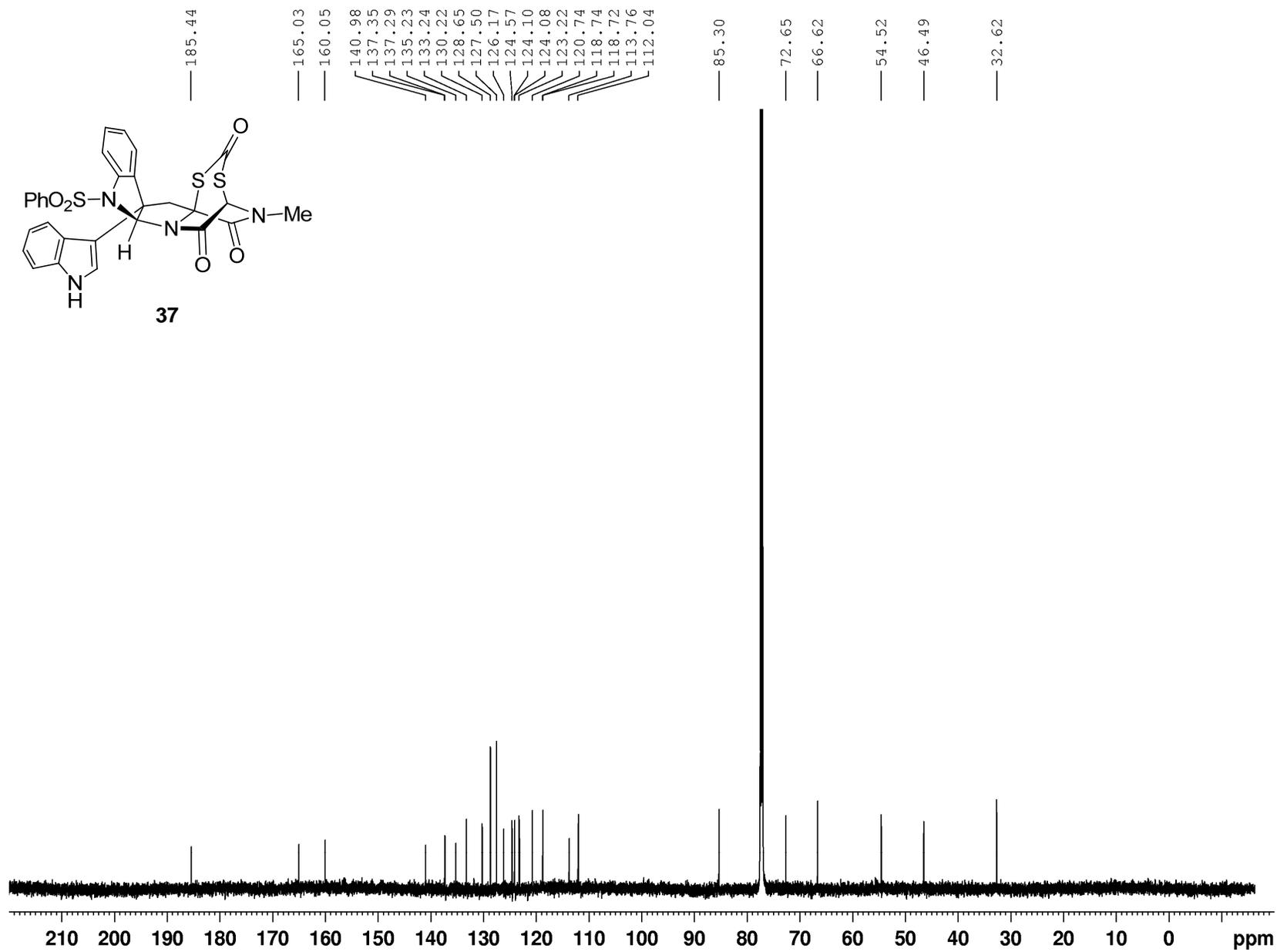


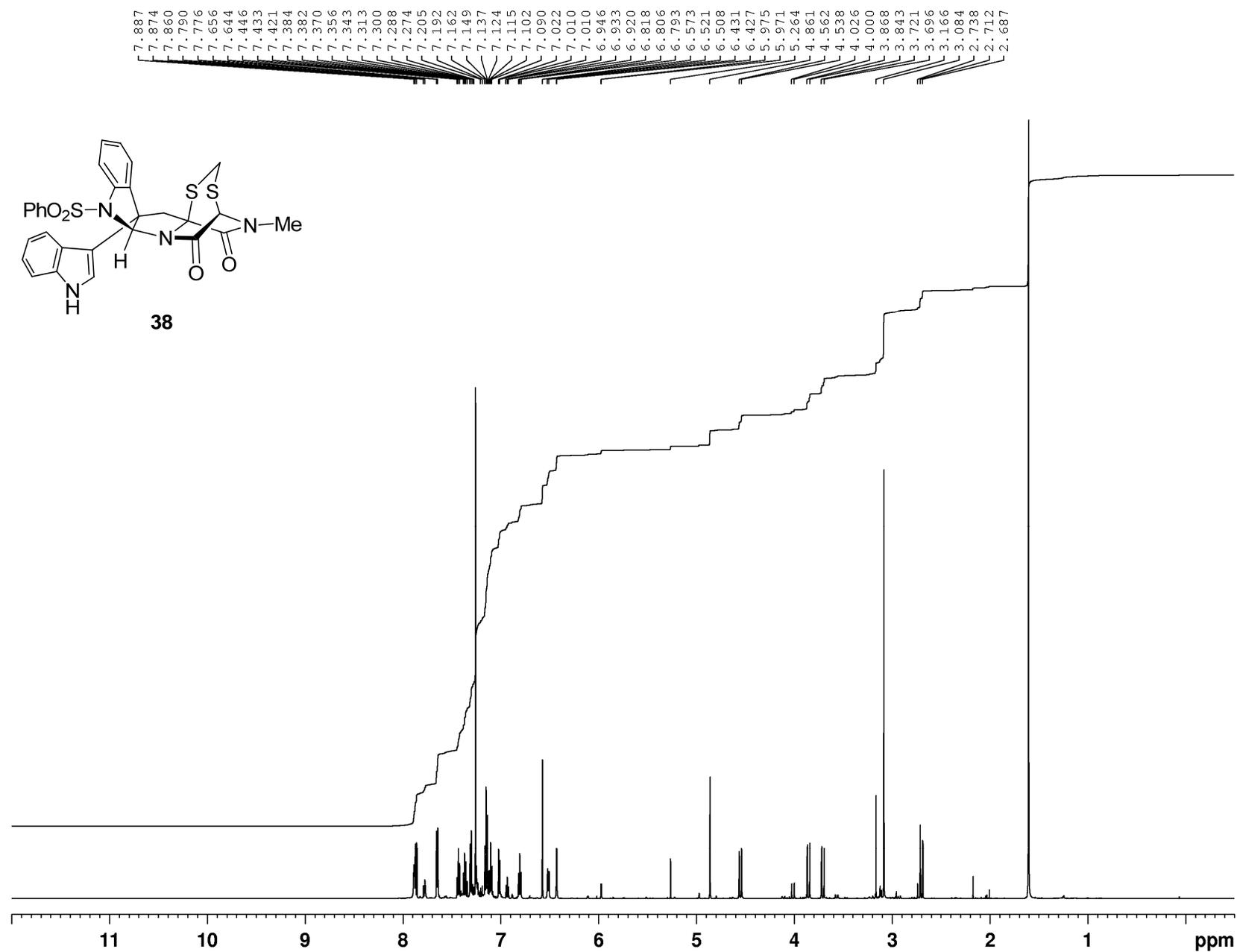


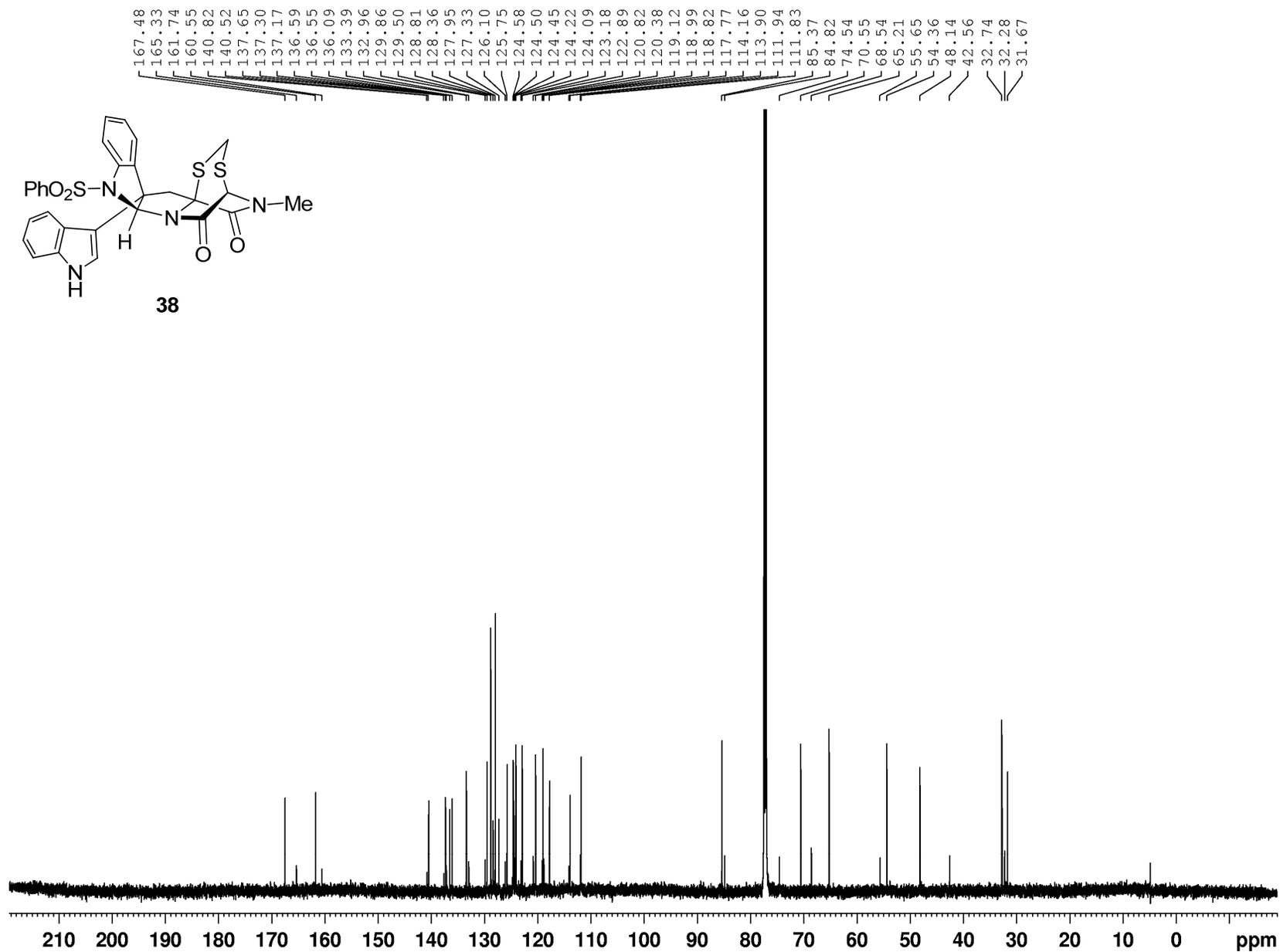


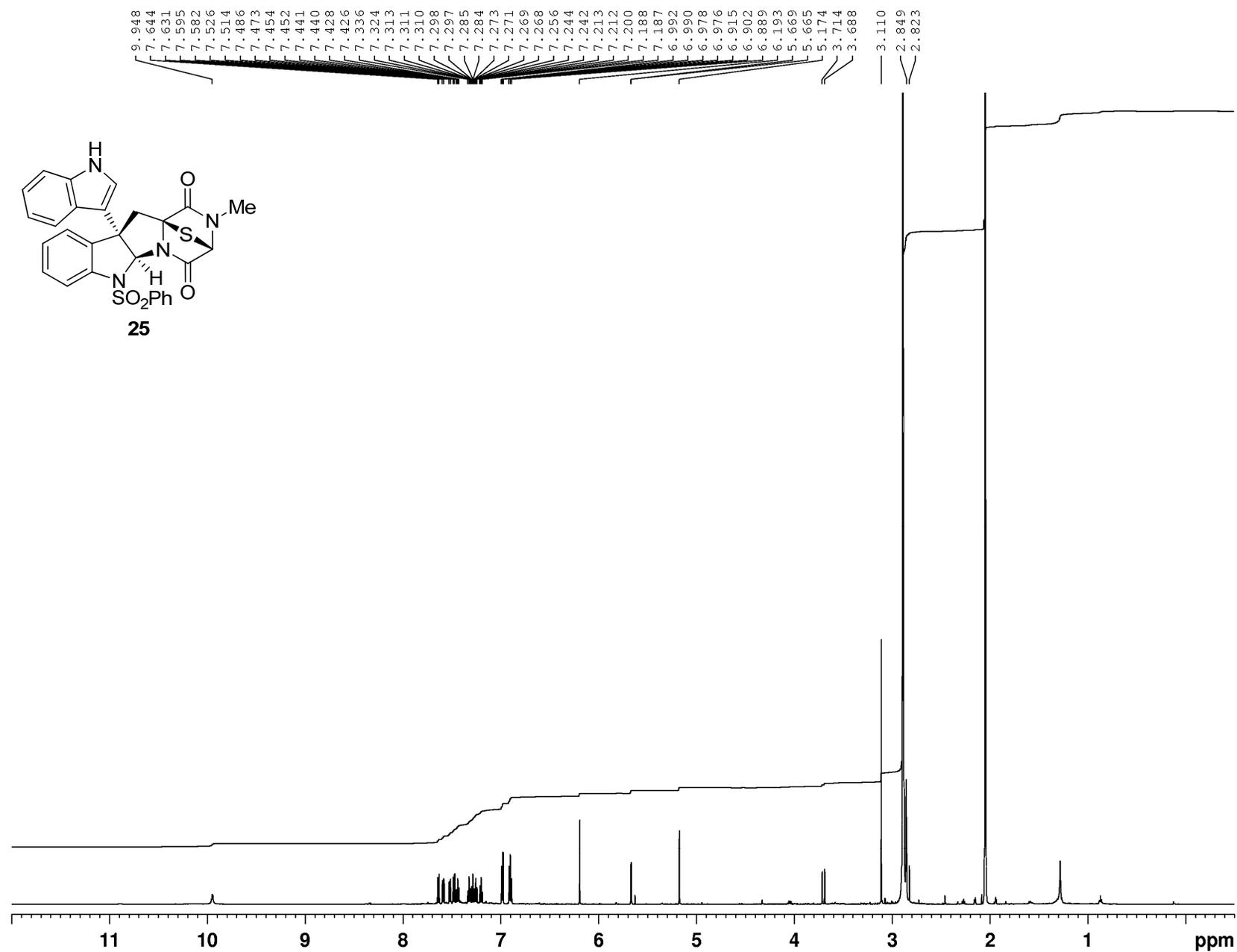


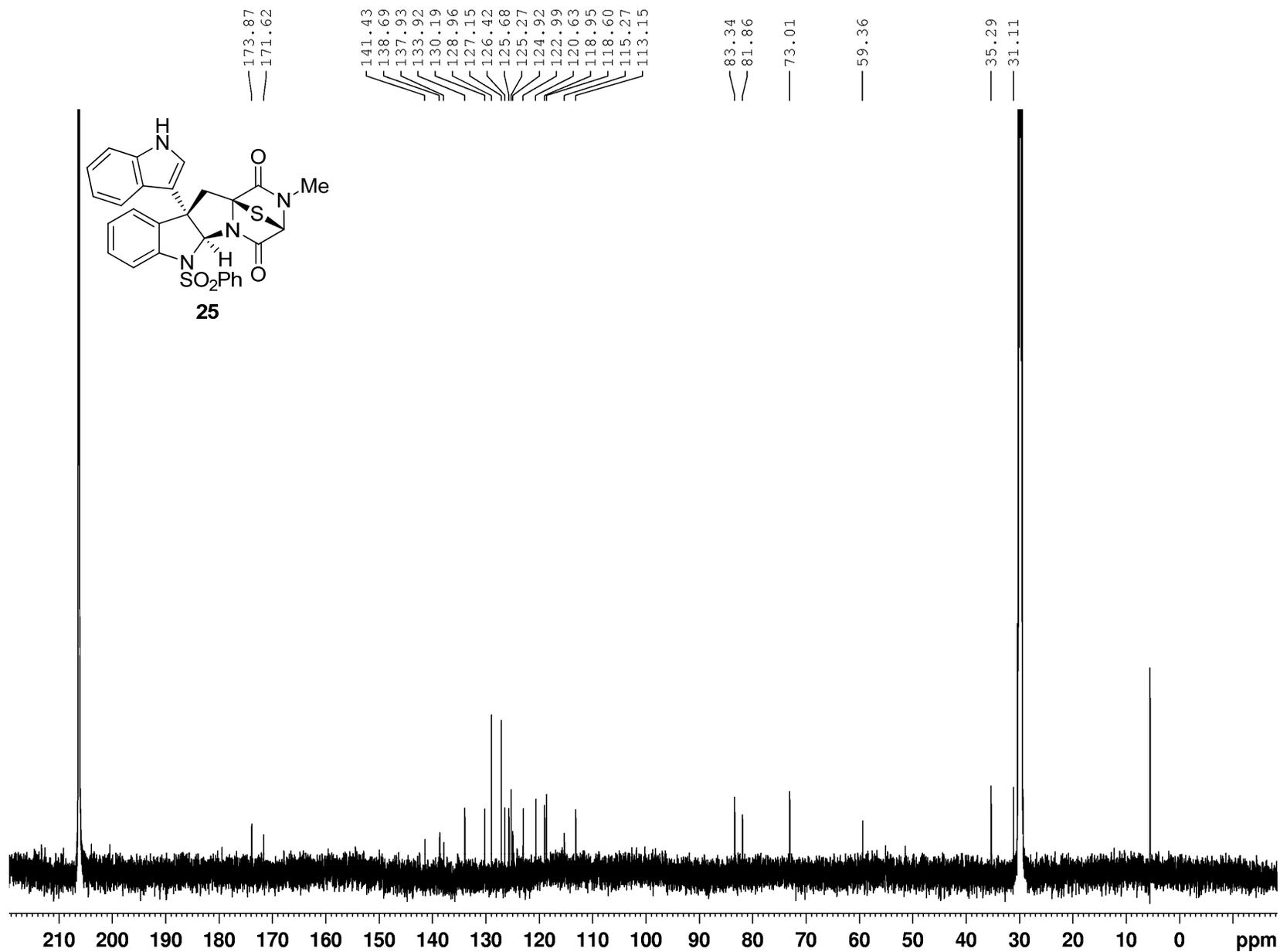


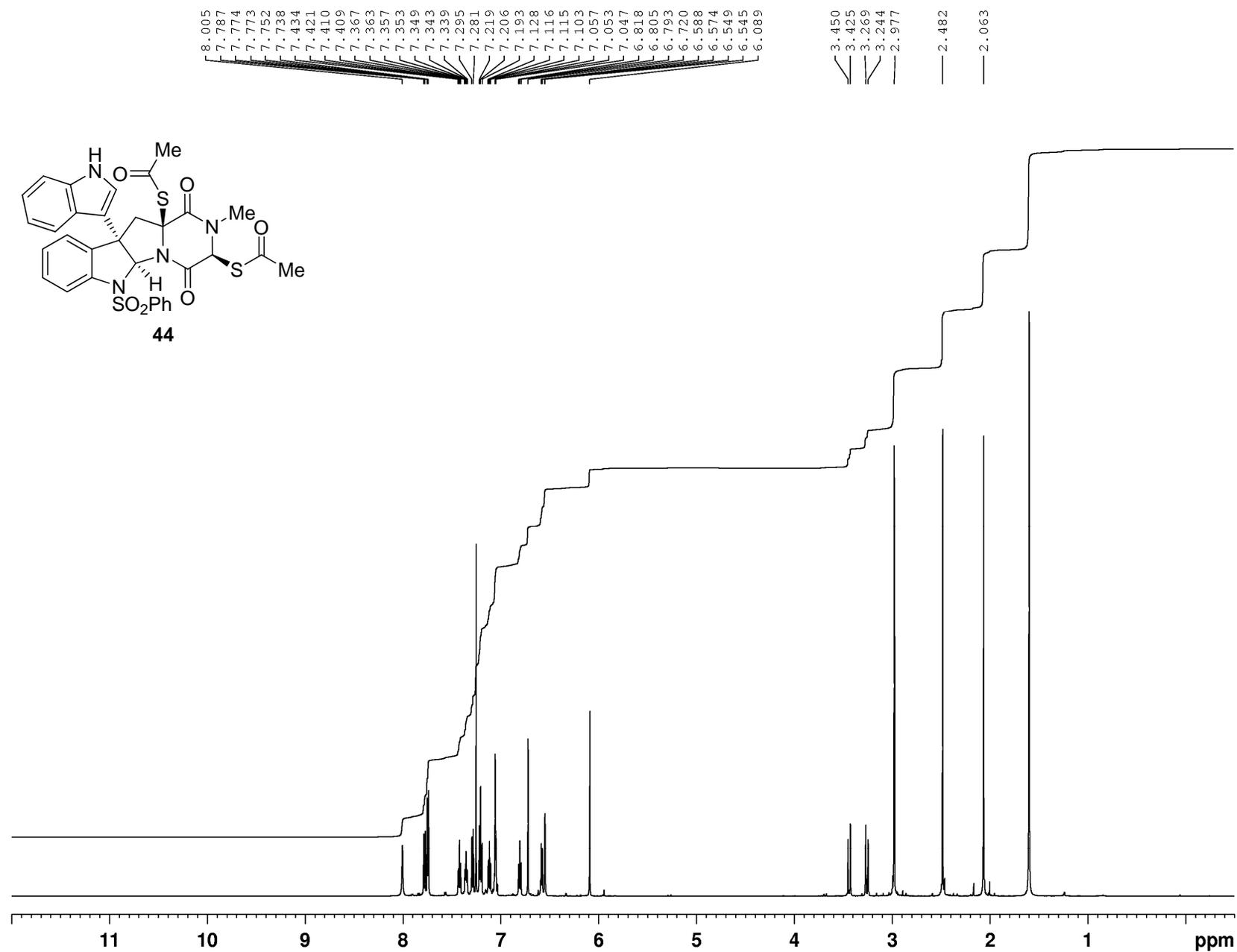


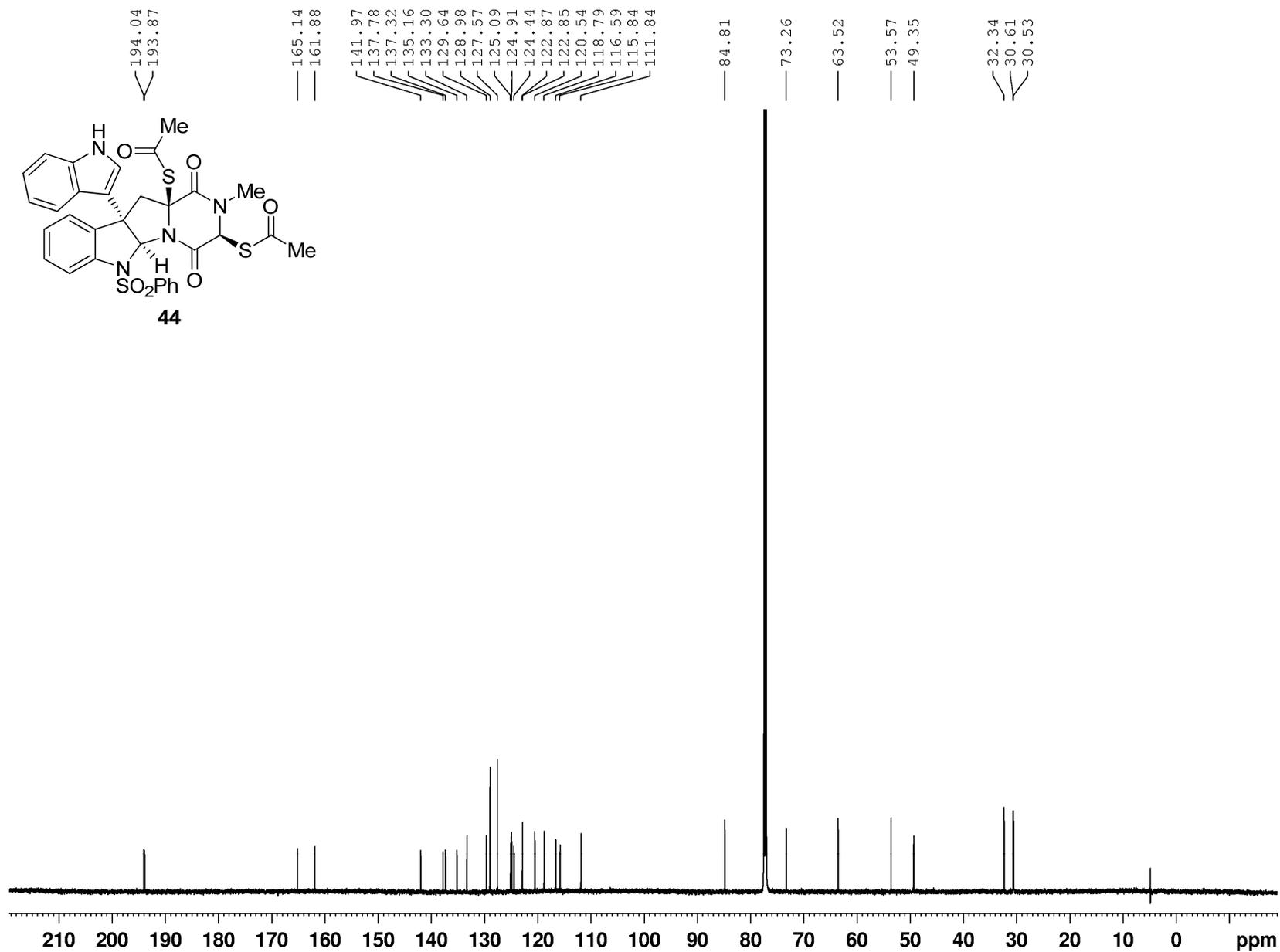


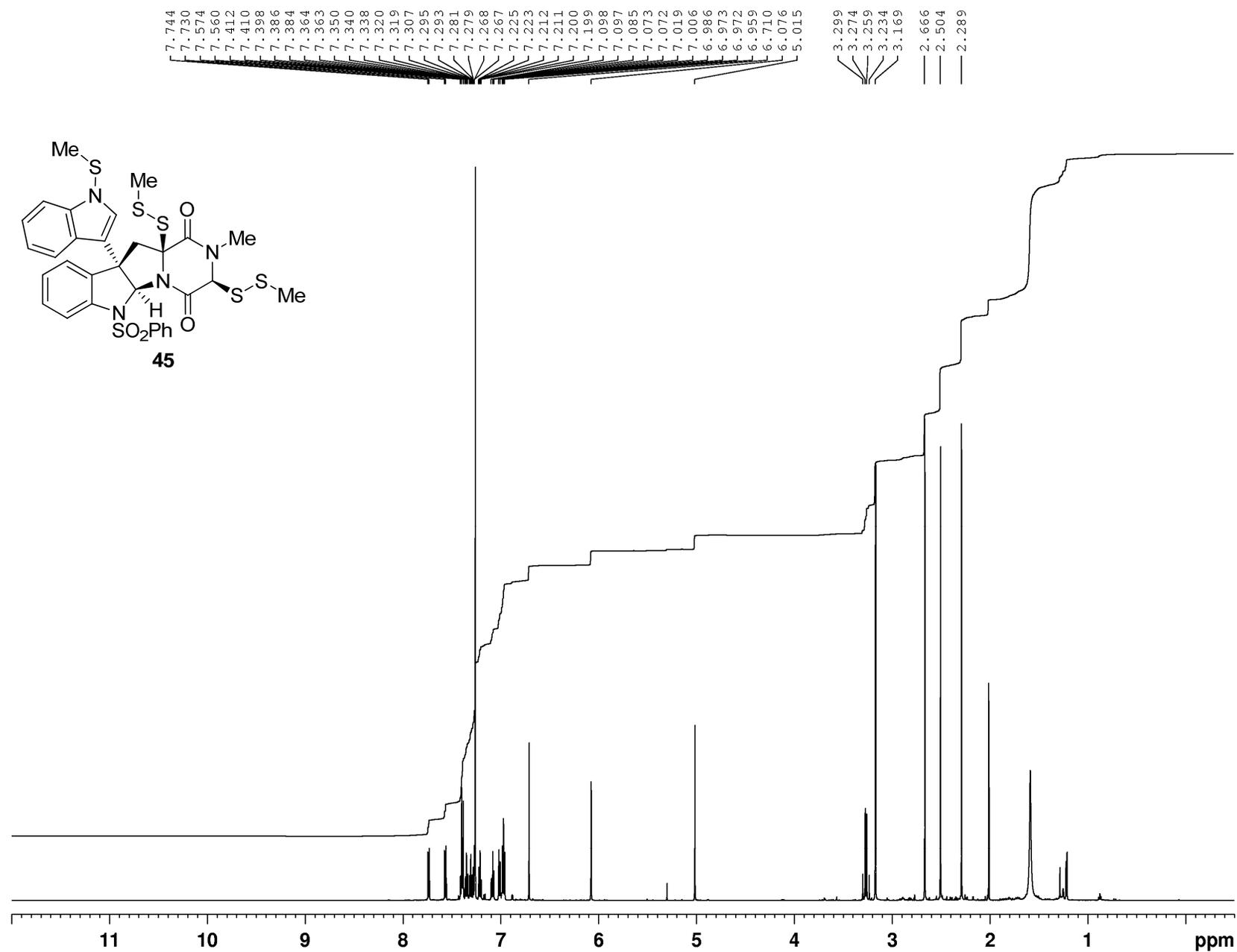


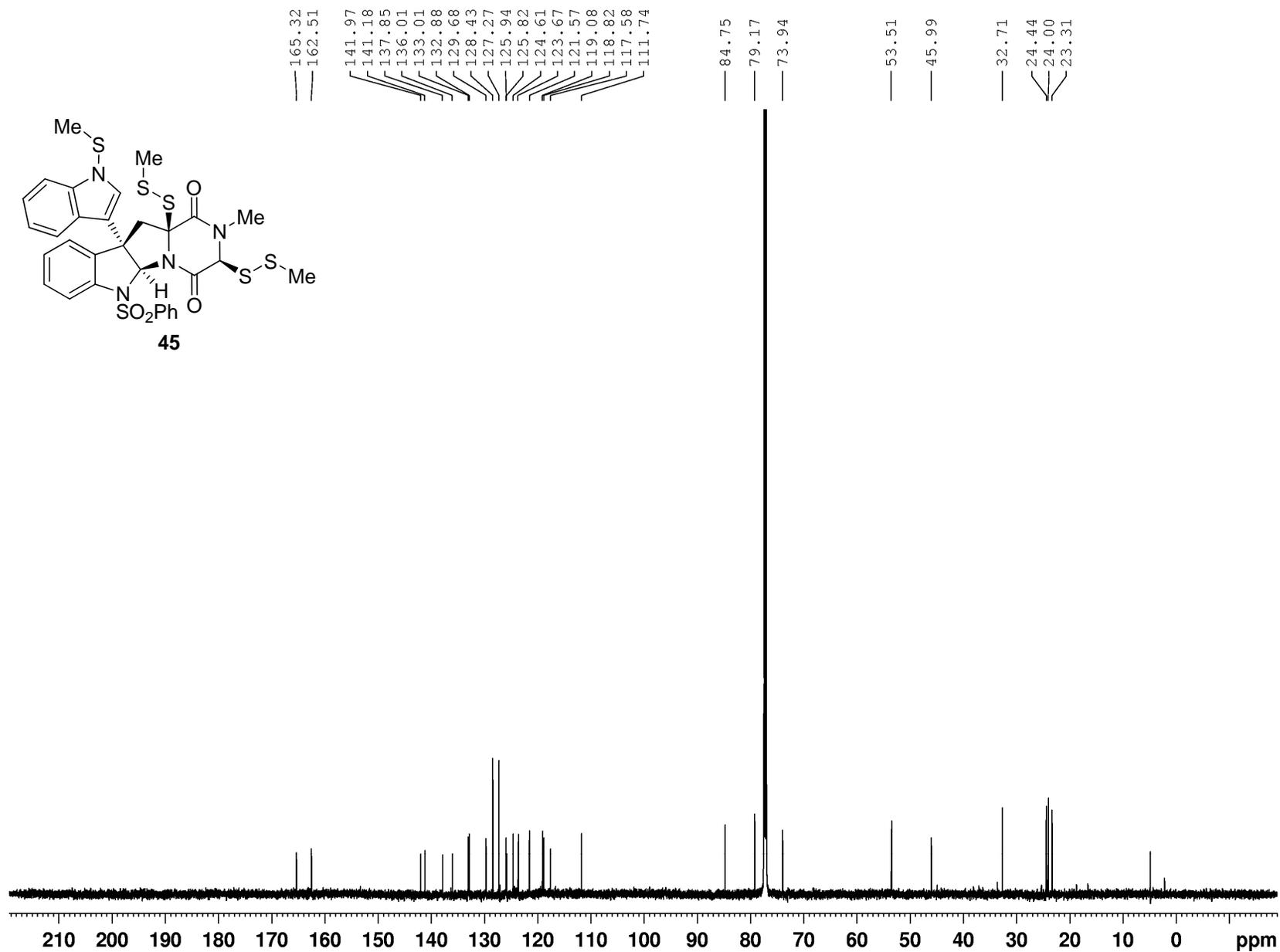


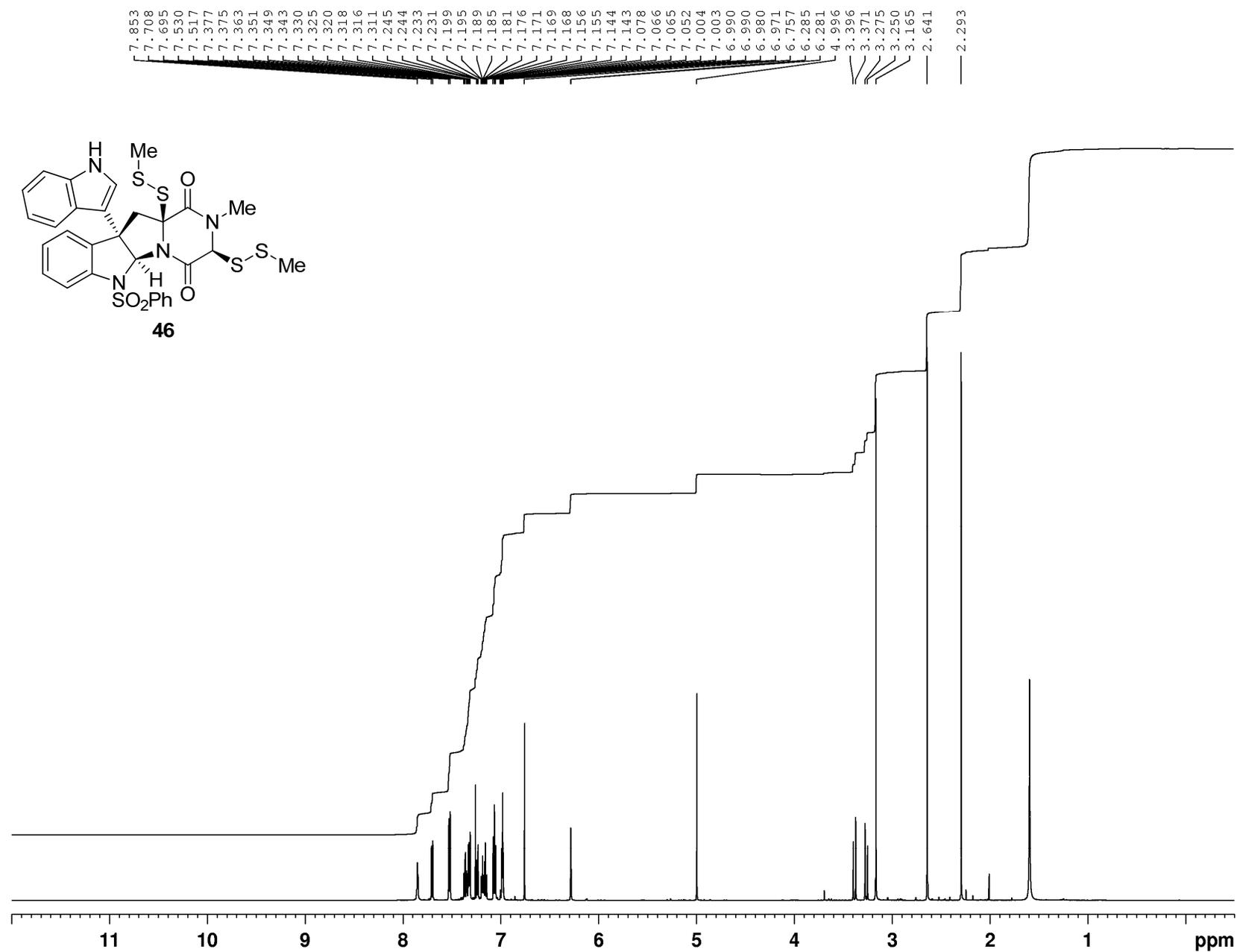


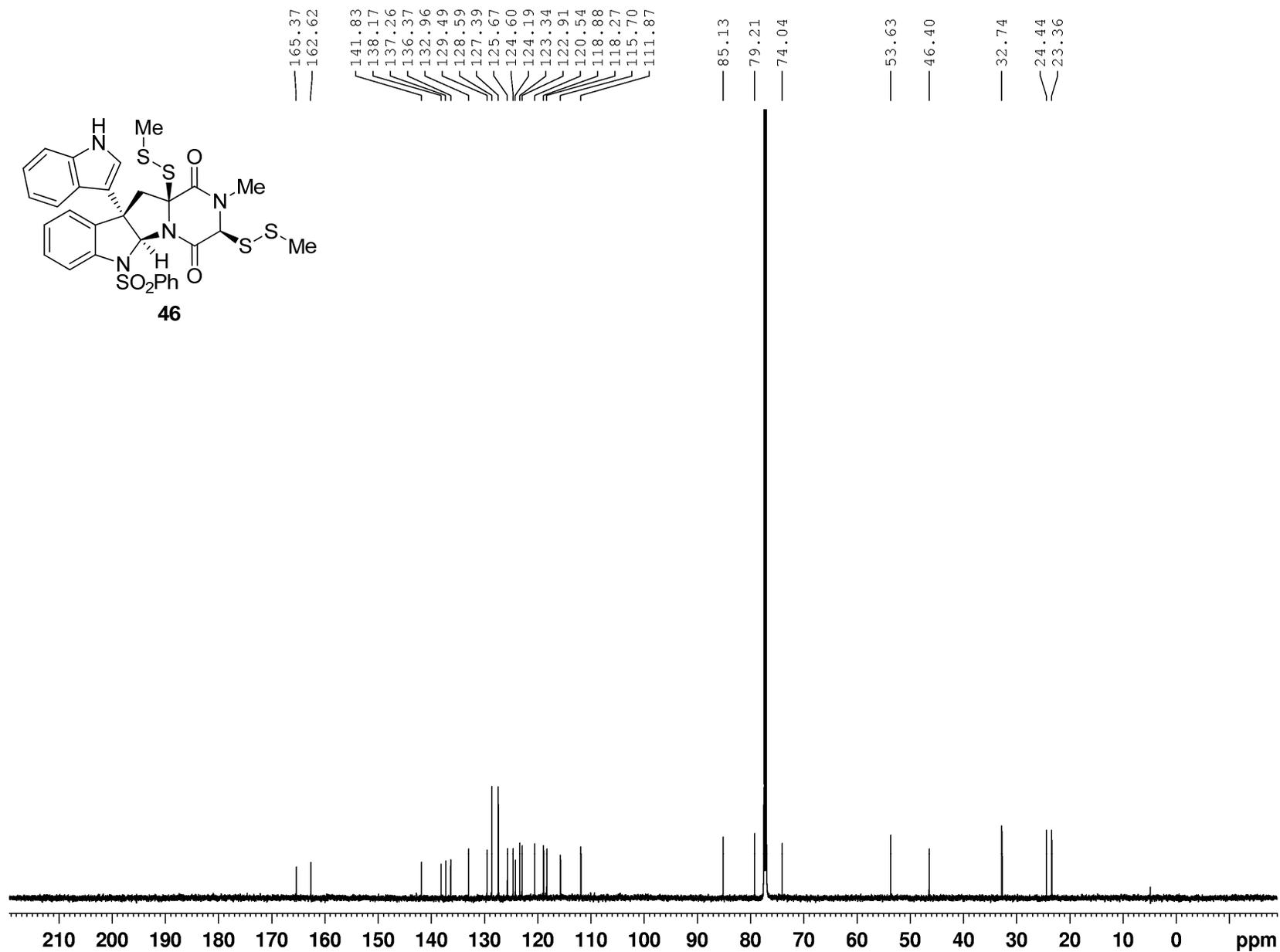


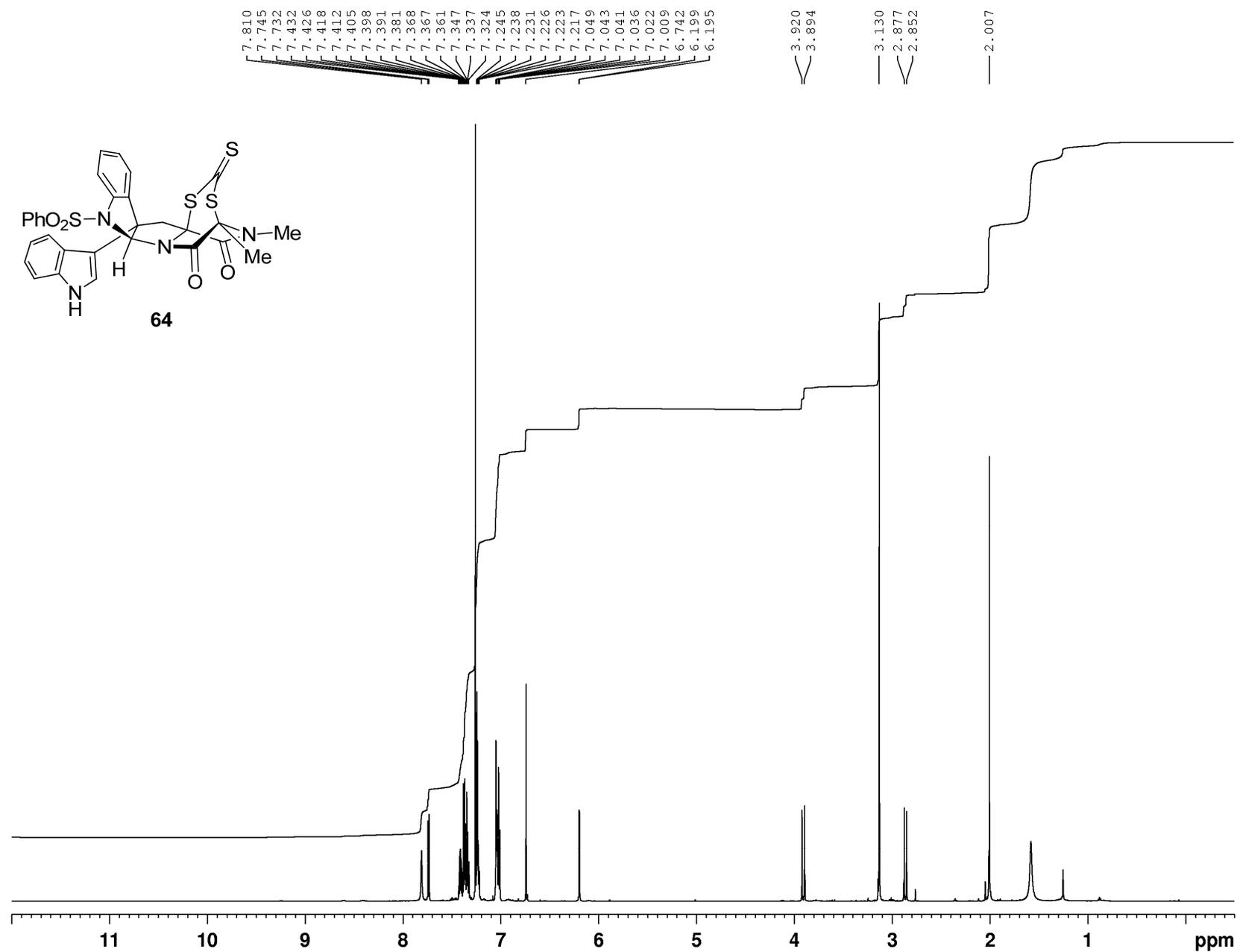


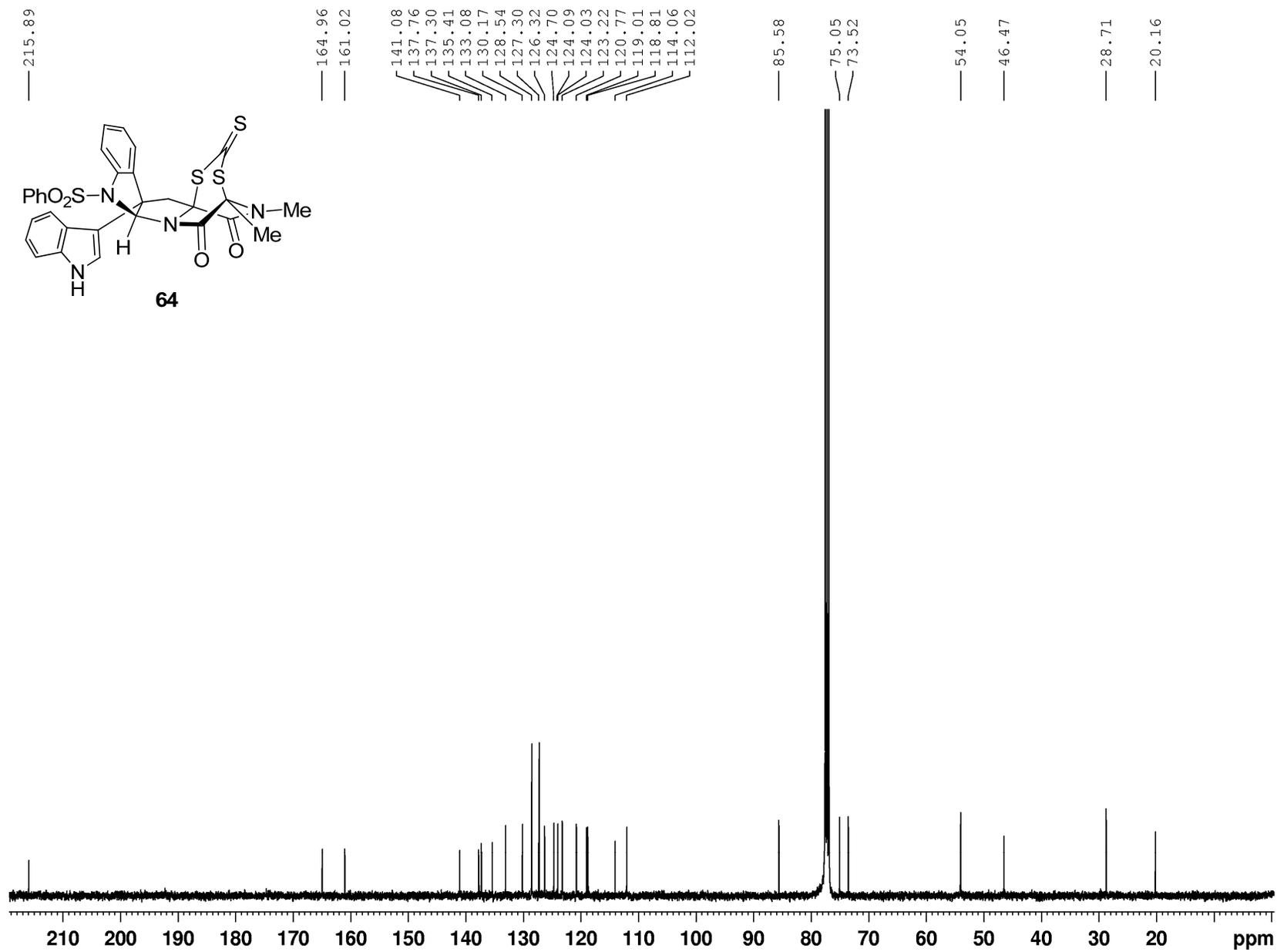


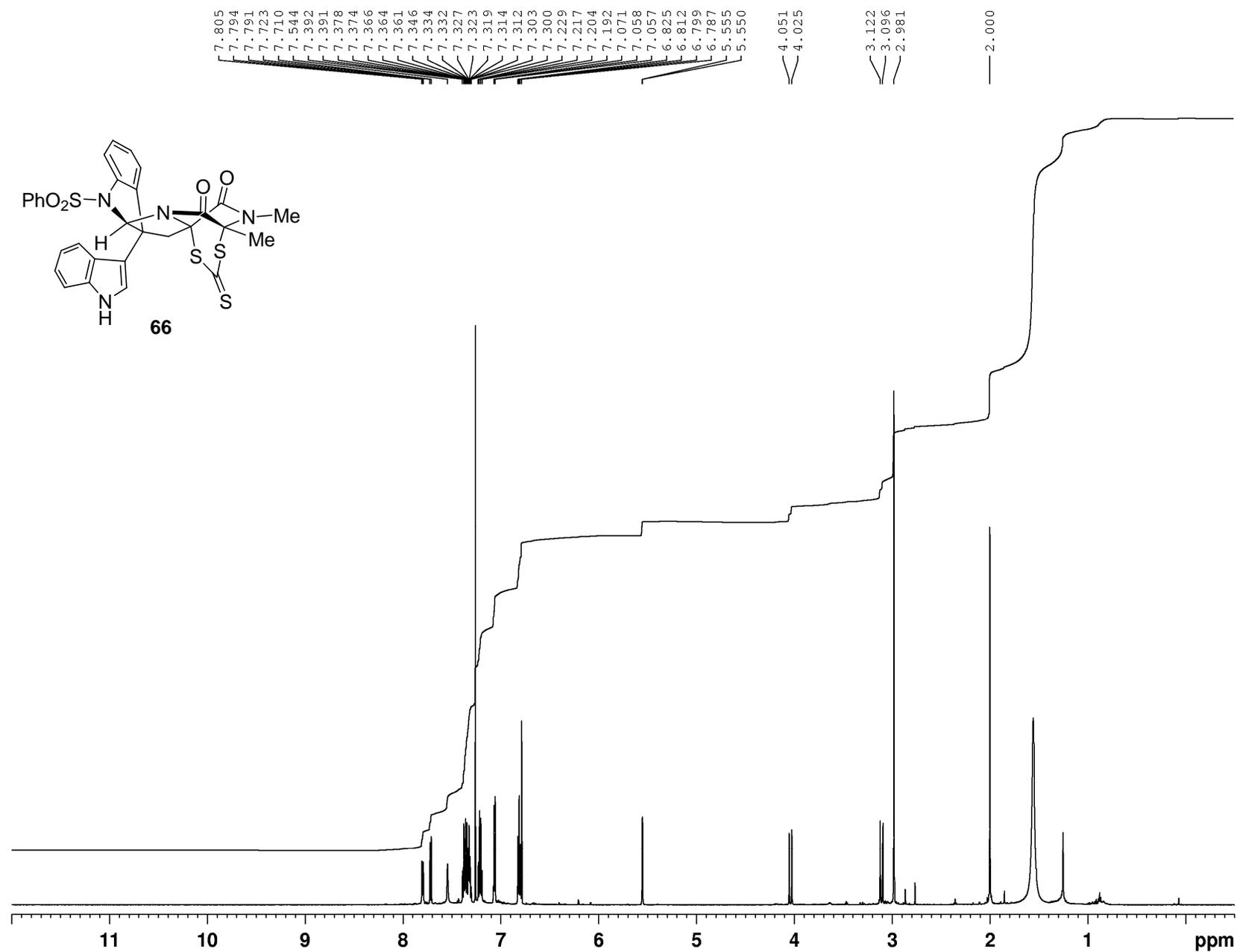


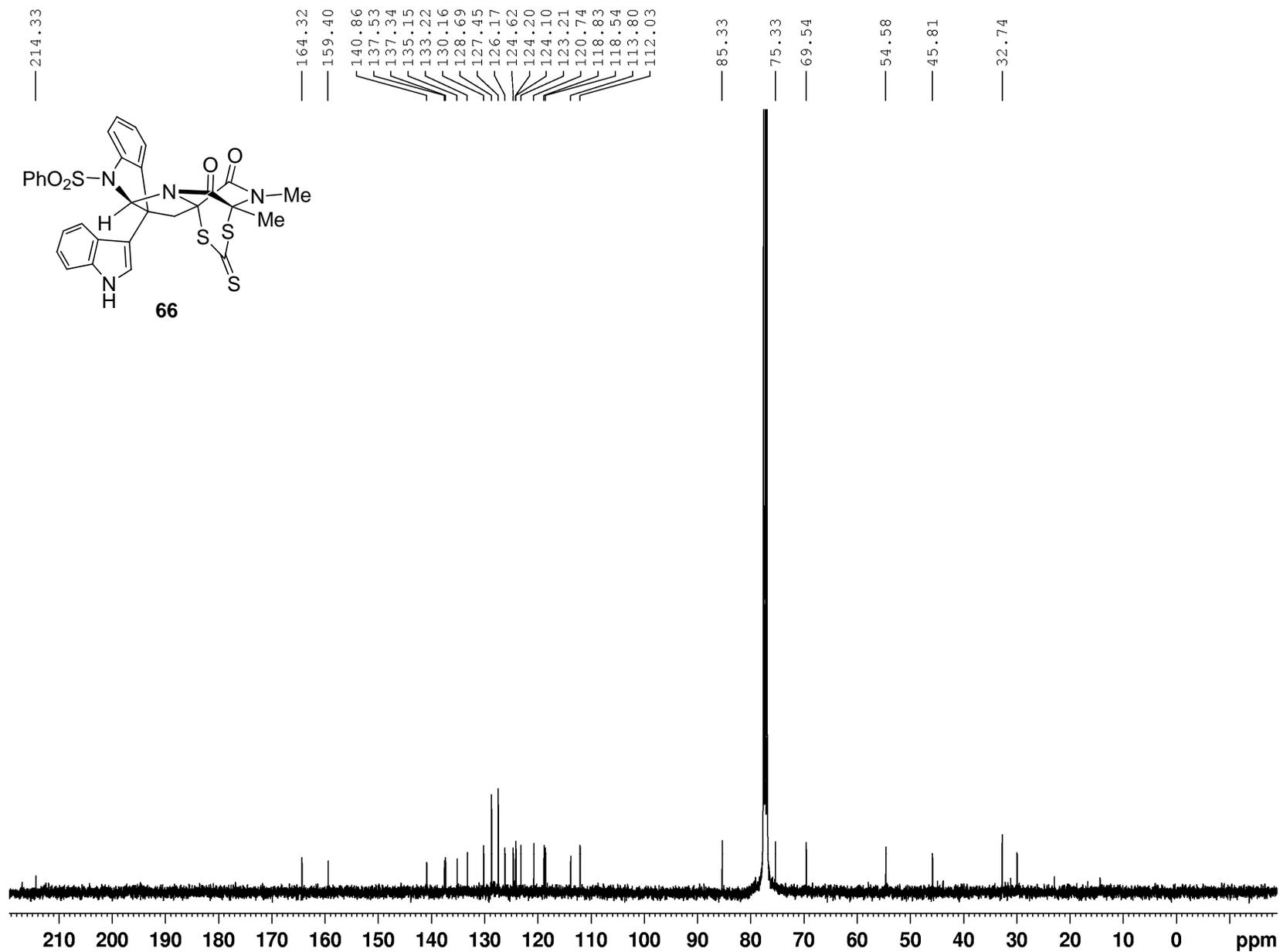


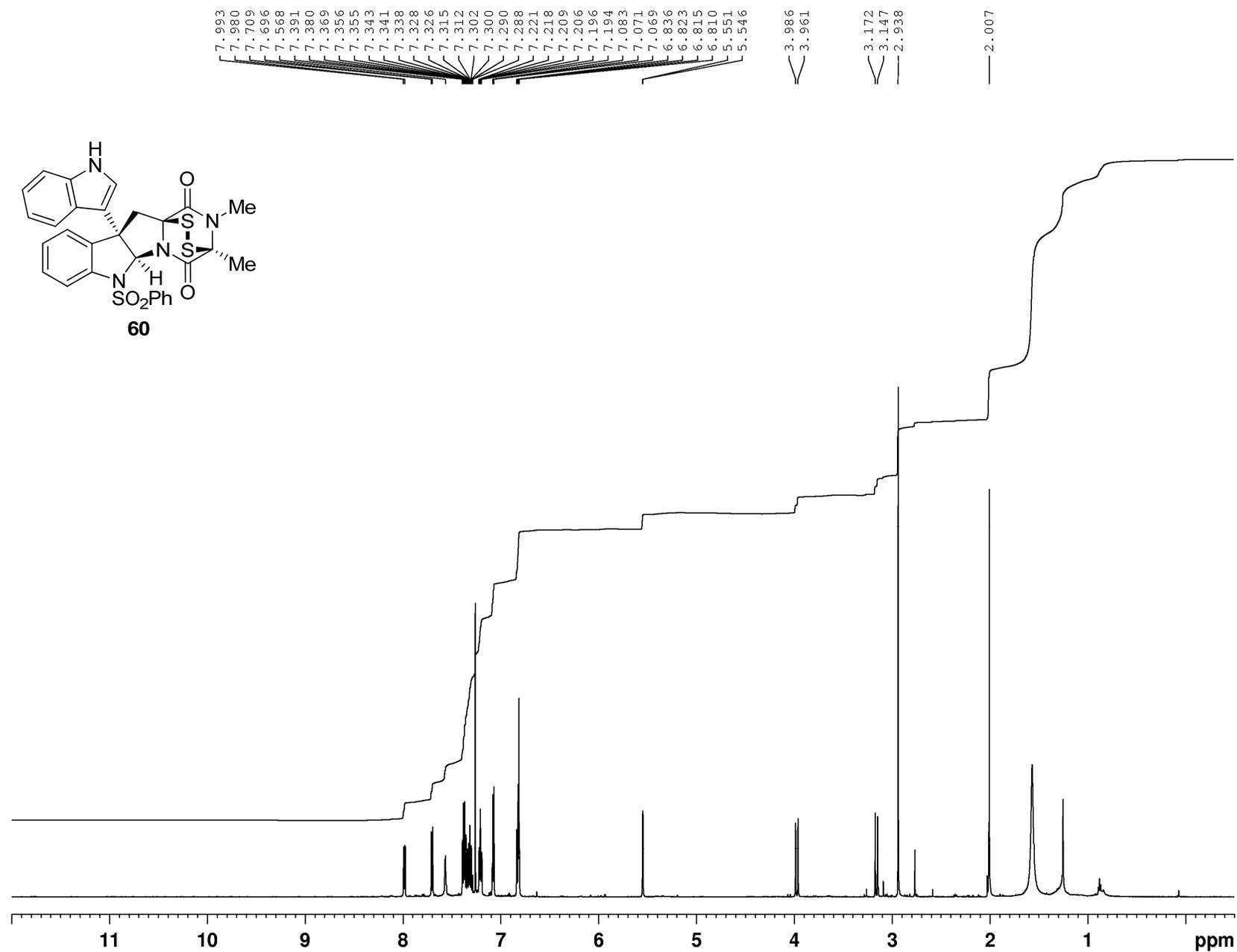


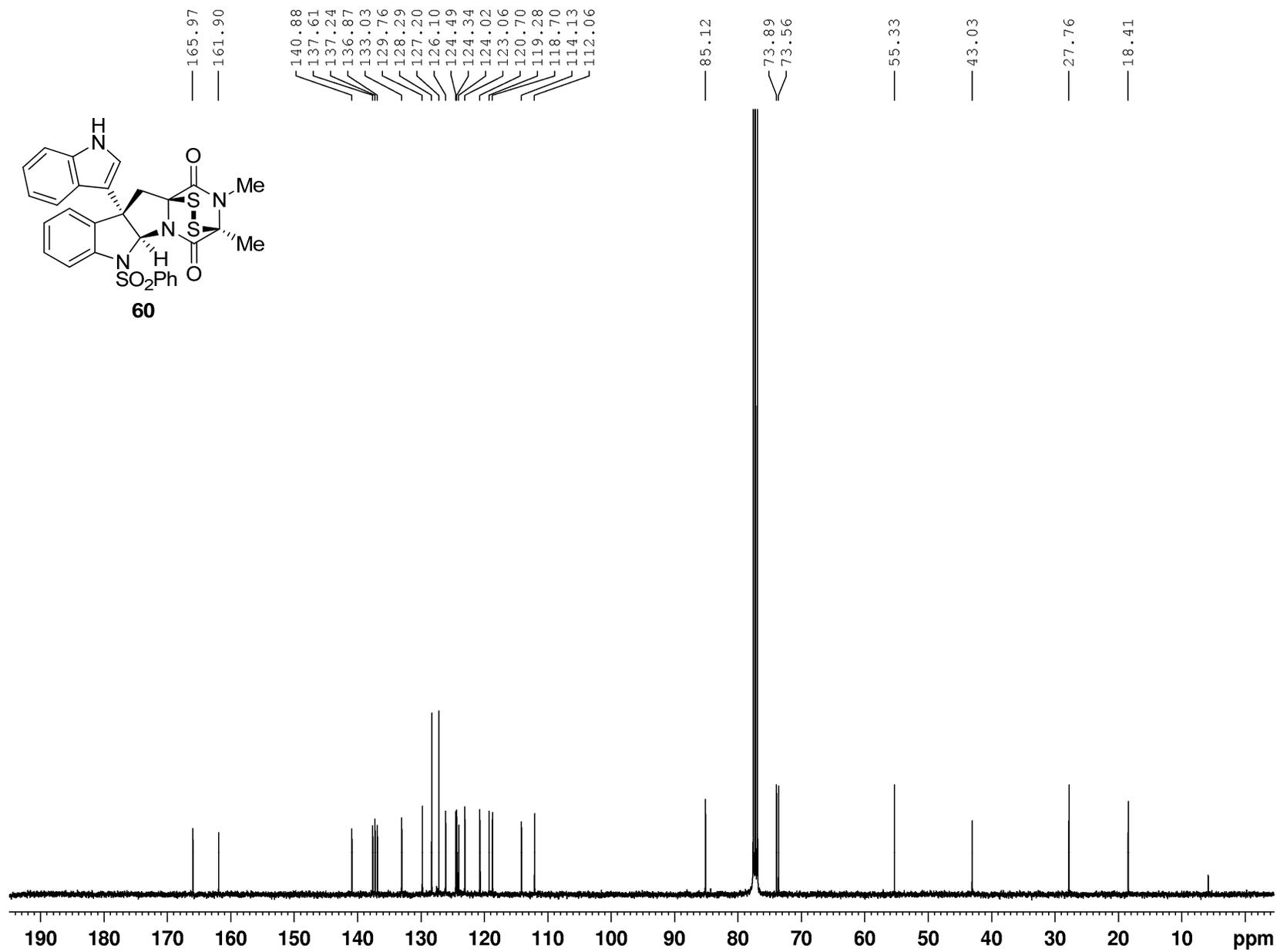


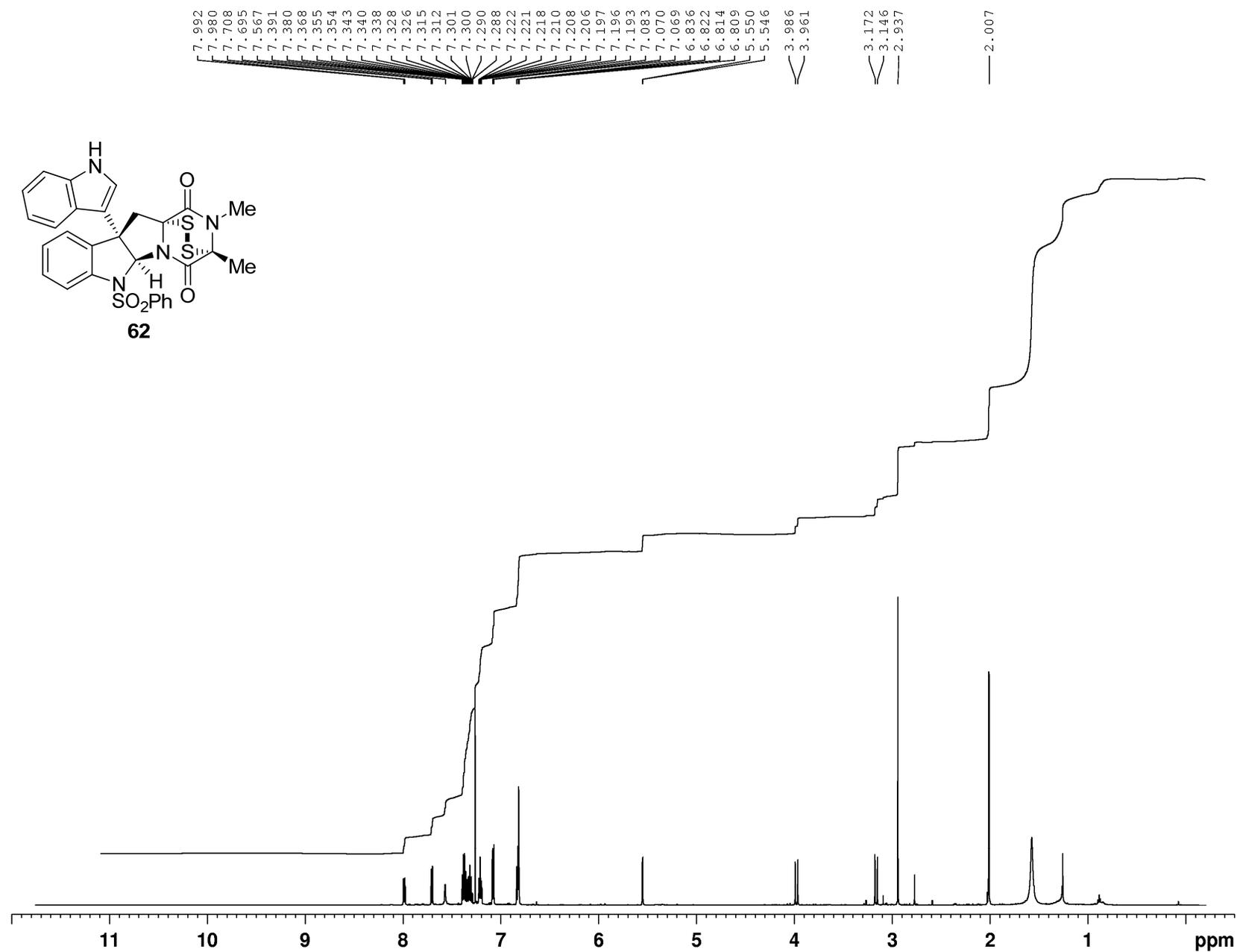


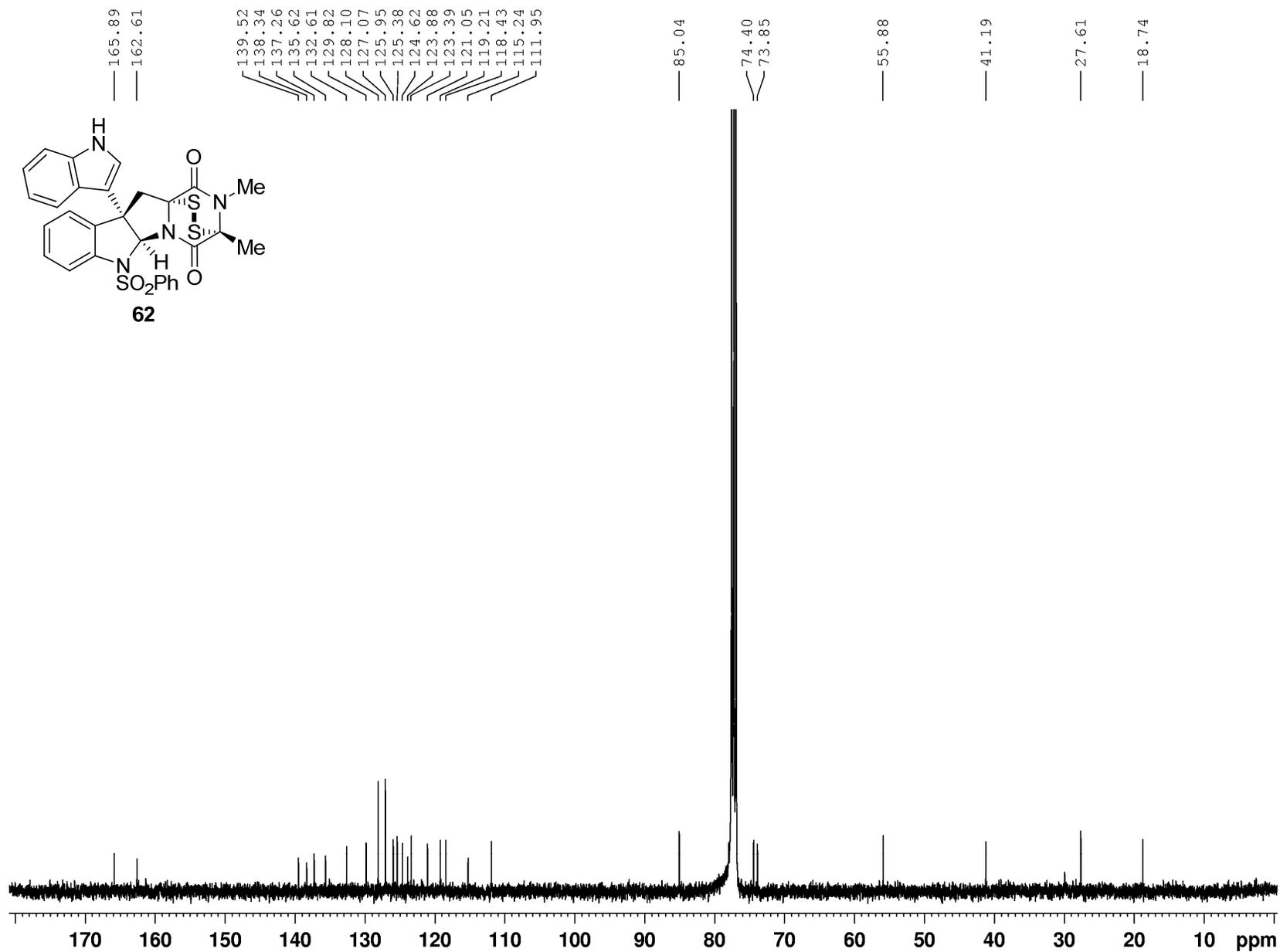






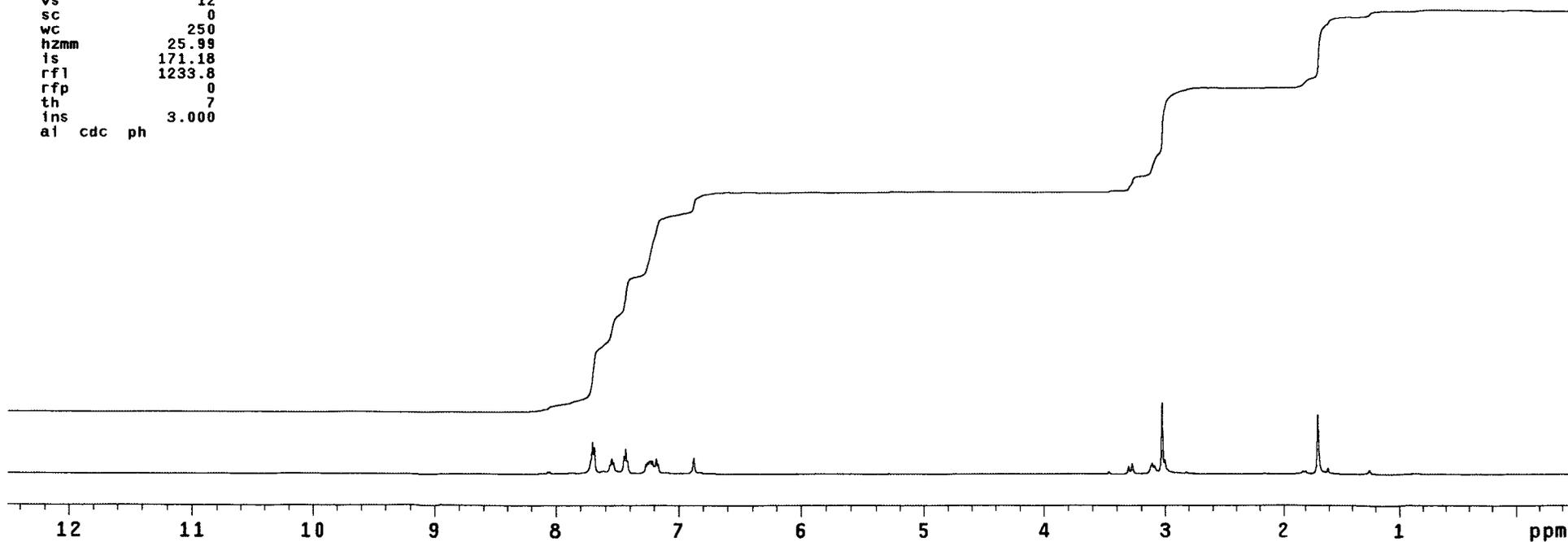
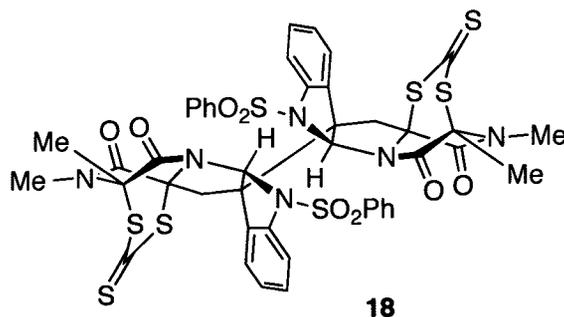






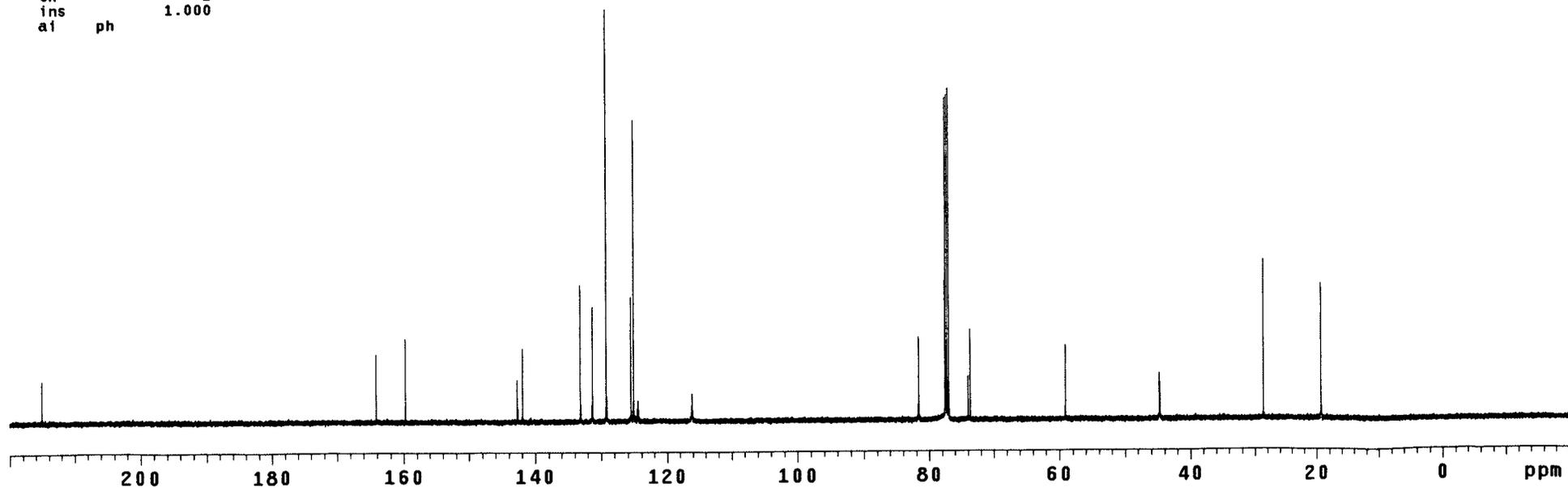
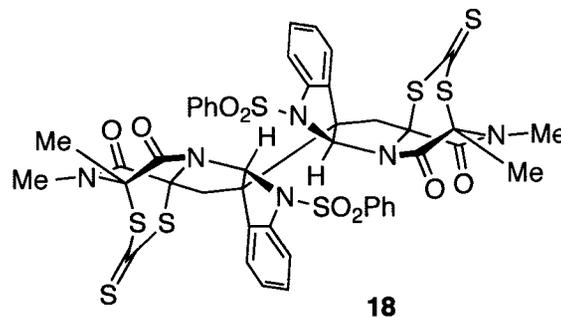
Synthesis and Anticancer Activity of Epipolythiodiketopiperazine Alkaloids
 Nicolas Boyer, Karen C. Morrison, Justin Kim, Paul J. Hergenrother, and Mohammad Movassaghi*

SAMPLE		DEC. & VT	
solvent	CDC13	dfrq	125.672
		dn	C13
		dpwr	30
		dof	0
		dm	nnn
		dmm	w
ACQUISITION		dmf	10000
sfrq	499.746	dseq	
tn	H1	dres	1.0
at	3.001	homo	n
np	63050	PROCESSING	
sw	10504.2	wtfile	
fb	not used	proc	ft
bs	1	fn	262144
tpwr	56	math	f
pw	8.6	werr	
d1	2.000	wexp	
tof	1519.5	wbs	
nt	11111	wnt	
ct	11	wft	
alock	n		
gain	not used		
FLAGS			
il	n		
in	y		
dp	y		
hs	nn		
DISPLAY			
sp	-249.9		
wp	6496.6		
vs	12		
sc	0		
wc	250		
hzmm	25.99		
is	171.18		
rfl	1233.8		
rfp	0		
th	7		
ins	3.000		
al	cdc ph		

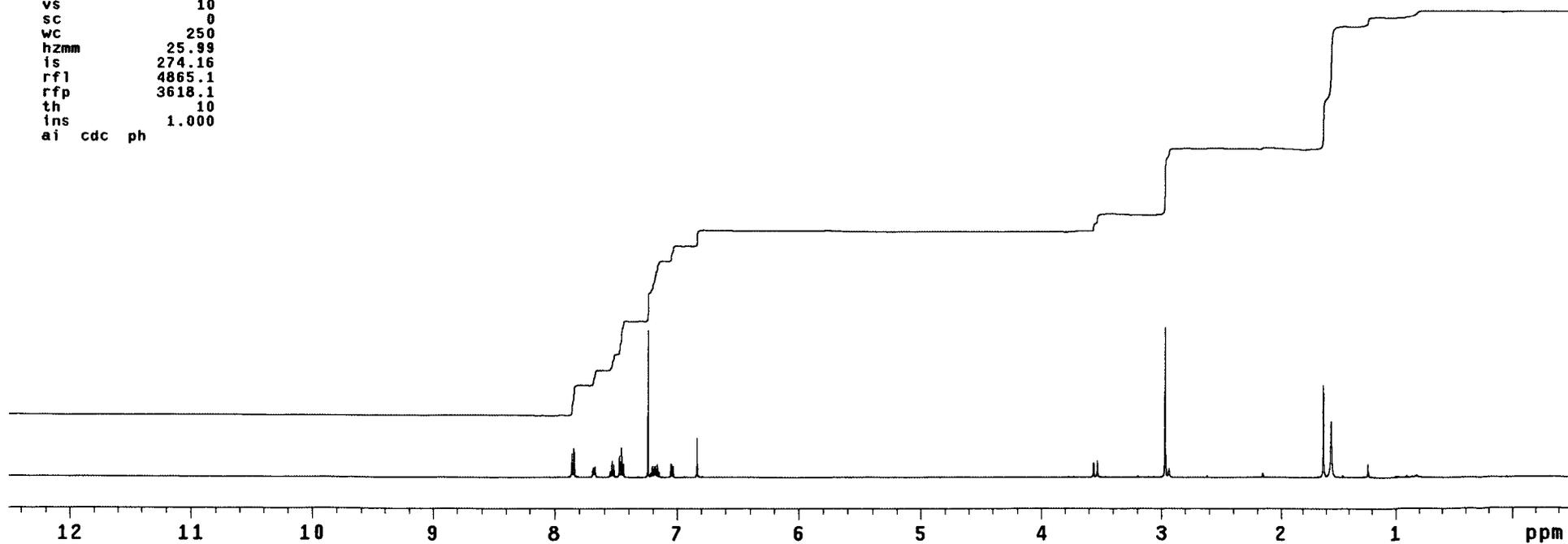
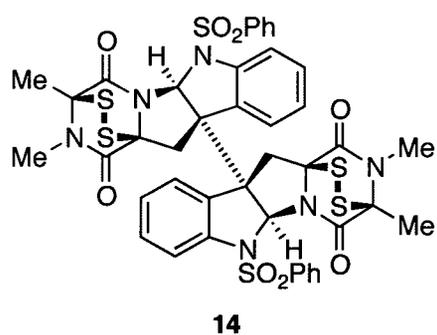


Synthesis and Anticancer Activity of Epipolythiodiketopiperazine Alkaloids
Nicolas Boyer, Karen C. Morrison, Justin Kim, Paul J. Hergenrother, and Mohammad Movassaghi*

SAMPLE		DEC. & VT	
solvent	CDC13	dfrq	500.229
		dn	H1
		dpwr	37
		dof	-500.0
		dm	y
		dmm	w
		dmf	10000
ACQUISITION		PROCESSING	
sfrq	125.795	dseq	1.0
tn	C13	dres	n
at	1.736	homo	n
np	131010	lb	0.30
sw	37735.8	wtfile	
fb	not used	proc	ft
bs	8	fn	131072
ss	1	math	f
tpwr	53		
pw	6.9		
d1	0.763	werr	
tof	631.4	wexp	
nt	1.11111e+07	wbs	
ct	3976	wnt	
alock	n		
gain	60		
FLAGS			
il	n		
in	n		
dp	y		
hs	nn		
DISPLAY			
sp	-2515.9		
wp	30187.6		
vs	1270		
sc	0		
wc	250		
hzmm	120.75		
is	500.00		
rfl	16003.9		
rfp	9714.2		
th	2		
ins	1.000		
ai	ph		

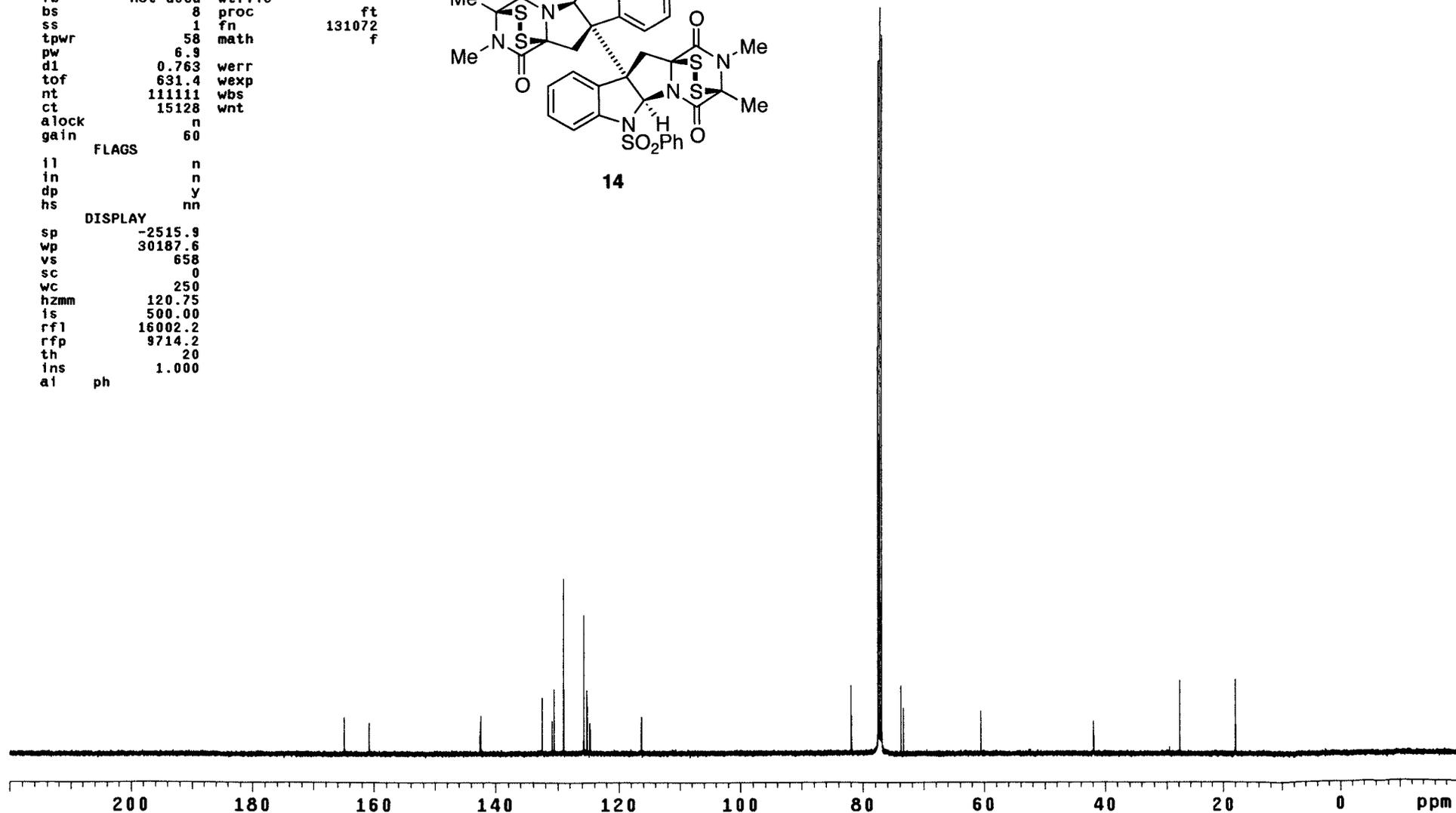
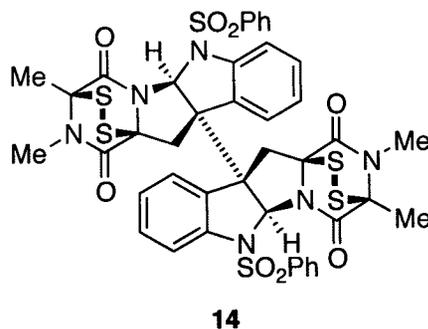


```
SAMPLE          DEC. & VT
solvent         CDC13    dfrq      125.672
                  dn       C13
                  dpwr      30
                  dof       0
                  dm       nnn
                  dmm       w
ACQUISITION     dmf      10000
sfrq           499.746 dseq
tn             H1      dres      1.0
at             3.001  homo      n
np             63050
sw             10504.2 wtfiler
fb             not used proc       ft
bs             1       fn       262144
tpwr           56     math      f
pw             8.6
d1             2.000 werr
tof            1519.5 wexp
nt             11111 wbs
ct             16     wnt
alock          n
gain           not used
                FLAGS
i1             n
in             n
dp             y
hs             nn
DISPLAY
sp            -249.9
wp            6496.6
vs            10
sc            0
wc            250
hzmm         25.99
is            274.16
rf1           4865.1
rfp           3618.1
th            10
ins           1.000
ai cdc ph
```

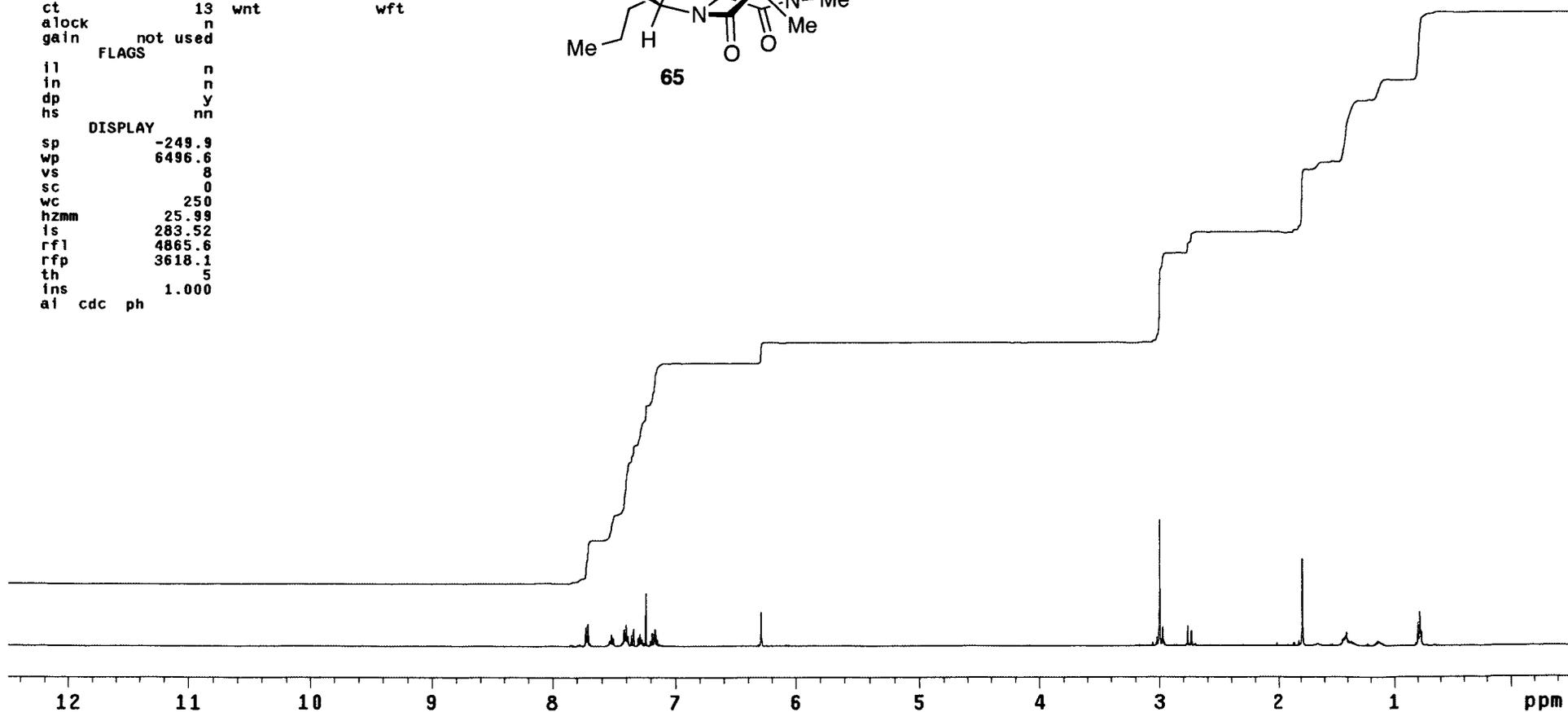
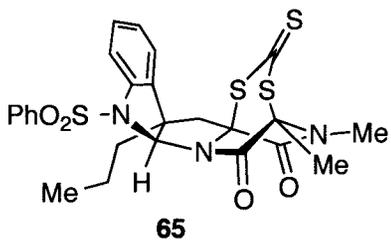


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Nicolas Boyer, Karen C. Morrison, Justin Kim, Paul J. Hergenrother, and Mohammad Movassaghi*

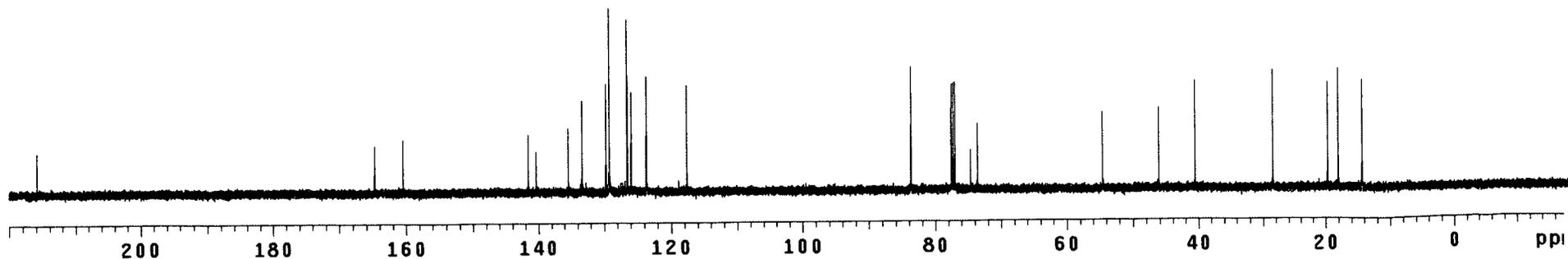
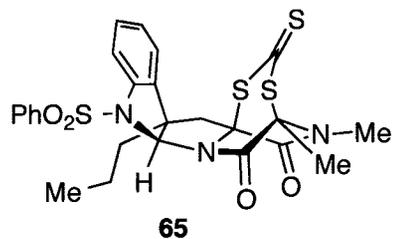
```
SAMPLE          DEC. & VT
solvent         CDC13    dfrq      500.229
                  dn       H1
                  dpwr      40
                  dof      -500.0
                  dm       y
                  dmm      w
ACQUISITION     dmf       10000
sfrq           125.795  dseq
tn             C13     dres      1.0
at            1.736   homo      n
np            131010
sw            37735.8  lb
fb            not used wtfile
bs            8       proc
ss            1       fn         131072
tpwr          58      math      f
pw            6.9
d1            0.763   werr
tof           631.4   wexp
nt            111111  wbs
ct            15128  wnt
alock         n
gain          60
FLAGS
il            n
in            n
dp            y
hs            nn
DISPLAY
sp            -2515.9
wp            30187.6
vs            658
sc            0
wc            250
hzmm         120.75
is            500.00
rf1          16002.2
rfp          9714.2
th            20
ins          1.000
ai           ph
```



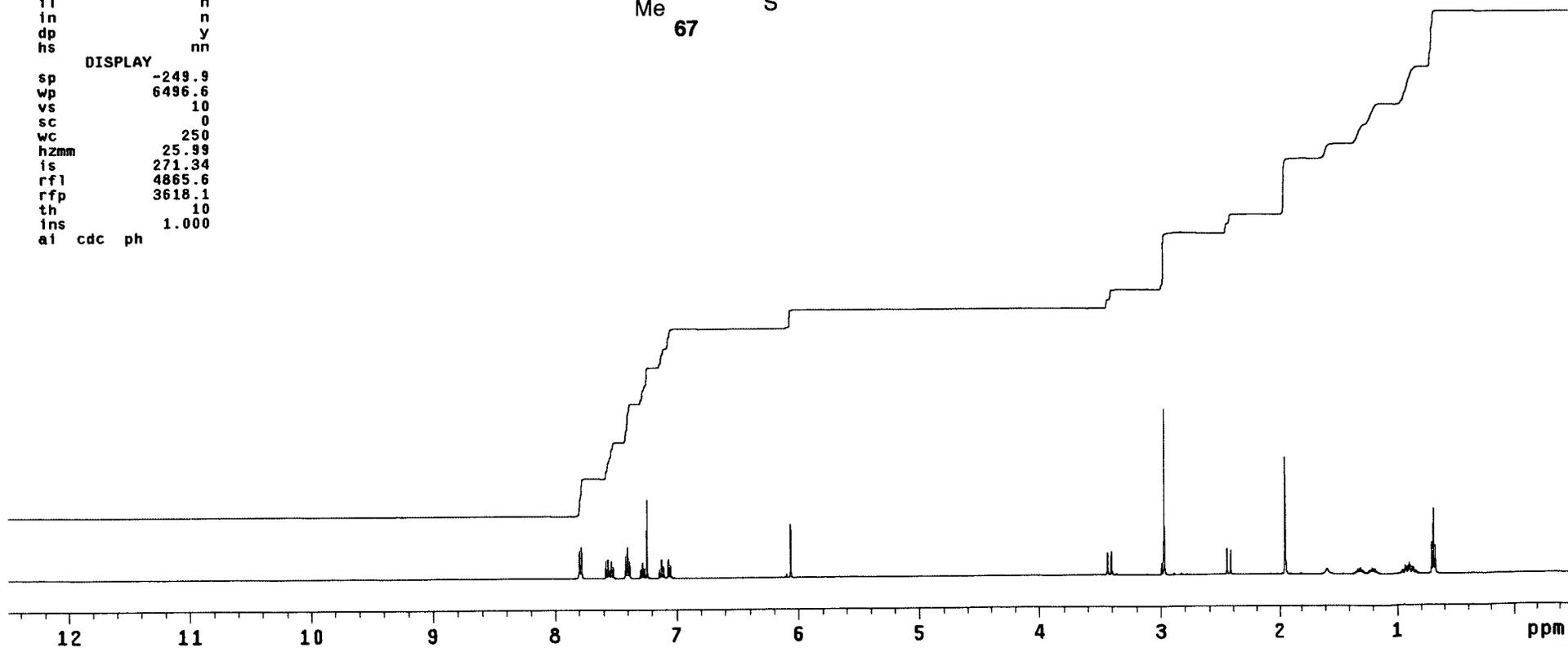
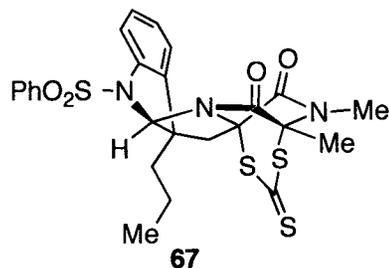
```
SAMPLE          DEC. & VT
solvent          CDC13    dfrq          125.672
                  dn           C13
                  dpwr          30
                  dof           0
                  dm           nnn
                  dmm          w
ACQUISITION      dmf          10000
sfrq            499.746  dseq
tn              H1       dres          1.0
at              3.001    homo          n
np              63050    wtfile
sw              10504.2  wfile
fb              not used proc            ft
bs              1       fn             262144
tpwr            56      math          f
pw              8.6     werr
d1              2.000   wexp
tof             1519.5  wbs
nt              11111   wnt
ct              13
alock           n
gain           not used
FLAGS
il              n
in              n
dp              y
hs              nn
DISPLAY
sp              -249.9
wp              6496.6
vs              8
sc              0
wc              250
hzmm            25.99
is              283.52
rfl             4865.6
rfp             3618.1
th              5
ins             1.000
ai cdc ph
```



```
SAMPLE          DEC. & VT
solvent          CDC13  dfrq      500.229
                  dn        H1
                  dpwr      37
                  dof      -500.0
                  dm        y
                  dmm       w
ACQUISITION     dmf      10000
sfrq           125.795 dseq
tn             C13     dres      1.0
at             1.736  homo      n
np            131010  PROCESSING
sw            37735.8  lb        0.30
fb            not used wtfile
bs             8      proc
ss             1      fn      131072
tpwr           53     math
pw             6.9
d1             0.763  werr
tof            631.4  wexp
nt            1.11111e+06 wbs
ct             104    wnt
alock          n
gain           60
FLAGS
il             n
in             n
dp             y
hs             nn
DISPLAY
sp            -2516.0
wp            30189.9
vs            393
sc             0
wc            250
hzmm          120.76
is            500.00
rfl           16009.9
rfp           9714.9
th             4
ins           1.000
ai            ph
```

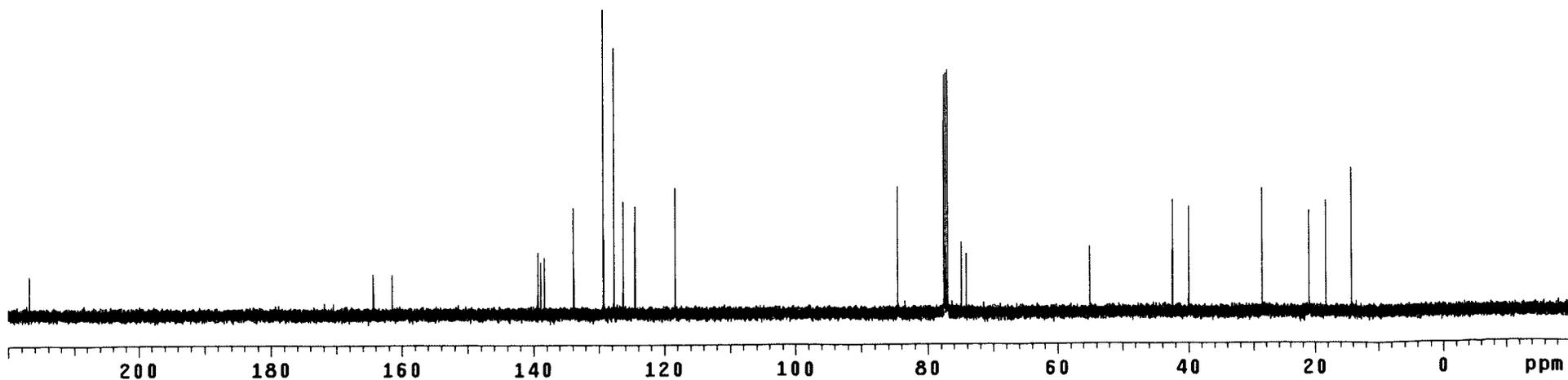
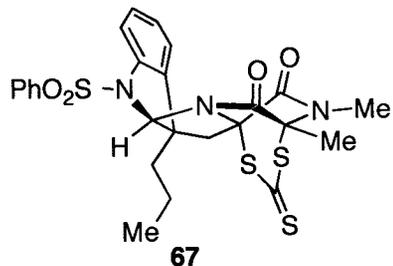


```
SAMPLE          DEC. & VT
solvent          CDC13  dfrq      125.672
                  dn       C13
                  dpwr      30
                  dof       0
                  dm       nnn
                  dmm       w
ACQUISITION     dmf      10000
sfrq           499.746  dseq
tn             H1      dres      1.0
at            3.001    homo
np            63050
sw           10504.2   wtfile
fb           not used  proc      ft
bs            1       fn      262144
tpwr         56      math     f
pw           8.6
d1           2.000   werr
tof          1519.5  wexp
nt           11111   wbs
ct            6     wnt
aLOCK         n
gain         not used
FLAGS
il            n
in            n
dp            y
hs            nn
DISPLAY
sp           -249.9
wp           6496.6
vs            10
sc            0
wc            250
hzmm         25.99
is            271.34
rfl          4865.6
rfp          3618.1
th            10
ins           1.000
ai cdc ph
```



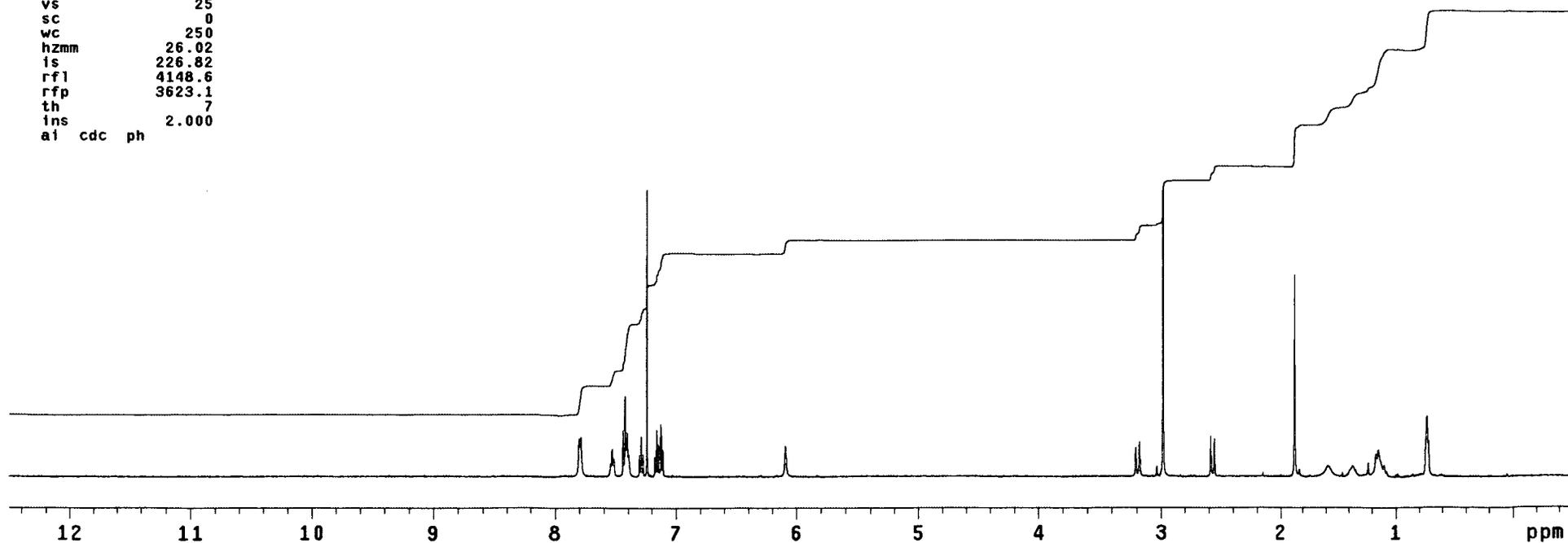
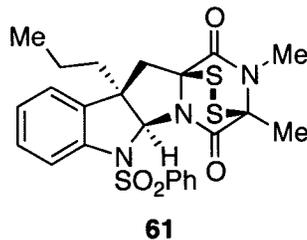
Synthesis and Anticancer Activity of Epipolythiodiketopiperazine Alkaloids
 Nicolas Boyer, Karen C. Morrison, Justin Kim, Paul J. Hergenrother, and Mohammad Movassaghi*

SAMPLE		DEC. & VT	
solvent	CDC13	dfrq	500.229
		dn	H1
		dpwr	37
		dof	-500.0
		dm	y
		dmm	w
		dmf	10000
ACQUISITION		PROCESSING	
sfrq	125.795	dseq	1.0
tn	C13	dres	n
at	1.736	homo	n
np	131010	lb	0.30
sw	37735.8	wtfile	
fb	not used	proc	ft
bs	8	fn	131072
ss	1	math	f
tpwr	53		
pw	6.9		
d1	0.763	werr	
tof	631.4	wexp	
nt	111111	wbs	
ct	224	wnt	
alock	n		
gain	60		
FLAGS			
il	n		
in	n		
dp	y		
hs	nn		
DISPLAY			
sp	-2516.0		
wp	30189.9		
vs	878		
sc	0		
wc	250		
hzmm	120.76		
is	500.00		
rf1	16005.3		
rfp	9714.9		
th	5		
ins	1.000		
ai	ph		

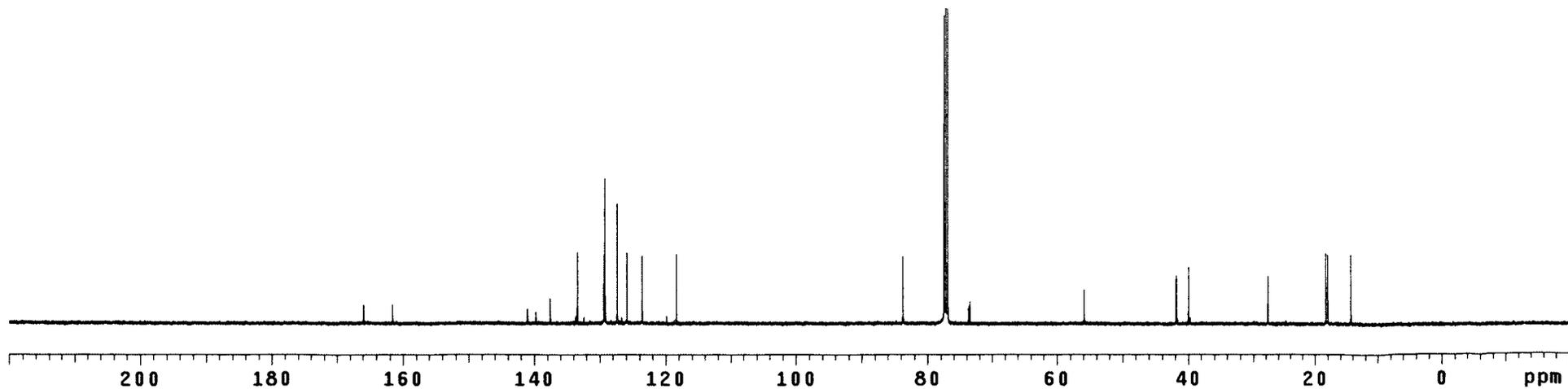
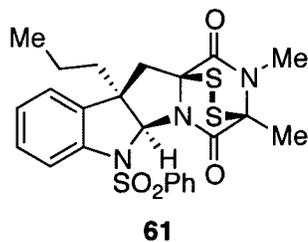


Synthesis and Anticancer Activity of Epipolythiodiketopiperazine Alkaloids
Nicolas Boyer, Karen C. Morrison, Justin Kim, Paul J. Hergenrother, and Mohammad Movassaghi*

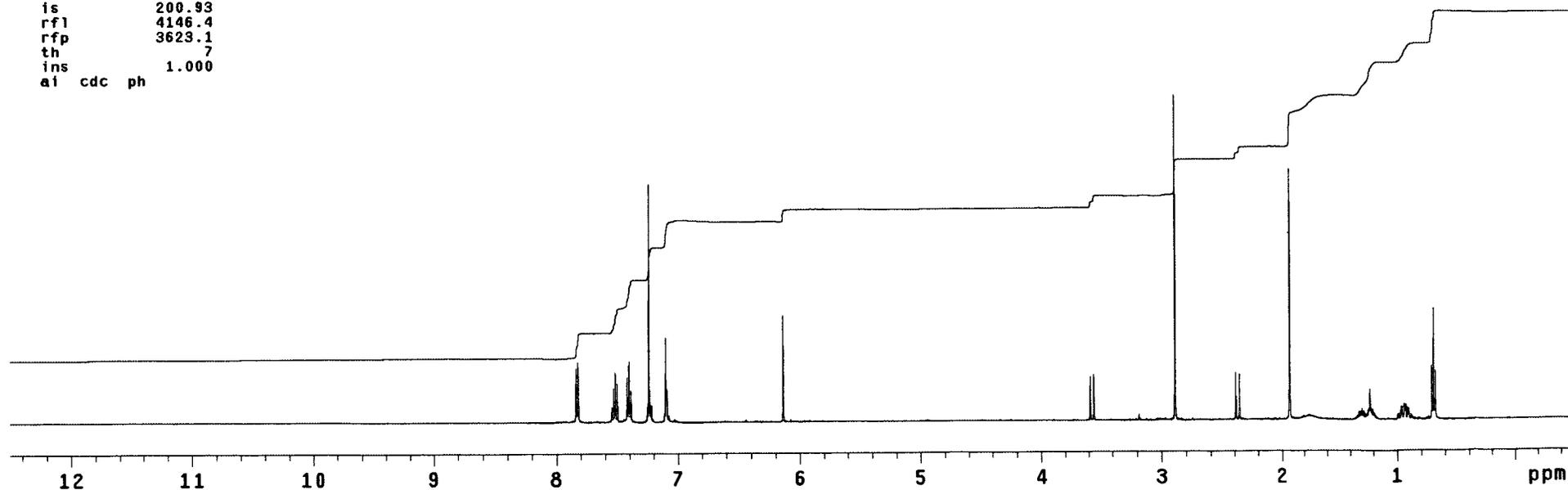
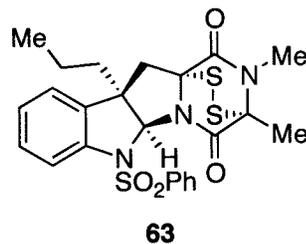
```
SAMPLE          DEC. & VT
date            dfrq      125.844
solvent         CDC13     dn       C13
                dpwr      30
                dof       0
                dm        nnn
                dmm       c
                dmf       200
ACQUISITION
sfrq           500.431  dseq
tn             H1      dres      1.0
at            4.999   homo      n
np            120102  wtfile
sw            12012.0 proc
fb            not used  fn       262144
bs            1      math      f
tpwr          60
pw            8.0
d1            0.100  werr
tof           3003.2  wexp
nt            11111  wbs
ct            8      wnt
alock         not used  wft
gain          n
FLAGS
il            n
in            n
dp            y
hs            nn
DISPLAY
sp            -250.3
wp            6505.5
vs            25
sc            0
wc            250
hzmm         26.02
is            226.82
rfl          4148.6
rfp          3623.1
th            7
ins          2.000
ai cdc ph
```



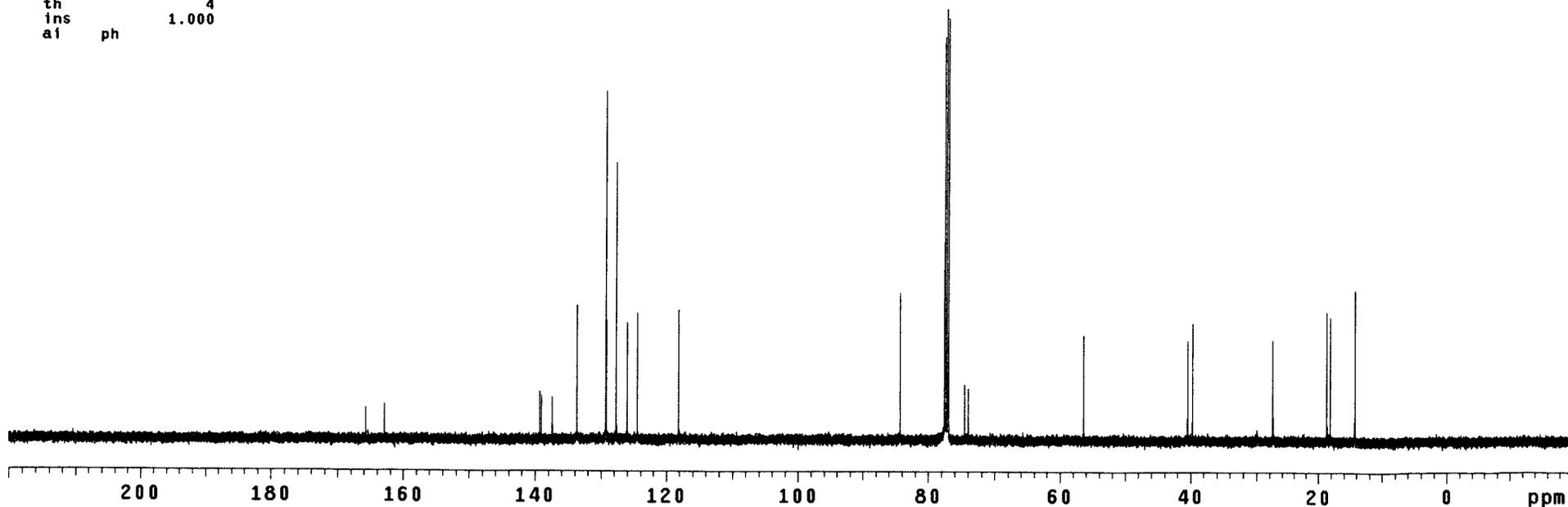
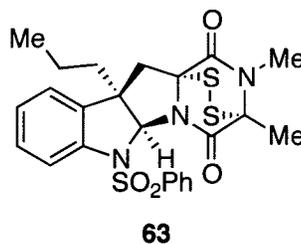
```
SAMPLE          DEC. & VT
solvent          CDC13    dfrq      500.229
                  dn        H1
                  dpwr      40
                  dof       -500.0
                  dm        y
                  dmm       w
                  dmf       10000
ACQUISITION
sfrq            125.795  dseq
tn              C13     dres      1.0
at              1.736   homo
np              131010  PROCESSING
sw              37735.8 lb         1.00
fb              not used wtfile
bs              8       proc
ss              1       fn         131072
tpwr            58      math
pw              6.9
d1              0.763   werr
tof             631.4   wexp
nt              1.1111e+06 wbs
ct              4112    wnt
alock           n
gain           60
FLAGS
il             n
in             n
dp             y
hs            nn
DISPLAY
sp            -2515.9
wp            30187.6
vs            354
sc            0
wc            250
hzmm         120.75
is            500.00
rf1          16002.7
rfp          9714.2
th           20
ins          1.000
ai           ph
```



SAMPLE		DEC. & VT	
solvent	CDC13	dfrq	125.844
		dn	C13
		dpwr	30
		dof	0
		dm	nnn
		dmm	c
		dmf	200
ACQUISITION		PROCESSING	
sfrq	500.431	dseq	1.0
tn	H1	dres	n
at	4.999	homo	n
np	120102	wtfile	
sw	12012.0	proc	ft
fb	not used	fn	262144
bs	1	math	f
tpwr	60		
pw	8.0	werr	
d1	0.100	wexp	
tof	3003.2	wbs	
nt	11111	wnt	wft
ct	8		
alock	n		
gain	not used		
FLAGS			
il	n		
in	n		
dp	y		
hs	nn		
DISPLAY			
sp	-250.3		
wp	6505.5		
vs	17		
sc	0		
wc	250		
hzmm	26.02		
is	200.93		
rfl	4146.4		
rfp	3623.1		
th	7		
ins	1.000		
al	cdc ph		

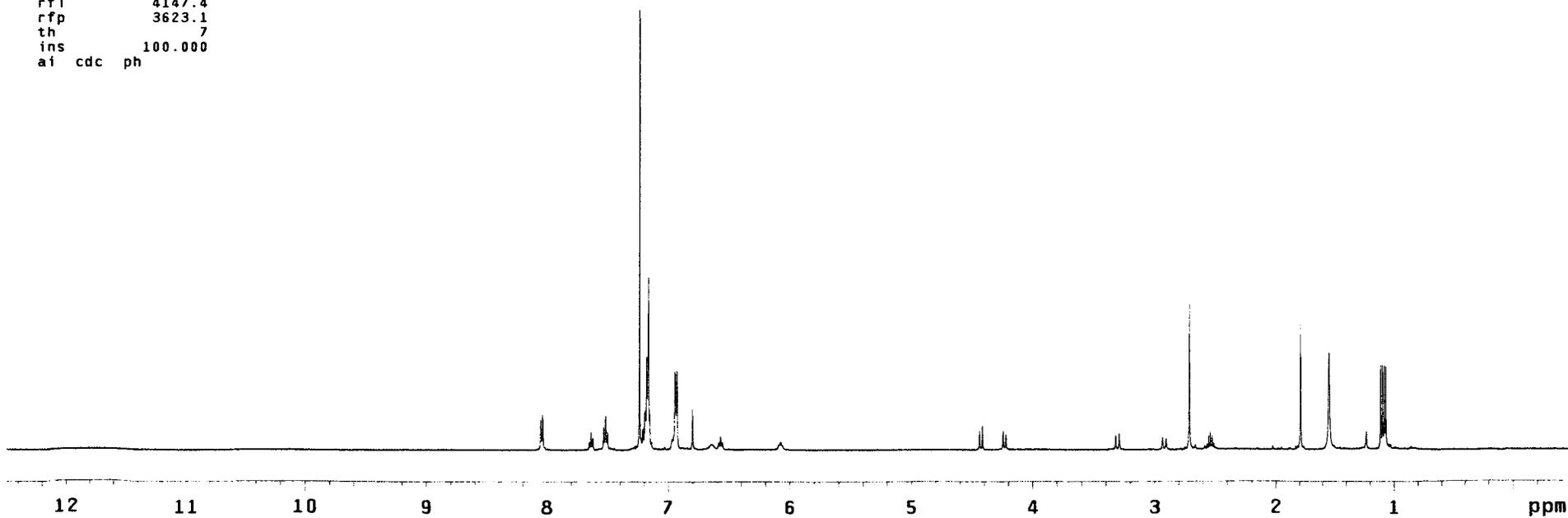
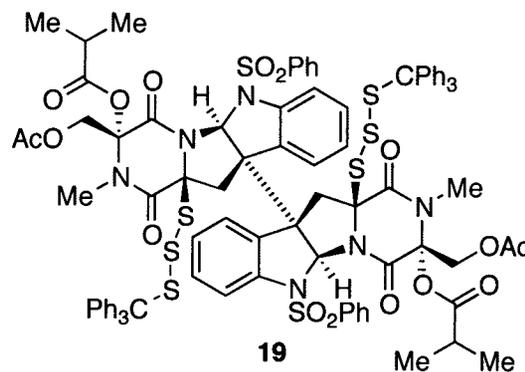


```
SAMPLE          DEC. & VT
solvent         CDC13    dfrq      500.229
                  dn        H1
                  dpwr      37
                  dof       -500.0
                  dm        y
                  dmm       w
ACQUISITION     dmf       10000
sfrq           125.795  dseq
tn             C13      dres      1.0
at             1.736    homo      n
np             131010   PROCESSING
sw             37735.8  lb        0.30
fb             not used wtfile
bs             8        proc      ft
ss             1        fn        131072
tpwr           53       math      f
pw             6.9
d1             0.763    werr
tof            631.4    wexp
nt             111111   wbs
ct             1592     wnt
alock          n
gain           60
FLAGS
il             n
in             n
dp             y
hs            nn
DISPLAY
sp            -2515.9
wp            30187.6
vs            1845
sc            0
wc            250
hzmm         120.75
is            500.00
rfl           16003.3
rfp           9714.2
th            4
ins           1.000
af            ph
```



```

SAMPLE          DEC. & VT
solvent         CDC13      dfrq      125.844
                  dn        C13
                  dpwr      30
ACQUISITION     dof        0
sfrq           500.431   dm         nnn
tn             H1        dmm         c
at            4.999     dmf        200
np            120102    dseq
sw            12012.0   dres       1.0
fb            not used homo
bs            1
tpwr          60       wtfile
pw            8.0      proc
dl            0.100    fn         262144
tof           3003.2   math
nt            1111
ct            60       werr
alock         n        wexp
gain          not used wbs
                FLAGS   wnt
il            n
in            n
dp            y
hs            nn
DISPLAY
sp            -250.3
wp            6505.5
vs            35
sc            0
wc            250
hzmm         26.02
is            33.57
rfl           4147.4
rfp           3623.1
th            7
ins           100.000
ai cdc ph
    
```



```
SAMPLE          DEC. & VT
solvent          CDC13      dfrq      500.229
                  dn        H1
                  dpwr      40
ACQUISITION      dof      -500.0
sfrq            125.795    dm         y
tn              C13       dmm        w
at              1.736     dmf        10000
np              131010    dseq
sw              37735.8   dres       1.0
fb              not used  homo       n
bs              8        PROCESSING
ss              1        lb          0.30
tpwr            58       wtfile
pw              6.9      proc
di              0.763    fn
tof             631.4    math       131072
nt              1.1111e+06  f
ct              13960    werr
alock           n        wexp
gain            60       wbs
                  wnt
FLAGS
il              n
in              n
dp              y
hs              nn
DISPLAY
sp              -2515.9
wp              30189.8
vs              884
sc              0
wc              250
hzmm            120.76
is              500.00
rf1             16002.4
rfp             9714.9
th              20
ins             1.000
ai              ph
```

