

Electronic Supplementary Information (ESI)

Novel *ortho*-OPE Metallofoldamers: Binding-Induced Folding Promoted by Nucleating Ag(I)-Alkyne Interactions

Ana Martín-Lasanta,^a Luis Álvarez de Cienfuegos,^a Alice Johnson,^b Delia Miguel,^a Antonio J. Mota,^c Angel Orte,^d María Jose Ruedas-Rama,^d María Ribagorda,^e Diego J. Cárdenas,^e M. Carmen Carreño,^e Antonio M. Echavarren^b and Juan M. Cuerva*,^a

^a Departamento de Química Orgánica, ^cInorgánica y ^dFísicoquímica, Universidad de Granada, Granada (Spain).

^b Institute of Chemical Research of Catalonia (ICIQ), Tarragona, Spain.

^e Departamento de Química y Química Orgánica, Universidad Autónoma de Madrid, Madrid (Spain).

Contents

SYNTHESIS AND CHARACTERIZATION

General details	2
General synthetic procedures	3
Synthesis and characterization of the new <i>o</i> -OPEs 6 and 12-16	4
Spectroscopical characterization of the Ag(I)-complexes	7

¹H-NMR TITRATIONS AND CONSTANT DETERMINATION

Additional supporting information figures S1-S3	14
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CRYSTAL STRUCTURE DETERMINATION

18

THEORETICAL STUDIES

21

PHOTOPHYSICAL STUDIES

Absorption and emission studies (Figures S4-S6)	27
Fluorescence quantum yield determinations	28
Time-resolved fluorescence measurements. Photokinetical study (Figures S7 and S8)	28
Estimation of the excited-state folding kinetic rate	30
CD titrations	30

¹ H- and ¹³ C-NMR spectra	31
Additional supporting information figures S10-S11	54

SYNTHESIS AND CHARACTERIZATION

General details

The following palladium catalysts, *trans*-dichlorobis(triphenylphosphine)palladium(II) ($\text{Pd}(\text{PPh}_3)_2\text{Cl}_2$) and *trans*-dichlorobis(acetonitrile)palladium(II) ($\text{Pd}(\text{CH}_3\text{CN})_2\text{Cl}_2$), were prepared from palladium(II) chloride (PdCl_2) according to previously described procedures.¹ Palladium(II) chloride (PdCl_2), copper(I) iodide (CuI), trimethylsilyl acetylene (TMSA), triethylamine (Et_3N), *N,N*-diisopropylamine (Pr_2NH), tri-tert-butylphosphonium tetrafluoroborate ($\text{P}^t\text{Bu}_3\text{HBF}_4$), silver tetrafluoroborate (AgBF_4) and all other reagents were used as purchased from standard chemical suppliers and used without further purification. TLC was performed on aluminium-backed plates coated with silica gel 60 (230-240 mesh) with F_{254} indicator. The spots were visualized with UV light (254 nm). All column chromatography were performed with silica gel 60 (35-70 μm) from VWR. The following known compounds were isolated as a pure samples and showed identical NMR spectra to the reported compound: **I**, **II**,² **III**,² **IV**,³ **V**,⁴ **1**,⁵ **2**,⁵ **3**,⁵ **4**,⁵ **5**,⁵ **7**,⁵ **8**,⁶ **9**,⁶ **10**,⁶ and **11**.⁶

NMR Spectra were measured at room temperature. ^1H NMR spectra were recorded at 300, 400 or 500 MHz. Chemical shifts are reported in ppm using residual solvent peak as reference. Data are reported as follows: chemical shift, multiplicity (s: singlet, d: doublet, t: triplet, q: quartet, quint: quintuplet; m: multiplet, dd: doublet of doublets, dt: doublet of triplets, dq: doublet of quartets, td: triplet of doublets, bs: broad singlet), coupling constant (J in Hz) and integration. ^{13}C NMR spectra were recorded at 75, 100 or 126 MHz using broadband proton decoupling and chemical shifts are reported in ppm using residual solvent peaks as reference. Carbon multiplicities were assigned by DEPT techniques.

High resolution mass spectra (HRMS) were recorded on a mass spectrometer using EI at 70eV, ESI and MALDI-TOF.

Absorption spectra were recorded at 25 °C with a Perkin-Elmer Lambda 650 UV/vis spectrophotometer equipped with a Peltier temperature-controlled cell holder. Steady-state fluorescence emission spectra were collected on a JASCO FP-6500 spectrofluorometer equipped with a 450 W xenon lamp for excitation, with temperature controller ETC-273T at 25 °C. All experiments were performed using 5×10-mm quartz cuvettes. Fluorescence decay traces were recorded in the single photon timing (SPT) mode using a FluoTime 200 fluorometer (PicoQuant, GmbH).

Single crystal X-ray diffraction data were recorded on a Bruker Kappa APEX II DUO diffractometer equipped with an APPEX 2 4K CCD area detector, a Microsource with Moka radiation and an Oxford Cryostream 700 low temperature device ($T = -173^\circ\text{C}$). Full-sphere data collection was used with w and j scans. Programs used: Data collection Apex2 V2009 1.0 (Bruker-Nonius 2008), data reduction Saint + Version 7.60A (Bruker AXS 2008) and absorption correction SADABS V. 2008-1 (2008). Structure solution and refinement were carried out using SIR2011 and SHELXTL V6.14, respectively. All-non hydrogen atoms were refined including anisotropic displacement parameters.

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General synthetic procedures

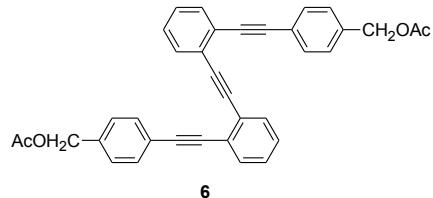
Sonogashira coupling of aryl iodides. (GP1)

A solution of the terminal alkyne (1.2 mmol) dissolved in the minimum volume of THF and with 2 mL of Et₃N was added dropwise to a carefully degassed solution of Pd(PPh₃)₂Cl₂ (5 mol%), CuI (10 mol%) and the aryl iodide (1 mmol) in 10 mL of Et₃N. The reaction was stirred between 3-16 h at room temperature under argon atmosphere. The mixture was then diluted with EtOAc or dichloromethane, washed with saturated aq NH₄Cl solution, dried over anhydrous Na₂SO₄ and the solvent was removed under reduced pressure. The residue was purified by flash chromatography (EtOAc/Hexane mixtures) to give the corresponding coupling product.

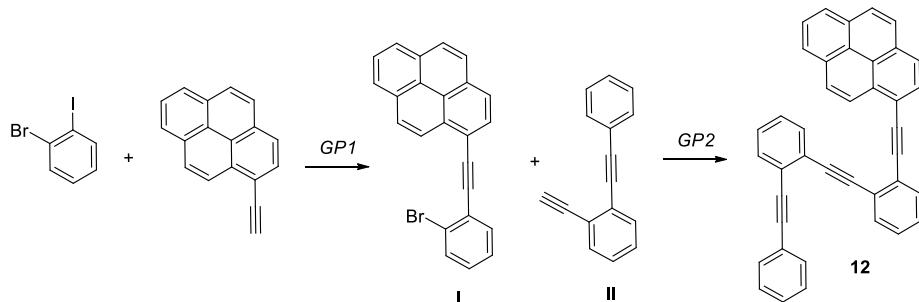
Sonogashira coupling of aryl bromides. (GP2)

A solution of the terminal alkyne (1.2 mmol) dissolved in the minimum volume of THF and with 2 mL of ⁱPr₂NH was added dropwise to a carefully degassed solution of Pd(CH₃CN)₂Cl₂ (3 mol%), P^tBu₃·HBF₄ (6 mol%), CuI (3 mol%) and the aryl bromide (1 mmol) in 10 mL of ⁱPr₂NH. The reaction was stirred between 3-16 h at room temperature under argon atmosphere. The mixture was then diluted with EtOAc or dichloromethane, washed with saturated aq NH₄Cl solution, dried over anhydrous Na₂SO₄ and the solvent was removed under reduced pressure. The residue was purified by flash chromatography (EtOAc/Hexane mixtures) to give the corresponding coupling product.

Synthesis and characterization of the new *o*-OPEs 6 and 12-16

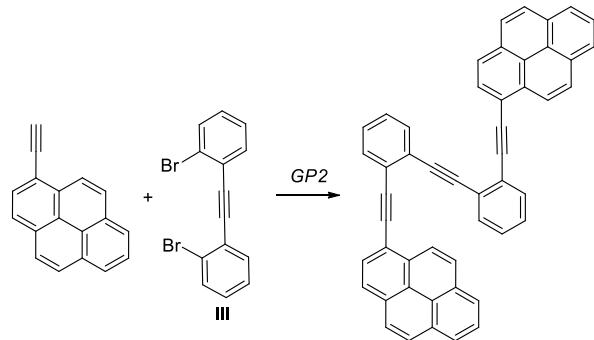


Compounds 6. A drop of acetic anhydride was added to a solution of **4** (60 mg, 0.14 mmol) and DMAP (17 mg, 0.14 mmol) in dry CH_2Cl_2 (10^{-3} M) and the reaction was stirring at room temperature for 1 hour. The solvent was then removed under reduced pressure and the residue was purified by flash chromatography (EtOAc/Hexane mixtures) to give **6** 49 % yield (36 mg) as white solid. ^1H NMR (600 MHz, CDCl_3) δ 7.60 – 7.56 (m, 4H), 7.47 (d, J = 8.2 Hz, 4H), 7.37–7.33 (m, 4H), 7.25 (d, J = 8.2 Hz, 4H), 5.03 (s, 4H), 2.05 (s, 6H). ^{13}C NMR (151 MHz, CDCl_3 ; DEPT) δ 171.0 (C), 137.2 (C), 132.6 (CH), 132.4 (CH), 132.2 (CH), 129.0 (CH), 128.8 (CH), 128.5 (CH), 126.1 (C), 126.0 (C), 123.3 (C), 93.8 (C), 92.6 (C), 88.9 (C), 66.1 (CH₂), 21.2 (CH₃). EI-HRMS calculated for $\text{C}_{36}\text{H}_{26}\text{O}_4$: 522.1831, found: 522.1834.

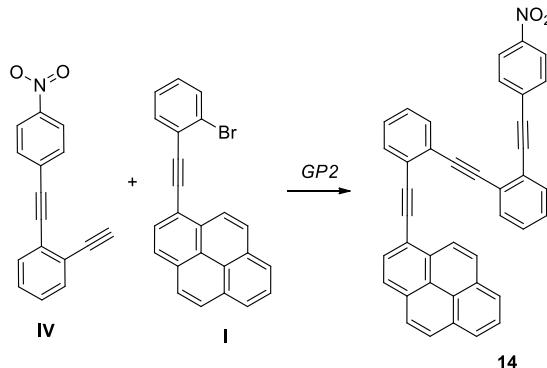


Compound I was prepared from 1-bromo-2-iodobenzene (200 mg, 0.71 mmol) and 1-ethynyl-pyrene (160 mg, 0.7 mmol) according to previously described GP1 to give **III** in 36 % yield (97 mg) as a bright yellow solid. ^1H NMR (300 MHz, CDCl_3) δ 8.84 (d, J = 9.1 Hz, 1H), 8.30-8.15 (m, 4H), 8.13-7.83 (m, 4H), 7.81-7.70 (m, 2H), 7.39 (t, J = 7.5 Hz, 1H), 7.26 (t, J = 7.7 Hz, 1H). ^{13}C NMR (75 MHz, CDCl_3) δ 133.5 (CH), 132.7 (CH), 132.2 (C), 131.6 (C), 131.3 (C), 131.1 (C), 129.8 (CH), 129.5 (CH), 128.6 (CH), 128.4 (CH), 127.31 (CH), 127.29 (CH), 126.3 (CH), 125.9 (C), 125.8 ($\text{CHx}2$), 125.7 (CH), 125.7 (C), 124.6 (CH), 124.5 (C), 124.3 (C), 117.4 (C), 93.7 (C), 93.5 (C). EI-HRMS calculated for $\text{C}_{24}\text{H}_{13}\text{Br}$: 380.0200, found: 380.0201.

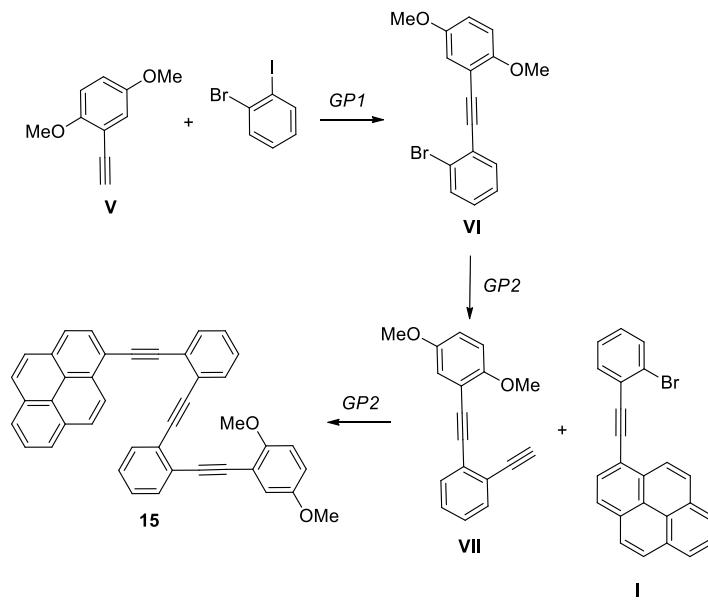
Compound 12 was prepared from intermediate **I** (200 mg, 0.71 mmol) and 1-ethynyl-2-(phenylethyynyl)benzene **II** (160 mg, 0.7 mmol) according to previously described GP2 to give **12** in 82 % yield brsm (39 mg) as a bright yellow solid. ¹H NMR (500 MHz, CDCl₃) δ 8.82 (d, *J* = 9.0 Hz, 1H), 8.21 (d, *J* = 7.9 Hz, 1H), 8.17 (d, *J* = 7.5 Hz, 1H), 8.11 (d, *J* = 7.5 Hz, 1H), 8.07 (d, *J* = 8.4 Hz, 2H), 8.04 – 7.97 (m, 2H), 7.81 – 7.75 (m, 2H), 7.72 (dd, *J* = 7.6, 1.4 Hz, 1H), 7.68 (d, *J* = 7.6 Hz, 1H), 7.63 (dd, *J* = 7.7, 1.1 Hz, 1H), 7.51 – 7.45 (m, 2H), 7.45 – 7.34 (m, 3H), 7.29 (td, *J* = 7.7, 1.4 Hz, 1H), 7.23 – 7.19 (m, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 132.6 (CH), 132.5 (CH), 132.2 (CH), 132.0 (CH), 131.8 (CHx2), 131.4 (C), 131.3 (C), 131.2 (C), 129.9 (CH), 128.5 (CH), 128.42 (CH), 128.40 (CH), 128.36 (CH), 128.1 (CH), 128.0 (CH), 127.3 (CH), 126.4 (C), 126.22 (CH), 126.20 (C), 126.1 (CH), 125.9 (C), 125.8 (C), 125.7 (CH), 125.6 (CH), 124.5 (CH), 124.5 (C), 124.3 (C), 123.3 (C), 117.9 (C), 94.1 (Cx2), 93.2 (C), 92.9 (C), 92.6 (C), 88.5 (C).



Compound 13 was prepared from **III** (30 mg, 0.08 mmol) and 1-ethynyl-pyrene (50 mg, 0.22 mmol) according to previously described GP2 to give **13** in 53 % yield (27 mg) as a bright yellow solid. ^1H NMR (500 MHz, CDCl_3) δ 8.57 (d, $J = 9.0$ Hz, 2H), 8.10 (d, $J = 7.0$ Hz, 2H), 8.03 (d, $J = 7.1$ Hz, 2H), 7.97 (d, $J = 7.7$ Hz, 2H), 7.96 (t, $J = 7.2$ Hz, 2H), 7.90 (d, $J = 8.9$ Hz, 2H), 7.83 (d, $J = 7.6$ Hz, 2H), 7.80–7.76 (m, 6H), 7.56 (d, $J = 9.1$ Hz, 2H), 7.47 (td, $J = 7.6, 1.2$ Hz, 2H), 7.38 (td, $J = 7.6, 1.2$ Hz, 2H); ^{13}C NMR (126 MHz, CDCl_3) δ 132.9 (CH), 132.2 (CH), 131.9 (C), 131.2 (Cx2), 131.1 (C), 131.0 (C), 129.8 (CH), 129.0 (C), 128.5 (CH), 128.1 (CHx2), 128.0 (CH), 127.1 (CH), 126.7 (C), 126.1 (CH), 125.9 (CH), 125.8 (C), 125.6 (CH), 125.4 (CH), 124.2 (CH), 124.1 (C), 117.7 (C), 94.1 (C), 93.5 (C), 93.0 (C); MALDI-HRMS calculated for $\text{C}_{50}\text{H}_{26}$: 626.2048, found: 626.2029.

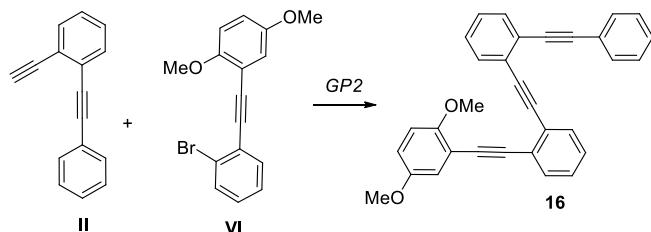


Compound 14 was prepared from **IV** (100 mg, 0.4 mmol) and **I** (111 mg, 0.29 mmol) according to previously described GP2 to give **14** in 57 % yield (91 mg) as a pale yellow solid. ^1H NMR (400 MHz, CDCl_3) δ 8.69 (d, $J = 9.1$ Hz, 1H), 8.19 (d, $J = 7.4$ Hz, 1H), 8.09 (dd, $J = 8.0, 4.4$ Hz, 3H), 8.04 – 7.97 (m, 3H), 7.79 (d, $J = 7.5$ Hz, 1H), 7.76 – 7.67 (m, 5H), 7.61 – 7.55 (m, 1H), 7.51 – 7.36 (m, 4H), 7.24 (d, $J = 8.4$ Hz, 2H); ^{13}C NMR (126 MHz, CDCl_3) δ 146.4 (C), 132.8 (CH), 132.5 (CH), 132.5 (CH), 132.3 (CH), 132.3 (CH), 132.1 (C), 132.0 (CH), 131.4 (C), 131.2 (C), 131.1 (C), 130.0 (C), 129.8 (CH), 129.0 (C), 128.7 (CH), 128.5 (CH), 128.4 (CH), 128.3 (CH), 128.3 (CH), 127.3 (CH), 126.4 (CH), 126.2 (C), 126.1 (CH), 125.8 (C), 125.7 (CH), 125.5 (CH), 125.2 (C), 124.4 (CH), 124.3 (C), 124.2 (C), 123.7 (CH), 123.0 (CH), 117.7 (C), 94.0 (C), 93.5 (C), 93.3 (C), 93.2 (C), 92.1 (C), 92.0 (C). ESIpos-HRMS calculated for $\text{C}_{42}\text{H}_{26}\text{O}_2\text{Na} [\text{M}+\text{Na}]^+$: 585.1847, found: 585.1825.



Compound VI was prepared from **V** (263 mg, 1.62 mmol) and 1-bromo-2-iodobenzene (458 mg, 1.62 mmol) according to previously described GP1 to give **VI** in 45 % yield (230 mg) as a transparent liquid. ^1H NMR (500 MHz, CDCl_3) δ 7.61 (dd, $J = 7.9, 1.4$ Hz, 1H), 7.60 (dd, $J = 7.9, 1.4$ Hz, 1H), 7.28 (td, $J = 7.6, 1.4$ Hz, 1H), 7.17 (td, $J = 7.8, 1.7$ Hz, 1H), 7.09 (d, $J = 3.1$ Hz, 1H), 6.89 (dd, $J = 9.0, 3.0$ Hz, 1H), 6.84 (d, $J = 9.0$ Hz, 1H), 3.89 (s, 3H), 3.79 (s, 3H); ^{13}C NMR (126 MHz, CDCl_3) δ 154.8 (C), 153.4 (C), 133.5 (CH), 132.5 (CH), 129.4 (CH), 127.1 (CH), 125.73 (C), 125.70 (C), 118.3 (CH), 116.3 (CH), 112.9 (C), 112.5 (CH), 92.0 (C), 90.5 (C), 56.8 (CH₃), 55.6 (CH₃); EI-HRMS calculated for $\text{C}_{16}\text{H}_{14}\text{BrO}_2 [\text{M}+\text{H}]^+$: 317.0165, found: 317.0177.

Compound 15. **VII** was prepared from **VI** (260 mg, 0.82 mmol) and TMSA (0.2 mL, 1.23 mmol) according to previously described GP2 in 45 % yield (230 mg) as a transparent liquid. This intermediate **VII** (23 mg, 0.09 mmol) was cross-coupled with **I** (32 mg, 0.09 mmol) according to previously described GP2 to give **16** in 51 % yield (25 mg) as a pale yellow solid. ¹H NMR (500 MHz, CDCl₃) δ 8.80 (d, *J* = 9.0 Hz, 1H), 8.20 – 8.17 (m, 2H), 8.12 – 8.06 (m, 3H), 8.02 (d, *J* = 8.9 Hz, 1H), 7.99 (t, *J* = 7.6 Hz, 1H), 7.76 – 7.73 (m, 3H), 7.68 (d, *J* = 7.8, 2H), 7.43–7.38 (m, 2H), 7.34 (td, *J* = 7.5, 1.3 Hz, 1H), 7.29 (td, *J* = 7.6, 1.3 Hz, 1H), 6.95 (d, *J* = 3.0 Hz, 1H), 6.70 (dd, *J* = 9.0, 3.0 Hz, 1H), 6.65 (d, *J* = 9.0 Hz, 1H), 3.65 (s, 3H), 3.55 (s, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 154.5 (C), 153.1 (C), 132.7 (CH), 132.5 (CH), 132.2 (C), 132.1 (CH), 132.0 (CH), 131.4 (C), 131.3 (C), 131.2 (C), 129.9 (CH), 128.5 (CH), 128.4 (CH), 128.3 (CH), 128.3 (CH), 128.0 (CH), 127.9 (CH), 127.4 (CH), 126.7 (C), 126.2 (CH), 126.2 (C), 126.2 (CH), 125.93 (C), 125.90 (C), 125.7 (CH), 125.5 (CH), 124.53 (CH), 124.51 (C), 124.4 (C), 117.9 (C), 117.7 (CH), 116.6 (CH), 112.9 (C), 112.1 (CH), 94.1 (C), 93.3 (C), 92.7 (C), 92.7 (C), 92.4 (C), 90.7 (C), 56.4 (CH₃), 55.6 (CH₃); ESIpos-HRMS calculated for C₄₂H₂₆O₂Na [M+Na⁺]: 585.1847, found: 585.1825.



Compound 16 was prepared from **VI** (60 mg, 0.19 mmol) and 1-ethynyl-2-(phenylethyynyl)benzene **II** (50 mg, 0.25 mmol) according to previously described GP2 to give **16** in 33 % yield (27 mg) as a transparent liquid. ¹H NMR (500 MHz, CDCl₃) δ 7.65 – 7.62 (m, 2H), 7.60 (dd, *J* = 7.5, 1.3 Hz, 1H), 7.58 – 7.55 (m, 1H), 7.53–7.50 (m, 2H), 7.35 – 7.27 (m, 7H), 7.05 (d, *J* = 3.0 Hz, 1H), 6.82 (dd, *J* = 9.0, 3.1 Hz, 1H), 6.77 (d, *J* = 9.0 Hz, 1H), 3.76 (s, 3H), 3.66 (s, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 154.7 (C), 153.3 (C), 132.4 (CH), 132.1 (CH), 132.0 (CH), 131.8 (CHx2), 131.8 (CH), 128.4 (CH), 128.4 (CHx2), 128.24 (CH), 128.22 (CH), 128.00 (CH), 127.98 (CH), 126.2 (C), 126.1 (C), 125.93 (C), 125.90 (C), 123.3 (C), 117.9 (CH), 116.7 (CH), 113.0 (C), 112.3 (CH), 93.9 (C), 92.6 (C), 92.3 (C), 92.3 (C), 90.4 (C), 88.4 (C), 56.6 (CH₃), 55.8 (CH₃); ESIpos-HRMS calculated for C₃₂H₂₂O₂Na [M+Na⁺]: 461.1518, found: 461.1512.

Spectroscopic characterization of the Ag(I)-complexes

Complex 1·Ag(I): ^1H NMR (500 MHz, 9:1 CD₂Cl₂:Acetone-*d*₆) δ 7.93–7.91 (m, 2H), 7.87–7.85 (m, 2H), 7.63–7.57 (m, 4H), 7.45 (d, *J* = 8.4 Hz, 4H), 7.22 (d, *J* = 8.4 Hz, 4H), 5.18 (s, 4H), 3.57 (s, 2H); ^{13}C NMR (126 MHz, 9:1 CD₂Cl₂:Acetone-*d*₆) δ 166.4 (C), 139.3 (C), 135.0 (CH), 134.0 (CH), 133.2 (CH), 131.2 (CH), 131.1 (CH), 129.0 (CH), 124.9 (C), 122.6 (C), 118.6 (C), 96.4 (C), 93.1 (C), 85.8 (C), 66.6 (CH₂), 42.5 (CH₂).

Complex 2·Ag(I): ^1H NMR (600 MHz, 9:1 CD₂Cl₂:Acetone-*d*₆) δ 7.92 (d, *J* = 8.9 Hz, 2H), 7.85 (d, *J* = 8.9 Hz, 2H), 7.64 – 7.56 (m, 4H), 7.44 (d, *J* = 8.2 Hz, 4H), 7.20 (d, *J* = 8.2 Hz, 4H), 5.15 (s, 4H), 2.81 (s, 4H). ^{13}C NMR (126 MHz, 9:1 CD₂Cl₂:Acetone-*d*₆) δ 172.3 (C), 140.0 (C), 134.7 (CH), 133.9 (CH), 133.1 (CH), 131.2 (CH), 131.0 (CH), 128.2 (CH), 124.9 (C), 122.9 (C), 118.5 (C), 96.2 (C), 92.9 (C), 85.7 (C), 65.8 (CH₂), 30.0 (CH₂).

Complex 3·Ag(I): ^1H NMR (400 MHz, 9:1 CD₂Cl₂:Acetone-*d*₆) δ 7.81 (dd, *J* = 6.7, 2.4 Hz, 2H), 7.72 (dd, *J* = 6.6, 2.3 Hz, 2H), 7.51 – 7.47 (m, 2H), 7.45–7.40 (m, 2H), 7.35 (d, *J* = 8.2 Hz, 4H), 7.05 (d, *J* = 8.2 Hz, 4H), 5.29 (s, 2H), 5.23 (d, *J* = 13.4 Hz, 2H), 4.91 (d, *J* = 13.4 Hz, 2H), 1.15 (s, 18H). A good quality ^{13}C NMR spectrum could not be obtained even using long acquisition period owing to the concentration used. Higher concentrations resulted in precipitation of the sample.

Complex 4·Ag(I): ^1H NMR (400 MHz, 9:1 CD₂Cl₂:Acetone-*d*₆) δ 7.78 – 7.75 (m, 2H), 7.70 – 7.67 (m, 2H), 7.47 – 7.44 (m, 2H), 7.34 (d, *J* = 8.0 Hz, 2H), 7.32 (d, *J* = 7.9 Hz, 1H), 7.21 (d, *J* = 7.9 Hz, 1H), 7.16 (s, 1H), 7.11 (t, *J* = 7.7 Hz, 1H), 6.96 (d, *J* = 8.0 Hz, 2H), 5.27 (q, *J* = 8.1 Hz, 2H), 5.04 (s, 2H), 4.91 (d, *J* = 13.4 Hz, 1H), 4.74 (d, *J* = 13.5 Hz, 1H), 1.17 (s, 9H), 1.15 (s, 9H). A good quality ^{13}C NMR spectrum could not be obtained even using long acquisition period owing to the concentration used. Higher concentrations resulted in precipitation of the sample.

Complex 5·Ag(I): ^1H NMR (400 MHz, 9:1 CD₂Cl₂:Acetone-*d*₆) δ 7.75 – 7.71 (m, 2H), 7.67 – 7.63 (m, 2H), 7.46 – 7.41 (m, 4H), 7.40 (d, *J* = 5.3 Hz, 2H), 7.23 (s, 2H), 7.18 (d, *J* = 7.7 Hz, 2H), 7.11 (t, *J* = 7.6 Hz, 2H), 5.24 (s, 2H), 4.95 (d, *J* = 13.0 Hz, 2H), 4.80 (d, *J* = 13.0 Hz, 2H), 1.13 (s, 18H). A good quality ^{13}C NMR spectrum could not be obtained even using long acquisition period owing to the concentration used. Higher concentrations resulted in precipitation of the sample.

Complex 6·Ag(I): ^1H NMR (500 MHz, 9:1 CD₂Cl₂:Acetone-*d*₆) δ 7.88 (dd, *J* = 6.1, 3.0 Hz, 2H), 7.80 (dd, *J* = 6.0, 3.1 Hz, 2H), 7.57–7.53 (m, 4H), 7.44 (d, *J* = 8.1 Hz, 4H), 7.20 (d, *J* = 8.1 Hz, 4H), 5.07 (s, 4H), 2.12 (s, 6H); ^{13}C NMR (126 MHz, 9:1 CD₂Cl₂:Acetone-*d*₆) δ 172.0 (C), 139.5 (C), 134.3 (CH), 133.7 (CH), 132.8 (CH_x2), 131.0 (CH), 130.6 (CH), 128.2 (CH_x2), 125.0 (C), 123.1 (C), 118.8 (C), 95.4 (C), 92.5 (C), 85.5 (C), 65.8 (CH₂), 21.1 (CH₃).

Complex 7·Ag(I): ^1H NMR (400 MHz, 9:1 CD₂Cl₂:Acetone-*d*₆) δ 7.89 (bd, *J* = 9.2 Hz, 2H), 7.83 (bd, *J* = 8.8 Hz, 2H), 7.63 – 7.53 (m, 4H), 7.42 (d, *J* = 8.0 Hz, 4H), 7.21 (d, *J* = 8.1 Hz, 4H), 4.68 (s, 4H); ^{13}C NMR (126 MHz, 9:1 CD₂Cl₂:Acetone-*d*₆) δ 144.7 (C), 134.2 (CH), 133.7 (CH), 132.7 (CH), 130.8 (CH), 130.5 (CH), 127.3 (CH), 125.2 (C), 123.3 (C), 118.4 (C), 95.7 (C), 92.6 (C), 85.4 (C), 64.8 (CH₂).

Complex 8·Ag(I): ^1H NMR (500 MHz, 9:1 CD₂Cl₂:Acetone-*d*₆) δ 7.87 (dd, *J* = 6.0, 3.1 Hz, 2H), 7.80 (dd, *J* = 5.9, 3.2 Hz, 2H), 7.59 – 7.54 (m, 4H), 7.46 – 7.41 (m, 4H), 7.37 (t, *J* = 7.6 Hz, 2H), 7.22 (t, *J* = 7.8 Hz, 4H). ^{13}C NMR (126 MHz, 9:1 CD₂Cl₂:Acetone-*d*₆) δ 134.4 (CH), 133.8 (CH), 132.7 (CH_x2), 131.1 (CH), 131.0 (CH), 130.7 (CH), 129.3 (CH_x2), 125.1 (C), 123.1 (C), 119.2 (C), 96.0 (C), 92.6 (C), 85.3 (C).

Complex 9·Ag(I): ^1H NMR (500 MHz, 9:1 CD₂Cl₂:Acetone-*d*₆) δ 7.94 – 7.88 (m, 2H), 7.71 (d, *J* = 7.8 Hz, 2H), 7.63 – 7.59 (m, 2H), 7.47 – 7.38 (m, 8H), 7.21 (t, *J* = 7.8 Hz, 2H), 7.11 (t, *J* = 7.8 Hz, 4H); ^{13}C NMR (126 MHz, 9:1 CD₂Cl₂:Acetone-*d*₆) δ 134.6 (CH), 133.6 (CH), 133.5 (CH), 132.7 (CH_x2), 131.4 (CH), 131.3 (CH), 130.9 (CH), 130.4 (CH), 129.2 (CH_x2), 125.3 (C), 123.9 (C), 122.4 (C), 120.2 (C), 97.4 (C), 93.6 (C), 91.8 (C), 87.0 (C).

Complex 10·Ag(I): ^1H NMR (500 MHz, 9:1 CD₂Cl₂:Acetone-*d*₆) δ 7.73 (bd, *J* = 7.5 Hz, 2H), 7.71 – 7.67 (m, 2H), 7.48 – 7.38 (m, 12H), 7.30 (td, *J* = 7.7, 1.0 Hz, 2H), 7.11 – 7.01 (m, 8H); ^{13}C NMR (126 MHz, 9:1 CD₂Cl₂:Acetone-*d*₆) δ 134.3 (CH), 134.0 (CH), 132.8 (CH), 132.5 (CH_x2), 132.4 (CH), 131.10 (CH), 131.09 (CH), 130.8 (CH), 130.7 (CH), 130.1 (CH), 129.0 (CH_x2), 125.1 (C), 123.6 (C), 122.9 (C), 122.4 (C), 120.5 (C), 97.6 (C), 94.3 (C), 92.5 (C), 92.0 (C), 87.7 (C).

Complex 11·Ag(I): ^1H NMR (600 MHz, 9:1 CD₂Cl₂:Acetone-*d*₆) δ 7.75 (d, *J* = 7.8 Hz, 2H), 7.61 (d, *J* = 7.7 Hz, 2H), 7.53 (t, *J* = 7.7 Hz, 2H), 7.39 (t, *J* = 7.7 Hz, 2H), 7.33 (t, *J* = 7.6 Hz, 2H), 7.31–7.27 (m, 6H), 7.21 (t, *J* = 7.8 Hz, 2H), 7.21–7.18 (m, 4H), 7.10 (d, *J*

= 7.7 Hz, 2H), 7.02 (t, J = 7.1 Hz, 2H), 6.99 (t, J = 7.2 Hz, 4H); ^{13}C NMR (151 MHz, 9:1 CD₂Cl₂:Acetone-*d*₆) δ 133.9 (CH), 133.7 (CH), 133.4 (CH), 133.1 (CH), 132.35 (CH), 132.30 (CH), 131.0 (CH), 130.9 (CH), 130.7 (CH), 130.4 (CH), 130.0 (CH), 129.4 (CH), 128.6 (CH), 125.7 (C), 123.9 (C), 122.9 (C), 122.8 (C), 122.7 (C), 121.3 (C), 97.6 (C), 95.3 (C), 92.9 (C), 92.8 (C), 92.1 (C), 88.1 (C).

Complex 12·Ag(I): ^1H NMR (500 MHz, 9:1 CD₂Cl₂:Acetone-*d*₆) δ 8.48 (d, J = 9.0 Hz, 1H), 8.34 (d, J = 7.6 Hz, 1H), 8.29 (d, J = 7.5 Hz, 1H), 8.25 (d, J = 8.9 Hz, 1H), 8.16 (d, J = 8.0 Hz, 1H), 8.13-8.08 (m, 3H), 8.01 (d, J = 8.0 Hz, 1H), 7.95-7.90 (m, 3H), 7.72 (d, J = 8.6 Hz, 1H), 7.62-7.54 (m, 4H), 6.84 (d, J = 7.3 Hz, 2H), 6.43 (t, J = 7.6 Hz, 1H), 6.27 (t, J = 7.8 Hz, 2H); ^{13}C NMR (126 MHz, 9:1 CD₂Cl₂:Acetone-*d*₆) δ 134.6 (CH), 134.5 (CH), 134.0 (CH), 133.8 (CH), 133.6 (C), 132.7 (C), 131.6 (CHx2), 131.5 (C), 131.15 (CH), 131.11 (C), 131.0 (CH), 130.8 (CH), 130.7 (CH), 130.1 (CH), 129.9 (CHx2), 128.1 (CHx2), 127.3 (CH), 127.2 (CH), 127.11 (CH), 127.10 (CH), 125.7 (C), 125.3 (CH), 125.2 (C), 124.7 (C), 124.3 (C), 124.2 (CH), 123.41 (C), 123.37 (C), 118.0 (C), 112.4 (C), 95.5 (C), 95.0 (C), 93.3 (C), 92.8 (C), 90.0 (C), 84.6 (C).

Complex 13·Ag(I): ^1H NMR (500 MHz, 9:1 CD₂Cl₂:Acetone-*d*₆) δ 8.11 (d, J = 9.0 Hz, 2H), 8.01 (bd, J = 9.0 Hz, 2H), 7.92 (bd, J = 7.5 Hz, 2H), 7.89 (d, J = 7.4 Hz, 2H), 7.85 (d, J = 7.5 Hz, 2H), 7.78 (d, J = 7.9 Hz, 2H), 7.76 (t, J = 7.5 Hz, 2H), 7.72 (d, J = 8.8 Hz, 2H), 7.71 (d, J = 9.0 Hz, 2H), 7.65 (td, J = 7.7, 1.5 Hz, 2H), 7.62 (td, J = 7.5, 1.5 Hz, 2H), 7.43 (d, J = 8.8 Hz, 2H), 7.38 (d, J = 7.9 Hz, 2H); ^{13}C NMR (126 MHz, 9:1 CD₂Cl₂:Acetone-*d*₆) δ 134.7 (CH), 134.0 (CH), 132.8 (C), 132.0 (C), 131.1 (CH), 130.9 (CH), 130.8 (C), 130.4 (C), 129.9 (CH), 129.7 (CH), 129.5 (CH), 126.9 (CH), 126.70 (CH), 126.68 (CH), 126.6 (CH), 125.9 (CH), 124.4 (CH), 123.8 (C), 123.6 (CH), 123.5 (C), 123.1 (C), 111.4 (C), 95.1 (C), 93.5 (C), 89.5 (C).

Complex 14·Ag(I): ^1H NMR (400 MHz, 9:1 CD₂Cl₂:Acetone-*d*₆) δ 8.56 (d, J = 9.0 Hz, 1H), 8.29 (d, J = 7.6 Hz, 1H), 8.24 (d, J = 7.7 Hz, 1H), 8.19 (d, J = 8.0 Hz, 1H), 8.15 (d, J = 8.9 Hz, 1H), 8.10 (t, J = 7.6 Hz, 1H), 8.05 (d, J = 8.0 Hz, 1H), 8.02 (d, J = 6.4 Hz, 1H), 8.00 (d, J = 6.4 Hz, 1H), 7.92 - 7.86 (m, 3H), 7.69 (d, J = 7.5 Hz, 1H), 7.62 - 7.50 (m, 4H), 7.29 (d, J = 8.5 Hz, 2H), 7.09 (d, J = 8.6 Hz, 2H); ^{13}C NMR (151 MHz, 9:1 CD₂Cl₂:Acetone-*d*₆) δ 147.0 (C), 133.4 (CH), 133.2 (CH), 132.9 (CH), 132.6 (C), 132.5 (CH), 132.41 (C), 132.39 (C), 131.5 (C), 131.2 (C), 130.6 (CH), 130.3 (CH), 130.1 (CH), 130.0 (CH), 129.7 (CH), 129.00 (CH), 128.97 (CH), 128.1 (C), 127.1 (CH), 126.7 (CH), 126.53 (CH), 126.50 (CH), 125.9 (C), 125.1 (CH), 125.0 (CH), 124.9 (C), 124.7 (C), 124.5 (C), 124.3 (C), 124.0 (CH), 123.2 (CH), 116.0 (C), 93.9 (C), 93.1 (C), 92.8 (C), 92.6 (C), 92.3 (C), 91.9 (C).

Complex 15·Ag(I): ^1H NMR (600 MHz, 9:1 CD₂Cl₂:Acetone-*d*₆) δ 8.56 (d, J = 9.0 Hz, 1H), 8.34 (d, J = 7.5 Hz, 1H), 8.29 (d, J = 7.5 Hz, 1H), 8.23 (d, J = 8.8 Hz, 1H), 8.21 (d, J = 7.9 Hz, 1H), 8.15 - 8.13 (m, 2H), 8.04 (d, J = 8.9 Hz, 1H), 8.02 (d, J = 8.0 Hz, 1H), 7.96-7.92 (m, 3H), 7.73 (d, J = 7.6 Hz, 1H), 7.65 - 7.55 (m, 4H), 6.16 (s, 1H), 5.81 (dd, J = 9.0, 2.7 Hz, 1H), 5.57 (d, J = 9.1 Hz, 1H), 3.35 (s, 3H), 3.13 (s, 3H); ^{13}C NMR (151 MHz, 9:1 CD₂Cl₂:Acetone-*d*₆) δ 152.9 (C), 151.3 (C), 134.5 (CH), 134.11 (CH), 134.10 (CH), 133.6 (CH), 133.6 (C), 132.5 (C), 131.5 (C), 131.2 (CH), 131.0 (CH), 130.9 (CH), 130.8 (CH), 130.6 (CH), 130.1 (CH), 129.8 (CH), 127.2 (CH), 127.2 (CH), 127.0 (CH), 126.9 (C), 125.5 (C), 125.1 (CH), 124.5 (C), 124.4 (C), 124.2 (C), 124.1 (CH), 123.3 (C), 122.8 (C), 117.4 (CH), 117.1 (CH), 112.5 (C), 110.8 (CH), 106.8 (C), 95.2 (C), 93.7 (C), 93.2 (C), 90.4 (C), 90.3 (C), 90.2 (C), 57.2 (CH₃), 55.4 (CH₃).

Complex 16·Ag(I): ^1H NMR (500 MHz, 9:1 CD₂Cl₂:Acetone-*d*₆) δ 7.88-7.83 (m, 4H), 7.59 - 7.55 (m, 4H), 7.44 (d, J = 7.2 Hz, 2H), 7.30 (t, J = 7.6 Hz, 1H), 7.15 (t, J = 7.8 Hz, 2H), 7.11 (d, J = 3.1 Hz, 1H), 6.90 (dd, J = 9.2, 3.1 Hz, 1H), 6.53 (d, J = 9.2 Hz, 1H), 3.77 (s, 3H), 3.58 (s, 3H); ^{13}C NMR (126 MHz, 9:1 CD₂Cl₂:Acetone-*d*₆) δ 154.3 (C), 152.9 (C), 134.5 (CH), 134.2 (CH), 134.0 (CH), 133.7 (CH), 132.7 (CHx2), 131.21 (CH), 131.18 (CH), 131.1 (CH), 130.9 (CH), 130.8 (CH), 129.3 (CHx2), 125.2 (C), 124.7 (C), 123.1 (C), 122.9 (C), 119.3 (CH), 118.7 (CH), 112.6 (CH), 96.5 (C), 93.3 (C), 92.9 (C), 91.0 (C), 90.8 (C), 85.5 (C), 57.2 (CH₃), 56.4 (CH₃), (two carbon signals were not observed).

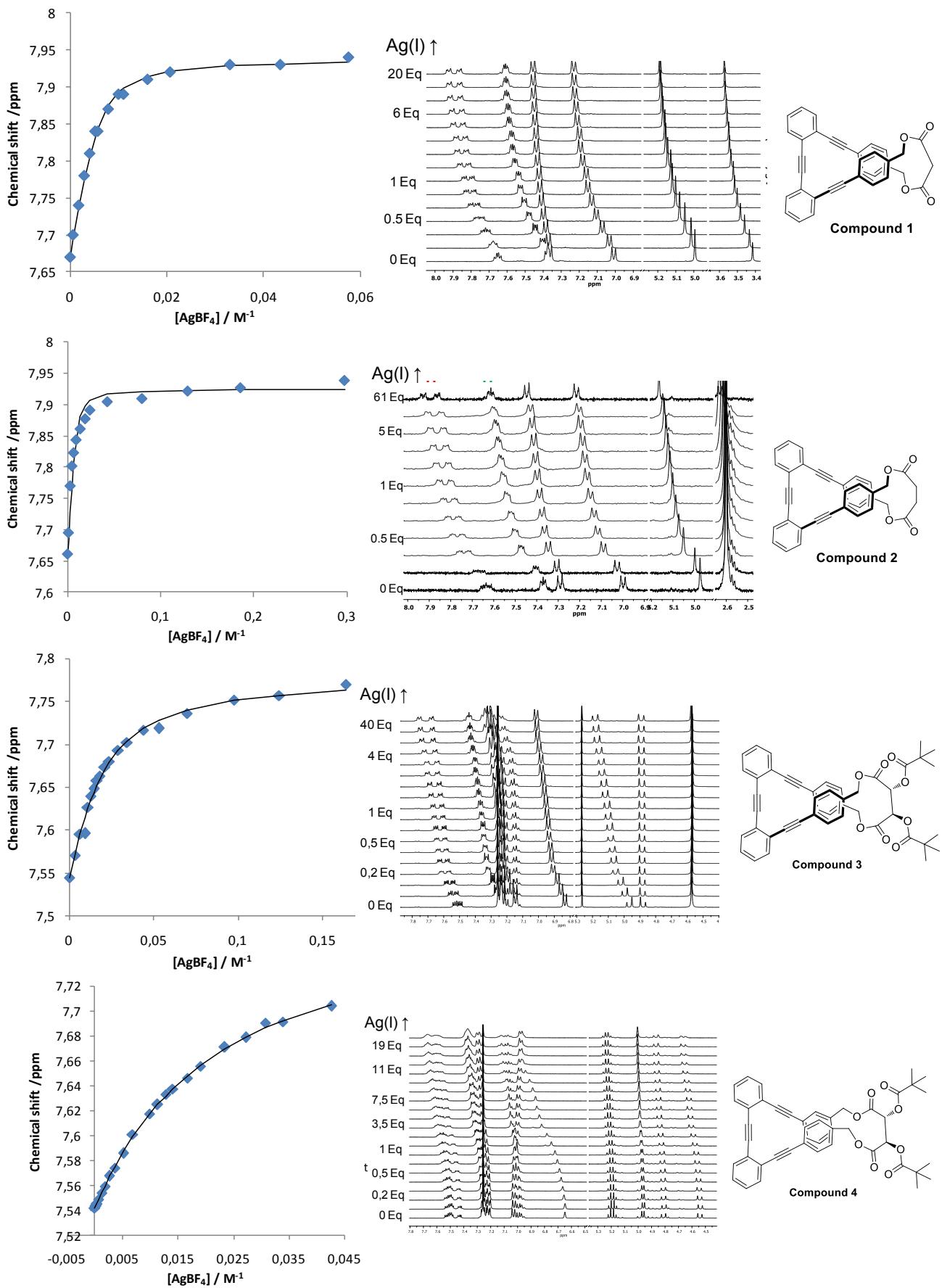
¹H-NMR titrations and binding constant determination

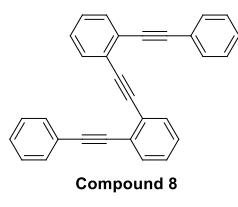
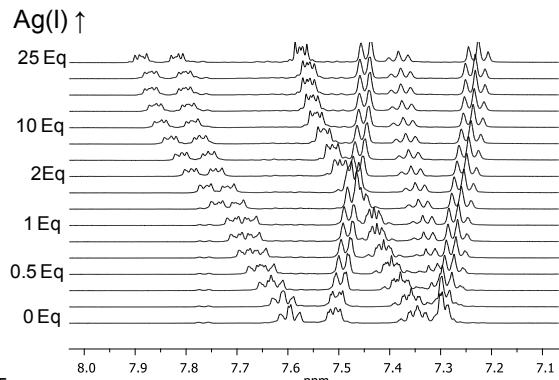
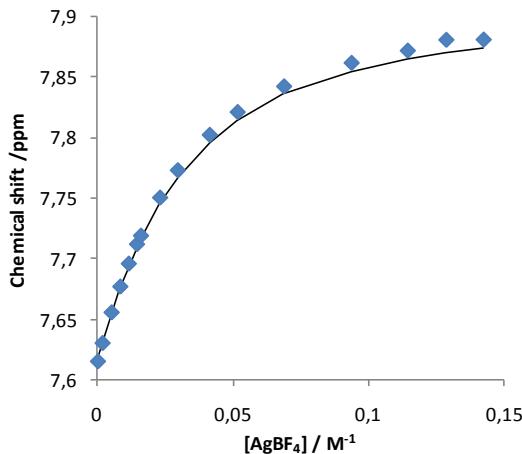
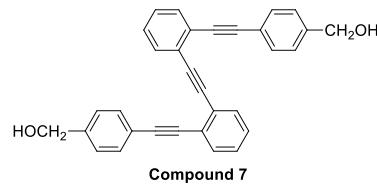
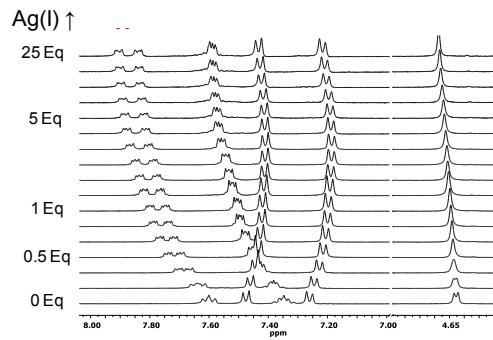
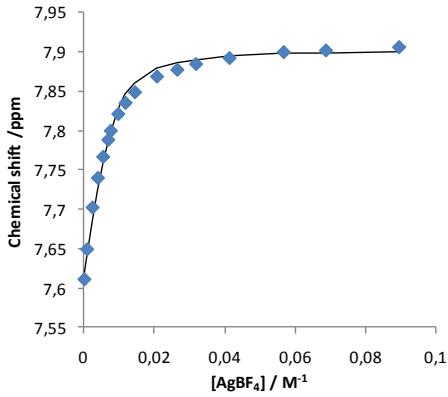
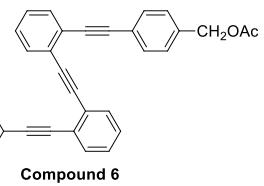
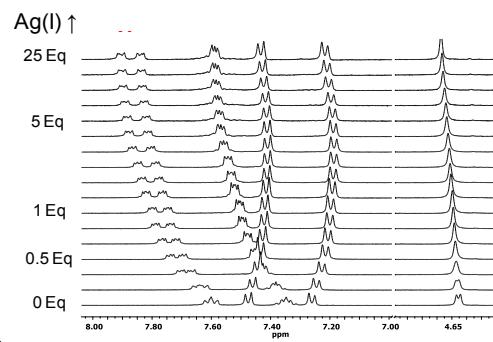
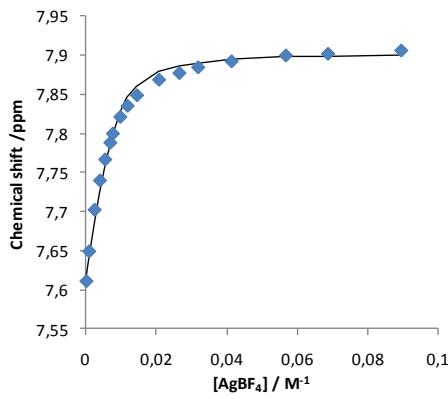
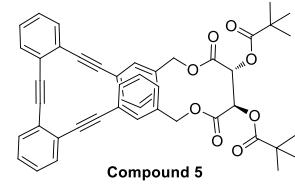
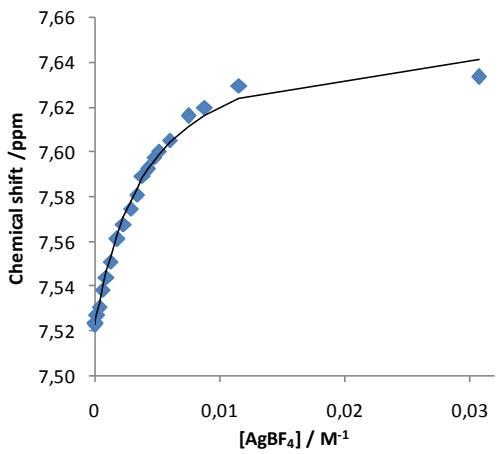
A stock solution of the ligands (10^{-2} - 10^{-3} M) in CD₂Cl₂/Acetone-d6 (v/v, 9:1) was firstly prepared. Using this solution as a solvent, a stock solution of AgBF₄ (20-200 mM depending on the ligand) was prepared. Then, 0.5 mL of the pure ligand solution was transferred to a NMR tube and initial spectra was registered. To this solution, small portions of the silver solution were added and the spectra recorded after each addition. Binding constants were determined by non-linear regression curve fitting methods using Program DynaFit⁷ plotting the differences measured between the initial chemical shifts of the selected protons⁸ against the concentration of the silver added.

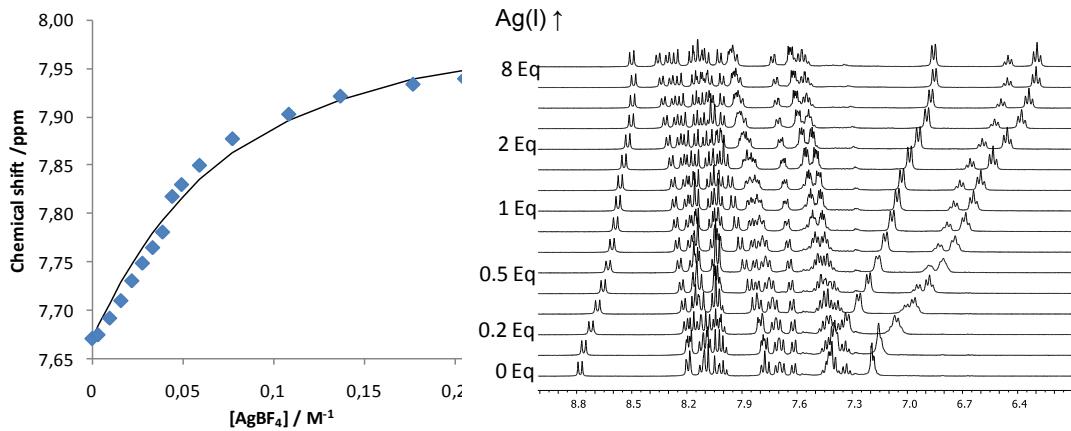
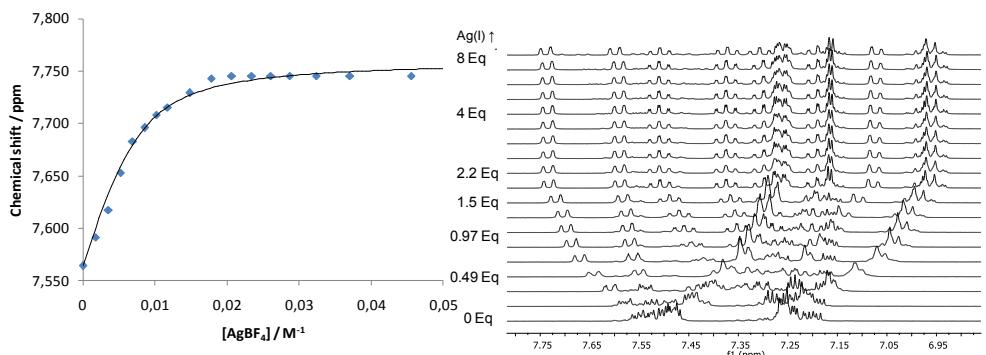
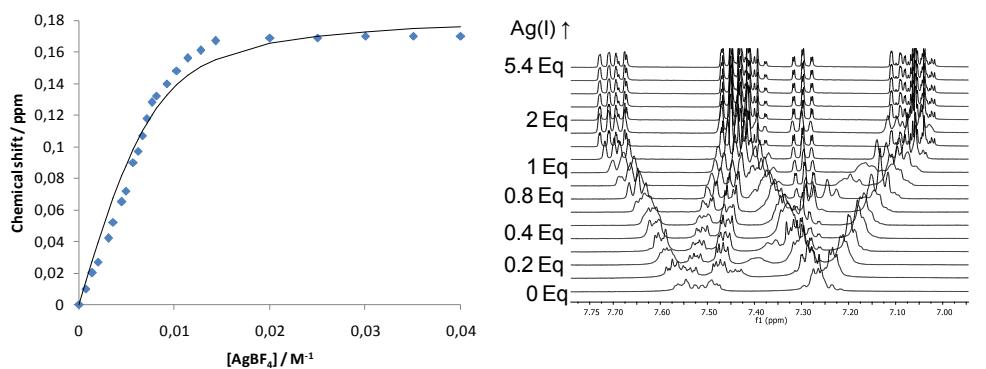
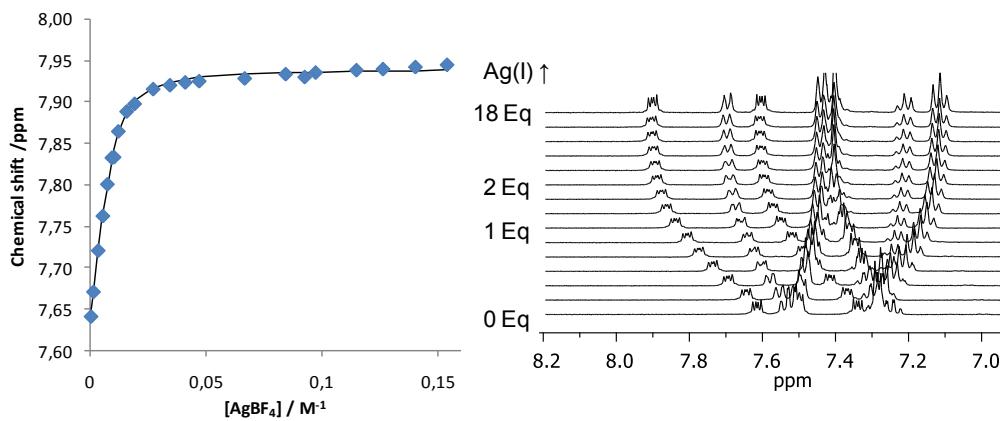
Compound	Ka (M ⁻¹) ± SD
1	1001±180
2	852±260
3	48.6±2.2
4	739±140
5	63.1±5.5
6	86.2±7.4
7	53.4±1.6
8	318±25
9	667±50
10	680 ± 24
11	559 ± 16
12	71±17
13	22.2±4.7
14	7.30±0.25
15	187±84
16	>10 ⁵ M ⁻¹

⁷ P. Kuzmic, *Analytical Biochemistry*, **1996**, 237, 260-273.

⁸ For the calculations all the identifiable signals were included in a global fitting except for the indicated cases.







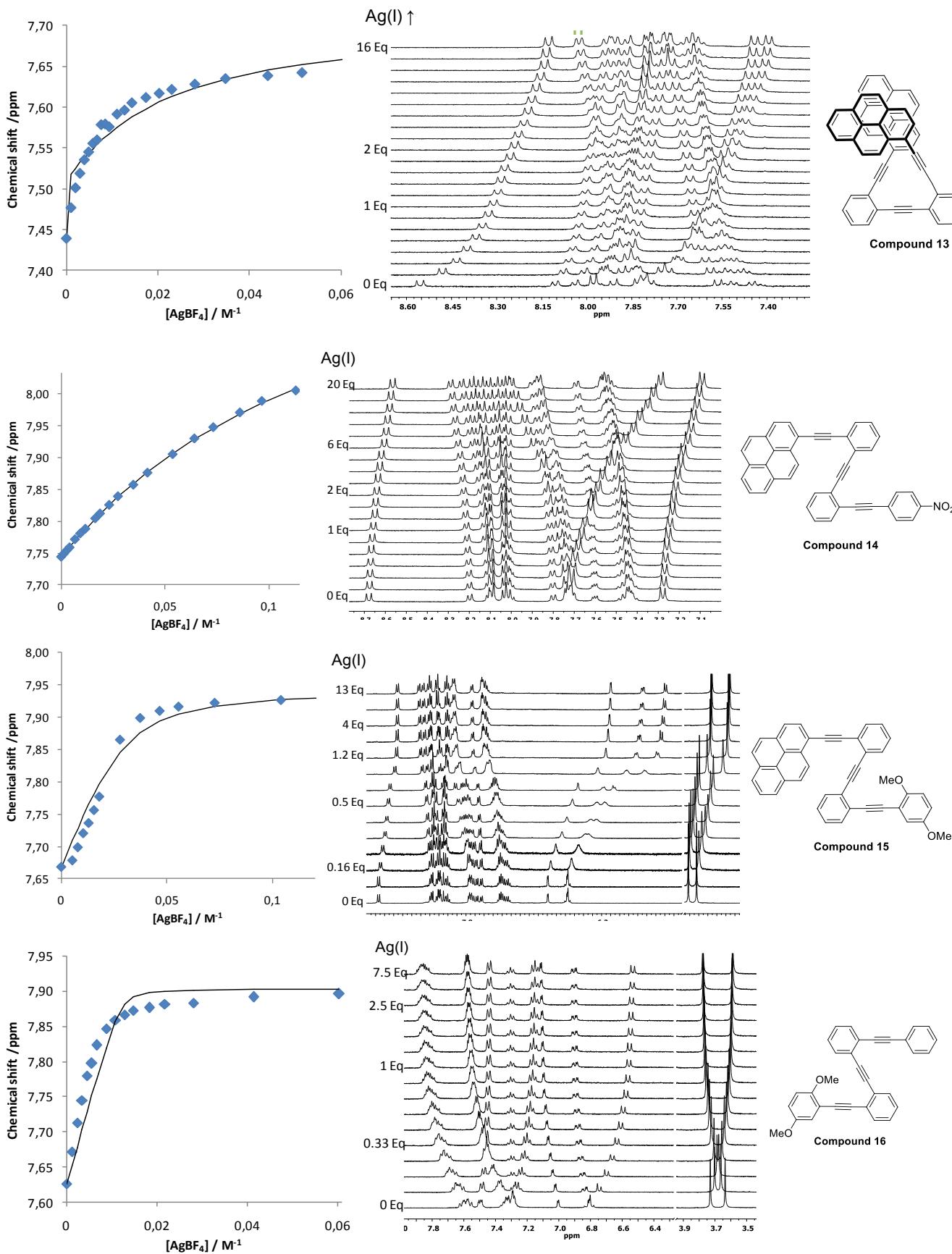


Figure S1: Influence of silver coordination evaluated in the ^1H and ^{13}C NMR data of 1 and 6.

In the work of Pale et al. (ref. 18 manuscript), three important aspects were used to evidence the *in situ* formation of a Ag(I)-alkyne complex: 1) a decrease on the coupling constant $^3\langle ^3\text{H}-^1\text{H} \rangle$ of the neighboring protons which indicated a loss of electron density of the triple bond.

- Interestingly, analogous hints of coordination were recorded in our case (See Figure S1).
 2) the deshielding of the alkyne neighboring protons and
 3) the shielding of the internal sp carbon which was accompanied by a deshielding of the terminal carbon.



Figure S2: ^1H -NMR spectra and titration curves for tolane, 1,2-bis(phenylethynyl)benzene and 1,3-bis((2-phenylethynyl)phenyl)ethynyl)benzene.

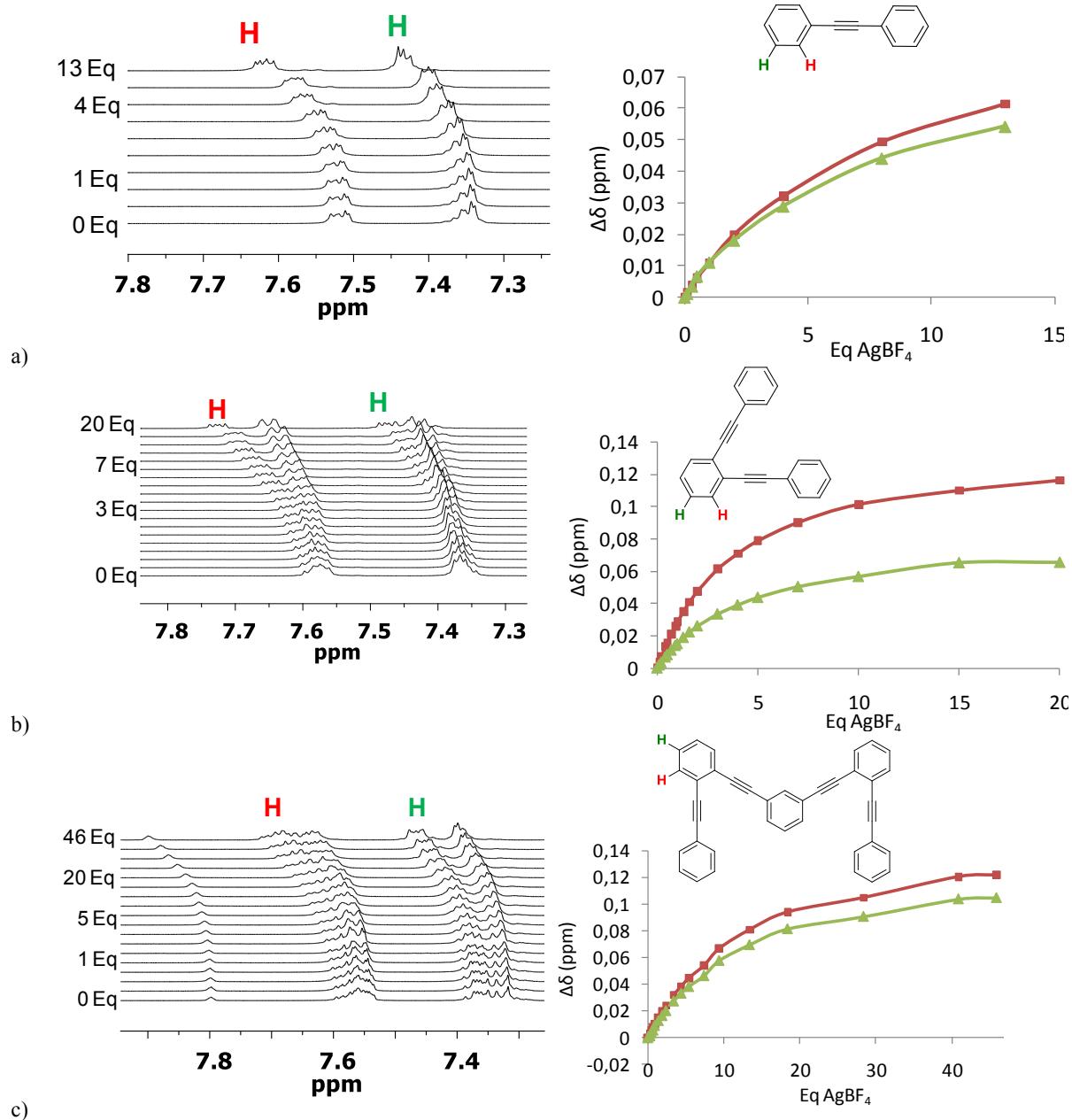
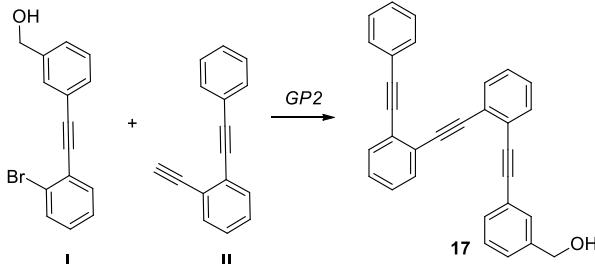


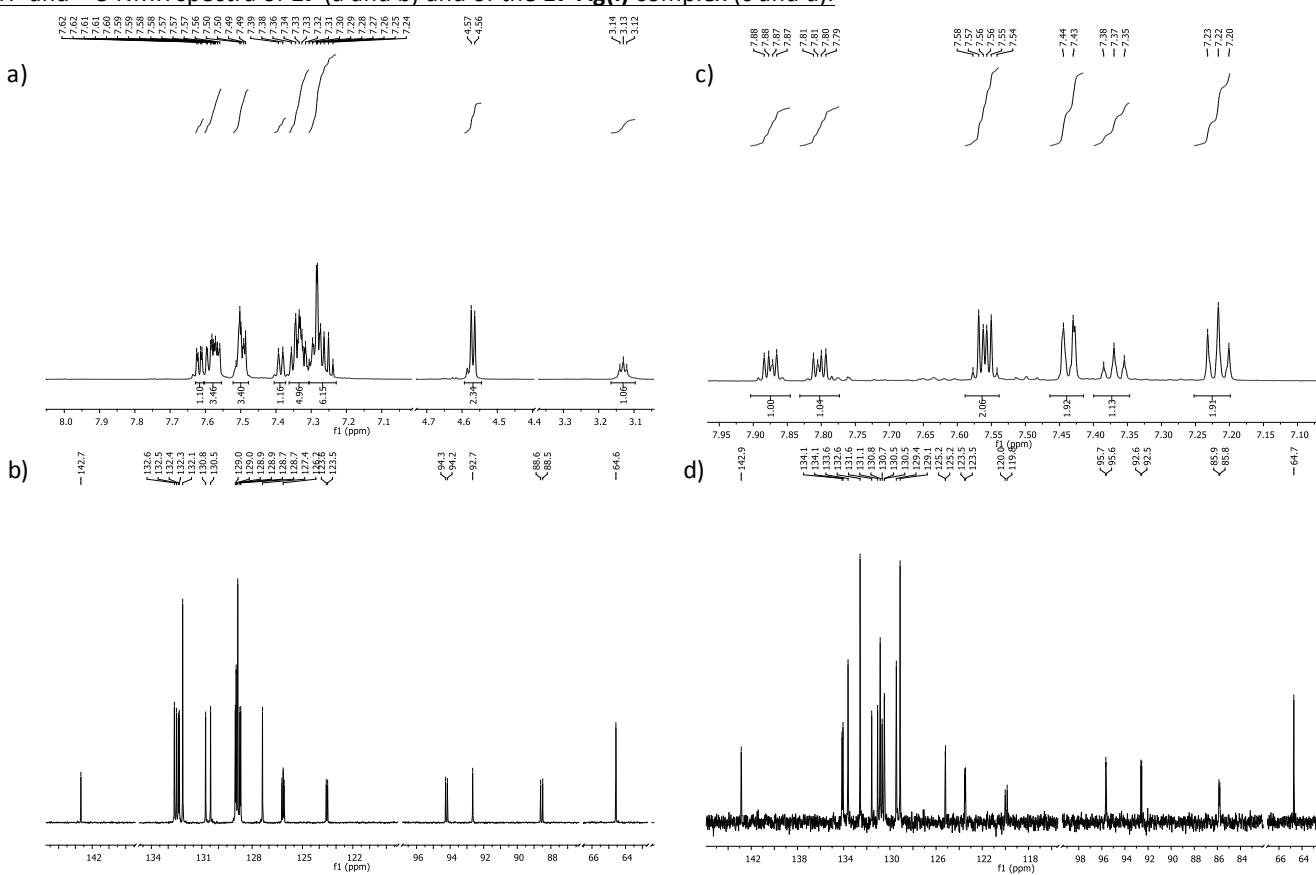
Figure S3: Synthesis, analytical characterization, NMR spectra and titration curves for the monohydroxylated ligand 17.

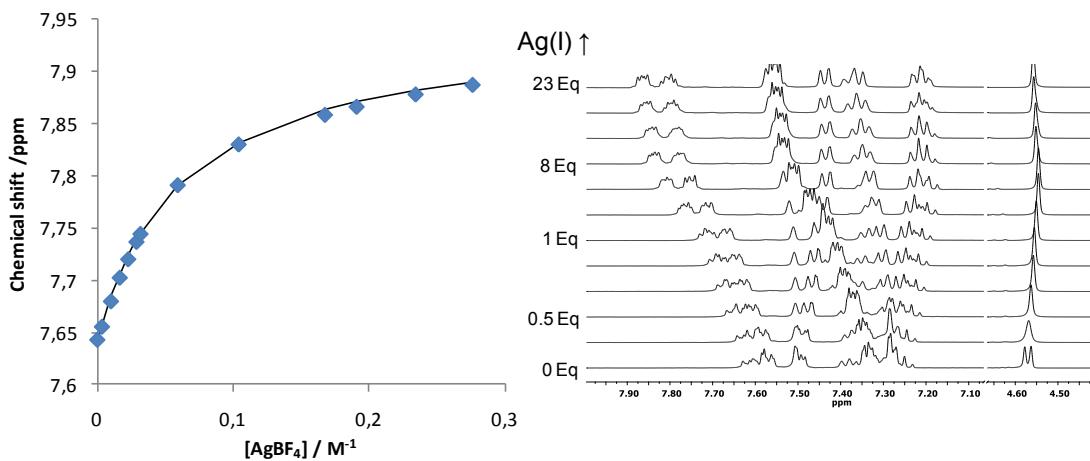


Compound 17 was prepared from I (300 mg, 1.15 mmol) and 1-ethynyl-2-(phenylethyynyl)benzene II (436 mg, 1.26 mmol) according to previously described GP2 to give 5 in 53 % yield (315 mg) as a pale yellow syrup. ^1H NMR (600 MHz, 9:1 CD₂Cl₂:Acetone-*d*₆) δ 7.64 – 7.55 (m, 4H), 7.52 – 7.48 (m, 3H), 7.39 (d, *J* = 7.4 Hz, 1H), 7.36 – 7.31 (m, 4H), 7.31 – 7.23 (m, 5H), 4.57 (d, *J* = 5.9 Hz, 2H), 3.13 (t, *J* = 5.8 Hz, 1H); ^{13}C NMR (151 MHz, 9:1 CD₂Cl₂:Acetone-*d*₆) δ 142.7 (C), 132.6 (CH), 132.5 (CH), 132.4 (CH), 132.3 (CH), 132.1 (CH), 130.8 (CH), 130.5 (CH), 129.0 (CH), 128.9 (CH), 128.70 (CH), 128.67 (CH), 127.4 (CH), 126.2 (C), 126.18 (C), 126.16 (C), 126.1 (C), 123.6 (C), 123.5 (C), 94.3 (C), 94.2 (C), 92.7 (C), 88.6 (C), 88.5 (C), 64.6 (CH₂), (some carbon signals were not observed); ESI-HRMS calculated for C₃₁H₂₁O [M+H⁺]: 409.1597, found: 409.1586.

Complex 17·Ag(I): ^1H NMR (500 MHz, 9:1 CD₂Cl₂:Acetone-*d*₆) δ 7.87 (dd, *J* = 6.0, 3.1 Hz, 2H), 7.80 (dd, *J* = 5.9, 3.2 Hz, 2H), 7.59 – 7.54 (m, 4H), 7.43 (bd, *J* = 7.5 Hz, 4H), 7.37 (t, *J* = 7.6 Hz, 2H), 7.22 (bt, *J* = 7.8 Hz, 4H). ^{13}C NMR (126 MHz, 9:1 CD₂Cl₂:Acetone-*d*₆) δ 142.9 (C), 134.1 (CH), 134.1 (CH), 133.6 (CH), 132.6 (CH), 131.6 (CH), 131.1 (CH), 130.8 (CH), 130.7 (CH), 130.5 (CH), 130.5 (CH), 129.4 (CH), 129.1 (CH), 125.2 (C), 125.2 (C), 123.5 (C), 123.5 (C), 120.0 (C), 119.9 (C), 95.7 (C), 95.6 (C), 92.6 (C), 92.6 (C), 85.9 (C), 85.8 (C), 64.7 (CH₂).

¹H- and ¹³C-NMR spectra of **17** (a and b) and of the **17·Ag(I)** complex (c and d).



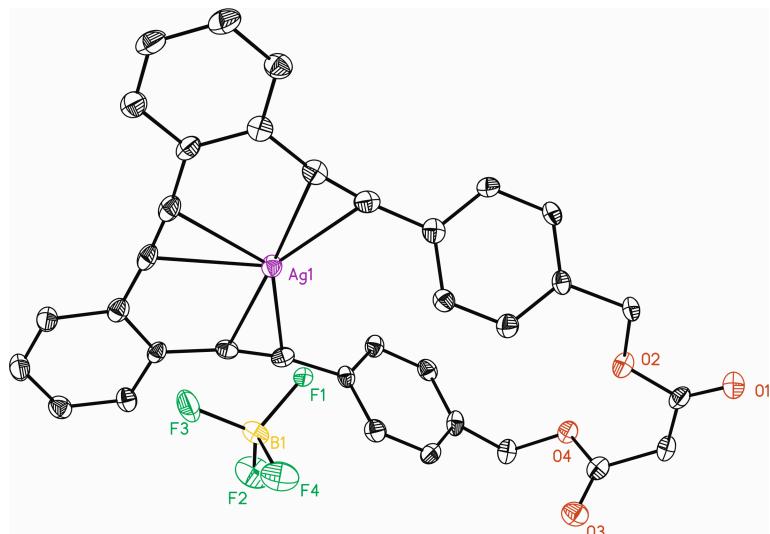


Compound	$K_a (M^{-1}) \pm SD$
17	16.82 ± 0.81

X-ray crystallographic analyses

Complex 1·Ag(I)

Under inert argon atmosphere, AgBF₄ (19.5 mg, 0.1 mmol) was added to a solution of **1** (50.7 mg, 0.1 mmol) in dry CH₂Cl₂ (10 ml). The mixture was stirred for 4 h with the exclusion of light. A pale brown precipitate formed which was collected by vacuum filtration, dissolved in acetone, filtered through celite and then the solvent removed under reduced pressure to give **1·Ag(I)** as a pale brown solid (16.4 mg). The filtrate from the initial filtration was filtered through celite and then solvent removed under reduced pressure to give **1·Ag(I)** as a brown solid (40.2 mg). X-Ray quality crystals of **1·Ag(I)** were obtained by slow diffusion of hexane into a solution of the complex in dichloromethane. Thermal ellipsoids are drawn at the 50% probability level. Hydrogen atoms and solvent molecules are omitted for clarity.



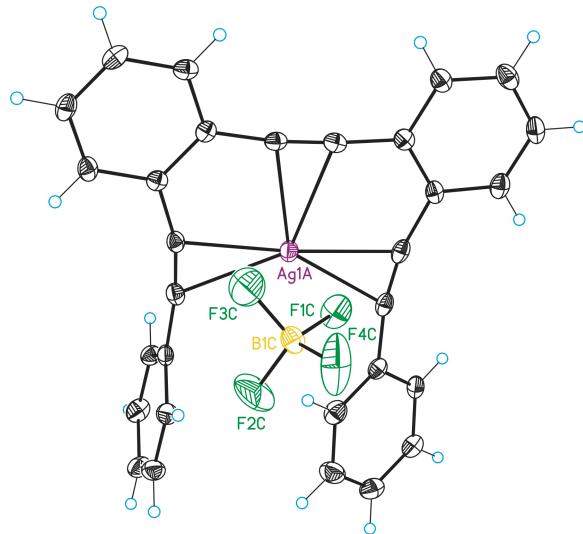
Crystal data and structure refinement for **1**·Ag(I)

Empirical formula	C35 H22 Ag B F4 O4	
Formula weight	701.21	
Temperature	100(2) K	
Wavelength	0.71073 Å	
Crystal system	Monoclinic	
Space group	Cc	
Unit cell dimensions	a = 11.3153(9) Å b = 14.4329(9) Å c = 17.5533(11) Å	a = 90.00 °. b = 90.232(4) °. g = 90.00 °.
Volume	2866.7(3) Å ³	
Z	4	
Density (calculated)	1.625 Mg/m ³	
Absorption coefficient	0.770 mm ⁻¹	
F(000)	1408	
Crystal size	0.01 x 0.01 x 0.005 mm ³	

Theta range for data collection	2.29 to 30.53 °.
Index ranges	-16 <=h<=15 , -20 <=k<=18 , -24 <=l<=24
Reflections collected	11863
Independent reflections	5616 [R(int) = 0.0515]
Completeness to theta =30.53 °	87.5%
Absorption correction	Empirical
Max. and min. transmission	0.9962 and 0.9923
Refinement method	Full-matrix least-squares on F ²
Data / restraints / parameters	5616 / 232 / 496
Goodness-of-fit on F ²	1.062
Final R indices [I>2sigma(I)]	R1 = 0.0471 , wR2 = 0.1137
R indices (all data)	R1 = 0.0594 , wR2 = 0.1212
Flack parameter	x = -0.04(3)
Largest diff. peak and hole	1.072 and -0.835 e.Å ⁻³

8·Ag(I)

Under inert argon atmosphere, AgBF₄ (19.5 mg, 0.100 mmol) was added to a solution of **8** (37.8 mg, 0.100 mmol) in dry CH₂Cl₂ (10 ml). The mixture was stirred overnight with the exclusion of light. The solution was filtered through celite and the filtrate evaporated to dryness to give **8·Ag(I)** as a pale brown solid (49.1 mg). X-Ray quality crystals were obtained by slow diffusion of pentane into a solution of the complex **8·Ag(I)** in dichloromethane. Thermal ellipsoids are drawn at the 50% probability level. Hydrogen atoms and solvent molecules are omitted for clarity.



Crystal data and structure refinement for **8·Ag(I)**

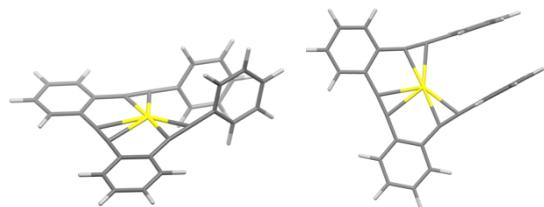
Identification code	mo_AJ116_P21c
Empirical formula	C ₃₀ H ₁₈ AgBF ₄

Formula weight	573.12
Temperature	100(2) K
Wavelength	0.71073 Å
Crystal system	Monoclinic
Space group	P2(1)/c
Unit cell dimensions	a = 17.2933(12) Å a= 90.00 °. b = 27.625(2) Å b = 104.778(2) °. c = 10.1403(7) Å g = 90.00 °.
Volume	4684.0(6) Å ³
Z	8
Density (calculated)	1.625 Mg/m ³
Absorption coefficient	0.910 mm ⁻¹
F(000)	2288
Crystal size	0.15 x 0.03 x 0.03 mm ³
Theta range for data collection	1.22 to 29.96 °.
Index ranges	-16 <=h<=24 , -31 <=k<=38 , -14 <=l<=13
Reflections collected	28590
Independent reflections	13454 [R(int) = 0.0361]
Completeness to theta =29.96 °	98.9%
Absorption correction	Empirical
Max. and min. transmission	0.9732 and 0.8756
Refinement method	Full-matrix least-squares on F ²
Data / restraints / parameters	13454 / 0 / 649
Goodness-of-fit on F ²	1.023
Final R indices [I>2sigma(I)]	R1 = 0.0386 , wR2 = 0.0752
R indices (all data)	R1 = 0.0677 , wR2 = 0.0839
Largest diff. peak and hole	0.730 and -0.642 e.Å ⁻³

Theoretical studies

All theoretical calculations were carried out using the M06 functional with the 6–31G* basis set for the ligands and the LANL2DZ relativistic effective core potential basis set for the Ag(I) ion as implemented in the Gaussian 09 program⁹

XYZ Coordinates of all the calculated structures

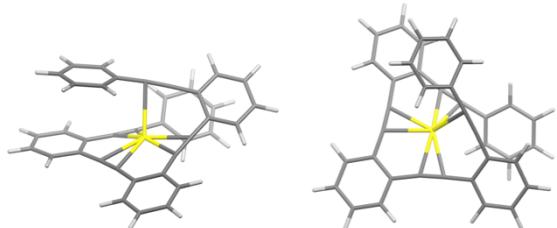


Complex 8·Ag(I) = AgC₃₀H₁₈

C	-3.666752000	1.237785000	1.028683000
C	-2.719431000	1.110322000	2.041971000
H	-3.015716000	0.755530000	3.026767000
C	-1.395724000	1.449014000	1.801515000
H	-0.646785000	1.367486000	2.587788000
C	-1.016856000	1.921776000	0.535591000
C	0.341286000	2.306096000	0.301812000
C	1.508697000	2.681249000	0.154017000
C	2.731947000	3.432420000	0.105562000
C	2.687885000	4.804506000	0.374839000
H	1.728797000	5.266925000	0.598572000
C	3.850122000	5.562198000	0.357094000
H	3.801187000	6.628131000	0.566626000
C	5.073589000	4.959370000	0.073257000
H	5.984807000	5.552976000	0.059670000
C	5.137355000	3.600274000	-0.196947000
H	6.089874000	3.125280000	-0.421570000
C	3.974074000	2.819704000	-0.188903000
C	4.071043000	1.426186000	-0.477513000
C	4.224230000	0.236879000	-0.738831000
C	4.495938000	-1.131628000	-1.034988000
C	3.455112000	-2.069460000	-1.243217000
C	2.072005000	-1.685886000	-1.187593000
C	0.839212000	-1.628773000	-1.236510000
C	-0.586388000	-1.594741000	-1.361684000
C	-1.164687000	-1.239854000	-2.590107000
H	-0.520882000	-0.994066000	-3.433349000
C	-2.545897000	-1.222631000	-2.721159000
H	-2.994437000	-0.958194000	-3.676369000

⁹ Gaussian 09, Revision B.01, M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, G. Scalmani, V. Barone, B. Mennucci, G. A. Petersson, H. Nakatsuji, M. Caricato, X. Li, H. P. Hratchian, F. Izmaylov, J. Bloino, G. Zeng, J. L. Sonneberg, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, T. Vreven, J. A. Montgomery, Jr., J. E. Peralta, F. Ogliaro, M. Bearpark, J. J. Heyd, E. Brothers, K. N. Kudin, V. N. Staroverov, T. Keith, R. Kobayashi, J. Normand, R. Raghavachari, A. Rendell, J. C. Burant, S. S. Iyengar, J. Tomasi, M. Cossi, N. Rega, J. M. Millam, M. Klene, J. E. Knox, J. B. Cross, V. Bakken, C. Adamo, J. Jaramillo, R. Gomperts, R. E. Stratmann, O. Yazyev, A. J. Austin, R. Cammi, C. Pomelli, J. W. Ochterski, R. L. Martin, K. Morokuma, V. G. Zakrzewski, G. A. Voth, P. Salvador, J. J. Dannenberg, S. Dapprich, A. D. Daniels, O. Farkas, J. B. Foresman, J. V. Ortiz, J. Cioslowski, D. J. Fox, Gaussian, Inc., Wallingford CT, 2010.

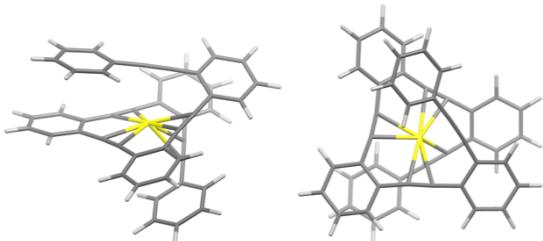
C	-3.353208000	-1.552982000	-1.635158000
C	-2.782683000	-1.898860000	-0.411950000
H	-3.417304000	-2.155250000	0.433935000
C	-1.403016000	-1.920504000	-0.268379000
H	-0.944059000	-2.191154000	0.681544000
C	3.777713000	-3.401522000	-1.524070000
H	2.972690000	-4.116136000	-1.682184000
C	5.103886000	-3.803922000	-1.597535000
H	5.339189000	-4.843394000	-1.813911000
C	6.128105000	-2.881232000	-1.395092000
H	7.166936000	-3.198095000	-1.452263000
C	5.828214000	-1.555809000	-1.117394000
H	6.623737000	-0.831532000	-0.956616000
C	-1.972010000	2.039293000	-0.485556000
H	-1.668075000	2.406367000	-1.465017000
C	-3.292656000	1.698026000	-0.232167000
H	-4.035580000	1.793514000	-1.021514000
Ag	1.725903000	0.487478000	-0.493525000
H	-4.436374000	-1.542615000	-1.742601000
H	-4.706103000	0.979427000	1.223375000



Complex 9·Ag(I) = AgC₃₈H₂₂

C	1.038405000	-1.567830000	-1.371896000
C	-0.166288000	-1.763062000	-1.505859000
C	-1.568082000	-2.028696000	-1.602747000
C	-2.083916000	-3.227371000	-1.088321000
H	-1.404187000	-3.956445000	-0.649852000
C	-3.448762000	-3.472223000	-1.142246000
H	-3.845627000	-4.404142000	-0.744712000
C	-4.307149000	-2.531304000	-1.706744000
C	-3.800046000	-1.343741000	-2.229931000
H	-4.471422000	-0.612115000	-2.674978000
C	-2.437757000	-1.086501000	-2.174926000
H	-2.031162000	-0.161212000	-2.585215000
H	-5.376646000	-2.728383000	-1.747179000
C	4.589262000	-2.659009000	-1.404249000
C	3.204824000	-2.671109000	-1.496469000
H	2.674415000	-3.589108000	-1.742253000
C	2.463699000	-1.504310000	-1.278456000
C	3.144703000	-0.304853000	-0.949816000
C	2.446300000	0.915615000	-0.704488000
C	1.917015000	2.006192000	-0.505055000
C	1.421427000	3.336222000	-0.345315000
C	2.313926000	4.404679000	-0.507571000
H	3.355552000	4.187669000	-0.734953000
C	1.877910000	5.715912000	-0.392044000
H	2.584945000	6.531234000	-0.526093000

C	0.540057000	5.985507000	-0.110796000
H	0.195031000	7.013014000	-0.022343000
C	-0.358069000	4.943049000	0.058889000
H	-1.403070000	5.145816000	0.283905000
C	0.062005000	3.611087000	-0.051734000
C	-0.911124000	2.581769000	0.150702000
C	-1.879038000	1.868460000	0.411372000
C	4.542889000	-0.311148000	-0.868119000
H	5.058956000	0.614357000	-0.620648000
C	5.259062000	-1.477907000	-1.092894000
H	6.344429000	-1.465238000	-1.024918000
Ag	0.041941000	0.394042000	-0.251965000
H	5.148335000	-3.575121000	-1.581315000
C	-3.028157000	1.086767000	0.731702000
C	-2.901610000	-0.182080000	1.356296000
C	-1.612796000	-0.693796000	1.680990000
C	-0.491368000	-1.099234000	1.939703000
C	0.849015000	-1.539084000	2.164182000
C	1.821772000	-0.620184000	2.591494000
H	1.523861000	0.403702000	2.818482000
C	3.144308000	-1.021021000	2.725008000
H	3.892926000	-0.307594000	3.063934000
C	3.509180000	-2.333705000	2.433763000
C	2.547261000	-3.250846000	2.014648000
H	2.833037000	-4.277232000	1.792104000
C	1.221425000	-2.861916000	1.883651000
H	0.462637000	-3.570418000	1.553919000
C	-4.056274000	-0.918075000	1.637601000
H	-3.953093000	-1.894037000	2.107770000
C	-5.311449000	-0.410824000	1.326498000
H	-6.199644000	-0.993808000	1.560476000
C	-5.434863000	0.838561000	0.723547000
H	-6.418509000	1.237692000	0.486603000
C	-4.301087000	1.580998000	0.423974000
H	-4.388906000	2.556200000	-0.051122000
H	4.547238000	-2.644735000	2.534709000

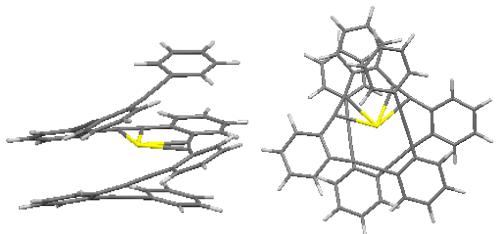


Complex 10·Ag(I) = AgC46H26

C	1.657503000	1.770220000	0.110282000
C	2.251370000	0.829477000	-0.413359000
C	3.107996000	-0.164260000	-0.979020000
C	4.484806000	0.099392000	-0.994801000
H	4.840982000	1.040914000	-0.582572000
C	5.380951000	-0.818073000	-1.520462000
H	6.444858000	-0.592051000	-1.526399000
C	4.916974000	-2.024148000	-2.040862000
H	5.615571000	-2.747125000	-2.456693000
C	3.559840000	-2.307975000	-2.029538000

H	3.188172000	-3.248951000	-2.430513000
C	2.636820000	-1.393880000	-1.503469000
C	1.257763000	-1.764957000	-1.492920000
C	0.124954000	-2.227804000	-1.537856000
Ag	-0.045955000	-0.008220000	-0.114663000
C	-1.200187000	-2.754559000	-1.550381000
C	-2.318398000	-1.886125000	-1.654051000
C	-2.120759000	-0.480715000	-1.773619000
C	-1.931696000	0.720412000	-1.886817000
C	-1.679313000	2.122832000	-1.972664000
C	-0.531216000	2.589930000	-2.632945000
H	0.127115000	1.873210000	-3.124159000
C	-0.247554000	3.948460000	-2.651647000
H	0.640732000	4.308173000	-3.167329000
C	-1.099929000	4.849061000	-2.015427000
C	-2.244532000	4.391638000	-1.365503000
H	-2.910859000	5.098540000	-0.874105000
C	-2.539052000	3.035455000	-1.343851000
H	-3.423211000	2.662740000	-0.828119000
C	-3.608612000	-2.422785000	-1.612163000
H	-4.459328000	-1.747526000	-1.682815000
C	-3.798557000	-3.792381000	-1.485653000
H	-4.808479000	-4.196370000	-1.460787000
C	-2.701686000	-4.646632000	-1.394242000
H	-2.851163000	-5.719944000	-1.299350000
C	-1.412093000	-4.132002000	-1.422843000
H	-0.551087000	-4.793253000	-1.345897000
H	-0.871154000	5.913140000	-2.026873000
C	-0.996910000	1.860731000	1.441375000
C	-1.818613000	0.969189000	1.610608000
C	-2.737285000	-0.110282000	1.759262000
C	-4.116933000	0.119833000	1.732859000
H	-4.480613000	1.141513000	1.637606000
C	-5.008394000	-0.940726000	1.827678000
H	-6.078916000	-0.748432000	1.809966000
C	-4.533226000	-2.245642000	1.944846000
C	-3.167483000	-2.494333000	1.975042000
H	-2.788978000	-3.510379000	2.069671000
C	-2.253922000	-1.439532000	1.891900000
H	-5.232835000	-3.075993000	2.015565000
C	0.161981000	5.374807000	1.740235000
C	-0.610022000	4.226715000	1.835760000
H	-1.586665000	4.255174000	2.315817000
C	-0.153743000	3.006120000	1.320141000
C	1.112730000	2.959734000	0.683114000
C	1.878227000	4.131488000	0.599319000
H	2.848706000	4.086087000	0.108735000
C	1.410034000	5.326540000	1.122623000
H	2.020547000	6.223634000	1.049248000
H	-0.210907000	6.310819000	2.150155000
C	0.362713000	-1.791932000	1.989839000
C	-0.849706000	-1.668698000	1.925393000
C	3.886518000	-0.768675000	2.551720000
C	2.499546000	-0.748650000	2.538511000
H	1.948759000	0.124449000	2.889241000

C	1.784758000	-1.861041000	2.063373000
C	2.481411000	-2.996210000	1.623985000
C	3.869246000	-3.008129000	1.645294000
H	4.407994000	-3.888745000	1.301229000
C	4.572582000	-1.894619000	2.100683000
H	5.660893000	-1.907029000	2.110283000
H	4.435750000	0.095884000	2.920095000
H	1.920611000	-3.855746000	1.258653000



Complex 11·Ag(I) = AgC₅₄H₃₀

C	0.998045000	-2.250330000	-1.816025000
C	-0.180102000	-2.209768000	-2.122285000
C	-1.556705000	-2.167573000	-2.479671000
C	-2.032932000	-1.152871000	-3.323917000
H	-1.331085000	-0.417484000	-3.716541000
C	-3.381335000	-1.097842000	-3.648934000
H	-3.746579000	-0.313267000	-4.309037000
C	-4.263226000	-2.047388000	-3.138065000
C	-3.795334000	-3.059758000	-2.302153000
H	-4.486150000	-3.802783000	-1.907741000
C	-2.448767000	-3.126149000	-1.975816000
H	-2.068003000	-3.908896000	-1.320687000
H	-5.319556000	-2.002410000	-3.396847000
C	-0.942348000	-1.946389000	1.219755000
C	-1.864652000	-1.250636000	0.809201000
C	-3.057613000	-0.591251000	0.386586000
C	-4.284703000	-1.163915000	0.749767000
H	-4.278045000	-2.070031000	1.353143000
C	-5.483081000	-0.589506000	0.355946000
H	-6.424129000	-1.049089000	0.650442000
C	-5.478840000	0.572617000	-0.412680000
H	-6.416529000	1.028114000	-0.723733000
C	-4.276156000	1.152042000	-0.785340000
H	-4.262794000	2.058177000	-1.388589000
C	-3.053316000	0.583237000	-0.402271000
C	-1.856282000	1.244877000	-0.809713000
C	-0.931885000	1.942419000	-1.213030000
C	1.865135000	-1.255430000	1.790447000
C	2.426211000	-0.171320000	1.738998000
C	3.087554000	1.076866000	1.576285000
C	4.467153000	1.112355000	1.339231000
H	5.027879000	0.178826000	1.359711000
C	5.099492000	2.314404000	1.055241000
H	6.171823000	2.332176000	0.872369000
C	4.361049000	3.496513000	1.002550000
C	2.996646000	3.482576000	1.257052000
H	2.414047000	4.402252000	1.226999000
C	2.343456000	2.282807000	1.559032000

H	4.855966000	4.438032000	0.773017000
C	1.823338000	-4.913786000	2.408205000
C	2.253276000	-3.600298000	2.296460000
H	3.289037000	-3.336688000	2.501763000
C	1.367552000	-2.583776000	1.914125000
C	0.018793000	-2.913665000	1.639261000
C	-0.396616000	-4.247495000	1.757344000
H	-1.436380000	-4.491876000	1.546939000
C	0.495264000	-5.238991000	2.137542000
H	0.154049000	-6.268205000	2.222806000
H	2.525835000	-5.687805000	2.709457000
C	-0.229853000	2.208876000	2.128867000
C	0.949523000	2.252673000	1.829108000
C	-3.432045000	1.075136000	3.636989000
C	-2.082190000	1.140216000	3.320311000
H	-1.377182000	0.410628000	3.717968000
C	-1.608134000	2.157412000	2.477722000
C	-2.504659000	3.108411000	1.967380000
C	-3.852773000	3.031796000	2.284918000
H	-4.546847000	3.768851000	1.885027000
C	-4.318228000	2.016927000	3.119152000
H	-5.375636000	1.964660000	3.372084000
H	-3.795204000	0.288957000	4.296395000
H	-2.126099000	3.893567000	1.313778000
Ag	0.226553000	-0.011412000	-0.017114000
C	0.026539000	2.914898000	-1.627244000
C	1.376774000	2.592574000	-1.904390000
C	1.882803000	1.267084000	-1.783716000
C	2.452284000	0.187360000	-1.729254000
C	3.123924000	-1.054537000	-1.559248000
C	2.390970000	-2.267223000	-1.539032000
C	3.053326000	-3.459471000	-1.226939000
H	2.479027000	-4.384207000	-1.194515000
C	4.416507000	-3.459249000	-0.965435000
C	5.144398000	-2.270789000	-1.021624000
H	6.215866000	-2.277555000	-0.833136000
C	4.502559000	-1.076135000	-1.315686000
H	5.054660000	-0.137723000	-1.340310000
C	2.257260000	3.615056000	-2.282955000
H	3.294523000	3.357681000	-2.488624000
C	1.820931000	4.926868000	-2.389251000
H	2.519547000	5.705517000	-2.687458000
C	0.491458000	5.244523000	-2.116372000
H	0.145155000	6.272404000	-2.197348000
C	-0.395180000	4.247251000	-1.739492000
H	-1.435864000	4.485837000	-1.526836000
H	4.919084000	-4.394664000	-0.727759000

Photophysical studies

Absorption and emission studies

Figure S4: Normalized absorption (dotted line) and emission (solid line, excitation wavelength between parenthesis) spectra of compounds **12** ($2.7 \cdot 10^{-5}$ M) and **13** ($4.4 \cdot 10^{-5}$ M)

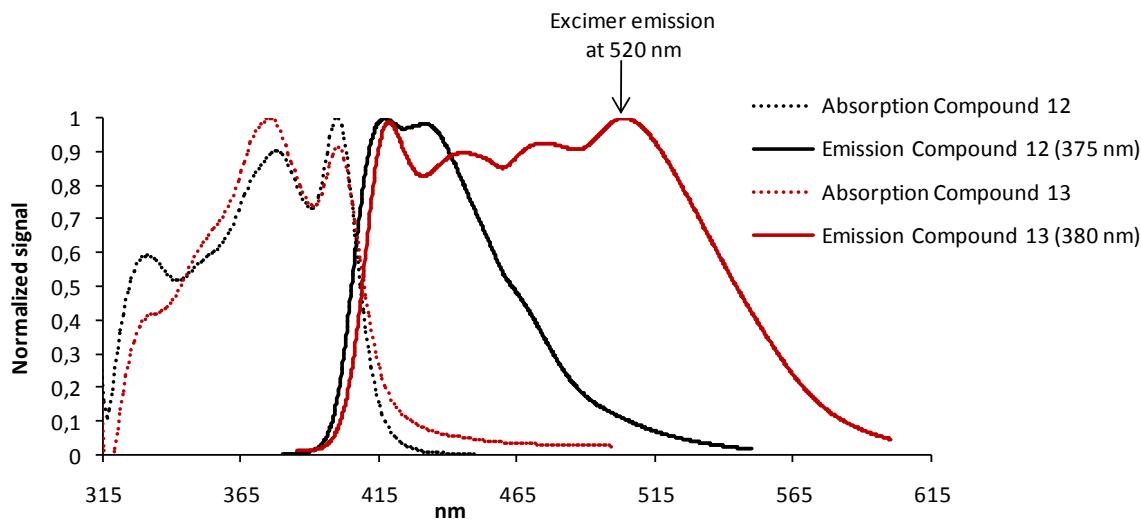


Figure S5: Normalized absorption and fluorescence spectra (375 nm) registered upon stepwise addition of AgBF_4 to a solution of **12** (measured at $2.7 \cdot 10^{-5}$ M, room temperature)

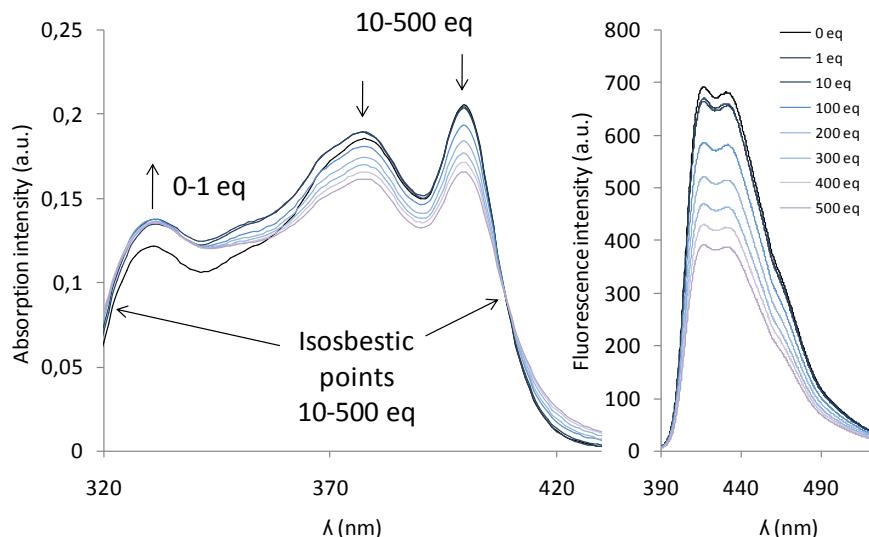
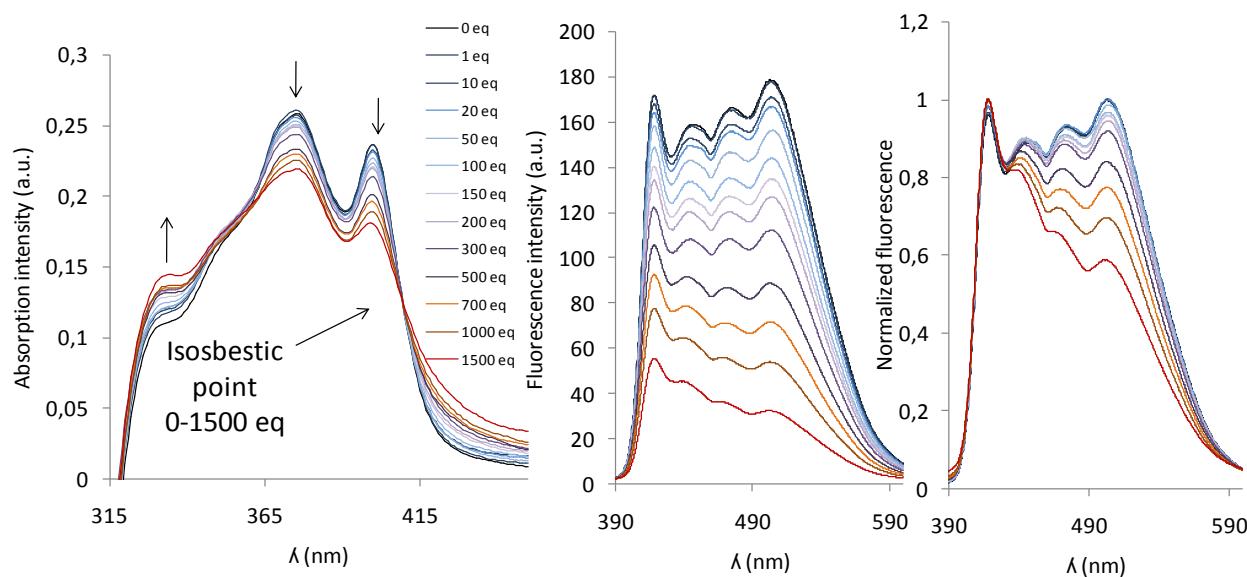


Figure S6: Normalized absorption and fluorescence spectra (380 nm) registered upon stepwise addition of AgBF_4 to a solution of **13** (measured at $4.4 \cdot 10^{-5}$ M, room temperature). Normalized fluorescence spectra (right) highlight a significant ratiometric change at the longer wavelength bands along the titration of **13** with Ag(I).



Fluorescence quantum yield determinations

The fluorescence quantum yields Φ were computed according to the following expression:

$$\Phi_S = \Phi_R \times \frac{F_S}{F_R} \times \frac{1 - 10^{-A_R(\lambda_{ex})}}{1 - 10^{-A_S(\lambda_{ex})}} \times \frac{n_S^2}{n_R^2}$$

where subscripts S and R refer respectively to sample and reference fluorophore (quinine sulfate, 0.1 M H₂SO₄) with known quantum yield (0.54), F stands for the *corrected*, integrated fluorescence spectra, A(λ_{ex}) denotes the absorbance at the used excitation wavelength λ_{ex} and n represents the refractive index of the solvent.

To minimize inner filter effects, the absorbance at the excitation wavelength λ_{ex} was kept under 0.1. All measurements were done on non-degassed samples at room temperature. The averages and standard uncertainties of Φ are computed from independent Φ measurements (two different concentrations of sample \times two different concentrations of reference \times two different excitation wavelengths λ_{ex}).

Time-resolved fluorescence measurements (Photokinetic study)

Fluorescence decay traces were recorded in the single photon timing (SPT) mode using a FluoTime 200 fluorometer (PicoQuant, GmbH). Briefly, the samples were excited by a 405 nm Picosecond Pulsed Diode Laser (Edinburgh EPL405) with a 10 MHz repetition rate. The full width at half maximum of the laser pulse was ~ 90 ps. The fluorescence was collected after crossing through a polarizer set at the magic angle, 2-mm slits and a 2 nm bandwidth monochromator. Fluorescence decay histograms were collected using a TimeHarp200 board, with a time increment per channel of 36 ps, at three or more different wavelengths. The histograms of the instrument response function (IRF) was determined using LUDOX scatterer, and sample decays were recorded until they typically reached 2×10^4 counts in the peak channel. Time-resolved emission spectroscopy (TRES) was performed using the same configuration; collecting fluorescence decay traces every 2 nm (from 420 to 510 nm) during a fixed amount of time to maintain the overall intensity information.

Time resolved fluorescence decay traces were deconvoluted from the signal and fitted using FluoFit 4.4 package (Picoquant GmbH). In all cases, fluorescence decay traces were recorded at different emission wavelengths, and all decay traces were fitted globally with the decay times linked as shared parameters, whereas the pre-exponential factors were local adjustable parameters. The experimental decay traces were fitted to multi-exponential functions via a Levenberg-Marquardt algorithm-based nonlinear least-squares error minimization deconvolution method. The quality of fittings was judged by the reduced chi-squared method, χ^2 , the weighted residuals and the correlation functions. The latter two were checked for random distributions.

For the TRES analysis and the estimation of the species-associated emission spectra (SAEMS), the same fitting procedure was performed. Then, SAEMS of each species i at any given emission wavelength (SAEMS $_{(i)}(\lambda_{em})$) is given by the fluorescence

intensity emitted by the species i ($A_{i,\lambda em} \times \tau_i$), normalized by the total intensity and corrected for the different detection sensitivity using the total intensity of the steady-state spectrum ($I_{ss,\lambda em}$):

$$SAEMS_i(\lambda_{em}) = \frac{A_{i,\lambda em} \times \tau_i}{\sum_i A_{i,\lambda em} \times \tau_i} \cdot I_{ss,\lambda em}$$

With the SAEMS, the approximate contribution of each species to the total spectrum is estimated. However, this is just an approximation because the promoted amount of each of the species to the excited state is unknown, as the molar absorption coefficients of each form are not separately accessible. Therefore, the approximated contribution of each form, assumes equal excitation rate for all the species.

Figure S7: Fluorescence decays registered at 420 nm for **13**

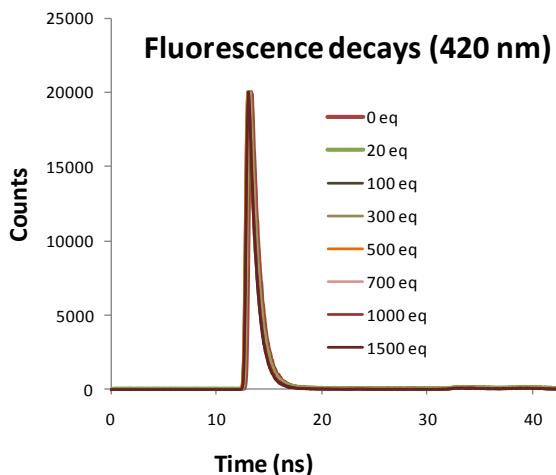
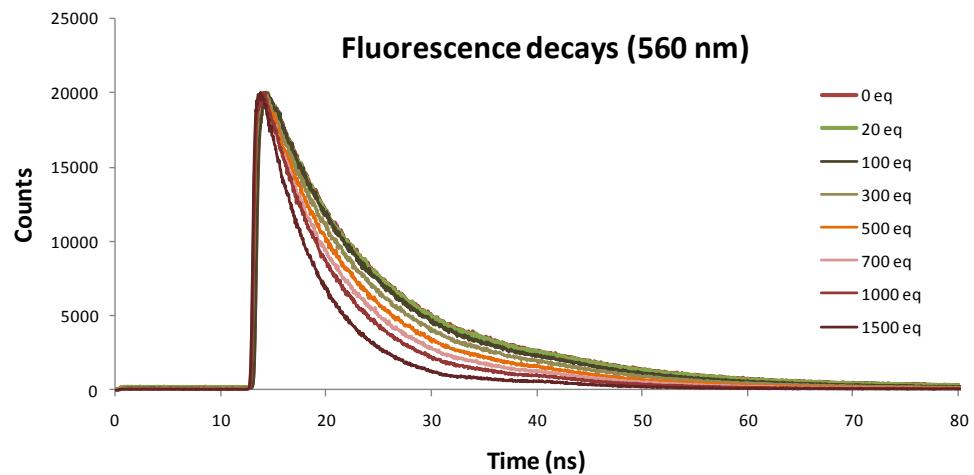
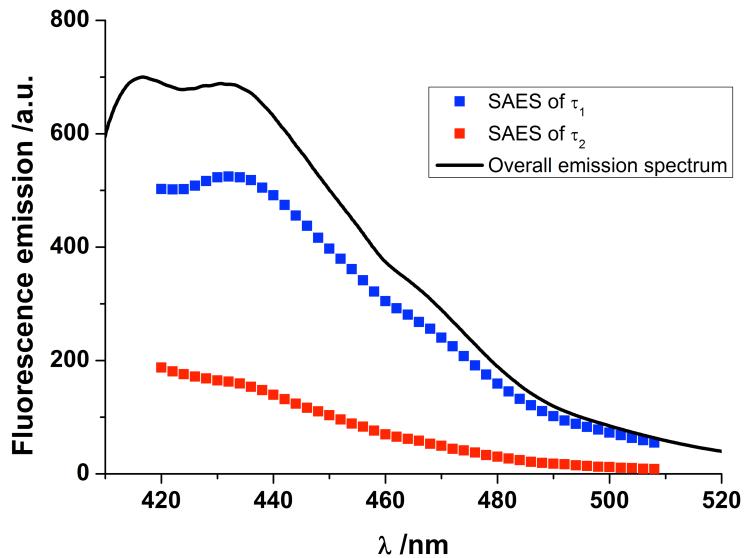


Figure S8: Fluorescence decays registered at 560 nm for **13**



Time-resolved emission spectroscopy (TRES) of compound **12** showed the presence of two different emissive species. The species-associated emission spectra (SAEMS) provide the spectral profile of each of the forms associated to each one of the decay times. The longest lifetime, τ_1 , (blue dots) shows a higher emission density in the spectral shoulder at 440 nm, whereas the short decay time, τ_2 , (red dots) is more prominent at the blue emission wavelengths (peak at 420 nm). This spectral distribution suggests that the longest lifetime, τ_1 , corresponds to the folded species as the π -stacking causes a bathochromic shift. Comparing the SAEMS with the overall steady-state emission spectra (black line) allowed to estimate an approximate 70%:30% ratio for the folded:unfolded forms. Note that this estimation takes into account the different emission efficiency of each form, as their quantum yield is proportional to the lifetime, but it does not take into account the different excitation efficiencies, as the individual extinction coefficients are not known. The estimation assumes equal, at least very similar, extinction coefficients of the two species at the excitation wavelength used.

Figure S9. SAEMS decomposition of **12**



Estimation of the excited-state folding kinetic rate

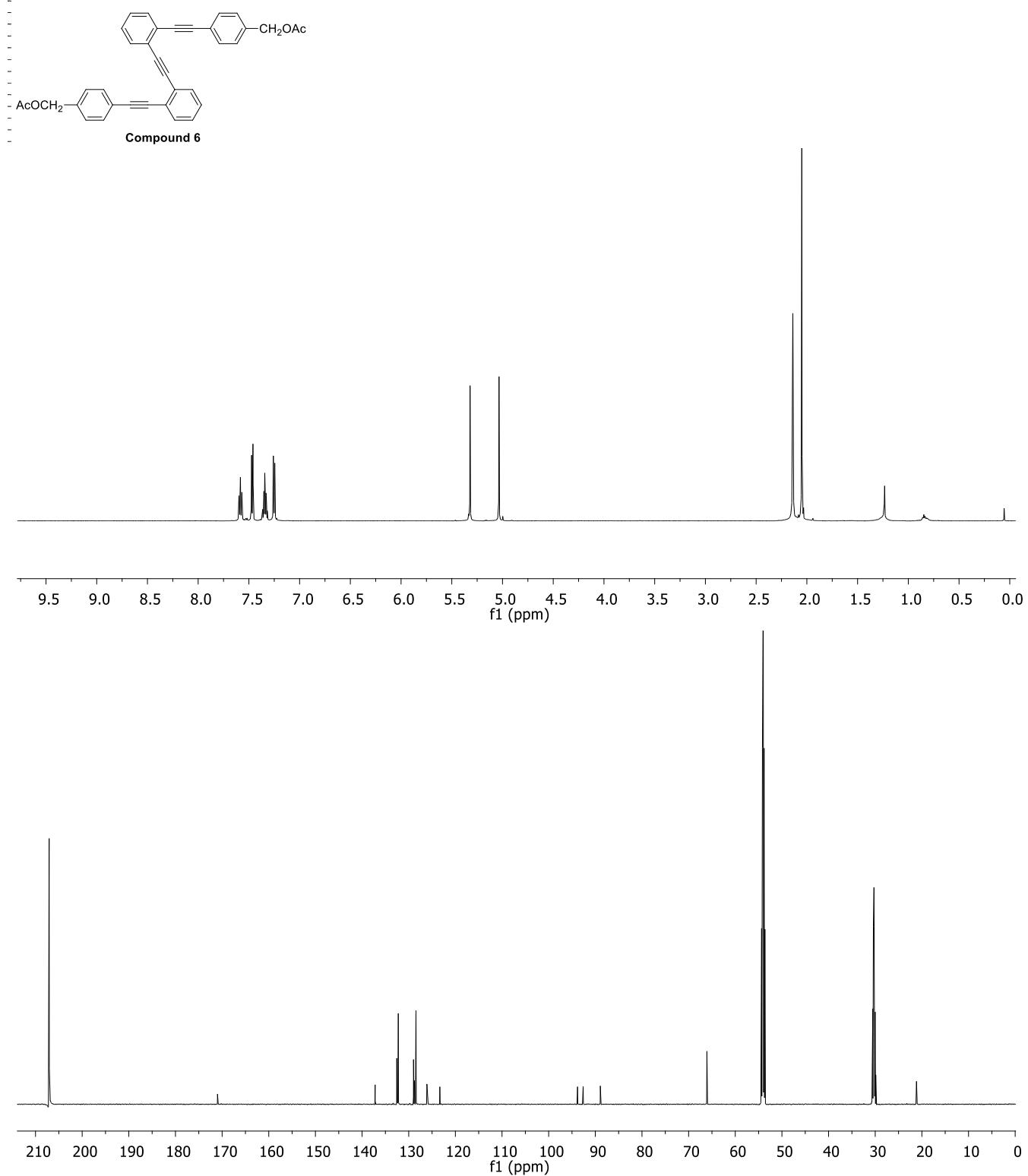
The kinetics of the excited-state folding reaction can be accessible through the fluorescence decay times. Indeed, the rate of the pyrene excimer formation, k_f , can be directly correlated with the excited-state folding kinetic constant. This rate constant can be obtained via the shortest decay time, τ_3 , by solving the kinetic differential equations:³⁹ $k_f = 1/\tau_3 - 1/\tau_{ref}$, where τ_{ref} is the fluorescence of a reference pyrene monomer, in the absence of excited-state reaction. We used **12** as the reference, because of its structural similarity with **13**, but lacking the second pyrene moiety, so the intramolecular excimer formation is avoided. The average fluorescence lifetime τ_{ref} was 2.36 ± 0.09 ns. In the absence of Ag(I), the value of k_f is $(9.8 \pm 0.3) \times 10^8$ s⁻¹, and represents one of the few examples of experimentally determined kinetic folding constant in OPE-based foldamers.⁴⁰ At increasing Ag(I) concentrations, the value of τ_3 provides the estimation of the excited-state folding rate. As expected, the presence of Ag(I) favors the excited-state folding process, and the k_f value increases (Figure 10c).

CD titrations

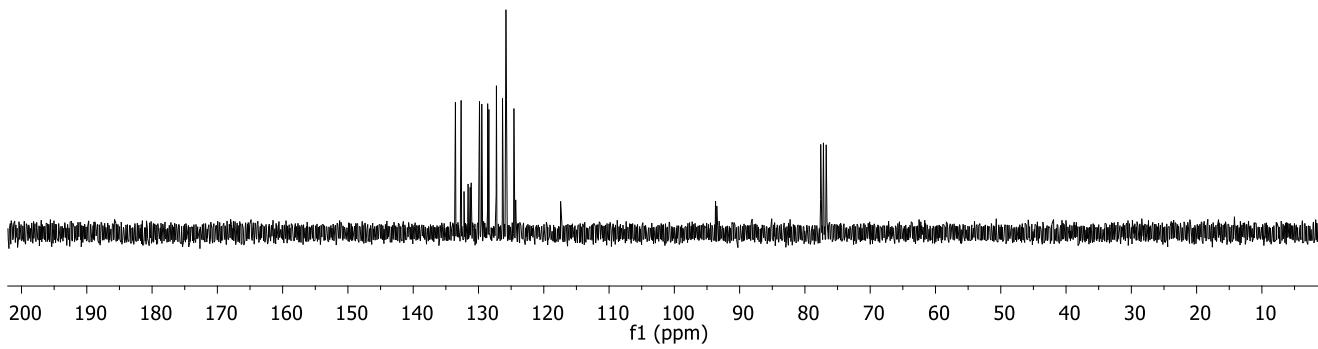
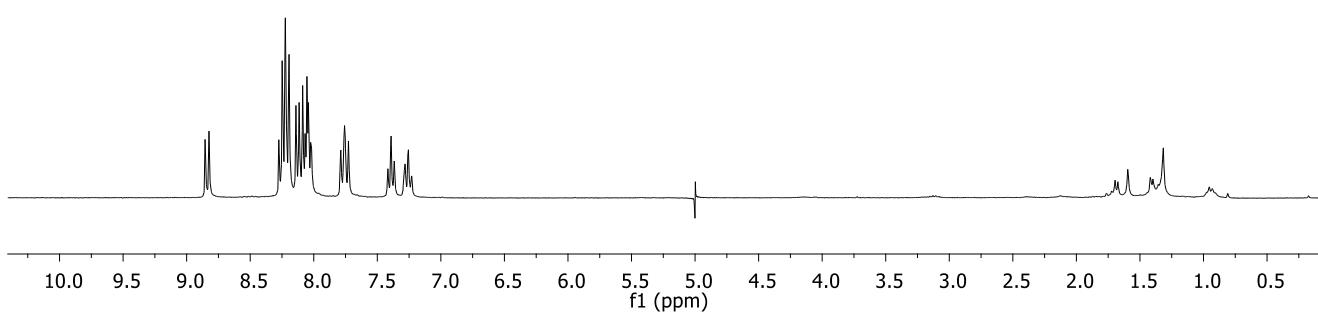
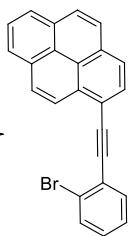
The experiments were conducted from stock solutions of the ligands (10^{-4} M) in CH₂Cl₂/Acetone (v/v, 9:1). Two or three solutions of different concentrations of the AgBF₄ were prepared in CD₂Cl₂/Acetone (v/v, 9:1). A small portion of the ligand was transferred to a cuvette and diluted with the solvent mixture to 10^{-5} - 10^{-6} M. After taking the initial spectra, small portions (from 1-100 μ L) of the silver solution were added and the spectra recorded after each addition.

¹H- and ¹³C-NMR spectra

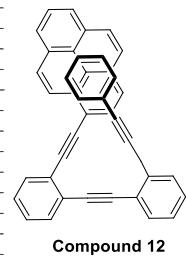
Compound 6:



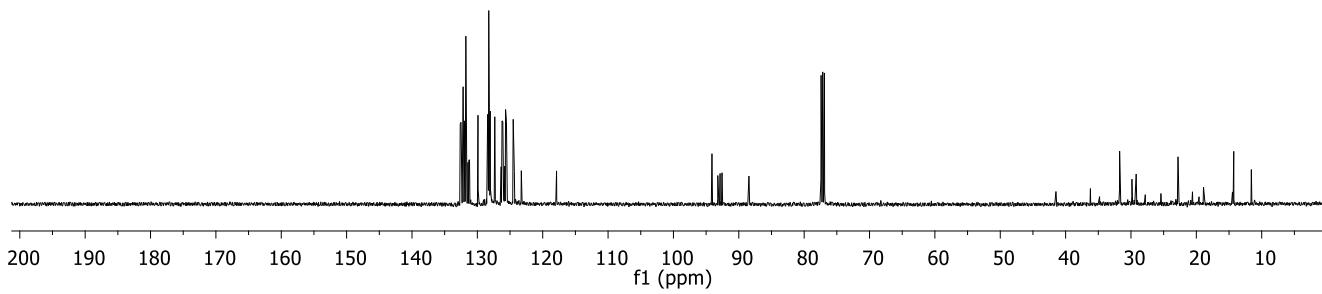
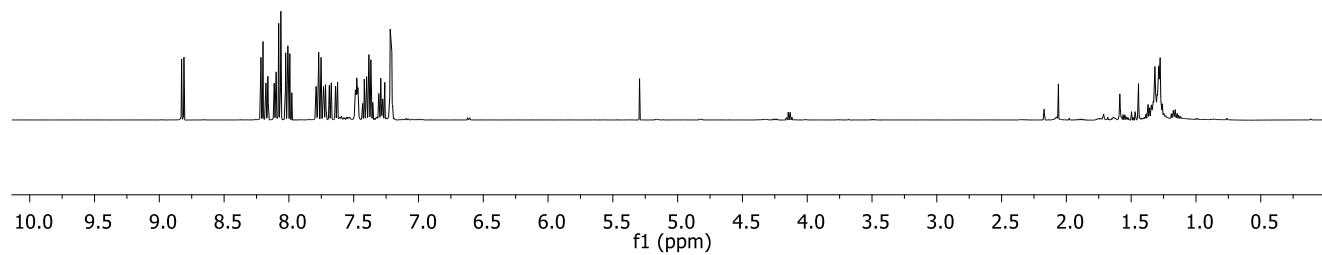
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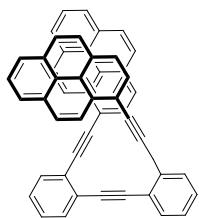
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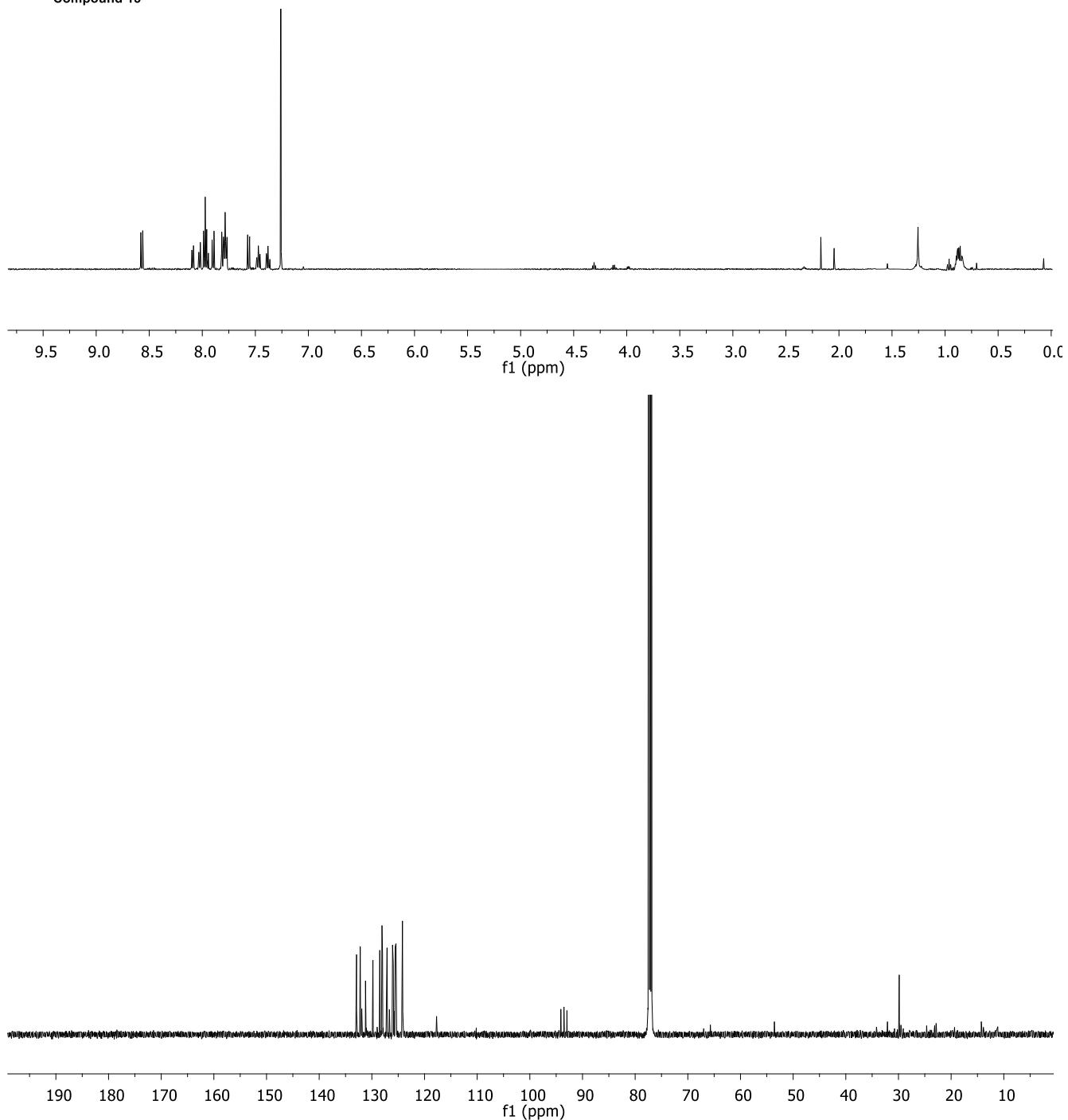
Compound 12



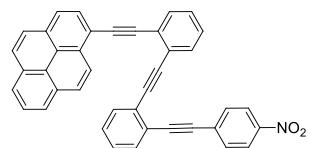
Compound 13:



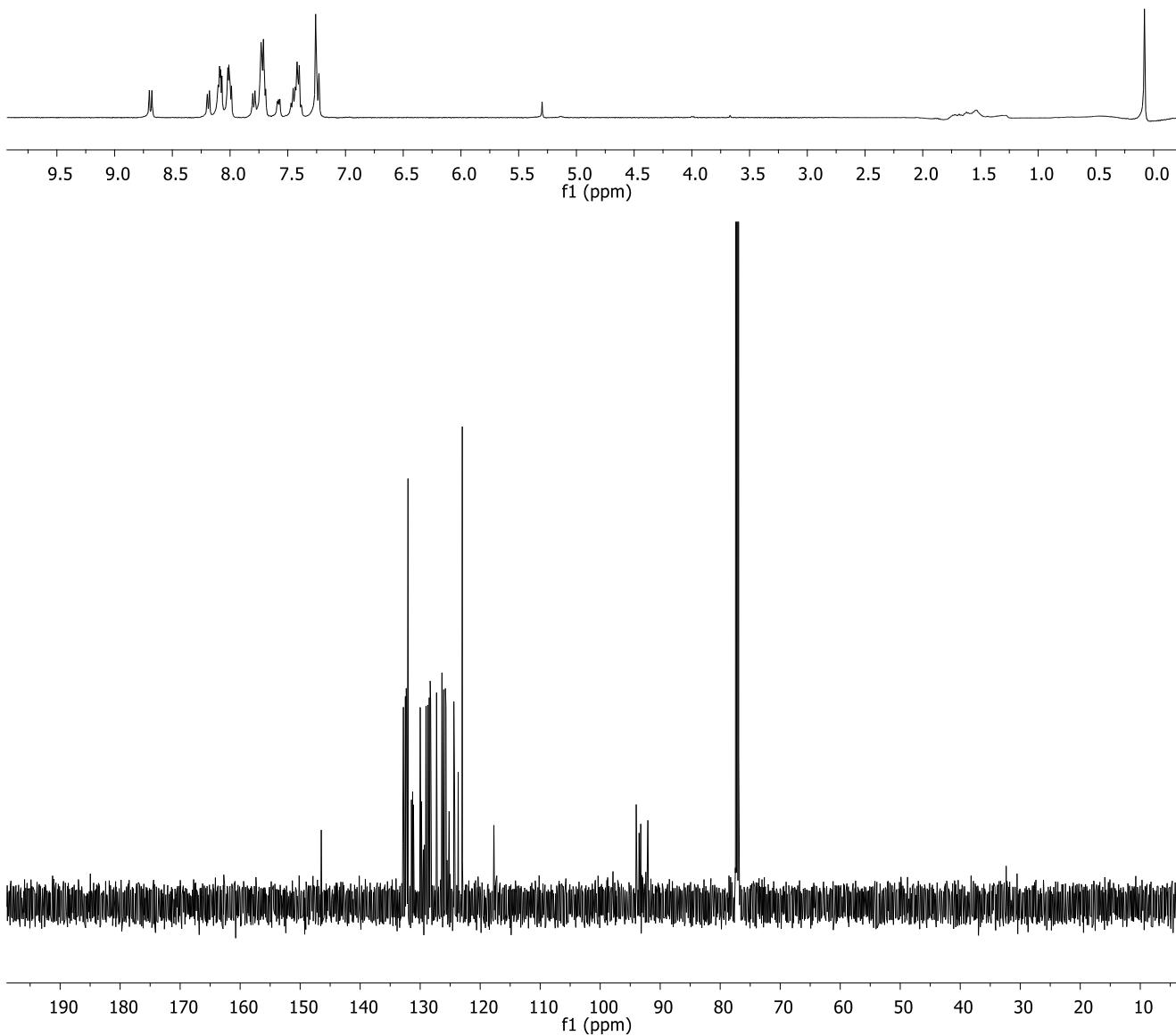
Compound 13



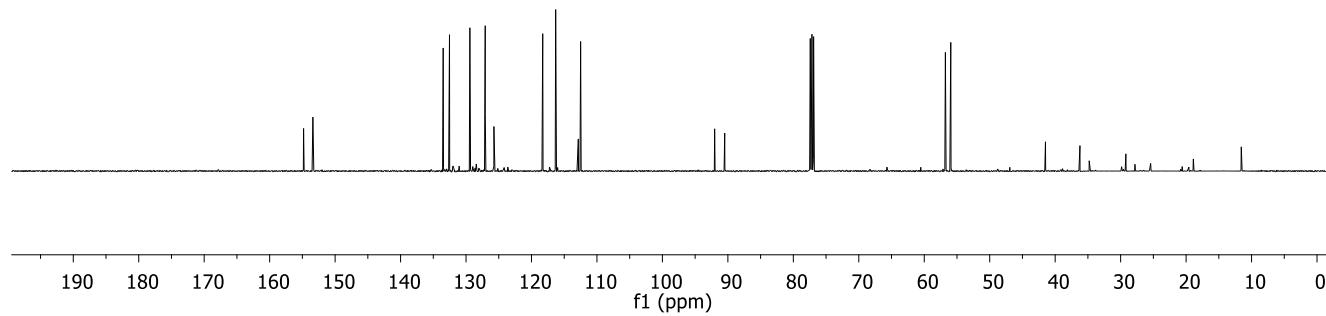
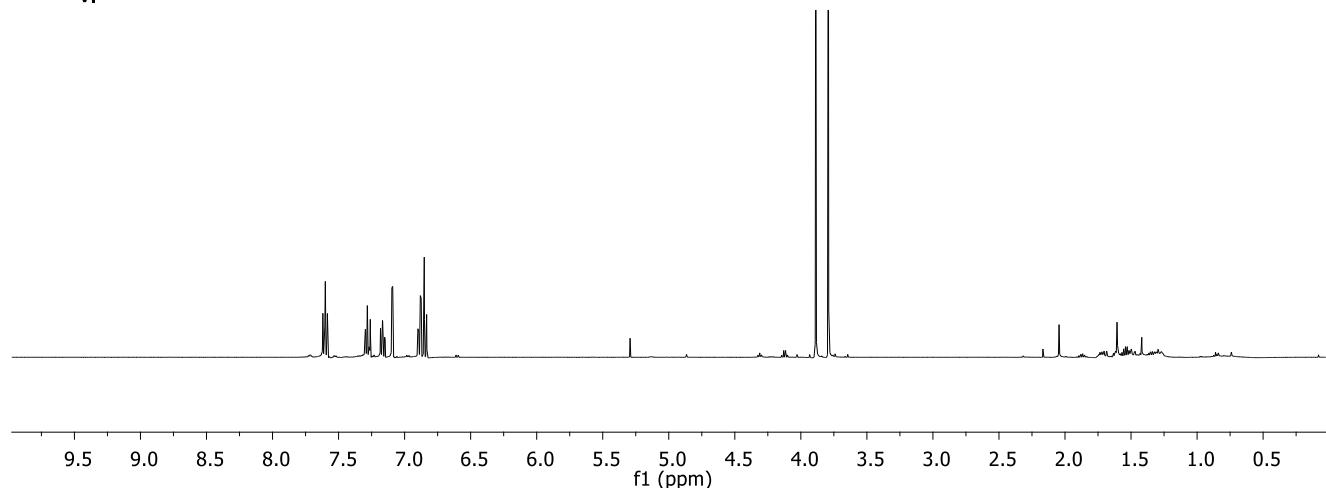
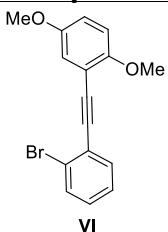
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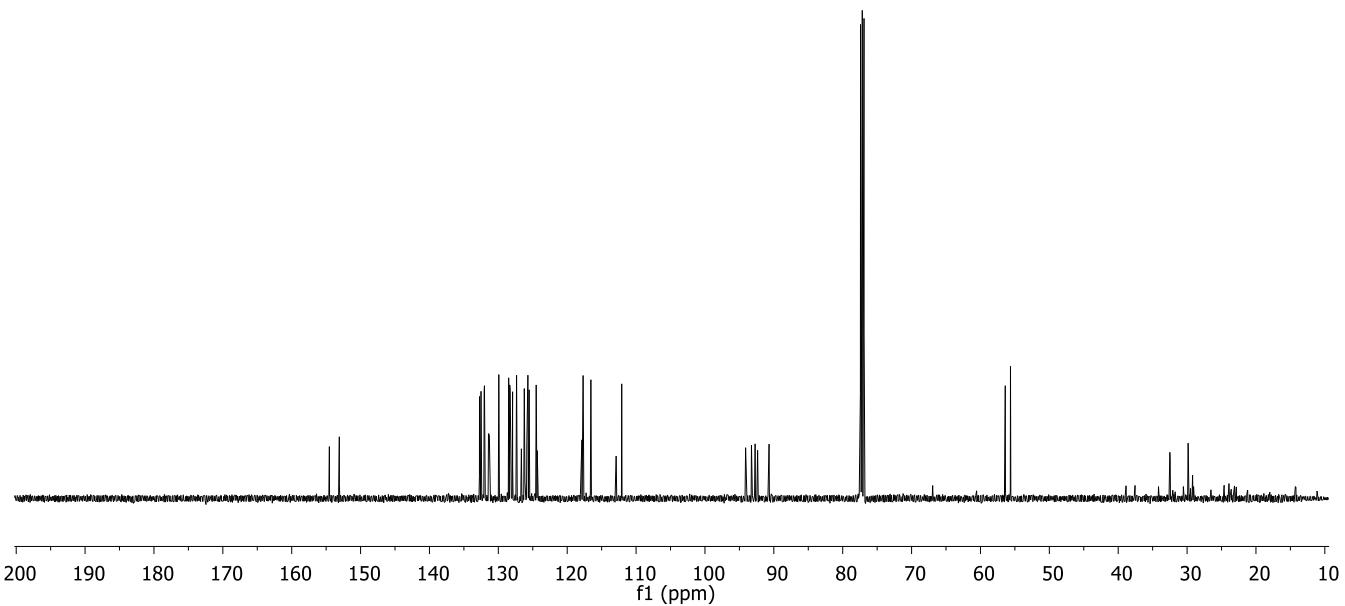
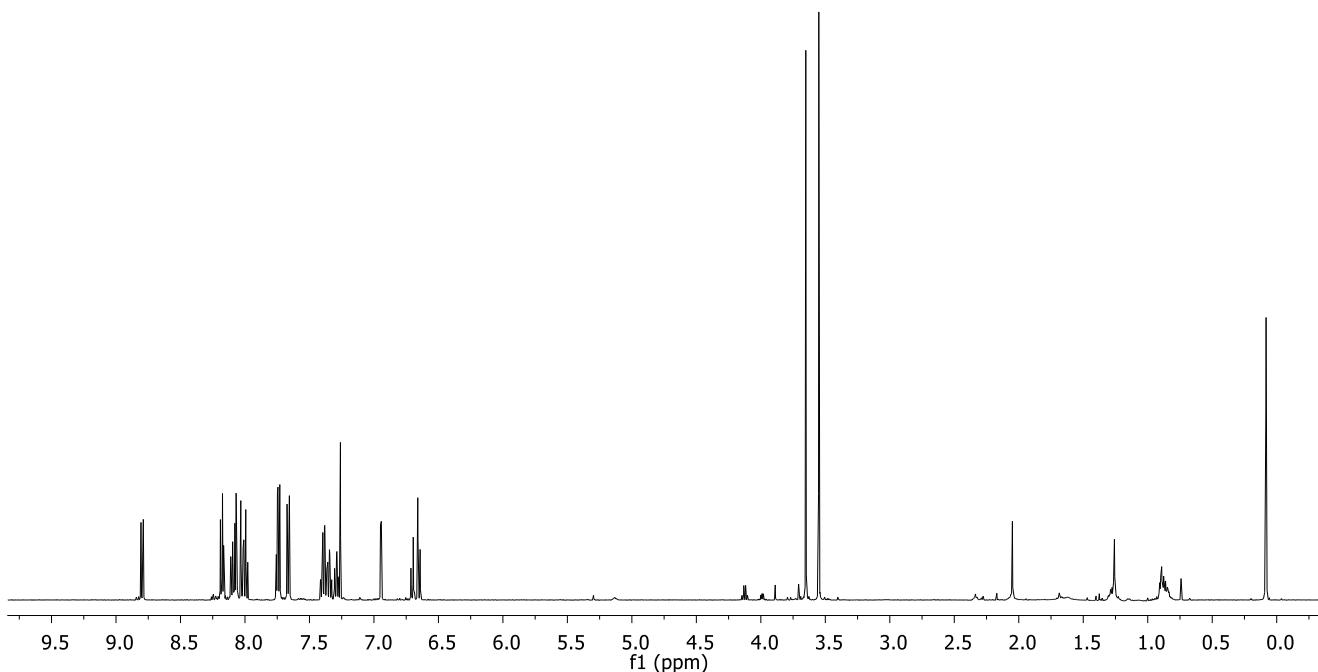
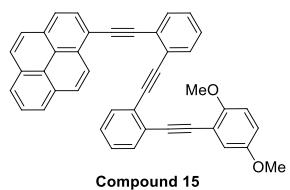
Compound 14



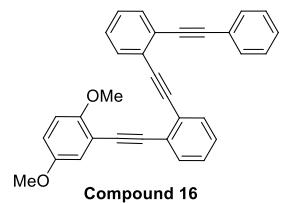
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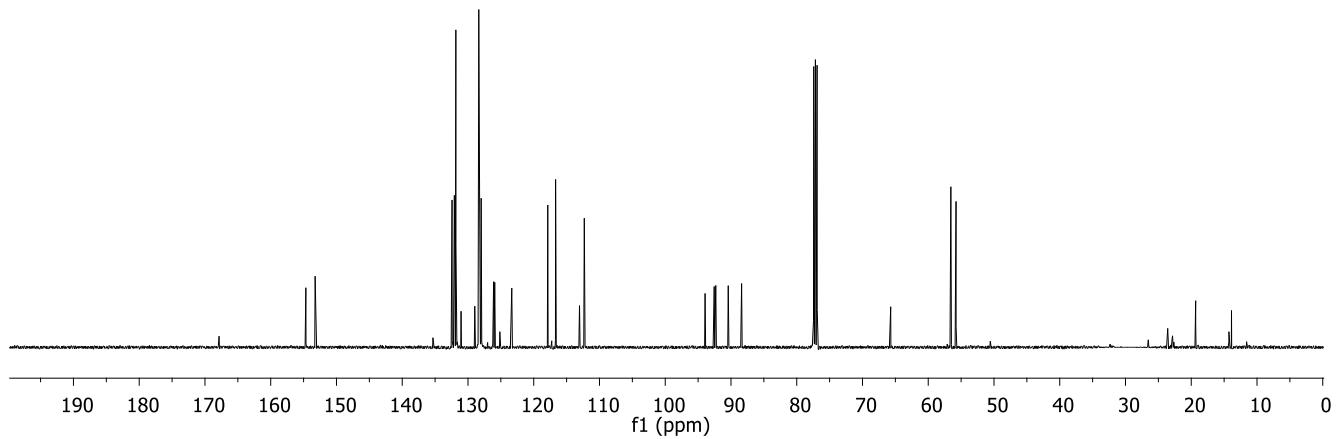
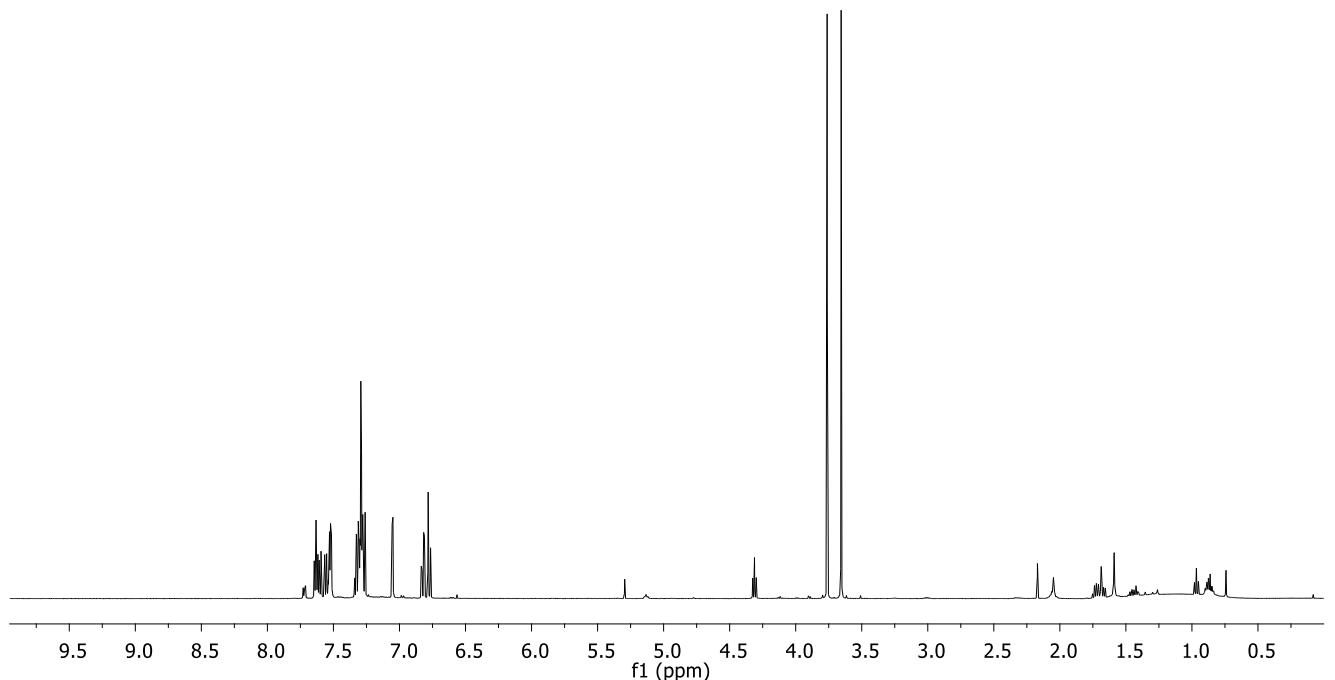
Compound 15:



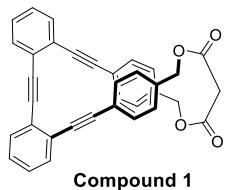
Compound 16:



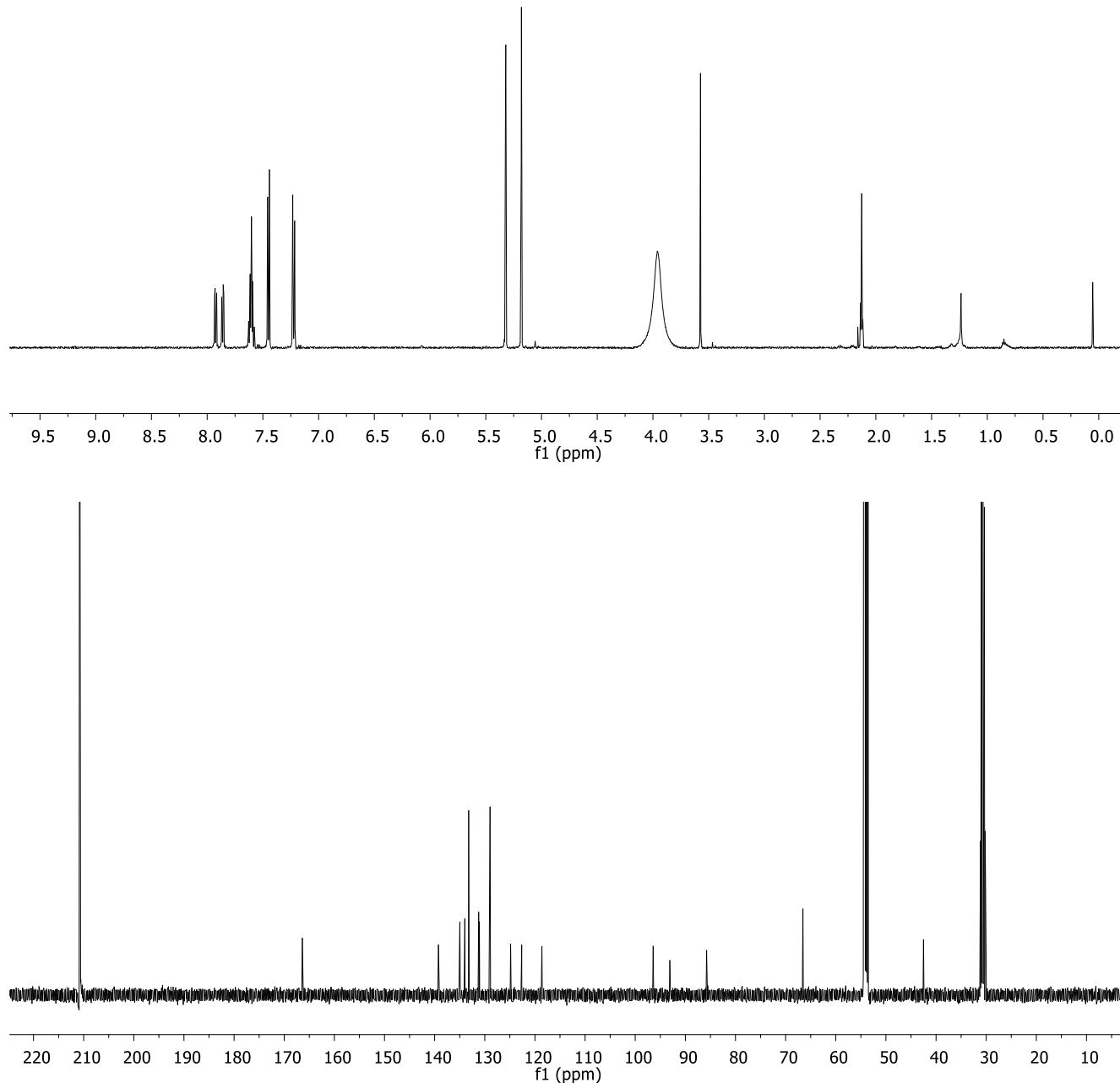
Compound 16



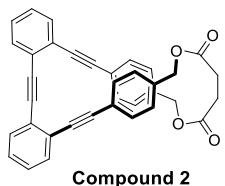
Complex 1·Ag(I):



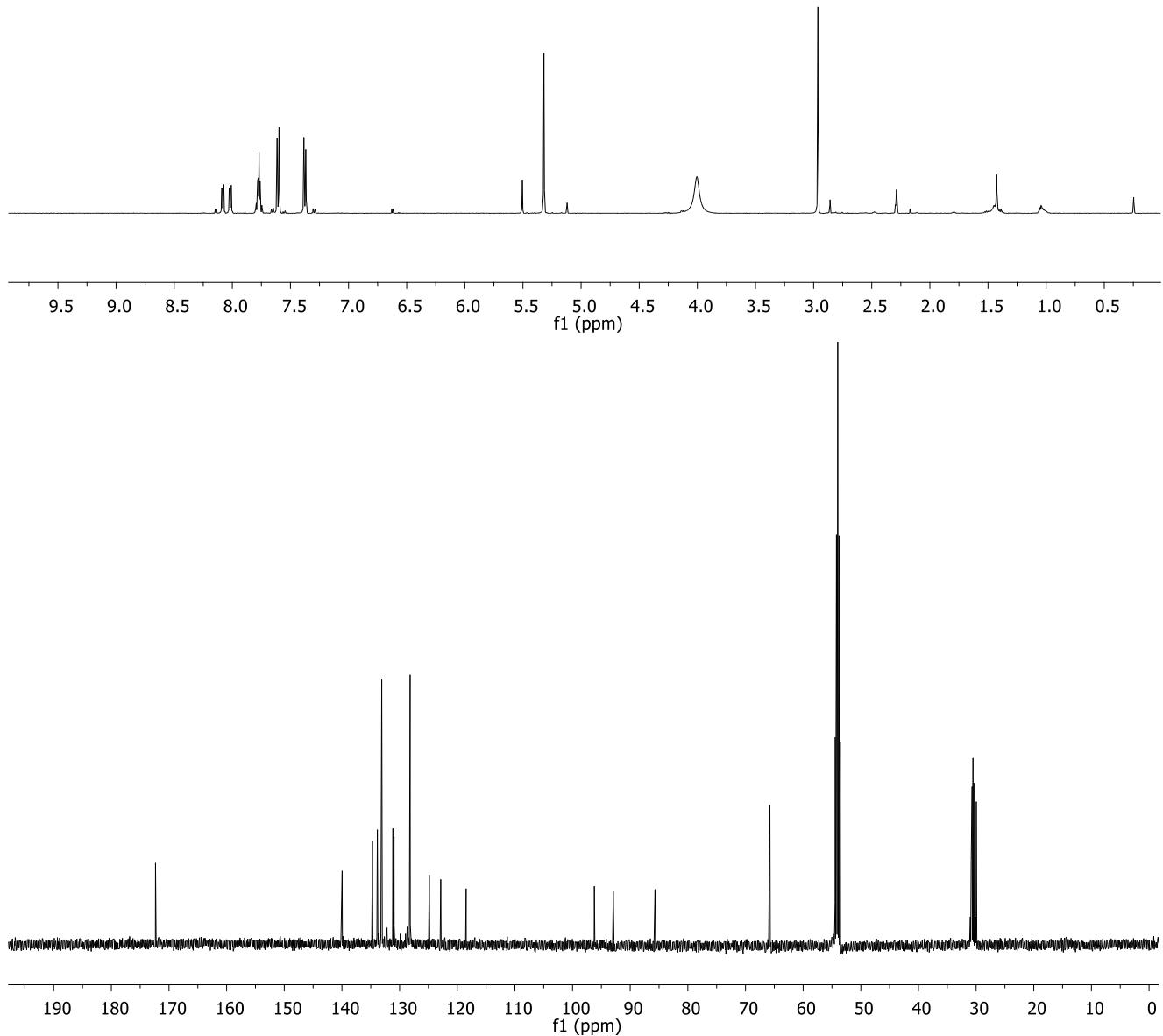
Compound 1



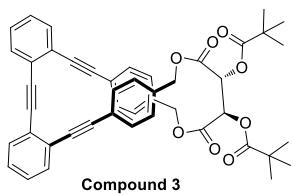
Complex 2·Ag(I):



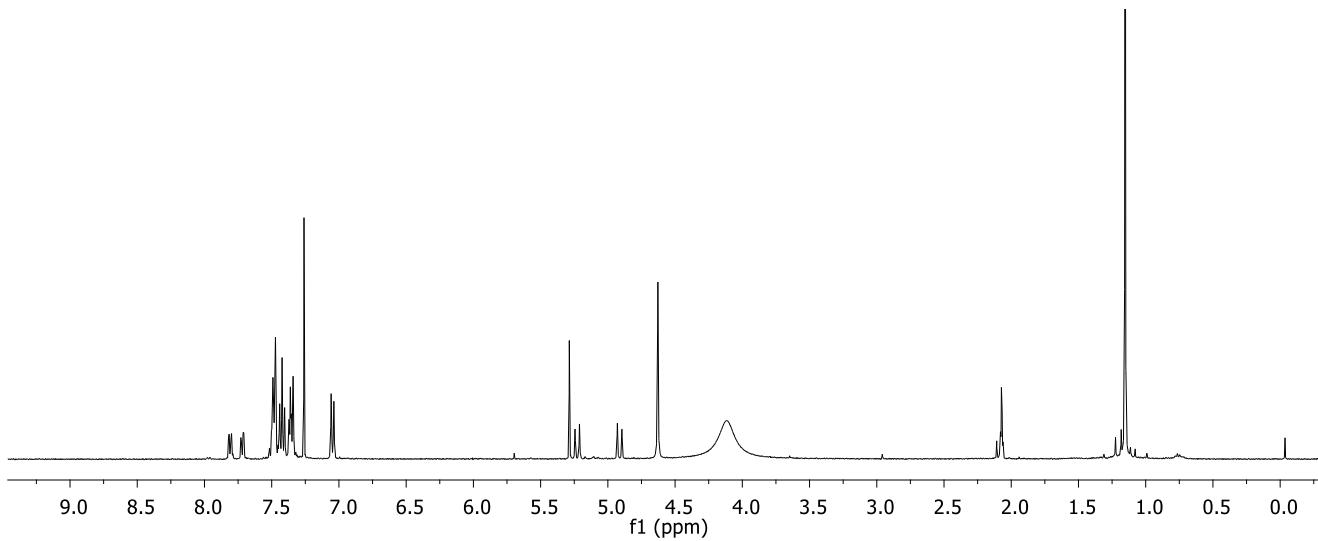
Compound 2



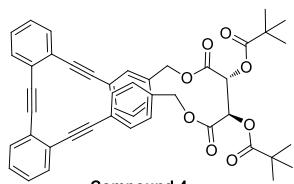
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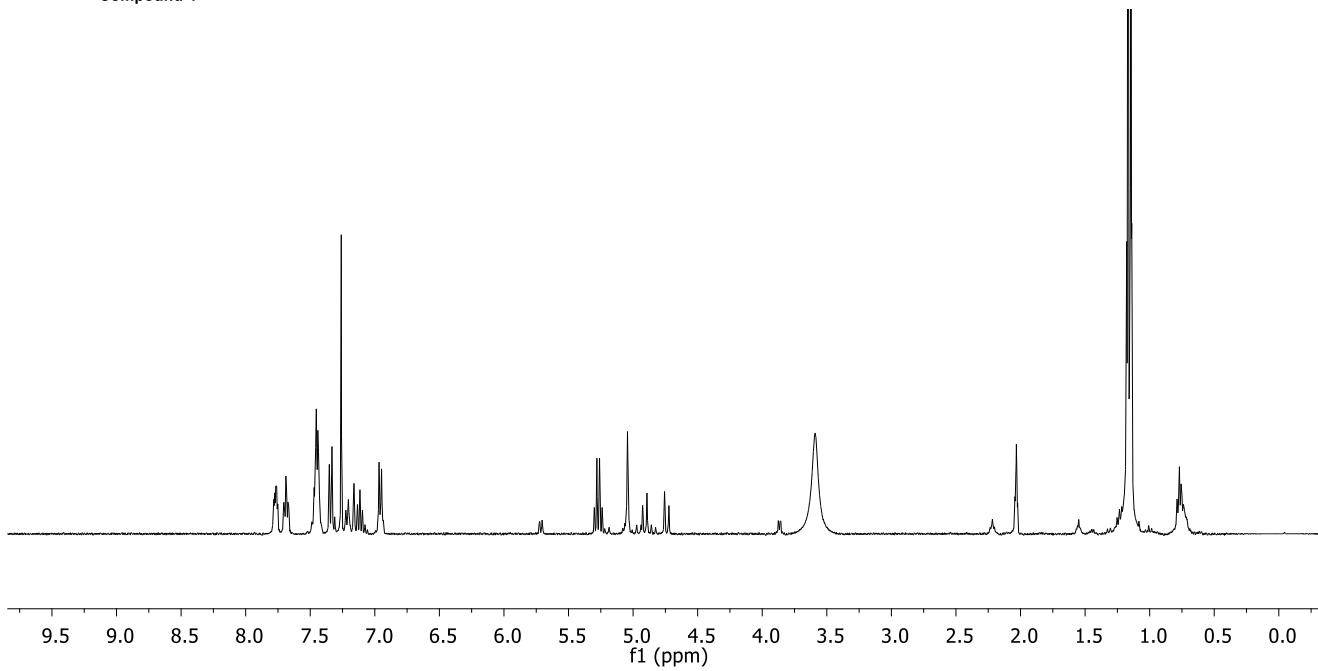
Compound 3



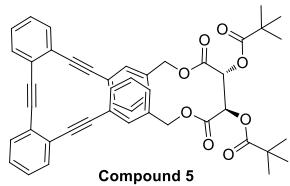
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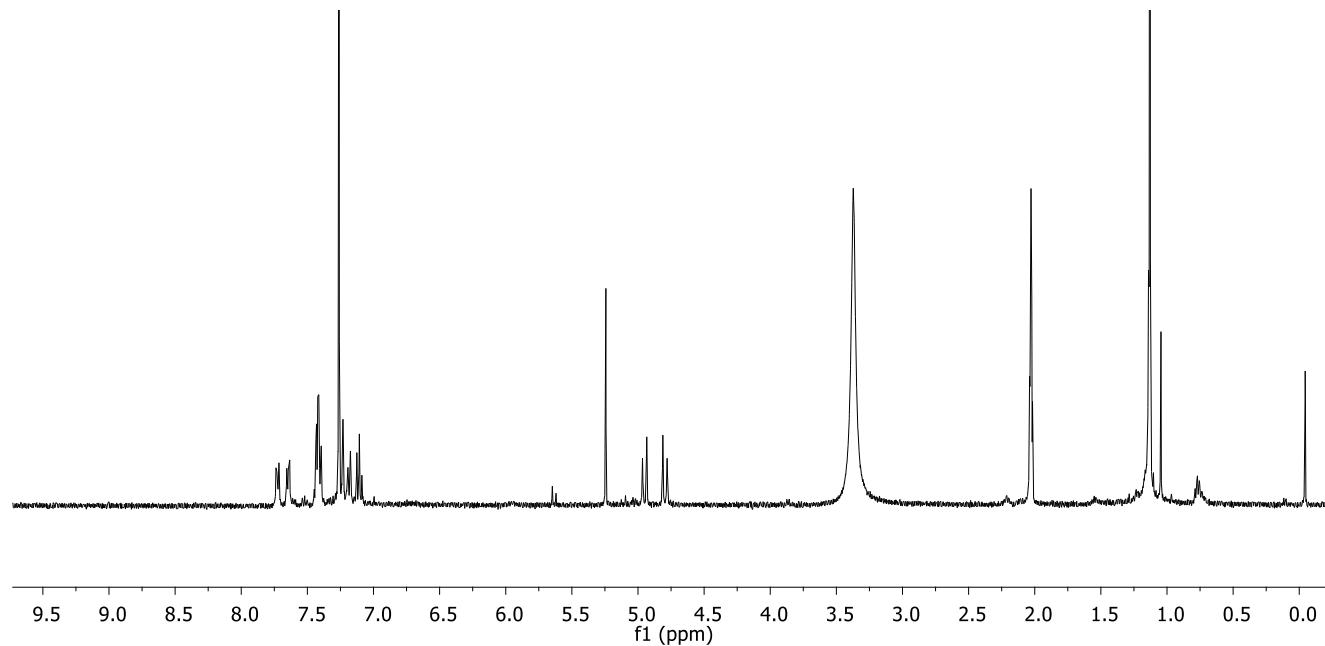
Compound 4



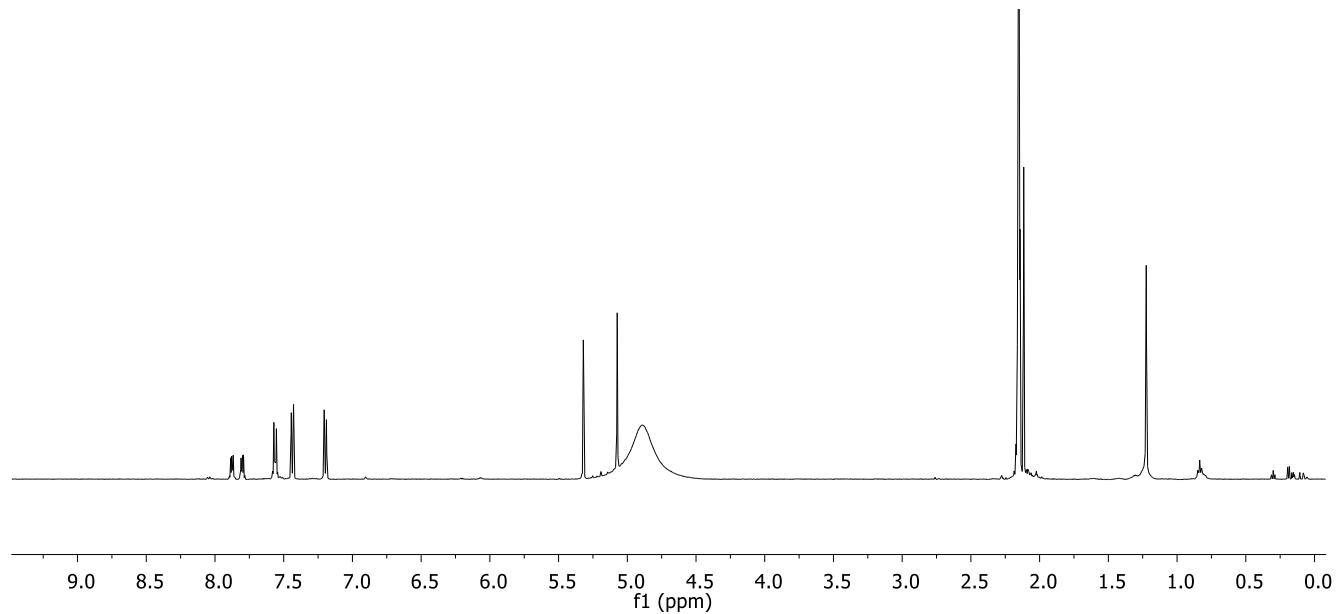
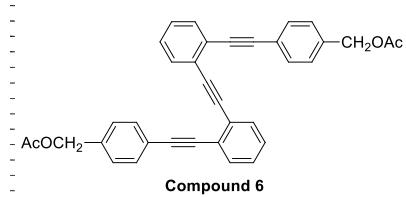
Complex 5·Ag(I):

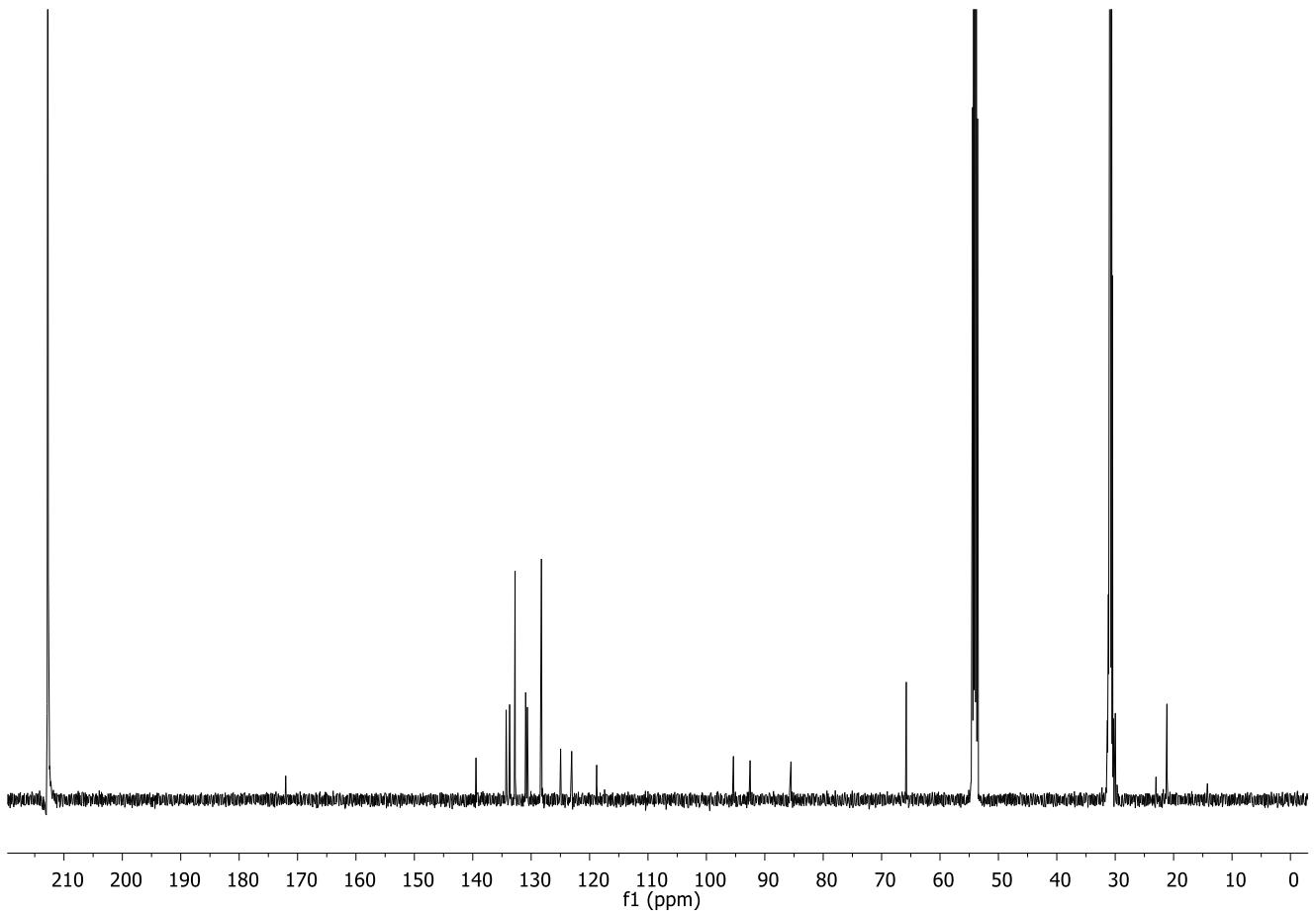


Compound 5

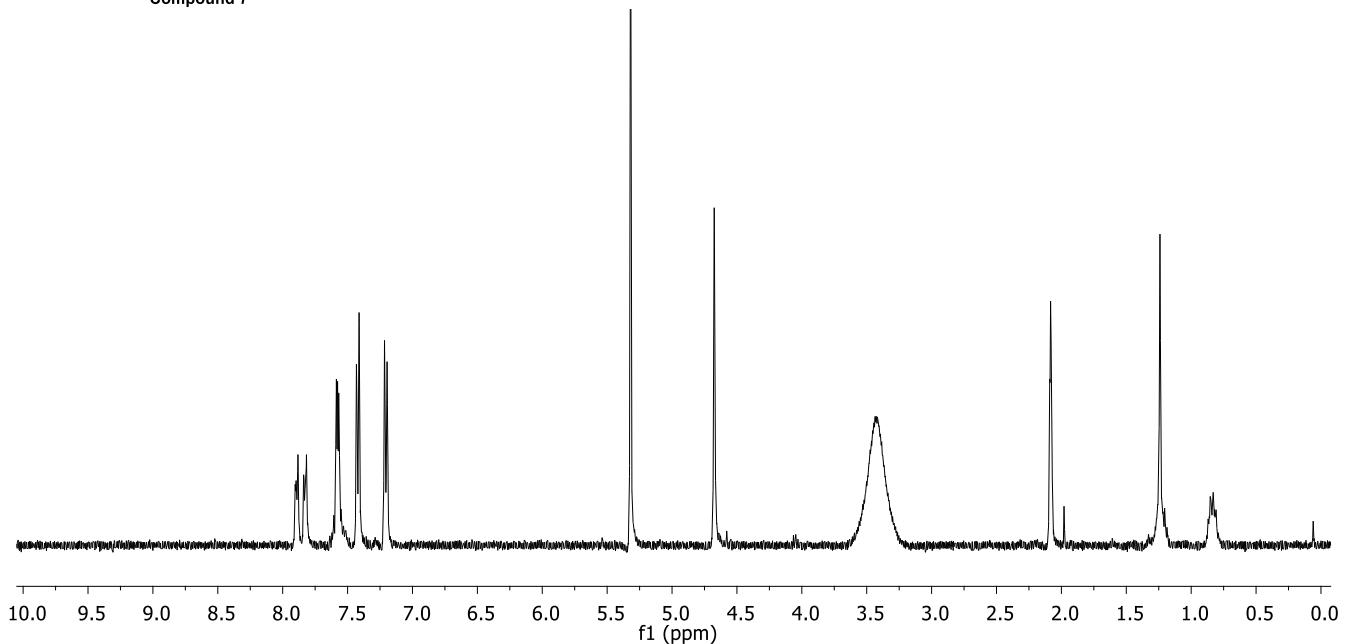
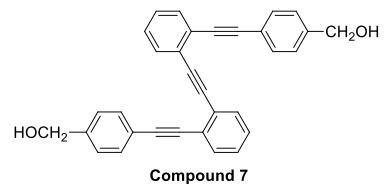


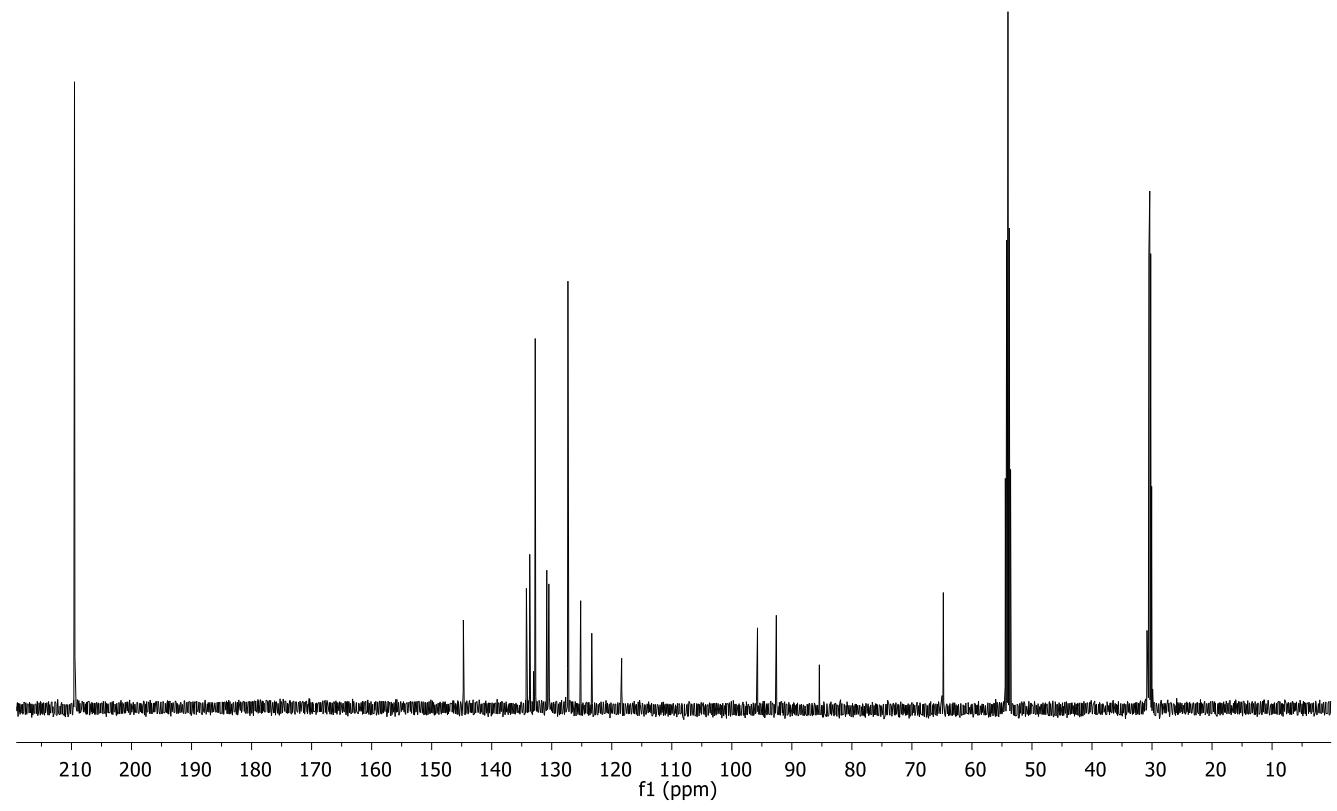
Complex 6·Ag(I):



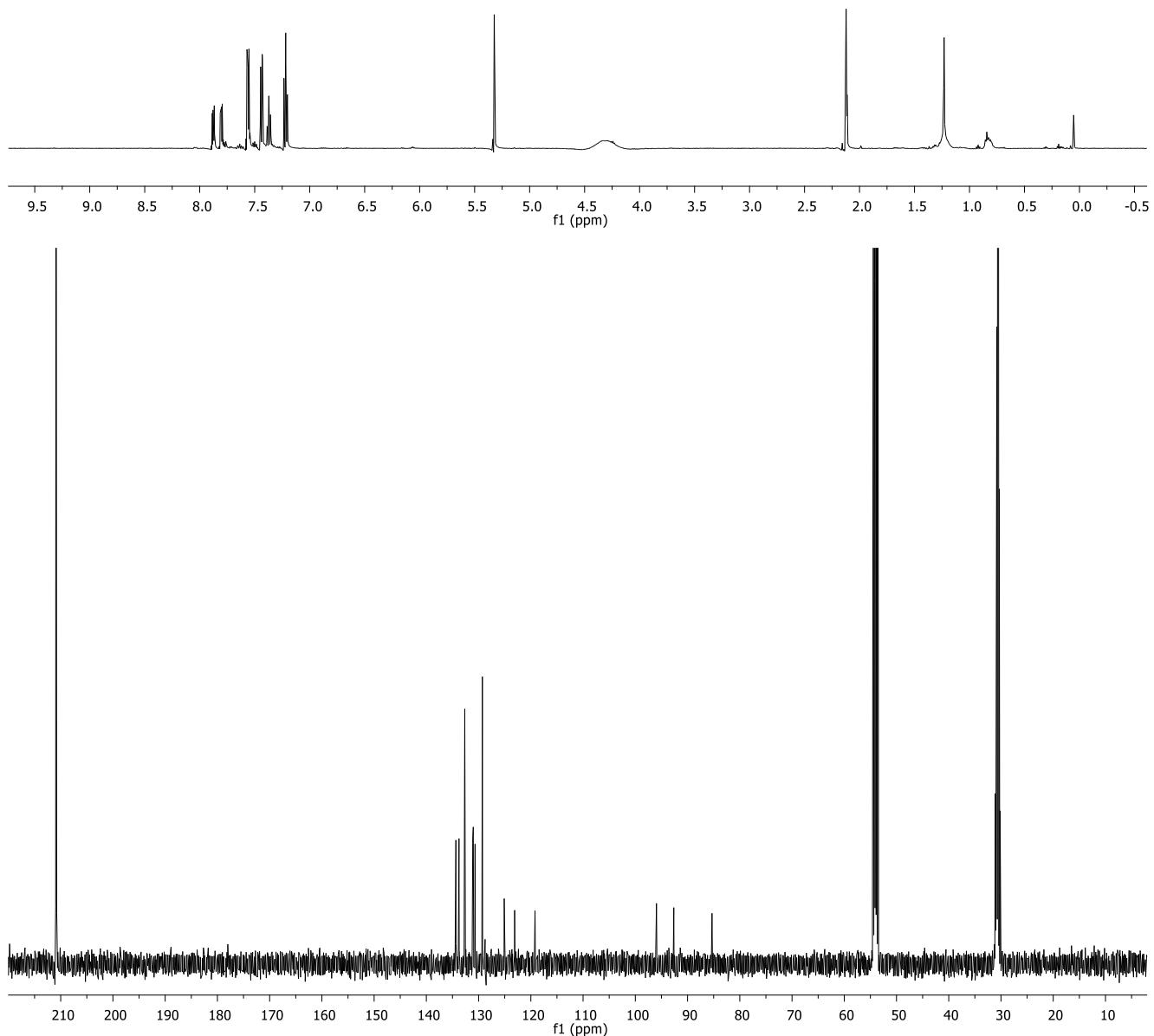
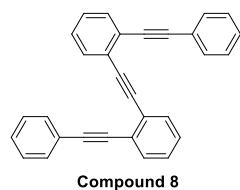


Complex 7·Ag(I):

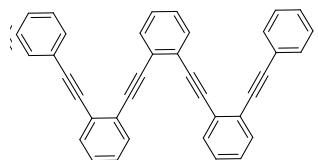




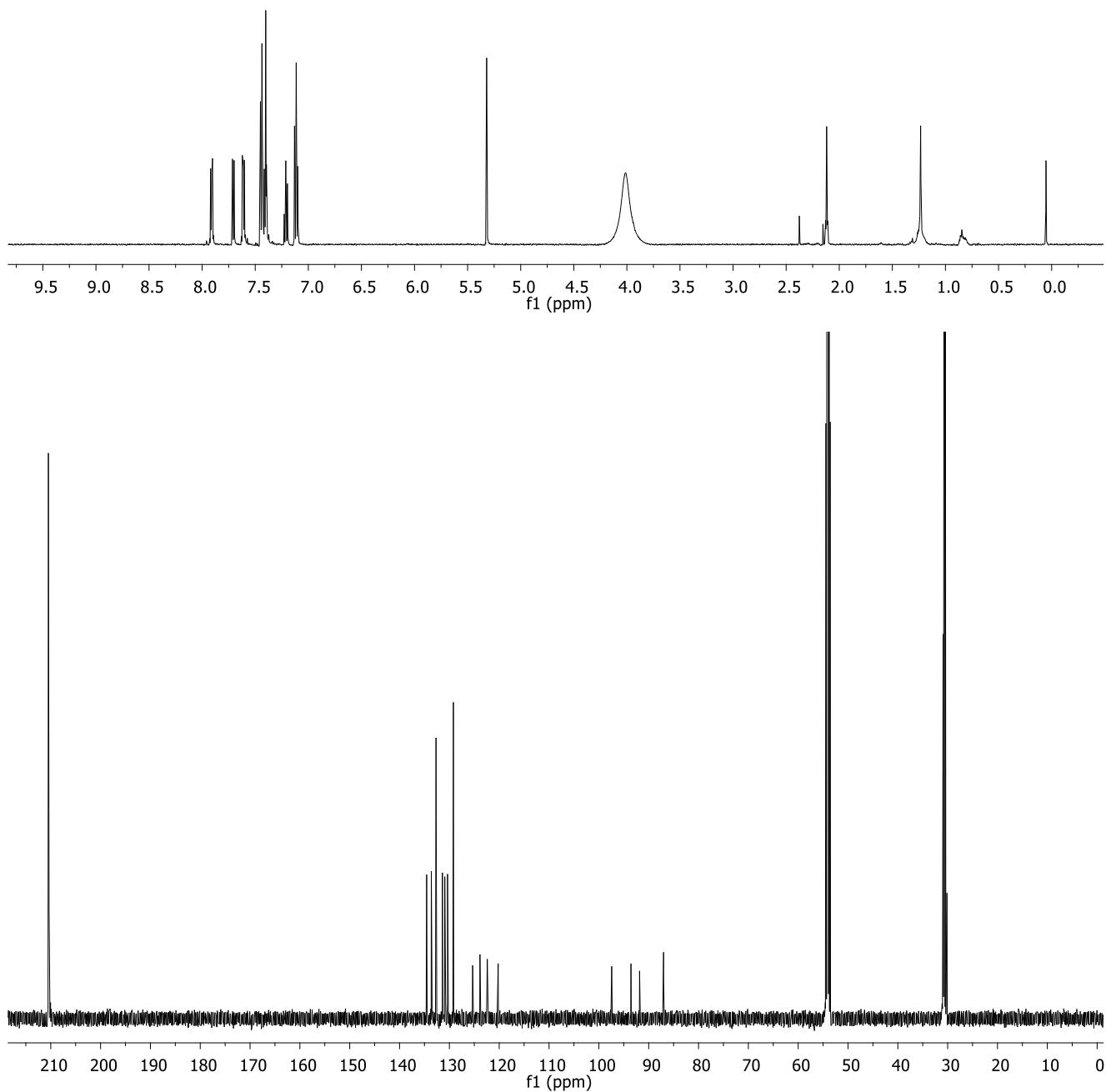
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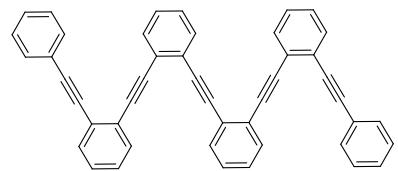
Complex 9·Ag(I):



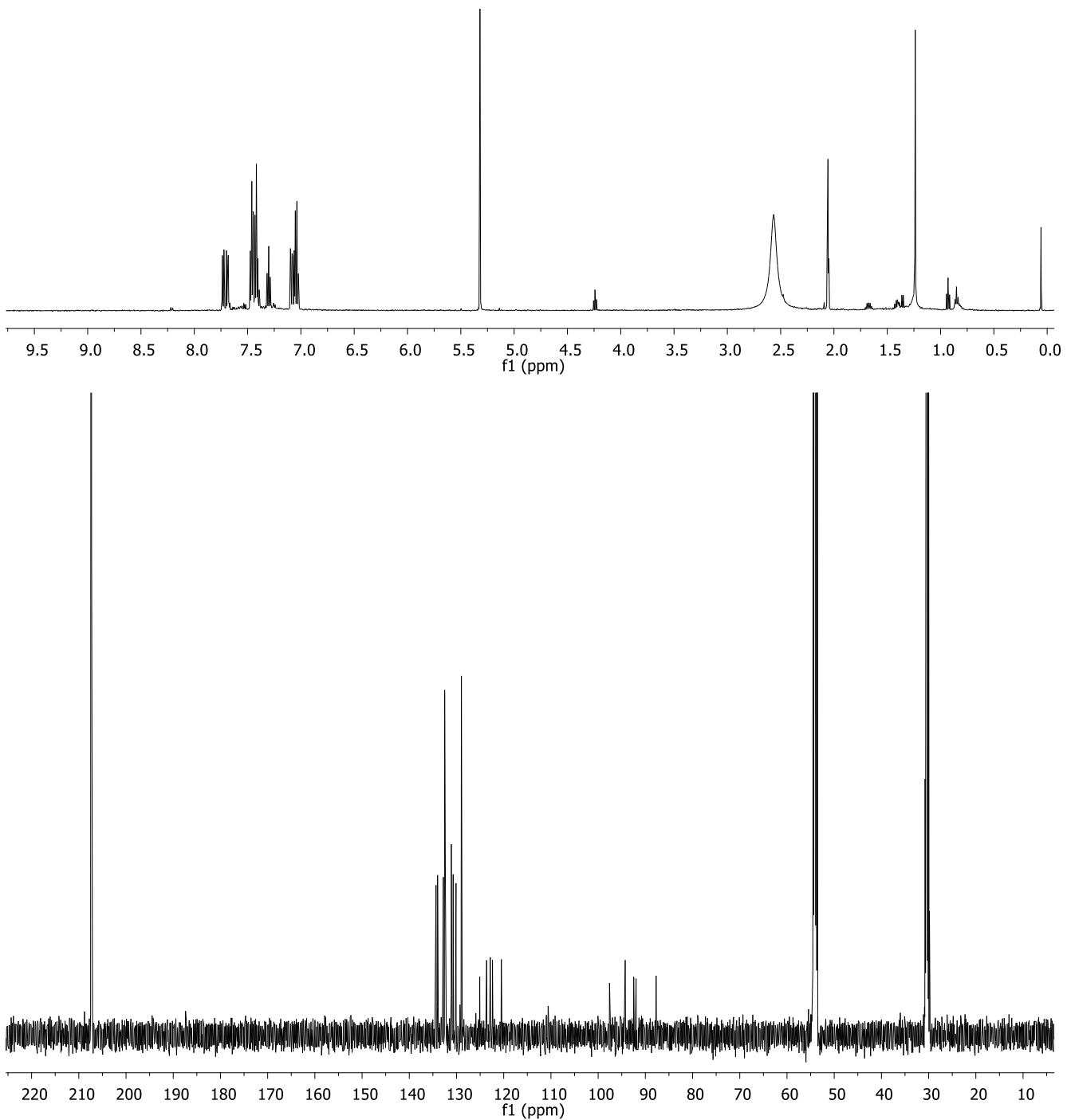
Compound 9



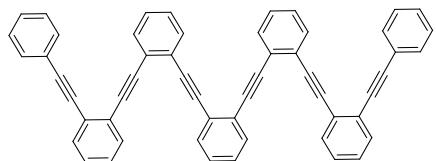
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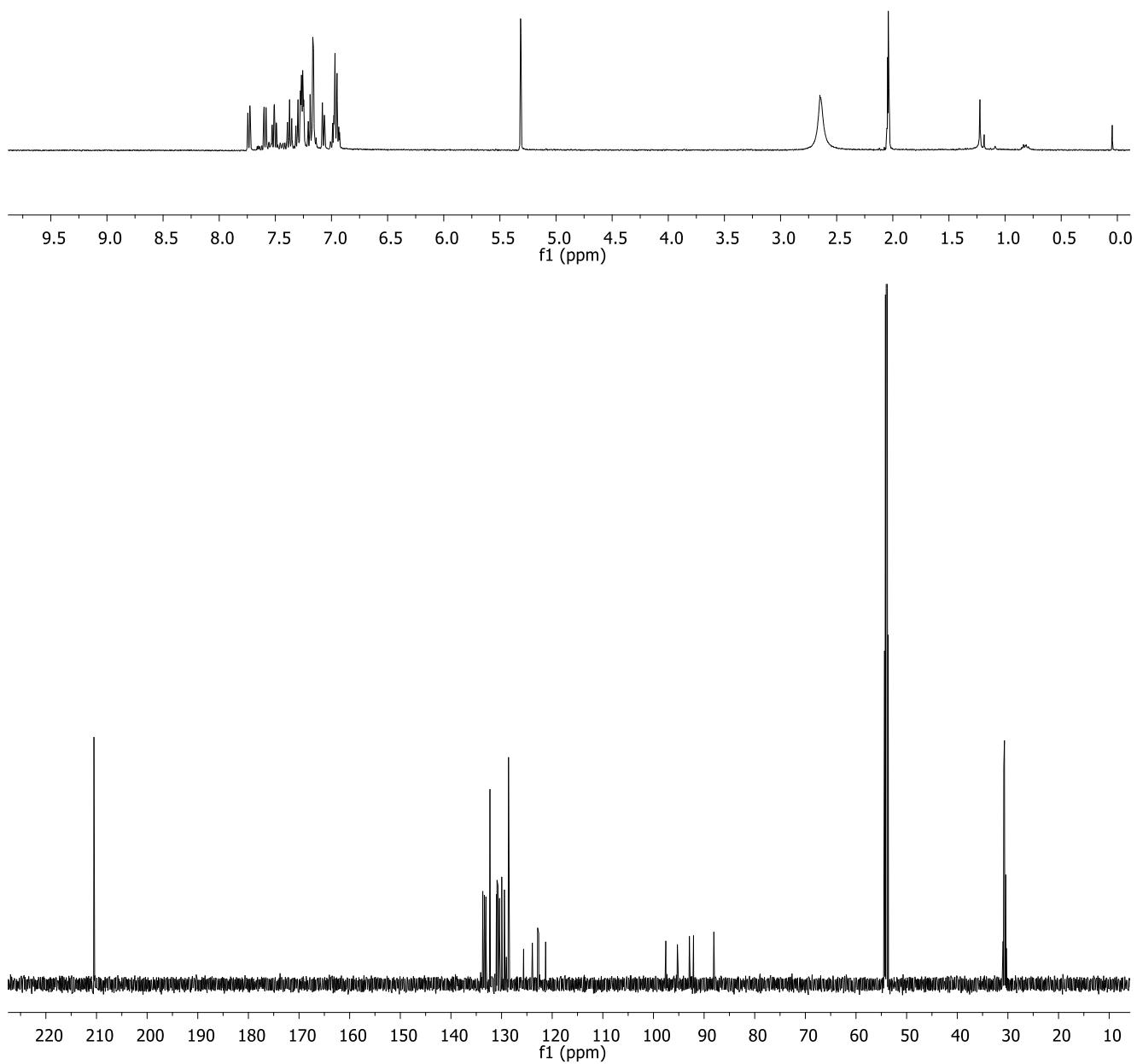
Compound 10



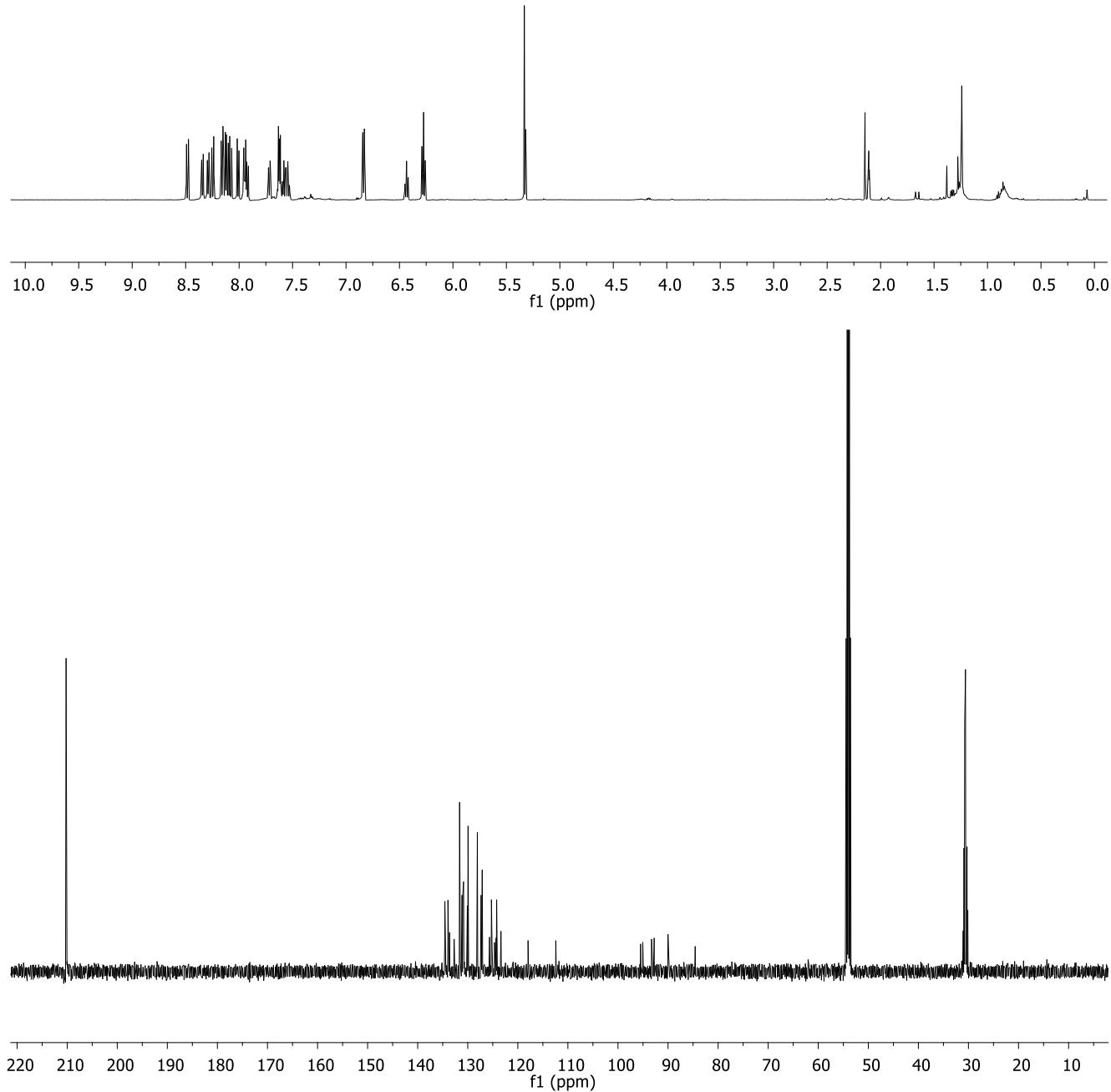
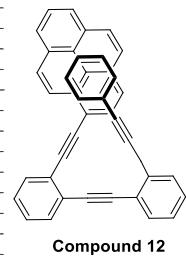
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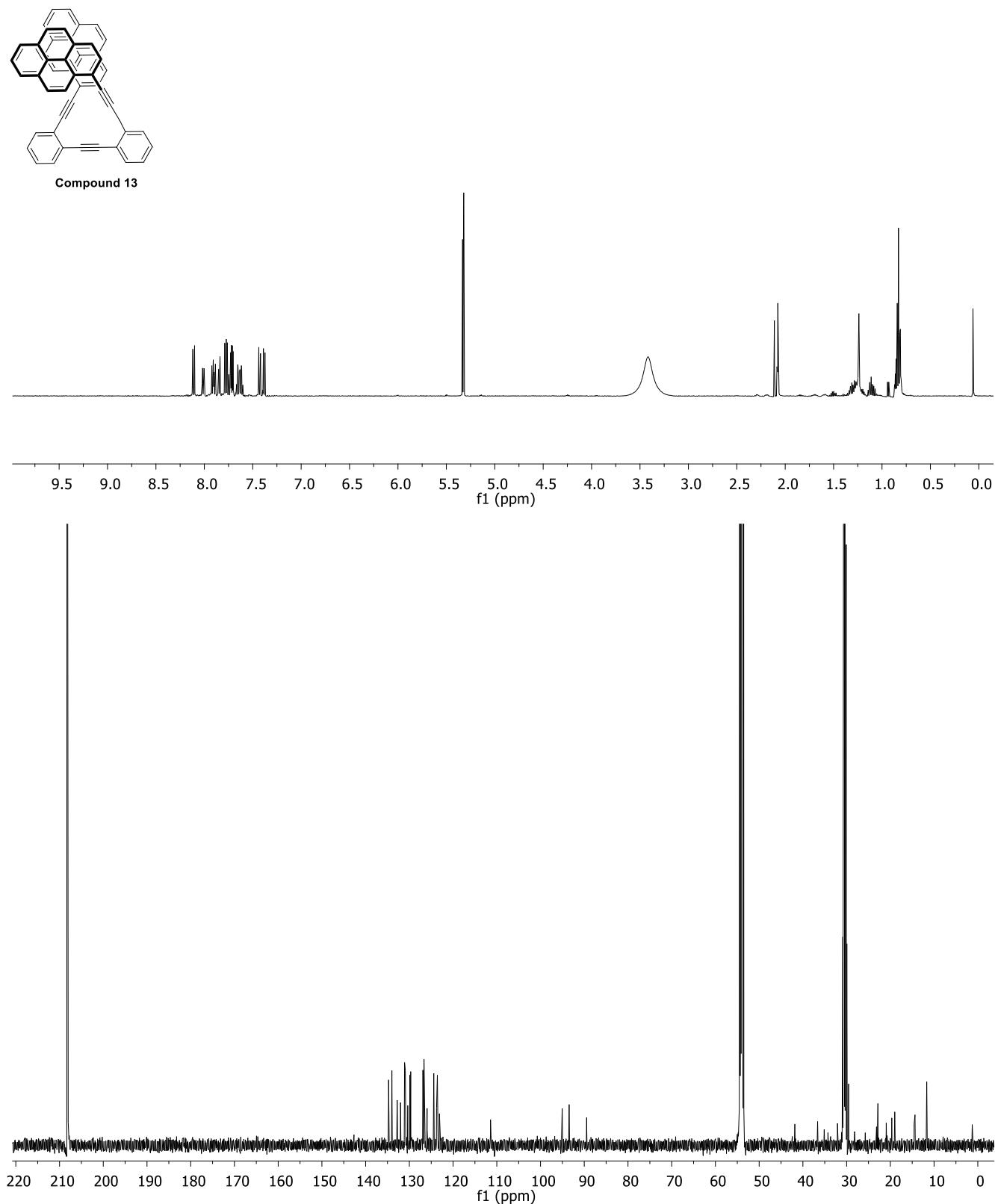
Compound 11



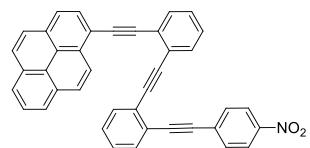
Complex 12·Ag(I):



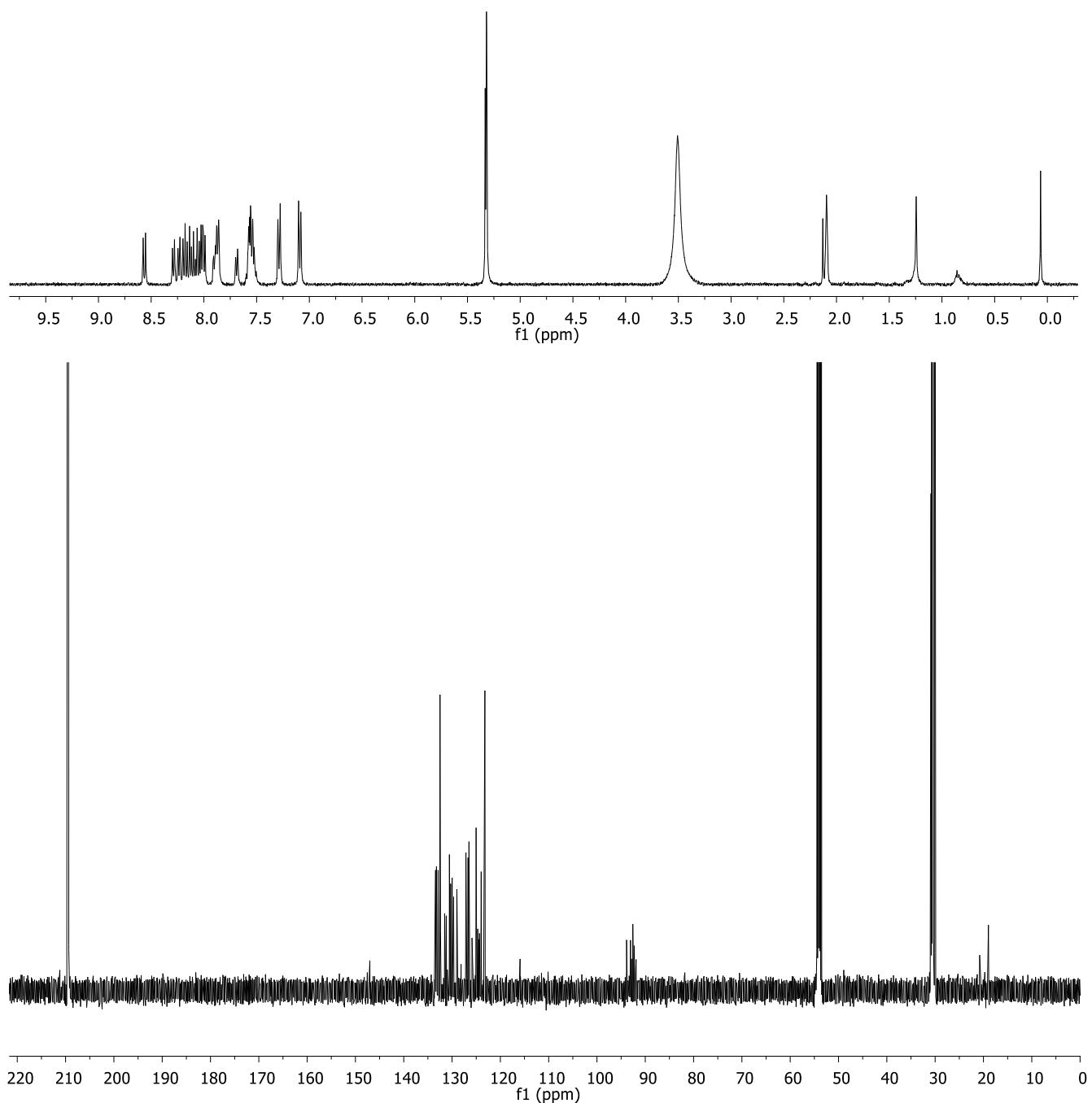
Complex 13·Ag(I):



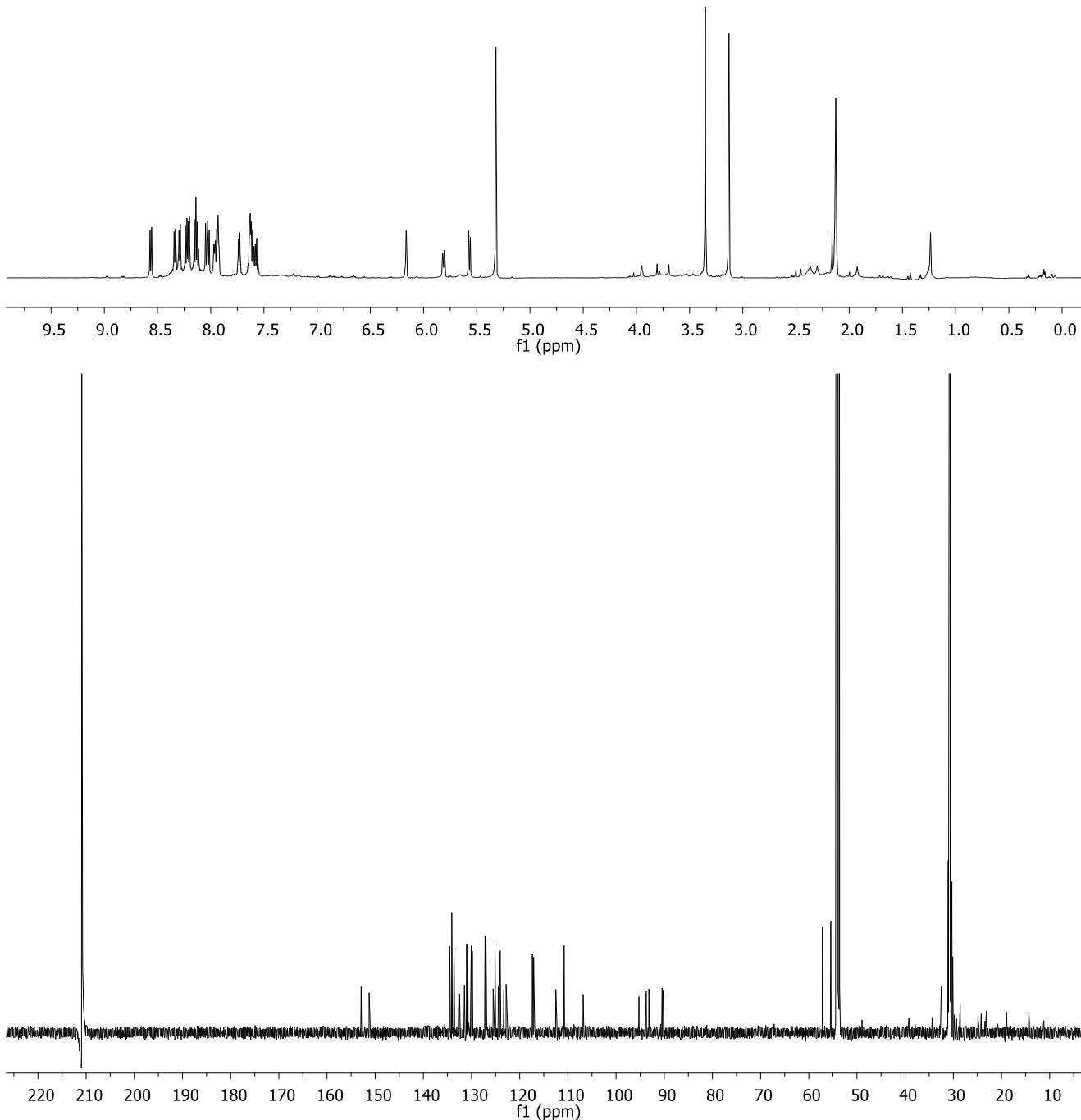
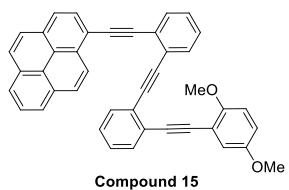
Complex 14·Ag(I):



Compound 14



Complex 15·Ag(I):



Complex 16·Ag(I):

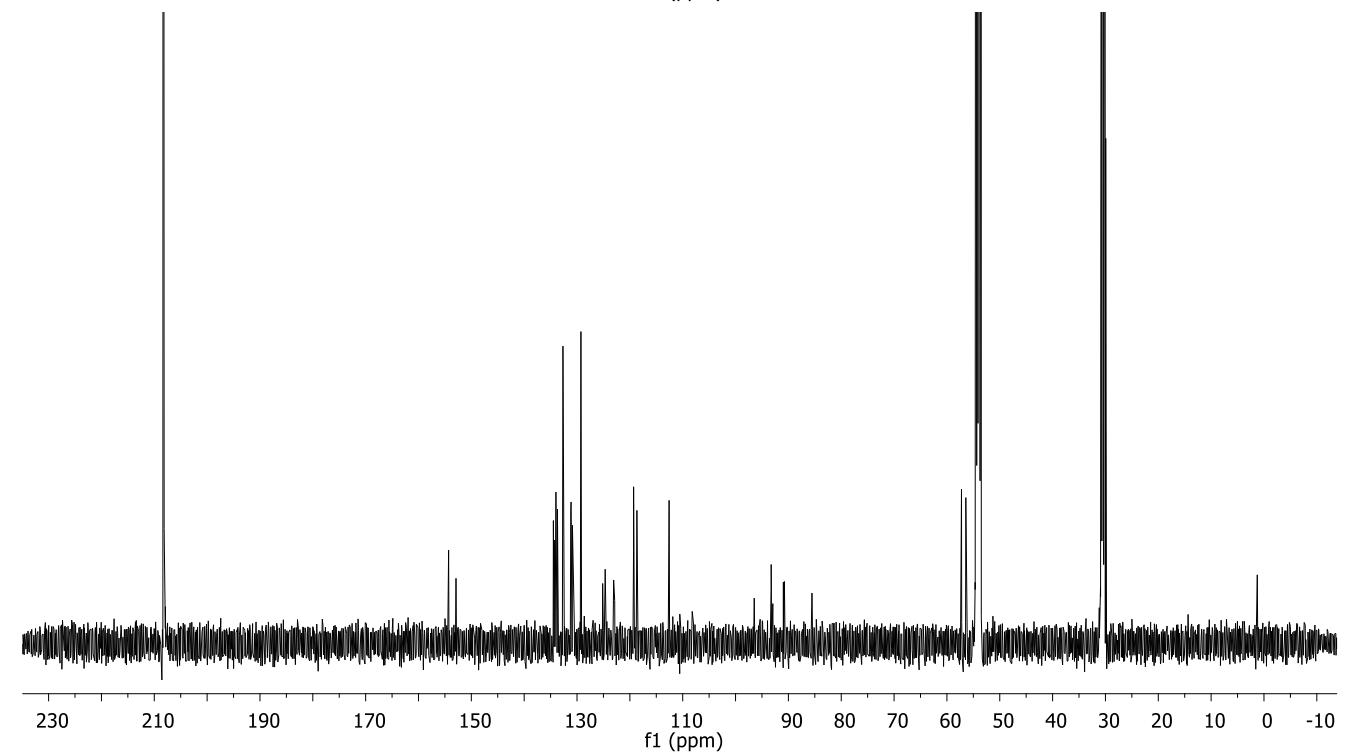
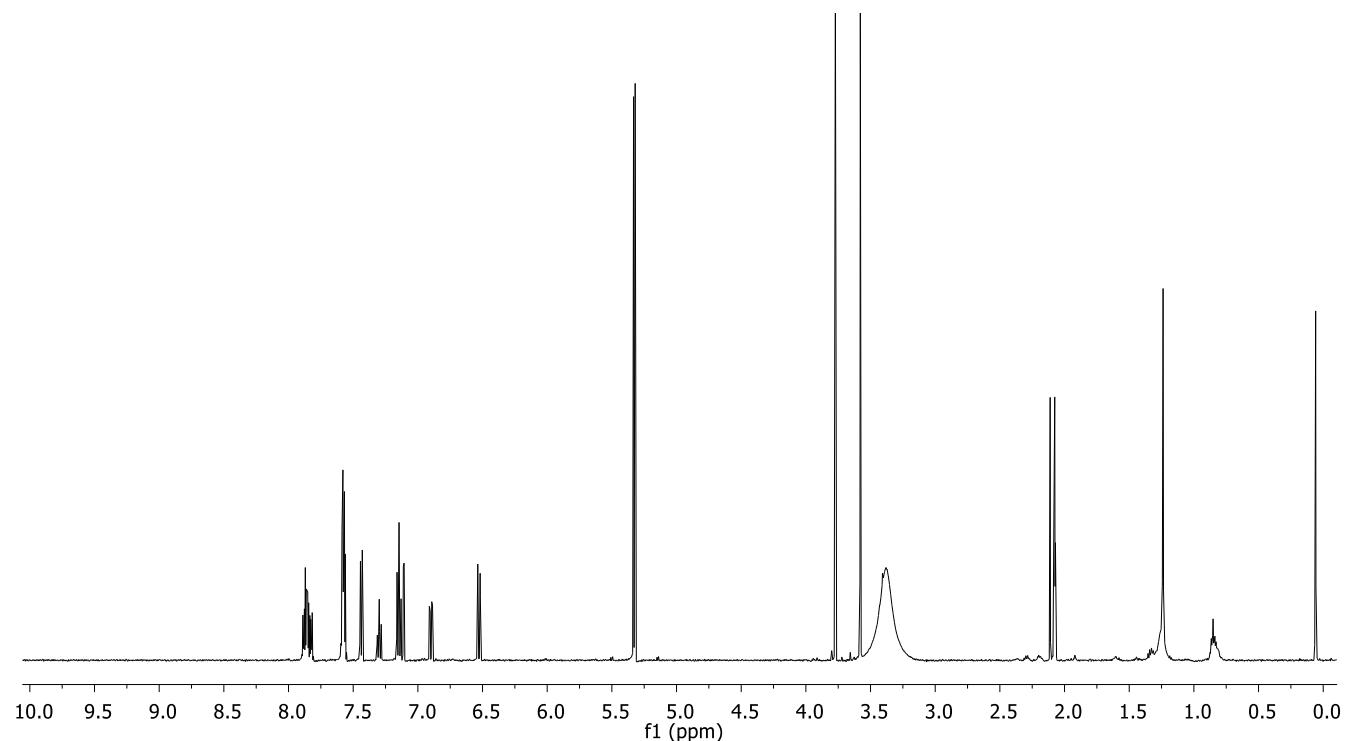
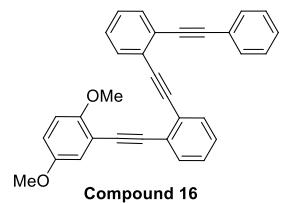


Figure S10: 2D-NOESY analysis of the complex 15·Ag(I)

The methoxy group located at the *ortho* position (Pink) respect to the alkyne (identify due to the cross peak A with H1 at 5.58 ppm) gave a strong NOE (crosspeak B) with the more deshielded H4 proton of the pyrene ring.

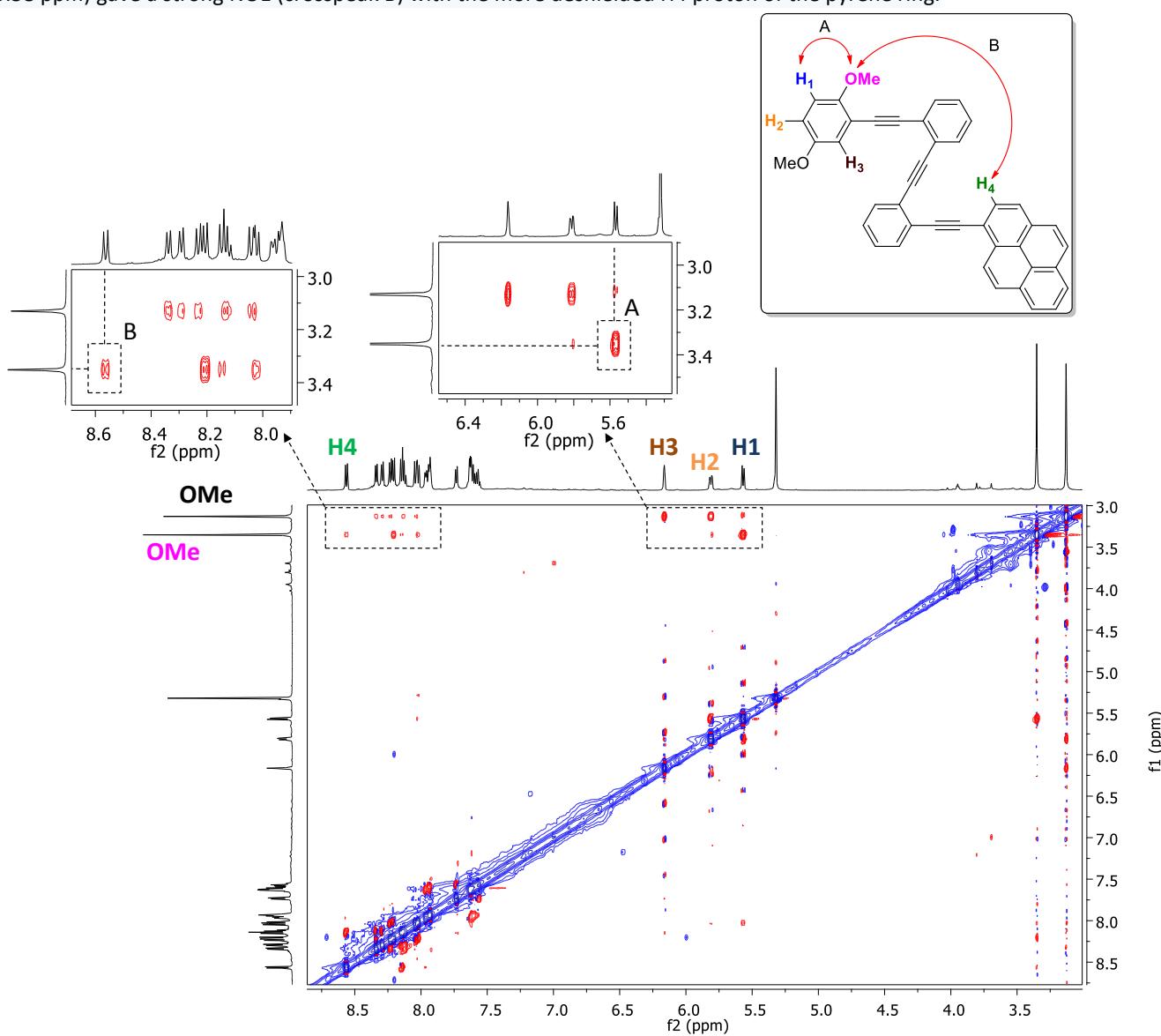


Figure S11: 2D-NOESY analysis of the complex 17·Ag(I)

The methoxy group located at the *ortho* position (Pink) respect to the alkyne (identify due to the cross peak A with H1 at 5.58 ppm) gave a strong NOE (crosspeak B) with the more deshielded H2 proton of the phenyl ring.

