Electronic Supplementary Information

Gelation of Amino acid-Based Amphiphiles in Water-Based Mixed Solvent Systems: Reusable Catalytic Templates for Nanostructured Silica and Silica-Zirconia Photocatalyst

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Synthesis of amino acid based amphiphiles. A series of different amino acid-based amphiphiles (conjugates of different chain length of fatty acids and tyrosine/phenylalanine/leucine) such as (C₁₇H₃₅-Tyr-OH/ C₁₃H₂₇-Tyr-OH/ C₉H₁₉-Tyr-OH/ $C_{17}H_{35}$ -Phe-OH or $C_{17}H_{35}$ -Leu-OH) are synthesized. The preparation of these amphiphiles involves the synthesis of their methyl ester (C17H35-Tyr-OMe/ C13H27-Tyr-OMe/ C9H19-Tyr-OMe/ $C_{17}H_{35}$ -Phe-OMe or $C_{17}H_{35}$ -Leu-OMe) followed by their saponification. The methyl esters of these amphiphiles are synthesized by conventional solution-phase method using a racemization-free fragmentation/condensation strategy. In general, the C terminus of amino acid is first protected via esterification with a methyl group, followed by coupling with a fatty acid by using DCC and HOBt. These esters were finally saponified to obtain the actual amphiphiles.

Synthesis of methyl ester of tyrosine (H₂N-Tyr-OMe), phenylalanine (H₂N-Phe-OMe) and leucine (H₂N-Leu-OMe). First, 20 mmol of either tyrosine (3.62 gm)/ phenylalanine (3.3 gm)/ leucine (2.62) is dissolved in of 20 ml ice cold dry methanol containing 2 ml SOCl₂ and allowed to stand for 8h to obtain the chloride salt of methyl ester of different amino acid (Cl⁻H₃N⁺-Tyr-OMe/ Cl⁻H₃N⁺-Phe-OMe/ Cl⁻H₃N⁺-Leu-OMe respectively). The subsequent neutralization of these compounds with saturated Na₂CO₃ solution and extraction with ethyl acetate results the required H₂N-Tyr-OMe/ H₂N-Phe-OMe/ H₂N-Leu-OMe. Using these purified esters, the following methyl esters of amphiphiles were synthesized.

Synthesis of methyl ester of amphiphiles (C₁₇H₃₅-Tyr-OMe/ C₁₃H₂₇-Tyr-OMe/ C₉H₁₉-Tyr-OMe/C₁₇H₃₅-Phe-OMe or C₁₇H₃₅-Leu-OMe): 12 mmol of different fatty acid [capric acid (2.06 g)/myristic acid (2.73 g)/ stearic acid (3.5 g)] is taken in DCM (10 ml) and cooled in an ice-water bath. DCM solution of purified methyl ester of amino acid (10 mmol) is then added separately to this fatty acid solution. Finally, DCC (2.06 g, 10 mmol) and HOBt (0.135 g, 1 mmol) are added consecutively to each of above reaction mixture and stirred magnetically for 3 days. Final reaction mixture taken in ethyl acetate (30 ml) was filtered through sintered funnel to remove the formed dicyclohexyl urea (DCU). The filtrate containing the actual compound is washed with 2N HCl (3×20 ml), brine (1×20 ml), saturated sodium carbonate (3×20 ml), water (1×20 ml) and dried over anhydrous sodium sulfate. The crude compound is isolated by evaporating the solvent in rotary evaporator and finally dried under vacuum. Final purification was done on a silica gel column (100-200 mesh) using mixture of ethyl acetate and toluene as eluent. Methyl ester of these amino acid based amphiphile (C17H35-Tyr-OMe/ C13H27-Tyr-OMe/ C9H19-Tyr-OMe/ C17H35-Phe-OMe or C₁₇H₃₅-Leu-OMe) is finally characterized through ESI mass spectroscopy, 300 MHz ¹H NMR spectroscopy, the details of which are given below.

C₁₇H₃₅-Tyr-OMe: ¹HNMR (300 MHz, CDCl₃, TMS): $\delta = 6.96-6.93$ (d, 2H, J = 9, Tyr ring H), 6.76-6.73 (d, 2H, J = 9, Tyr ring H), 5.89-5.86 (d, 1H, J = 9, amide H), 4.90-4.83 (q, 1H, J = 3, Tyr C^{α} Hs), 3.73 (s, 3H, -OCH₃), 3.12-2.97 (m, 2H, Tyr C^{β} Hs), 2.20-2.14 (t, 2H, J = 7.5, -CO-CH₂), 1.61 (br., 8H, -(CH₂)₂-), 1.25 (br., 22H, -(CH₂)₂-), 0.89-0.85 (t, 3H, J = 6.5, -CH₃).

MS (35eV): m/z (%): 484 [M + Na⁺]

C₁₃H₂₇-Tyr-OMe: ¹HNMR (300 MHz, CDCl₃, TMS): $\delta = 6.96-6.93$ (d, 2H, J = 9, Tyr ring H), 6.76-6.73 (d, 2H, J = 9, Tyr ring H), 5.89-5.86 (d, 1H, J = 9, amide H), 4.90-4.83 (q, 1H, J = 3, Tyr C^{α}), 3.73 (s, 3H, OCH₃), 3.12-2.97 (m, 2H, J = 5.16, Tyr C^{β}), 2.20-2.14 (t, 2H, J = 7.5, -CO-CH₂), 1.61 (br., 8H, -(CH₂)₂-), 1.25 (br., 14H, -(CH₂)₂-), 0.89-0.85 (t, 3H, J = 6.5, -CH₃).

MS (35eV): m/z (%): 428 [M + Na⁺]

C₉H₁₉-Tyr-OMe: ¹HNMR (300 MHz, CDCl₃, TMS): $\delta = 6.95-6.92$ (d, 2H, J = 9, Tyr ring H), 6.75-6.72 (d, 2H, J = 9, Tyr ring H), 5.89-5.86 (d, 1H, J = 9, amide H), 4.89-4.83 (m, 1H,

Tyr C^{α} Hs), 3.73 (s, 3H, -OCH₃), 3.06-2.98 (m, 2H, Tyr C^{β} Hs), 2.19-2.14 (t, 2H, J = 7.5, -CO-CH₂), 1.91-1.80 (m, 14H, -(CH₂)₂-), 0.89-0.84 (t, 3H, J = 6.5, -CH₃).

MS (35 eV): m/z (%): 372 [M+Na⁺]

C₁₇H₃₅-Phe-OMe: ¹HNMR (300 MHz, CDCl₃, TMS): $\delta = 7.099$ (br, 5H, Phe ring H), 5.82 (s, 1H, amide H), 4.90-4.83 (m, 1H, Phe C^{α}), 3.73 (s, 3H, -OCH₃), 3.139 (br, 2H, Phe C^{β}), 2.18 (br, 2H, -CO-CH₂-), 1.60 (br., 8H, -(CH₂)₂-), 1.25 (br., 24H, -(CH₂)₂-), 0.89-0.85 (t, 3H, J = 6.5, -CH₃).

MS (35 eV): m/z (%): 468 [M + Na⁺]

 $C_{17}H_{35}$ -Leu-OMe:¹HNMR (300 MHz, CDCl₃, TMS): $\delta = 0.96$ -0.95 (d, 6H, J = 9, Leu C^{δ}), 5.9-5.88 (d, 1H, J = 9, amide -NH), 4.62-4.58 (m, 1H, Leu C^{α} H), 1.63 (m, 2H, Leu C^{β} H),1.70 (m, 1H, Leu C^{γ}), 3.73 (s, 3H, -OCH₃), 2.18-2.13 (t, 2H, J = 9, -CO-CH₂-), 1.55 (br., 2H,-CH₂-), 1.26 (br., 28H, -(CH₂)₁₄-), 0.89-0.85 (t, 3H, J = 6, -CH₃).

MS (35 eV): m/z (%): 444 [M + Na⁺]

Synthesis of amphiphiles ($C_{17}H_{35}$ -Tyr-OH/ $C_{13}H_{27}$ -Tyr-OH/ $C_{9}H_{19}$ -Tyr-OH/ $C_{17}H_{35}$ -Phe-OH or $C_{17}H_{35}$ -Leu-OH). Typically, for the synthesis of amino acid based amphiphile ($C_{17}H_{35}$ -Tyr-OH), an aqueous NaOH (10 ml, 2N) solution is added dropwise to the methanolic solution (25 ml) of purified ester of amphiphile ($C_{17}H_{35}$ -Tyr-OMe) and the progress of this basic hydrolysis is monitored via thin layer chromatography (TLC). The reaction mixture is then stirred (magnetically) further for 12 h and subsequently methanol is removed. The rest alkaline aqueous part containing sodium salt of amphiphiles is mixed with 30 ml of water followed by washing with diethyl ether (2 × 20 ml) to remove any unreacted ester. Then the pH of the aqueous layer was adjusted to 2 using 2N HCl and organic compound (amphiphile) was further extracted with ethyl acetate (3 × 20 ml). The extract was dried over anhydrous sodium sulphate, and evaporated by rotary evaporator and finally dried under vacuum to isolate the amphiphile. Other amphiphiles ($C_{13}H_{27}$ -Tyr-OH, $C_{9}H_{19}$ -Tyr-OH, $C_{17}H_{35}$ -Phe-OH and $C_{17}H_{35}$ -Leu-OH) are prepared by similar procedure. ¹H NMR spectroscopy, ESI mass spectroscopy are used to characterize the resultant amphiphiles, the details of which are mentioned below.

C₁₇**H**₃₅-**Tyr-OH:** Yield = (79%); ¹HNMR (300MHZ, CDCl₃ + (D₆)DMSO, TMS): δ = 6.93-6.90 (d, 2H, J = 9, Tyr ring-Hs), 6.70-6.67 (d, 2H, J = 9, Tyr ring Hs), 6.03-6.00 (d, 1H, J = 9, amide -NH), 4.71-4.69 (m, 1H, Tyr C^αH), 3.07-3.05 (d, 2H, J = 6, Tyr C^βH), 2.12-2.07 (t, 2H, J = 9, -CO-CH₂), 1.52 (br., 2H, -CH₂), 1.21 (br., 28H, -(CH₂)₄-), 0.83-0.79 (t, 3H, J = 6, -



¹³CNMR (500MHZ, (D₆)DMSO, TMS): $\delta = 129.87$ (d, 2C,Tyr ortho), 114.84 (d,2C,Tyr meta),127.68 (s,1C,ipso),155.02 (s,1C,Tyr para),173.26(s,1C,CO₂H),172.06 (s,1C, -CONH-), 53.57(s,1C,Tyr C^{α}),36.01(s,1C,Tyr C^{β}),35.03(s,1C,Acid C^{α}),29-28.47(m,12C,Alkyl chain), 25.14 (s,1C, Acid C^{β}), 22.03(s,1C, Acid C¹⁷), 31.24 (s, 1C,Acid C¹⁶), 13.87(s,1C,C¹⁸)





C₁₃**H**₂₇-**Tyr-OH:** Yield = (81%); ¹HNMR (300MHZ, CDCl₃ + (D₆)DMSO, TMS): δ = 6.93-6.90 (d, 2H, J = 9, Tyr ring-Hs), 6.70-6.67 (d, 2H, J = 9, Tyr ring Hs), 6.03-6.00 (d, 1H, J = 9, amide -NH), 4.71-4.69 (m, 1H, Tyr C^αH), 3.07-3.05 (d, 2H, J = 6, Tyr C^β), 2.12-2.07 (t, 2H, J = 9, -CO-CH₂), 1.52 (br., 2H, -CH₂), 1.21 (br., 28H, -(CH₂)₄-), 0.83-0.79 (t, 3H, J = 6, -CH₃).



¹³CNMR (300MHZ, CDCl₃ + (D₆)DMSO, TMS): δ = 128.44 (d, 2C,ortho), 113.45 (d,2C, meta),126.04 (s,1C,ipso),154.45 (s,1C,para),171.83(s,1C,CO₂H),170.98 (s,1C, -CONH-),

52.04 (s,1C,Tyr C^α),34.04(s,1C,Tyr C^β),33.96(s,1C,Acid C^α),27.75-27.33(m,8C,Alkyl chain), 23.85 (s,1C, Acid C^β), 20.77(s,1C, Acid C¹³), 29.99 (s, 1C,Acid C¹²), 12.47(s,1C,C¹⁴)



MS (35eV): m/z (%):392 [M+H⁺],414 [M+Na⁺]

C₉**H**₁₉-**Tyr-OH:** Yield = (74%); ¹HNMR (300MHZ, CDCl₃ + (D₆)DMSO, TMS): $\delta = 6.99$ -6.96 (d, 2H, J = 9, Tyr ring-Hs), 6.73-6.71 (d, 2H, J = 6, Tyr ring Hs), 6.66-6.65 (d, 1H, J = 9, amide -NH), 4.71-4.64 (m, 1H, Tyr C^αH), 3.07-3.05 (d, 2H, J = 6, Tyr C^β), 2.18-2.13 (t, 2H, J = 9, -CO-CH₂), 1.55 (br., 2H, -CH₂), 1.26 (br., 12H, -(CH₂)₄-), 0.89-0.85 (t, 3H, J = 6, -CH₃).



¹³CNMR (300MHZ, CDCl₃ + (D₆)DMSO, TMS): δ = 120.57 (d, 2C,ortho), 113.63 (d,2C, meta),126.12 (s,1C,ipso),154.58 (s,1C,para),171.96(s,1C,CO₂H),171.10 (s,1C, -CONH-), 52.13(s,1C,Tyr C^α),35(s,1C,Tyr C^β),34.15(s,1C,Acid C^α),27.71-27.46(m,12C,Alkyl chain), 23.99 (s,1C, Acid C^β), 20.91(s,1C, Acid C⁹), 30.10 (s, 1C,Acid C⁸), 12.6(s,1C,C¹⁰).





C₁₇**H**₃₅-**Phe-OH:** Yield = (82%); ¹HNMR (300 MHz, CDCl₃, TMS): δ = 7.099 (br, 5H, Phe ring H), 5.82 (s, 1H, amide H), 4.90-4.83 (m, 1H, Phe C^{α}), 3.139 (br, 2H, Phe C^{β}), 2.18 (br, 2H, -CO-CH₂-), 1.60 (br., 8H, -(CH₂)₂-), 1.25 (br., 24H, -(CH₂)₂-), 0.89-0.85 (t, 3H, J = 6.5, - CH₃).



¹³CNMR (300MHZ, CDCl₃, TMS): δ = 128.71 (d, 2C, ortho), 129.51 (d,2C, meta),135.92 (s,1C,ipso),127.26 (s,1C,para),174.73(s,1C,CO₂H),174.11(s,1C, -CONH-), 53.24(s,1C,Phe C^α),37.45(s,1C,Phe C^β),36.58(s,1C,Acid C^α),29.83-29.29(m,12C,Alkyl chain), 25.71 (s,1C, Acid C^β), 22.81(s,1C, Acid C¹⁷), 32.05 (s, 1C,Acid C¹⁶), 14.22(s,1C,C¹⁸)





C₁₇**H**₃₅-Leu-OH: Yield = (75%); ¹HNMR (300MHZ, CDCl₃, TMS): δ = 0.96-0.95 (d, 6H, J = 9, Leu C^δ), 5.9-5.88 (d, 1H, J = 9, amide -NH), 4.62-4.58 (m, 1H, Leu C^αH), 1.63 (m, 2H, Leu C^βH), 1.70 (m, 1H, Leu C^γ), 2.18-2.13 (t, 2H, J = 9, -CO-CH₂-), 1.55 (br., 2H, -CH₂-), 1.26 (br., 28H, -(CH₂)₁₄-), 0.89-0.85 (t, 3H, J = 6, -CH₃).



¹³CNMR (300MHZ, CDCl₃, TMS): $\delta = 22.95$ (d, 2C,Leu C^δ), 22.81 (d, 1C,Leu C^γ),127.68 (s,1C,ipso),155.02(s,1C,Tyr para),176.73(s,1C,CO₂H),174.19(s,1C,-CONH-), 51.02 (s,1C,Leu C^α), 41.36(s,1C,Leu C^β),36.62(s,1C,Acid C^α),31.02-29.22(m,12C,Alkyl chain), 25.75 (s,1C, Acid C^β), 22.03(s,1C, Acid C¹⁷), 31.24 (s, 1C,Acid C¹⁶), 14.22(s,1C,C¹⁸).







Preparation of salt of the amphiphile. Typically, 10 mg of $C_{17}H_{35}$ -Tyr-OH was first dissolved in 3 mL of methanol in a small beaker; subsequently water (5 mL) was added with swirling. 0.025 N NaOH was added to it dropwise with subsequent pH measurement. At certain point, pH abruptly increases to 8.3. The salt of the amphiphile is collected after solvent removal under vacuum.

Table S1. Gelation study of the salts of the amphiphiles.

Amphiphile	Solvent	Solvent composition (v/v)	Concentration (wt %)	Appearance
C ₁₇ H ₃₅ -Tyr-ONa				TS
C ₁₃ H ₂₇ -Tyr-ONa				TS
C ₉ H ₁₉ -Tyr-ONa	Water/DMSO	4:1	0.5	OS
C ₁₇ H ₃₅ -Phe-ONa				OS
C ₁₇ H ₃₅ -Leu-ONa				OS

TS = transparent solution, OS = opaque solution.

Table S2: Reusability of gel in three cycles to prepare silica gel and its corresponding yield.

No. of cycles	%yield	
1 st	96	
2^{nd}	85.4	
3 rd	76.8	



Figure S1. Photograph showing the solution of salts of amphiphiles in water/DMSO (4:1) mixed solvent.



Figure S2. FTIR spectra of (A) $C_{17}H_{35}$ -Phe-OH xerogel obtained from water/DMSO and (B) $C_{17}H_{35}$ -Phe-OH in CHCl₃ solution.



Figure S3. UV-vis spectra of (A) $C_{17}H_{35}$ -Tyr-OH gel in (4:1) water/DMSO and (B) its solution in (2:1) water/DMSO.



Figure S4. XRD patterns of (A) xerogel of $C_{17}H_{35}$ -Tyr-OH obtained from water/DMSO at 0.5%, (B) neat $C_{17}H_{35}$ -Tyr-OH obtained by casting from its DMSO solution.



Figure S5. FTIR spectra of (A) composite gel and (B) silica aerogel obtained from $C_{17}H_{35}$ -Tyr-OH gel in water/DMSO.



Figure S6. FESEM image of silica nanostructures obtained using C₉H₁₉-Tyr-OH gel as template in water/DMSO (4:1) mixed solvent.



Figure S7: TGA thermograms of (A) dried composite gel and (B) silica aerogel.

Calculation of washable organic moiety present in the composite gel.

We assume that after heating to 600 °C, we obtain materials of similar nature for the dried silica gel and composite gel. We observe that 13% of physisorbed materials are removed after heating the dried silica gel at 600 °C. The remaining 87% material is silica. The weight loss of composite gel is 46%. Therefore, for the composite gel is the amount of physisorbed material is (13/87x54) = 8%. Therefore the total amount of organic moiety removed after methanol washing is 38%.



Figure S8: N₂ adsorption/desorption isotherm of silica aerogel.



Figure S9. FESEM image of silica nanostructures obtained after using $C_{17}H_{35}$ -CO₂H as catalyst in water/DMSO (4:1) mixture as solvent.



Figure S10: NMR spectrum of recovered $C_{17}H_{35}$ -Tyr-CO₂H after silica formation.

Recovered C₁₇H₃₅-Tyr-OH after gel based catalysis: ¹HNMR (300MHz, CDCl₃ + (D₆)DMSO, TMS): $\delta = 6.99-6.96$ (d, 2H, J = 9, Tyr ring-Hs), 6.63-6.60 (d, 2H, J = 9, Tyr ring Hs), 9.15 (d, 1H, amide -NH), 4.32-4.27 (m, 1H, Tyr C^aH), 2.92-2.86 (s, 2H, J = 6, Tyr C^β), 2.07-1.99 (t, 2H, J = 9, -CO-CH₂), 1.40-1.37 (br., 2H, -CH₂-), 1.22 (br., 30H, -(CH₂)-), 0.84-0.81 (t, 3H, J = 6, -CH₃).



Figure S11: FESEM image of the silica gel formed after 2^{nd} cycle using recovered $C_{17}H_{35}$ -Tyr-OH after 1^{st} cycle in water/DMSO at 0.5%.



Figure S12: FTIR spectra of SiO₂-ZrO₂ and pure ZrO₂ nanostructures



Figure S13: FESEM-EDX analysis of SiO₂-ZrO₂ nanostructures.



Figure S14: XRD pattern of (A) SiO₂-ZrO₂ and (B) neat ZrO₂ maintaining at their normalized intensity.



Figure S15: FESEM image of (A) SiO_2 -Zr O_2 and (B)Zr O_2 obtained using $C_{17}H_{35}$ -Tyr-OH gel (0.5%) in (4:1) water/DMSO mixture.



Figure S16: N₂ adsorption/desorption isotherm of SiO₂-ZrO₂ nanostructures



Figure S17: (A) Successive UV-vis absorption spectra of the blank photodegradation of methylene blue and using (B) neat ZrO_2 photocatalyst and (C) SiO_2 - ZrO_2 mixed oxide.