

# 1 Supplementary Information

## 1.1 Synthesis

### 1.1.1 Synthesis of cholesteryl-succinic acid propargylamide

2.5 g (5.1 mmol, 1.1 eq) of cholesteryl-hemisuccinate (Aldrich), 1.81 g (5.6 mmol, 1.3 eq) of *p*-toluenesulfonic anhydride (Aldrich) and 1.26 g (10 mmol, 2.2 eq) of DMAP are dissolved in 25 ml of chloroform and after 15 min 0.26 g (4.6 mmol, 1 eq) propargylamine (Aldrich) are added. After 1.5 h the mixture is quenched with 3 ml of a saturated solution of sodium bicarbonate (NaHCO<sub>3</sub>). The solution is extracted with ethyl acetate and the combined organic layers are washed two times with 50 ml of sat. NaHCO<sub>3</sub> and two times with 50 ml of brine. The organic layer is dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>. The crude product is purified by recrystallisation in ethyl acetate and freeze dried from cyclohexane.

- **Yield:** 1.25 g (55 %).
- **<sup>1</sup>H-NMR** (300MHz, CDCl<sub>3</sub>): 6.2 ppm (1H, -NHCO-); 5.35 ppm (1H, H-6 Cholesterol); 4.6 ppm (1H, H-3 Cholesterol); 4.0 ppm (2H, -CH<sub>2</sub> propargyl); 2.2 ppm (1H, H-alkyne); 0.65 ppm (9H, -CH<sub>3</sub> cholesterol).

### 1.1.2 Synthesis of the SAPs

Prior to the experiment, solutions of CuSO<sub>4</sub> (*c* = 0.13 mol/l) and a THPTA (*c* = 63 mmol/l) are prepared with millipore water. The polyrotaxane (3 μmol) and the cholesteryl succinic acid propargylamide (6 μmol, 2 eq (per azide)) are dissolved in a mixture of 1.5 ml tBuOH/millipore water 8:2, sonicated for 5 min and heated for several minutes to ensure complete solubilisation of the compounds. Then the ligand solution (1 μmol, 0.3 eq) and the CuSO<sub>4</sub> solution (0.2 μmol, 0.06 eq) are added to the mixture to give a transparent solution. Sodium ascorbate (2.5 μmol, 0.8 eq) is added and the solution is left under stirring over night at room temperature. The transparent solution is diluted with 5 ml of Millipore water and dialysed (cut-off 2 kg/mol) twice with 2 l of millipore water for 24 h and freeze dried. The crude product is taken up in 5 ml of ether and centrifuged 3 times to eliminate the residual cholesteryl alkyne. The residue is dissolved in 10 ml of tBuOH/H<sub>2</sub>O 8:2 and freeze dried.

- **Yield:** 50-95 % (depending on MW of polyrotaxane and used stopper).
- **<sup>1</sup>H-NMR** (300MHz, CDCl<sub>3</sub>) DMPE stopper: 8.3 ppm (s, 1H, -NHCO-); 7.8 ppm (s, 1H, H-triazol); 6.96 - 6.74 ppm (6H, aromatic H<sub>O</sub>s DMPE); 5.86 ppm (4H, H-urea); 5.6 ppm - 5.4 ppm (12H, OH-2 and OH-3 CD); 5.3 ppm (1H, CH sp<sup>2</sup> cholesterol); 5.0 ppm (1H, H-1 modified glucose unit CD); 4.8 ppm (5H, H-1 CD<sub>0</sub>); 4.5 ppm (6H, OH-6 CD); 3.1-3.9 (nH, -OCH<sub>2</sub>CH<sub>2</sub>- PEG and H-2, H-5, CH<sub>2</sub>-6 CD); 2 - 0.8 ppm (H cholesteryl moiety); 0.65 ppm (9H, -CH<sub>3</sub> cholesterol).

### 1.1.3 Synthesis of Cholesteryl α-CD

100 mg (50 μmol, 1 eq) ACDN3 and 29 mg (60 μmol, 1.2eq) cholesteryl succinic acid propargylamide are dissolved in 1.5 ml of ultrapure DMF and 160 μl of a CuI/PMDETA 1:1 solution (*c* = 57 mmol/l in DMF) are added under argon atmosphere. The solution is stirred for 24h. The solvent is evaporated and the compound is suspended in 4 ml of phosphate buffer solution (20 mM, *pH* = 6.5). Then the compound is centrifuged in 3 ml of millipore water, as well as 3 times in 2 ml of acetone. Finally the compound is taken up in 10 ml of millipore water and freeze dried.

- **Yield:** 35 mg (35 %).
- **<sup>1</sup>H-NMR** (300MHz, DMSO-D<sub>6</sub>): 8.30 (1H, -NHCO-); 7.8 ppm (1H, H-triazol); 5.6 ppm - 5.4 ppm (12H, OH-2 and OH-3 CD); 5.0 ppm (1H, H-1 modified glucose unit CD); 4.8 ppm (5H, H-1 CD); 4.4 ppm (6H, OH-6 CD); 3.2-3.7 (H-2 and H-5, CH<sub>2</sub>-6 CD); 2.5 ppm (residual H<sub>2</sub>O); 2 - 0.8 ppm (H cholesteryl moiety); 0.65 ppm (9H, -CH<sub>3</sub> cholesterol).

## 1.2 Monolayer in-plane morphology

### 1.2.1 Brewster Angle Microscopy

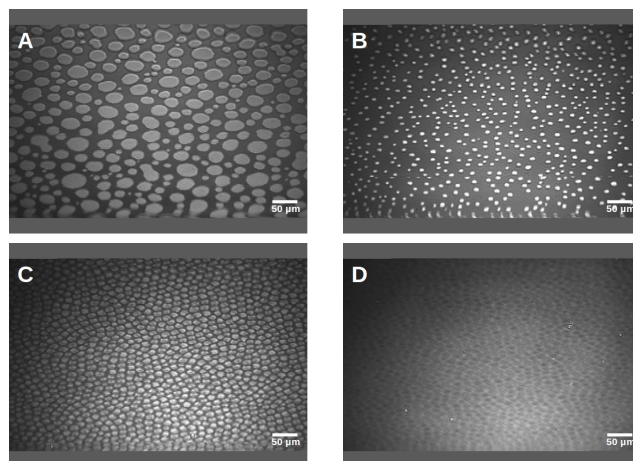


Figure 1: BAM images for the mixture 3 mol% SAP-6k/DPPC at: A) 5 mN/m; B) 9 mN/m; C) 13 mN/m and D) 30 mN/m.

The Brewster Angle Microscope (BAM), type PI, C-138K003, Optrel GBR, Berlin, coaligned with the Langmuir trough is based on the Hoenig and Moebius setup [1]. A green laser (Las-Nova series 50) with a wavelength of 532 nm is directed onto the water surface at the Brewster Angle ( $53.1^\circ$ ). The reflected light from the surface is imaged by means of a CCD camera (EHD®kamPro02) to give images of the monolayer morphology with a size of  $480 \mu\text{m} \times 599 \mu\text{m}$  and a resolution of  $480 \times 640$  pixel.

### 1.2.2 Atomic force microscopy

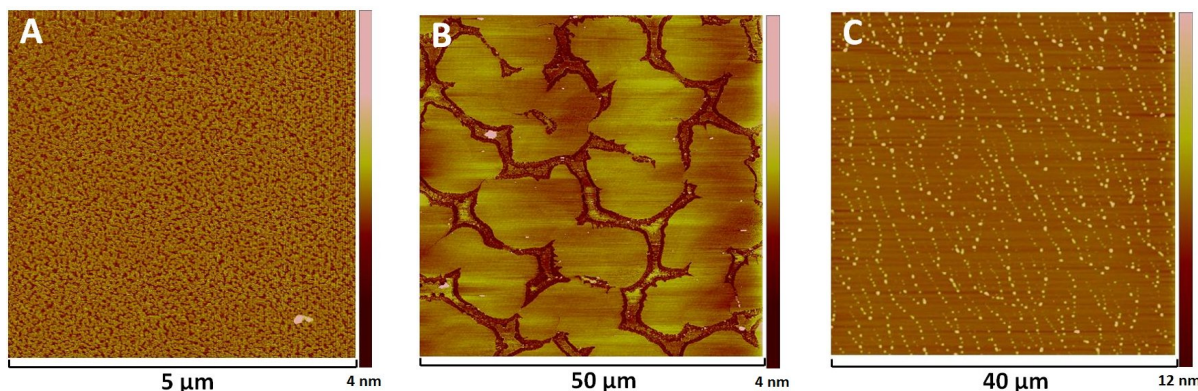


Figure 2: AFM images for monolayers of the mixture 3 mol% SAP-6k/DPPC deposited on mica at: A) 5 mN/m; B) 13 mN/m and C) 35 mN/m.

Mixed SAP/DPPC monolayers at several surface pressures were transferred from the air-water interface onto freshly cleaved, hydrophilic mica wafers ( $11 \times 11 \times 0.15 \text{ mm}$ , purchased from Agar Scientific), using the Langmuir-Blodgett (LB) technique (dipper speed: 1 mm/min). Transfer ratios close to 1 were obtained for good monolayer depositions.

The films are then examined in tapping mode with a Nanoscope V (Veeco) AFM. Standard cantilevers with a conical silicon etched probe tip (NSC15,  $\mu\text{masch}$ ) with typical spring constants in the order of 40 N/m, as determined by the thermal resonance method [2], and typical resonance frequencies in the order of 350 kHz are used. Images with scan sizes of  $1 \mu\text{m} \times 1 \mu\text{m}$  and  $10 \mu\text{m} \times 10 \mu\text{m}$  have been recorded with scan rates of 1 Hz and 0.5 Hz, respectively.

### 1.2.3 Discussion

Using BAM and AFM the in-plane film morphology can be investigated from  $\mu\text{m}$  to nm scale. Mixtures with SAPs of different MW give comparable results for similar surface densities. At

low surface pressures large bright domains of  $\sim 50 \mu\text{m}$  in diameter are visible (Figure 1 A). Complementary AFM images show that there is also a heterogeneity at much smaller scale in the order of tens of nanometers (Figure 2 A). With further compression those domains vanish and starting at  $\Pi > 8 \text{ mN/m}$  small ( $10 \mu\text{m}$ ) brighter domains appear, which grow in number (Figure 1 B). These brighter domains, visible for all molar ratios and regardless the MW of the SAPs, correspond to the LE-LC phase transition, typical for DPPC [3]. These domains are also displayed in corresponding AFM images (Figure 2 B).

The domains become denser with further rise of  $\Pi$  (Figure 1 C and Figure 2 C), and an inversion of contrast occurs for  $\Pi \sim 20 \text{ mN/m}$ , resulting in a honeycomb-like pattern, which prevails up to very high surface pressures (Figure 1 D). This is in contrast to pure DPPC, where a uniform surface is obtained. For higher SAP molar ratios the bright domains visible at  $13 \text{ mN/m}$  are less dense and the inversion of contrast occurs at lower surface pressure to give the same honeycomb-like pattern only with higher contrast. Interestingly for molar ratios  $> 30 \text{ mol\%}$  BAM images do not display a contrast for high surface pressures. The AFM images for high surface pressures do not display the patterns observed in BAM, but many aggregates are visible (Figure 2 D). The aggregates (height  $5 - 10 \text{ nm}$ ) increase in number with SAP molar ratios and they are oriented in the direction of deposition.

### 1.3 Neutron reflectivity

Table 1: Selected SLDs, taken from references [4–7], for the different components. The SLD for the  $\alpha$ -CD part of the SAP is calculated, using the molecular volume  $V_m = 1000 \text{ \AA}^3$  and the scattering length  $b = 189 \text{ fm}$ . To account for the complexation of  $\alpha$ -CD and PEG in the SAP the SLD of the inserted PEG is added to the  $\alpha$ -CD.

material	$SLD[10^{-6}\text{\AA}^{-2}]$
Si	2.07
SiO <sub>2</sub>	3.47
D <sub>2</sub> O	6.34
4MW	4
ZMW	0
H <sub>2</sub> O	-0.56
DPPC-alkyl tail	-0.4
DPPC-d <sub>62</sub> alkyl tail	6.82
DPPC-PC head	1.74
DSPE-alkyl tail	-0.4
DSPE-PE head	2.66
PEG	0.6
$\alpha$ -CD head + inserted PEG	2.4
$\beta$ -CD head	1.9
cholesteryl anchor	0.5

		bilayer 10 mol% SAP-6k		bilayer 20 mol% SAP-6k	
layer		25°C	50°C	25°C	50°C
water	thickness [Å]	4.6 ± 1	5.3 ± 1	4.0 ± 1	6.3 ± 1
	SLD [Å <sup>-2</sup> ]	-	-	-	-
	water [v/v%]	100	100	100	100
	roughness [Å]	7.1 ± 2	7.1 ± 2	6.3 ± 2	6.5 ± 2
heads DPPC	thickness [Å]	9.1 ± 1	9.12 ± 1	9.0 ± 1	9.2 ± 1
	SLD [Å <sup>-2</sup> ]	1.7	1.7	1.7	1.7
	water [v/v%]	30 ± 5	35 ± 5	28 ± 5	28 ± 5
	roughness [Å]	7.8 ± 2	6.6 ± 2	7.0 ± 2	7.5 ± 2
tails	thickness [Å]	30 ± 1	25 ± 1	29.7 ± 1	25.5 ± 1
	SLD [Å <sup>-2</sup> ]	6.81 ± 0.2	6.6 ± 0.2	6.55 ± 0.2	6.37 ± 0.2
	water [v/v%]	10 ± 5	10 ± 5	10 ± 5	10 ± 5
	roughness [Å]	8.5 ± 2	8.3 ± 2	9.0 ± 2	8.0 ± 2
heads mixed	thickness [Å]	10.2 ± 1	10.0 ± 1	12.2 ± 1	12.0 ± 1
	SLD [Å <sup>-2</sup> ]	1.8	1.8	1.8	1.8
	water [v/v%]	25 ± 5	25 ± 5	22 ± 5	21 ± 5
	roughness [Å]	8.5 ± 2	8.5 ± 2	8.0 ± 2	8.5 ± 2
polymer	$H$ [Å]	84 ± 10	75 ± 10	110 ± 10	95 ± 10
	SLD [Å <sup>-2</sup> ]	0.6	0.6	0.6	0.6
	$\Phi_0$ [v/v]	0.05 ± 0.02	0.041 ± 0.02	0.11 ± 0.02	0.11 ± 0.02
	roughness [Å]	9 ± 2	9.2 ± 2	9.5 ± 2	10 ± 2

Table 2: Neutron reflectivity results for supported bilayers with a first monolayer DSPE as well as a second mixed monolayer DPPC/cholesteryl  $\beta$ -CD and DPPC/SAP-10k, respectively.

## References

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