

## Controlling the number of dendrimers in dendrimicelle nanoconjugates from 1 to more than 100

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2. Determination of the micellar mass and aggregation number (Figure S1 & Table S2)
3. Determination of CMC (Figure S2 & Table S3)
4. CONTIN analysis of micellar size and size distribution; (Figure S3)
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## Experiment section

### 1. Materials

The diblock copolymer, poly(N-methyl-2-vinyl-pyridinium iodide)-*b*-poly(ethylene oxide) (P2MVP<sub>128</sub>-*b*-PEO<sub>477</sub>), was obtained by quaternization of poly(2-vinylpyridine)-*b*-poly(ethylene oxide) (P2MVP<sub>128</sub>-*b*-PEO<sub>477</sub>) (Polymer Source,  $M_w/M_n = 1.10$ ,  $M_n = 34.5$  k) following a procedure described elsewhere.<sup>1</sup> The degree of quaternization is about 87% determined by titration with poly(acrylic acid) (PAA, Polymer Source,  $M_w/M_n = 1.16$ ,  $M_n = 2.2$  k) in 20 mM NaHCO<sub>3</sub>/Na<sub>2</sub>CO<sub>3</sub> buffer at pH 10. The PAMAM dendrimers with sodium carboxylate surface groups were purchased from Sigma Aldrich (G0-G7) or Dendritech Inc. (G8, G9). The original solvent methanol was evaporated carefully under N<sub>2</sub> and the obtained compound was dissolved in nanopure water. The micelles were prepared by mixing solutions of P2MVP<sub>128</sub>-*b*-PEO<sub>477</sub> and PAMAM dendrimer in 20 mM NaHCO<sub>3</sub>/Na<sub>2</sub>CO<sub>3</sub> buffer at pH 10 ± 0.1.

### 2. Method

#### Dynamic and static light scattering

Light scattering at an angle of 90° was performed with an ALV light scattering-apparatus, equipped with a 300 mW cobalt samba DPSS laser operating at a wavelength of 532.0 nm. All measurements were performed at room temperature. Titrations were carried out using a Schott-Geräte computer-controlled titration setup to control sequential addition of titrant and cell stirring. After every dosage, the laser light-scattering intensity ( $I$ ) and the correlation function were recorded. The hydrodynamic radius and the scattered intensity are studied as a function of the mole fraction of positive charge fraction,  $f^+$ , which is defined as follows:

$$f^+ = \frac{n^+}{n^+ + n^-} \quad (1)$$

where  $n^+$  and  $n^-$  are the numbers of positive and negative charges.

The light scattering intensity is expressed as the excess Rayleigh ratio  $R_\theta$  divided by the total polymer concentration.  $R_\theta$  is obtained as

$$R_\theta = \frac{I_{sample} - I_{solvent}}{I_{toluene}} \times R_{toluene} \times \frac{n_{solvent}^2}{n_{toluene}^2} \quad (2)$$

where  $I_{sample}$  is the scattering intensity of the micellar solution and  $I_{solvent}$  is the intensity of the solvent.  $I_{toluene}$  is the scattering intensity of toluene,  $R_{toluene}$  is the known Rayleigh ratio of toluene ( $2.1 \cdot 10^{-2} \text{ m}^{-1}$ ) and  $n$  is the refractive of solvent (1.333) and toluene (1.497). The total polymer concentration is the sum of the concentrations of dendrimer and diblock copolymer contributing to micelle formation. The CUMULANT method<sup>2</sup> was used to analyze the mean apparent hydrodynamic radius ( $R_h$ ) as

$$R_h = kTq^2 / 6\pi\eta\Gamma \quad (3)$$

where  $q$  is the scattering vector,  $k$  is the Boltzman constant,  $T$  is the absolute temperature,  $\eta$  is the viscosity of the solvent, and  $\Gamma$  is the measured decay rate (second order cumulant) of the correlation function. The CONTIN method was<sup>3, 4</sup> used to analyze the distribution of particle (C3Ms) radii. The data were analyzed with AfterALV program (AfterALV 1.0d, Dullware), which provides  $\Gamma_i W_i$  as default output for each size fraction. Here, the intensity weighted contribution  $W_i$  is multiplied by  $\Gamma$ , as suggested by Petr Stepanek for the “equal-area representation”.<sup>5</sup> To facilitate comparisons, the absolute  $\Gamma_i W_i$  was normalized with the highest value of  $\Gamma_i W_i$  for each sample. For angular-dependent light scattering, the intensity was recorded at  $30^\circ \leq \theta \leq 140^\circ$  in intervals of  $5^\circ$ .

The Rayleigh ratio can be linked to the concentration and mass of the scattering objects:

$$\frac{K_R C}{R_\theta} = \frac{I}{M} \times \frac{I}{P(qR)} \times \frac{I}{S(q)} \quad (4)$$

where  $C$  is the weight concentration of micelles,  $M$  is their molecular mass, and  $R$  is the radius of the object that contribute to scatter light.  $P(qR)$  and  $S(q)$  are the form factor and the structure factor, respectively.  $K_R$  is an optical constant defined as:

$$K_R = \frac{4\pi^2 n^2}{N_{Av} \lambda_0^4} \left( \frac{dn}{dc} \right)^2 \quad (5)$$

where  $n$  is the refractive index of solvent,  $N_{Av}$  is Avogadro's number,  $\lambda_0$  is the wavelength of the incoming beam (532.0 nm), and  $dn/dc$  is the refractive index increment of the micelles.  $dn/dc$  of the micelles formed with PAMAM dendrimers at different generation is measured by a differential refractive index detector (Shodex RI-71), see the following table,

Generation	2	3	4	5	6	7	8	9
$dn/dc \cdot 10^4$ $m^3 kg^{-1}$	1.97	1.97	2.02	2.05	1.95	1.94	1.90	1.89

Table S1 Measured  $dn/dc$  of Gn-C3Ms formed at different dendrimer generation.

In our experiments, the scattering vector  $q = (4\pi n/\lambda_0) \sin(\theta/2)$  is approximately  $0.023 \text{ nm}^{-1}$  ( $\theta = 90^\circ$ ), so that  $qR$  is small for the micelles (which have a radius on the order of 25 nm). We therefore assume that  $P(qR)=1$ . At low concentrations, the structure factor can be approximated as

$$\frac{I}{S(q)} = I + 2B_2 \frac{C}{M} \quad (6)$$

where  $B_2$  is the second virial coefficient. Substitution into equation 4, we get

$$\frac{K_R C}{R_\theta} = \frac{I}{M} + 2B_2 \frac{C}{M^2} \quad (7)$$

By plotting  $K_R C/R_\theta$  versus  $C$ , we can obtain the molar mass  $M$  from the intercept. In our study,  $M$  corresponds to the molar mass of micelles,  $M_{micelle}$ , from which we can calculate the aggregation number of the micelles, see Figure S1.

### **Depolarized light scattering**

The depolarized light scattering experiments were carried out on the same DLS instrument mentioned above but with a Glan-Laser prism polarizer (Melles Griot, 03 PGL 301/A, extinction ratio  $< 5 \times 10^{-5}$ ) in front of the detector. The detection angle was fixed at  $90^\circ$  and the polarizer was adjusted to transmit only vertically ( $I_{vv}$ ) or only horizontally ( $I_{vh}$ ) polarized light, allowing to determine the depolarization ratio

$$\Delta_{vh} = \frac{I_{vh} - I_{vh}^0}{I_{vv} - I_{vv}^0} \quad (8)$$

with  $I$  and  $I^0$  denoting the scattering intensity from sample and solvent, respectively.

### **Small angle X-ray scattering**

Small angle X-ray scattering experiments were performed on a Ganesha lab instrument equipped with a GeniX-Cu ultra low divergence source producing X-rays with a wavelength of  $1.54 \text{ \AA}$  and a flux of  $1 \times 10^8 \text{ ph/s}$ . Scattering patterns were collected on a Pilatus 300K silicon pixel detector ( $487 \times 619$  pixels of  $172 \text{ \mu m}^2$ ) at two sample-to-detector distances corresponding to 730 and at 1530 mm covering a  $q$  range of  $6.7 \times 10^{-2} < q < 4.45 \text{ nm}^{-1}$ . The position of the beam center and the  $q$  range were calibrated using the diffraction peaks of silver behenate. The liquid samples were contained in 2 mm quartz capillaries sealed and fixed in a stainless steel holder kept at room temperature. The sample concentration was fixed at a dendrimer charge

concentration of 2mM in all cases. The scattering data were corrected for background contributions (such as scattering from the buffer solution), detector response and primary beam intensity fluctuations. The SAXS data were treated and analyzed using the software packages SAXSGUI and SASVIEW. The experimental scattering profiles given in Figure S3 are well described by a polydisperse core-shell sphere model, from which we extract the micellar core size  $R_{\text{core}}$  and shell thickness  $H_{\text{shell}}$ .

## Determination of the micellar mass and aggregation number

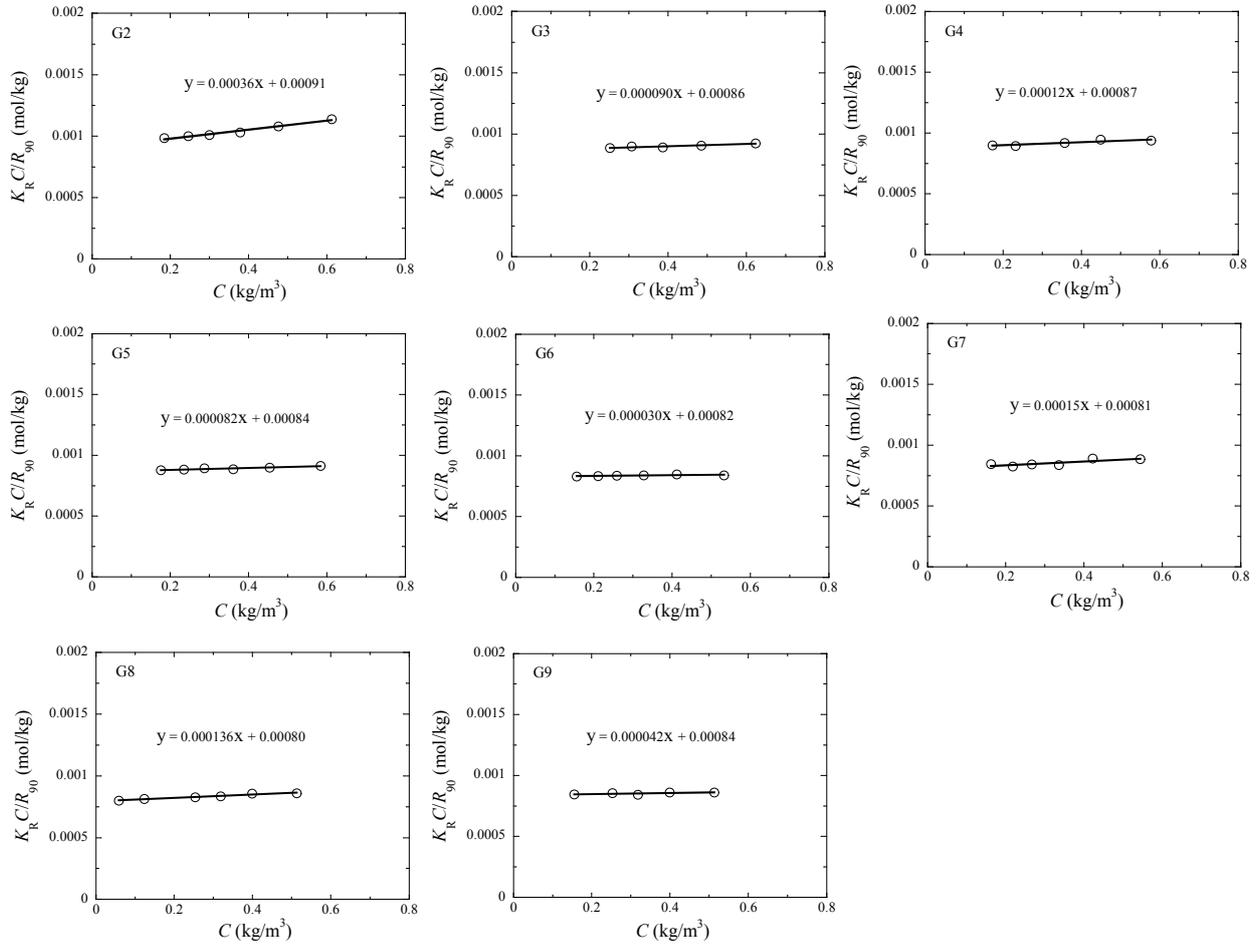


Figure S1  $K_R C/R_{90}$  is plotted as a function of total concentration of P2MVP<sub>128</sub>-*b*-PEO<sub>477</sub> polymer and dendrimer. ( $C = C_{\text{overall}} - \text{CMC}$ , and CMC is obtained from Table S3)

The intercept of the line gives the micellar mass,  $M_{\text{micelle}}$ . Here we take G5 line as an example, to show how to calculate the aggregation number:

$$M_{\text{micelle}} = 1/0.00084 = 1190.476 \text{ kg/mol} = 1190476 \text{ g/mol}$$

The charge numbers,  $Z$  and molecular weights of dendrimer and polymer are

$$Z_{G5} = 128, \quad M_{G5} = 26252, \text{ (g/mol)}$$

$$Z_{\text{polymer}} = 128 \cdot 0.87 = 111, \quad M_{\text{polymer}} = 50313 \text{ (g/mol)}$$

By taking into account of the PMC  $f^+$  shift, we get

$$N_{\text{polymer}} * 111 / (N_{G5} * 128 + N_{\text{polymer}} * 111) = 0.48 \quad (9)$$

Moreover, the molar masses of dendrimer, polymer and micelle are recorded as follows:

$$N_{G5} * 26252 + N_{\text{polymer}} * 50313 = 1190476 \quad (10)$$

Finally, we combine and solve equation 9 and 10, we find that

$N_{G5} = 15.0$  and  $N_{\text{polymer}} = 15.8$  Then we take 15 and 16 as average dendrimer and polymer numbers within one micelle.

The calculations with other generations follow the same strategy, and micellar mass and aggregation numbers are included in the below table:

Generation	2	3	4	5	6	7	8	9
$N_{\text{PAMAM}}$	108.2	56.1	28.5	15.0	8.0	4.0	2.1	1.0
$N_{\text{polymer}}$	15.5	16.1	15.5	15.8	15.8	16.0	16.0	15.0

Table S2 Micellar mass and calculated aggregations numbers of Gn-C3Ms.

### Determination of the CMC of G<sub>n</sub>-C3Ms

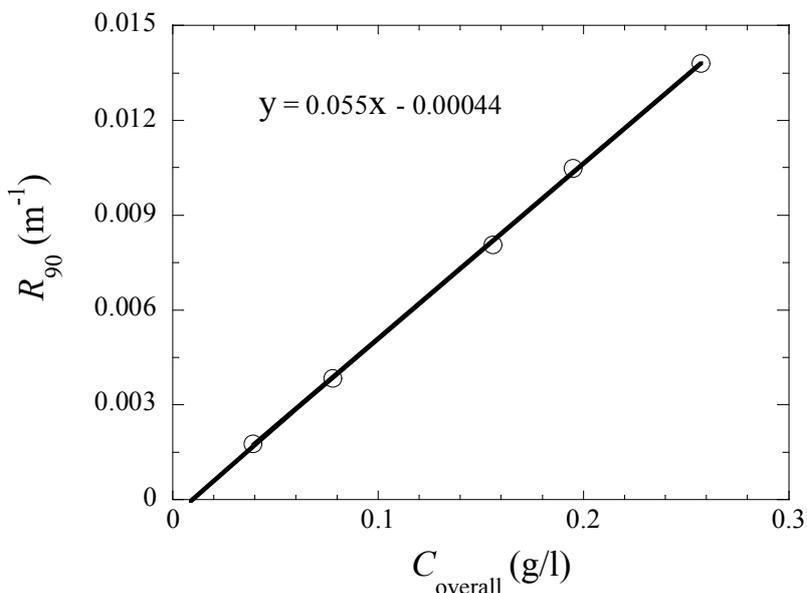


Figure S2 Intensity decay of G<sub>2</sub>-C<sub>3</sub>Ms solution upon diluting with NaHCO<sub>3</sub>/Na<sub>2</sub>CO<sub>3</sub> buffer. The open circles are the experimental data and the solid line is the fitting curve.

The intensity is represented as Rayleigh ratio  $R_{\theta}$ , which is subtracted with the intensity from buffer solution and corrected by the intensity of toluene as a reference, see experimental section, method, equation 2. The CMC is determined by extrapolating the decay line to zero intensity, and calculated from the fitting formula. The CMCs of the micelles formed at other generations are obtained following same strategy, and the numbers are included in the table below :

Generation	2	3	4	5	6	7	8	9
CMC g/l	0.008	0.010	0.010	0.007	0.010	0.010	0.006	0.007

Table S3 CMC of G<sub>n</sub>-C<sub>3</sub>Ms formed at different dendrimer generation.

## CONTIN analysis of micellar size and size distribution

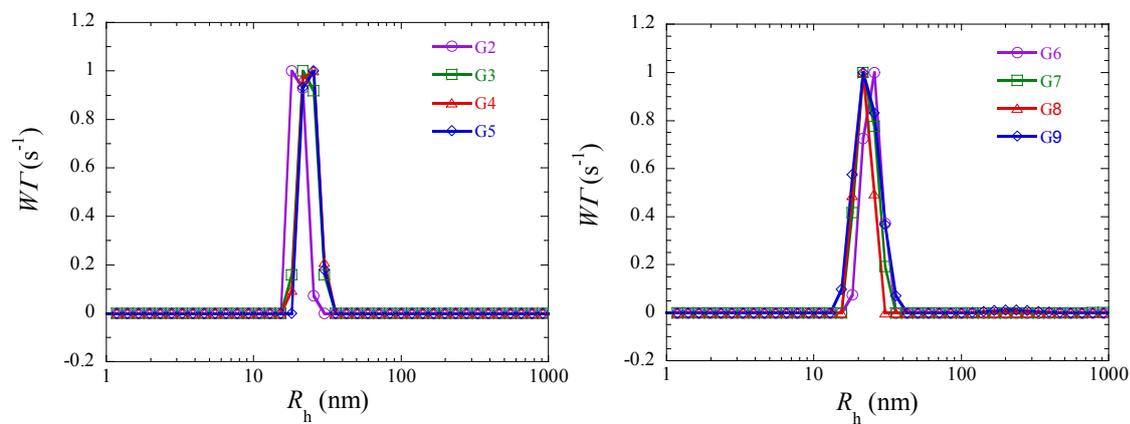


Figure S3 size and size distribution of micelles obtained by CONTIN analysis. For clarity reason, we plot the curve in two plots. In fact, there is no big difference between  $G_n$ -C3Ms at different generations.

## Determination of the micellar core size and shell thickness

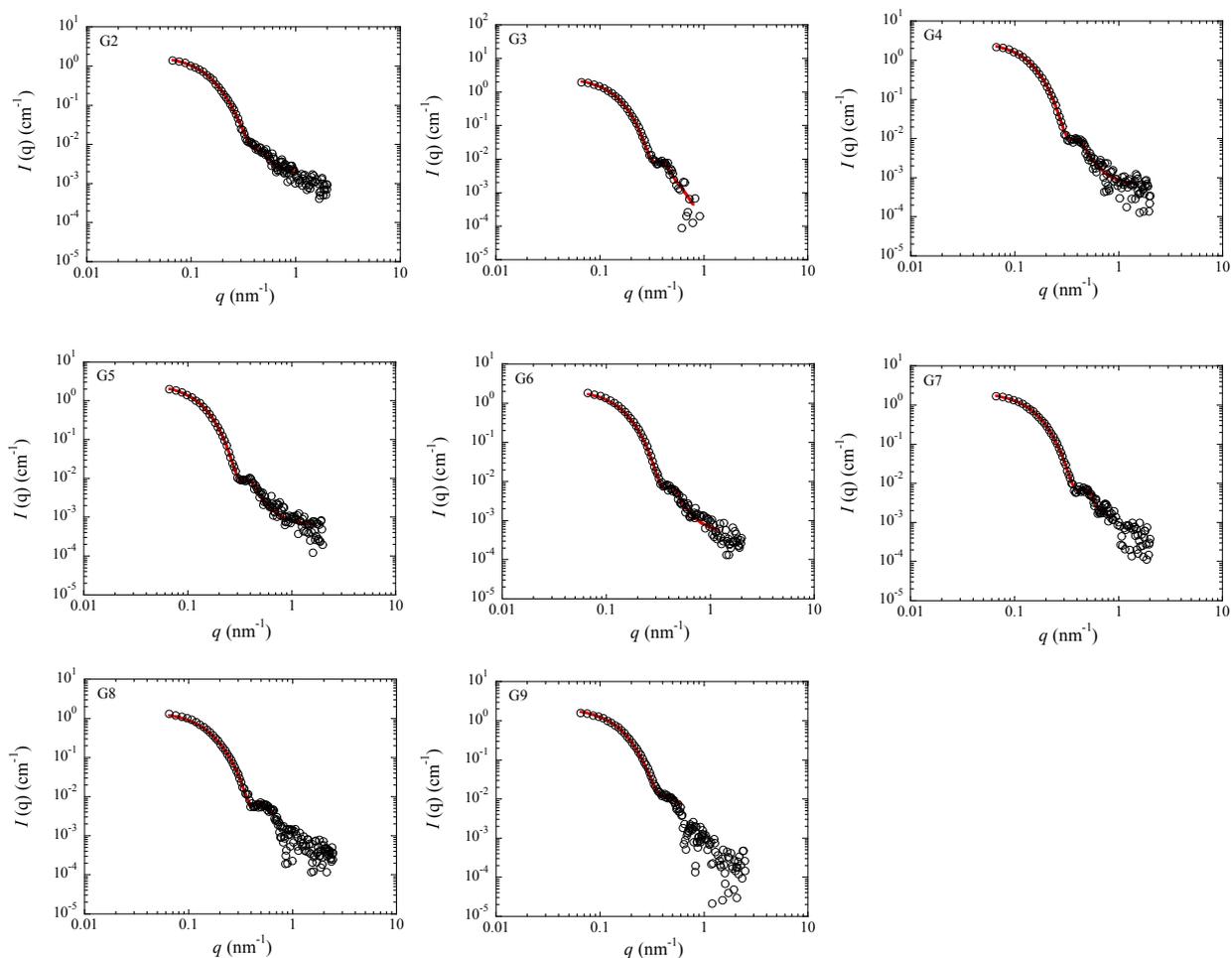


Figure S4 SAXS profiles of Gn-C3Ms at different dendrimer generations. The open circles shows the experimental data and the solid red line corresponds to the fits with a form factor for polydisperse (Gaussian distribution) core-shell spheres. The obtained core radius  $R_{\text{core}}$  and shell thickness  $H_{\text{shell}}$  are shown in main text, Table 1.

The fitting parameters are adapted from literature:<sup>6</sup>

$\rho_{\text{solvent}}$ ( $10^{10} \text{ cm}^{-2}$ )	$\rho_{\text{shell}}$ ( $10^{10} \text{ cm}^{-2}$ )	$\rho_{\text{core}}$ ( $10^{10} \text{ cm}^{-2}$ )	PD ratio (%) (core and shell)
9.37	9.45	11.11	16

Table S4 Fitting parameters applied in SasView program.  $\rho$ : scattering length density;

PD: polydispersity.

## Reference:

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