Supporting Information

A Recyclable Carbon Nanoparticles-Based Fluorescent Probe for Highly Selective and Sensitive Detection of Mercapto Biomolecules

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| Methods | Respone | pН | PL | PL | Circu- | Ref. |
|-------------------------|---------|----------|-------------------------|--------------------------|----------------|-----------|
| | time | range | Quenching efficiency | recovering efficiency | larity test | |
| C-dots-Hg ²⁺ | 5 min | 7.5~9.5 | ~60% | ~30% | N/A | [R1] |
| C-dots-Au NPs | 5 min | N/A | N/A | ~25% | N/A | [R2] |
| C-dots-Cu ²⁺ | 1 min | 5.0~6.0 | ~90% | ~80% | N/A | [R3] |
| GQDs-Hg ²⁺ | N/A | 7.0~7.4 | ~93% | ~95% | N/A | [R4] |
| GQDs-Ag NPs | 5 min | 6.5~9.5 | ~40% | N/A | N/A | [R5] |
| CNPs-Hg ²⁺ | 10 s | 5.0~10.0 | 98.7% | 93.2% | 10 | This work |

Table S1. CNPs based fluorescence sensor toward thiols compounds.

[R1] L. Zhou, Y. H. Lin, Z. Z. Huang, J. S. Ren and X. G. Qu, *Chem. Commun.*, 2012, **48**, 1147.

[R2] Y. P. Shi, Y. Pan, H. Zhang, Z. M. Zhang, M. J. Li, C. Q. Yi and M. S. Yang, *Biosens. Bioelectron.*, 2014, **56**, 39.

[R3] H. Z. Zheng, W. J. Bai, Y. J. Long, M. Gao and L. Y. Zhang, *Scientia Sinica Chimica.*, 2011, **41**, 1031.

[R4] Z. Z. Wu, W. Y. Li, J. Chen and C. Yu, *Talanta*, 2014, 119, 538.

[R5] X. Ran, H. J. Sun, F. Pu, J. S. Ren and X. G. Qu, Chem. Commun., 2013, 49, 1079.

| | Reaction temperature (°C) | Reaction time (h) | Citrate trisodium /Melamine | Quenching efficiency (%) | Recovering efficiency (%) | QY @330 nm | QY @340 nm | QY @350 nm |
|--------|---------------------------------|----------------------|-----------------------------------|--------------------------------|---------------------------------|------------------|------------------|------------------|
| CNPs-A | 160 | 12 | 2 | 97.7 | 77.8 | 0.31 | 0.31 | 0.33 |
| CNPs-B | 180 | 3 | 2 | 95.4 | 73.1 | 0.31 | 0.32 | 0.34 |
| CNPs-C | 180 | 6 | 2 | 97.6 | 74.5 | 0.35 | 0.35 | 0.36 |
| CNPs-D | 180 | 12 | 2 | 98.7 | 93.2 | 0.30 | 0.31 | 0.34 |
| CNPs-E | 180 | 24 | 2 | 95.0 | 91.1 | 0.27 | 0.29 | 0.31 |
| CNPs-F | 180 | 12 | 1 | 97.6 | 75.6 | 0.33 | 0.35 | 0.36 |
| CNPs-G | 180 | 12 | 0.5 | 94.4 | 81.8 | 0.28 | 0.30 | 0.32 |

Table S2. The influence of synthesis parameters on the properties of CNPs.

Data notes:

1. Quenching efficiency and recovering efficiency were calculated from the following equations:

Quenching efficiency (%) = $(I_0-I_{Hg})*100/I_0$;

Recovering efficiency (%) = I_{Cys} *100/ I_0

where I_0 is the PL intensity of CNPs at 437 nm in the absence of Hg²⁺ and Cys. I_{Hg} and I_{Cys} refer to the PL intensity of CNPs at 437 nm in the presence of 10 μ M Hg²⁺ and 20 μ M Cys, respectively.

2. The QY was determined by the reference of quinine sulfate (QY = 0.54 when in 0.1 M H₂SO₄ aqueous solution) at different excitation wavelengths (330, 340 and 350 nm).



Fig. S1. Zeta-potential spectrum of CNPs.



Fig. S2. PL spectra of CNPs aqueous solution under excitation with different wavelengths from 275 to 450 nm.



Fig. S3. The PL intensities of CNPs aqeuous solution at 437 nm at different NaCl concentrations (0, 0.2, 0.4, 0.6, 0.8, 1.0 mol/L).



Fig. S4. Photostability of CNPs aqueous solution evaluated by the 350 nm xenon lamp equipped in the FL spectrometer. F_0 and F represent the fluorescent intensity of CNPs at 437 nm before and after irradiation, respectively.



Fig. S5. Fluorescence decay curve (yellow line) of CNPs recorded at 437 nm with the excitation of 355 nm. Blue line: the instrument noise; and red line: fitting of the fluorescence decay curve. Fit = $A+B_1exp(-t/\tau_1)$, ($\tau_1=7.0$ ns).



Fig. S6. Time-dependent PL spectra of CNPs upon addition of 10 μ M Hg²⁺ in HEPES buffer solution at room temperature (λ_{ex} =350 nm).



Fig. S7. Time-dependent PL spectra of CNPs-Hg²⁺ system upon addition of 10 μ M Cys in HEPES buffer solution at room temperature (λ_{ex} =350 nm).



Fig. S8. Normalized UV-vis spectra of CNPs prepared under different conditions.



Fig. S9A. (a) PL spectra of CNPs-A aqueous solution under excitation with different wavelengths from 300 to 400 nm. (b) Dependence of PL spectra of CNPs-A in HEPES buffer solution (10 mM, pH 7.2) on the gradual addition of Hg²⁺ from 0 to 10 μ M and (c) following addition of Cys from 4 to 20 μ M. (λ_{ex} =350 nm). (d) Dependence of PL intensity at 437 nm with incremental addition of Hg²⁺ and Cys.



Fig. S9B. (a) PL spectra of CNPs-B aqueous solution under excitation with different wavelengths from 300 to 400 nm. (b) Dependence of PL spectra of CNPs-B in HEPES buffer solution (10 mM, pH 7.2) on the gradual addition of Hg²⁺ from 0 to 10 μ M and (c) following addition of Cys from 4 to 20 μ M. (λ_{ex} =350 nm). (d) Dependence of PL intensity at 437 nm with incremental addition of Hg²⁺ and Cys.



Fig. S9C. (a) PL spectra of CNPs-C aqueous solution under excitation with different wavelengths from 300 to 400 nm. (b) Dependence of PL spectra of CNPs-C in HEPES buffer solution (10 mM, pH 7.2) on the gradual addition of Hg²⁺ from 0 to 10 μ M and (c) following addition of Cys from 4 to 20 μ M. (λ_{ex} =350 nm). (d) Dependence of PL intensity at 437 nm with incremental addition of Hg²⁺ and Cys.



Fig. S9D. (a) PL spectra of CNPs-D aqueous solution under excitation with different wavelengths from 300 to 400 nm. (b) Dependence of PL spectra of CNPs-D in HEPES buffer solution (10 mM, pH 7.2) on the gradual addition of Hg²⁺ from 0 to 10 μ M and (c) following addition of Cys from 4 to 20 μ M. (λ_{ex} =350 nm). (d) Dependence of PL intensity at 437 nm with incremental addition of Hg²⁺ and Cys.



Fig. S9E. (a) PL spectra of CNPs-E aqueous solution under excitation with different wavelengths from 300 to 400 nm. (b) Dependence of PL spectra of CNPs-E in HEPES buffer solution (10 mM, pH 7.2) on the gradual addition of Hg²⁺ from 0 to 10 μ M and (c) following addition of Cys from 4 to 20 μ M. (λ_{ex} =350 nm). (d) Dependence of PL intensity at 437 nm with incremental addition of Hg²⁺ and Cys.



Fig. S9F. (a) PL spectra of CNPs-F aqueous solution under excitation with different wavelengths from 300 to 400 nm. (b) Dependence of PL spectra of CNPs-F in HEPES buffer solution (10 mM, pH 7.2) on the gradual addition of Hg²⁺ from 0 to 10 μ M and (c) following addition of Cys from 4 to 20 μ M. (λ_{ex} =350 nm). (d) Dependence of PL intensity at 437 nm with incremental addition of Hg²⁺ and Cys.



Fig. S9G. (a) PL spectra of CNPs-G aqueous solution under excitation with different wavelengths from 300 to 400 nm. (b) Dependence of PL spectra of CNPs-G in HEPES buffer solution (10 mM, pH 7.2) on the gradual addition of Hg²⁺ from 0 to 10 μ M and (c) following addition of Cys from 4 to 20 μ M. (λ_{ex} =350 nm). (d) Dependence of PL intensity at 437 nm with incremental addition of Hg²⁺ and Cys.



Fig. S10. (a) Absorption spectra of CNPs (black line), CNPs-Hg²⁺ (red line) and CNPs-Hg²⁺-Cys mixture (blue line) in HEPES buffer solutions (10 mM, pH 7.2). (b) Absorption spectra of 10 μ M HgCl₂ (black line), 20 μ M Cys (red line) and the mixture of HgCl₂ and Cys (blue line) in HEPES buffer solutions (10 mM, pH 7.2).



Fig. S11. (a) PL spectra and (b) the corresponding PL intensities at 437 nm of CNPs-Hg²⁺ solutions with the addition of amino acids, DNA and RNA in HEPES buffer solution (λ_{ex} =350 nm). The concentration of these analytes was 20 µM.



Fig. S12. Selectivity of CNPs-Hg²⁺ system to Cys in the presence of 20 μ M Cl⁻, histidine, Ag⁺ and S²⁻ in HEPES buffer solution (10 mM, pH 7.2). Blank refer to the free CNPs-Hg²⁺ aqueous solution.



Fig. S13. Confocal laser scanning microscopy z-stack images of A549 cell incubated with CNPs (λ_{ex} =405 nm). (a) The overlay of bright field and fluorescence images. (b)~(l) Z-stack scanning mode images from the top to the bottom of cells. The scale bar is 50 µm.



Fig. S14. Confocal microscopy images of multiple cross-sections exhibiting various locations of CNPs within A549 cells.