# Anionic Deep Cavitands Control the Adhesion of Unmodified Proteins at a Membrane Bilayer

Yoo-Jin Ghang,<sup>†</sup> Lizeth Perez,<sup>†</sup> Melissa A. Morgan,<sup>†</sup> Fang Si,<sup>‡</sup> Omar M. Hamdy,<sup>†</sup> Consuelo N. Beecher,<sup>†</sup> Cynthia K. Larive,<sup>†</sup> Ryan R. Julian,<sup>†</sup> Wenwan Zhong,<sup>†</sup> Quan Cheng<sup>†</sup> and Richard J. Hooley<sup>†</sup>\*

<sup>†</sup>Department of Chemistry, University of California, Riverside, CA 92521, United States <sup>‡</sup>Donghua University, College of Chemistry, Chemical and Biological Engineering, Shanghai, 201620, China

richard.hooley@ucr.edu

## **Electronic Supplementary Information**

## 1. SPR Binding Analysis (sensorgrams not shown in the text)

1) Bovine Serum Albumin (BSA)



Figure S-1. SPR sensorgrams of the binding event between POPC:cavitand 1 and BSA in 20 mM PBS (left) or 100 mM PBS (right).

## b. POPC:cavitand 2 bilayer



Figure S-2. SPR sensorgrams of the binding event between POPC:2% cavitand 2 and BSA in water (left) or 100mM PBS (right).

c. POPC:sodium palmitate bilayer



Figure S-3. SPR sensorgram of the binding event between POPC:sodium palmitate and BSA in water.

## 2) Cytochrome c (cyt c)

a. POPC:cavitand 1 bilayer



Figure S-4. SPR sensorgram of the binding event between POPC:cavitand 1 and cyt c in 100 mM PBS.

## b. POPC:cavitand 2 bilayer



Figure S-5. SPR sensorgram of the binding event between POPC:2% cavitand 2 and cyt c in 100 mM PBS.

#### 3) Myoglobin

## a. POPC:cavitand 1 bilayer



**Figure S-6.** SPR sensorgrams of the binding event between POPC:cavitand 1 and myoglobin in water (left) or 100 mM PBS (right).

#### b. Control experiment (POPC bilayer alone)



Figure S-7. SPR sensorgram of the binding event between POPC membrane and myoglobin in water.

## c. POPC:cavitand 2 bilayer



**Figure S-8.** SPR sensorgrams of the binding event between POPC:2% cavitand **2** and myoglobin in water (left) or 100 mM PBS (right).

#### 4) Trypsin

a. POPC:cavitand 1 bilayer



**Figure S-9.** SPR sensorgrams of the binding event between POPC:cavitand 1 and trypsin in water (left) or 100 mM PBS (right).

#### b. Control experiment (POPC bilayer alone)



Figure S-10. SPR sensorgram of the binding event between POPC membrane and trypsin in water.

## 5) TPCK-trypsin

a. POPC:cavitand 1 bilayer



**Figure S-11.** SPR sensorgrams of the binding event between POPC:cavitand 1 and TPCK-trypsin in water (left) or 100 mM PBS (right).

b. Control experiment (POPC bilayer alone)



Figure S-12. SPR sensorgram of the binding event between POPC membrane and trypsin in water.



# 2. Capillary Electrophoresis Binding Analysis

**Figure S-13.** Electropherograms for cyt *c* incubated with cavitand **1** as running buffer at 191 nm,  $[1] = 3-30 \mu$ M, [Cyt *c*] = 3  $\mu$ M).



Sample ID (Cavitand: cyt C molar ratio)	Mobility 1	Mobility 2	Mobility 3	Average Mobility	Stdev Mobility
Cyt C control	-4.64E-08	-4.59E-08	-4.62E-08	-4.62E-08	2.43E-10
3uM:3uM	-4.26E-08	-4.20E-08	-4.16E-08	-4.21E-08	4.60E-10
7.5uM:3uM	-4.03E-08	-4.00E-08	-4.07E-08	-4.03E-08	3.42E-10
15uM:3uM	-3.92E-08	-3.98E-08	-3.90E-08	-3.94E-08	3.98E-10
22.5uM:3uM	-3.98E-08	-3.99E-08	-3.91E-08	-3.96E-08	4.36E-10
30uM:3uM	-3.92E-08	-3.98E-08	-3.99E-08	-3.97E-08	3.64E-10

**Figure** *S***-14.** Mobility shift of cyt *c* vs. [1] and binding constant calculation:  $K_d = 2.5 \times 10^{-6} M$ .



**Figure S-15.** Electropherograms for cyt *c* incubated with cavitand **1** and POPC lipids as running buffer at 191 nm,  $[1] = 3-30 \ \mu\text{M}$ ,  $[Cyt c] = 3 \ \mu\text{M}$ ,  $[POPC] = 15.8 \ \mu\text{M}$ ).



**Figure S-16.** Mobility shift of cyt *c* vs. [1] in the presence of POPC lipid vesicles and binding constant calculation:  $K_d = 7.59 \times 10^{-6} M$ .

## 3. Trypsin digestion on bioreactive surface



#### a) Unreacted Oxidized Insulin chain B

Figure S-17. ESI-MS analysis of oxidized insulin chain B.

## b) Trypsin digestion of Insulin B in solution



**Figure S-18.** ESI-MS analysis of trypsin (7.5  $\mu$ M) digestion of insulin chain B (150  $\mu$ M) for 1 h at 298 K in aqueous solution.



## c) Trypsin digestion of Insulin B at the Bioreactive surface

**Figure S-19.** SPR sensorgram of trypsin digestion of insulin chain B for 1 h at 298 K at the POPC:1:trypsin surface.



Figure S-20. SPR sensorgram of trypsin digestion of insulin chain B for 10 min at 298 K in SPR.



**Figure S-21.** HPLC and ESI-MS analysis of trypsin digestion of insulin chain B for 1 h at 298 K in SPR. a) HPLC-ESI mass spectra of collected fractions from HPLC: b) fraction 1; c) fraction 2; and d) fraction 3.



Figure S-22. ESI-MS analysis of trypsin digestion of insulin chain B for 10 min in SPR.



## d) Surface Reusability: multiple trypsin digestions at the same surface

Figure S-23. SPR sensorgram of multiple trypsin digestions of insulin chain B for 1 h at 298 K in SPR.



**Figure S-24.** ESI-MS analysis of multiple trypsin digestions of insulin chain B for 1 h at 298 K in SPR: a) first digestion, b) second digestion.

#### e) Inhibition of trypsin digestion by addition of benzamidine hydrochloride



Figure S-25. SPR sensorgram of trypsin digestion of insulin chain B with 100 mM benzamidine hydrochloride for 1 h at 298 K in SPR.



Figure S-26. ESI-MS analysis of trypsin digestion of insulin chain B with 100 mM benzamidine hydrochloride for 1 h at 298 K in SPR.

#### f) TPCK-trypsin digestion



**Figure S-27.** ESI-MS analysis of TPCK-trypsin (7.5  $\mu$ M) digestion of insulin chain B (150  $\mu$ M) for 1 h at 298 K in aqueous solution.



Figure S-28. SPR sensorgram of TPCK-trypsin digestion of insulin chain B for 1 h at 298 K in SPR.



Figure S-29. HPLC analysis of TPCK-trypsin digestion of insulin chain B for 1 h at 298 K in SPR.



Figure S-30. ESI-MS analysis of TPCK-trypsin digestion of insulin chain B for 1 h at 298 K in SPR.

# 4. CD Analysis of injected protein structure



Figure S-31: CD spectrum of 2 µM BSA in water (solid line) or 100 mM PBS (dotted line).



Figure S-32: CD spectrum of 2 µM cyt c in water (solid line) or 100 mM PBS (dotted line).



Figure S-33: CD spectrum of 2 µM myoglobin in water (solid line) or 100 mM PBS (dotted line).