

Supporting Information

Injectable and Biodegradable Supramolecular Hydrogels formed by Nucleobases-terminated Poly(ethylene oxide)s and α -Cyclodextrin

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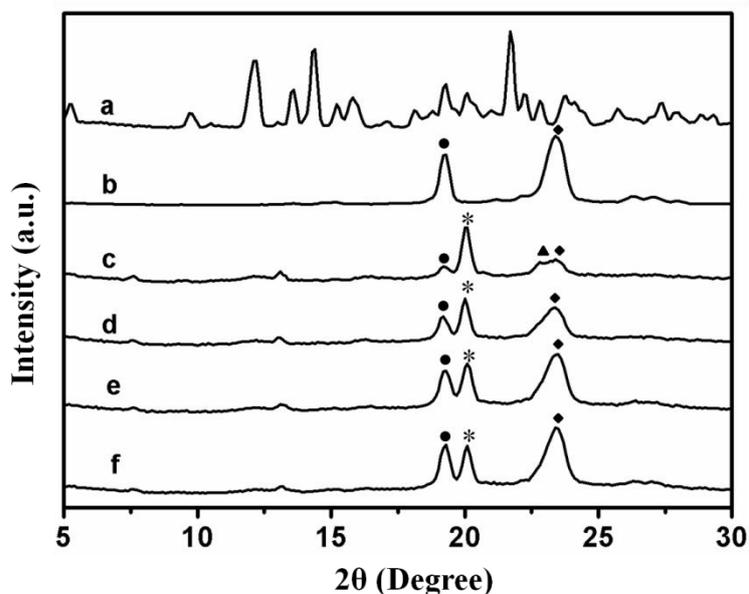


Figure S1. Powder X-ray diffraction of (a) α -CD, (b) PEG20k, (c) G5, (d) G6, (e) G7 and (f) G8. The characteristic X-ray diffraction peaks of crystalline columnar α -CD at around $2\theta = 19.9$ and 22.6° as well as PEG at around $2\theta = 19.3$ and 23.5° are labeled with *, \blacktriangle , \bullet and \blacklozenge , respectively.

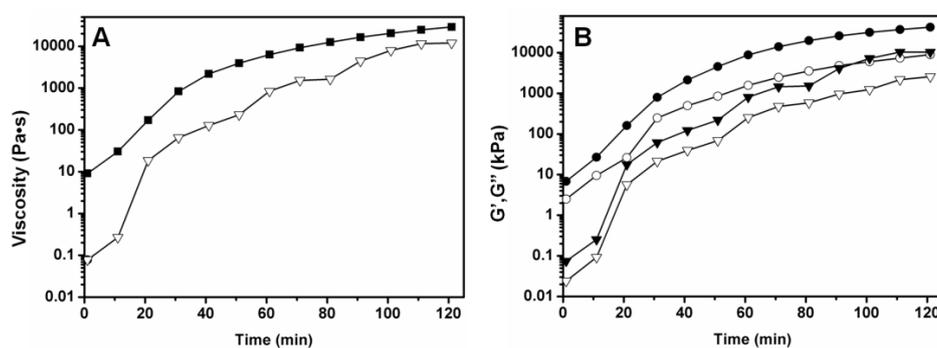


Figure S2. (A) Changes of viscosities of hydrogels as a function of time. (\blacksquare , viscosity of G4 hydrogel; \square , viscosity of PEG10k/ α -CD (20 wt %/10 wt %) hydrogel); (B) Time sweep measurements for the viscoelastic properties of hydrogels as a function of time. (\bullet , G' and \circ , G'' of G4 hydrogel; \blacktriangledown , G' and \triangledown , G'' of PEG10k/ α -CD (20 wt %/10 wt %) hydrogel).

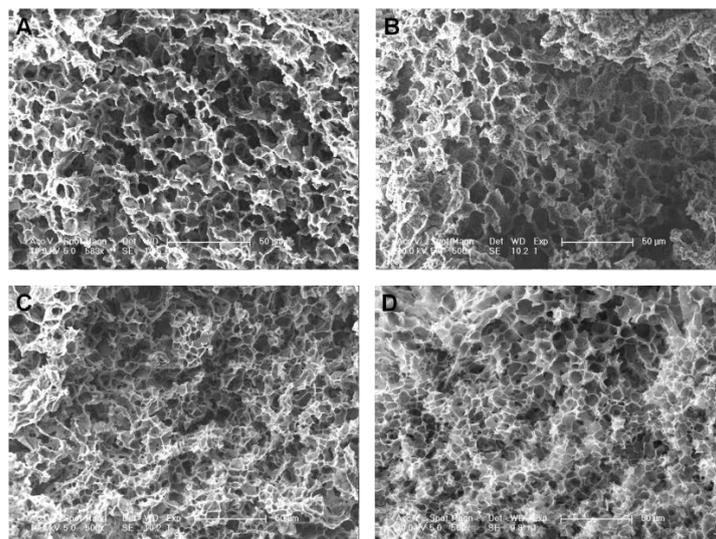


Figure S3. The SEMs of the lyophilized hydrogels. (A) G2, (B) G4, (C) G6, (D) G8. Scale bar = 50 μm.

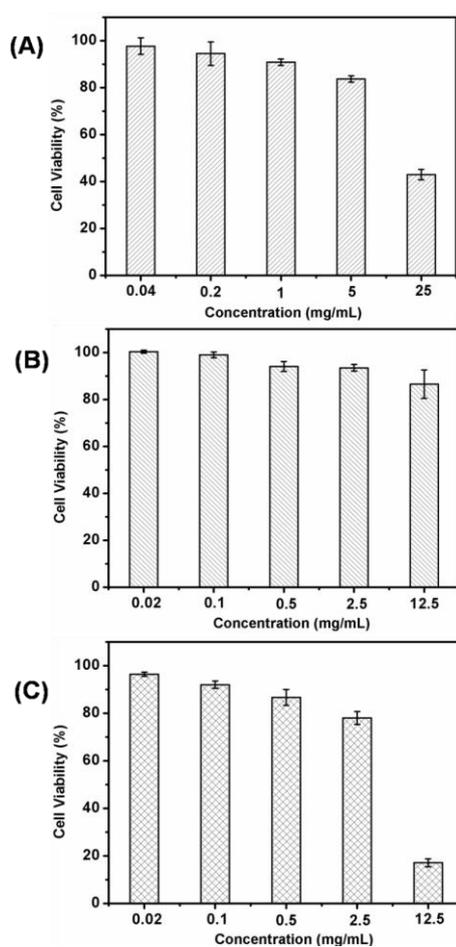


Figure S4. Cell viability of the freeze-dried powder of G2 hydrogel (A) at various concentrations as well as B-PEG10k-B (B) and α -CD (C) at equivalent concentrations against L929 cell line for 48 h.

Table S1. Preparation of PEG/ α -CD Hydrogels

gel precursor ^a	gel composition ^b	results		
	polymer (wt %)	gels	gelation time ^c (min)	gelation time ^d (min)
PEG10k	5	G1'	24	33
	10	G2'	7	20
	15	G3'	5	16
	20	G4'	4	12
PEG20k	5	G5'	10	15
	10	G6'	5	11
	15	G7'	2	9
	20	G8'	1	6

^a The gel precursor was composed of PEG.

^b The concentration of α -CD solution was kept at 10 wt % for all the studies.

^c The gelation time was measured at room temperature.

^d The gelation time was measured at 37 °C.