# **Electronic Supplementary Material**

## Hydrophobin: fluorosurfactant-like properties without fluorine

Roberto Milani,<sup>\*a,b</sup> Evanthia Monogioudi,<sup>a</sup> Michele Baldrighi,<sup>c</sup> Gabriella Cavallo,<sup>b,c</sup> Valentina Arima,<sup>d</sup> Lucia Marra,<sup>d</sup> Alessandra Zizzari,<sup>e</sup> Ross Rinaldi,<sup>d,e</sup> Markus Linder,<sup>\*a,f</sup> Giuseppe Resnati<sup>\*b,c</sup> and Pierangelo Metrangolo<sup>\*a,b,c</sup>

<sup>a</sup> VTT Technical Research Centre of Finland, Tietotie 2, Espoo, FI-02044 VTT, Finland. E-mail: roberto.milani@vtt.fi

<sup>b</sup> Center for Nano Science and Technology@PoliMi, Istituto Italiano di Tecnologia, Via Giovanni Pascoli, 70/3, I-20133 Milano, Italy

<sup>c</sup> NFMLab-DCMIC "Giulio Natta", Politecnico di Milano, Via Mancinelli 7, I-20131 Milano, Italy. E-mail: pierangelo.metrangolo@polimi.it

<sup>d</sup> NNL - Institute of Nanoscience (NANO), CNR, Via per Arnesano 16, I-73100 Lecce, Italy

<sup>e</sup> University of Salento, Via Provinciale Lecce-Monteroni, I-73100 Lecce, Italy

<sup>f</sup> Aalto University, School of Chemical Technology, Kemistintie 1, P.O. Box 16100, FI-00076 AALTO, Espoo, Finland.

#### 1. Measurement of HFBI isoelectric point

The isoelectric point of HFBI was measured experimentally with a Nano-ZS zetasizer (Malvern instruments, UK) by performing titrations with 0.01M and 0.1M NaOH on 0.10 mg ml<sup>-1</sup> protein solutions in mQ water.

The measurements were highly reproducible, and the experimental pI obtained as an average of three different measurements was 6.08. For comparison, the theoretical pI calculated with Expasy software was 5.7.

In Fig. S1 is reported a typical experimental titration curve.



**Fig. S1.** Titration curve for the measurement of HFBI pI. The final result was obtained as an average of three different measurements.

### 2. Tensiometric measurements at varying pH values

Tensiometric measurements at several pH values were performed by the pendant drop method. A 15  $\mu$ l droplet of GSV90 was injected into 900  $\mu$ l of freshly prepared 0.10 mg ml<sup>-1</sup> HFBI solutions in 20 mM aqueous buffers (citrate buffer, pH 4; sodium acetate buffer, pH5.5; phosphate buffer, pH 7; boric acid buffer, pH 10) in a quartz cuvette, and the measurement was started immediately. Droplet shape fitting was performed by the Young–Laplace method. The curves reported in **Fig. S2** illustrate the interface tension vs. time.

The results are very similar in all cases.



**Fig. S2.** Plot of interface tension vs. time for GSV90 droplets immersed in buffered aqueous solutions of HFBI (0.10 mg/ml) at different pH values.

#### 3. Detailed description of the microfluidic circuit

The microreactor was a glass-based microdevice with overall dimension of (2.5x6) cm. A B270 glass sheet with photoresist and chrome layers (TELIC, USA) was used as substrate. Uncoated glass was chosen for its chemical inertness, hydrophilicity and well established fabrication methods. The fabrication process consisted of UV exposure, heat treatment and HF etching. Two glass plates were etched using specular mask designs and fused by thermal bonding so that an approximately circular cross section of the orifice was obtained. One of the two glass sides was then drilled before bonding, in order to obtain 700  $\mu$ m diameter holes. HPLC connectors were used to flow liquids inside the reactor.

Droplets were generated from flow focusing geometry, consisting of three channels converging into a main channel via a narrow orifice. In this configuration the dispersed phase, contained in the middle channel, is squeezed by continuous phase flows from the two opposing side channels. Both phases pass through the small orifice that is located downstream of the three channels. Finally, the stream of the dispersed phase becomes narrow and breaks into droplets. The droplet size is determined by the flow rates of the two phases and by the flow rate ratio, in addition to the channel geometries and the viscosities of the two phases.

The reactor geometry consisted of several parts:

- two inlets, one for water (continuous phase) and one for oil, FC-70, (dispersed phase) injection (width  $w_1$ = 420µm, depth  $d_1$  = 100 µm). The water channel had a bifurcation which converged at the nozzle and wa responsible of shear forces and generation of oil droplets;
- a classical flow focusing nozzle ( $w_2 = 100 \mu m$ ,  $d_2 = 80 \mu m$ ) enabling the production of oil droplets in a continuous water phase;
- a serpentine channel ( $w_3 = 420 \mu m$ ,  $d_3 = 100 \mu m$  and length  $L_3 \sim 15 cm$ );
- a collecting chamber ( $w_4 = 2cm$ ,  $d_4 = 100 \ \mu m$  and  $L_4 = 1.6cm$ ), for droplet coalescence evaluation.



Fig. S3. Schematic representation of the flow focusing microfluidic circuit employed.

Further images of the microfluidic circuit in the nozzle area and film formation on the walls of the circuit are reported below.



**Fig. S4.** Optical images at the nozzle for  $H_2O/FC-70$  at a) pH 10.4 and b) pH 13.0 without surfactant; for HFBI in  $H_2O/FC-70$  at c) pH 10.4 and d) pH 13.0; for Zonyl FSN-100 in  $H_2O/FC-70$  at e) pH 7.0 and f) pH 13.0. The scale bar in the figures is 350 µm.



**Fig. S5** Optical images in the serpentine region for HFBI in  $H_2O/FC-70$  at pH 10.4 a) frame 1 and b) frame 2 (acquired 0.13 s after frame 1); optical images of the serpentine region c) formation of HFBI film on microchannel walls at pH 10.4. Optical images in the serpentine region for HFBI in  $H_2O/FC-70$  at pH 13.0: d) frame 1, e) frame 2 (acquired 0.13 s after frame 1), and f) frame 3 (acquired 0.13 s after frame 2). The scale bar in the figures is 350 µm.