Electronic Supplementary Material (ESI) for Journal of Materials Chemistry B. This journal is © The Royal Society of Chemistry 2014

## **Electronic Supplementary Information for**

## Peptide Amphiphiles with Multifunctional Fragments Promoting Cellular Uptake and Endosomal Escape as Efficient Gene Vectors

Liang Luan<sup>a</sup>, Qingbin Meng<sup>a</sup>, Liang Xu<sup>a</sup>, Zhao Meng<sup>a</sup>, Husheng Yan<sup>\*b</sup> and Keliang Liu<sup>\*a</sup>

<sup>a</sup> Beijing Institute of Pharmacology & Toxicology, Beijing, 100850, PR China. Fax: +86-10-68211656; Tel.: +86-10-68169363; E-mail: keliangliu55@126.com (K. Liu)

<sup>b</sup> Key Laboratory of Functional Polymer Materials (Ministry of Education) and Institute of Polymer Chemistry,

Nankai University, and Collaborative Innovation Center of Chemical Science and Engineering (Tianjin), Tianjin

300071, PR China. Fax: +86-22-23503510; Tel.: +86-22-23503509; E-mail: yanhs@nankai.edu.cn (H. Yan)



**Fig. S1.** Agarose gel electrophoresis retardation assay with different N/P ratios for (A) TAT, (B) C<sub>18</sub>-TAT, (C) G(LLKK)<sub>3</sub>G, (D) C<sub>18</sub>- G(LLKK)<sub>3</sub>G, (E) C(LLKK)<sub>3</sub>C, and (F) C<sub>18</sub>-C(LLKK)<sub>3</sub>C peptides.



**Fig. S2**. Size distribution analysis of peptide/DNA nanoparticles. (A) G(LLKK)<sub>3</sub>G-TAT/DNA, (B) C(LLKK)<sub>3</sub>C-TAT/DNA, (C) C<sub>18</sub>-G(LLKK)<sub>3</sub>G-TAT/DNA, and (D) C<sub>18</sub>-C(LLKK)<sub>3</sub>C-TAT/DNA.



**Fig. S3**. Confocal microscopy images of the cellular uptake of the following peptide/plasmid DNA (pDNA) complexes labeled with YOYO-1 at a N/P ratio of 4 in 293T cells: (A) G(LLKK)<sub>3</sub>G-TAT/DNA, (B) C(LLKK)<sub>3</sub>C-TAT/DNA, (C) C<sub>18</sub>-G(LLKK)<sub>3</sub>G-TAT/DNA, and (D) C<sub>18</sub>-C(LLKK)<sub>3</sub>C-TAT/DNA. Nuclei are stained with DAPI. Scale bar: 20 μm.



**Fig. S4**. Confocal microscopy images of the cellular uptake of the following peptide/plasmid DNA (pDNA) complexes labeled with YOYO-1 at a N/P ratio of 4 in NIH-3T3 cells: (A) G(LLKK)<sub>3</sub>G-TAT/DNA, (B) C(LLKK)<sub>3</sub>C-TAT/DNA, (C) C<sub>18</sub>-G(LLKK)<sub>3</sub>G-TAT/DNA, and (D) C<sub>18</sub>-C(LLKK)<sub>3</sub>C-TAT/DNA. Nuclei are stained with DAPI. Scale bar: 20 μm.



**Fig. S5.** *In vitro* luciferase expression levels detected in 293T cells (A) and NIH-3T3 cells (B) treated with C(LLKK)<sub>3</sub>C-TAT and C<sub>18</sub>-C(LLKK)<sub>3</sub>C-TAT at N/P ratios ranging from 2 to 8 in the presence or absence of chloroquine (CQ). Data are shown as the mean  $\pm$  SD (n = 3).



**Fig. S6.** Liposome leakage assays were performed with peptide and peptide/DNA complexes (at a N/P ratio of 4 and a final concentration of 0.5  $\mu$ M) and calcein liposomes (POPC/cholesterol = 3:1) in 200 mM NaCl/citrate buffer (PH 5). Fluorescence due to calcein release from liposomes was measured at 515 nm and was plotted as a percentage of total fluorescence detected following treatment with 1% TritonX-100 (positive control). Data shown are the mean  $\pm$  SD (n = 3).

Peptide	α-helix content	
(each at 50 $\mu\text{M})$	50% TFE (%)	PBS (%)
G(LLKK)₃G	90.66	16.99
C(LLKK) <sub>3</sub> C	93.16	27.98
G(LLKK)₃G-TAT	70.33	9.24
C(LLKK) <sub>3</sub> C-TAT	86.48	19.47
C <sub>18</sub> -G(LLKK) <sub>3</sub> G	65.56	66.00
C <sub>18</sub> -C(LLKK) <sub>3</sub> C	84.28	89.82
C₁8-G(LLKK)₃G-TAT	70.93	42.60
C <sub>18</sub> -C(LLKK) <sub>3</sub> C-TAT	76.23	69.42

**Table S1.** The  $\alpha$ -helix content of the peptides studied in a 50% trifluoroethanol (TFE)/PBS solution and a PBS solution.