

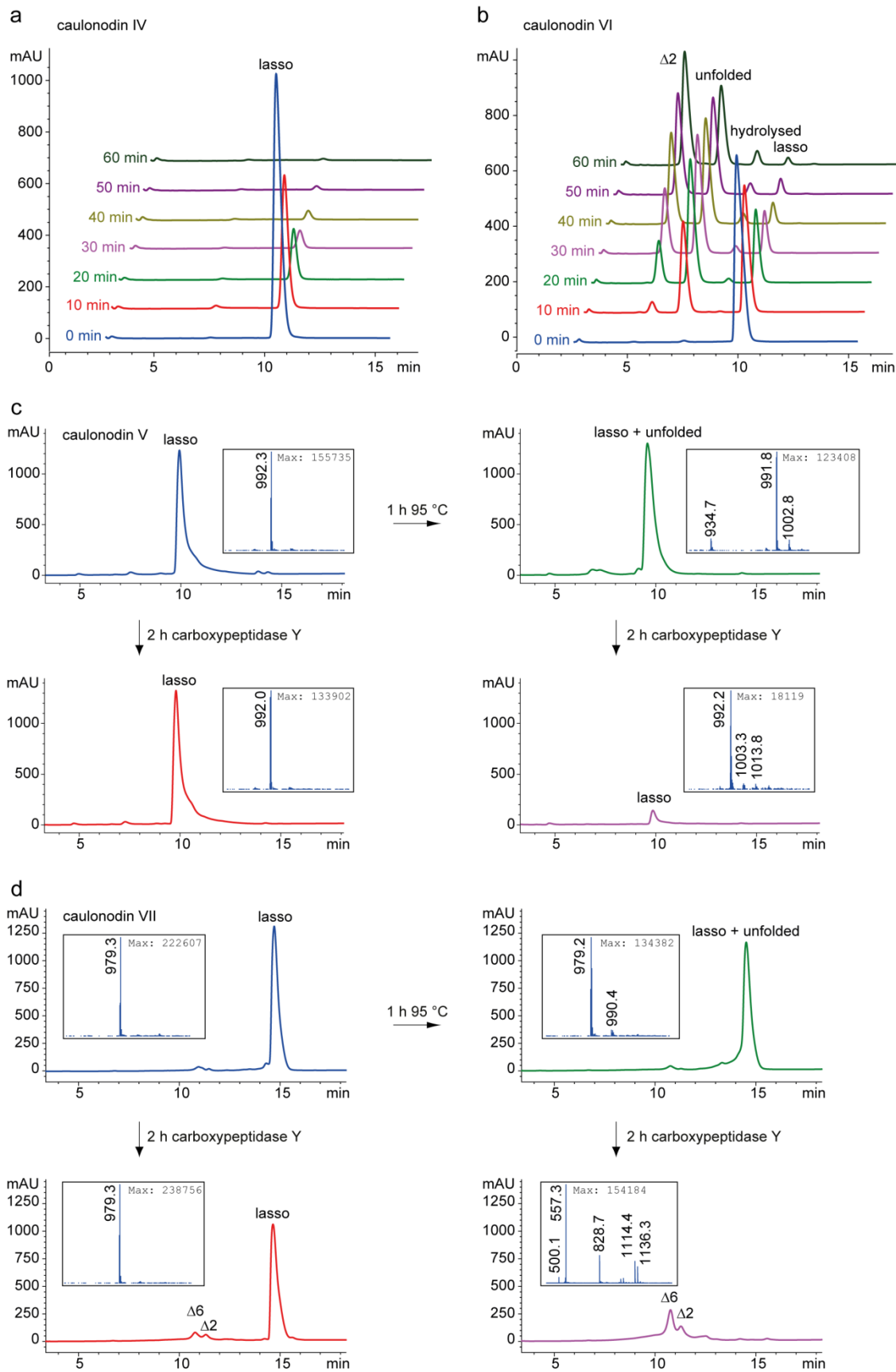
Characterization of Caulonodin Lasso Peptides Revealed
Unprecedented N-Terminal Residues and a Precursor Motif
Essential for Peptide Maturation

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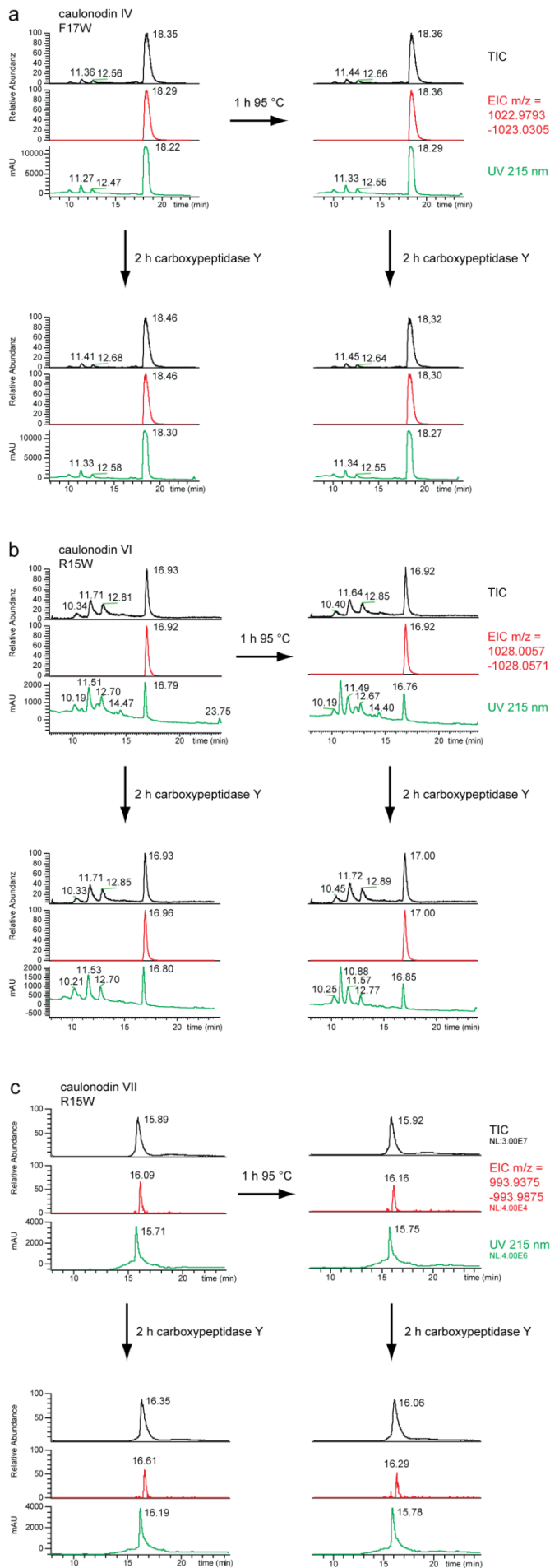
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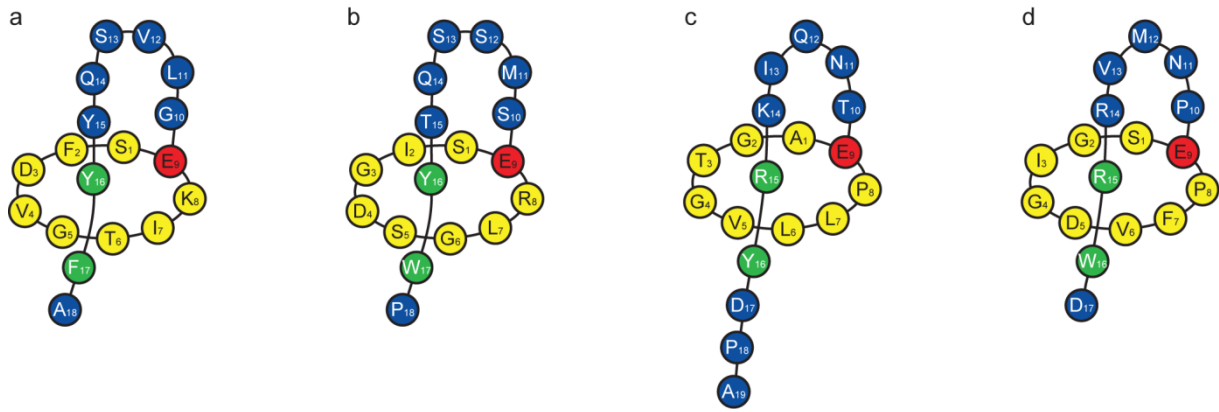
Supporting Information



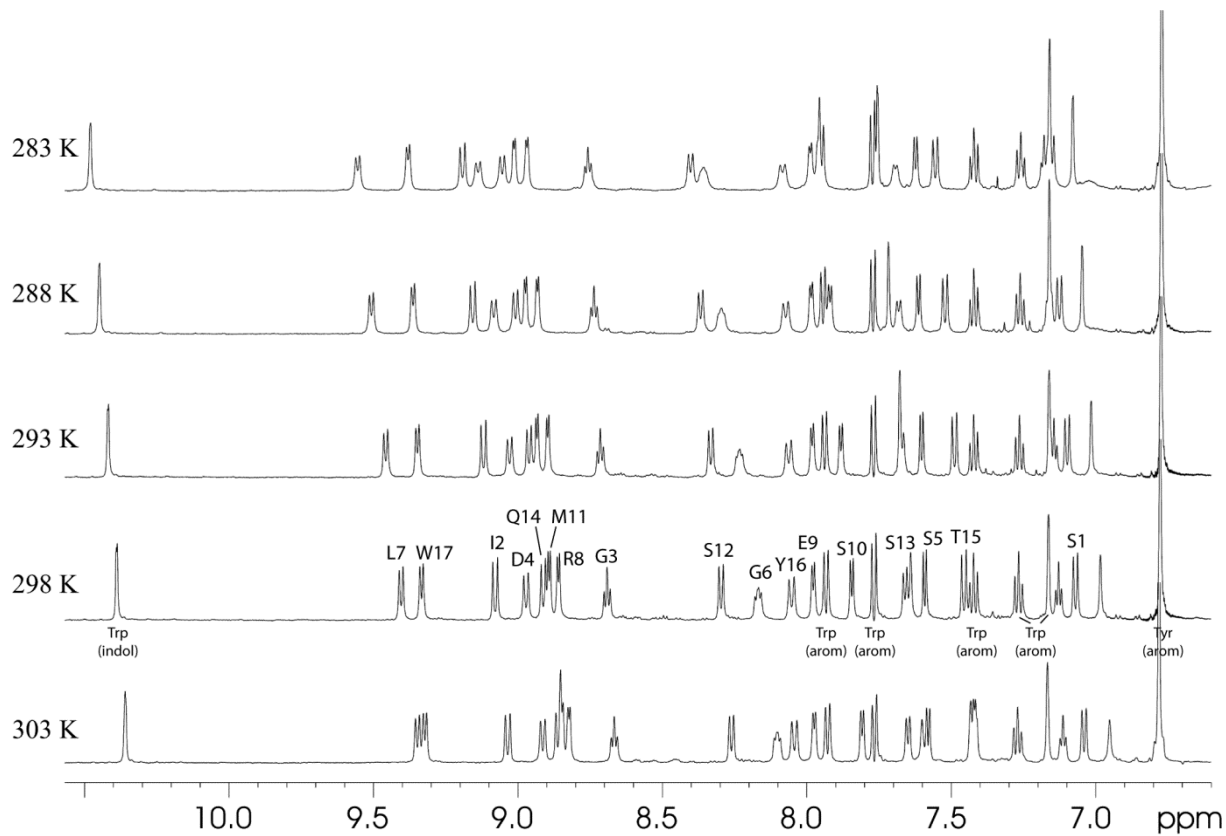
Supporting figure 1 – heat stability of **(a)** caulonodin IV and **(b)** caulonodin VI, heat and carboxypeptidase stability of **(c)** caulonodin V and **(d)** caulonodin VII shown as UV traces and mass signals measured on the low-resolution HPLC-MS system



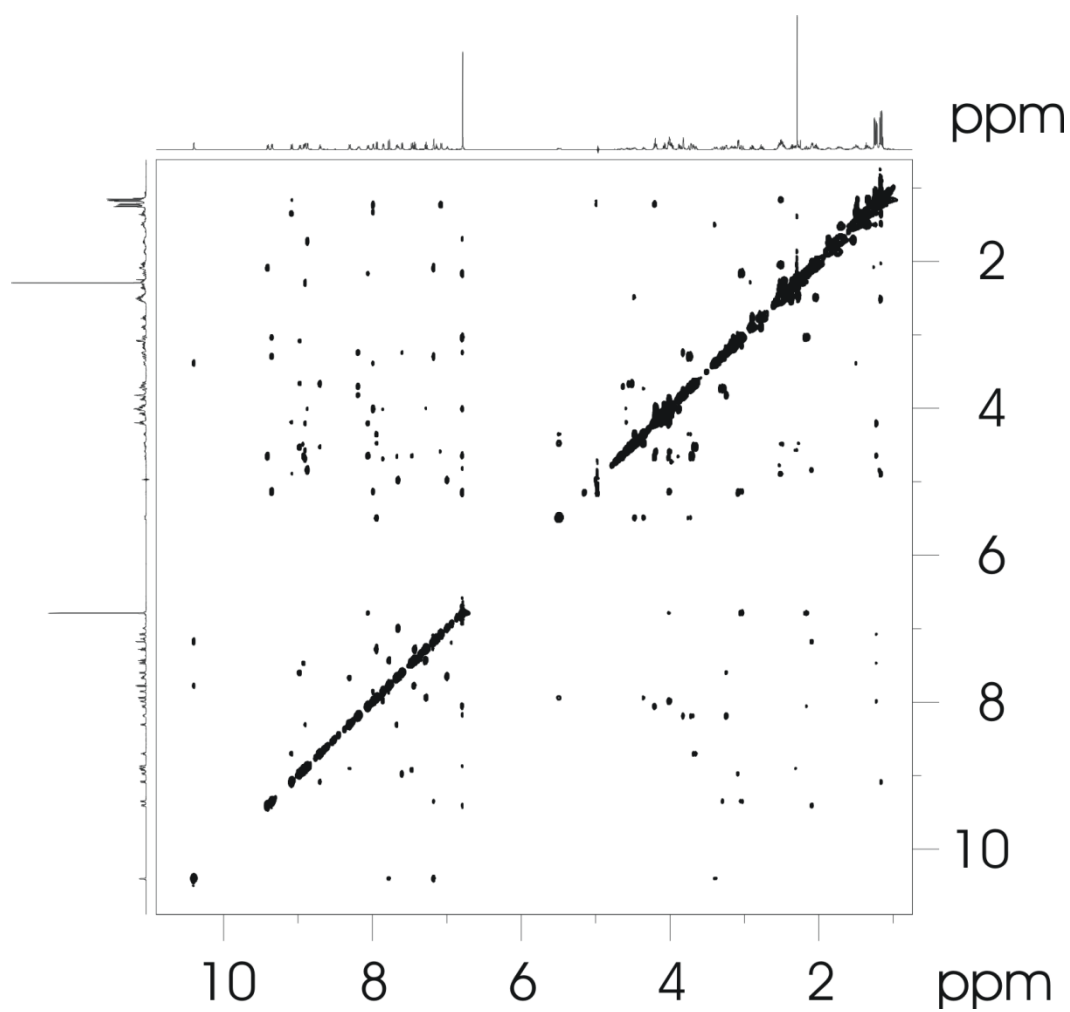
Supporting figure 2 – heat stable mutants of **(a)** caulonodin IV, **(b)** caulonodin VI and **(c)** caulonodin VII shown as total ion current (TIC), extracted ion current (EIC) and UV traces measured on the high-resolution HPLC-MS system



Supporting figure 3 – 3D fold predictions of the caulonodins **(a)** IV, **(b)** V, **(c)** VI and **(d)** VII in a schematic representation based on the plug mutants; aa involved in the formation of the macrolactam ring shown in red, other aa of the ring shown in yellow, aa in the tail shown in blue, plug aa shown in green.

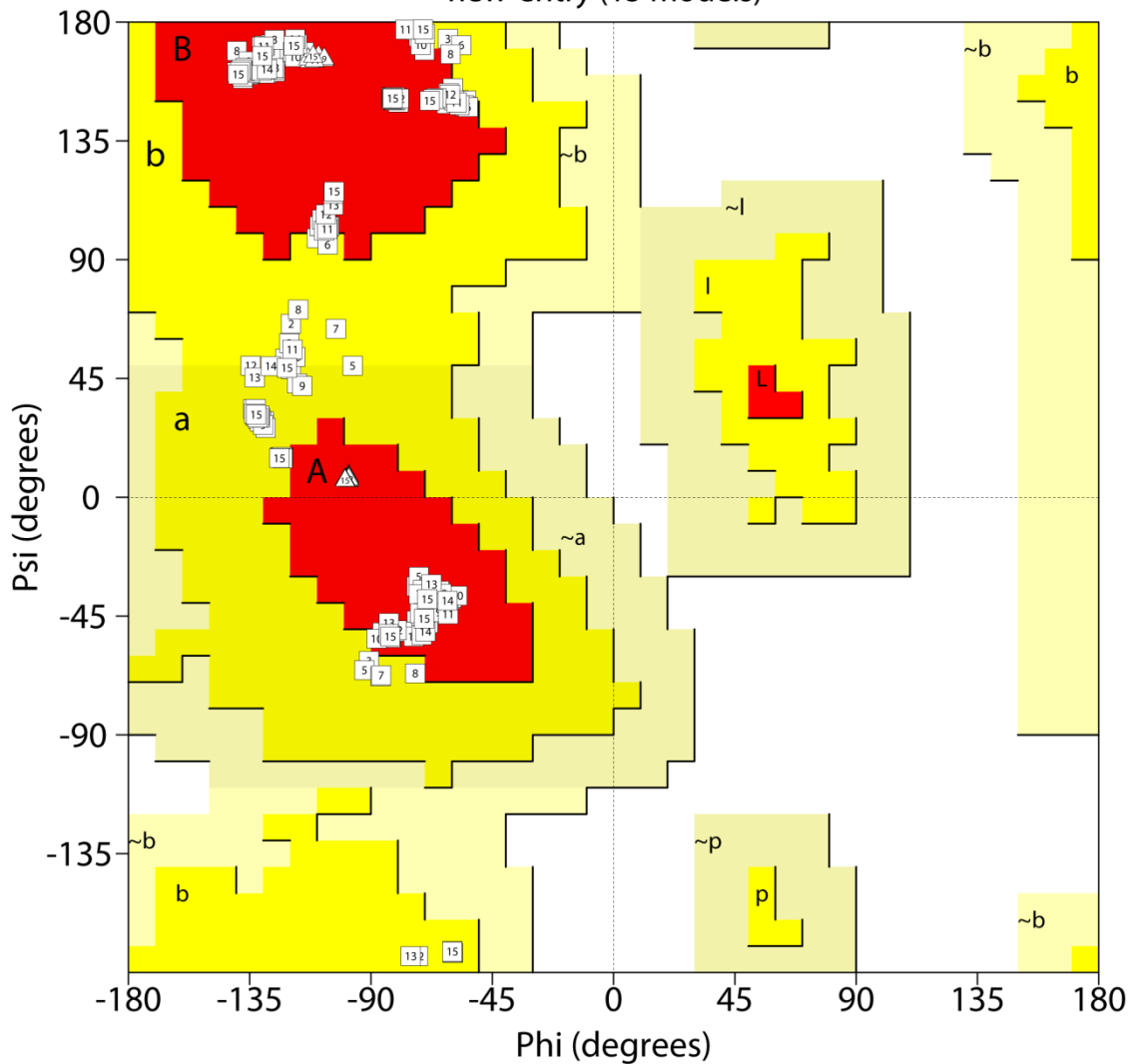


Supporting figure 4 – ^1H NMR spectra of caulonodin V in $\text{H}_2\text{O}/\text{D}_2\text{O}$ (9:1) at variable temperatures in the range from 10.6 to 6.6 ppm



Supporting figure 5 – 2D ¹H NOESY spectrum of caulonodin V at 300 ms mixing time

Ramachandran Plot new-entry (15 models)

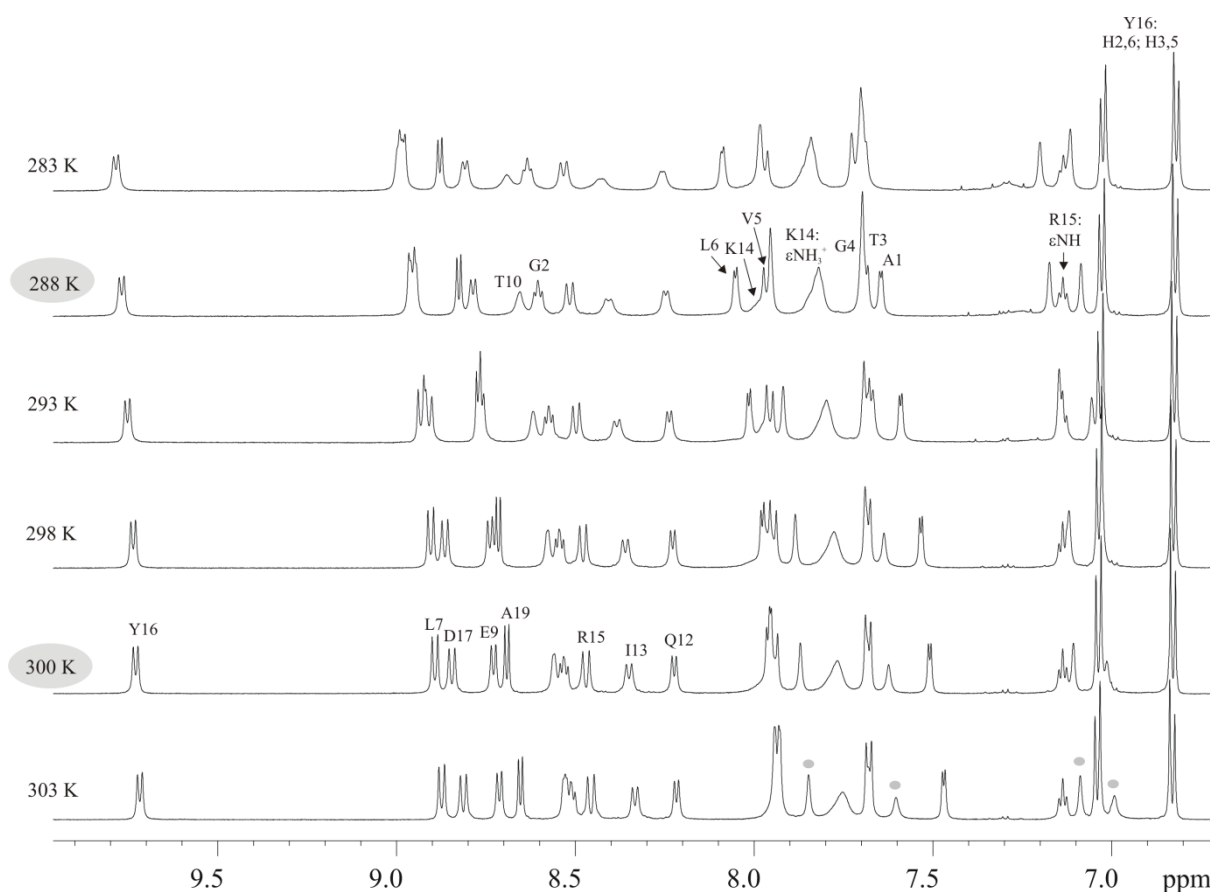


Plot statistics

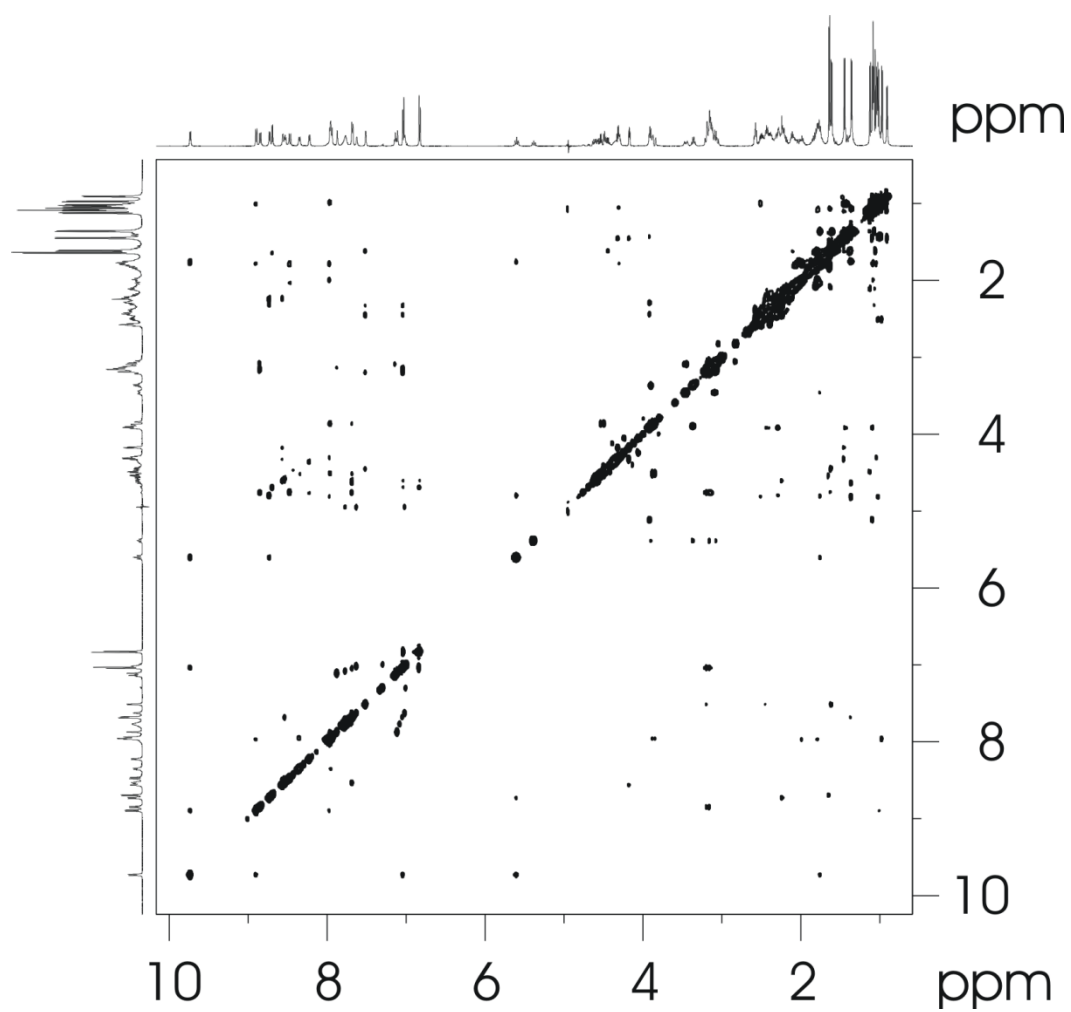
Residues in most favored regions [A,B,L]	132	62.9%
Residues in additional allowed regions [a,b,l,p]	78	37.1%
Residues in generously allowed regions [~a,~b,~l,~p]	0	0.0%
Residues in disallowed regions	0	0.0%
	----	----
Number of non-glycine and non-proline residues	210	100.0%
Number of end-residues (excl. Gly and Pro)	15	
Number of glycine residues (shown as triangles)	30	
Number of proline residues	15	
	----	----
Total number of residues	270	

Based on an analysis of 118 structures of resolution of at least 2.0 Angstroms and R-factor no greater than 20%, a good quality model would be expected to have over 90% in the most favoured regions.
Model numbers shown inside each data point.

Supporting figure 6 – Quality of the structure determination. Ramachandran Plot of the 15 minimum energy structures showing all residues in most favored and additionally allowed regions.



Supporting figure 7 – ^1H NMR spectra of caulonodin VI in $\text{H}_2\text{O}/\text{D}_2\text{O}$ (9:1) at variable temperatures in the range from 10.0 to 6.7 ppm; labeling according to signal assignments at 288 and 300 K; circles denote side chain NH_2 of N11 and Q12.



Supporting figure 8 – 2D ^1H NOESY spectrum of caulonodin VI at 300 ms mixing time

a

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1984  -MTPIQS-KFCLLRVGSAKRLTQS-FDVGTIKEGLVSQY----YFA- 39
198x  -MTQVSPSPRLRLIRVGRALDLTRS-IGDSGLRESMSSQT----YWP- 40
2238  ----MNTLKTRLIRFGSAKRLTRAGTGVL-LPETNQIKR----YDPA 38
2239  ----MTPKFRLIRLGSAKRLTRSGIGDV-FPEPNMVRP----WD-- 36
5194  MERIEDHIDDELIDLGAASVETQG--DVLNAPEPGIGRE-PTGLSRD 44
5193  MQRIIDETTDGLIELGAASVETQG--DVLFAPEPGVGRP-PMGLSED 44
519x  MEFEGIPSPDARIDLGLASEETCG--QIYDHPEVIGAYGCEGLQR- 44

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b

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1984  MTPIQ-SKFCLLRVGSAKRLTQSFDVGTIKEGLVSQYYFA- 39
198x  MTQVSPSPRLRLIRVGRALDLTRSIGDSGLRESMSSQTYWP- 40
2238  MNTLK---TRLIRFGSAKRLTRAGTGVLLPETNQIKRYDPA 38
2239  MTPK---FRLIRLGSAKRLTRSGIGDVFPEPNMVRWD-- 36

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similarity 

Supporting figure 9 – Multiple alignments using the gonnet protein weight matrix of **(a)** all seven caulonodin precursors and **(b)** the four chromosomal precursors; coloring according to similarity from red for identical and orange to yellow and white for different; arrow marks proteolytic cleavage site between leader and core peptide

Name	Start	p-value	Sites ?
Caulonodin_I	10	8.15e-15	MERIEDHID DELIDLGAASVETQ
Caulonodin_II	10	7.81e-14	MQRIIDETT DGLIELGAASVETQ
Caulonodin_VII	6	2.77e-12	MTTPK FRLIRLGS AKRLTR
Sphingopyxin_II	8	5.99e-12	MERTEVI EEVIDLGKASVETK
Caulosegnin_II	5	1.24e-11	MTKT HRLIRLGD AQRLTQ
Sphingopyxin_I	5	1.43e-11	MKDF NELIDLGAISVETR
Astexin-1	15	1.43e-11	I ISETVQPKT AGLIVLGKASAETR
Sphingonodin_I	5	5.42e-11	MERD NDVIELGAVSVETK
Xanthomonin_III	16	8.89e-11	DSAPSQEDHG KDIIVLG IASVETQ
Caulosegnin_I	10	1.61e-10	MTKKNATQA PRLVRVGD AHRLTQ
Astexin-3	12	2.28e-10	TKRTTIAARR VGLIDL GKATRQTK
Caulonodin_IV	8	3.19e-10	MTPIQSK FCLLRVGS AKRLTQ
Syanodin	10	4.43e-10	MERNSEDRR DDVVELGAVSVETK
Sphingonodin_II	10	4.94e-10	MDRHDNSEV DEIIDLGTASAVTQ
Xanthomonin_II	15	5.49e-10	NNDARTTALD QDLIVLGVASLDTQ
Caulonodin_VI	6	5.49e-10	MNTLK TRLIRFGS AKRLTR
→ Rhodanodin	21	1.03e-09	TNENIRSNAQ DDVIELSVASVETK
Rubrivinodin	12	1.26e-09	KEFAMDEELE LEIVDLGDAKELTQ
Caulosegnin_III	5	1.54e-09	MTSR FQLRLGKADRLTR
SSV-2083	23	1.87e-09	MTSTDELYEA PELIEIGDYAELTR
Caulonodin_III	10	2.51e-09	MEFEGIPSP DARIDLGLASEETC
Xanthomonin_I	13	2.16e-08	SNDTTHSDAS NEITVLGVASTDTK
→ Caulonodin_V	9	6.72e-08	MTQVSPSP LRLIRVGRALDLTR
Astexin-2	12	1.81e-07	RTYNRSLPAR AGLTDLGKVTTHTK
Zucinodin	10	4.61e-07	MTRLLNLS VRLGFGS AKAAATN
Lariatn	13	8.55e-07	SQPSKKTINA PSLVQRGKFARTTA
SRO15-2005	15	4.01e-06	IKQQKKAYVK PSMFQQGDFSKKTA

Supporting figure 10 – Alignment of the 27 leader peptides from the 30 known, functional lasso peptide biosynthetic gene clusters, wherein the motif was identified using the MEME algorithm; leader peptide of caulonodin V marked with a blue arrow, of rhodanodin with a red arrow

Supporting table 1 – Oligonucleotide primers

name	sequence
CK31_198x_A1_NdeI_FP	GGATTAC <u>CAT ATG</u> ACC CCG ATC CAA TCC AAG TTC TGC CTT CTG
CK31_198x_C_XhoI_RP	ATAT <u>CTC GAG</u> TCA TGA GAC CCT GAG CTT GGC TTC CCA GG
CK31_2238_A1_NdeI_FP	GGATTAC <u>CAT ATG</u> AAT ACG CTC AAG ACC CGG CTG ATC CG
CK31_2241_C_XhoI_RP	ATAT <u>CTC GAG</u> TTA GGC GCC GCG CGC TTC CCA G
CK31_198xB_SLIM_FPS	CTT TGG CTC GCC CAG GAC GTC CAT GC
CK31_198xB_rbs_SLIM_FPL	AGAGGAGAAATTAACC ATG CCC CTT TGG CTC GCC CAG GAC GTC CAT GC
CK31_198xA2_SLIM_RPS	TTA GGG CCA GTA GGT CTG CGA GCT CAT G
CK31_198xA2_rbs_SLIM_RPL	GGG CAT GGTAAATTTCTCCTCT TTA GGG CCA GTA GGT CTG CGA GCT CAT G
CK31_198xA2Del_SLIM_RPS	TCA GGC GAA GTA GTA TTG GCT GAC GAG ACC
CK31_198xA2Del_SLIM_RPL	GGG CAT GGTAAATTTCTCCTCT TCA GGC GAA GTA GTA TTG GCT GAC GAG ACC
CK31_198xA1Del_SLIM_FPS	ATG ACT CAA GTC TCT CCC TCG CCC CTG C
CK31_198xA1Del_SLIM_FPL	CTT TAA GAA GGA GAT ATA CAT ATG ACT CAA GTC TCT CCC TCG CCC CTG C
CK31_bothA1Del_SLIM_RPS	TTA AAC AAA ATT ATT TCT AGA GGG GAA TTG TTA TCC GCT CAC
CK31_bothA1Del_SLIM_RPL	ATG TAT ATC TCC TTC TTA AAG TTA AAC AAA ATT ATT TCT AGA GGG GAA TTG TTA TCC GCT CAC
CK31_22xxB_SLIM_FPS	CTA ACC TGG CGG CCC GGC GTT CAC
CK31_22xxB_SLIM_FPL	AGAGGAGAAATTAACC ATG ACG CTA ACC TGG CGG CCC GGC GTT CAC
CK31_22xxA2_SLIM_RPS	TCA GTC CCA ACG GCG AAC CAT GTT GGG C
CK31_22xxA2_SLIM_RPL	CGT CAT GGTAAATTTCTCCTCT TCA GTC CCA ACG GCG AAC CAT GTT GGG C
CK31_22xxA2Del_SLIM_RPS	TCA GGC CGG GTC GTA CCG CTT GAT CTG
CK31_22xxA2Del_SLIM_RPL	GGG CAT GGTAAATTTCTCCTCT TCA GGC CGG GTC GTA CCG CTT GAT CTG
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CK31_22xxA1Del_SLIM_FPL	CTT TAA GAA GGA GAT ATA CAT ATG ACG ACT CCC AAG TTT CGA CTG ATC CGT CTG
198xA12_SLIM_FPS	TGA AGA GGA GAA ATT AAC CAT GCC CCT TTG GCT C
1984A1_SLIM_RPS	GAG ACC TTC CTT GAT GGT GCC GAC ATC GAA G
1984A1_Y15A_FPL	GTC AGC CAA GCC TAC TTC GCC TGA AGA GGA GAA ATT AAC CAT GCC CCT TTG GCT C
1984A1_Y15A_RPL	GGC GAA GTA GGC TTG GCT GAC GAG ACC TTC CTT GAT GGT GCC GAC ATC GAA G
1984A1_Y16A_FPL	GTC AGC CAA TAC GCC TTC GCC TGA AGA GGA GAA ATT AAC CAT GCC CCT TTG GCT C
1984A1_Y16A_RPL	GGC GAA GGC GTA TTG GCT GAC GAG ACC TTC CTT GAT GGT GCC GAC ATC GAA G
1984A1_F17A_FPL	GTC AGC CAA TAC TAC GCC GCC TGA AGA GGA GAA ATT AAC CAT GCC CCT TTG GCT C
1984A1_F17A_RPL	GGC GGC GTA GTA TTG GCT GAC GAG ACC TTC CTT GAT GGT GCC GAC ATC GAA G
1984A1_Y15W_FPL	GTC AGC CAA TGG TAC TTC GCC TGA AGA GGA GAA ATT AAC CAT GCC CCT TTG GCT C
1984A1_Y15W_RPL	GGC GAA GTA CCA TTG GCT GAC GAG ACC TTC CTT GAT GGT GCC GAC ATC GAA G
1984A1_Y16W_FPL	GTC AGC CAA TAC TGG TTC GCC TGA AGA GGA GAA ATT AAC CAT GCC CCT TTG GCT C
1984A1_Y16W_RPL	GGC GAA CCA GTA TTG GCT GAC GAG ACC TTC CTT GAT GGT GCC GAC ATC GAA G
1984A1_F17W_FPL	GTC AGC CAA TAC TAC TGG GCC TGA AGA GGA GAA ATT AAC CAT GCC CCT TTG GCT C

name	sequence
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198xA2_SLIM_RPS	CAT GGA CTC CCG CAG ACC GGA GTC G
198xA2_T15A_FPL	AGC TCG CAG GCC TAC TGG CCC TGA AGA GGA GAA ATT AAC CAT GCC CCT TTG GCT C
198xA2_T15A_RPL	GGG CCA GTA GGC CTG CGA GCT CAT GGA CTC CCG CAG ACC GGA GTC G
198xA2_Y16A_FPL	AGC TCG CAG ACC GCC TGG CCC TGA AGA GGA GAA ATT AAC CAT GCC CCT TTG GCT C
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198xA2_Y16F_FPL	AGC TCG CAG ACC TTC TGG CCC TGA AGA GGA GAA ATT AAC CAT GCC CCT TTG GCT C
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name	sequence
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name	sequence
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CK31_2238_A1V_RPL	CCC GGT GCC GAC GCG GGT CAG GCG CTT GGC CGA GCC GAA ACG G
CK31_2238_A1T_FPL	CTG ACC CGC ACC GGC ACC GGG GTG CTT CTC CCG GAA ACC AAC CAG ATC AAG C
CK31_2238_A1T_RPL	CCC GGT GCC GGT GCG GGT CAG GCG CTT GGC CGA GCC GAA ACG G
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198xLeaderDel3FPL	ATC CGT GTC ACC CGT TCG ATC GGC GAC TCC GGT CTG CGG GAG TCC
198xLeaderDel3RPL	CGA ACG GGT GAC ACG GAT GAG CCG CAG GGG CGA GGG AGA GAC TTG
198xLeaderBlock1 SLIM FPS	GAT CTG ACC CGT TCG ATC GGC GAC TCC
198xLeaderBlock1 SLIM RPS	CCG CAG GGG CGA GGG AGA GAC TTG
198xLeaderLIVAtoRRRR FPL	CGC CGT CGT CGC GGG CGC CGT CTC GAT CTG ACC CGT TCG ATC GGC GAC TCC
198xLeaderLIVAtoRRRR RPL	GAG ACG GCG CCC GCG ACG ACG GCG CCG CAG GGG CGA GGG AGA GAC TTG
198xLeaderLIVAtoEEEE FPL	GAA GAG CGT GAA GGG CGC GAA CTC GAT CTG ACC CGT TCG ATC GGC GAC TCC
198xLeaderLIVAtoEEEE RPL	GAG TTC GCG CCC TTC ACG CTC TTC CCG CAG GGG CGA GGG AGA GAC TTG
198x_LI-11-12toRR FPL	CGC CGT CGT GTC GGG CGC GCC CTC GAT CTG ACC CGT TCG ATC GGC GAC TCC
198x_LI-11-12toRR RPL	GAG GGC GCG CCC GAC ACG ACG GCG CCG CAG GGG CGA GGG AGA GAC TTG
198x_LI-11-12toEE FPL	GAA GAG CGT GTC GGG CGC GCC CTC GAT CTG ACC CGT TCG ATC GGC GAC TCC
198x_LI-11-12toEE RPL	GAG GGC GCG CCC GAC ACG CTC TTC CCG CAG GGG CGA GGG AGA GAC TTG
198x_LI-11-12toQQ FPL	CAA CAG CGT GTC GGG CGC GCC CTC GAT CTG ACC CGT TCG ATC GGC GAC TCC
198x_LI-11-12toQQ RPL	GAG GGC GCG CCC GAC ACG CTG TTG CCG CAG GGG CGA GGG AGA GAC TTG
198x_VA-9-6toRR FPL	CTC ATC CGT CGC GGG CGC CGT CTC GAT CTG ACC CGT TCG ATC GGC GAC TCC

name	sequence
198x_VA-9-6toRR RPL	GAG ACG GCG CCC GCG ACG GAT GAG CCG CAG GGG CGA GGG AGA GAC TTG
198x_VA-9-6toEE FPL	CTC ATC CGT GAA GGG CGC GAA CTC GAT CTG ACC CGT TCG ATC GGC GAC TCC
198x_VA-9-6toEE RPL	GAG TTC GCG CCC TTC ACG GAT GAG CCG CAG GGG CGA GGG AGA GAC TTG
198x_VA-9-6toQQ FPL	CTC ATC CGT CAG GGG CGC CAG CTC GAT CTG ACC CGT TCG ATC GGC GAC TCC
198x_VA-9-6toQQ RPL	GAG CTG GCG CCC CTG ACG GAT GAG CCG CAG GGG CGA GGG AGA GAC TTG
198x_V-9R FPL	CTC ATC CGT CGC GGG CGC GCC CTC GAT CTG ACC CGT TCG ATC GGC GAC TCC
198x_V-9R RPL	GAG GGC GCG CCC GCG ACG GAT GAG CCG CAG GGG CGA GGG AGA GAC TTG
198x_V-9E FPL	CTC ATC CGT GAA GGG CGC GCC CTC GAT CTG ACC CGT TCG ATC GGC GAC TCC
198x_V-9E RPL	GAG GGC GCG CCC TTC ACG GAT GAG CCG CAG GGG CGA GGG AGA GAC TTG
198x_A-6R FPL	CTC ATC CGT GTC GGG CGC CGT CTC GAT CTG ACC CGT TCG ATC GGC GAC TCC
198x_A-6R RPL	GAG ACG GCG CCC GAC ACG GAT GAG CCG CAG GGG CGA GGG AGA GAC TTG
198x_A-6E FPL	CTC ATC CGT GTC GGG CGC GAA CTC GAT CTG ACC CGT TCG ATC GGC GAC TCC
198x_A-6E RPL	GAG TTC GCG CCC GAC ACG GAT GAG CCG CAG GGG CGA GGG AGA GAC TTG
198x_G-8F FPL	CTC ATC CGT GTC TTC CGC GCC CTC GAT CTG ACC CGT TCG ATC GGC GAC TCC
198x_G-8F RPL	GAG GGC GCG GAA GAC ACG GAT GAG CCG CAG GGG CGA GGG AGA GAC TTG
198x_G-8R FPL	CTC ATC CGT GTC CGT CGC GCC CTC GAT CTG ACC CGT TCG ATC GGC GAC TCC
198x_G-8R RPL	GAG GGC GCG ACG GAC ACG GAT GAG CCG CAG GGG CGA GGG AGA GAC TTG
198x_G-8E FPL	CTC ATC CGT GTC GAA CGC GCC CTC GAT CTG ACC CGT TCG ATC GGC GAC TCC
198x_G-8E RPL	GAG GGC GCG TTC GAC ACG GAT GAG CCG CAG GGG CGA GGG AGA GAC TTG
198x_G-8A FPL	CTC ATC CGT GTC GCG CGC GCC CTC GAT CTG ACC CGT TCG ATC GGC GAC TCC
198x_G-8A RPL	GAG GGC GCG CGC GAC ACG GAT GAG CCG CAG GGG CGA GGG AGA GAC TTG
198x_G-8V FPL	CTC ATC CGT GTC GTG CGC GCC CTC GAT CTG ACC CGT TCG ATC GGC GAC TC
198x_G-8V RPL	GAG GGC GCG CAC GAC ACG GAT GAG CCG CAG GGG CGA GGG AGA GAC TTG
198x_G-8P FPL	CTC ATC CGT GTC CCG CGC GCC CTC GAT CTG ACC CGT TCG ATC GGC GAC TCC
198x_G-8P RPL	GAG GGC GCG CGG GAC ACG GAT GAG CCG CAG GGG CGA GGG AGA GAC TTG
198x_R-10A FPL	CTC ATC GCT GTC GGG CGC GCC CTC GAT CTG ACC CGT TCG ATC GGC GAC TCC
198x_R-10A RPL	GAG GGC GCG CCC GAC AGC GAT GAG CCG CAG GGG CGA GGG AGA GAC TTG
198x_R-10F FPL	CTC ATC TTT GTC GGG CGC GCC CTC GAT CTG ACC CGT TCG ATC GGC GAC TCC
198x_R-10F RPL	GAG GGC GCG CCC GAC AAA GAT GAG CCG CAG GGG CGA GGG AGA GAC TTG
198x_R-10E FPL	CTC ATC GAA GTC GGG CGC GCC CTC GAT CTG ACC CGT TCG ATC GGC GAC TCC

name	sequence
198x_R-10E_RPL	GAG GGC GCG CCC GAC TTC GAT GAG CCG CAG GGG CGA GGG AGA GAC TTG
198xLeaderBlock 2SLIM FPS	TCC GGT CTG CGG GAG TCC ATG AGC TC
198xLeaderBlock 2SLIM RPS	GGC GCG CCC GAC ACG GAT GAG C
198xR-1A_FPL	CTC GAT CTG ACC GCC TCG ATC GGC GAC TCC GGT CTG CGG GAG TCC ATG AGC TC
198xR-1A_RPL	GTC GCC GAT CGA GGC GGT CAG ATC GAG GGC GCG CCC GAC ACG GAT GAG C
198xR-1Q_FPL	CTC GAT CTG ACC CAG TCG ATC GGC GAC TCC GGT CTG CGG GAG TCC ATG AGC TC
198xR-1Q_RPL	GTC GCC GAT CGA CTG GGT CAG ATC GAG GGC GCG CCC GAC ACG GAT GAG C
198xR-1N_FPL	CTC GAT CTG ACC AAC TCG ATC GGC GAC TCC GGT CTG CGG GAG TCC ATG AGC TC
198xR-1N_RPL	GTC GCC GAT CGA GTT GGT CAG ATC GAG GGC GCG CCC GAC ACG GAT GAG C
198xR-1K_FPL	CTC GAT CTG ACC AAG TCG ATC GGC GAC TCC GGT CTG CGG GAG TCC ATG AGC TC
198xR-1K_RPL	GTC GCC GAT CGA CTT GGT CAG ATC GAG GGC GCG CCC GAC ACG GAT GAG C
198xR-1H_FPL	CTC GAT CTG ACC CAC TCG ATC GGC GAC TCC GGT CTG CGG GAG TCC ATG AGC TC
198xR-1H_RPL	GTC GCC GAT CGA GTG GGT CAG ATC GAG GGC GCG CCC GAC ACG GAT GAG C
198xR-1F_FPL	CTC GAT CTG ACC TTC TCG ATC GGC GAC TCC GGT CTG CGG GAG TCC ATG AGC TC
198xR-1F_RPL	GTC GCC GAT CGA GAA GGT CAG ATC GAG GGC GCG CCC GAC ACG GAT GAG C
198xR-1E_FPL	CTC GAT CTG ACC GAA TCG ATC GGC GAC TCC GGT CTG CGG GAG TCC ATG AGC TC
198xR-1E_RPL	GTC GCC GAT CGA TTC GGT CAG ATC GAG GGC GCG CCC GAC ACG GAT GAG C
198xT-2A_FPL	CTC GAT CTG GCC CGT TCG ATC GGC GAC TCC GGT CTG CGG GAG TCC ATG AGC TC
198xT-2A_RPL	GTC GCC GAT CGA ACG GGC CAG ATC GAG GGC GCG CCC GAC ACG GAT GAG C
FP_Rhothi_Leader_SLIM	GGA GTA TTG CCC ATC GGC AAT GAG TTC ATG GGC CAC G
RP_Rhothi_Leader_SLIM	GAG CTC GAT GAC ATC GTC CTG CGC GTT GCT G
FPTail_Rhothi_S-8G_SLIM	GGC GTC GCC AGT GTC GAG ACC AAA GGA GTA TTG CCC ATC GGC AAT GAG TTC ATG GGC CAC G
RPTail_Rhothi_S-8G_SLIM	TTT GGT CTC GAC ACT GGC GAC GCC GAG CTC GAT GAC ATC GTC CTG CGC GTT GCT G

Introduced restriction enzyme sites are underlined; introduced ribosomal binding sites are shown in italics; introduced mutations are shown in bold

Supporting table 2 – M9 Vitamin Mix

component	amount
biotin	0.2 g
choline chloride	1.0 g
disodium adenosine 5'-triphosphate	0.3 g
folic acid	1.0 g
myo-inositol	2.0 g
nicotinamide	1.0 g
panthothenic acid	1.0 g
pyridoxal hydrochloride	1.0 g
riboflavin	0.1 g
thiamine	1.0 g
H ₂ O	ad 300 mL

10 M NaOH were added to the resulting suspension until everything was dissolved. The solution was then sterile filtered and stored at 4°C

Supporting Table 3 – Tail sequences of the caulonodins with possible plug residues highlighted

peptide	14	15	16	17	18	19
caulonodin IV	Q	Y	Y	F	A	-
caulonodin V	Q	T	Y	W	P	-
caulonodin VI	K	R	Y	D	P	A
caulonodin VII	R	R	W	D	-	-

Positions picked for mutation in bold black, most likely plugs in red

Supporting Table 4 – Production amounts of all constructed caulonodin and rhodanodin mutants; mutant numbering according to figure 1a, positive numbers for mutations in the core peptide, negative numbers for mutations in the leader peptide.

lasso peptide mutant	relative production to corresponding wild type in %	heat stable
caulonodin IV WT	100 ± 15	no
Y15A	49 ± 2	no
Y15W	167 ± 12	no
Y16A	14.3 ± 0.7	no
Y16W	84 ± 6	no
F17A	not detected	-
F17W	132 ± 9	yes
caulonodin V WT	100 ± 23	no
S1G	4.0 ± 0.3	-
S1A	132.2 ± 1.6	-
S1C	detected	-
S1V	detected	-
S1T	detected	-
S1F	detected	-
T15A	2.7 ± 0.1	-
Y16A	detected	-
Y16F	detected	-
W17A	detected	-
W17F	detected	-
W17Y	8.5 ± 0.6	-
Del1	7.8 ± 0.5	-
Del2	detected	-
Del3	detected	-
Del12	not detected	-
Del123	not detected	-
L-12R_I-11R_V-9R_A-6R	not detected	-
L-12E_I-11E_V-9E_A-6E	not detected	-
L-12R_I-11R	0.3 ± 0.1	-
L-12E_I-11E	0.4 ± 0.1	-
L-12Q_I-11Q	7.6 ± 0.1	-
V-9R_A-6R	not detected	-
V-9E_A-6E	not detected	-
V-9Q_A-6Q	not detected	-
R-10A	97.8 ± 1.8	-
R-10F	71.2 ± 1.5	-
R-10E	63.0 ± 1.4	-
V-9E	2.0 ± 0.1	-
V-9R	3.6 ± 0.2	-
G-8A	detected	-

lasso peptide mutant	relative production to corresponding wild type in %	heat stable
G-8F	6.0 ± 0.5	-
G-8E	4.7 ± 0.1	-
G-8V	0.5 ± 0.1	-
G-8R	not detected	-
G-8P	0.3 ± 0.1	-
A-6E	detected	-
A-6R	detected	-
T-2A	detected	-
R-1A	80 ± 15	-
R-1F	1.0 ± 0.1	-
R-1E	33 ± 14	-
R-1Q	84 ± 3	-
R-1N	62 ± 12	-
R-1K	90 ± 8	-
R-1H	15.7 ± 1.0	-
caulonodin VI WT	100 ± 7	no
A1G	7.1 ± 0.5	-
A1S	122 ± 30	-
A1C	detected	-
A1V	detected	-
A1T	1.3 ± 0.1	-
A1F	detected	-
K14A	38 ± 6	no
K14W	22.9 ± 1.4	no
R15A	6.1 ± 0.5	no
R15W	2.0 ± 0.1	yes
Y16A	detected	-
Y16W	99 ± 14	no
caulonodin VII WT	100 ± 29	no
R14A	29.7 ± 0.6	no
R15A	0.8 ± 0.1	no
R15K	123 ± 5	no
R15W	detected	yes
W16A	not detected	-
W16F	detected	-
W16Y	73 ± 12	no
rhodanodin WT	100	-
S-8G	1460	-

Supporting table 5 – Chemical shift Assignment of ¹H signals (ppm) of caulonodin V in H₂O/D₂O (9:1) at 298 K

aa	NH	αH	βH	others
Ser1	7.075	4.560	4.190; 3.995	/
Ile2	9.084	4.891	2.509	γCH ₃ : 1.159 γCH ₂ : 1.478; 1.342 δCH ₃ : 1.151
Gly3	8.702	4.528; 3.664	/	/
Asp4	8.973	5.155	3.082	/
Ser5	7.600	3.240	3.821	/
Gly6	8.185	4.651; 3.701	/	/
Leu7	9.406	4.841	2.089	γCH: 2.044 δCH ₃ : 1.244; 1.172
Arg8	8.872	4.007	1.863; 1.737	γCH ₂ : 1.700; 1.513 δCH ₂ : 3.170; 3.132 εNH: 7.140
Glu9	7.989	3.386	1.485; 1.327	γCH ₂ : 1.225
Ser10	7.854	4.691	4.200; 4.008	/
Met11	8.900	4.574	2.304	γCH ₂ : 2.894; 2.763 εCH ₃ : 2.290
Ser12	8.305	4.730	4.078; 3.969	/
Ser13	7.671	4.653	4.013; 3.871	/
Gln14	8.921	4.488	2.039	γCH ₂ : 2.494 εNH ₂ : 7.649; 6.992
Thr15	7.468	4.643	4.204	γCH ₃ : 1.221
Tyr16	8.058	5.136	3.033; 2.165	2. 6H: 6.787 3. 5H: 6.787
Trp17	9.350	5.488	3.731; 3.293	2H: 7.176; 4H: 7.941 5H: 7.278; 6H: 7.434 7H: 7.779; NH: 10.401
Pro18	/	4.774	2.516; 2.355	γCH ₂ : 2.267; 2.468 δCH ₂ : 4.483; 4.351

Supporting table 6 – Structural statistics for the family of 15 structures selected to represent the structure of caulonodin V in H₂O/D₂O (9:1)

Restraining Constraints	Constraints Violations
Total: 166	Distance violations, >0.5 Å: 0
distance, i=j: 79	RMS deviations: 0.02 Å
distance, i-j =1: 35	Dihedral violations, > 5°: 0
distance, i-j >1: 22	RMS deviation: 1.8°
dihedral: 22	Average pairwise RMS deviation (Ile ² -Trp ¹⁷)
hydrogen bond: 0	Backbone atoms: 0.06 Å
Constraints/residue: 9.2	All heavy atoms: 0.45 Å

Supporting table 7 – Chemical shift Assignment of ^1H signals (ppm) of caulonodin VI in $\text{H}_2\text{O}/\text{D}_2\text{O}$ (9:1) at 288 K

aa	NH	αH	βH	others
Ala1	7.654	4.426	1.617	/
Gly2	8.608	4.576; 4.602		
Thr3	7.695	4.623	4.437	γCH_3 : 1.358
Gly4	7.700	3.844; 4.524		
Val5	7.960	4.829	2.516	γCH_3 : 0.964; 1.008
Leu6	8.027	4.289	amb.*	γCH_2 : amb. δCH_3 : 1.038; 1.078
Leu7	8.962	5.131	amb.	γCH_2 amb. δCH_3 : 0.911; 1.096
Pro8	/	4.776	amb.	Amb.
Glu9	8.790	4.586	2.242	γCH_2 : 2.275; 2.424
Thr10	8.659	4.293	4.146	γCH_3 : 1.444
Asn11	amb.	amb.	amb.	amb.
Gln12	8.252	4.384	2.340; 2.547	γCH_2 : 2.505; 2.575 ϵNH_2 : amb.
Ile13	8.412	4.494	amb.	γCH_2 : amb. γCH_3 : 1.120; δCH_3 : 1.056
Lys14	7.989	4.756	amb.	γCH_2 : amb.; δCH_2 : amb. ϵCH_2 : 3.162; ϵNH_3^+ : 7.825
Arg15	8.521	5.593	amb.	γCH_2 : amb. δCH_2 : 3.077; 3.479 NH: 7.143
Tyr16	9.772	4.727	3.135; 3.183	2, 6H: 7.027 3, 5H: 6.825
Asp17	8.959	5.363	amb.	
Pro18	/	4.693	amb.	amb.
Ala19	8.825	4.532	1.644	

*amb. = ambiguous