Supporting Information

Conductance of Amyloid β Based Peptide Filaments: Structure-Function Relations

Moran Amit^a, Ge Cheng^b, Ian W. Hamley^b, and Nurit Ashkenasy^{*,a,c}

^a Department of Materials Engineering, Ben Gurion University of the Negev, Beer-Sheva, Israel

^b School of Chemistry, University of Reading, Reading, UK

^c The Ilse Katz Institute for Nanoscale Science and Technology, Ben Gurion University of the Negev, Beer-Sheva, Israel

Correspondence should be addressed to: nurita@bgu.ac.il

Synthesis of (3-Thi)(3-Thi)VLKAA

Fmoc-alanine-OH, Fmoc-lysine(Boc)-OH, Fmoc-leucine-OH and Fmoc-valine-OH, Fmocalanine–Wang resin (0.72 mmol g⁻¹ substitution), HBTU (2-(1H-benzotriazol-1-yl)-1,1,3,3tetramethyluronium hexafluorophosphate) were purchased from Novabiochem (UK). Fmoc-L-3-Thienylalanine-OH, trifluoroacetic acid (TFA), piperidine and triisopropylsilane were purchased from Sigma-Aldrich. HOBt/DMF (a mixture of 1-hydroxybenzotriazole and dimethylformamide), DIEA/NMP (a mixture of diisopropylethylamine and Nmethylpyrrolidone), and NMP were obtained from Applied Biosystems(UK). Water (HPLC grade), acetonitrile (HPLC grade) and diethyl ether were purchased from Fisher Scientific (UK).

The peptide (3-Thi)(3-Thi)VLKAA (Scheme 1) was synthesized by solid phase methods using standard FastMoc chemistry [Fmoc (9-fluorenylmethyloxycarbonyl) protecting group and activation by HBTU /HOBt] on a 0.25 mmol scale with a fully automated peptide synthesizer (433A Applied Biosystems), which allowed for direct conductivity monitoring of Fmoc deprotection. The crude peptide was purified with reverse phase HPLC (Perkin Elemer 200 with a Macherey-Nagel C18 semi-preparative column and a Perkin Elemer Series 200 UV/VIS Detector), followed by lyophilization to give a white solid. ESIMS m/z for $C_{37}H_{58}N_8O_8S_2 [M+H]^+$ calcd 807.38, found $[M+H]^+$, 807.39; $[M+2H]^{2+}/2$ calcd 404.20, found 404.20.¹H NMR (700 MHz methanol-d₄, ppm): 0.96(d, J = 7.0 Hz, 3H), 0.99 (t, J = 7.0 Hz, 9H), 1.42 (dd, J = 19.6 Hz, 7.7 Hz, 6H), 1.49 (m, 2H), 1.58-1.75 (m, 6H), 1.85 (m, 1H), 2.06 (sep, J = 7.0 Hz, 1H), 2.95 (t, J = 7.0 Hz, 2H), 3.03 (dd, J = 14.7 Hz, 8.6 Hz, 1H), 3.14 (dd, J = 14.4 Hz, 7.7 Hz, 1H), 3.18 (dd, J = 14.4 Hz, 5.6 Hz, 1H), 3.28 (dd, J = 15 Hz, 4.2 Hz, 1H), 4.10 (dd, J = 7.7 Hz, 4.9 Hz, 1H), 4.21 (t, J = 7 Hz, 1H), 4.37 (ABq, $J_{AB} = 7$ Hz, 3H), 4.43 (ABq, $J_{AB} = 5.6$ Hz, 1H), 4.76 (m, 1H), 7.04 (t, J = 4.2 Hz, 2H), 7.17 (d, J = 2.2 Hz, 1H), 7.27 (d, J = 2.2 Hz, 1H), 7.34 (dd, J = 4.9 Hz, 3.5 Hz, 1H), 7.43 (dd, J = 4.9 Hz, 3.5 Hz, 1H), 8.14, 8.19, and 8.26-8.59(NH, NH₂ 10 H).



(3-Thi)(3-Thi)VLKAA

Supporting figures



Figure S1. I-V curves of 0.03%wt (2-Thi)(2-Thi)VLKAA assembled in MeOH carried out on **a**) chip 1 and **b**) chip 2 under pressure of 10⁻³ mbar.





Figure S2. a) AFM height (z scale 88 nm) and the corresponding cross-sections across the blue and green lines of 0.03%wt (2-Thi)(2-Thi)VLKAA assembled in MeOH, showing fibers with diameter of ~45 nm and ~13 nm, respectively. **b)** AFM height (z scale 33 nm) and the corresponding cross-section across the blue line of 0.03%wt (2-Thi)(2-Thi)VLKAA assembled in TDW, showing bundled fibers with diameter of ~6 nm. **c)** AFM height (z scale 86 nm) and the corresponding cross-section across the blue line of 0.03%wt (2-Thi)(2-Thi)VLKAA assembled in TDW, showing bundled fibers with diameter of ~6 nm. **c)** AFM height (z scale 86 nm) and the corresponding cross-section across the blue line of 0.03%wt AAKLVFF assembled in MeOH, showing fibers with diameter of ~30 nm. **d)** AFM height (z scale 330 nm) and the corresponding cross-section across the blue line of 0.03%wt (3-Thi)(3-Thi)VLKAA assembled in MeOH, showing wide tapes with height of ~150 nm (blue line), either folded or twisted (green and purple arrows, respectively), and bundled fibers (blue arrow) with diameter of ~70 nm (green line). **f)** AFM height (z scale 58 nm) and the corresponding cross-section across the blue line of ~23 nm.