

Electronic supplementary information (ESI)

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**Facile Fabrication of Stable Monolayer and Few-layer Graphene
Nanosheets as Superior Sorbents for Persistent Aromatic Pollutant
Management in Water**

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Supporting Information consists of 6 pages, including this one.

There are four Figures and two Tables.

Contents

1. Synthesis of Graphene Oxide (GO) and Graphene	S2
2. Synthesis of Aminosilica Particles (NH ₂ -SiO ₂)	S3
3. Fabrication of Aminosilica-Support-Graphene (ASG)	S3
4. Structural Characterization	S3
5. Batch Adsorption Experiment	S4
Table S-1. Elemental analysis results of SiO ₂ , NH ₂ -SiO ₂ and ASG.	S6
Table S-2. BET-N ₂ surface area of SiO ₂ , pristine graphene and ASG.	S6
Table S-3. Regression parameters of adsorption isotherms of phenanthrene onto SiO ₂ , NH ₂ -SiO ₂ , ASG and pristine graphene.	S6
Figure S-1. Selected FT-IR spectra curves of SiO ₂ and NH ₂ -SiO ₂ nanoparticles (A), and whole FT-IR spectra of SiO ₂ , GO, ASGO, ASG nanoparticles (B).	S7
Figure S-2. Zeta potential curves of NH ₂ -SiO ₂ , GO and ASG at different pH values.	S8
Figure S-3. The Raman spectra of original graphene and ASG.	S9
Figure S-4. The adsorption kinetic of phenanthrene onto ASG	S10

16 Synthesis of Graphene Oxide (GO) and Graphene

17 GO was synthesized from natural graphite flakes (325 mesh, 99.8%, Alfa Aesar)
18 according to a modified Hummers method. $K_2S_2O_8$ (8.33 g) and P_2O_5 (8.33 g) were dispersed
19 in concentrated H_2SO_4 (30 mL), then graphite flake powder (10 g) was added gradually upon
20 continuous stirring, and the mixture was kept at 80 °C for 4.5 h under stirring. After that, the
21 product was collected through filtration and washing with water until the pH became neutral;
22 the product was then dried in the oven overnight at 60 °C to obtain the preoxidized graphite.
23 The preoxidized graphite powder (10 g) and $NaNO_3$ (5 g) were placed into cold (0 °C)
24 concentrated H_2SO_4 (230 mL) in an ice bath, and $KMnO_4$ (30 g) was slowly added under
25 continuous stirring to keep the temperature below 4 °C. Then, the mixture was stirred at 35 °C
26 for 2 h; deionized water (460 mL) was then gradually added, producing a large amount of
27 heat. The mixture was further stirred for 15 min at 98 °C to increase the oxidation level. The
28 reaction was terminated by adding deionized water (460 mL) and 30% H_2O_2 (25 mL) to
29 obtain graphite oxide. The resultant bright yellow mixture was rinsed with a 10% HCl
30 solution (3.6 L) to remove the residual SO_4^{2-} (verified using 0.01 mol/L $BaCl_2$) and metal ions,
31 followed by centrifugation at 8000 rpm. Then, the solid phase was redispersed in deionized
32 water and peeled by ultrasonication for 30 min at a power of 250 W. The centrifugation and
33 ultrasonication were repeated 3 times, and the solution was then subjected to dialysis to
34 remove the acid and other impurities. After being vacuum dried at 60 °C, GO was obtained as
35 a powder.

36 Graphene was prepared by the hydrazine reduction of the GO nanosheets. Briefly, a 1L
37 GO aqueous dispersion (1 mg/mL) was ultrasonicated (250 W) until it became clear and
38 contained no visible particulate matter. The dispersion was then centrifuged at 3000 rpm for 5
39 min to remove any unpeeled sheets. The supernatant was loaded in a 1-L round-bottom flask,
40 followed by adjusting the solution pH to 10 using ammonium hydroxide (28%). Then, 10 mL
41 of hydrazine hydrate (98%, Aladdin Co. Ltd.) was added, and the solution was heated in an
42 water bath at 98 °C under a water-cooled condenser for 24 h. The resultant black solid was
43 isolated by filtration through a 0.22 μm membrane filter, washed by deionized water and

methanol to remove the excess hydrazine, and finally freeze dried. The black powder was graphene.

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Synthesis of Aminosilica Particles ($\text{NH}_2\text{-SiO}_2$)

Nanosilica particles (SiO_2 , 50 ± 5 nm) were purchased from Aladdin Industrial Corporation. $\text{NH}_2\text{-SiO}_2$ was synthesized from SiO_2 by binding $-\text{NH}_2$ to the surface through a simple one-step process. SiO_2 (10 g) was added gradually to ethyl alcohol (300 mL) under continuous stirring and ultrasonication. Then, APTES (1 mL) was added to the mixture. The reaction was processed for 12 h at 50°C , and the product was collected by filtration, washed with alcohol and water several times and dried under vacuum at 60°C .

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Fabrication of Aminosilica-Support-Graphene (ASG)

The fabrication of ASG included two brief steps. First, 100 mL of 0.1 mg/mL GO solution was completely peeled off under ultrasonication. $\text{NH}_2\text{-SiO}_2$ (2 g) was dispersed in deionized water (100 mL) under stirring and ultrasonication to ensure good dispersion. Then, the GO solution was gradually added under stirring to the $\text{NH}_2\text{-SiO}_2$ mixture. Due to the electrostatic interactions between them, the GO layers immediately attached to the surface of the $\text{NH}_2\text{-SiO}_2$. The wrapped nanoscale particles were obtained by filtration and dried under vacuum. Second, ASGO (800 mg) was added to water (800 mL) with the addition of hydrazine hydrate (0.04 mL). The reaction was processed in a water bath at 98°C . The final ASG particles were collected by filtration and dried in a vacuum at 60°C .

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Structural Characterization

Elemental (C, H, N) analyses were conducted using an EA112 CHN elemental analyzer (Thermo Finnigan). The Brunauer-Emmett-Teller (BET) nitrogen specific surface areas were estimated by nitrogen adsorption-desorption at -196°C with a NOVA-2000E surface area analyzer. Four data points, with relative pressures of 0.05 to 0.3, were used to determine the monolayer adsorption capacity. Raman spectra were obtained using a LabRamHRUV Raman

spectrometer (Jobin-Yvon, FR); the laser excitation was provided by an Ar⁺ laser at a wavelength of 514 nm. The surface functional groups were examined using Fourier transform infrared spectroscopy (FTIR). The FTIR spectra were recorded in the 4000-400 cm⁻¹ region with a resolution of 4 cm⁻¹ using a Bruker Vector 22 FTIR spectrometer. The surface morphologies were characterized using an S-4800 field-emission scanning electron microscope (FE-SEM) (Hitachi, Tokyo) and a JEM-2010 transmission electron microscope (TEM) (JEOL, Japan). The surface charge properties of the biochars were evaluated using zeta potential measurements, which were conducted at a different equilibrium pH values using a Nano-ZS90 Zetasizer (Malvern Instruments Ltd.). The samples were pre-equilibrated for 24 h in different pH solutions. The zeta potential was measured three times at each pH, and the average was calculated. The isoelectric point (pH_{IEP}), defined as the pH value at which the electrokinetic potential equals zero, of the biochars was determined by plotting the zeta potentials vs. pH.

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86 **Batch Adsorption Experiment**

Phenanthrene (98%+, J&K Chemical) was selected as a model aromatic organic pollutant. Adsorption kinetics and batch adsorption experiments were performed in PTFE screw-cap vials sealed with tin foil at 25±1 °C. Concerning the adsorption kinetics of ASG, the water-to-solid ratio was 1.5 mg per 8 mL, and the initial phenanthrene concentration was 5.6 µmol/L. The reaction time ranged from 10 min to 1440 min. The adsorption capacities of SiO₂, NH₂-SiO₂ and the original graphene obtained at various times (36 h and 72 h) were the same, suggesting that equilibration was achieved before 36 h.

Batch adsorption experiments with SiO₂, NH₂-SiO₂, pristine graphene and ASG were conducted in background solution at a water-to-solid ratio of 200 mg per 8 mL, 200 mg per 8 mL, 1 mg per 240 mL, and 1 mg per 8 mL, respectively. The background solution contained 0.01 mL/L CaCl₂ and 200 mg/L NaN₃ to control the ionic strength and eliminate the effect of microorganisms. The phenanthrene in acetonitrile was dissolved in the background solution (pH=7) to prepare the working solution; the volume of acetonitrile in the background solution

was maintained at less than 0.1% to avoid cosolvent effects. The initial concentrations of the phenanthrene solution were controlled to obtain equilibrium concentration ranges over 3 orders of magnitude including 0.0056, 0.017, 0.034, 0.056, 0.17, 0.34, 0.56, 1.7, 3.4, 5.6 $\mu\text{mol/L}$. Each point, either the blank or calibration control, was evaluated in duplicate. The vials were placed on a shaker and agitated in the dark at 120 rpm for 36 h to reach apparent equilibrium. The solution was separated from the solid. Then, 500 μL of the supernatants was mixed with 500 μL of acetonitrile to determine the PAH concentration on an Agilent 1200 HPLC (Agilent Eclipse XDB-C 18 column, 4.6 mm \times 250 mm \times 5 μm) equipped with a G1321A fluorescence detector. The mobile phases were acetonitrile:water (90:10, v:v) and a flow rate of 1 $\text{mL}\cdot\text{min}^{-1}$ was used. The determination of phenanthrene was performed using an excitation wavelength of 244 nm and an emission wavelength of 360 nm. The results revealed no noticeable mass loss, and the amount of PAH adsorbed by the materials was calculated from the difference between the initial and equilibrium concentrations.

Table S-1. Elemental analysis results of SiO₂, NH₂-SiO₂ and ASG.

Sample	Relative atomic percentage (%)		
	Nitrogen	Carbon	Hydrogen
SiO ₂	0	0.0650	0.6916
NH ₂ -SiO ₂	0.4185	1.1110	0.7886
ASG	0.3189	1.6986	0.7011

Table S-2. BET-N₂ surface area of SiO₂, pristine graphene and ASG.

Sample	Surface area (m ² /g)
SiO ₂	120.041
NH ₂ -SiO ₂	108.5
pristine graphene	715.8
ASG	118.7

Table S-3. Regression parameters of adsorption isotherms of phenanthrene onto SiO₂, NH₂-SiO₂, ASG and pristine graphene.

Freundlich			
Adsorbent	K_f	N	R^2
SiO ₂	0.0060 ± 0.00034	1.0598 ± 0.0994	0.944
NH ₂ -SiO ₂	0.0048 ± 0.00062	0.8708 ± 0.1634	0.866
pristine graphene	149.26 ± 20.24	0.9450 ± 0.0721	0.967
ASG	1.7371 ± 0.05054	0.3239 ± 0.0186	0.976

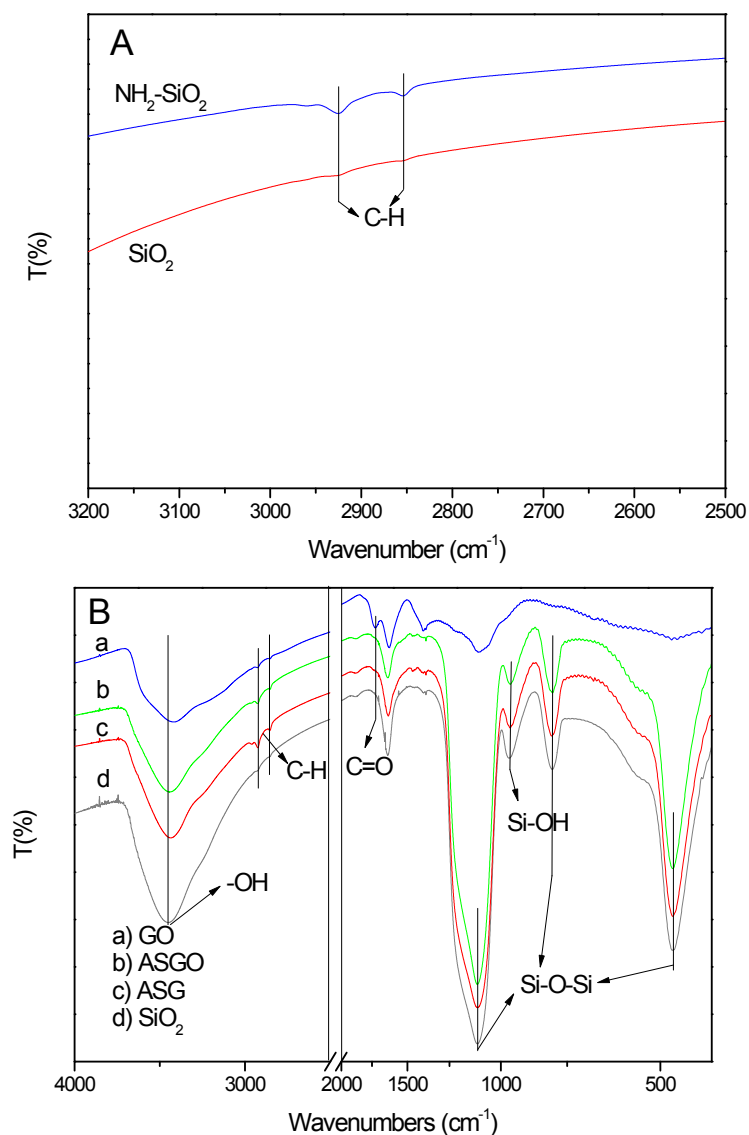


Figure S-1. Selected FT-IR spectra curves of SiO_2 and $\text{NH}_2\text{-SiO}_2$ nanoparticles (A), and whole FT-IR spectra of SiO_2 , GO, ASGO, ASG nanoparticles (B).

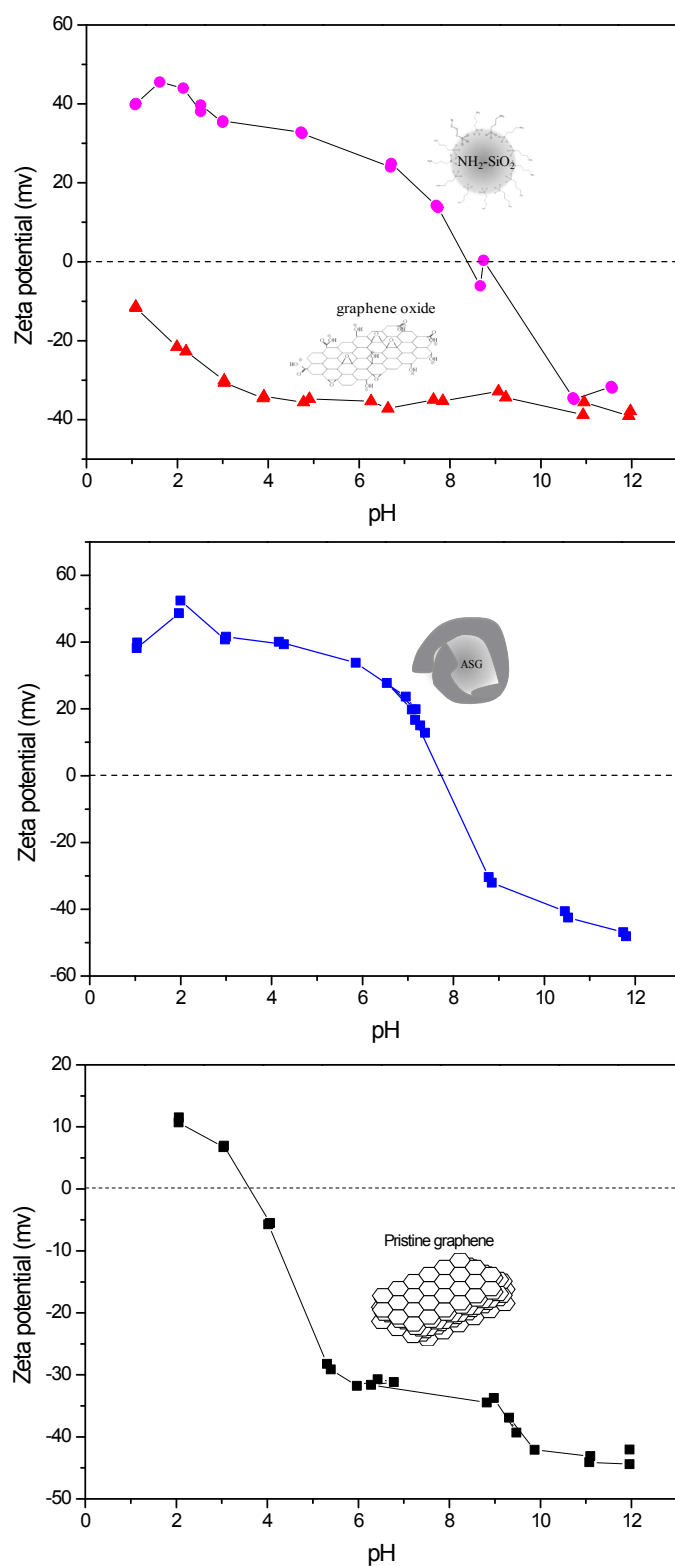


Figure S-2. Zeta potential curves of NH₂-SiO₂, GO and ASG at different pH values.

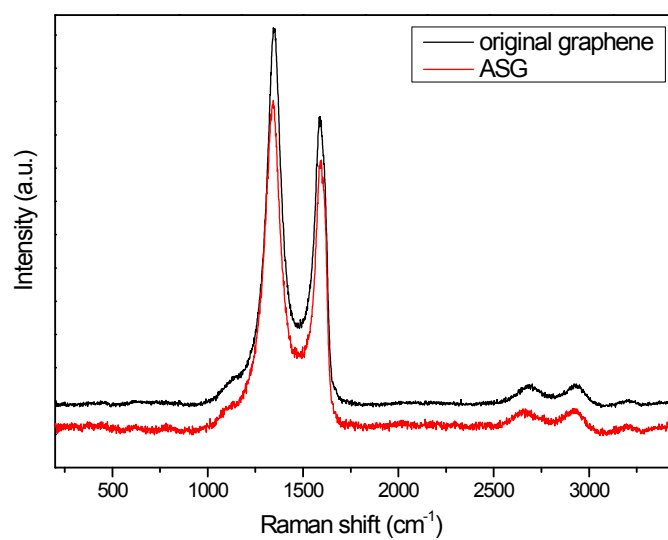
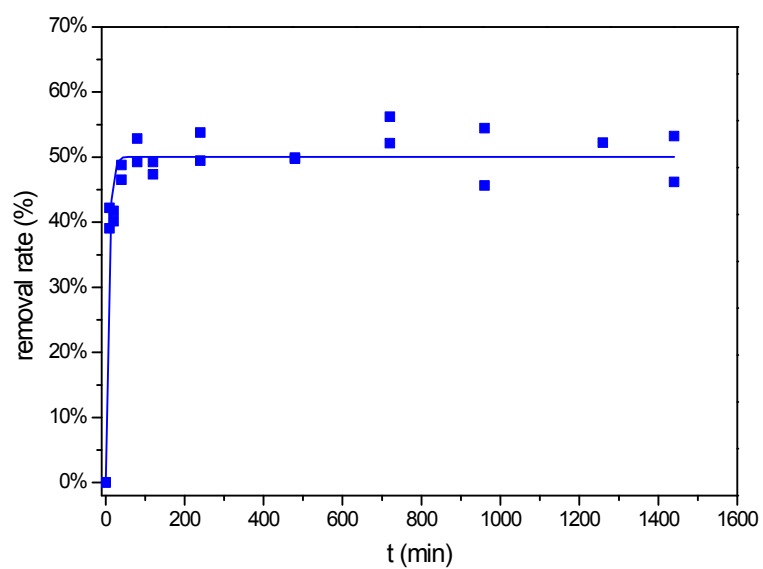


Figure S-3. The Raman spectra of original graphene and ASG.



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Figure S-4. The adsorption kinetic of phenanthrene onto ASG.