

Supplementary Information
for *In-Situ* Generation of Ion-Paired Chiral Ligands: Rapid Identification of Optimal Ligand for
Palladium-Catalyzed Asymmetric Allylation

Kohsuke Ohmatsu,^a Yoshiyuki Hara,^a and Takashi Ooi^{*a,b}

^a *Institute of Transformative Bio-Molecules (WPI-ITbM), and Department of Applied Chemistry,*
Graduate School of Engineering, Nagoya University, Nagoya, 464-8603, Japan.

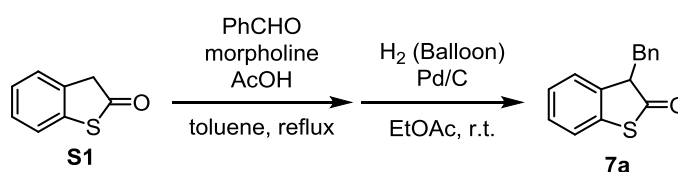
^b *CREST, Japan Science and Technology Agency (JST), Nagoya, 464-8603, Japan.*

too@apchem.nagoya-u.ac.jp

General Information: Infrared spectra were recorded on a Shimadzu IRAffinity-1 spectrometer. ¹H NMR spectra were recorded on a JEOL JNM-ECS400 (400 MHz) spectrometer. Chemical shifts are reported in ppm from the tetramethylsilane (0.0 ppm) resonance as the internal standard (CDCl₃). Data are reported as follows: chemical shift, integration, multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, sept = septet, m = multiplet) and coupling constants (Hz). ¹³C NMR spectra were recorded on a JEOL JNM-ECS400 (101 MHz) spectrometer with complete proton decoupling. Chemical shifts are reported in ppm from the solvent resonance as the internal standard (CDCl₃; 77.16 ppm). ³¹P NMR spectra were recorded on a JEOL JNM-ECS400 (162 MHz) spectrometer with complete proton decoupling. Chemical shifts are reported in ppm from H₃PO₄ (0.0 ppm) resonance as the external standard. Optical rotations were measured on a HORIBA SEPA-500 polarimeter. The high resolution mass spectra were measured on a Thermo Fisher Scientific Exactive (ESI). Analytical thin layer chromatography (TLC) was performed on Merck precoated TLC plates (silica gel 60 GF254, 0.25 mm). Flash column chromatography was performed on PSQ60AB (spherical, 40-50 μm; FUJI SILYSIA CHEMICAL Co., Inc.). Enantiomeric excesses were determined by HPLC analysis using chiral columns (φ 4.6 mm x 250 mm, DAICEL CHIRALCEL OD-3 (OD3), CHIRALCEL OZ-3 (OZ3), CHIRALPAC IC-3 (IC3) and CHIRALPAC AD-3 (AD3)) with hexane (H) and isopropyl alcohol (IPA) as eluent.

All air- and moisture-sensitive reactions were performed under an atmosphere of argon (Ar) in dried glassware. The manipulations for Pd-catalyzed reactions were carried out with standard Schlenk techniques under Ar. Toluene was supplied from Kanto Chemical Co., Inc. as “Dehydrated” and further purified by both A2 alumina and Q5 reactant using a GlassContour solvent dispensing system. Allylic carbonates **5** were synthesized from the corresponding allylic alcohols.¹ Other simple chemicals were purchased and used as such.

Representative Procedures for Synthesis of Benzothiophenones 7:

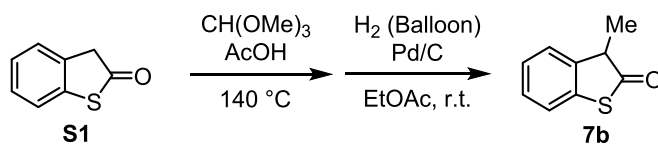


To a solution of **S1**² (891 mg, 3.74 mmol) in toluene (10 mL) were added benzaldehyde (0.76 mL, 7.48 mmol), morpholine (0.032 mL, 0.37 mmol), and acetic acid (0.021 mL, 0.37 mmol), and whole reaction

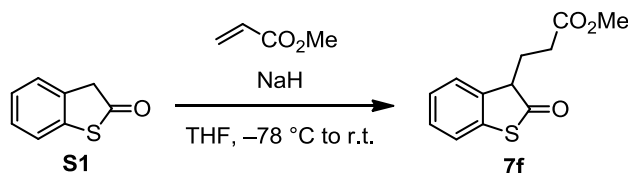
¹ R. Matsubara, T. F. Jamison, *J. Am. Chem. Soc.* **2010**, *132*, 6880.

² W. Chen, Y. Shi, H. Feng, M. Du, J. Z. Zhang, J. Hu, D. Yang, *J. Phys. Chem. B*, **2012**, *116*, 9231.

mixture was refluxed with stirring for 12 h. After cooling to room temperature, the reaction mixture was filtered through a short pad of silica gel with the aid of EtOAc, and then concentrated. The resulting residue was dissolved in EtOAc (10 mL), and the flask was degassed by alternating vacuum evacuation/Ar backfill. After successive addition of 10% Pd/C (400 mg) at 0 °C, the reaction flask was evacuated again and refilled with H₂ three times. The resulting suspension was stirred for 12 h at room temperature. After flushing the remaining H₂ out with Ar, the mixture was filtered to remove Pd/C and the filtercake was rinsed with EtOAc. The filtrate was evaporated and the residue was purified by silica gel column chromatography (H/EtOAc = 30:1 as eluent) to give **7a** (737 mg, 3.07 mmol, 82% yield) as an off-yellow oil. Benzothiophenones (**7c**, **7d**, **7e**, **7g**, **7h** and **7i**) were synthesized by following the same procedure. **7a**: ¹H NMR (400 MHz, CDCl₃) δ 7.28-7.18 (5H, m), 7.12 (1H, ddd, *J* = 7.8, 6.8, 1.9 Hz), 7.08-7.05 (2H, m), 6.90 (1H, d, *J* = 7.8 Hz), 4.07 (1H, dd, *J* = 7.8, 4.8 Hz), 3.42 (1H, dd, *J* = 14.0, 4.8 Hz), 3.16 (1H, dd, *J* = 14.0, 7.8 Hz); ¹³C NMR (101 MHz, CDCl₃) δ 206.1, 136.7, 136.2, 136.1, 129.7, 128.5, 128.5, 127.0, 125.9, 125.5, 123.1, 58.3, 38.9; IR (film) 3063, 3028, 2922, 1703, 1466, 1449, 1022, 748, 737, 696 cm⁻¹; HRMS (ESI, negative ion mode) Calcd for C₁₅H₁₁O₁S₁⁻ ([M-H]⁻) 239.0525. Found 239.0531.



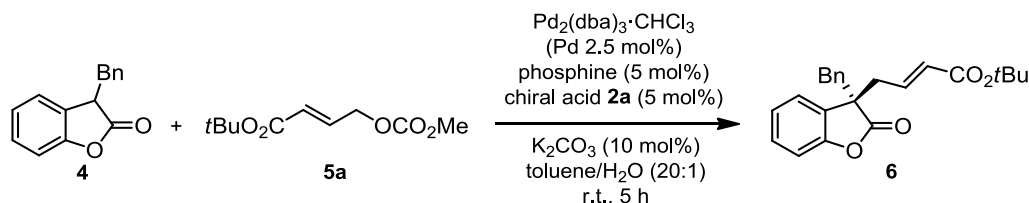
A round-bottomed flask was charged with **S1** (751mg, 5.0 mmol) and trimethyl orthoformate (2.5 mL, 15 mmol) in acetic acid (1.9 mL, 20 mmol). The solution was stirred at 140 °C in an oil bath for 12 h. After cooling to room temperature, the reaction mixture was concentrated in vacuo. Extractive work-up was conducted with EtOAc three times, and the combined organic phases were washed with brine, dried over Na₂SO₄, and evaporated. To a solution of the resulting residue in EtOAc (3 mL) was added 10% Pd/C (87 mg) at 0 °C under Ar, the reaction flask was evacuated again and refilled with H₂ three times. The resulting suspension was stirred for 12 h at room temperature. After flushing the remaining H₂ out with Ar, the mixture was filtered to remove Pd/C and the filtercake was rinsed with EtOAc. The filtrate was evaporated and the residue was purified by silica gel column chromatography (H/EtOAc = 10:1 as eluent) to give **7b** (95 mg, 0.58 mmol, 71% yield) as a colorless oil. **7b**: ¹H NMR (400 MHz, CDCl₃) δ 7.36 (1H, d, *J* = 7.8 Hz), 7.32-7.22 (3H, m), 3.83 (1H, q, *J* = 7.8 Hz), 1.55 (3H, d, *J* = 7.8 Hz); ¹³C NMR (101 MHz, CDCl₃) δ 207.0, 138.3, 135.5, 128.5, 126.4, 124.6, 123.1, 52.2, 17.4; IR (film) 2978, 2932, 1711, 1468, 1445, 1101, 1005, 924, 750, 696 cm⁻¹; HRMS (ESI, negative ion mode) Calcd for C₉H₇O₁S₁⁻ ([M-H]⁻) 163.0223. Found 163.0211.



A dried two neck flask was charged with NaH (67 mg, 1.68 mmol) in THF (2 mL) under Ar. To this suspension was dropwised a solution of **S1** (253 mg, 1.68mmol) in THF (2 mL) at 0 °C, and the resulting mixture was stirred for 30 min at the same temperature. The reaction solution was then slowly transferred via cannula into a cooled solution of methyl acrylate (0.076 mL, 0.84 mmol) in THF (2 mL) at -78 °C. After stirring for 15 min, the reaction mixture was allowed to warm gradually to room temperature and quenched

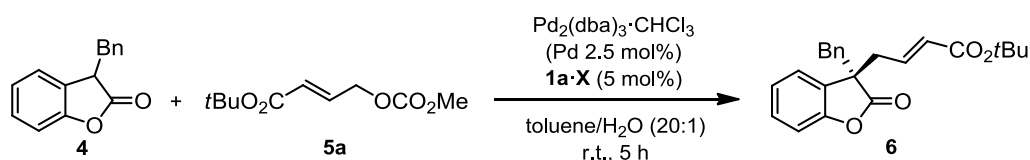
with saturated aqueous NH_4Cl (10 mL). Extractive work-up was conducted with EtOAc three times. The combined organic extracts were dried over MgSO_4 and concentrated in vacuo. The residue was purified by flash chromatography on silica gel (10:1 H/EtOAc) to afford **7f** (79 mg, 0.33 mmol, 40% yield) as a red liquid. **7f**: ^1H NMR (400 MHz, CDCl_3) δ 7.35 (1H, d, $J = 7.3$ Hz), 7.33-7.26 (2H, m), 7.24 (1H, dd, $J = 8.2, 1.2$ Hz), 3.90 (1H, dd, $J = 6.0, 4.1$ Hz), 3.63 (3H, s), 2.51-2.2.40 (2H, m), 2.34-2.23 (2H, m); ^{13}C NMR (101 MHz, CDCl_3) δ 206.0, 173.2, 136.1, 135.9, 128.7, 126.5, 124.9, 123.2, 55.7, 51.8, 29.5, 27.6; IR (film) 2951, 1732, 1703, 1593, 1447, 1437, 1211, 1175, 1009, 748 cm^{-1} ; HRMS (ESI, negative ion mode) Calcd for $\text{C}_{12}\text{H}_{11}\text{O}_3\text{S}_1^-$ ($[\text{M}-\text{H}]^-$) 235.0423. Found 235.0427.

General Procedure for *In-Situ* Generation of Ion-Paired Chiral Ligand



To a Schlenk flask were added 3-benzylbenzofuranone **4** (22.4 mg, 0.1 mmol), $\text{Pd}_2(\text{dba})_3 \cdot \text{CHCl}_3$ (1.29 mg, 0.0013 mmol), ammonium phosphine **1a**· HSO_4 (2.5 mg, 0.005 mmol), chiral phosphoric acid **2a** (3.12 mg, 0.005 mmol) and K_2CO_3 (1.38 mg, 0.01 mmol) and the flask was degassed by alternating vacuum evacuation/Ar backfill. Then, toluene (1 mL) was added, and the resulting catalyst mixture was evacuated and refilled with Ar three times. After addition of H_2O (0.05 mL), allylic carbonate **5a** (21.6 mg, 0.1 mmol) was successively introduced at room temperature. After stirring for 5 h at the same temperature, the reaction mixture was directly subjected to the purification by column chromatography on silica gel (H/EtOAc = 10:1 as eluent) to afford **6** (34.6 mg, 0.095 mmol, 95% yield) as a colorless liquid.

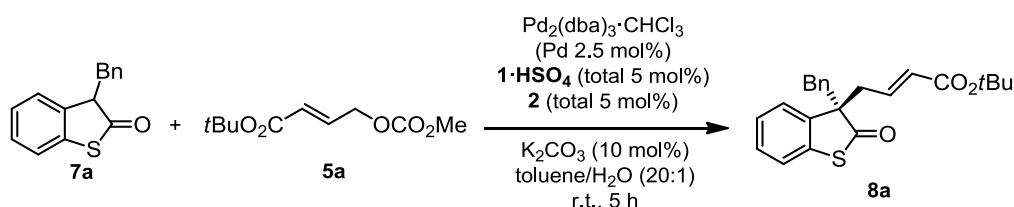
Additional Control Experiments: To verify the activity of the catalyst prepared from $\text{Pd}_2(\text{dba})_3 \cdot \text{CHCl}_3$ and precursors **1a**·**X**, the reactions of **4** with **5a** were conducted under the similar conditions described in Table 1 without chiral acid **2a** and K_2CO_3 . The results were summarized in Table S1. The reason why the reaction using **1a**·**I** solely as a ligand showed higher conversion than that of the reaction with **1a**·**I**, **2a**, and K_2CO_3 (Table S1, entry 2 vs Table 1, entry 3) is unclear at present. While bond formation did not take place at all when **1a**· HSO_4 was used as a ligand (entry 4), an attempted reaction with the same ligand in the absence of H_2O furnished allylated benzofuranone **6** in moderate yield (entry 5). These results suggested that **1a**· HSO_4 would dissolve in water and thus could not form the corresponding Pd complex without ion-exchange event.

Table S1. Palladium-catalyzed allylation of **4** with **5a** using achiral **1a·X** as a ligand.^[a]

entry	phosphine	yield [%] ^[b]
1	1a·Br	62
2	1a·I	88
3	1a·OAc	90
4	1a·HSO₄	0
5 ^[c]	1a·HSO₄	47

^[a] Reactions were carried out with 0.10 mmol of **4** and 0.10 mmol of **5a** in the presence of Pd₂(dba)₃·CHCl₃ (Pd 2.5 mol%) and **1a·X** (5 mol%) in toluene (1.0 mL)/H₂O (0.05 mL) at room temperature. ^[b] Isolated yield of **6**. ^[c] The reaction was conducted without H₂O.

General Procedure for Asymmetric Allylation – Combinatorial Catalyst Screening



To a Schlenk flask were added 3-benzylbenzothiophenone **7a** (the amount is shown below), Pd₂(dba)₃·CHCl₃, ammonium phosphines **1·HSO₄**, chiral phosphoric acids **2** and K₂CO₃, and the flask was degassed by alternating vacuum evacuation/Ar backfill. Then, toluene was added, and the resulting catalyst mixture was evacuated and refilled with Ar three times. After addition of H₂O, allylic carbonate **5a** was successively introduced at room temperature. After stirring for 5 h at the same temperature, the reaction mixture was directly subjected to the purification by column chromatography on silica gel (H/EtOAc = 10:1 as eluent) to afford **8a** as a colorless liquid.

Step 1:

benzothiophenone 7a	0.4 mmol	1.0 equiv
allylic carbonate 5a	0.4 mmol	1.0 equiv
Pd ₂ (dba) ₃ ·CHCl ₃	0.0052 mmol	2.5 mol%
4 × ammonium phosphine 1·HSO₄	each 0.005 mmol	5 mol%
6 × phosphoric acid 2	each 0.0033 mmol	5 mol%
K ₂ CO ₃	0.04 mmol	10 mol%
toluene 4.0 mL, H ₂ O 0.2 mL		

Step 2:

benzothiophenone 7a	0.2 mmol	1.0 equiv
allylic carbonate 5a	0.2 mmol	1.0 equiv
Pd ₂ (dba) ₃ ·CHCl ₃	0.0026 mmol	2.5 mol%
2 × ammonium phosphine 1·HSO₄	each 0.005 mmol	5 mol%

3 × phosphoric acid **2** each 0.0033 mmol 5 mol%
 K₂CO₃ 0.02 mmol 10 mol%
 toluene 2.0 mL, H₂O 0.1 mL

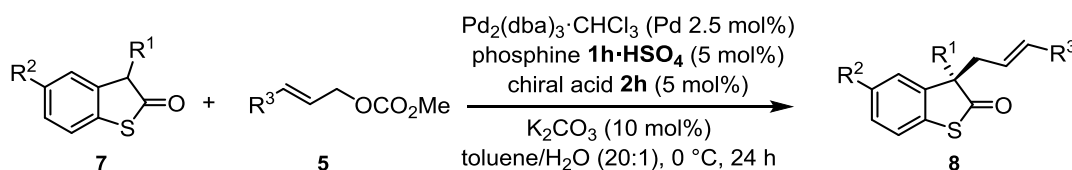
Step 3:

benzothiophenone **7a** 0.1 mmol 1.0 equiv
 allylic carbonate **5a** 0.1 mmol 1.0 equiv
 Pd₂(dba)₃·CHCl₃ 0.0013 mmol 2.5 mol%
 ammonium phosphine **1**·H₂SO₄ 0.005 mmol 5 mol%
 phosphoric acid **2** 0.005 mmol 5 mol%
 K₂CO₃ 0.01 mmol 10 mol%
 toluene 1.0 mL, H₂O 0.05 mL

Table S2. The results of all possible 144 combinations of **1**·H₂SO₄ and **2**

XX% ee (yield)	1a	1b	1c	1d	1e	1f	1g	1h	1i	1j	1k	1l
2a	79% ee (99%)	73% ee (99%)	83% ee (92%)	90% ee (91%)	88% ee (94%)	85% ee (96%)	90% ee (96%)	88% ee (99%)	80% ee (88%)	55% ee (86%)	81% ee (92%)	78% ee (99%)
2b	84% ee (94%)	74% ee (99%)	85% ee (93%)	92% ee (94%)	89% ee (96%)	88% ee (92%)	94% ee (99%)	94% ee (97%)	80% ee (88%)	57% ee (94%)	82% ee (90%)	70% ee (93%)
2c	70% ee (94%)	60% ee (90%)	76% ee (90%)	58% ee (89%)	67% ee (92%)	43% ee (86%)	86% ee (95%)	84% ee (83%)	64% ee (85%)	22% ee (88%)	55% ee (85%)	77% ee (84%)
2d	65% ee (94%)	52% ee (90%)	50% ee (90%)	68% ee (89%)	74% ee (99%)	66% ee (88%)	52% ee (98%)	78% ee (82%)	65% ee (85%)	13% ee (99%)	58% ee (85%)	35% ee (89%)
2e	56% ee (91%)	54% ee (93%)	62% ee (99%)	64% ee (90%)	71% ee (96%)	60% ee (88%)	65% ee (96%)	67% ee (86%)	60% ee (87%)	49% ee (96%)	52% ee (84%)	62% ee (92%)
2f	56% ee (86%)	77% ee (84%)	61% ee (92%)	68% ee (83%)	69% ee (90%)	62% ee (90%)	66% ee (96%)	67% ee (81%)	42% ee (83%)	56% ee (91%)	60% ee (80%)	65% ee (81%)
2g	70% ee (95%)	77% ee (90%)	79% ee (99%)	84% ee (93%)	88% ee (90%)	90% ee (88%)	89% ee (95%)	92% ee (89%)	83% ee (89%)	62% ee (96%)	83% ee (95%)	66% ee (90%)
2h	72% ee (99%)	83% ee (96%)	73% ee (92%)	87% ee (95%)	88% ee (99%)	92% ee (91%)	78% ee (89%)	94% ee (99%)	80% ee (93%)	64% ee (89%)	76% ee (95%)	86% ee (99%)
2i	74% ee (83%)	57% ee (93%)	67% ee (99%)	85% ee (99%)	65% ee (97%)	67% ee (88%)	70% ee (90%)	79% ee (86%)	69% ee (93%)	57% ee (83%)	67% ee (80%)	70% ee (90%)
2j	59% ee (88%)	61% ee (92%)	57% ee (89%)	71% ee (94%)	69% ee (97%)	71% ee (82%)	71% ee (90%)	78% ee (87%)	60% ee (88%)	39% ee (96%)	66% ee (87%)	40% ee (89%)
2k	50% ee (99%)	54% ee (92%)	31% ee (91%)	47% ee (91%)	52% ee (99%)	54% ee (90%)	59% ee (95%)	47% ee (93%)	35% ee (94%)	47% ee (90%)	42% ee (94%)	53% ee (90%)
2l	62% ee (74%)	65% ee (82%)	58% ee (83%)	62% ee (80%)	58% ee (85%)	64% ee (88%)	50% ee (80%)	61% ee (84%)	57% ee (86%)	61% ee (80%)	58% ee (82%)	40% ee (93%)

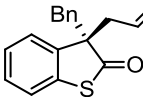
General Procedure for Asymmetric Allylation of Benzothiophenones **7**:

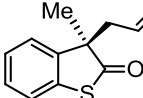


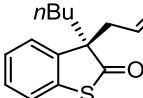
To a Schlenk flask were added benzothiophenone **7a** (22.4 mg, 0.1 mmol), Pd₂(dba)₃·CHCl₃ (1.29 mg, 0.0013 mmol), ammonium phosphine **1h**·H₂SO₄ (2.5 mg, 0.005 mmol), chiral phosphoric acid **2h** (3.12 mg, 0.005 mmol) and K₂CO₃ (1.38 mg, 0.01 mmol), and the flask was degassed by alternating vacuum

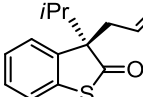
evacuation/Ar backfill. Then, toluene (1 mL) was added, and the resulting catalyst mixture was evacuated and refilled with Ar three times. After addition of H₂O (0.05 mL), allylic carbonate **5a** (21.6 mg, 0.1 mmol) was successively introduced at 0 °C. After stirring for 24 h at the same temperature, the reaction mixture was directly subjected to the purification by column chromatography on silica gel (H/EtOAc = 10:1 as eluent) to afford **8a** (36.1 mg, 0.095 mmol, 95% yield) as a colorless liquid.

Characterization Data for the Alkylated Product 8:

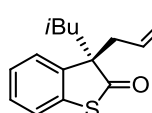
 **8a**: $[\alpha]_D^{21} = -27.8$ ($c = 0.94$, CHCl₃) for 97% ee; ¹H NMR (400 MHz, CDCl₃) δ 7.29-7.16 (4H, m), 7.13-7.04 (3H, m), 6.79 (1H, dd, $J = 8.2, 1.8$ Hz), 6.43 (1H, ddd, $J = 15.6, 8.4, 6.8$ Hz), 5.73 (1H, d, $J = 15.6$ Hz), 3.24 (1H, d, $J = 13.3$ Hz), 3.07 (1H, d, $J = 13.3$ Hz), 2.92 (1H, dd, $J = 14.2, 8.4$ Hz), 2.76 (1H, dd, $J = 14.2, 6.8$ Hz), 1.40 (9H, s); ¹³C NMR (101 MHz, CDCl₃) δ 208.6, 165.2, 140.1, 137.9, 135.7, 134.6, 130.1, 128.8, 128.0, 127.2, 127.0, 126.2, 125.0, 123.2, 80.5, 64.6, 46.0, 41.7, 28.2; IR (film) 2978, 2930, 1705, 1653, 1368, 1152, 980, 922, 735, 700 cm⁻¹; HRMS (ESI, positive ion mode) Calcd for C₂₃H₂₄O₃S₁Na⁺ ([M+Na]⁺) 403.1338. Found 403.1340.; HPLC OZ3, H/IPA = 10:1, flow rate = 0.5 mL/min, $\lambda = 210$ nm, 10.3 min (minor), 11.7 min (major).

 **8b**: $[\alpha]_D^{21} = -50.5$ ($c = 1.0$, CHCl₃) for 93% ee; ¹H NMR (400 MHz, CDCl₃) δ 7.37 (1H, dd, $J = 7.3, 1.8$ Hz), 7.29 (1H, dt, $J = 7.3, 1.8$ Hz), 7.26 (1H, dt, $J = 7.3, 1.8$ Hz), 7.18 (1H, dd, $J = 7.3, 1.8$ Hz), 6.47 (1H, ddd, $J = 15.6, 8.6, 7.0$ Hz), 5.70 (1H, dd, $J = 15.6, 1.4$ Hz), 2.74 (1H, ddd, $J = 14.2, 8.6, 1.4$ Hz), 2.63 (1H, ddd, $J = 14.2, 7.0, 1.4$ Hz), 1.46 (3H, s), 1.42 (9H, s); ¹³C NMR (101 MHz, CDCl₃) δ 208.7, 165.3, 140.5, 140.4, 134.4, 128.7, 127.1, 126.7, 124.2, 123.3, 80.5, 59.2, 42.4, 28.2, 25.3; IR (film) 2976, 2930, 1707, 1653, 1368, 1337, 1155, 984, 955, 758 cm⁻¹; HRMS (ESI, positive ion mode) Calcd for C₁₇H₂₀O₃S₁Na⁺ ([M+Na]⁺) 327.1025. Found 327.1024.; HPLC IC3, H/IPA = 10:1, flow rate = 0.5 mL/min, $\lambda = 210$ nm, 18.0 min (major), 29.0 min (minor).

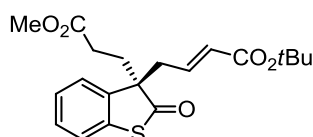
 **8c**: $[\alpha]_D^{21} = +41.4$ ($c = 1.0$, CHCl₃) for 94% ee; ¹H NMR (400 MHz, CDCl₃) δ 7.37 (1H, dd, $J = 7.4, 1.8$ Hz), 7.30 (1H, dt, $J = 7.4, 1.8$ Hz), 7.27 (1H, dt, $J = 7.4, 1.8$ Hz), 7.15 (1H, dd, $J = 7.4, 1.8$ Hz), 6.44 (1H, ddd, $J = 15.8, 8.6, 6.9$ Hz), 5.68 (1H, brd, $J = 15.8$ Hz), 2.71 (1H, ddd, $J = 14.1, 8.6, 0.9$ Hz), 2.60 (1H, ddd, $J = 14.1, 6.9, 1.4$ Hz), 2.01 (1H, dt, $J = 12.9, 4.4$ Hz), 1.79 (1H, dt, $J = 12.9, 4.4$ Hz), 1.41 (9H, s), 1.28-1.05 (3H, m), 0.92-0.78 (1H, m), 0.77 (3H, t, $J = 7.3$ Hz); ¹³C NMR (101 MHz, CDCl₃) δ 209.1, 165.3, 140.2, 139.2, 135.6, 128.6, 127.0, 126.6, 124.2, 123.3, 80.4, 63.6, 42.8, 39.7, 28.2, 26.1, 22.9, 13.8; IR (film) 2959, 2932, 1703, 1653, 1368, 1254, 1153, 980, 920, 743 cm⁻¹; HRMS (ESI, positive ion mode) Calcd for C₂₀H₂₆O₃S₁Na⁺ ([M+Na]⁺) 369.1495. Found 369.1494.; HPLC IC3, H/IPA = 19:1, flow rate = 1.0 mL/min, $\lambda = 210$ nm, 15.8 min (major), 16.5 min (minor).

 **8d**: $[\alpha]_D^{21} = +97.1$ ($c = 1.0$, CHCl₃) for 97% ee; ¹H NMR (400 MHz, CDCl₃) δ 7.36 (1H, dd, $J = 7.6, 1.4$ Hz), 7.30 (1H, dt, $J = 7.6, 1.4$ Hz), 7.25 (1H, dt, $J = 7.6, 1.4$ Hz), 7.17 (1H, dd, $J = 7.6, 1.4$ Hz), 6.35 (1H, ddd, $J = 15.6, 8.7, 6.7$ Hz), 5.68 (1H, dt, $J = 15.6, 1.4$ Hz), 2.83 (1H, ddd, $J = 14.1, 8.7, 1.4$ Hz), 2.71 (1H, ddd, $J = 14.1, 6.7, 1.4$ Hz), 2.19 (1H, sep, $J = 7.1$ Hz), 1.39 (9H, s), 0.95 (3H, d, $J = 7.1$ Hz), 0.94 (3H, d, $J = 7.1$ Hz); ¹³C NMR (101 MHz, CDCl₃) δ 208.8, 165.3, 140.9, 138.6, 136.1, 128.5, 126.9, 126.3, 124.7, 123.1, 80.4, 66.2, 39.6, 38.2, 28.2, 17.4, 16.9; IR (film) 2974, 2934, 1703, 1653, 1368, 1327, 1152, 982, 910, 750 cm⁻¹; HRMS (ESI, positive ion mode) Calcd for

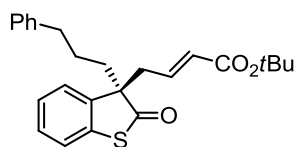
$C_{19}H_{24}O_3S_1Na^+$ ($[M+Na]^+$) 355.1338. Found 355.1337.; HPLC OZ3, H/IPA = 10:1, flow rate = 0.5 mL/min, λ = 210 nm, 10.0 min (minor), 10.9 min (major).



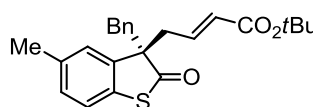
8e: $[\alpha]_D^{21} = +28.9$ ($c = 0.75$, $CHCl_3$) for 95% ee; 1H NMR (400 MHz, $CDCl_3$) δ 7.38 (1H, dd, $J = 7.6, 1.6$ Hz), 7.30 (1H, dt, $J = 7.6, 1.6$ Hz), 7.27 (1H, dt, $J = 7.6, 1.6$ Hz), 7.15 (1H, dd, $J = 7.6, 1.6$ Hz), 6.44 (1H, ddd, $J = 15.6, 8.6, 7.0$ Hz), 5.66 (1H, dt, $J = 15.6, 1.4$ Hz), 2.64 (1H, ddd, $J = 13.9, 8.6, 1.4$ Hz), 2.52 (1H, ddd, $J = 13.9, 7.0, 1.4$ Hz), 2.01 (1H, dd, $J = 14.1, 8.0$ Hz), 1.84 (1H, dd, $J = 14.1, 5.5$ Hz), 1.49-1.35 (1H, m), 1.42 (9H, s), 0.75 (3H, d, $J = 6.7$ Hz), 0.64 (3H, d, $J = 6.7$ Hz); ^{13}C NMR (101 MHz, $CDCl_3$) δ 209.0, 165.3, 139.9, 139.1, 135.4, 128.6, 127.2, 126.4, 124.7, 123.3, 80.5, 63.2, 47.6, 44.7, 28.2, 25.5, 24.1, 23.6; IR (film) 2959, 2932, 1705, 1653, 1468, 1368, 1333, 1153, 982, 743 cm^{-1} ; HRMS (ESI, positive ion mode) Calcd for $C_{20}H_{26}O_3S_1Na^+$ ($[M+Na]^+$) 369.1495. Found 369.1495.; HPLC OZ3, H/IPA = 10:1, flow rate = 0.5 mL/min, λ = 210 nm, 8.0 min (minor), 8.7 min (major).



8f: $[\alpha]_D^{21} = +59.6$ ($c = 0.97$, $CHCl_3$) for 92% ee; 1H NMR (400 MHz, $CDCl_3$) δ 7.39 (1H, dd, $J = 7.3, 1.6$ Hz), 7.32 (1H, dt, $J = 7.3, 1.6$ Hz), 7.29 (1H, dt, $J = 7.3, 1.6$ Hz), 7.18 (1H, dd, $J = 7.3, 1.6$ Hz), 6.42 (1H, ddd, $J = 15.6, 8.7, 7.1$ Hz), 5.70 (1H, dt, $J = 15.6, 1.5$ Hz), 3.56 (3H, s), 2.75 (1H, ddd, $J = 14.1, 8.7, 1.5$ Hz), 2.63 (1H, ddd, $J = 13.9, 7.1, 1.5$ Hz), 2.36-2.15 (3H, m), 1.96-1.84 (1H, m), 1.41 (9H, s); ^{13}C NMR (101 MHz, $CDCl_3$) δ 208.1, 172.8, 165.1, 139.5, 137.7, 135.5, 129.1, 127.5, 126.9, 124.4, 123.5, 80.6, 62.7, 51.8, 42.5, 34.2, 28.8, 28.2; IR (film) 2978, 2953, 1738, 1703, 1655, 1447, 1368, 1152, 984, 745 cm^{-1} ; HRMS (ESI, positive ion mode) Calcd for $C_{20}H_{24}O_5S_1Na^+$ ($[M+Na]^+$) 399.1237. Found 399.1226.; HPLC OZ3, H/IPA = 10:1, flow rate = 0.5 mL/min, λ = 210 nm, 19.7 min (minor), 23.2 min (major).

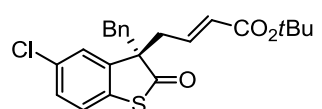


8g: $[\alpha]_D^{21} = +24.9$ ($c = 0.82$, $CHCl_3$) for 95% ee; 1H NMR (400 MHz, $CDCl_3$) δ 7.36 (1H, dd, $J = 7.6, 1.6$ Hz), 7.28 (1H, dt, $J = 7.6, 1.6$ Hz), 7.26-7.20 (3H, m), 7.17-7.12 (1H, m), 7.07 (1H, dd, $J = 7.6, 1.6$ Hz), 7.02 (1H, d, $J = 8.2$ Hz), 7.02 (1H, d, $J = 8.2$ Hz), 6.41 (1H, ddd, $J = 15.7, 8.6, 7.0$ Hz), 5.67 (1H, dt, $J = 15.7, 1.4$ Hz), 2.70 (1H, ddd, $J = 14.1, 8.6, 1.4$ Hz), 2.57 (1H, ddd, $J = 14.1, 7.0, 1.4$ Hz), 2.54 (1H, ddd, $J = 14.4, 8.7, 6.4$ Hz), 2.43 (1H, ddd, $J = 14.4, 8.7, 6.4$ Hz), 2.06 (1H, ddd, $J = 13.3, 12.8, 4.5$ Hz), 1.82 (1H, ddd, $J = 13.3, 12.8, 4.5$ Hz), 1.54-1.42 (1H, m), 1.41 (9H, s), 1.25-1.14 (1H, m); ^{13}C NMR (101 MHz, $CDCl_3$) δ 208.9, 165.3, 141.4, 140.1, 138.8, 135.5, 128.7, 128.4, 128.4, 127.1, 126.7, 126.0, 124.1, 123.4, 80.5, 63.5, 42.8, 39.3, 35.8, 28.2, 25.6; IR (film) 2976, 2934, 2361, 1703, 1601, 1368, 1150, 982, 746, 698 cm^{-1} ; HRMS (ESI, positive ion mode) Calcd for $C_{25}H_{28}O_3S_1Na^+$ ($[M+Na]^+$) 431.1651. Found 431.1650.; HPLC OZ3, H/IPA = 10:1, flow rate = 0.5 mL/min, λ = 210 nm, 9.5 min (minor), 10.6 min (major).

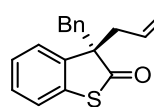


8h: $[\alpha]_D^{21} = -39.1$ ($c = 1.0$, $CHCl_3$) for 96% ee; 1H NMR (400 MHz, $CDCl_3$) δ 7.14-7.05 (5H, m), 6.99 (1H, s), 6.79-6.77 (2H, m), 6.42 (1H, ddd, $J = 15.6, 8.7, 6.7$ Hz), 5.74 (1H, d, $J = 15.6$ Hz), 3.21 (1H, d, $J = 13.3$ Hz), 3.06 (1H, d, $J = 13.3$ Hz), 2.89 (1H, dd, $J = 14.2, 8.7$ Hz), 2.74 (1H, dd, $J = 14.2, 6.7$ Hz), 2.38 (3H, s), 1.41 (9H, s); ^{13}C NMR (101 MHz, $CDCl_3$) δ 209.0, 165.3, 140.3, 137.8, 136.1, 134.7, 132.2, 130.2, 129.6, 127.9, 127.1, 127.0, 125.7, 122.9, 80.5, 64.5, 46.1, 41.6, 28.2, 21.5; IR (film) 2976, 2920, 1697, 1653, 1325, 1150, 978, 922, 731, 700

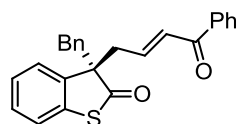
cm⁻¹; HRMS (ESI, positive ion mode) Calcd for C₂₄H₂₆O₃S₁Na⁺ ([M+Na]⁺) 417.1495. Found 417.1491.; HPLC AD3, H/IPA = 19:1, flow rate = 0.5 mL/min, λ = 210 nm, 12.4 min (minor), 14.1 min (major).



8i: $[\alpha]_D^{19} = -42.6$ ($c = 1.0$, CHCl₃) for 95% ee; ¹H NMR (400 MHz, CDCl₃) δ 7.25 (1H, dd, $J = 8.0, 1.8$ Hz), 7.17 (1H, d, $J = 1.8$ Hz), 7.15-7.08 (3H, m), 7.12 (1H, dd, $J = 8.0, 1.8$ Hz), 6.81 (2H, m), 6.42 (1H, ddd, $J = 15.6, 8.7, 6.6$ Hz), 5.75 (1H, brd, $J = 15.6$ Hz), 3.25 (1H, d, $J = 13.3$ Hz), 3.05 (1H, d, $J = 13.3$ Hz), 2.93 (1H, ddd, $J = 14.2, 8.7, 0.9$ Hz), 2.74 (1H, ddd, $J = 14.2, 6.6, 1.4$ Hz), 1.41 (9H, s); ¹³C NMR (101 MHz, CDCl₃) δ 207.6, 165.1, 139.7, 139.4, 134.1, 132.3, 130.1, 129.1, 128.1, 127.5, 127.3, 125.3, 124.2, 80.7, 64.9, 46.1, 41.5, 28.2, one peak for aromatic carbon was not found probably due to overlapping; IR (film) 2978, 2927, 1709, 1655, 1456, 1368, 1152, 980, 922, 700 cm⁻¹; HRMS (ESI, positive ion mode) Calcd for C₂₃H₂₃O₃Cl₁S₁Na⁺ ([M+Na]⁺) 437.0949. Found 437.0948.; HPLC OD3, H/IPA = 10:1, flow rate = 0.5 mL/min, λ = 210 nm, 12.2 min (major), 13.5 min (minor).

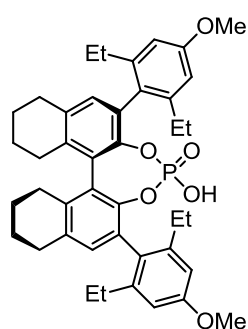


8j: $[\alpha]_D^{21} = -27.0$ ($c = 0.96$, CHCl₃) for 93% ee; ¹H NMR (400 MHz, CDCl₃) δ 7.30-7.23 (2H, m), 7.21-7.17 (2H, m), 7.14-7.05 (3H, m), 6.79 (2H, d, $J = 7.3$ Hz), 6.53 (1H, ddd, $J = 15.8, 8.3, 7.3$ Hz), 5.79 (1H, d, $J = 15.8$ Hz), 3.64 (3H, s), 3.25 (1H, d, $J = 13.3$ Hz), 3.08 (1H, d, $J = 13.3$ Hz), 2.96 (1H, dd, $J = 14.2, 8.3$ Hz), 2.79 (1H, dd, $J = 14.2, 7.3$ Hz); ¹³C NMR (101 MHz, CDCl₃) δ 208.5, 166.3, 141.7, 137.7, 135.7, 134.5, 130.2, 128.9, 128.0, 127.1, 126.3, 125.1, 125.0, 123.3, 64.6, 51.6, 46.2, 41.7; IR (film) 3030, 2949, 1703, 1659, 1437, 1325, 1275, 1167, 737, 700 cm⁻¹; HRMS (ESI, positive ion mode) Calcd for C₂₀H₁₈O₃S₁Na⁺ ([M+Na]⁺) 361.0869. Found 361.0862.; HPLC OZ3, H/IPA = 10:1, flow rate = 0.5 mL/min, λ = 210 nm, 17.7 min (minor), 22.2 min (major).

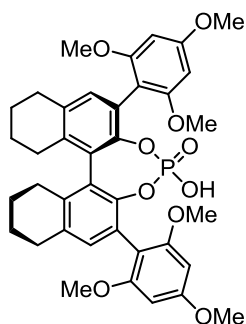


8k: $[\alpha]_D^{21} = +17.1$ ($c = 1.0$, CHCl₃) for 89% ee; ¹H NMR (400 MHz, CDCl₃) δ 7.72-7.70 (2H, m), 7.51 (1H, t, $J = 7.8$ Hz), 7.39 (2H, t, $J = 7.8$ Hz), 7.33-7.24 (3H, m), 7.21 (1H, dd, $J = 7.8, 1.7$ Hz), 7.16-7.07 (3H, m), 6.83-6.81 (2H, m), 6.76 (1H, d, $J = 15.6$ Hz), 6.53 (1H, ddd, $J = 15.6, 8.2, 7.1$ Hz), 3.30 (1H, d, $J = 13.3$ Hz), 3.14 (1H, d, $J = 13.3$ Hz), 3.08 (1H, dd, $J = 14.2, 8.2$ Hz), 2.90 (1H, dd, $J = 14.2, 7.1$ Hz); ¹³C NMR (101 MHz, CDCl₃) δ 208.7, 190.7, 141.4, 137.9, 137.5, 135.7, 134.5, 132.9, 130.3, 130.2, 128.9, 128.7, 128.6, 128.0, 127.1, 126.4, 125.2, 123.3, 65.0, 46.2, 42.4; IR (film) 3061, 3030, 2920, 1703, 1670, 1622, 1447, 1281, 922, 698 cm⁻¹; HRMS (ESI, positive ion mode) Calcd for C₂₅H₂₀O₂S₁Na⁺ ([M+Na]⁺) 407.1076. Found 407.1063.; HPLC OZ3, H/IPA = 10:1, flow rate = 0.5 mL/min, λ = 210 nm, 27.9 min (minor), 44.2 min (major).

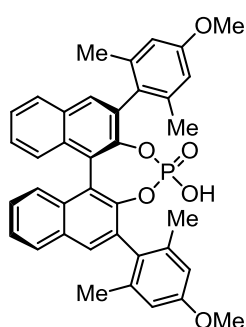
Characterization Data for Chiral Phosphoric Acids 2



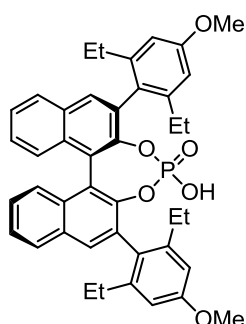
2b: $[\alpha]_D^{21} = -35.2$ ($c = 1.0$, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 6.89 (2H, s), 6.58 (2H, s), 6.57 (2H, s), 3.69 (6H, s), 2.87-2.69 (6H, m), 2.43-2.19 (10H, m), 1.90-1.80 (6H, m), 1.71-1.64 (2H, m), 1.11 (6H, t, $J = 7.6$ Hz), 0.97 (6H, t, $J = 7.6$ Hz); ¹³C NMR (101 MHz, CDCl₃) δ 159.1, 144.5, 144.0, 143.9 (d, $J_{P-C} = 8.6$ Hz), 136.5, 134.4, 132.5, 129.6 (d, $J_{P-C} = 3.9$ Hz), 128.0, 126.9, 110.9, 110.6, 55.0, 29.4, 27.9, 27.1, 26.9, 23.0, 22.9, 15.6, 14.6; ³¹P NMR (162 MHz, CDCl₃) δ 2.1; IR (film) 2959, 2930, 1601, 1578, 1190, 1157, 1016, 953, 901, 748 cm⁻¹; HRMS (ESI, negative ion mode) Calcd for C₄₂H₄₈O₆P₁⁻ ([M-H]⁻) 679.3179. Found 679.3179.



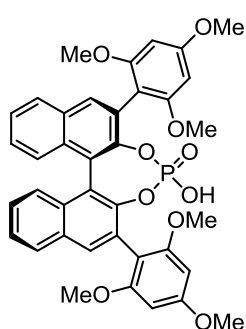
2c: $[\alpha]_D^{20} = -69.6$ ($c = 1.0$, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 6.99 (2H, s), 6.08 (4H, s), 3.72 (6H, s), 3.55 (6H, s), 3.53 (6H, s), 2.87-2.79 (4H, m), 2.78-2.71 (2H, m), 2.39-2.30 (2H, m), 1.86-1.74 (6H, m), 1.71-1.61 (2H, m); ^{13}C NMR (101 MHz, CDCl_3) δ 160.8, 158.8, 158.7, 144.7 (d, $J_{\text{P-C}} = 8.7$ Hz), 136.6, 134.2, 133.0, 126.7, 124.1 (d, $J_{\text{P-C}} = 2.9$ Hz), 107.8, 91.2, 90.7, 56.0, 55.9, 55.3, 29.4, 28.1, 22.9, 22.8; ^{31}P NMR (162 MHz, CDCl_3) δ 1.1; IR (film) 2934, 2837, 1585, 1454, 1414, 1204, 1123, 1016, 903, 725 cm^{-1} ; HRMS (ESI, negative ion mode) Calcd for $\text{C}_{38}\text{H}_{40}\text{O}_{10}\text{P}_1^-$ ($[\text{M}-\text{H}]^-$) 687.2354. Found 687.2363.



2g: $[\alpha]_D^{21} = -60.8$ ($c = 1.0$, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.92 (2H, d, $J = 8.2$ Hz), 7.79 (2H, s), 7.50 (2H, t, $J = 7.6$ Hz), 7.39-7.30 (4H, m), 6.56 (2H, d, $J = 2.3$ Hz), 6.49 (2H, d, $J = 2.3$ Hz), 3.39 (6H, s), 2.12 (6H, s), 1.99 (6H, s); ^{13}C NMR (101 MHz, CDCl_3) δ 158.9, 145.7 (d, $J_{\text{P-C}} = 8.7$ Hz), 138.5 (d, $J_{\text{P-C}} = 2.9$ Hz), 132.8, 132.1, 132.0, 131.6, 128.6, 128.3, 127.2, 126.5, 125.8, 122.3, 113.0, 112.8, 54.9, 21.5, 20.8, one peak for aromatic carbon was not found probably due to overlapping; ^{31}P NMR (162 MHz, CDCl_3) δ 5.1; IR (film) 2955, 2920, 1605, 1312, 1194, 1150, 1020, 905, 750, 729 cm^{-1} ; HRMS (ESI, negative ion mode) Calcd for $\text{C}_{38}\text{H}_{32}\text{O}_6\text{P}_1^-$ ($[\text{M}-\text{H}]^-$) 615.1931. Found 615.1932.



2h: $[\alpha]_D^{21} = -23.6$ ($c = 1.0$, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.91 (2H, d, $J = 8.5$ Hz), 7.81 (2H, s), 7.50 (2H, ddd, $J = 8.5, 6.4, 2.2$ Hz), 7.36-7.30 (4H, m), 6.62 (2H, d, $J = 2.5$ Hz), 6.56 (2H, d, $J = 2.5$ Hz), 3.58 (6H, s), 2.39-2.20 (8H, m), 1.11 (6H, t, $J = 7.7$ Hz), 0.98 (6H, t, $J = 7.7$ Hz); ^{13}C NMR (101 MHz, CDCl_3) δ 159.3, 146.1 (d, $J_{\text{P-C}} = 8.7$ Hz), 144.9, 144.4, 132.4 (d, $J_{\text{P-C}} = 2.9$ Hz), 132.4, 132.3, 131.2, 128.3, 127.9, 127.3, 126.3, 125.7, 122.3, 110.8, 110.7, 55.1, 27.2, 27.0, 15.6, 14.7; ^{31}P NMR (162 MHz, CDCl_3) δ 3.8; IR (film) 2963, 2932, 1601, 1578, 1192, 1148, 1018, 995, 962, 748 cm^{-1} ; HRMS (ESI, negative ion mode) Calcd for $\text{C}_{42}\text{H}_{40}\text{O}_6\text{P}_1^-$ ($[\text{M}-\text{H}]^-$) 671.2557. Found 671.2558.



2i: $[\alpha]_D^{21} = -93.2$ ($c = 1.0$, CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 7.90 (2H, d, $J = 8.6$ Hz), 7.89 (2H, s), 7.46-7.42 (4H, m), 7.29-7.24 (2H, m), 6.15 (2H, d, $J = 1.8$ Hz), 6.11 (2H, d, $J = 1.8$ Hz), 3.75 (6H, s), 3.59 (6H, s), 3.54 (6H, s); ^{13}C NMR (101 MHz, CDCl_3) δ 161.4, 159.2, 158.8, 146.4 (d, $J_{\text{P-C}} = 10.6$ Hz), 133.4, 132.2, 131.5, 128.5, 127.6, 127.0 (d, $J_{\text{P-C}} = 2.9$ Hz), 126.0, 125.3, 121.9 (d, $J_{\text{P-C}} = 1.9$ Hz), 107.7, 91.3, 91.2, 56.2, 56.1, 55.5; ^{31}P NMR (162 MHz, CDCl_3) δ 3.3; IR (film) 2938, 2837, 1609, 1585, 1410, 1225, 1204, 1123, 1018, 750 cm^{-1} ; HRMS (ESI, negative ion mode) Calcd for $\text{C}_{38}\text{H}_{32}\text{O}_{10}\text{P}_1^-$ ($[\text{M}-\text{H}]^-$) 679.1728. Found 679.1740.

Crystallographic Structure Determination:

Recrystallization of 8a (CCDC 995705): A single crystal of **8a** was obtained from a solution of hexane and diethyl ether at room temperature. The single crystal thus obtained was mounted on CryoLoop. Data of X-ray diffraction were collected at 103 K on a Bruker SMART APEX CCD diffractometer with graphite-monochromated Mo K α radiation ($\lambda = 0.71073$ Å). An absorption correction was made using SADABS. The structure was solved by direct methods and Fourier syntheses, and refined by full-matrix least

squares on F^2 by using SHELXTL.³ All non-hydrogen atoms were refined with anisotropic displacement parameters. All hydrogen atoms were placed in calculated positions. The crystallographic data are summarized in Table S3 and ORTEP diagram is shown in Fig. S1.

Table S3. Crystal data and structure refinement for **8a**.

Empirical formula	C ₂₃ H ₂₄ O ₃ S	
Formula weight	380.48	
Temperature	103(2) K	
Wavelength	0.71073 Å	
Crystal system	Orthorhombic	
Space group	P2(1)2(1)2(1)	
Unit cell dimensions	a = 6.5465(17) Å	$\alpha = 90^\circ$.
	b = 7.844(2) Å	$\beta = 90^\circ$.
	c = 38.945(10) Å	$\gamma = 90^\circ$.
Volume	1999.9(9) Å ³	
Z	4	
Density (calculated)	1.264 Mg/m ³	
Absorption coefficient	0.182 mm ⁻¹	
F(000)	808	
Crystal size	0.3 x 0.4 x 0.8 mm ³	
Theta range for data collection	2.09 to 29.14°.	
Index ranges	-8 ≤ h ≤ 8, -10 ≤ k ≤ 10, -44 ≤ l ≤ 53	
Reflections collected	12232	
Independent reflections	5092 [R(int) = 0.0215]	
Completeness to theta = 29.14°	94.8 %	
Absorption correction	Empirical	
Refinement method	Full-matrix least-squares on F ²	
Data / restraints / parameters	5092 / 0 / 247	
Goodness-of-fit on F ²	0.662	
Final R indices [I > 2σ(I)]	R1 = 0.0235, wR2 = 0.0420	
R indices (all data)	R1 = 0.0559, wR2 = 0.0426	
Absolute structure parameter	0.00(5)	
Largest diff. peak and hole	0.168 and -0.182 e.Å ⁻³	

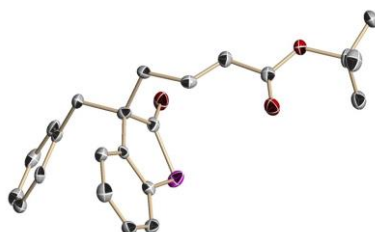
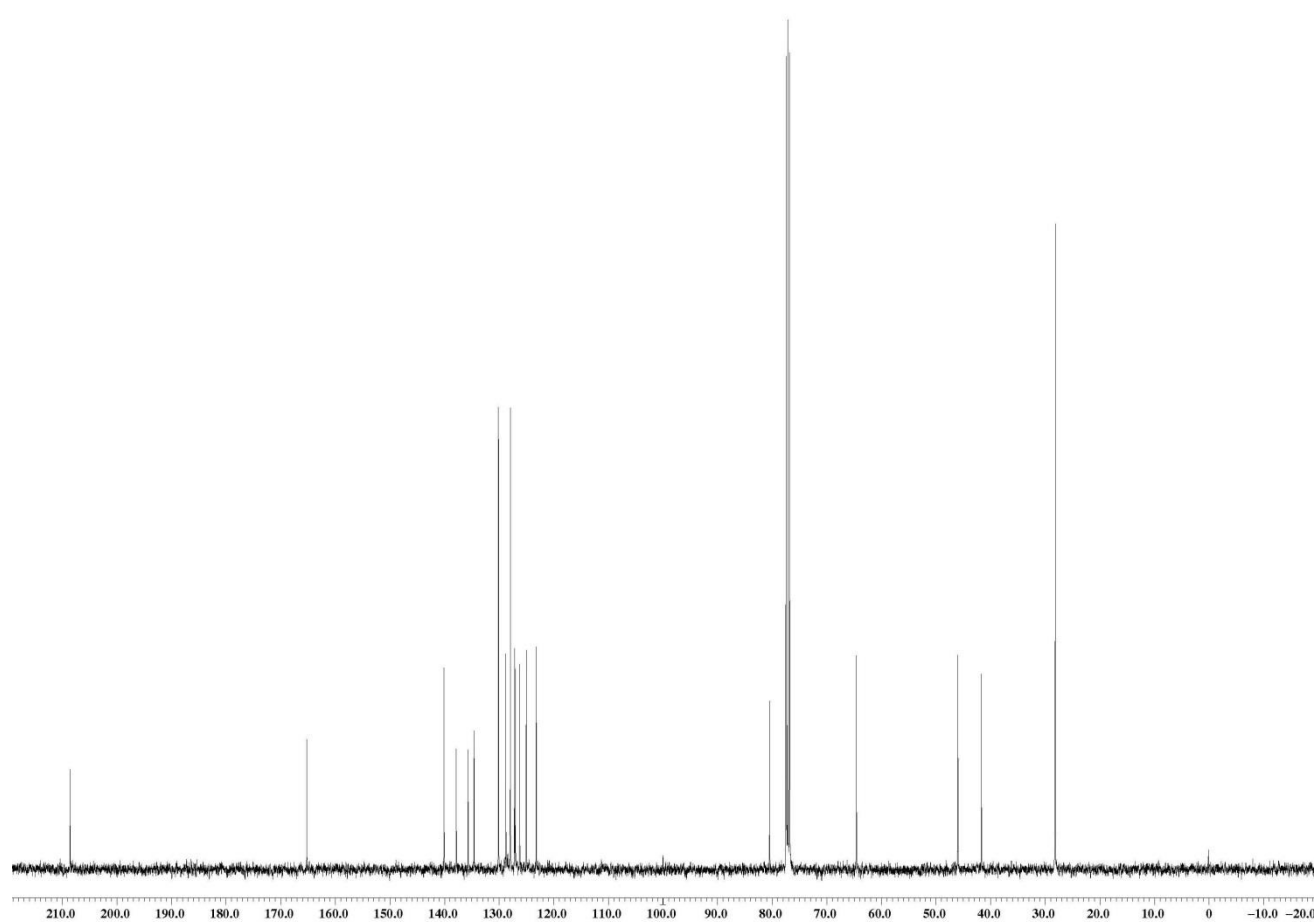
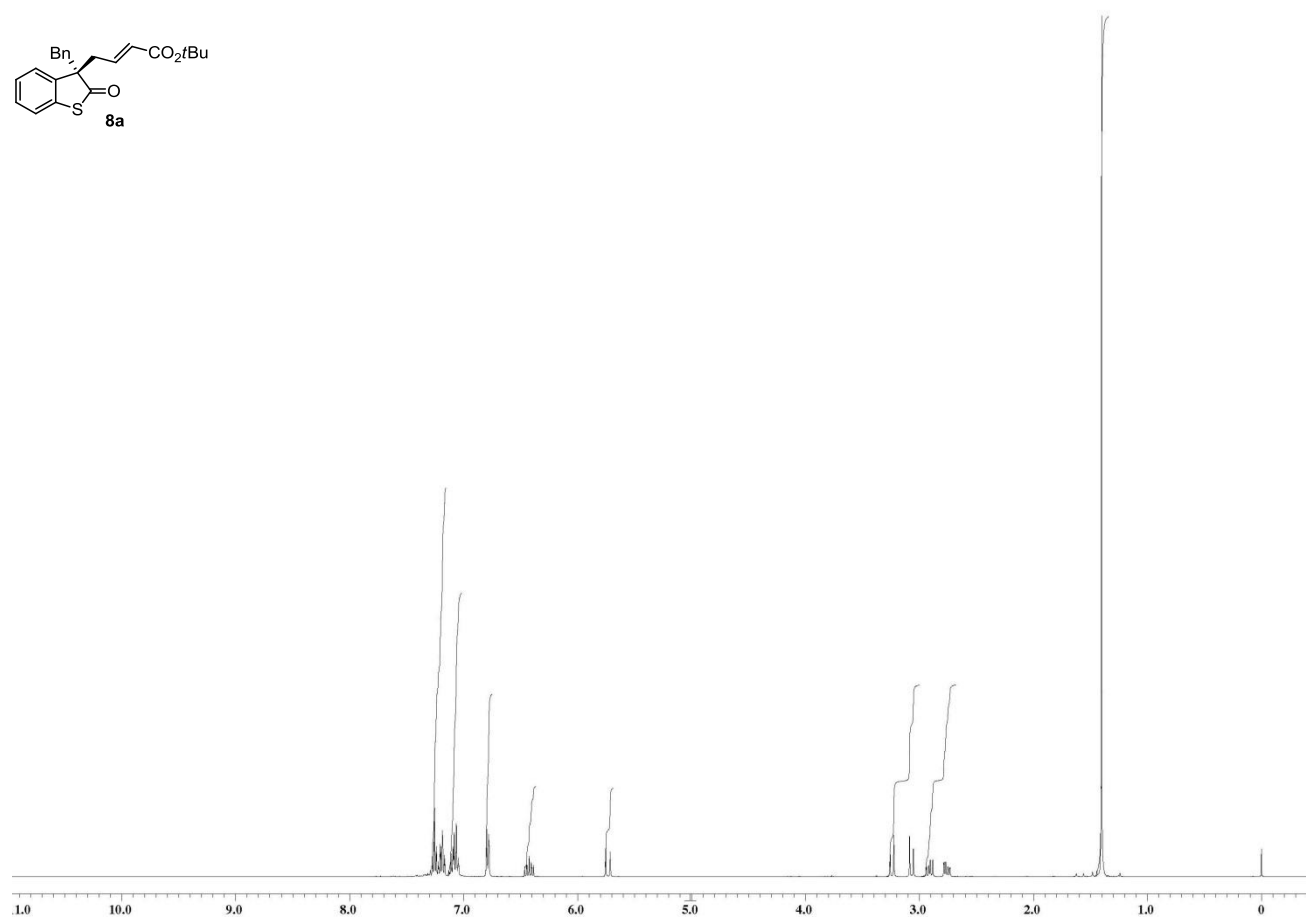
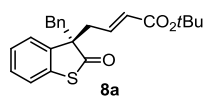
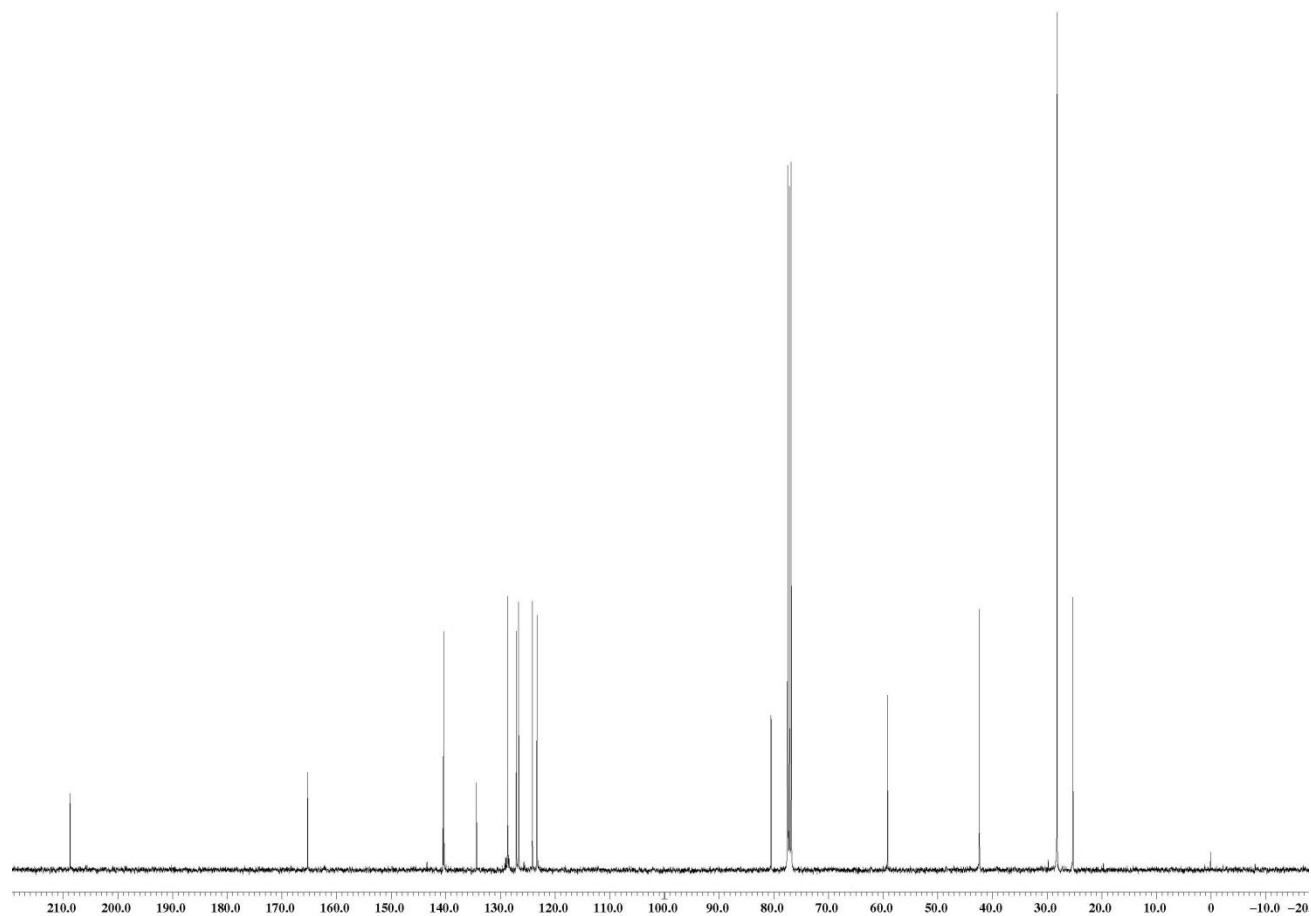
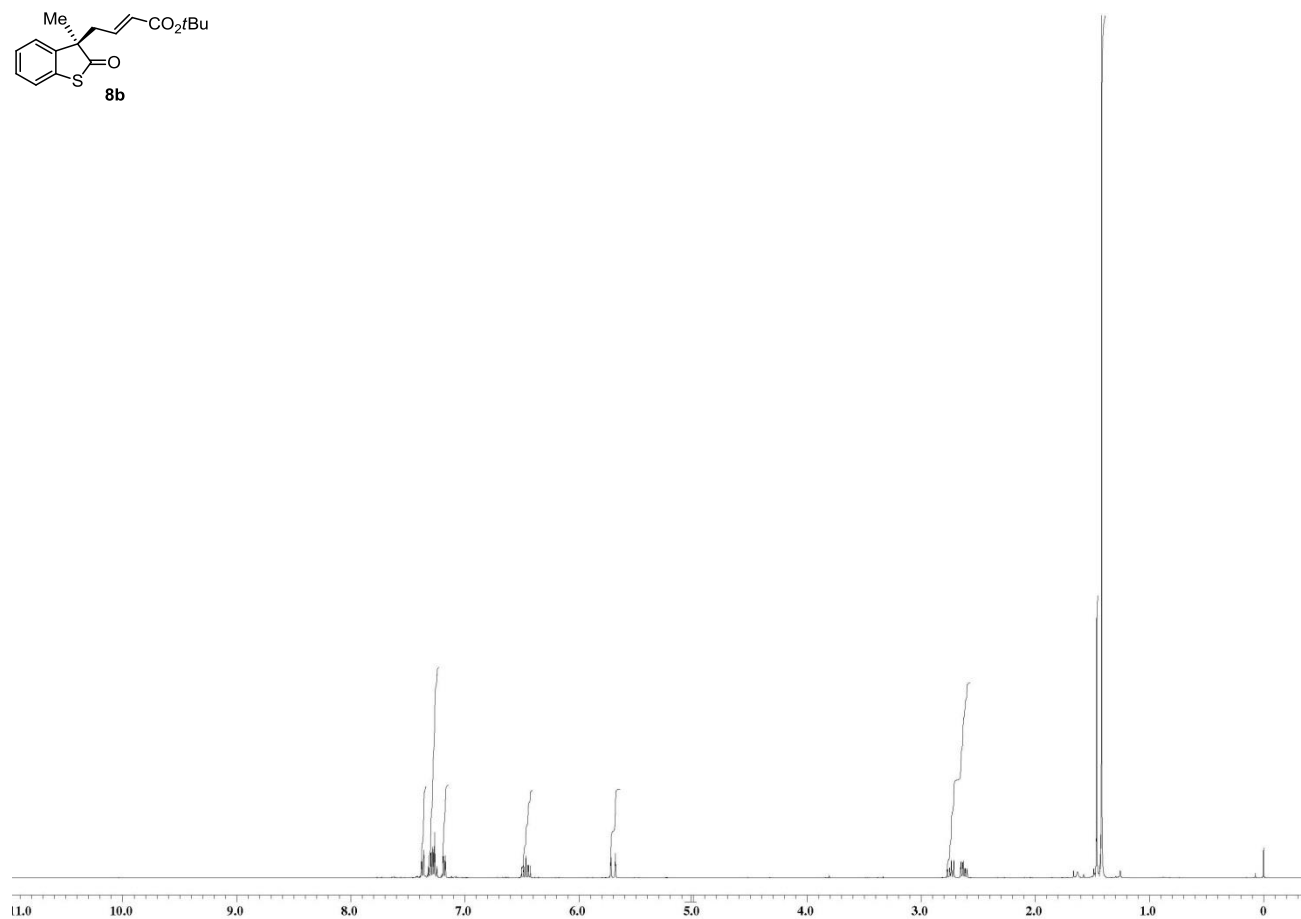
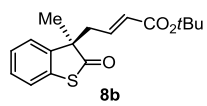


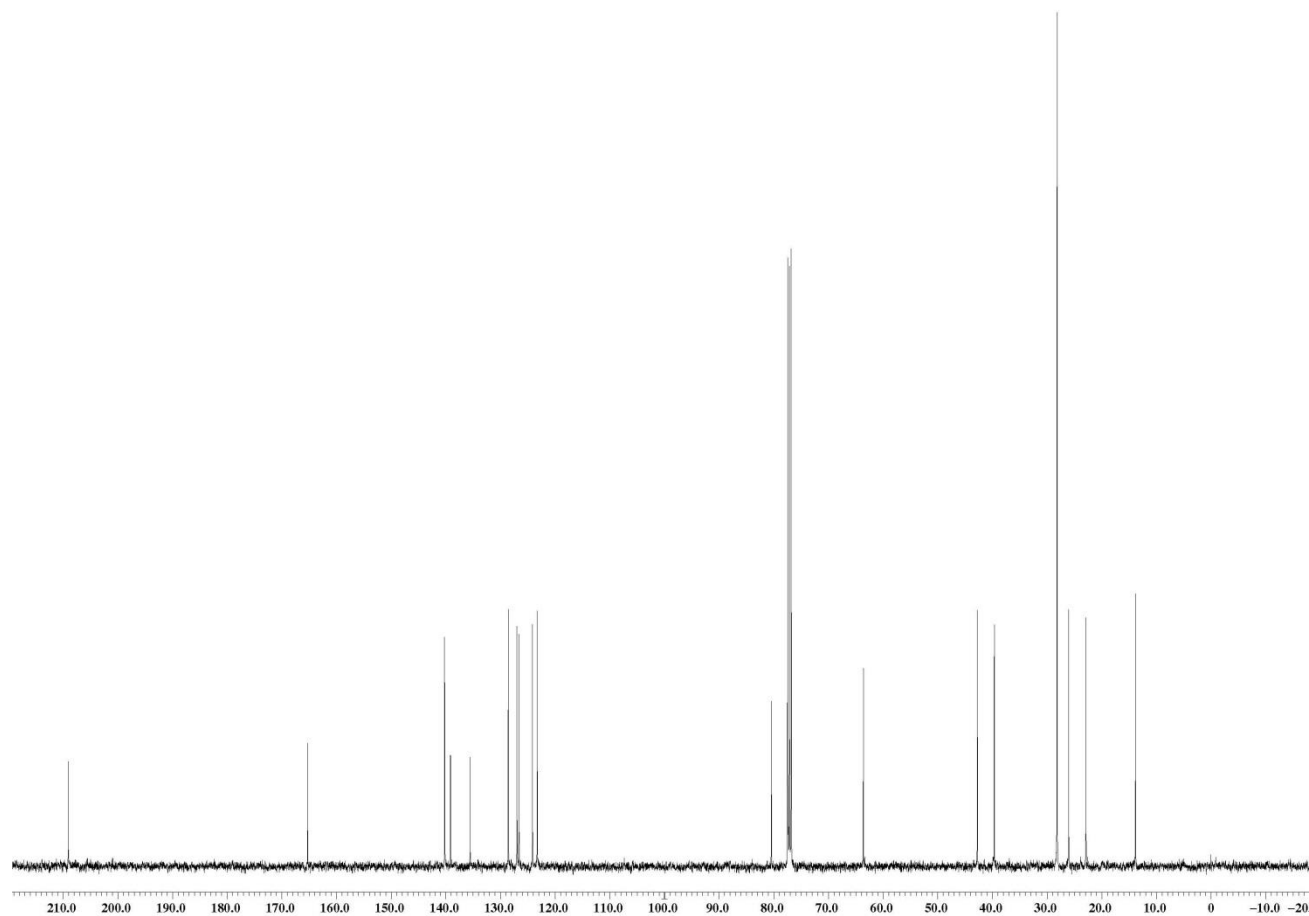
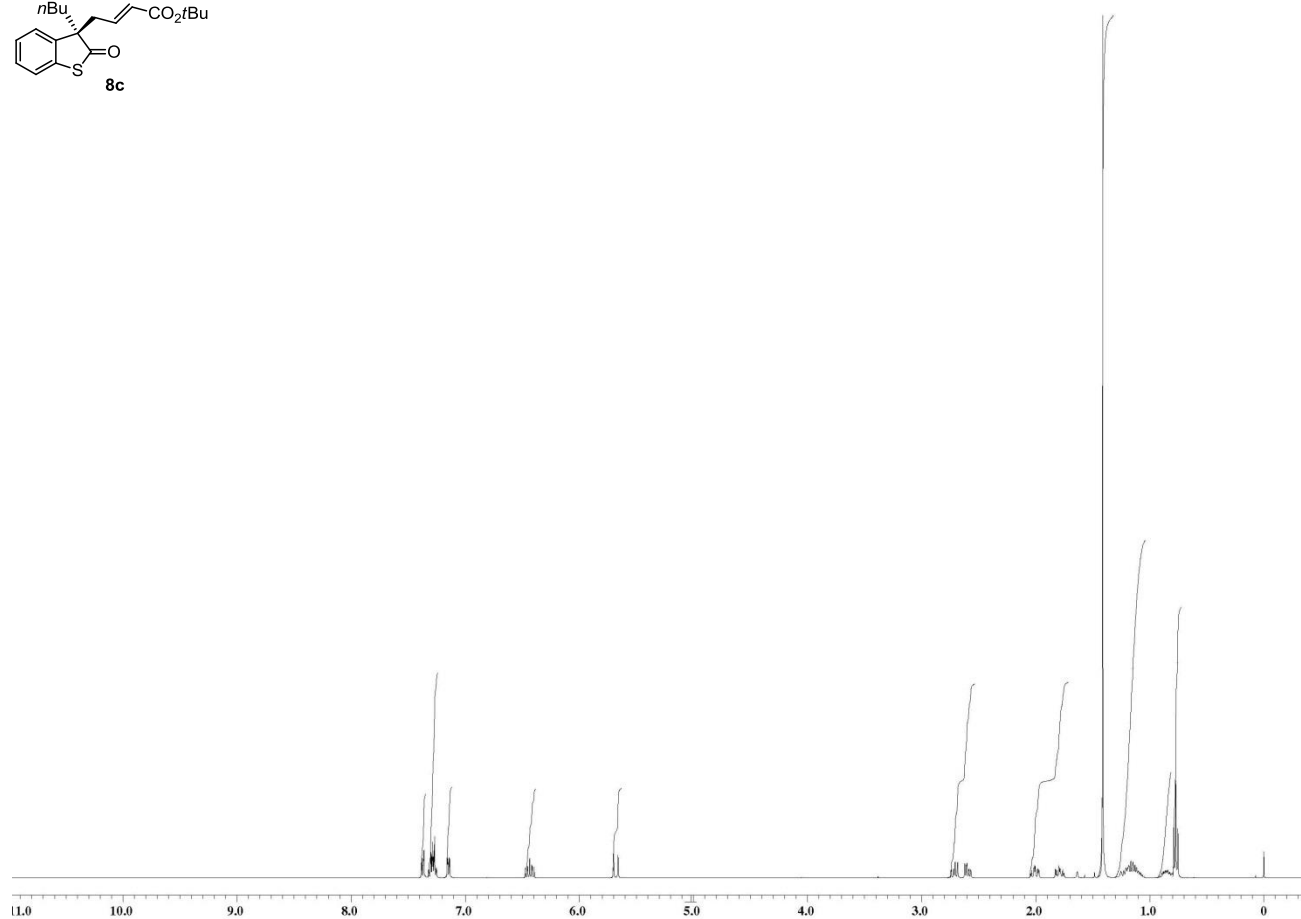
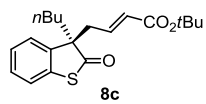
Figure S1. Molecular structure of **8a**. All calculated hydrogen atoms are omitted for clarity. purple = sulfur, red = oxygen, black = carbon.

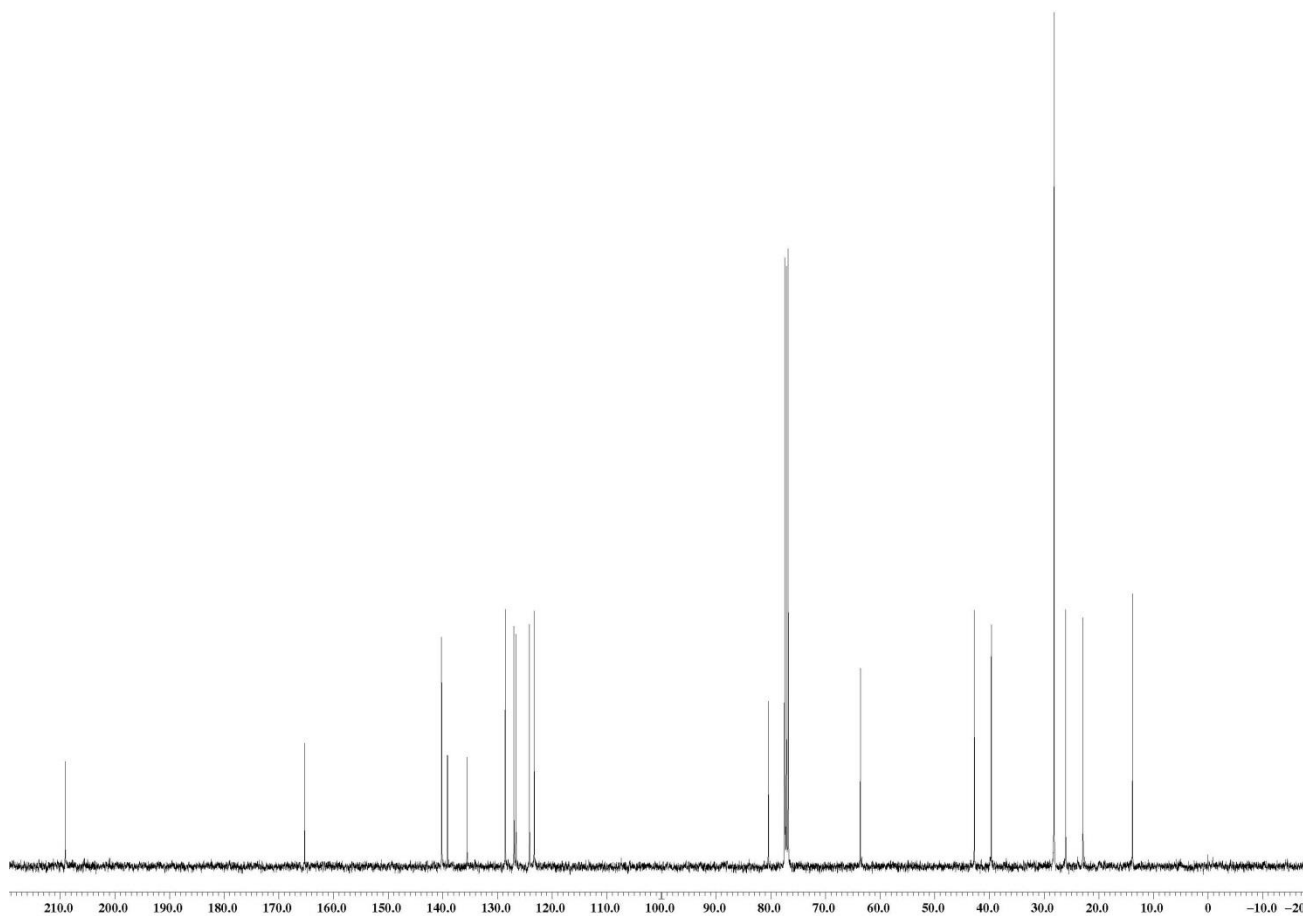
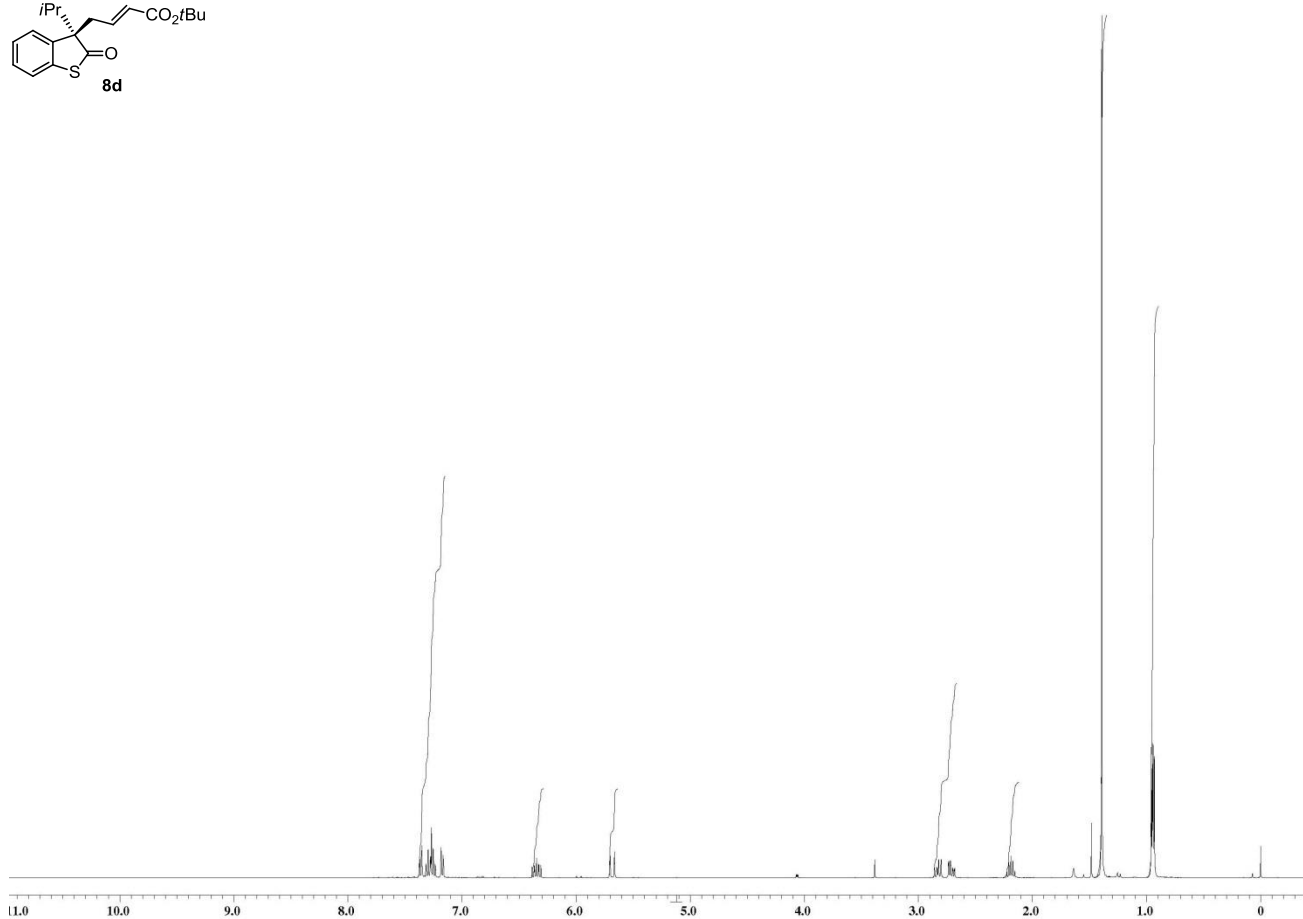
³ Sheldrick, G. M. SHELXTL 5.1, Bruker AXS Inc., Madison, Wisconsin, 1997.

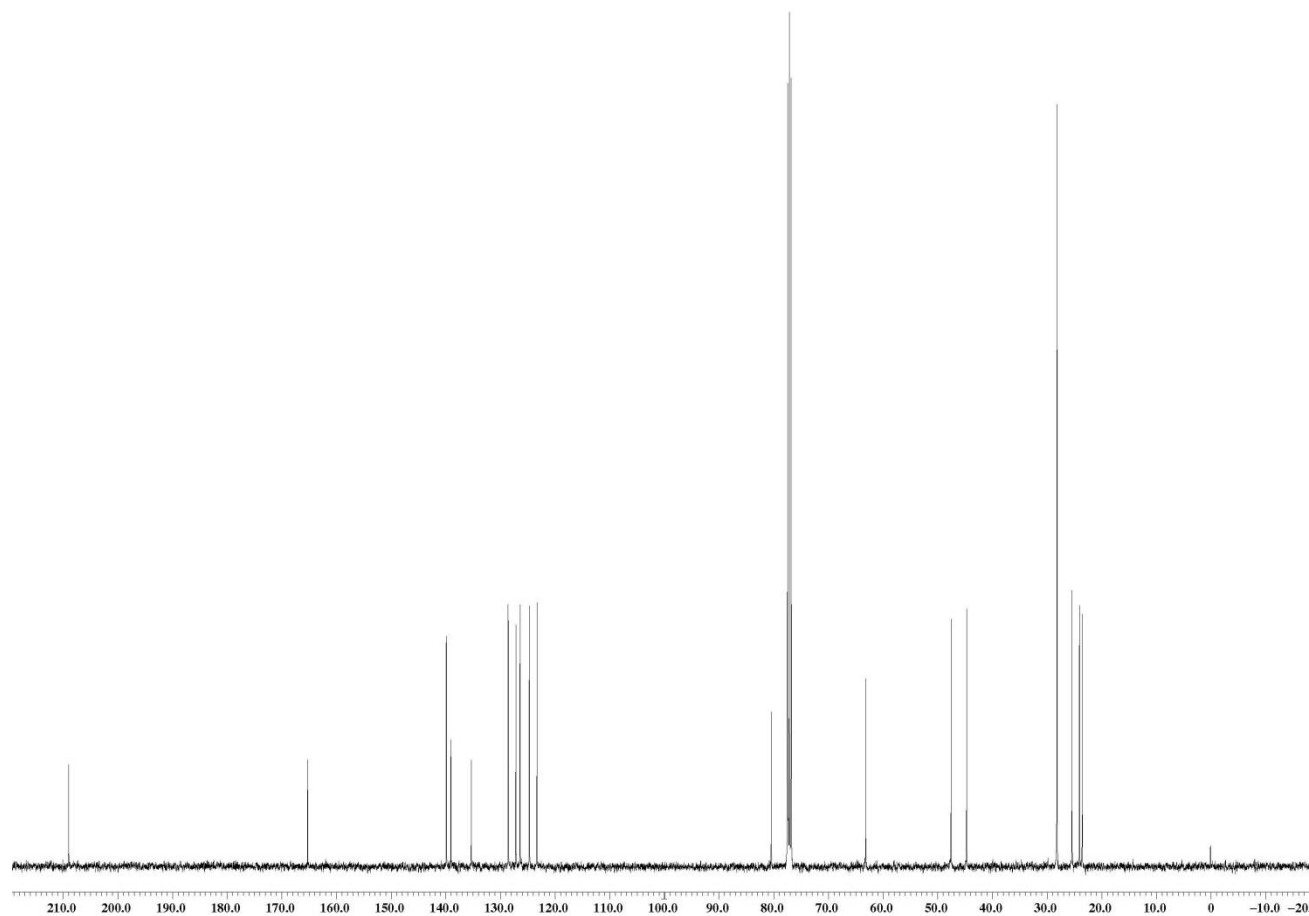
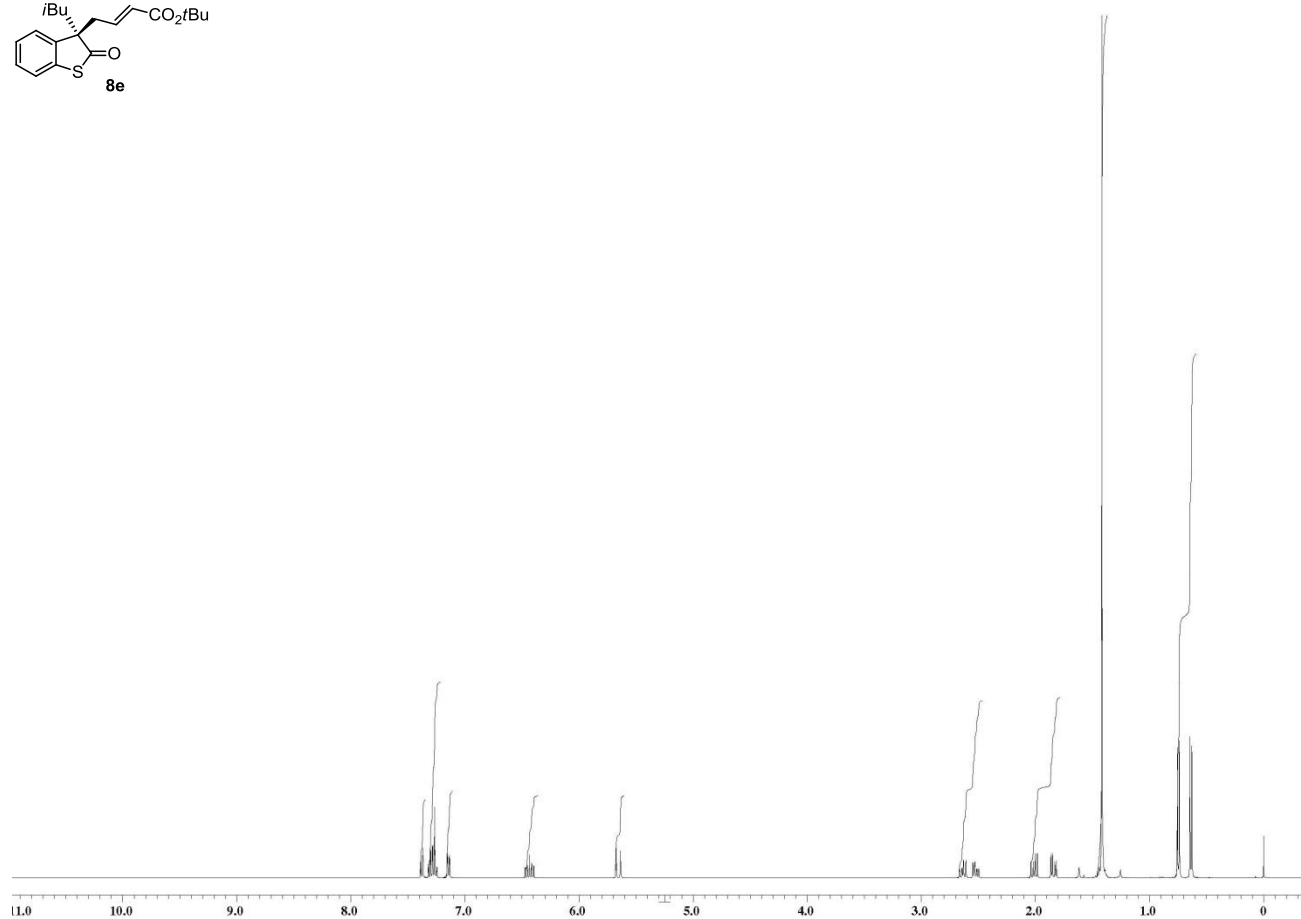
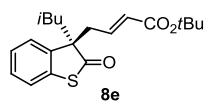
Copies of ^1H and ^{13}C NMR Spectra:

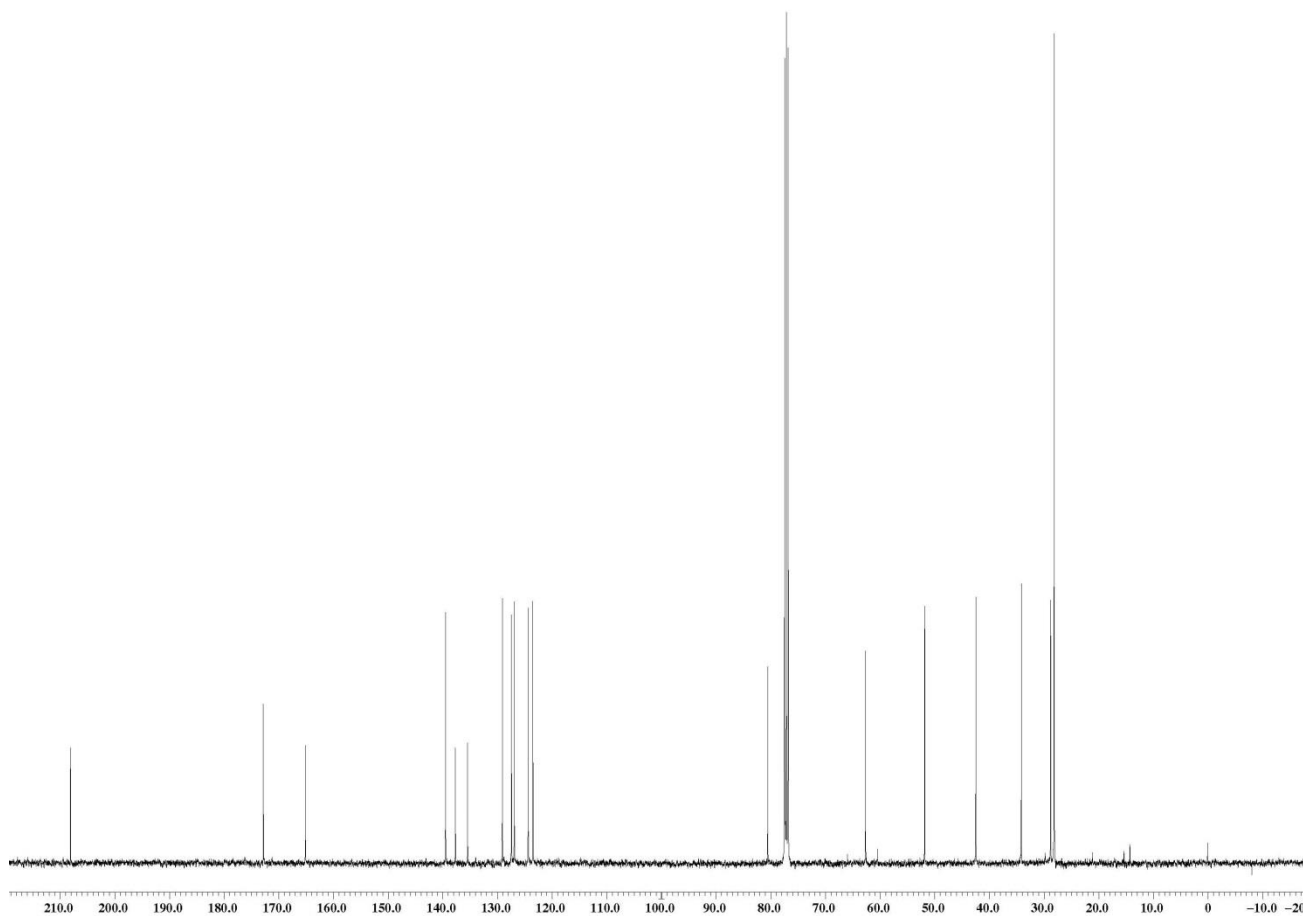
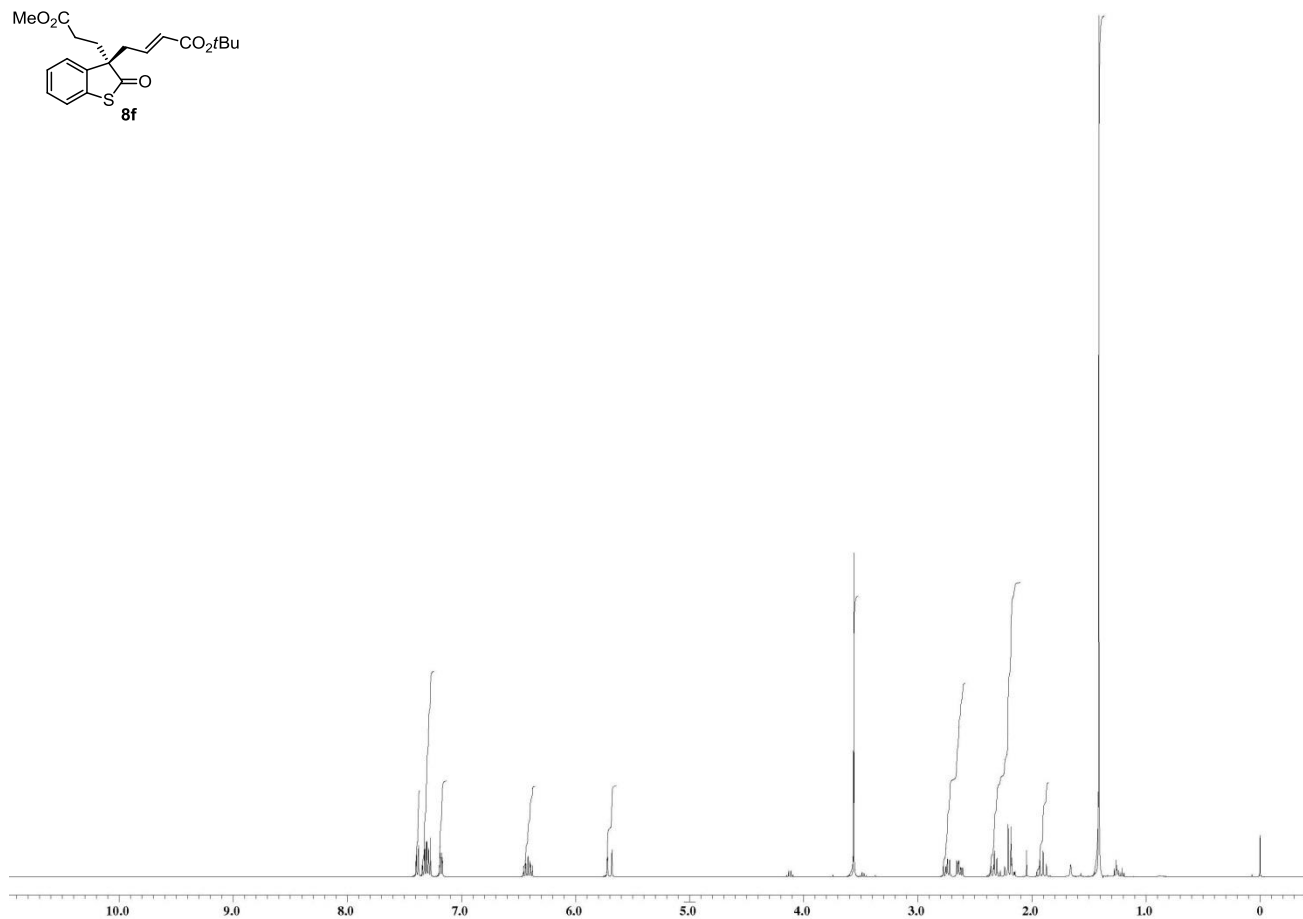
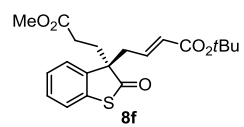


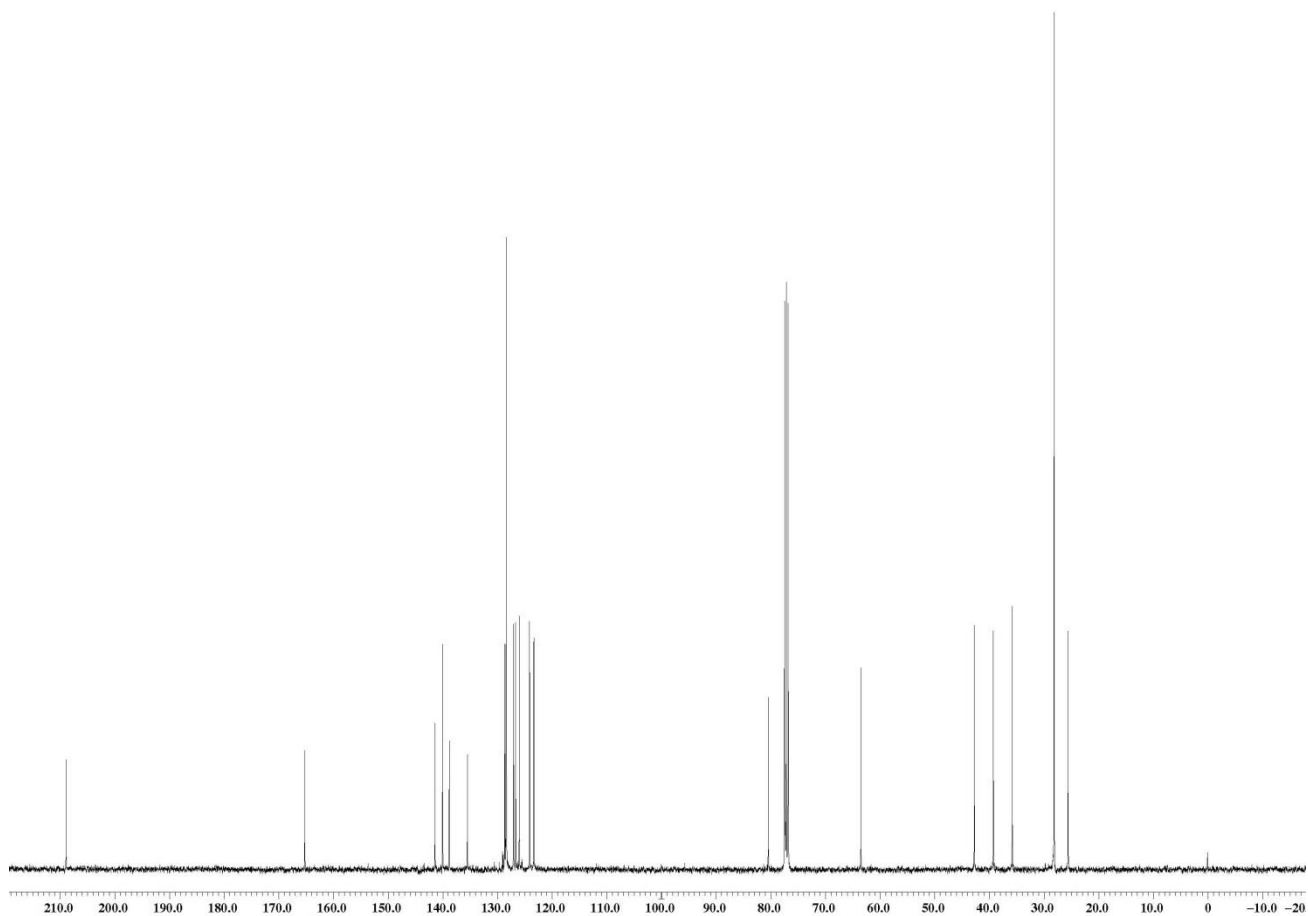
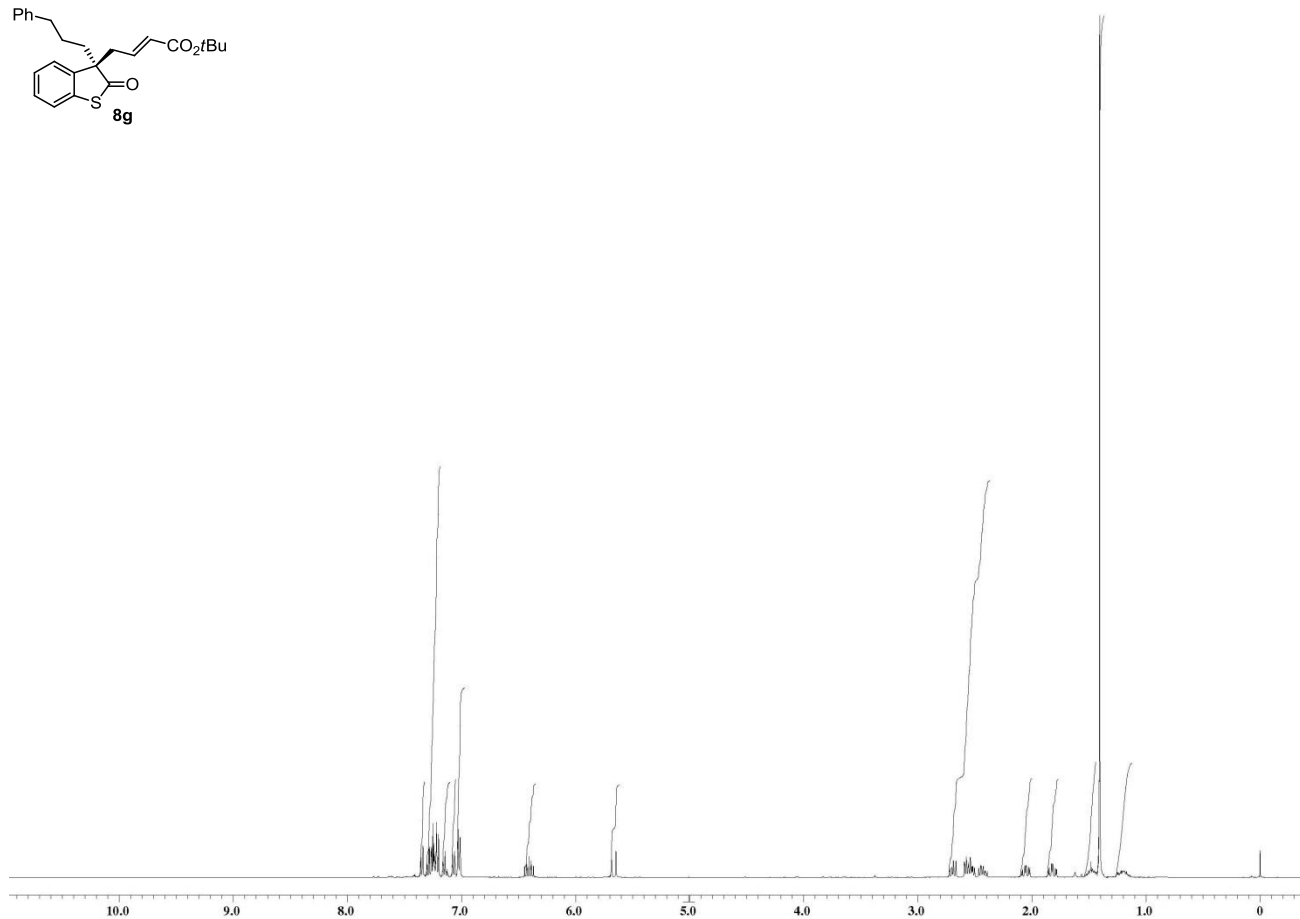
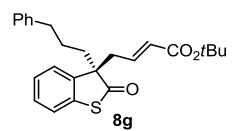


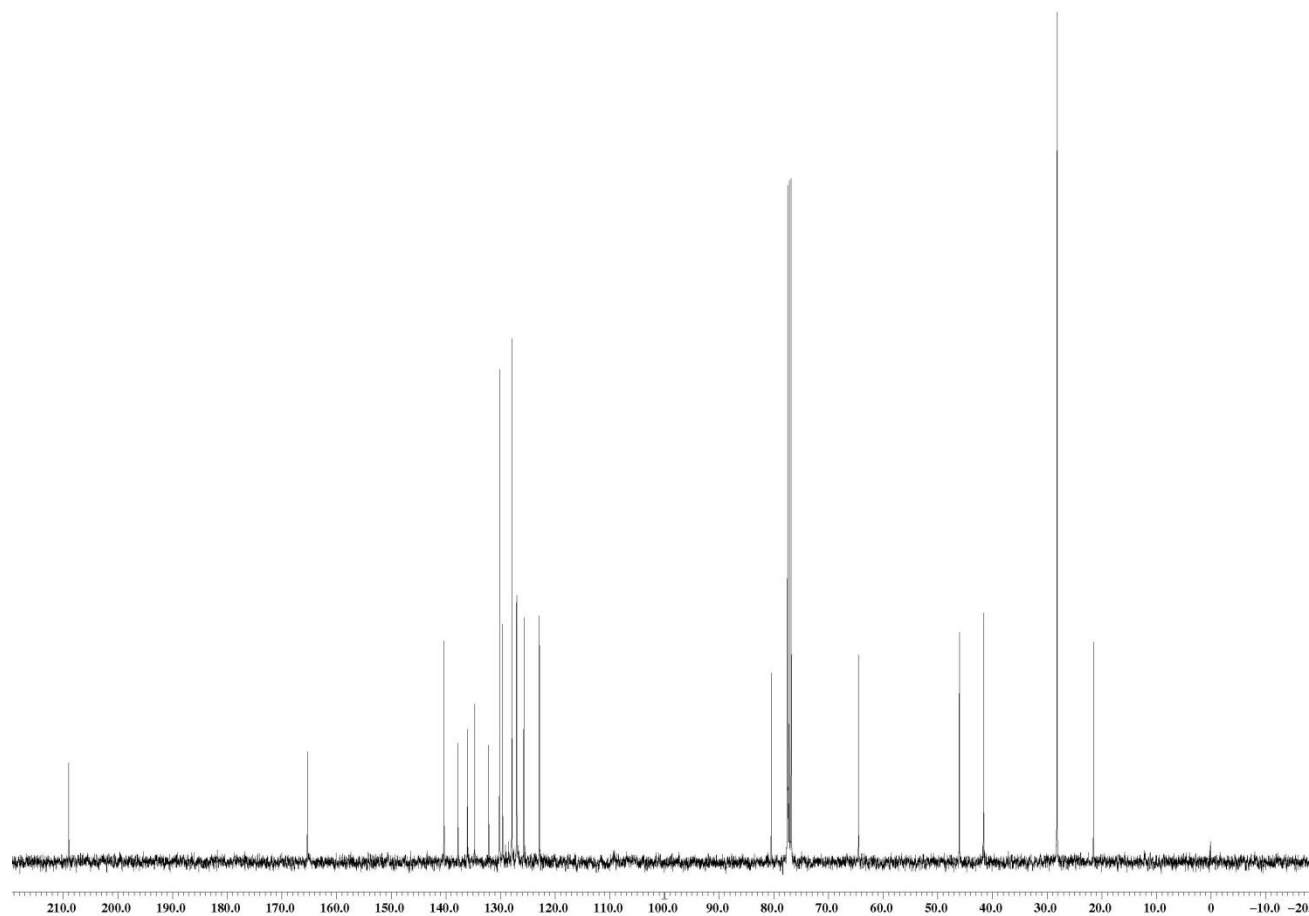
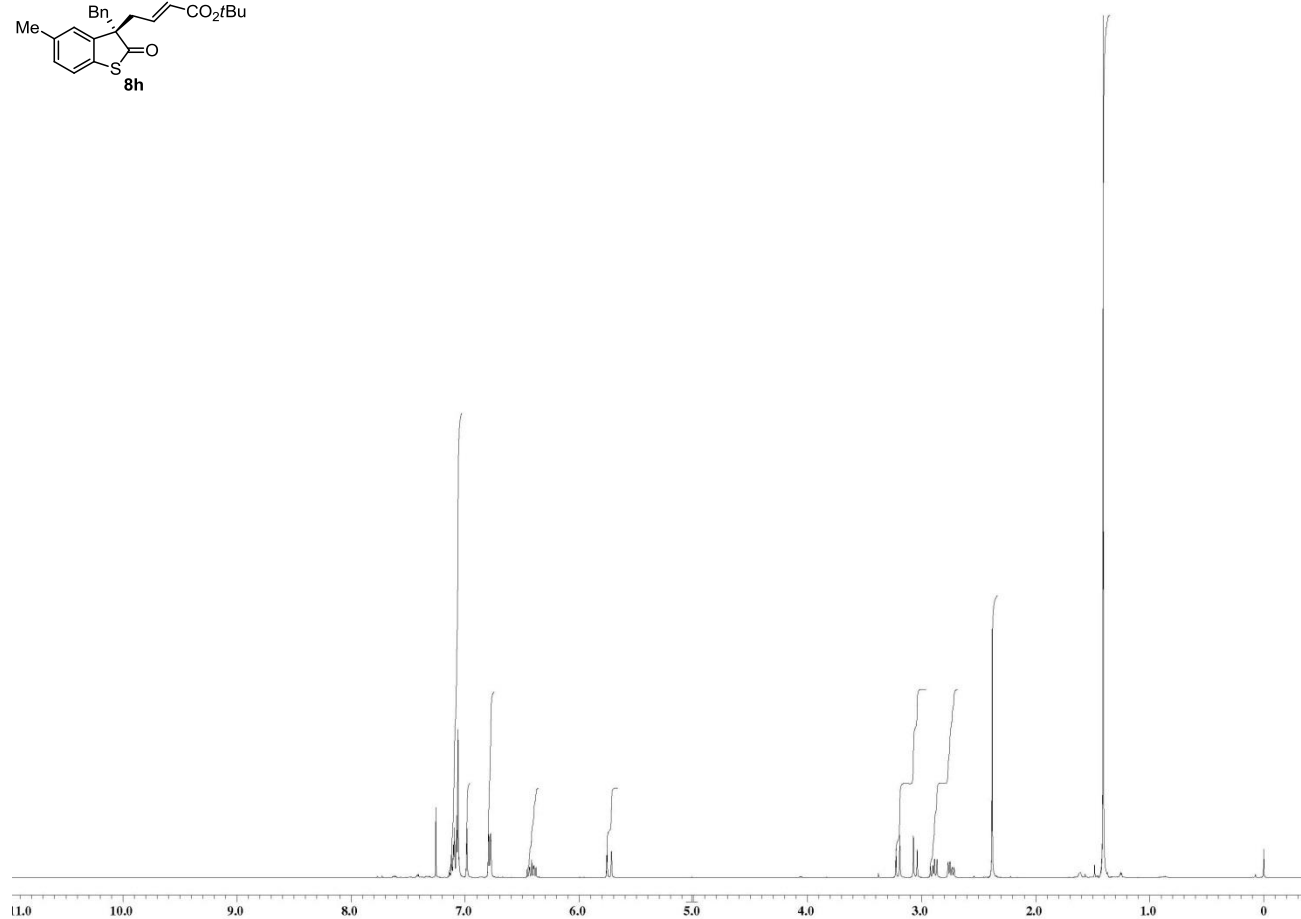
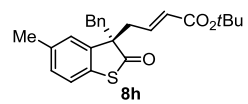


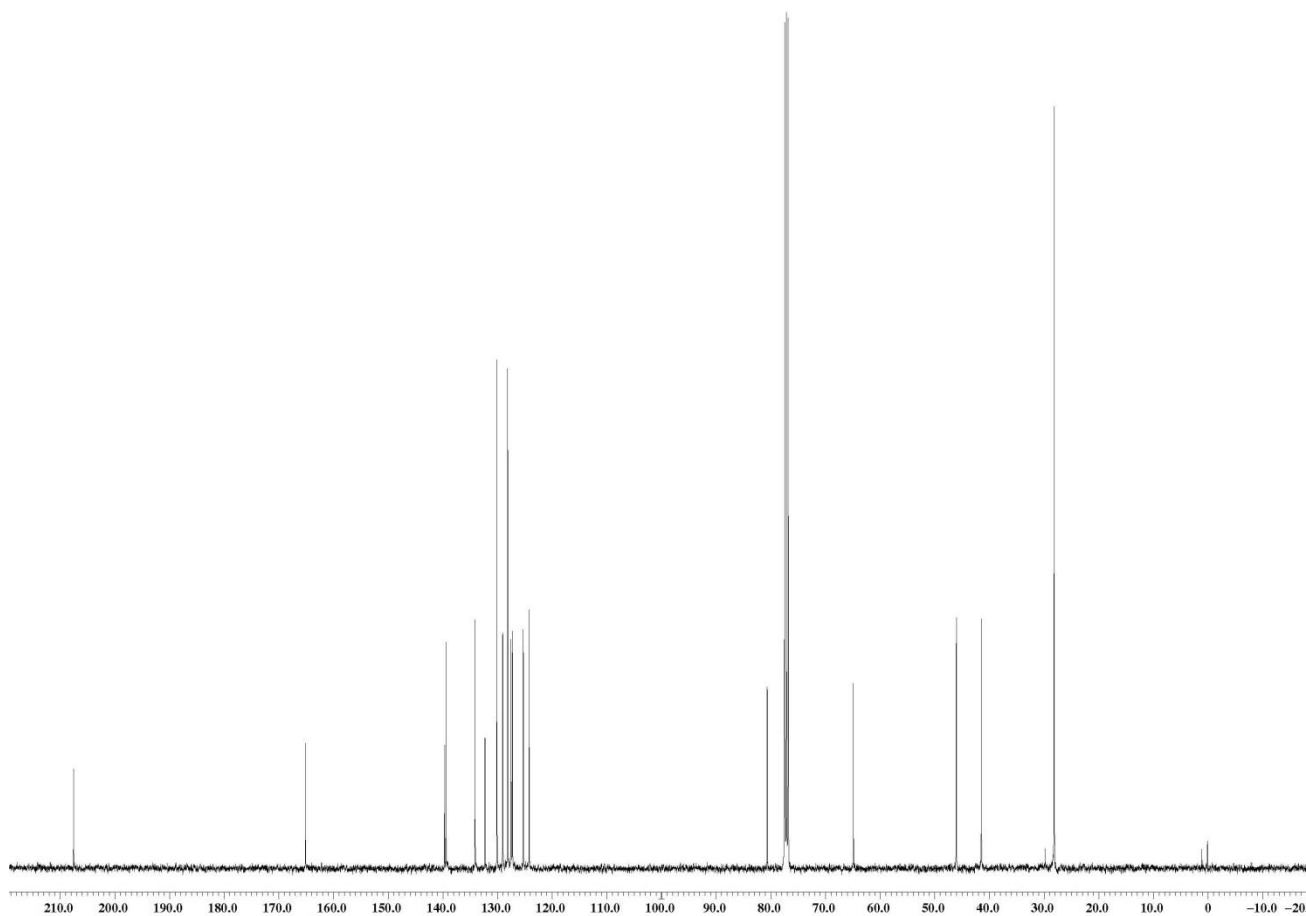
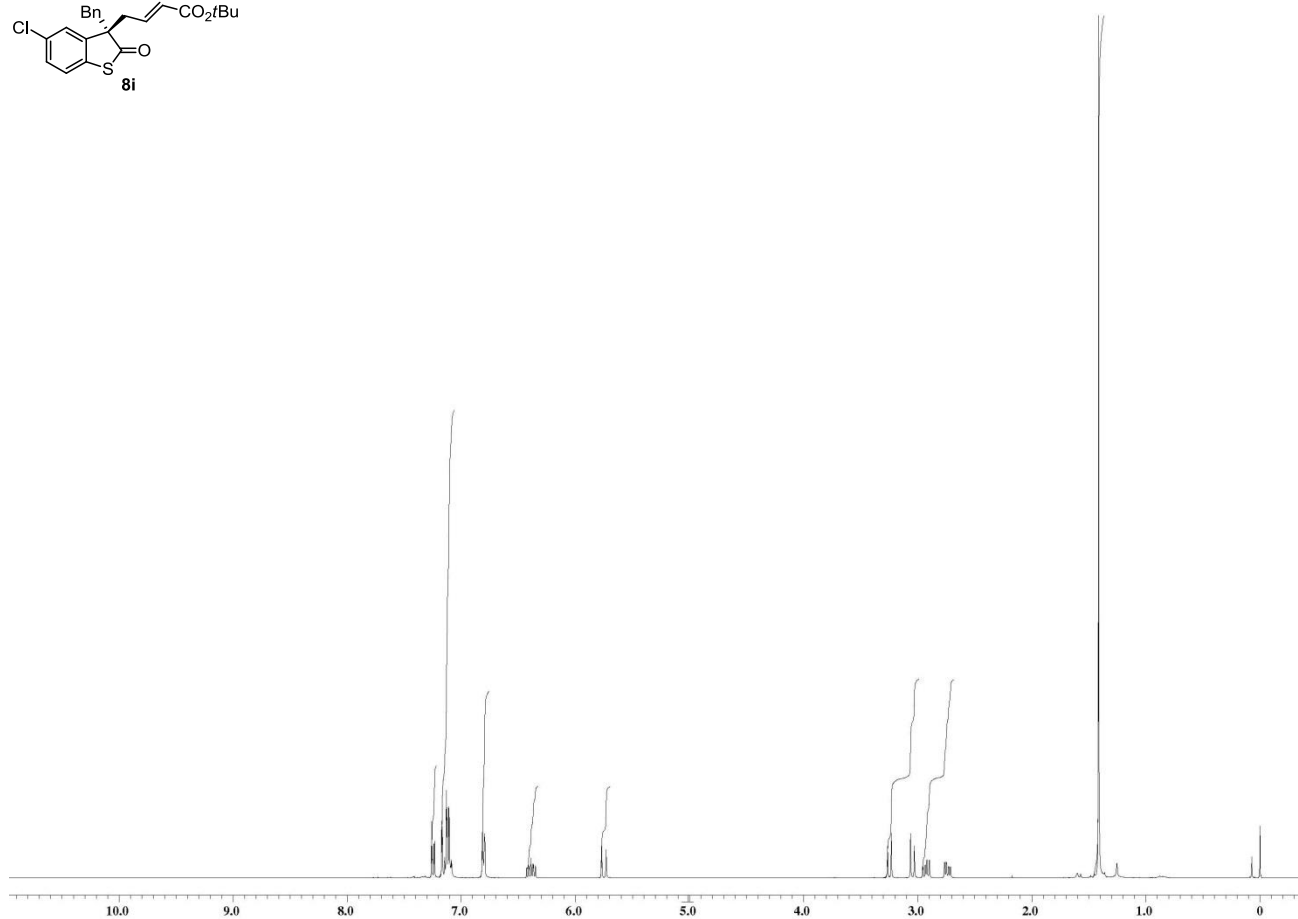
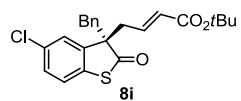


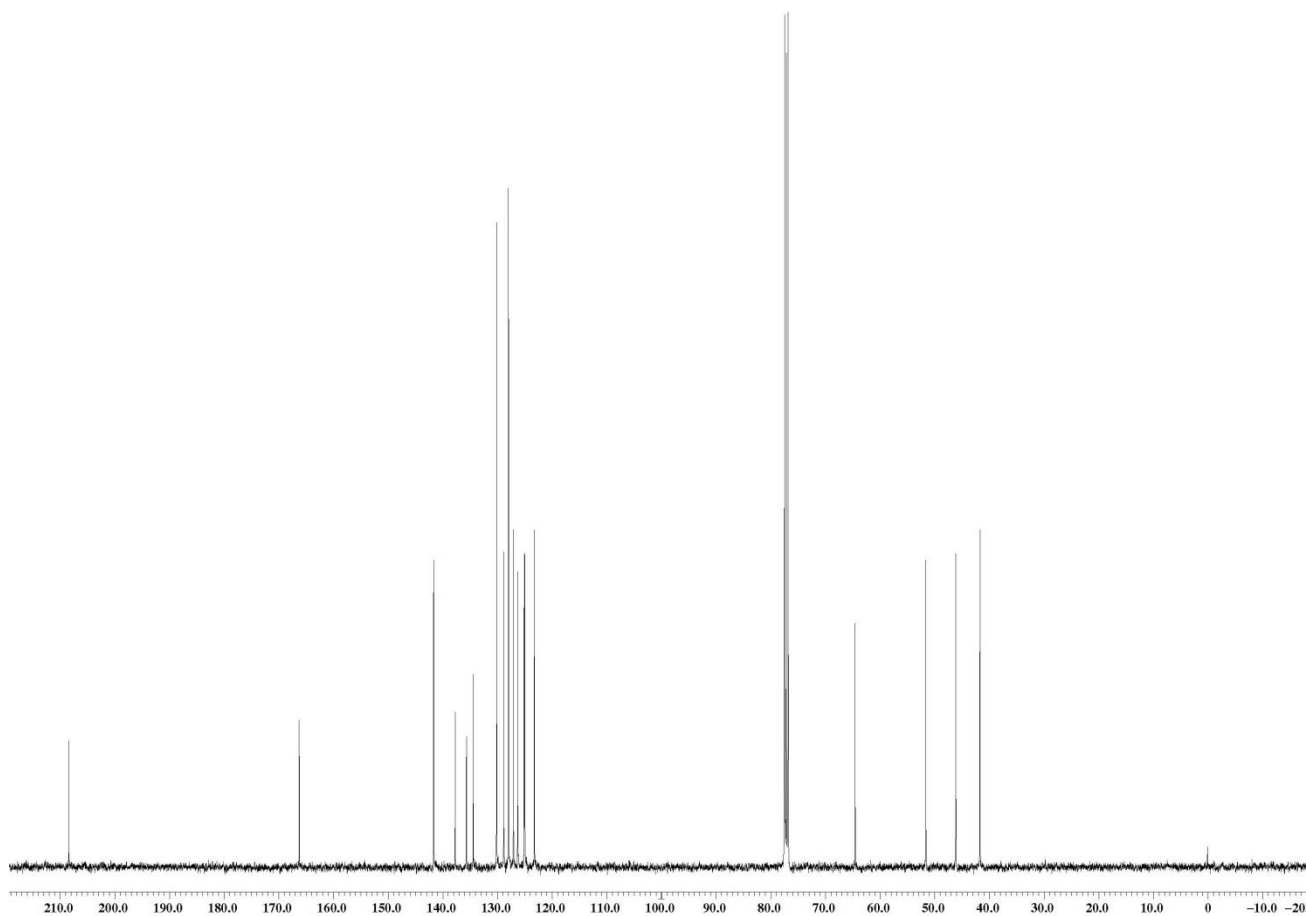
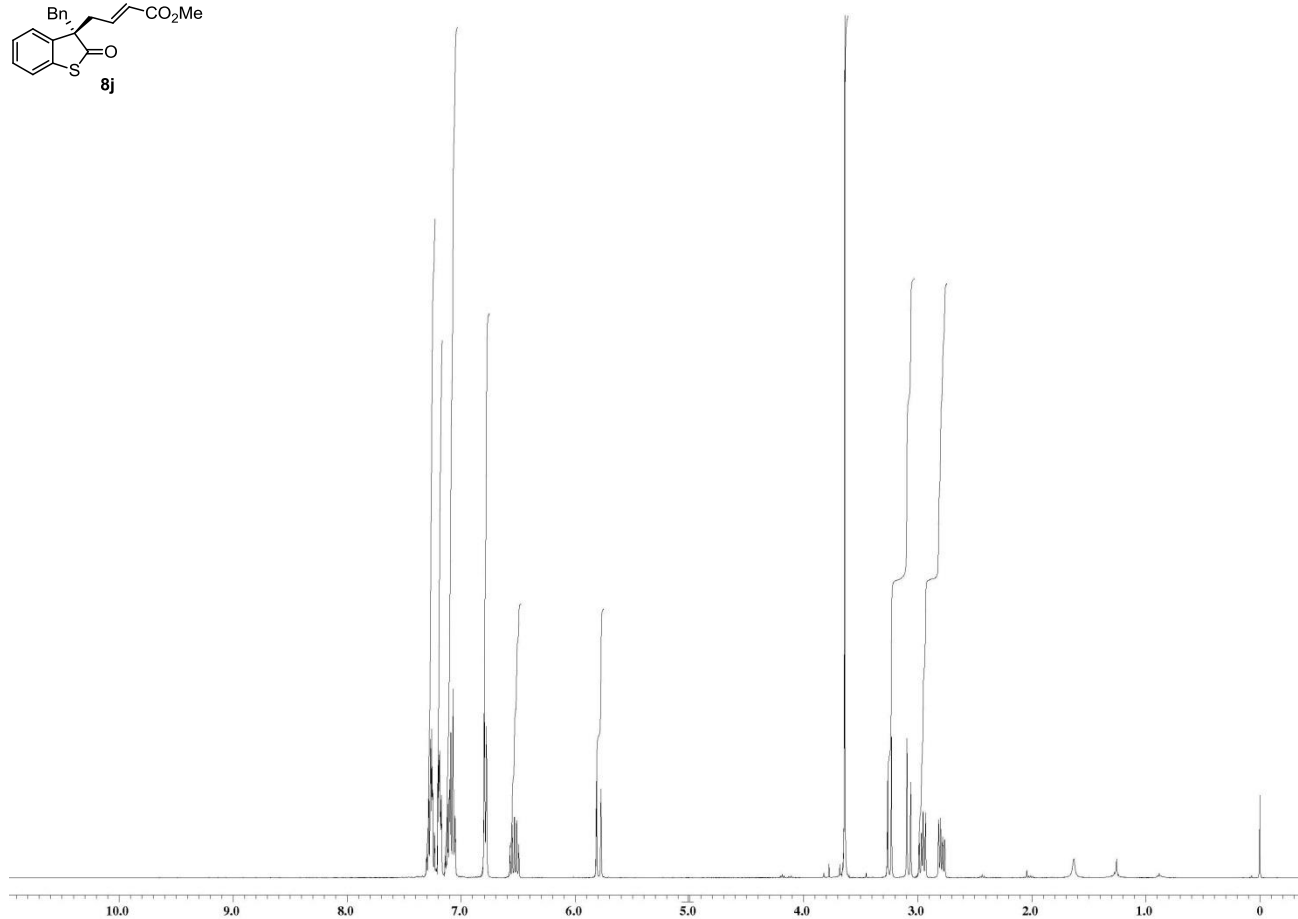
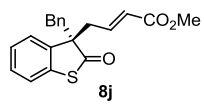


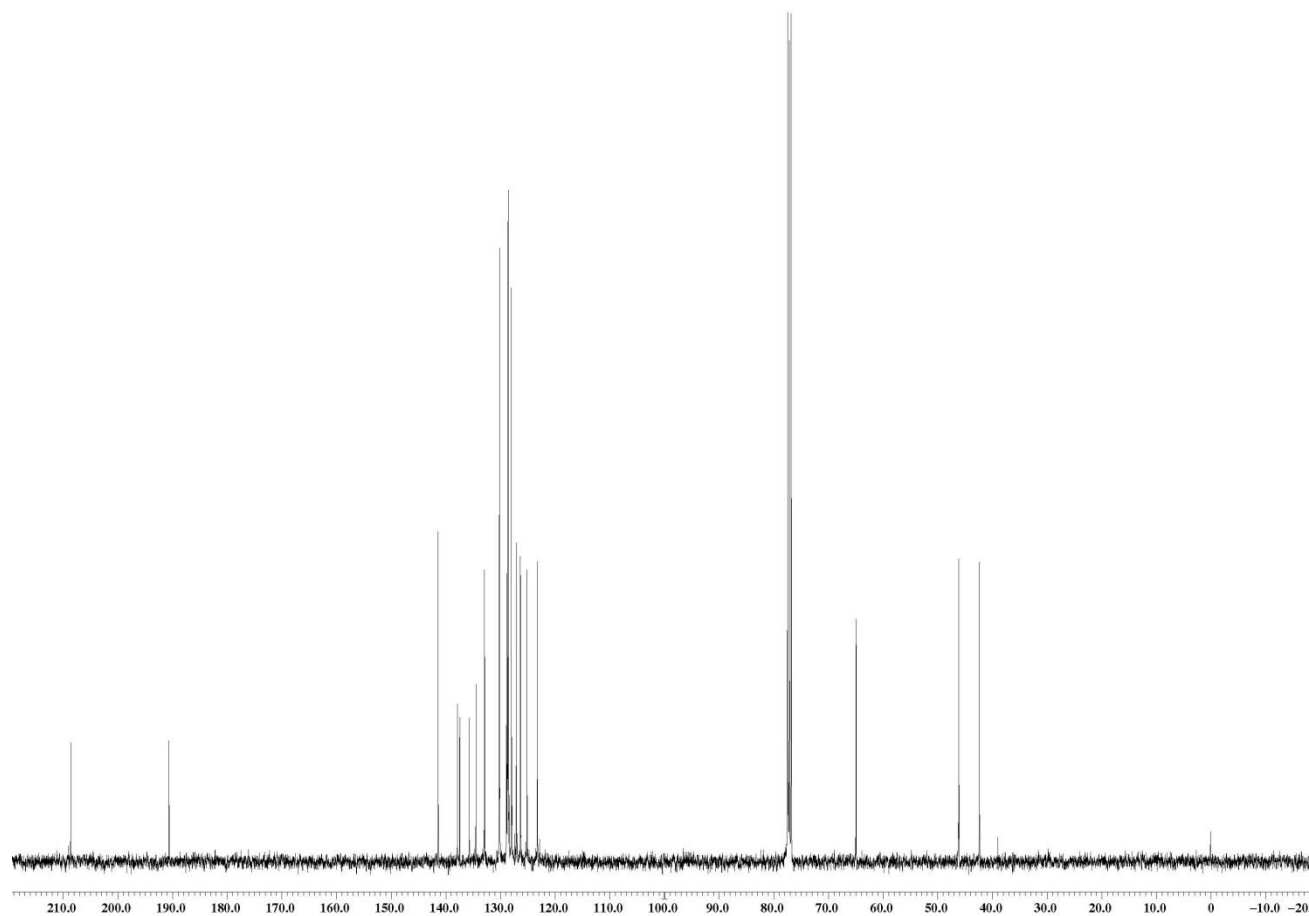
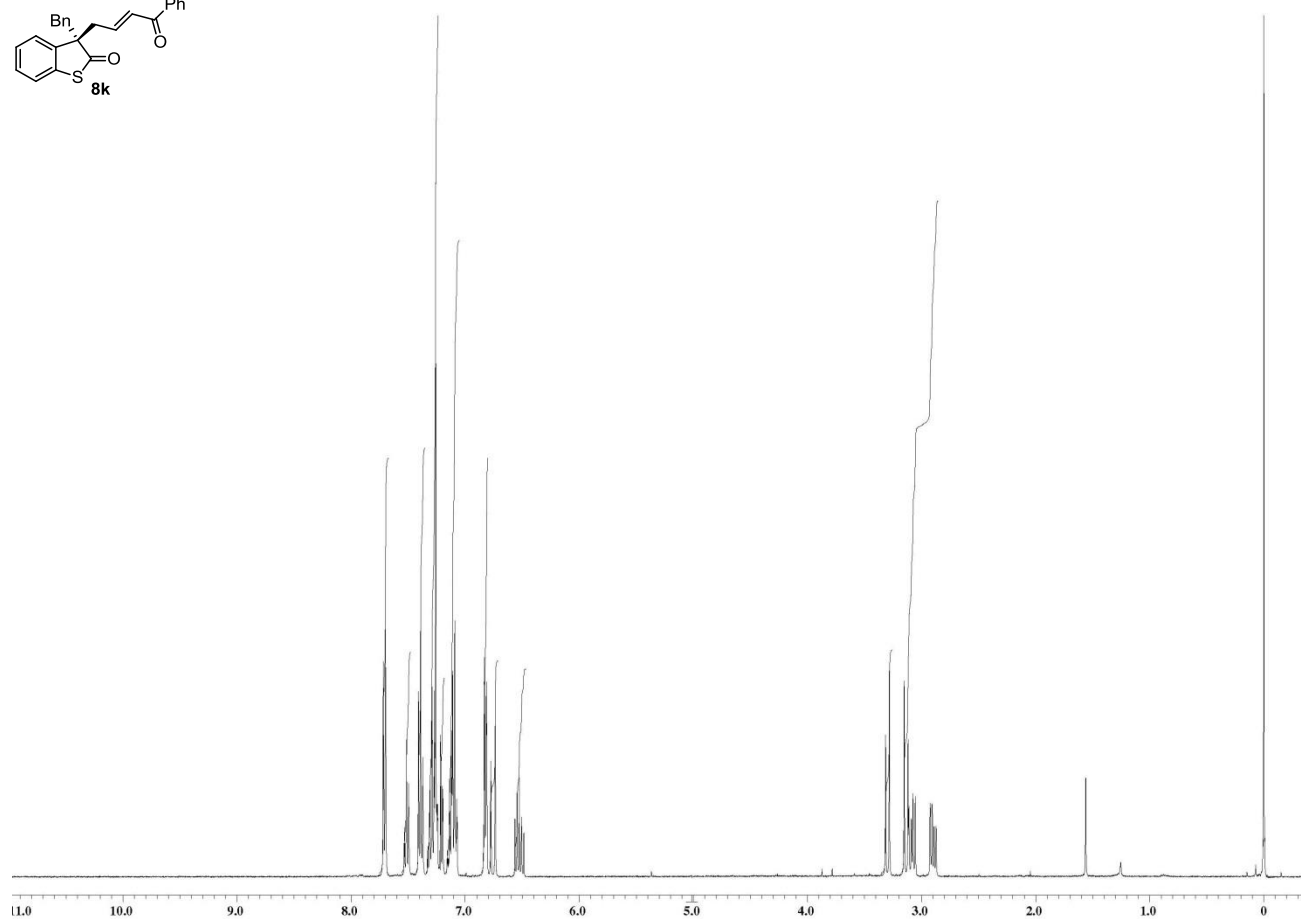
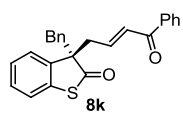






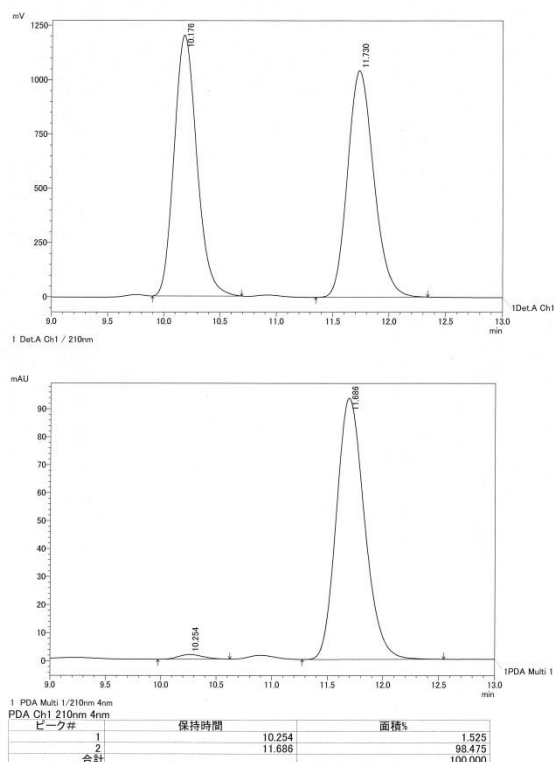




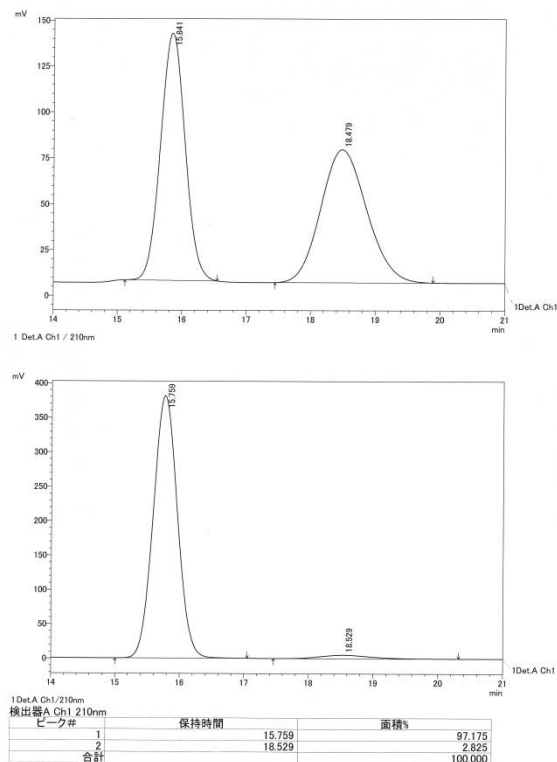


Copies of HPLC Chromatograms:

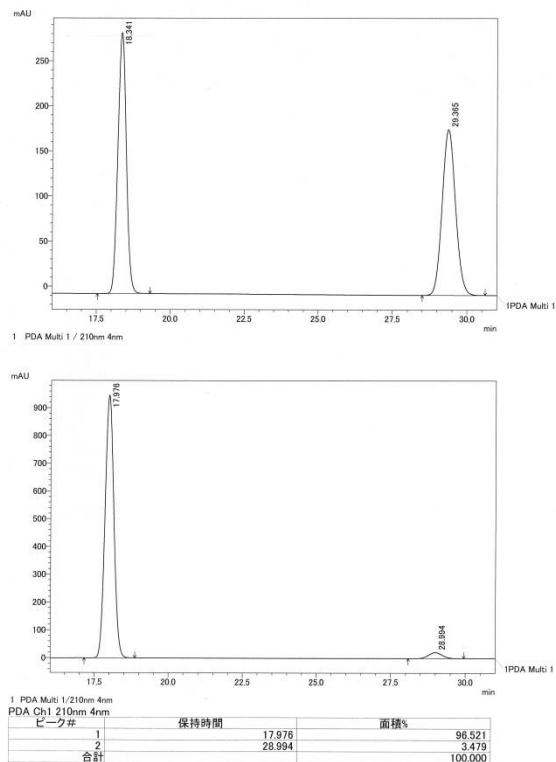
8a



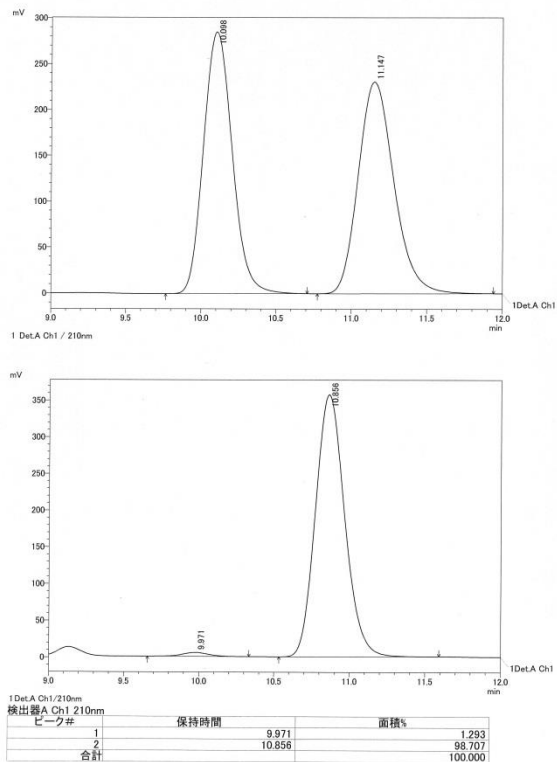
8c



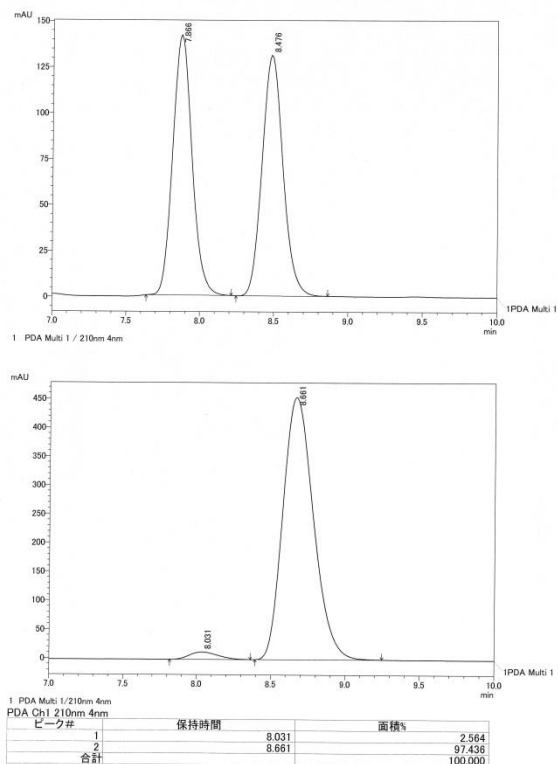
8b



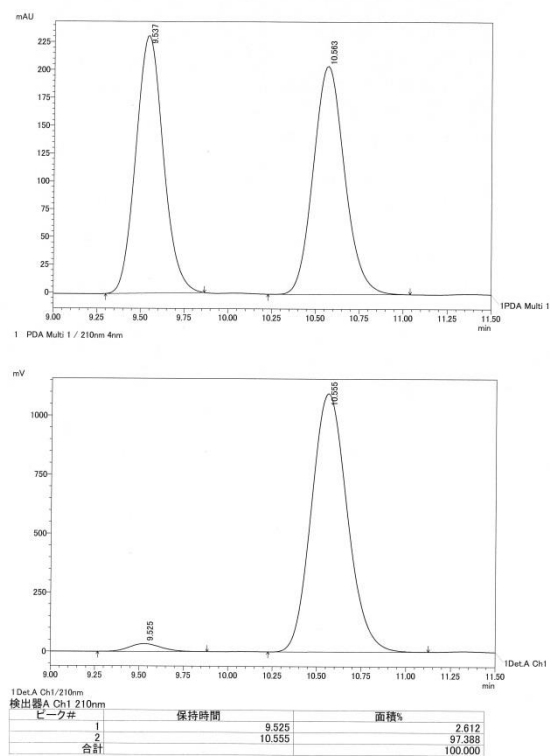
8d



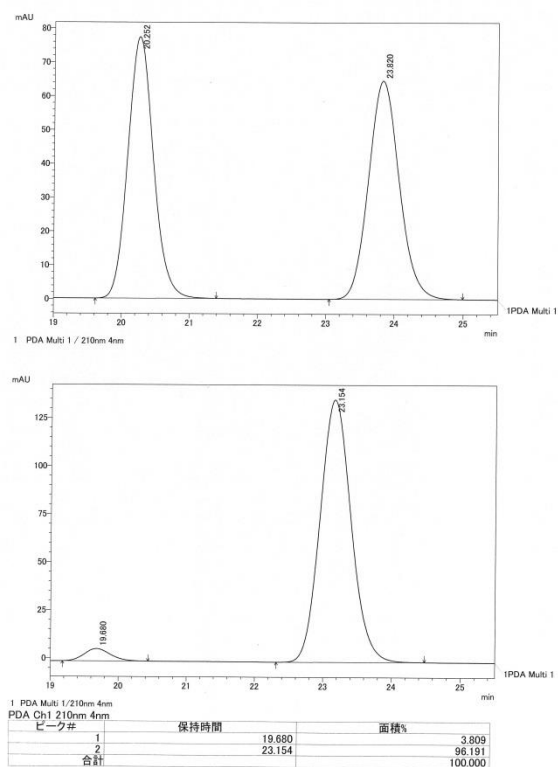
8e



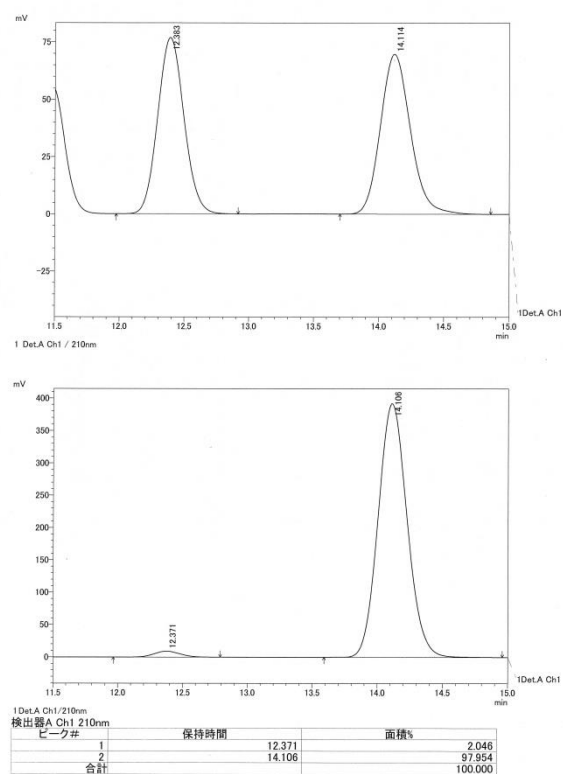
8g



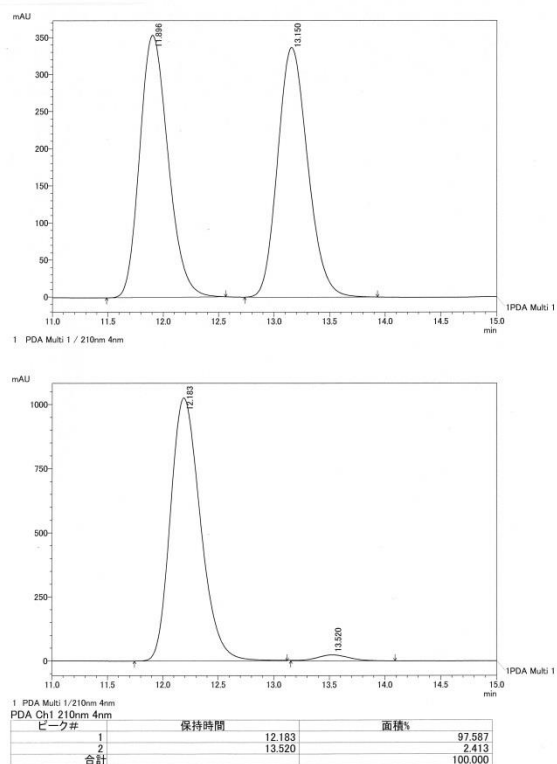
8f



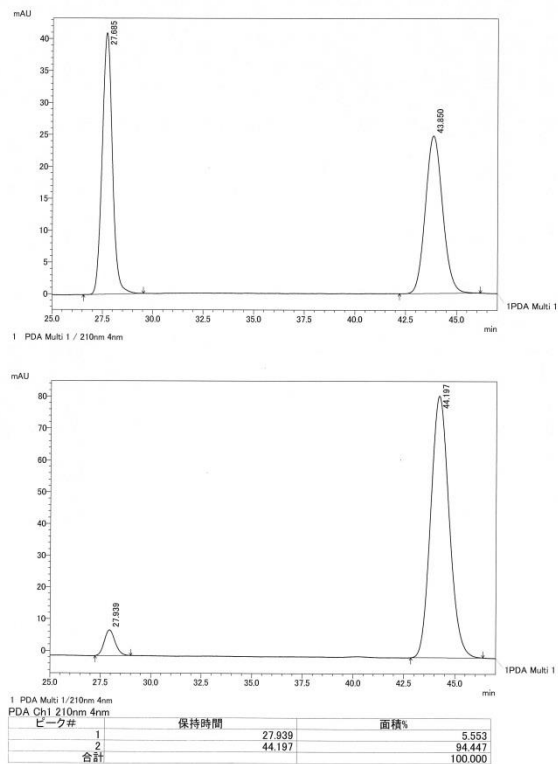
8h



8i



8k



8j

