

# Colloidal Potentials Mediated by Specific Biomolecular Interactions

Gregg Duncan and Michael A. Bevan\*

Chemical & Biomolecular Engineering, Johns Hopkins University, Baltimore, MD 21218

## Supporting Information

### Calculating $K_D$ for Harmonic Well Potentials

The procedure for determining  $K_D$  was developed based on the work by Luo and Sharp.<sup>1</sup> For the case of a simple harmonic well potential,  $U_H$ , has the form,

$$U_H = U_M + 0.5\Delta q \cdot \mathbf{F} \cdot \Delta q \quad (\text{S1})$$

where  $\Delta q$  are the translational and rotational coordinates of reactant A, B, and their complex AB, and  $\mathbf{F}$  is the force constant matrix defined as,

$$\mathbf{F} = kT [\sigma_{ij}^2]^{-1} = k_s J_3 \quad (\text{S2})$$

where  $\sigma_{ij}$  is the coordinate fluctuation covariance matrix,  $k_s$  is the spring constant, and  $J_3$  is a 3x3 unit matrix (a matrix of ones). The association constant,  $K_A$ , can be computed for a harmonic well potential as,

$$K_A = \frac{1}{8\pi^2} \int H(r) e^{-U_H(r)/kT} dr \quad (\text{S3})$$

where  $H(r) = 1$  in the bound state and 0 otherwise. The integral is solved analytically as,

$$\int e^{-U_H(r)/kT} dr = e^{-U_M/kT} \sqrt{(2\pi)^n |\sigma_{ij}^2|} \quad (\text{S4})$$

where  $|\sigma_{ij}| = (kT/k_s)^3$  is the determinant of  $\sigma_{ij}$  and  $n$  is the degrees of freedom in the system. With biomolecules allowed to diffuse in 3 dimensions and non-orientation dependent binding, substituting Eq. (S4) into Eq. (S3) yields,

$$K_A^{-1} = K_D = (2\pi kT/k_s)^{(-3/2)} \exp(|U_M|/kT) \quad (\text{S5})$$

allowing for direct computation of  $K_D$  as a function of  $k_s$  and  $U_M$ .

### Calculating $K_D$ for Harmonic Well + Hard Sphere Potentials

As described in the Theory section, the second virial coefficient,  $B_2$ , is calculated for ligands with hard core + harmonic well (HCHW) potentials (*i.e.*,  $a_L > 0$ ) to estimate their effective interaction strength. The second virial coefficient for ligands with hard cores,  $B_{2,HCHW}$ , is calculated with Eq. (4) as,

$$B_{2,HCHW} = (16/3)\pi a_L^3 + 12 \int_{a_L}^{a_L+\delta} [1 - \exp[-U_{RL}(r, a_L)/kT]] r^2 dr \quad (\text{S6})$$

An equivalent harmonic well potential is then found by determining a simple harmonic well (HW) potential (*i.e.*,  $a_L = 0$ ) with the same  $B_2$  as the ligand including a hard core. The second virial

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\* To whom correspondence should be addressed. email: mabevan@jhu.edu

coefficient of a HW potential (without a hard core),  $B_{2,HW}$ , is calculated with Eq. (4) as,

$$B_{2,HW} = 12 \int_0^{\delta} [1 - \exp[-U_{RL}(r, 0)/kT]] r^2 dr \quad (S7)$$

To find an equivalent HW potential,  $U_M$  in  $U_{RL}$  (Eq. 1) is adjusted at constant  $k_s$  until  $B_{2,HW}$  is equal to  $B_{2,HCHW}$ . These two potentials are then considered equivalent and the  $K_D$  of the harmonic well calculated from Eq. (3) assigned to the ligand including a hard core.

### 3D Rotational Receptor Moves

In order to maintain uniform spacing between the receptors and the surface while making 3D receptor rotational moves, the rotations made must be uniform in Euclidean space. The following algorithm was developed by Arvo to make random, uniform rotational moves<sup>2</sup>:

1. Rotate randomly about the z axis by applying the rotation matrix,  $R$ :

$$R = \begin{vmatrix} \cos(2\pi\delta_1) & \sin(2\pi\delta_1) & 0 \\ -\sin(2\pi\delta_1) & \cos(2\pi\delta_1) & 0 \\ 0 & 0 & 1 \end{vmatrix} \quad (S8)$$

where  $\delta_1$  is a random variable that ranges from 0 to the maximum rotational step size,  $s_{rot}$ .

2. Rotate randomly from the position  $(0, 0, a + h_R)$  to a random position by using the Householder matrix,  $H$ :

$$H = I - 2vv^T \quad (S9)$$

where  $v$  is the matrix,

$$v = \begin{vmatrix} \cos(2\pi\delta_2)\sqrt{\delta_3} \\ \sin(2\pi\delta_2)\sqrt{\delta_3} \\ \sqrt{1-\delta_3} \end{vmatrix} \quad (S10)$$

3. The final rotation can be defined with matrix,  $M$  as:

$$M = -HR \quad (S11)$$

The random variables  $\delta_2$  and  $\delta_3$  also range from 0 to  $s_R$ . A maximum value of  $1 \times 10^{-3}$  for  $s_R$  was found empirically to not disturb the equilibrium ligand coverage and allowed for unbiased configurational sampling as shown in Figure S1.

### Colloid-Ligand Cluster Moves

In order to allow our colloids to freely diffuse with dense coverages of ligands on their surface, cluster moves were used to translate all interacting colloids and ligands when attempting colloidal MC moves. The following algorithm was used to decide whether to accept or reject the cluster moves:

1. Determine number of ligands,  $n_L$ , bound to a receptor on the colloid ( $r < a_L + \delta$ )

2. Move  $n_L$  ligands on colloid with same translation and rotation as attempted by the colloid and receptors in that step.
3. Calculate total change in energy due to the cluster move,  $\Delta E$ , for colloid and the  $n_L$  ligands bound.

$$\Delta E = \Delta E_C + \sum_{i=1}^{n_L} E_{L,i} \quad (\text{S12})$$

where  $\Delta E_C$  is the change in energy of the colloid and  $\Delta E_L$  is the change an energy of a ligand.

4. If  $\Delta E$  is less than or equal to zero or  $e^{(-\Delta E)}$  is greater than a random integer, the cluster move is accepted.

By applying the metropolis criterion to these cluster moves, the potential for biasing in these moves was avoided. Figure S2 shows on a single diffusing colloid with coverages up to ~99%, MC move efficiency of the colloid (open triangles down) and ligands was still maintained with a minimum of ~78% acceptance rates (open circles).

#### *Calculating Number, Energy, and Orientation of Ligand Bridges*

The interaction of ligands with receptors is strictly limited to one receptor on each particle. If a ligand comes within range of a receptor ( $r < a_L + \delta$ ), the net interaction is zero if it is currently occupying a receptor on the same particle. All ligands interacting with two receptors (one on each particle) are identified as a bridge. The number of bridges,  $N_B$ , was tracked in each step as a function of  $L$  in all MC-US simulations. The average number of bridges as a function of  $L$ ,  $\langle N_B \rangle(L)$ , can be calculated as,

$$\langle N_B \rangle(L) = \sum^{n(L)} N_B(L) / n(L) \quad (\text{S13})$$

where  $n(L)$  is the total number of bridging events at separation  $L$ . The energy of each ligand bridge is calculated as,

$$U_B = U_{RL,i}(r_i) + U_{RL,j}(r_j) \quad (\text{S14})$$

which is simply the sum of the interactions with the receptors that make up the bridge, receptor  $i$  and receptor  $j$ , that can be calculated with Eq. (3). Since colloids only make translational moves in the  $x$ -direction, we can determine the bridge orientation with respect to the colloidal surfaces as the orientation of the bridge in the  $x$ -direction. The orientation of each bridge is calculated with respect to the colloids in the  $x$ -direction by first determining the root mean-square distance between the receptors  $i$  and  $j$ ,  $r_{ij}$ , calculated as,

$$r_{ij} = \sqrt{(x_i - x_j)^2 + (y_i - y_j)^2 + (z_i - z_j)^2} \quad (\text{S15})$$

and the root mean-square distance between receptors  $i$  and  $j$  in the  $x$ -direction,  $r_{ij,x}$ , calculated as,

$$r_{ij,x} = \sqrt{(x_i - x_j)^2} \quad (\text{S16})$$

We can then calculate the bridge orientation,  $\phi$ , between receptors  $i$  and  $j$  with respect to the colloidal surfaces as,

$$\phi = 2 \arcsin\left(r_{ij,x}/r_{ij}\right) \quad (\text{S17})$$

Histograms of  $U_B$  and  $\phi$  were then populated as a function of  $L$  from each of the 12 bins of the MC-US simulation results.

### Supplementary Figure Captions

**Figure S1.** Schematic of receptor position (green circle) in spherical coordinates where  $\alpha$  and  $\psi$  are the polar and azimuthal angles of each receptor with respect to the center of the colloid (blue circle). Histograms of sampled (A)  $\alpha$  and (B)  $\psi$  in 2000 representative configurations for MC colloidal surface adsorption simulation at  $[C] = 10 \mu\text{M}$ ,  $a_L = 5\text{nm}$ , and  $U_M = 11kT$  ( $\theta \approx 0.99$ ).

**Figure S2.** MC move efficiency (open circles) and fraction coverage,  $\theta$ , (open triangles down) for MC colloidal surface adsorption simulation at  $[C] = 10 \mu\text{M}$ ,  $a_L = 5\text{nm}$ , and  $U_M = 11kT$ .

### Supplementary References

1. Luo, H.; Sharp, K. On the calculation of absolute macromolecular binding free energies. *Proceedings of the National Academy of Sciences of the United States of America* **2002**, *99*, 10399-404.
2. Arvo, J. Fast Random Rotation Matrices. *Graphics Gems III* **1992**, 117-120.

Figure S1

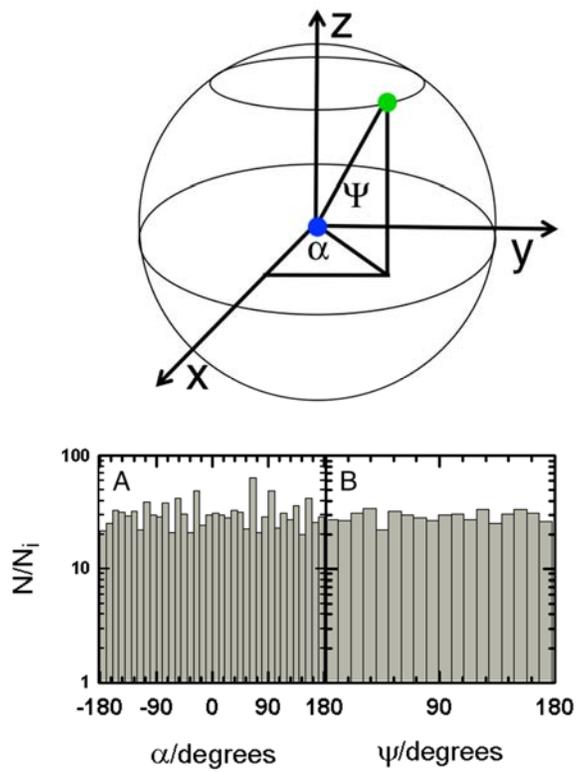


Figure S2

