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Electronic Supporting Information

for

Azidophenyl as a click-transformable redox label of DNA suitable for electrochemical footprinting of DNA-protein interactions

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1 Single incorporation of functionalized dNTPs



Figure S1. PEX single incorporations of a $dN^{A}TP$ into 19-nt DNA using temp^{*C*} or temp^{*A*} template and PWO DNA polymerase.



Figure S2. PEX single incorporations of a $dN^{TP}TP$ and $dN^{TNO2}TP$ into 19-nt DNA using temp^{*C*} or temp^{*A*} template and Pwo DNA polymerase.

2 Kinetics study



Figure S3. Kinetics of PEX using $dC^{A}TP$ in comparison with natural dCTP (+). Time intervals are given in minutes.



Figure S4. Kinetics of PEX using $dA^{A}TP$ in comparison with natural dATP (+). Time intervals are given in minutes.

3 Multiple incorporations of functionalized dNTPs



Figure S5. PEX incorporations of a $dN^{A}TP$ into 31-nt DNA using temp^{*rnd16*} template, KOD XL and PWO DNA polymerase.



Figure S6. PEX incorporations of a $dN^{A}TP$ into 31-nt DNA using temp^{*rnd16*} template, Vent (*exo-*) DNA polymerase.



Figure S7. PEX incorporations of a $dN^{A}TP$ into 50-nt DNA using template temp^{2CON4} and KOD XL DNA polymerase.

4 Study of DNA-protein interaction



Figure S8. Native PAGE analysis of 50-mer DNA^{2CON4}_p53CD_GST complex. Lane 1: natural DNA; 2: 0.4 equiv.; 3: 0.7 equiv.; 4: 1.2 equiv.; 5: 1.8 equiv. of protein p53CD_GST to DNA; Lane 6: DNA^A; 7: 0.4 equiv.; 8: 0.7 equiv.; 9: 1.2 equiv.; 10: 1.8 equiv. of protein p53CD_GST to DNA.



Figure S9. Native PAGE analysis of stability of DNA^{2CON4}_p53CD_GST complex after click reaction. Lane 1: DNA^A; lane 2: protein/DNAcomplex; lane 3: protein/DNAcomplex, 0.5 mM 4-nitrophenylacetylene, 5 μ M CuBr; 25 μ M TBTA ligand, 65 μ M Na ascorbate, 20 °C, 1h.

5 Thermal stability of complex DNA-protein



Figure S10. Native PAGE analysis of thermal stability of DNA^{1a2G}_p53CD_GST complex. Lane 1: DNA^A; lanes 2-5: 1.2 equiv. of protein p53CD_GST to DNA. Conditions: DNA and proteins were mixed together in binding buffer and created protein/DNA complexes were incubated at mentioned temperatures for 1h.



Figure S11. Native PAGE analysis of thermal stability of DNA^{2CON4}_p53CD_GST **complex.** Lane 1: DNA^A; lanes 2-5: 1.2 equiv. of protein p53CD_GST to DNA. **Conditions:** DNA and proteins were mixed together in binding buffer and created protein/DNA complexes were incubated at mentioned temperatures for 1h.

6 Cu^I concentration dependence of stability of complex DNA-protein



Figure S12. Native PAGE analysis of Cu^I concentration dependence of stability of DNA^{1a2G}_p53CD_GST complex. Lane 1: DNA^A; lanes 2-9: 1.2 equiv. of protein p53CD_GST to DNA; lane 2: protein/DNAcomplex; lane 3: protein/DNAcomplex, 20°C; lane 4: protein/DNAcomplex, 5 μ M CuBr, 20°C; lane 5: protein/DNAcomplex, 5 μ M CuBr, 25 μ M TBTA ligand, 20°C; lane 6: protein/DNAcomplex, 10 μ M CuBr, 20°C; lane 7: protein/DNAcomplex, 10 μ M CuBr; 50 μ M TBTA ligand, 20°C; lane 8: protein/DNAcomplex, 20 μ M CuBr, 20°C; lane 9: protein/DNAcomplex, 20 μ M CuBr; 100 μ M TBTA ligand, 20°C. Conditions: DNA and proteins were mixed together in binding buffer and created protein/DNAcomplexes were incubated with various concentration of CuBr in/without presence of the TBTA ligand at 20°C for 1h.



Figure S13. Native PAGE analysis of Cu^I concentration dependence of stability of DNA^{2CON4}_p53CD_GST complex. Lane 1: DNA^A; lanes 2-9: 1.2 equiv. of protein p53CD_GST to DNA; lane 2: protein/DNAcomplex; lane 3: protein/DNAcomplex, 20°C; lane 4: protein/DNAcomplex, 5 μ M CuBr, 20°C; lane 5: protein/DNAcomplex, 5 μ M CuBr, 25 μ M TBTA ligand, 20°C; lane 6: protein/DNAcomplex, 10 μ M CuBr, 20°C; lane 7: protein/DNAcomplex, 10 μ M CuBr; 50 μ M TBTA ligand, 20°C; lane 8: protein/DNAcomplex, 20 μ M CuBr; 20°C; lane 9: protein/DNAcomplex, 20 μ M CuBr; 100 μ M TBTA ligand, 20°C. Conditions: DNA and proteins were mixed together in binding buffer and created protein/DNAcomplexes were incubated with various concentration of CuBr in/without presence of the TBTA ligand at 20°C for 1h.

7 Electrochemistry



Figure S14. CV responses at HMDE of PEX products synthesized with temp^{*rnd16*} template and dNTP mixes containing $dA^{A}TP$ (as specified in legend) complemented with three natural dNTPs and PEX products after click reaction with (nitro)phenyltriazole. CA – peak due to reduction of cytosine and adenine, G – peak due to guanine, N₃^{red} – azide reduction, NO₂^{red} – nitrogroup reduction.



Figure S15. CV responses at HMDE of PEX products synthesized with temp^{*rnd16*} template and dNTP mixes containing $dC^{A}TP$ (as specified in legend) complemented with three natural dNTPs and PEX products after click reaction with (nitro)phenyltriazole.



Figure S16. CV responses at HMDE of PEX products synthesized with temp^{*rnd16*} template and dNTP mixes containing $dA^{TX}TP$ (as specified in legend) complemented with three natural dNTPs.



Figure S17. CV responses at HMDE of PEX products synthesized with temp^{*rnd16*} template and dNTP mixes containing $dC^{TX}TP$ (as specified in legend) complemented with three natural dNTPs.



Figure S18. CV responses at HMDE of PEX products synthesized with temp^{la2G} template and **dA^ATP** complemented with three natural dNTPs (red curve) and PEX products after click reaction with nitrophenylacetylene (green curve), DNA-p53 complex after click reaction followed by denaturation (violet curve), control with BSA (black curve).



Figure S19. CV responses at HMDE of PEX products synthesized with temp^{2CON4} template and **dA^ATP** complemented with three natural dNTPs (red curve) and PEX products after click reaction with nitrophenylacetylene (green curve), DNA-protein complex after click reaction followed by denaturation (violet curve), control with BSA (black curve).



Figure S20. Detail of CV responses at HMDE of PEX products synthesized with $temp^{2CON4}$ template and $dA^{A}TP$ complemented with three natural dNTPs (red curve) and PEX products after click reaction with nitrophenylacetylene (green curve), DNA-protein complex after click reaction followed by denaturation (violet curve), the control with BSA (black curve). For full CV scans see Fig. S19.

8 Selected copies of NMR spectra

¹H NMR and ¹³C spectra of dC^A .



¹H NMR and ¹³C spectra of dA^A .



¹H NMR and ¹³C spectra of dC^{TP} .



¹H NMR and ¹³C spectra of dA^{TP} .



¹H NMR and ¹³C spectra of dC^{TNO2} .















¹H NMR, ¹³C and ³¹P spectra of $dC^{TP}TP$.









¹H NMR, ¹³C and ³¹P spectra of $dC^{TNO2}TP$.











9 Copies of Maldi-TOF spectra of DNA after click reaction

Figure S21. MALDI-TOF MS spectrum of temp^{*rnd16*} with A^{TP} modification (31 nt).



Figure S22. MALDI-TOF MS spectrum of temp^{*rnd16*} with A^{TNO2} modification (31 nt)



Figure S23. MALDI-TOF MS spectrum of temp^{rnd16} with C^{TP} modification (31 nt).



Figure S24. MALDI-TOF MS spectrum of temp^{rnd16} with C^{TNO2} modification (31 nt).



45000-M [Da] ON^{CTNO2} 40000 10674.3 9420.0 35000 30000-25000 9438.5-9458.0 20000-15000-9293.0 9497 10000-9169.6 9709.3 9980.5 5000 10627.9 7000 8500 12000 m/z 6500 7500 8000 10500 6000 9000 9500 10000 11000 11500

Figure S26. MALDI-TOF MS spectrum of temp^{rnd16} with C^{TNO2} modification (31 nt).

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Figure S27. MALDI-TOF MS spectrum of temp^{*rnd16*} with A^A modification (31 nt).



Figure S28. MALDI-TOF MS spectrum of temp^{*rnd16*} with C^A modification (31 nt).