

Host-Guest Interactions in Polycationic Human Serum Albumin

Bioconjugates

- Supporting Information -

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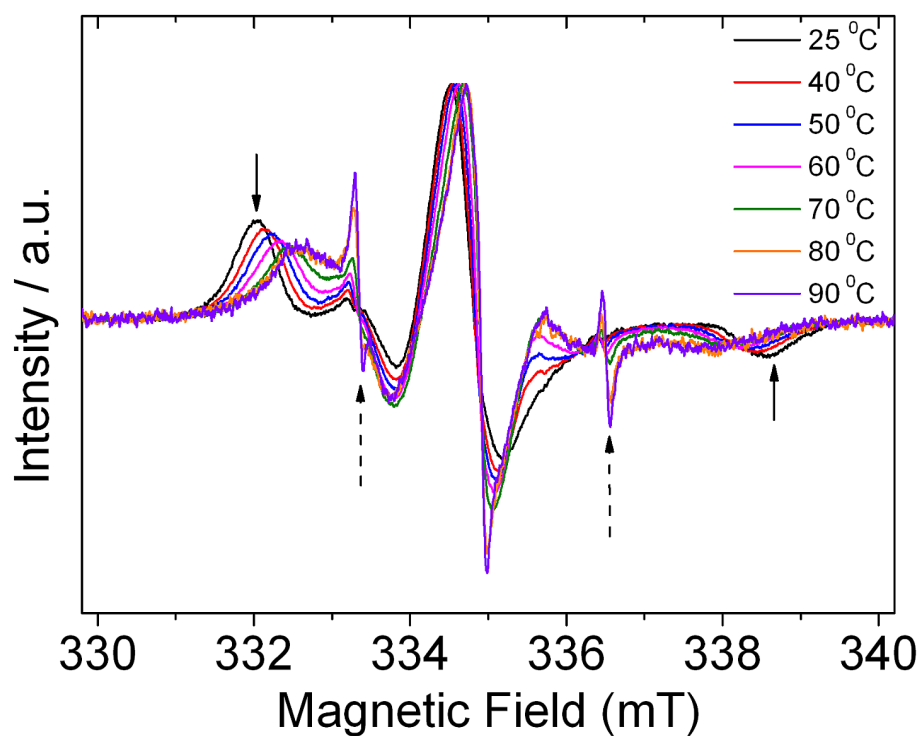


Figure S1. CW X-band EPR spectra of 5-DSA in HSA/buffer solution at an FA/HSA ratio of 4 at the temperatures of 25 °C (black), 40 °C (red), 50 °C (blue), 60 °C (magenta), 70 °C (green), 80 °C (orange) and 90 °C (violet). The characteristic signals of 5-DSA bound to albumin are marked by solid arrows and signals of free 5-DSA are marked by dashed arrows.

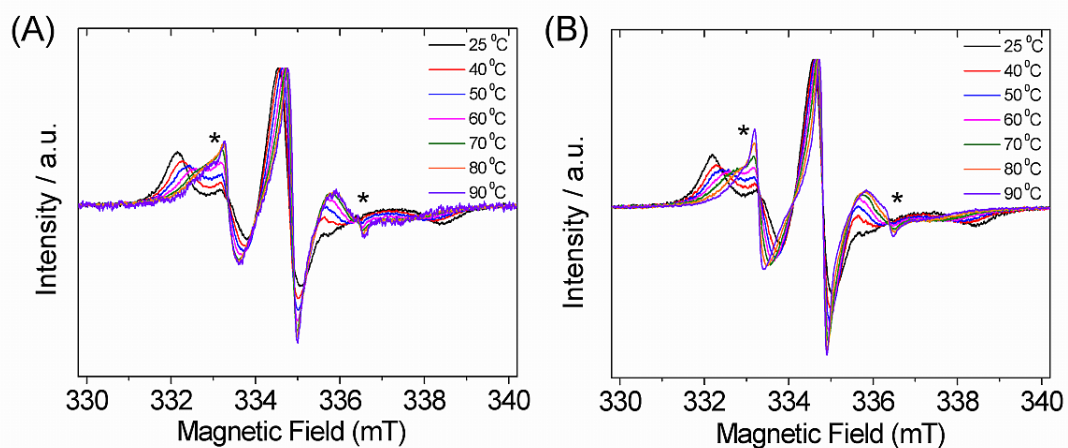


Figure S2. CW X-band EPR spectra of 5-DSA in cHSA/buffer (A) and in cHSA-PEO(2000)₁₉/buffer (B) solutions at an FA/HSA ratio of 4 at the temperatures of 25 °C (black), 40 °C (red), 50 °C (blue), 60 °C (magenta), 70 °C (green), 80 °C (orange) and 90 °C (violet). Asterisks mark a new spectral component corresponding to 5-DSA bound to low affinity electrostatic binding sites of cHSA, which is not observed in HSA.

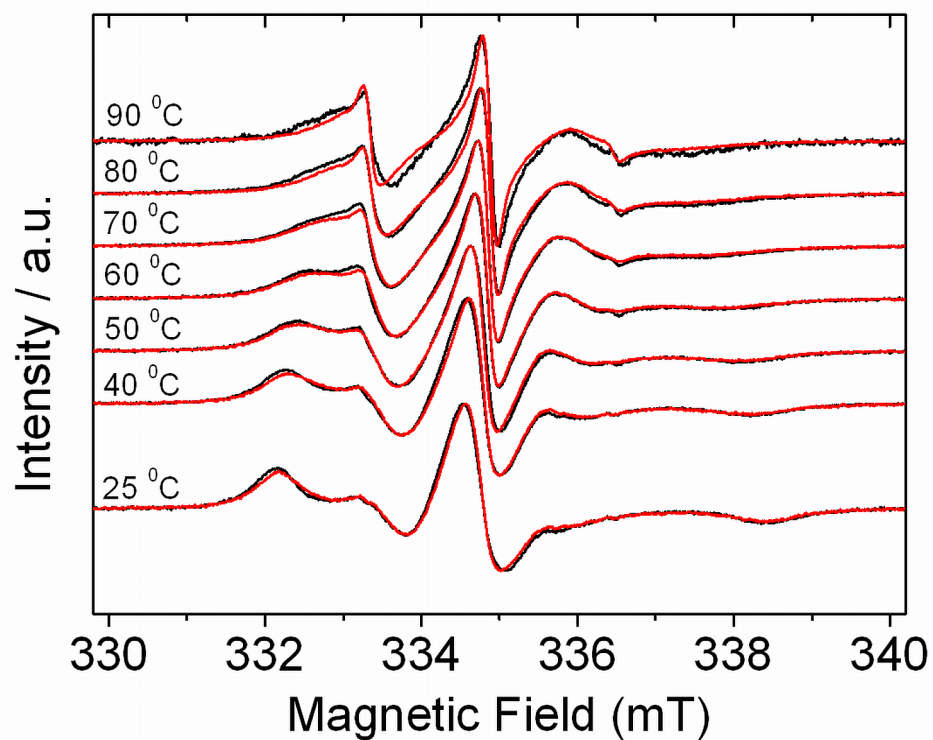


Figure S3. CW X-band EPR spectra of 5-DSA in cHSA/buffer (black) and in cHSA-PEO(2000)₁₉/buffer (red) solutions at a FA/HSA ratio of 4 at the temperatures of 25 °C, 40 °C, 50 °C, 60 °C, 70 °C, 80 °C and 90 °C.

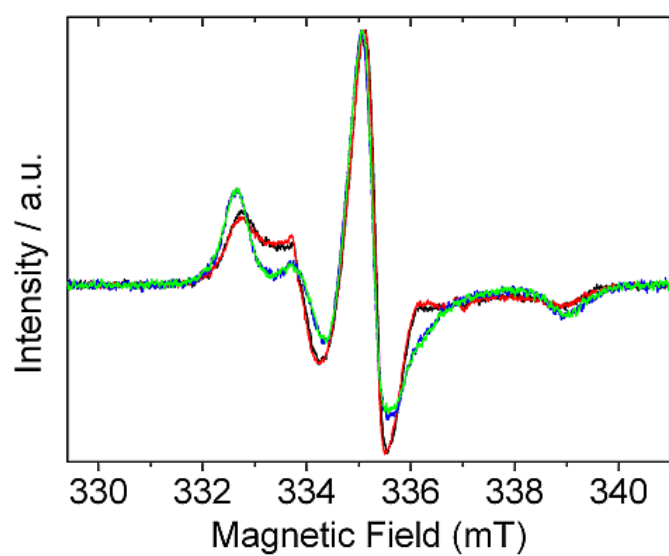


Figure S4. CW X-band EPR spectra of 5-DSA in cHSA/buffer (blue), in cHSA/PEO(750)₂₂/buffer (green) solutions and 16-DSA in cHSA/buffer (black), in cHSA/PEO(750)₂₂/buffer (red) solutions at a FA/HSA ratio of 2 at room temperature.

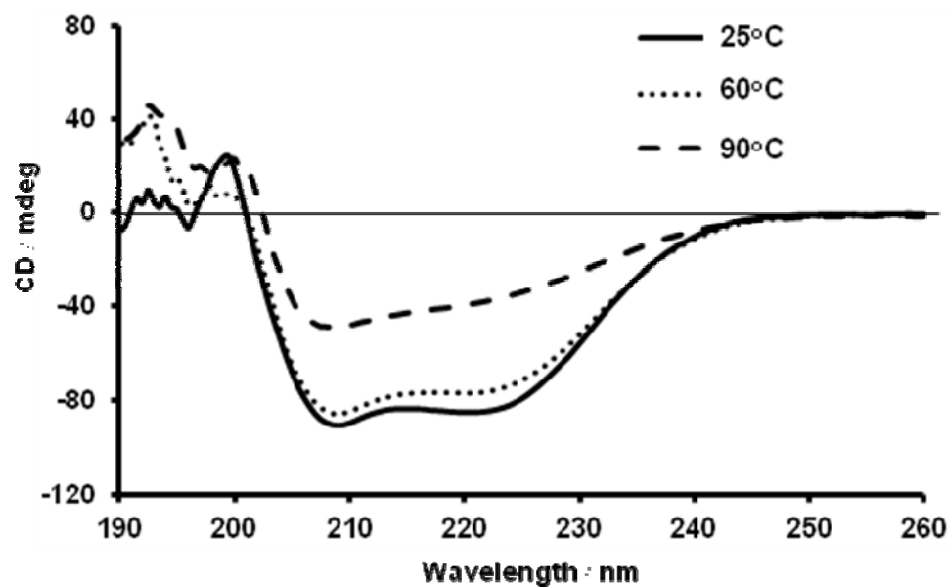


Figure S5. Circular dichroism spectra of HSA at different temperatures.

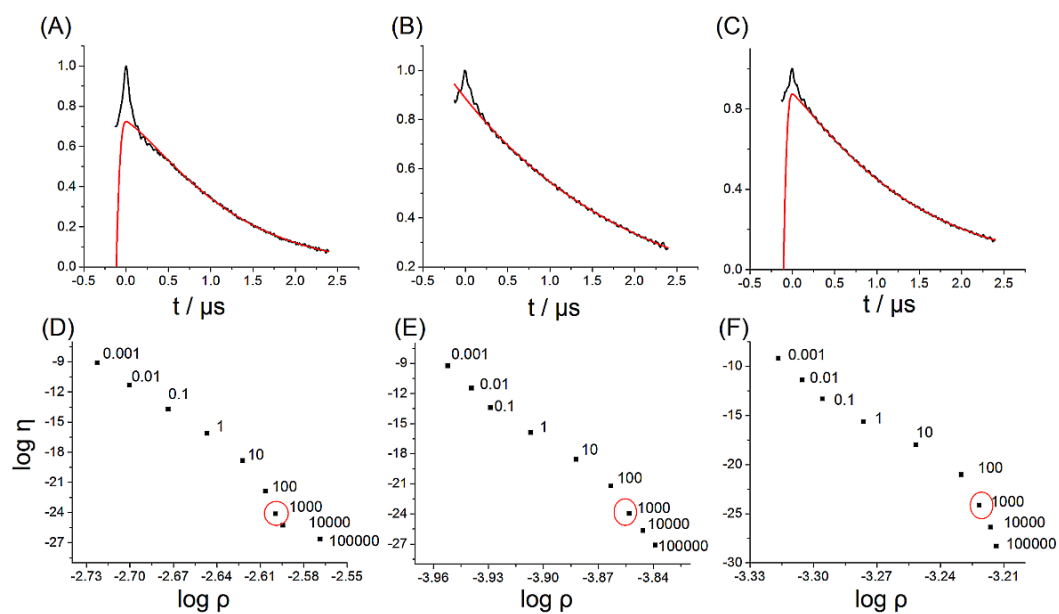


Figure S6. Time domain DEER data (black) and background fits (red) of 5-DSA in HSA (A), in cHSA (B) and in cHSA-PEO (750) (C), and the L-curves of 5-DSA in HSA (D), in cHSA (E) and in cHSA-PEO (750) (F).

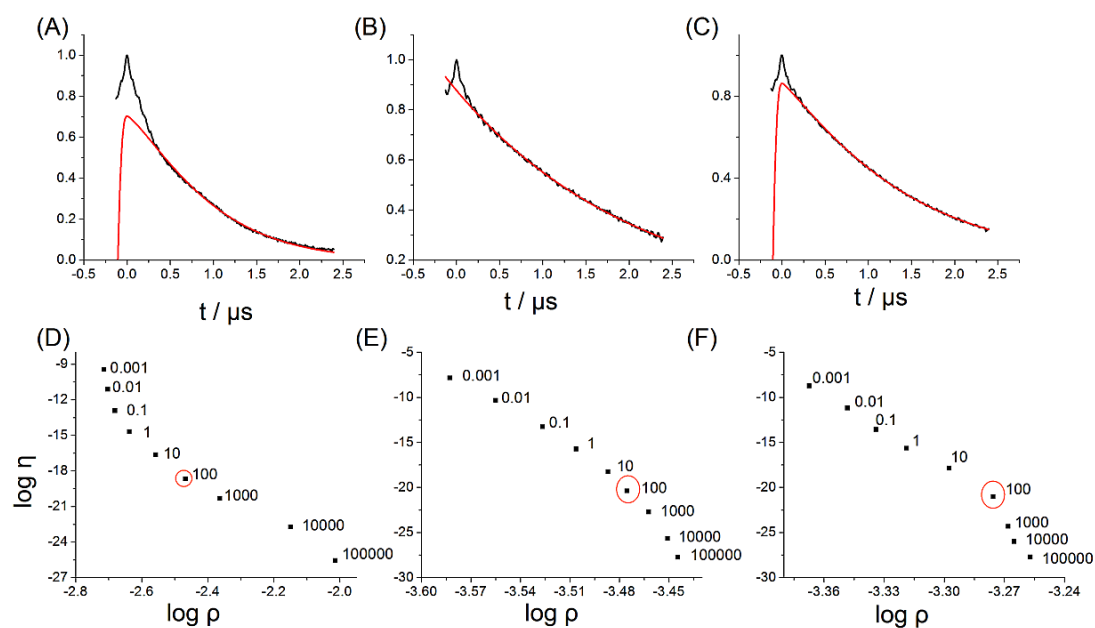


Figure S7. Time domain DEER data (black) and background fits (red) of 16-DSA in HSA (A), in cHSA (B) and in cHSA-PEO (750) (C), and the L-curves of 16-DSA in HSA (D), in cHSA (E) and in cHSA-PEO (750) (F).

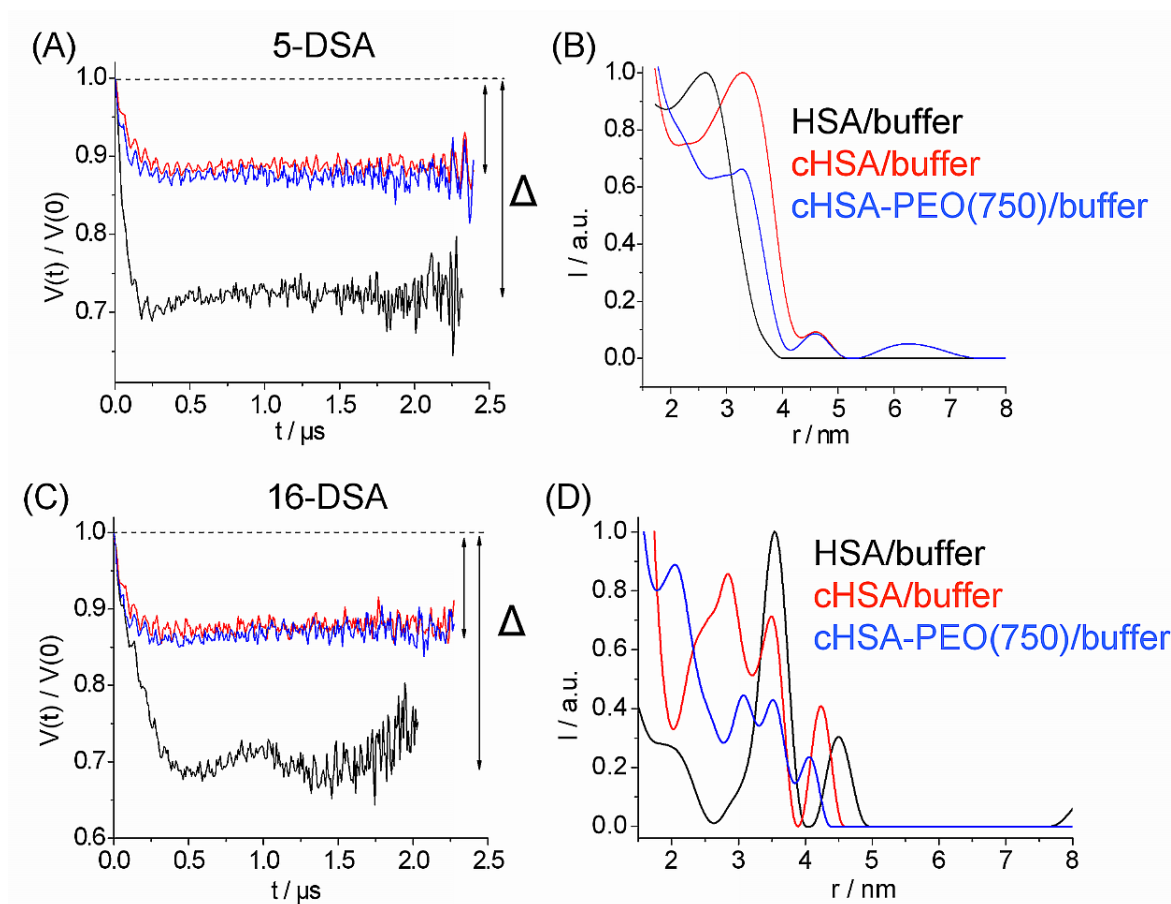


Figure S8. (A) Intramolecular part of the DEER time-domain data and the distance distribution obtained from Tikhonov regularization for 5-DSA (A, B) and 16-DSA (C, D), respectively, in HSA/buffer (black), in cHSA/buffer (red) and in cHSA-PEO(750)/buffer (blue) solutions. On average, each protein molecule is loaded with two spin-labeled FAs. The modulation depths Δ are marked. 20% (v/v) of glycerol was added to the aqueous solutions. $d = 3.8$ for HSA, $d = 3.4$ for cHSA-PEO(750)₂₂ and $d = 3.0$ for cHSA (see Fig. S6 and S7).

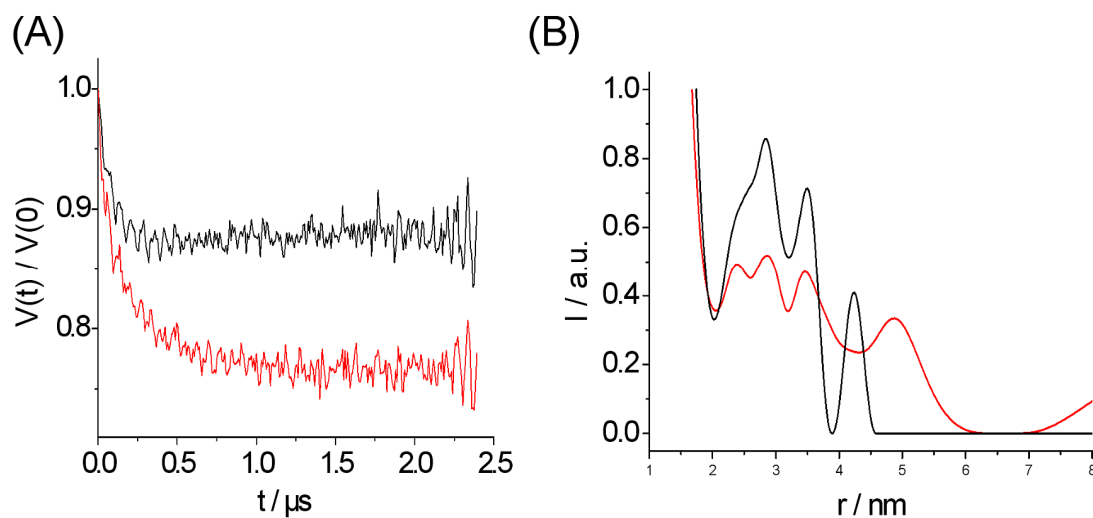


Figure S9. Intramolecular part of the DEER time-domain data (A) and extracted distance distributions by Tikhonov regularization (B) of two paramagnetic 16-DSA complexed in cHSA/buffer (black) and additional four diamagnetic analogues of 16-DSA (nitroxides reduced to hydroxylamines) in cHSA/buffer (red). 20% (v/v) of glycerol was added to the aqueous solutions.

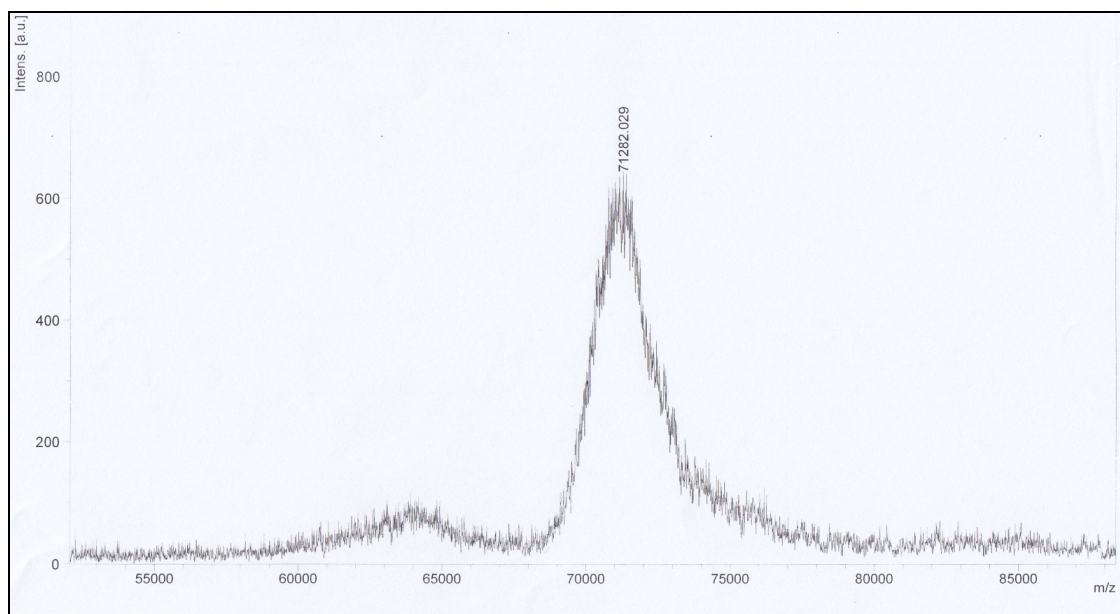


Figure S10new. MALDI-ToF spectrum of cHSA.

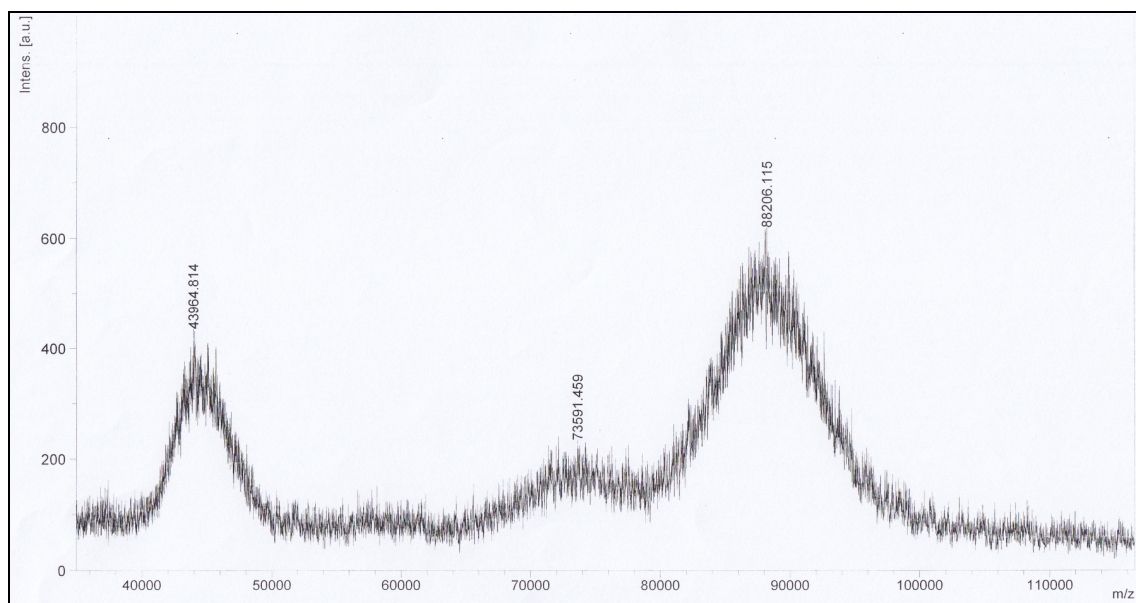


Figure S11new. MALDI-ToF spectrum of cHSA-PEO(750)₂₂.

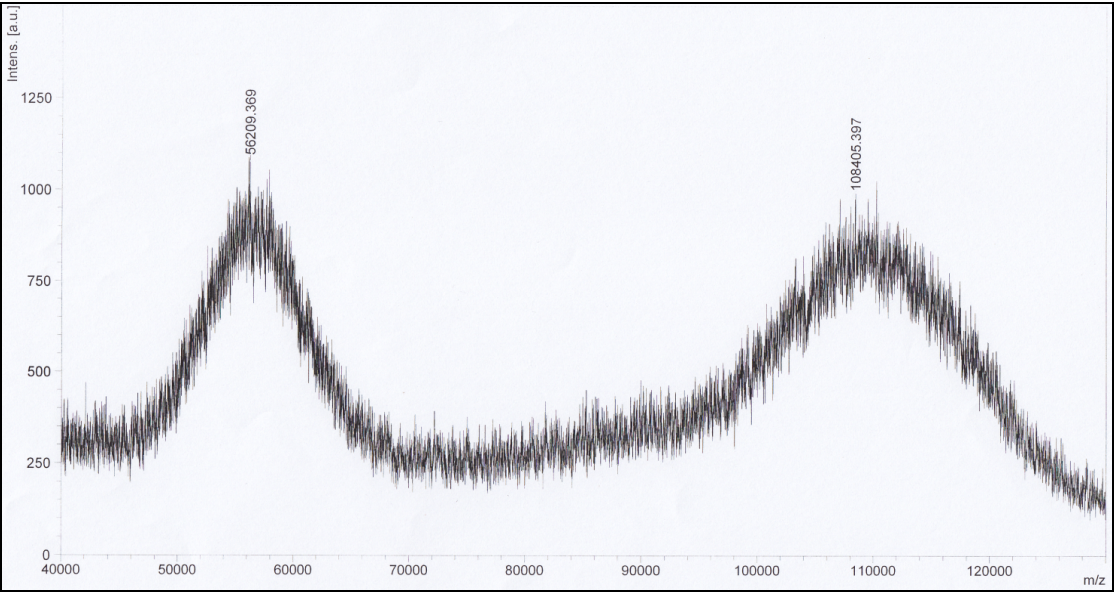


Figure S12new. MALDI-ToF spectrum of cHSA-PEO(2000)₁₉.