Electronic Supplementary Material (ESI) for Journal of Materials Chemistry B. This journal is © The Royal Society of Chemistry 2014

DNA-templated silver nanoclusters as label-free, sensitive detection probes for potassium ion and nitric oxide

Jihyun Lee, ^{‡,a,b} Junghong Park, ^{‡,a,b}, Honghee Lee^b, Hugh I. Kim^b and Won Jong Kim^{*,a,b}

^aCenter for Self-assembly and Complexity, Institute for Basic Science, Pohang 790-784,

Korea

^bDepartment of Chemistry, Polymer Research Institute, Pohang University of Science and Technology (POSTECH), Pohang 790-784, Korea

*Corresponding author. Tel.: +82 54 279 2104; Fax: : +82 54 279 3399.

E-mail address: wjkim@postech.ac.kr (W. J. Kim)

[‡] These authors contributed equally to this work.

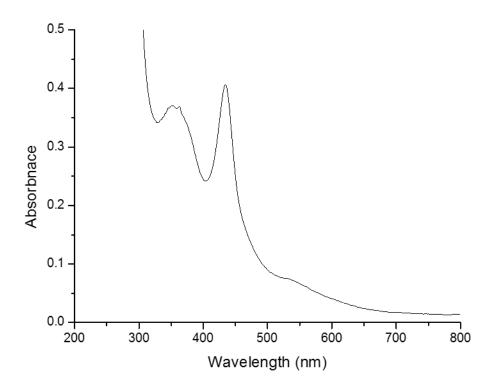


Fig S1. Absorption spectrum of DNA1-AgNCs (10 μ M DNA, 120 μ M Ag⁺, and 120 μ M BH₄⁻) acquired 3h after adding BH₄⁻.

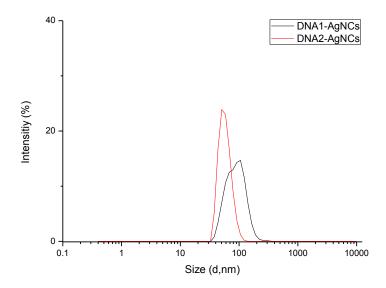


Fig S2. DLS measurement of DNA1-AgNCs, DNA2-AgNCs (5 $\mu M)$ by Malvern Nano-S90 spectrometer.

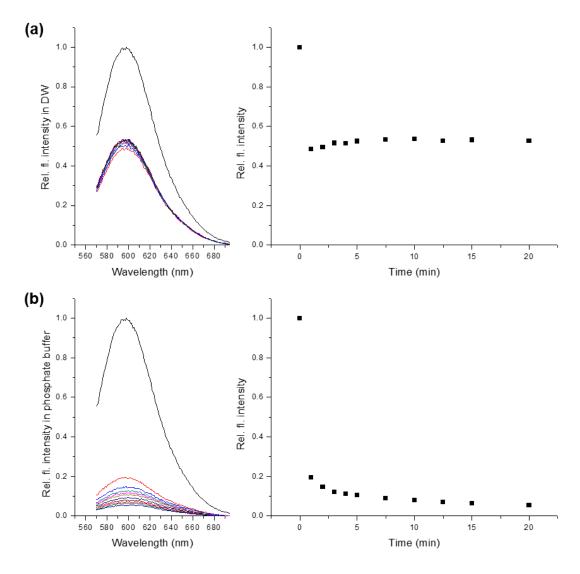


Fig S3. Fluorescence emission spectra of DNA1-AgNCs affected by different dilution solvents as (a) DW and (b) phosphate buffer (Na $^+$ 20 mM, Mg $^{2+}$ 1 mM, pH 7.0). In left, fluorescence emission spectra of DNA1-AgNCs diluted with either DW or phosphate buffer (1 μ M in DNA, coloured lines) depending on time (from 1 min to 20 min) are plotted when maximum emission intensity of initial DNA1-AgNCs (2 μ M in DNA, black line) set as 1. Plots of the relative fluorescence intensity (I/I $_0$) at 595 nm excited at 540 nm during 20 min for different dilution solvent show in right.

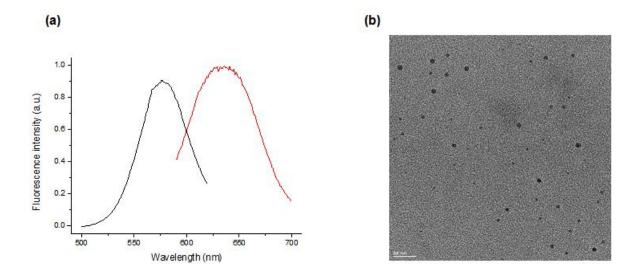
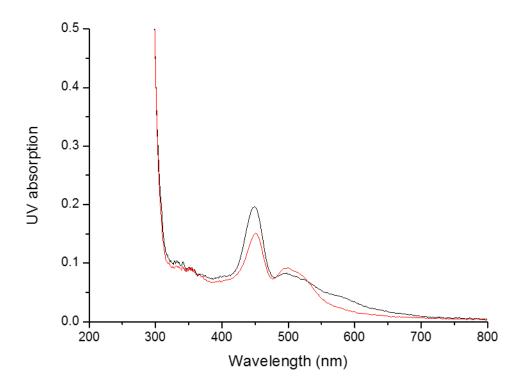


Fig S4. (a) Maximum fluorescence of DNA2-AgNCs with ex/em=570/615 nm (b) TEM image of DNA2-AgNCs

.



g S5. Absorption spectrum of DNA2-AgNCs (black, 15 μ M DNA, 90 μ M Ag⁺, and 90 μ M BH₄⁻), and DNA2-AgNCs+NO (red, 15 μ M DNA, 90 μ M Ag⁺, and 90 μ M BH₄⁻, NO 5.64 μ M). The DNA2-AgNCs was prepared 3 h after adding BH₄⁻.

Fi

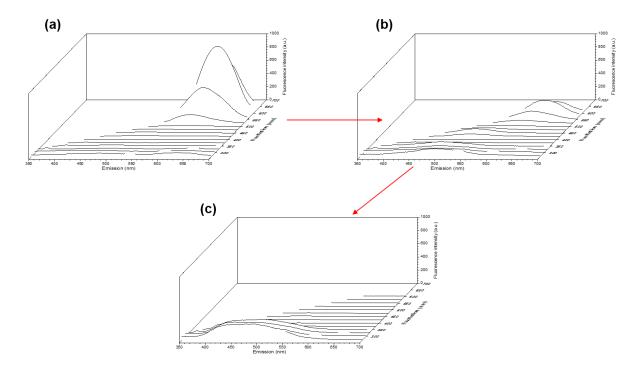


Fig S6. Fluorescence emission spectra as a function of excitation wavelengths. The prepared DNA-AgNCs was obtained using DNA2 (1 μ M) with Ag⁺ (6 μ M) and BH₄⁻ (6 μ M). The emission wavelengths are on the bottom axis, and the excitation wavelengths are incremented by 50 nm starting at 270 nm on the right axis. All the data was obtained 3h after adding BH₄⁻. (a) DNA2-AgNCs (b) DNA2-AgNCs+NO (5.64 μ M which concentration in dynamic range) and (c) DNA2-AgNCs+NO (1.88 mM in excess) at 23°C

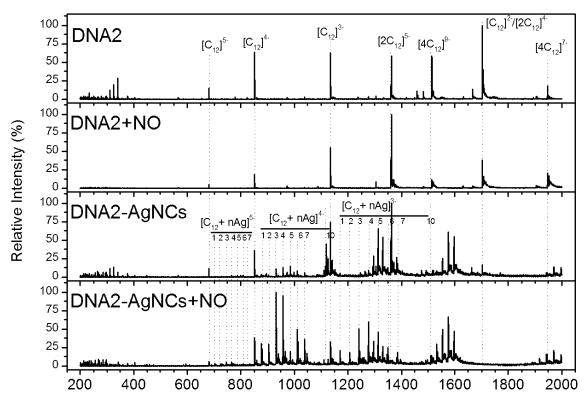


Fig S7. ESI-MS spectrum of DNA2, DNA2+NO, DNA2-AgNCs, and DNA2-AgNCs+NO.

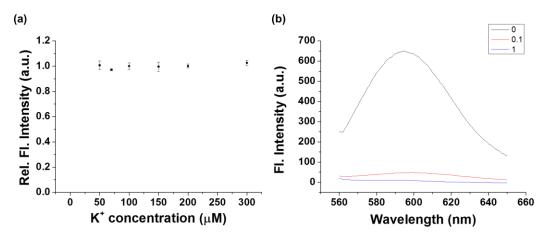


Fig S8. Change of DNA sequence for detection each K^+ by DNA2-AgNCs and NO by DNA1-AgNCs. Relative fluorescence intensity of (a) DNA2-AgNCs(5 μM) after addition of K^+ (0, 50, 70, 100, 150, 200, 300 μM), and (b) DNA1-AgNCs(5 mM) after addition of nitric oxide (0, 0.1, 1 mM). (a) The result shows the fluorescence intensity is not interfered by existence of K^+ . Because in the case of DNA1, K^+ is used for inducing a DNA1 to form G-quadruplex. (b) The result shows the fluorescence intensity is immediately quenched even low concentration of NO. Because in the case of DNA2, it has 12 nts of cytosine which can be reacted with NO to uracil by deamination to make AgNCs unstable for inducing structural change.