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## **Supporting Information**

## Dual-functional carbon dots-silver@ zinc oxide nanocomposite:

## In vitro evaluation of cellular uptake and apoptosis induction

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## Fluorescence quantum yield determination

Fluorescence quantum yield of samples in water was calculated according to the following equation using quinine sulphate as a standard ( $Q_R = 0.54$  at 360 nm):

 $Q = Q_R \times \underline{I} \times \underline{A}_R \times \underline{\eta}^2$  $I_R \quad A \quad \eta^2_R$ 

Where Q represents the quantum yield of desired sample, I is the measured integrated emission intensity (area under the curve), refractive index being  $\eta$  and A stands for optical density. Subscript R signifies the reference fluorophore of known quantum yield. To eliminate the likelihood of re-absorption effects, the absorption in the 10 mm cuvette was always kept under 0.1 at the excitation wavelength. The emission range of the samples used for area calculation was kept between 375-700 nm.

Sample	Integrated emission intensity <i>(I)</i>	Absorbance at 360 nm <i>(A)</i>	Refractive index of solvent (η)	Quantum yield at 360 nm <i>(Q)</i>
Quinine sulphate	658172	0.0728	1.33	0.54 (known)
CDs	92856	0.0823	1.33	0.0674
CD-Ag@ZnO	88239	0.0821	1.33	0.0642

Table	<b>S1</b> .	Quantum	yield	measurements.
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A



Fig. S1. Fluorescence decay curve of (A) CDs and (B) CD-Ag@ZnO ( $\lambda_{ex} = 320$  nm;  $\lambda_{em} = 400$  nm).

Sample	<b>a</b> <sub>1</sub>	$\tau_1(ns)$	<b>a</b> <sub>2</sub>	$\tau_2(ns)$	<b>a</b> <sub>3</sub>	$\tau_3(ns)$	$\tau_{av}(ns)$	$\chi^2$
CD	0.5409	3.306	0.1521	0.590	0.3070	10.833	5.204	1.169
CD-Ag@ZnO	0.5323	3.334	0.1634	0.644	0.3043	10.369	5.035	1.157

 Table S2. Tabular representation of fluorescence lifetime calculation.

Average lifetime  $(\tau_{av})$  was calculated by using the following equation:

$$\tau_{av} = a_1\tau_1 + a_2\tau_2 + a_3\tau_3$$

where  $\tau_1$ ,  $\tau_2$ ,  $\tau_3$  were the first, second and third component of the decay time of CDs and  $a_1$ ,  $a_2$ ,  $a_3$  were the corresponding relative weightings( emission %) of these components, respectively.



Fig. S2. EDAX spectrum and elemental composition of CD-Ag@ZnO NC.



Fig. S3. N<sub>2</sub> adsorption–desorption isotherms for Ag@ZnO.



Fig. S4. TEM image of CDs.



Fig. S5. XRD spectra of CD-Ag@ZnO NC.



Fig. S6. FTIR spectra of CD-Ag@ZnO NC.



Fig. S7. Bright field microscopic images. Scale bar:  $400 \ \mu m$ .



**Fig. S8.** Fluorescence microscopic images depicting the cellular uptake in L-132 cells. a,e) CDs and b,f) 20  $\mu$ g/mL, c,g) 50  $\mu$ g/mL , d,h) 70  $\mu$ g/mL CD-Ag@ZnO NC treated L-132 cells. The images in the upper panel are corresponding bright field images.



Intensity\_MC\_Ch02

**Fig. S9.** Flow cytometric analysis of ROS production in MCF-7 and A549 cells. Upper panel: (a) untreated and (b) 20  $\mu$ g/mL, (c) 50  $\mu$ g/mL, (d) 70  $\mu$ g/mL Ag@ZnO treated MCF-7 cells. Lower panel: (e) untreated and (f) 20  $\mu$ g/mL, (g) 50  $\mu$ g/mL, (h) 70  $\mu$ g/mL Ag@ZnO treated A549 cells.

Gene	Primers
Beta-actin	Forward: 5' CTGTCTGGCGGCACCACCAT 3' Reverse : 5' GCAACTAAGTCATAGTCCGC 3'
p53	Forward: 5' TGGCCCCTCCTCAGCATCTTAT 3' Reverse : 5' GTTGGGCAGTGCTCGCTTAGTG 3'
Caspase 3	Forward : 5' TTCAGAGGGGGATCGTTGTAGAAGTC 3' Reverse : 5' CAAGCTTGTCGGCATACTGTTTCAG 3'
C-myc	Forward : 5' CCAGGACTGTATGTGGAGCG 3' Reverse : 5' CTTGAGGACCAGTGGGCTGT 3'
Bax	Forward : 5' AAGCTGAGCGAGTGTCTCAAGCGC 3' Reverse : 5' TCCCGCCACAAAGATGGTCACG 3'
Bad	Forward : 5' CCTTTAAGAAGGGACTTCCTCGCC 3' Reverse : 5' ACTTCCGATGGGACCAAGCCTTCC 3'
Bcl-xl	Forward : 5' ATGGCAGCAGTAAAGCAAGC 3' Reverse : 5' CGGAAGAGTTCATTCACTACCTGT 3'
Bcl-2	Forward : 5' TCCGCATCAGGAAGGCTAGA 3' Reverse : 5' AGGACCAGGCCTCCAAGCT 3'

Table S3. Forward and reverse primer sequences for various apoptotic signaling genes.