Supplementary Information

# Light-triggered strand exchange reaction using the change in the hydrogen bonding pattern of a nucleobase analogue

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# 1. <sup>1</sup>H, <sup>13</sup>C and <sup>31</sup>P spectra of new compounds

## 1-1. <sup>1</sup>H spectrum of compound 2



# 1-2. <sup>13</sup>C spectrum of compound 2



## 1-3. <sup>1</sup>H spectrum of compound 4



# 1-4. <sup>13</sup>C spectrum of compound 4



#### 1-5. <sup>1</sup>H spectrum of compound 5



# 1-6. <sup>13</sup>C spectrum of compound 5



## 1-7. <sup>1</sup>H spectrum of compound 6



# 1-8. <sup>13</sup>C spectrum of compound 6



## 1-9. <sup>1</sup>H spectrum of compound 7



# 1-10. <sup>13</sup>C spectrum of compound 7



## 1-11. <sup>1</sup>H spectrum of compound 8



# 1-12. <sup>13</sup>C spectrum of compound 8



## 1-13. <sup>31</sup>P spectrum of compound 8



#### 2. Molecular modeling for base pairing

Molecular modeling for base-pairing was performed by consecutive molecular mechanics and molecular orbital calculations with Mac Spartan 10' (Wavefunction Inc.). First, a double strand DNA (dsDNA) containing **SB**<sup>NV</sup> was build as a B-type helix using conditions during which only **SB**<sup>NV</sup>-nucleotides atoms could move. Next, every structure generated randomly by rotating every torsion angles concerning **SB**<sup>NV</sup>-nucleotide were optimized under MMFF (aq) force field, which was carried out with the end base-pairings constrained. Finally, every **SB**<sup>NV</sup>-base molety extracted from the optimized structure was further optimized under HF/6-31G\* level. Pictures were drawn with PyMol ver.0.99.



**Fig. S1.** Comparison of **SB**<sup>NV</sup>:G, **SB**:A and natural T:A pair as space filling models.

## 3. HPLC and MALDI-TOF MS analysis of SB<sup>NV</sup>-modified ODNs

#### 3-1. ODN1

**RP-HPLC** 

Column: Waters XBridge<sup>™</sup> OST C18 2.5 µm, 4.6 x 50 mm

Gradient: 6-15% MeCN (over 15 min) in triethylammonium acetate buffer (pH 7.0, 0.1 M)

Flow rate: 1.0 mL/min

Column temperature: 50 °C



#### MALDI-TOF MS

Calcd. 3881.7 [M-H]



#### 3-2. ODN2

**RP-HPLC** 

Column: Waters XBridge<sup>™</sup> OST C18 2.5 μm, 4.6 x 50 mm

Gradient: 6-15% MeCN (over 15 min) in triethylammonium acetate buffer (pH 7.0, 0.1 M)

#### Flow rate: 1.0 mL/min

Column temperature: 50 °C



#### MALDI-TOF MS

#### Calcd. 3867.6 [M-H]



#### 3-3. ODN3

**RP-HPLC** 

Column: Waters XBridge<sup>™</sup> OST C18 2.5 µm, 4.6 x 50 mm

Gradient: 8-24% MeCN (over 15 min) in triethylammonium acetate buffer (pH 7.0, 0.1 M)

#### Flow rate: 1.0 mL/min

Column temperature: 50 °C



#### MALDI-TOF MS

Calcd. 3737.7 [M-H]<sup>-</sup>



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4. Photoreaction of SB<sup>NV</sup>-modified ODNs



**Fig. S2.** Time course of the photoreaction (37 °C) of (A) **ODN1** and (B) **ODN1** with complementary DNA. Error bars indicate standard deviation (n = 3).



Fig. S3. HPLC analysis of photoreaction of ODN2.

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**Fig. S4.** Influence of the reaction temperature on the efficiency of the photoreaction. (A) **ODN1** (B) **ODN1** with complementary DNA. Error bars indicate standard deviation (n = 3).

#### 5. UV melting experiments



Fig. S5. Changes in UV melting curves for the duplexes with DNA targets triggered by light.

Table S1. Melting temperatures of duplex					Table S2.
between <b>ODN1</b> and RNA targets					between O
<b>ODN1:</b> 5'-d(AGCAAAXAACGC)-3'					ODN2: 5
3'-r(UCGUUUYUUGCG)-5'					
<i>T</i> <sub>m</sub> (°C)					
	X:Y	-UV	+UV		X:Y
	SB <sup>NV</sup> :G	25	16		SB <sup>NV</sup> :G
	SB <sup>NV</sup> :A	19	25		SB <sup>NV</sup> :A
	SB <sup>NV</sup> :C	ND	ND		SB <sup>NV</sup> :C
	SB <sup>NV</sup> :U	ND	16		SB <sup>NV</sup> :U
	A:U	19	19		T:A
	G:C	31	31		C:G
				=	

 Table S2. Melting temperatures of duplex

 between ODN2 and RNA targets

 ODN2:
 5'-d(GCGTTXTTTGCT)-3'

3′-	3′-r(CGCAAYAAACGA)-5′			
	T <sub>m</sub> (°C)			
X:Y	-UV	+UV		
SB <sup>NV</sup> :G	36	32		
SB <sup>NV</sup> :A	30	40		
SB <sup>NV</sup> :C	26	29		
SB <sup>NV</sup> :U	28	32		
T:A	40	40		
C:G	46	46		

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Fig. S6. Changes in UV melting curves for the duplexes with RNA targets triggered by light.



Fig. S7. Changes in UV melting curves for the duplexes ODN3/bad and ODN3/bcl-xL triggered by light.

6. Direct observation of light-triggered strand exchange reaction using fluorescent changes



**Fig. S8.** The fluorescence changes of a liquid drop containing **ODN3**-BHQ2, *bad* and *bcl-xL* upon photoirradiation. Conditions: **ODN3**-BHQ2 (12  $\mu$ M), *bad* (8  $\mu$ M), *bcl-xL* (8  $\mu$ M), NaCl (20 mM), sodium phosphate buffer (10 mM, pH 7.2). Irradiation (365 nm) was performed at room temperature. The fluorescence was observed on a UV transilluminator.