Supplementary Information:

Secondary structure and rigidity in model proteins

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п	dry	wet	literature
GFP	1.473	1.475	1.34 -1.46 ¹
LYS	1.472	1.484	$1.48^{2} \\ 1.51^{3} \\ 1.538 - 1.575^{4} \\ 1.45^{5}$
BSA	1.480	1.487	$\frac{1.582^{6}}{1.4^{7}}$
МУО	1.478	1.483	1.549 ⁶
proteins			1.35-1.55 ⁸ 1.45 ⁵

Table S1: Refractive index of proteins measured in our experiments. A comparison with literature values is also provided.

In agreement with terahertz (THz) time domain spectroscopy data reported for Cytocrome C samples⁹ the refractive index of hydrated proteins is slightly higher than that of the dry system. The variation of n with temperature can be neglected^{10, 11}

ρ	ρ wet [g/cm ³]	% Δρ dry to wet	% Δρ T _r (room) to T _c (cryogenic)
GFP	1.220	7.3%	$T_r=300K, T_c=100K$ -4.5% ¹²
LYS	1.227 ¹³	(298K) 7.3% ¹³	$T_r=298K, T_c=100K$ -1.8% ¹⁴
BSA	1.170 ¹⁵	7.3%	$T_r=300K, T_c=100K$ -1.8% ¹⁶
MYO	0.976 ¹⁷	7.3%	$T_r=277K, T_c=77K$ -2.9% ¹⁷

Table S2. Density values used to calculate the Young's modulus of hydrated samples. Variation of density of protein crystals as a function of hydration and temperature (literature data) used to calculate density of dry samples and the temperature dependence of ρ .

Table S3 Poisson's ratio σ reported in literature for different proteins

protein	technique	σ
Tetragonal lysozyme	Brillouin spectroscopy	0.33 as observed in many polymers ^{3, 18}
Tetragonal lysozyme	Ultrasonic pulse-echo	0.37 ¹³
Collagen	Brillouin spectroscopy	II 0.42 ; $\perp 0.26^{19}$
lysozyme	Derived from X-Ray	0.25^{20}
Tetragonal lysozyme	Ultrasonic pulse-echo	0.42^{21}
Lysozyme, haemoglobin, myoglobin	ultrasound	0.47 ²²

Table S4 Literature data at room temperature (~295K) for longitudinal sound velocity c_L and Young's modulus *E* of biological systems with different secondary structures. c_L or *E* values for dry and hydrated proteins are reported in separated columns. Data in each column refer to samples with comparable hydration levels. For the hydrated samples the hydration ratio h=g water/g protein (similar to our samples) is given in parenthesis.

nrotein	Secondary	tochniquo	c_L ,[m/s]		Young's modulus <i>E</i> [GPa]	
protein	structure	teeninque	DRY	HYDR.	DRY	HYDR.
Bombyx mori silk (dry, in oil)	30% pleated β-sheet	Brillouin scattering	4700 ¹⁹		30.9 ¹⁹	
β-Keratin 28% from feather rachis (dry in oil)	β structure	Brillouin scattering	3710 ¹⁹		19.3 ¹⁹	
β-lactoglobulin	β barrel	X-ray scattering		$2900 \pm 160 \\ (h=0.5) \\ 2860 \pm 110^{23} \\ (h=1)$		
Maltose Binding Protein (MBP)	α/β	Brillouin neutron scattering	3780 ± 130^{24}			
Tetragonal lysozyme	α/β	Ultrasonic pulse-echo method	[110] v1 3097 [001] v4 3149 ²¹	v1 2070 \pm 34 v4 2001 \pm 33 ²¹ (h=0.47)	7.25 ²¹	3.21 ²¹ (h=0.47)
Triclinic lysozyme	α/β	Resonance method			$\begin{matrix} [011] \ 10 \pm 1.1 \\ [01\overline{1}] \ 11.2 \pm 1.3^{25} \end{matrix}$	
monoclinic P2, lysozyme						$[010] \\ 2.5-3.5^{26} \\ (h=0.3)$
Ribonuclease crystal	α/β	Ultrasound		$1784 \pm 72^{22} \\ (h{\sim}0.5)$		
α-Keratin 25% from Porcupine quill (dry in oil)	α-helix	Brillouin scattering	3660 ¹⁹		18.8 ¹⁹	
Cytochrome C	α-helix	X-ray scattering	$3458 \pm 42^{\ 27}$			
Paramyosin from Molluscan catch muscle	supercoiled α-helix	Brillouin scattering	3530 ²⁸		17.4 ²⁸	
Hemoglobin crystal	α-helix	Ultrasound ²² / Resonance method		1828 ²² (h~0.5)		$[110] \text{ or} \\ [\overline{1}10] 5 \pm 1^{25} \\ (h=0.3)$
myoglobin monoclinic	α-helix	Resonance method				$ [001] 3.6\pm1.3^{25} (h=0.3) $
Whale myoglobin amorphous films	α-helix	Resonance method				2-3.6 ²⁶ (h=0.3)

Morozov et al.²⁶ reported also measurements of *E* at low temperature for hydrated samples of amorphous whale myoglobin and cross-linked BSA amorphous films. Their results are in agreement with our estimates: at 150K MYO systems (E=7.8 GPa) are softer than the BSA (E=13.2 GPa) ones.

	Number of residues in helices	Total HBs in helices	Number of HBs/residues in helices
BSA	450	375	0.83
MYO	123	102.5	0.83
	Number of residues in β-barrel	Total HBs in β-barrel	Number of HBs/residues in β-barrel
GFP	121	109 29	0.9

Table S5 Number of HBs per residue in the secondary structural units for MYO, BSA (helices) as well as for GFP (β-barrel)

References

- 1. K. Suhling, J. Siegel, D. Phillips, P. M. W. French, S. Lévêque-Fort, S. E. D. Webb and D. M. Davis, *Biophysical journal*, 2002, **83**, 3589-3595.
- 2. J. Vörös, *Biophysical journal*, 2004, **87**, 553-561.
- 3. S. Speziale, F. Jiang, C. L. Caylor, S. Kriminski, C. S. Zha, R. E. Thorne and T. S. Duffy, *Biophysical journal*, 2003, **85**, 3202-3213.
- 4. B. Cervelle, F. Cesbron, J. Berthou and P. Jolles, *Acta Crystallographica Section A*, 1974, **30**, 645-648.
- 5. K. L. Prime and G. M. Whitesides, *Journal of the American Chemical Society*, 1993, **115**, 10714-10721.
- 6. M. Andersen and S. Nir, *Polymer*, 1977, 18, 867-870.
- 7. A. G. Markelz, A. Roitberg and E. J. Heilweil, *Chemical Physics Letters*, 2000, **320**, 42-48.
- 8. G. B. Sigal, C. Bamdad, A. Barberis, J. Strominger and G. M. Whitesides, *Analytical Chemistry*, 1996, **68**, 490-497.
- 9. Y. He, J. Y. Chen, J. R. Knab, W. Zheng and A. G. Markelz, *Biophysical journal*, 2011, **100**, 1058-1065.
- 10. A. V. Svanidze, S. G. Lushnikov and S. Kojima, Jetp Lett., 2007, 84, 551-555.
- 11. A. V. Svanidze, V. P. Romanov and S. G. Lushnikov, Jetp Lett., 2011, 93, 409-414.
- 12. F. Yang, L. G. Moss and G. N. Phillips, *Nat Biotech*, 1996, **14**, 1246-1251.
- 13. H. Koizumi, M. Tachibana and K. Kojima, *Phys Rev E*, 2006, **73**, 041910.
- 14. A. C. M. Young, R. F. Tilton and J. C. Dewan, *Journal of molecular biology*, 1994, **235**, 302-317.
- 15. R. J. McClure and B. M. Craven, Journal of molecular biology, 1974, 83, 551-555.
- 16. A. Merlino, private communication, antonello.merlino@unina.it.
- 17. P. Urayama, G. N. Phillips Jr and S. M. Gruner, *Structure*, 2002, 10, 51-60.
- 18. M. Tachibana, K. Kojima, R. Ikuyama, Y. Kobayashi and M. Ataka, *Chemical Physics Letters*, 2000, **332**, 259-264.
- 19. S. Cusack and A. Miller, *Journal of molecular biology*, 1979, **135**, 39-51.
- 20. A. A. Chernov, Journal of structural biology, 2003, 142, 3-21.
- 21. H. Koizumi, M. Tachibana and K. Kojima, *Phys Rev E*, 2009, **79**, 061917.
- 22. C. Edwards, S. B. Palmer, P. Emsley, J. R. Helliwell, I. D. Glover, G. W. Harris and D. S. Moss, *Acta Crystallographica Section A*, 1990, **46**, 315-320.
- 23. K. Yoshida, S. Hosokawa, A. Q. R. Baron and T. Yamaguchi, *The Journal of chemical physics*, 2010, **133**, 134501..
- 24. A. Paciaroni, A. Orecchini, M. Haertlein, M. Moulin, V. Conti Nibali, A. De Francesco, C. Petrillo and F. Sacchetti, *The Journal of Physical Chemistry B*, 2012, **116**, 3861-3865.
- 25. V. N. Morozov and T. Y. Morozova, *Biopolymers*, 1981, 20, 451-467.
- 26. V. N. Morozov and S. G. Gevorkian, *Biopolymers*, 1985, 24, 1785-1799.

- 27. B. M. Leu, A. Alatas, H. Sinn, E. E. Alp, A. H. Said, H. Yavas, J. Zhao, J. T. Sage and W. Sturhahn, The Journal of chemical physics, 2010, 132, 085103
- 28. R. Harley, D. James, A. Miller and J. W. White, *Nature*, 1977, 267, 285-287.
- 29. V. Helms, T. P. Straatsma, J. A. McCammon J. Phys. Chem. B 1999, 103, 3263-3269